

Catalyst-free visible-light mediated selective oxidation of sulfides into sulfoxides under clean conditions

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General information

Experimental section

All chemicals were purchased from commercial sources and used without further purification. Thin-layer chromatography (TLC) was conducted on silica-gel 60 F254 plates (Merck KGaA). ^1H NMR spectra were recorded at 500 MHz and ^{13}C NMR spectra were recorded at 125 MHz using a Bruker Avance 400 spectrometer at ambient temperature (25 °C) with TMS as an internal standard. GC-MS were obtained by EI on a Shimadzu GC-MS 2010, Conditions: Flow ripples:1 mL/min; column oven initial temperature 50 °C; injection port temperature: 300 °C; temperature range: 50~300 °C. XRD experiments were conducted by using an XRD-6000 with a copper-K α radiation source ($\lambda = 1.54056 \text{ \AA}$). TGA was performed on a TGA Q500 thermal analyses system with a heating rate of 10 °C/min from 25 to 800 °C under N_2 atmosphere. The UV-Vis DRS spectra were recorded on a UH4150 Spectrophotometer and fluorescence spectra was recorded on a F-7000 FL Spectrophotometer. The UV–Vis absorption spectrum of thioanisole was recorded in a mix solution of MeOH and H_2O ($v/v=2/1$) using a Shimadzu 1800 PC spectrophotometer in the spectral region of 250–800 nm.

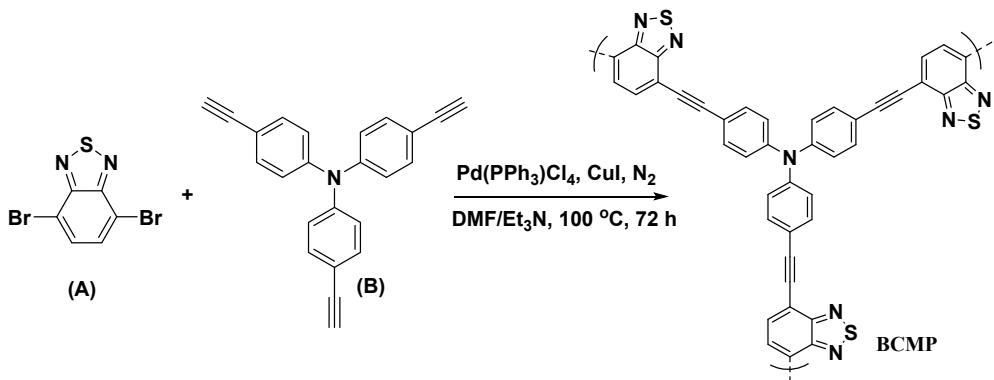
Computational details

Geometrical optimization of all stationary points along the reaction paths was performed at the B3LYP-D3/6-311+G (d) theoretical level which dispersion corrections are comprised using the Grimme's D3 model¹⁻⁵. Frequency analysis were carried out at the same level to classify the located stationary points as minima (no imaginary frequency) and transition states (only one imaginary frequency) and also to obtain zero point energies (ZPES) and the thermodynamic correctional data. Moreover, the intrinsic reaction coordinate (IRC)⁶ pathways computations were also carried out for all transition states to determine whether these transition states can connect the reactants, intermediates or products. The polarized continuum model (PCM)⁷ was used was used in self-consistent reaction field (SCRF)⁸ for simulating the solvent environment consisted of MeOH and H_2O ($v/v=2/1$), where the solvent parameters eps and epsinf were set as 47.860 and 1.7697, respectively. The relax scanning was performed using the PM6-D3 method⁹ with the distance decreasing from 3.0 Å to 1.5

Å (stepsize=-0.1 Å). It should be noted that the thermal corrected data of **1a*** was carried out in gas state due to huge time consuming for frequency analysis of excited state in simulated solvent environment. Besides, the geometrical optimization and frequency analysis of triplet **1a**^{3,*} were performed using the unrestricted DFT method. The enthalpy of singlet oxygen was obtained by adding the experimental singlet-triplet energy difference (22.5 kcal/mol)¹⁰ to the calculated enthalpy of triplet oxygen, as previously suggested by McKee¹¹, since calculation of ¹O₂ required an extremely complex wavefunction. All the energies listed in this paper were corrected by thermal corrections, wherein the scale factors 0.9882, 1.0124 and 1.0174 were used for the corrections of zero-point energy, enthalpy and entropy, respectively¹². All of the quantum chemical calculations were performed using the Gaussian 16 program package¹³. The S₁/T₁ minimal energy crossing point (MECP) of **1a*** was located by the sobMECP program at the B3LYP-D3/6-311+G (d) theoretical level with the default settings^{14,15}.

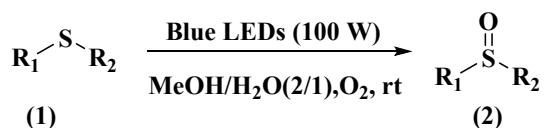
General procedures for preparing BCMP and sulfoxides

Procedures for the Synthesis of BCMP: To a 50 mL Pyrex tube added a solution of **A** (147 mg, 0.5 mmol), **B** (159 mg, 0.5 mmol), tetrakis(triphenylphosphine)-palladium(0) (40 mg) and copper(I) iodide (20 mg) in anhydrous DMF (5 mL) and Et₃N (5 mL) under nitrogen atmosphere. The reaction mixture was heated to 100 °C and stirred for 72 h. Afterwards, the reaction system was cooled to room temperature, and the precipitates (crude products) was filtered and removed the unreacted monomers or catalyst by washing the residues with CHCl₃, H₂O, MeOH, and acetone successively. The resulted solids were further purified by Soxhlet extraction with MeOH and CHCl₃ for 24 h respectively. The obtained product was dried under vacuum for 24 h at 60 °C and afforded the desired products as dark brown powder.



Scheme S1 Synthetic route of preparing BCMP

General procedures for the synthesis of sulfoxides: To a 25 mL shrek tube charged with O₂ balloon added a solution of sulfides (**1**, 0.2 mmol) in MeOH (6 mL) and H₂O (3 mL), the resulting mixture was subjected to visible-light irradiation (100 W blue LEDs) until totally conversion of **1** (monitored by TLC). Subsequently, organic solvent was removed under vacuum and the resulting mixture was extracted with CH₂Cl₂ (3*10 mL). The organic layers were combined and dried with anhydrous Na₂SO₄. The desired sulfoxide was afforded after removing Na₂SO₄ and CH₂Cl₂ by evaporation under vacuum or by flash chromatography (200-300 mesh silica gel) with ethyl acetate/hexane(*v/v*=1:3) as eluent.



Phenyl methyl sulfoxide(2a): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.72-2.73 (3H, d, -CH₃), 7.51-7.66 (5H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 43.93, 123.47, 129.36, 131.05, 145.60.

4-NH₂-Phenyl methyl sulfoxide (2b): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.69 (3H, d, -CH₃), 3.63-4.11 (2H, br, -NH₂), 6.73-6.75 (2H, d, Ph-H), 7.42-7.44 (2H, d, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 43.59, 115.02, 125.77, 132.50, 149.99.

4-OMe-Phenyl methyl sulfoxide (2c): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.60 (3H, s, -CH₃), 3.74 (3H, s, -OCH₃), 6.92-6.94 (2H, d, Ph-H), 7.49-7.51 (2H, d, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 43.80, 55.46, 114.78, 125.41, 136.30, 161.87.

3-OMe-Phenyl methyl sulfoxide (2d): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.72-2.73

(3H, t, -CH₃), 3.84-3.85 (3H, t, -OCH₃), 7.00-7.43 (4H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 43.81, 55.44, 107.90, 115.40, 117.17, 130.26, 146.95, 160.32.

4-F-Phenyl methyl sulfoxide (2e): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.65 (3H, s, -CH₃), 7.14-7.17 (2H, m, Ph-H), 7.58-7.60 (2H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 44.09, 116.59 (J=90.0 Hz), 125.82 (J=35.0 Hz), 141.03 (J=10.0 Hz), 163.28, 165.27.

4-Cl-Phenyl methyl sulfoxide (2f): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.73 (3H, d, -CH₃), 7.50-7.53 (2H, m, Ph-H), 7.59-7.61 (2H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 44.06, 124.99, 129.66, 137.23, 144.20.

4-Br-Phenyl methyl sulfoxide (2g): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.73 (3H, s, -CH₃), 7.52-7.54 (2H, m, Ph-H), 7.67-7.69 (2H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 44.01, 125.17, 125.46, 132.58, 144.85.

4-NO₂-Phenyl methyl sulfoxide (2h): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.82 (3H, s, -CH₃), 7.85-7.87 (2H, d, Ph-H), 8.40-8.42 (2H, m, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 43.85, 124.51, 124.71, 149.49, 153.18.

2-Cl-Phenyl methyl sulfoxide (2i): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.83 (3H, s, -CH₃), 7.39-7.46 (2H, m, Ph-H), 7.47-7.56 (1H, t, Ph-H), 7.95-7.97 (1H, t, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 41.69, 125.30, 128.18, 129.77, 129.79, 132.02, 143.58.

2-Br-Phenyl methyl sulfoxide (2j): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 2.71 (3H, s, -CH₃), 7.27-7.28 (1H, m, Ph-H), 7.44-7.49 (2H, m, Ph-H), 7.82-7.85 (1H, t, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 41.79, 118.31, 125.53, 128.67, 132.27, 132.84, 145.13.

Phenyl ethyl sulfoxide (2k): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 1.09-1.12 (3H, t, -CH₃), 2.66-2.84 (2H, m, -CH₂CH₃), 7.40-7.45 (3H, m, Ph-H), 7.52-7.53 (2H, d, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 5.91, 50.19, 124.11, 129.12, 130.92, 143.12.

Phenyl cyclopropyl sulfoxide (2l): ¹H NMR (500 MHz, CDCl₃): δ (ppm) 0.88-1.01 (3H, m, -cyclopropyl-H), 1.18-1.19 (1H, m, -cyclopropyl-H), 2.23-2.26 (1H, m, -cyclopropyl-H), 7.48-7.50 (3H, m, Ph-H), 7.65 (2H, s, Ph-H); ¹³C NMR (125 MHz, CDCl₃): δ (ppm) 2.25, 3.33, 33.59, 123.86, 129.07, 130.84, 144.69.

Phenyl benzyl sulfoxide (2m): ^1H NMR (500 MHz, CDCl_3): δ (ppm) 3.97-4.08 (2H, m, $-\text{PhCH}_2-$), 6.96-6.97 (2H, d, Ph- \underline{H}), 7.21-7.26 (3H, m, Ph- \underline{H}), 7.35-7.44 (5H, m, Ph- \underline{H}); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 63.48, 124.43, 128.86, 128.45, 128.88, 129.11, 130.39, 131.21, 142.67.

(4-(phenylsulfinyl)phenyl)methanol (2n): ^1H NMR (500 MHz, CDCl_3): δ (ppm) 4.67 (2H, s, $-\text{PhCH}_2-$), 7.40-7.45 (5H, m, Ph- \underline{H}), 7.53-7.55 (2H, d, Ph- \underline{H}), 7.58-7.60 (2H, m, Ph- \underline{H}); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 64.20, 124.76, 125.04, 127.58, 129.39, 131.18, 143.84, 144.94, 145.10.

Dibenzyl sulfoxide (2o): ^1H NMR (500 MHz, CDCl_3): δ (ppm) 3.86-3.94 (4H, q, $-\text{PhCH}_2-$), 7.28-7.30 (4H, m, Ph- \underline{H}), 7.35-7.40 (6H, m, Ph- \underline{H}); ^{13}C NMR (125 MHz, CDCl_3): δ (ppm) 57.31, 128.41, 129.00, 130.12, 130.18.

Sulindac (3a): ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ (ppm) 2.16 (3H, s, $-\text{CH}_3$), 2.83 (3H, s, $-\text{SOCH}_3$), 3.59 (2H, s, $-\text{CH}_2-$), 6.71-7.04 (3H, Ph- \underline{H}), 7.17 (1H, s, $\text{C}=\text{CH}$), 7.19-7.80 (4H, Ph- \underline{H}), 12.46 (1H, s, $-\text{COOH}$); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ (ppm) 10.73, 31.55, 43.58, 106.45 ($J=95.0$ Hz), 110.82 ($J=90.0$ Hz), 124.4, 130.41, 133.01, 138.97, 140.78, 146.74, 162.02, 163.96, 172.05.

Oxfendazole (3b): ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ (ppm) 3.80 (3H, s, $-\text{OCH}_3$), 7.41-7.56 (5H, m, Ph- \underline{H}), 7.68-7.80 (3H, m, Ph- \underline{H}), 11.98 (2H, br, $-\text{NH}$); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ (ppm) 53.13, 118.19, 124.49, 129.80, 131.15, 138.29, 147.14, 149.38, 154.74.

Oxfendazole (3c): ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ (ppm) 0.96 (3H, t, $-\text{CH}_3$), 1.47-1.63 (2H, m, $-\text{CH}_2-$), 2.77-2.84 (2H, m, $-\text{CH}_2-$), 2.80 (3H, s, $-\text{OCH}_3$), 7.34-7.73 (3H, m, $-\text{Ar}-\underline{H}$), 11.95 (2H, br, $-\text{NH}$); ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ (ppm) 13.42, 15.89, 53.11, 58.65, 117.25, 136.59, 149.08, 154.85.

Structure characterization spectra of BCPM and sulfoxides

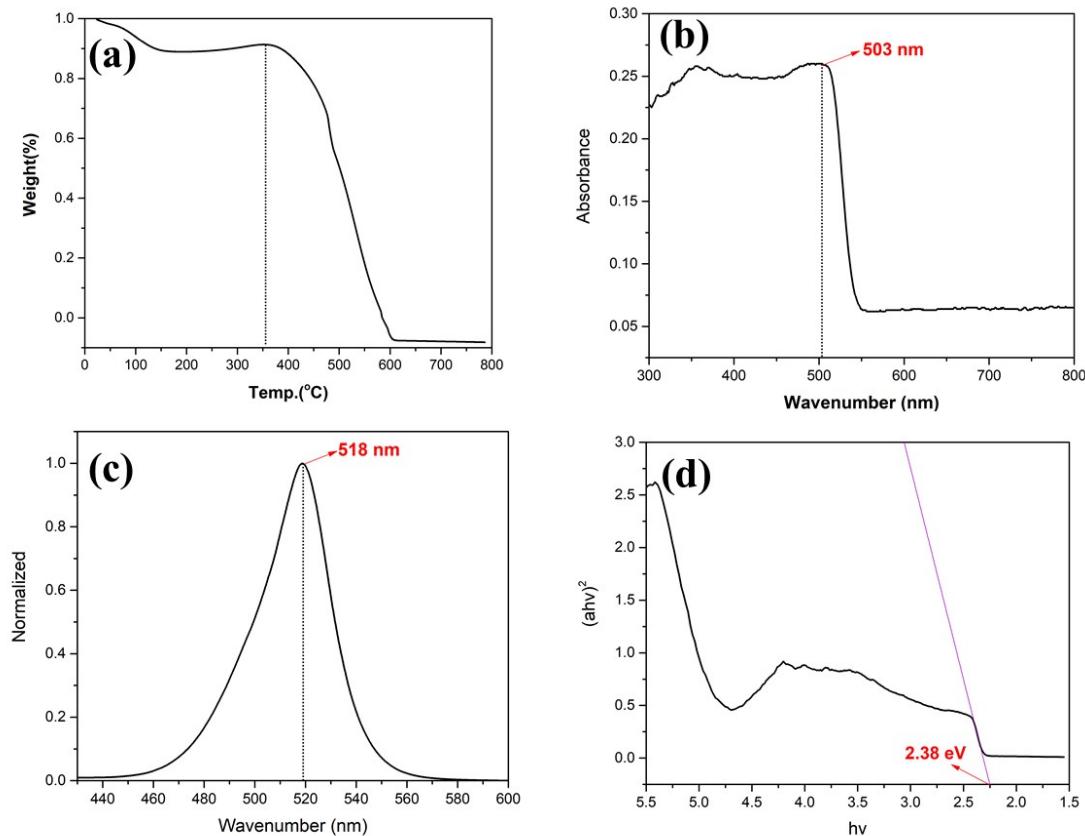


Fig. S1 Structure characterization spectra of BCPM. (a) TGA curve; (b)UV-Vis DRS spectra; (c) PL emission spectra; (d) Tauc fitting curve.

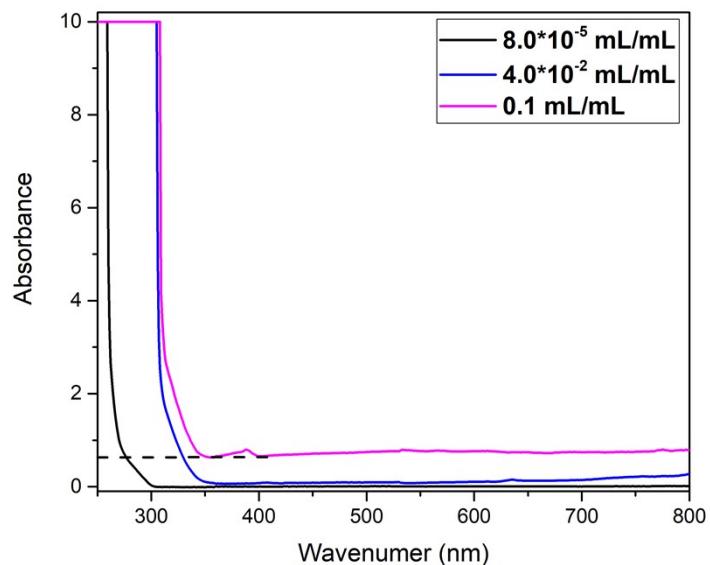
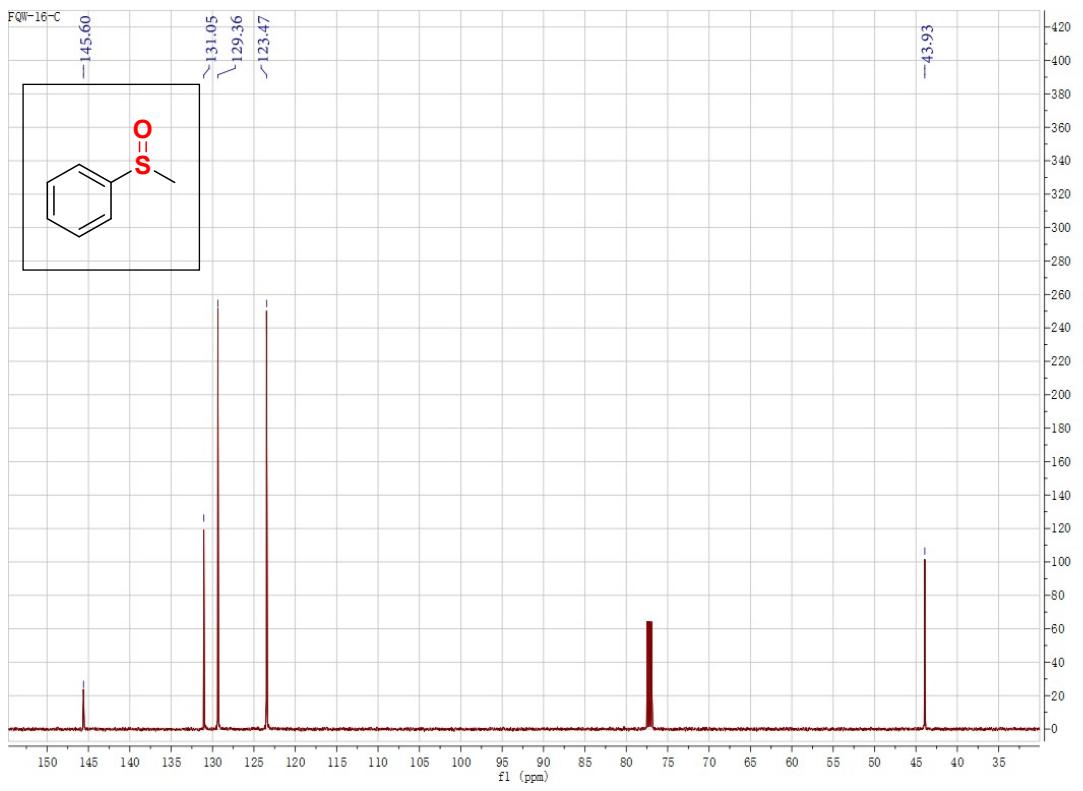
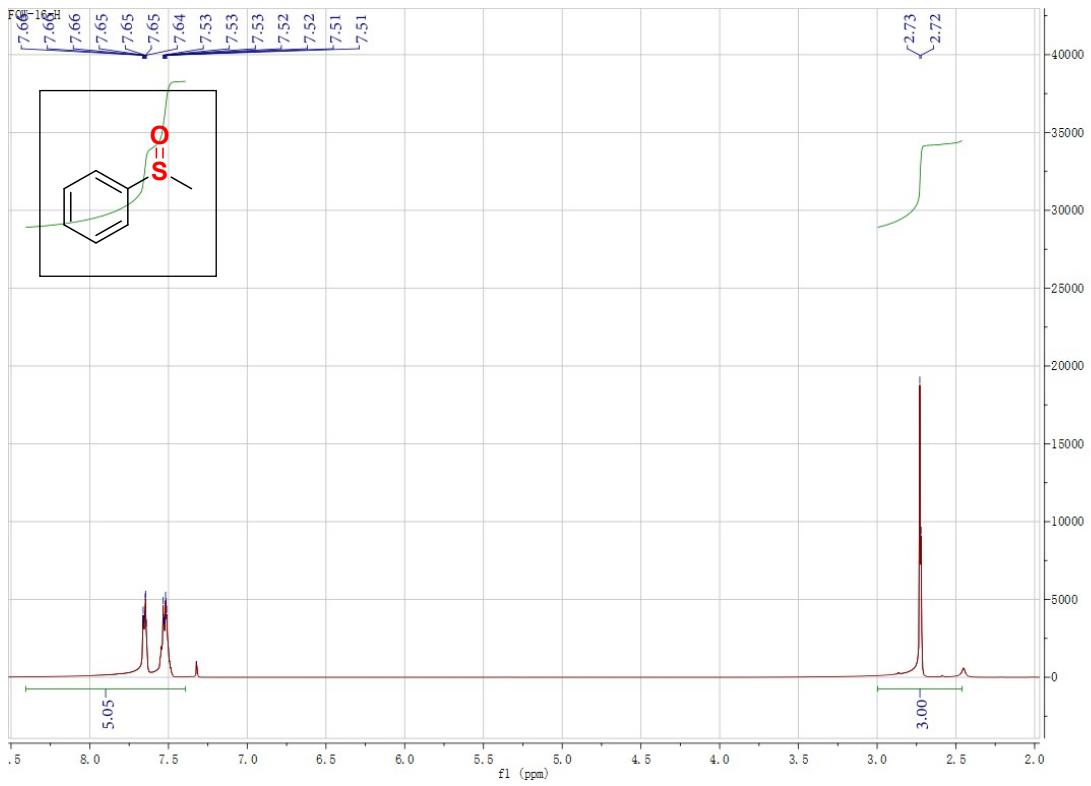
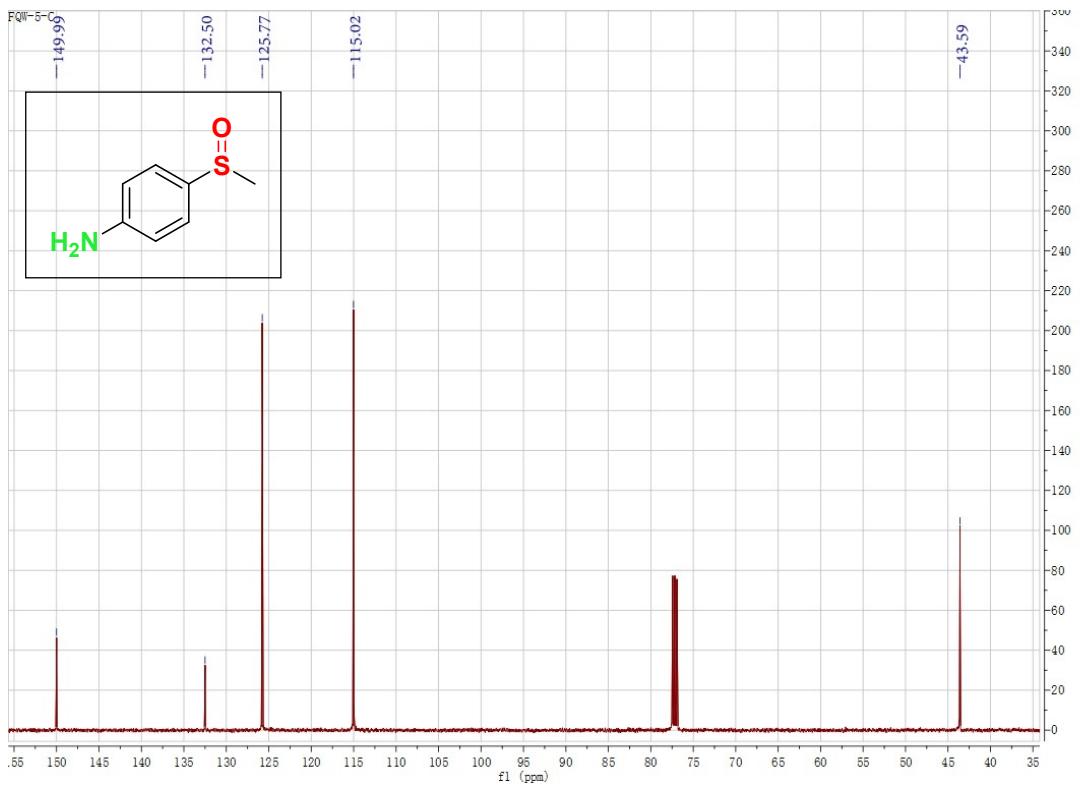
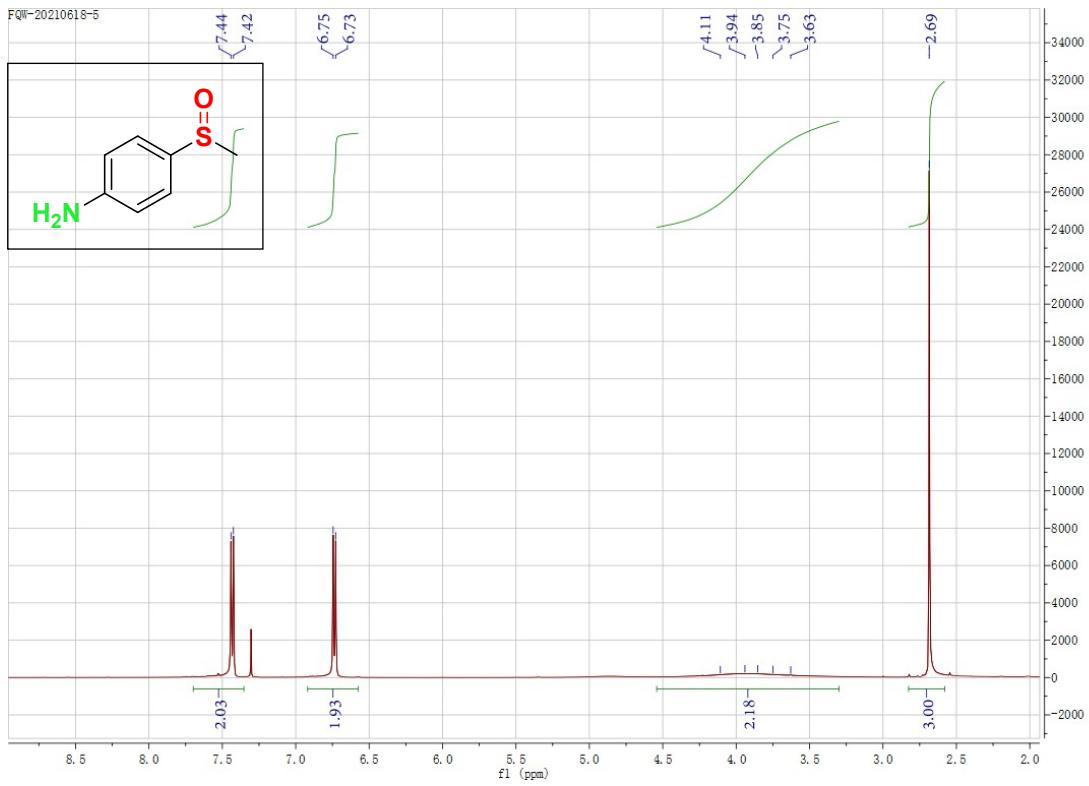
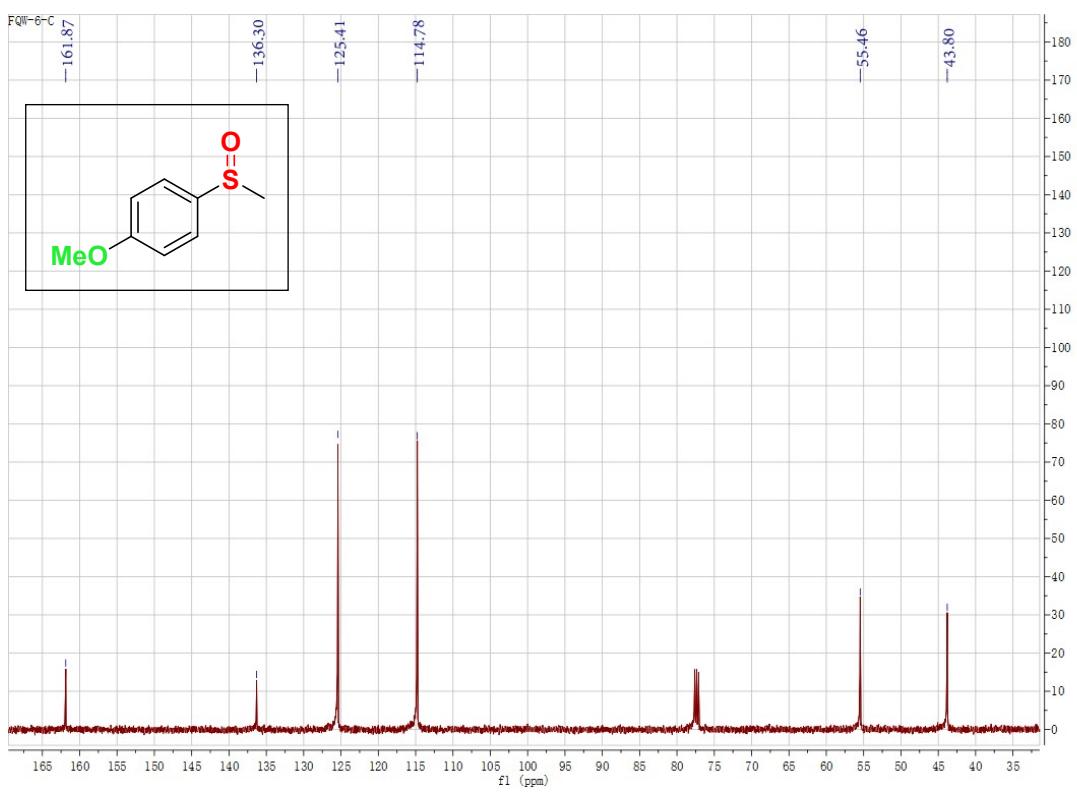
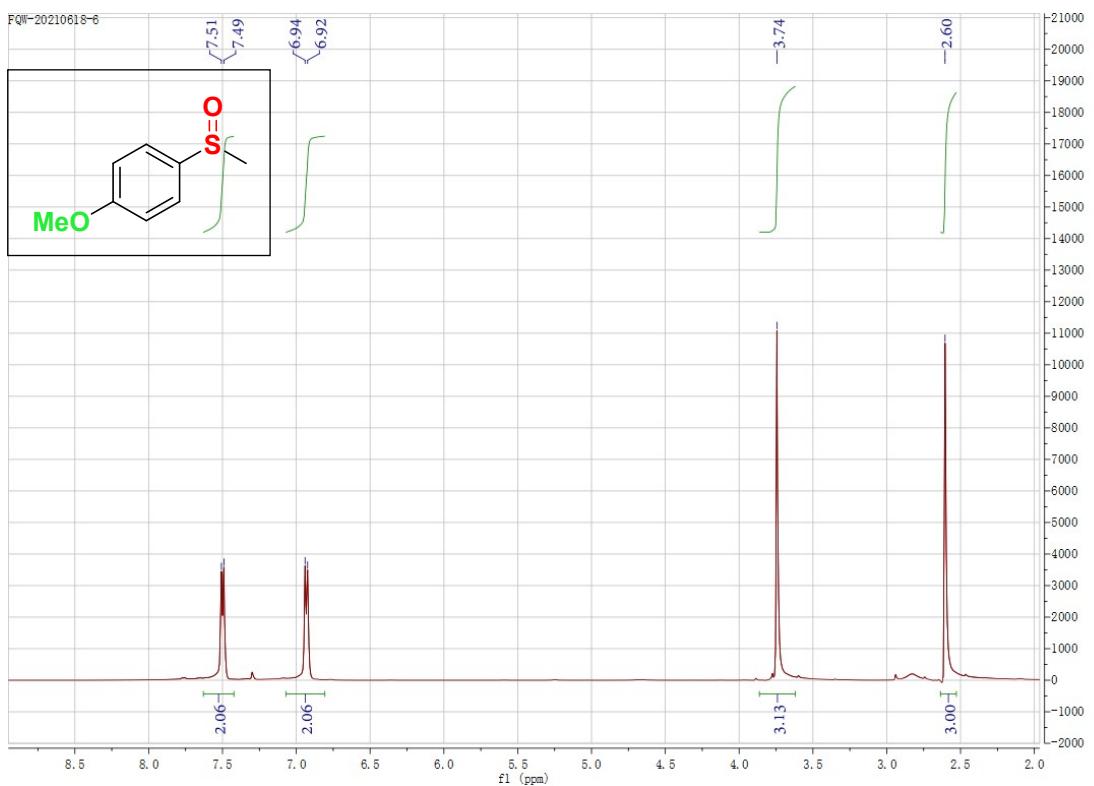
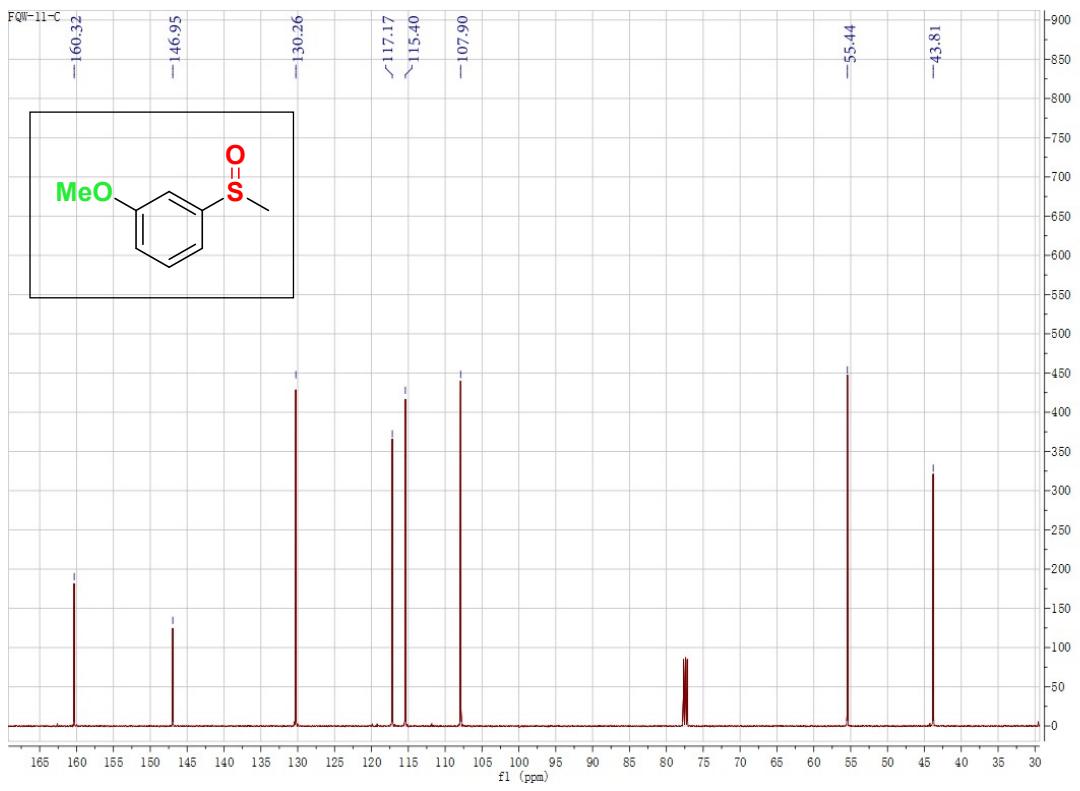
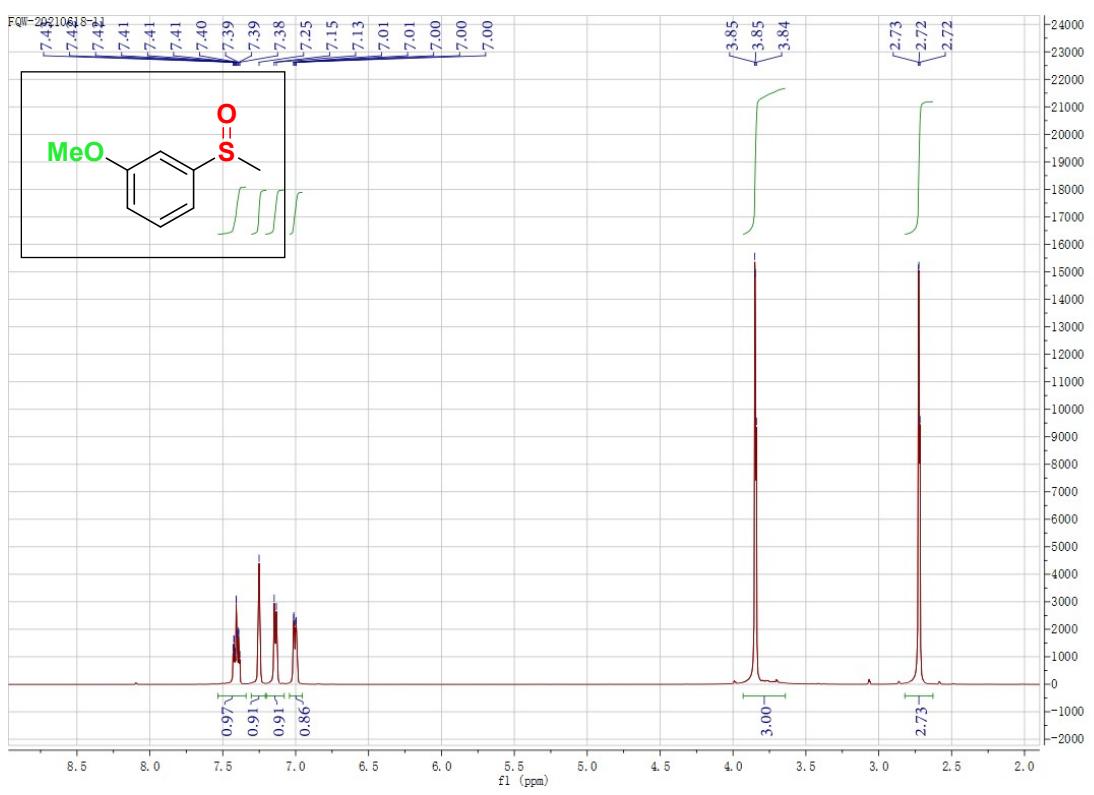


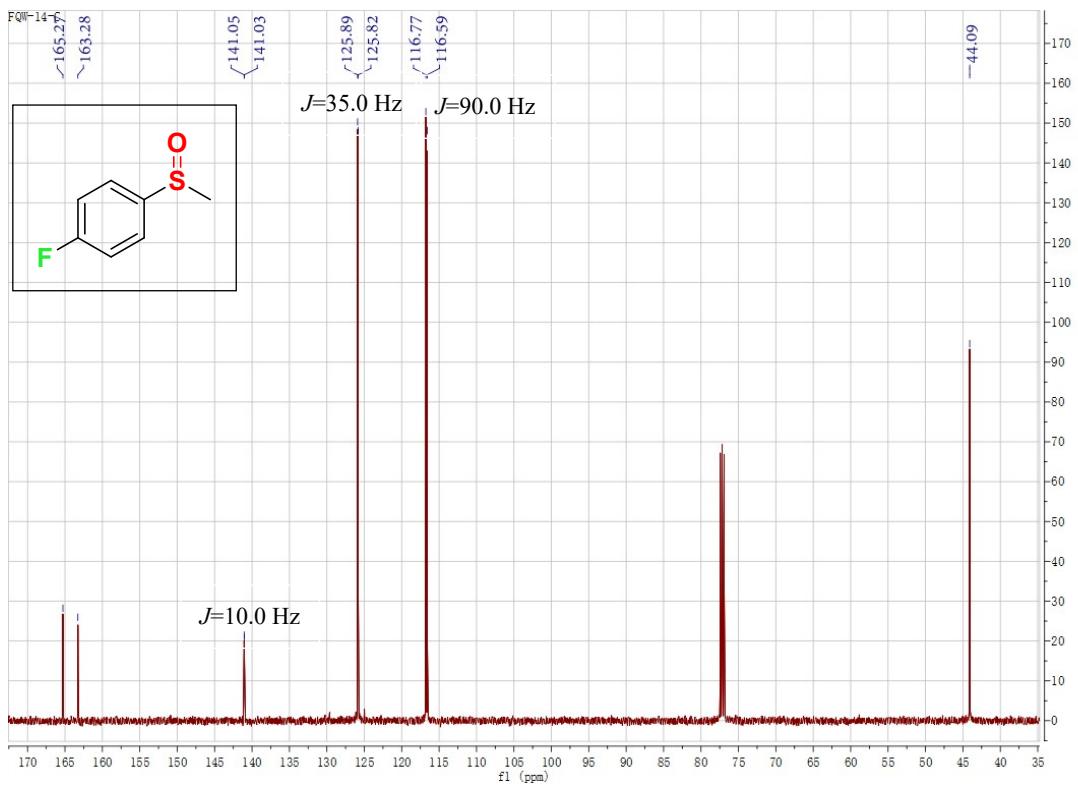
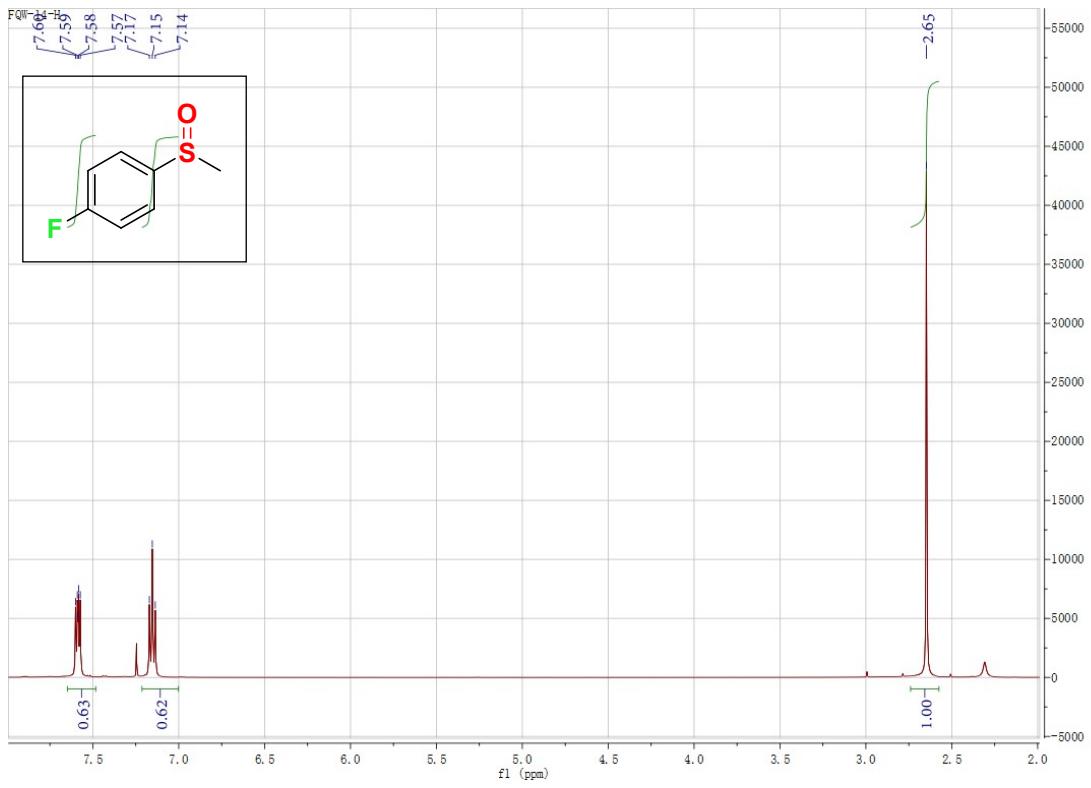
Fig.S2 UV-Vis spectra of thioanisole with different concentrations

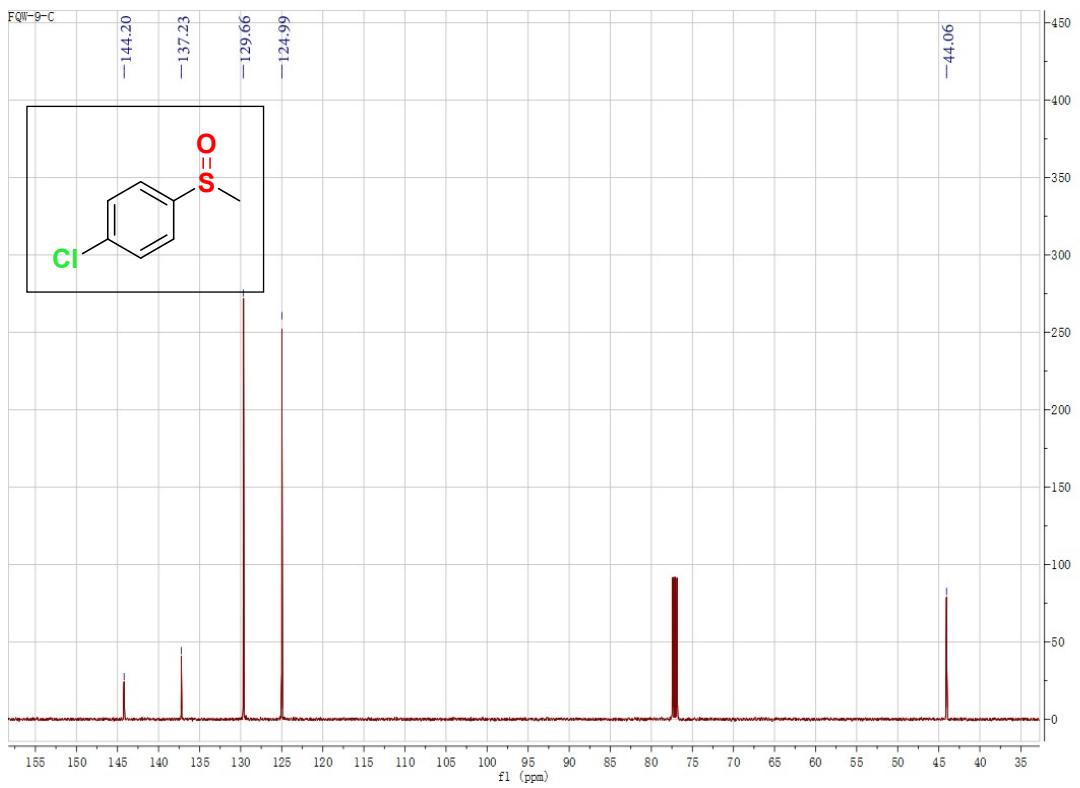
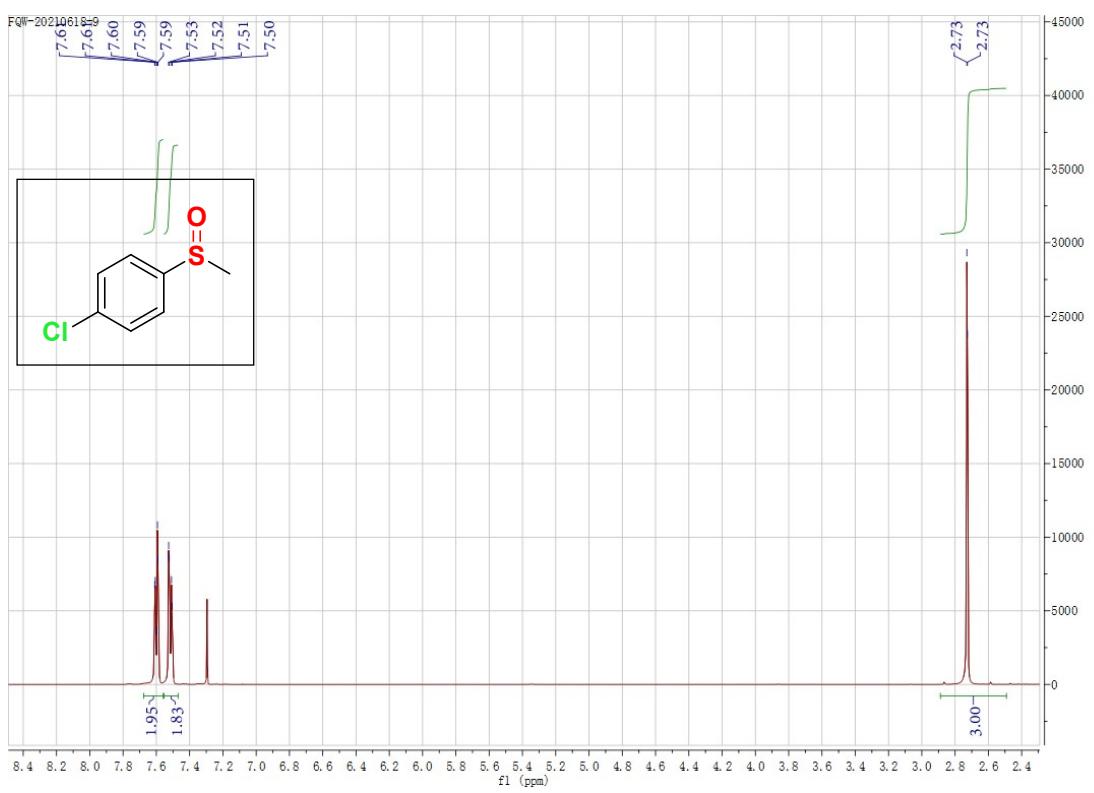


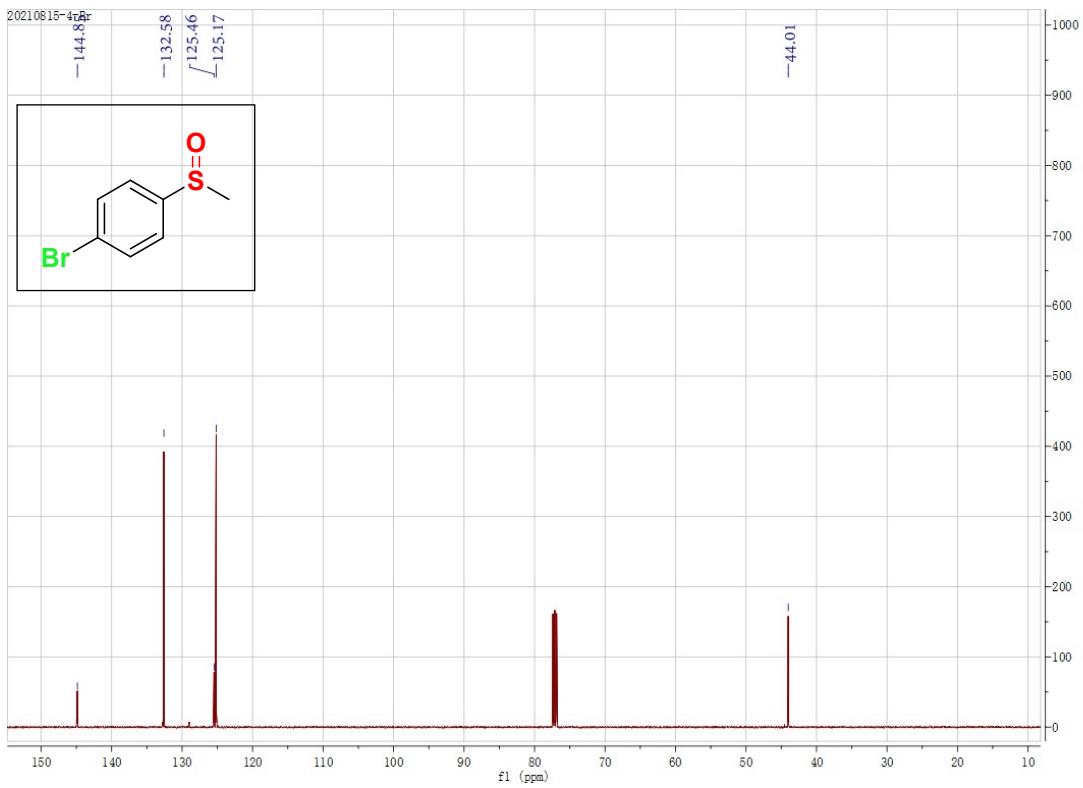
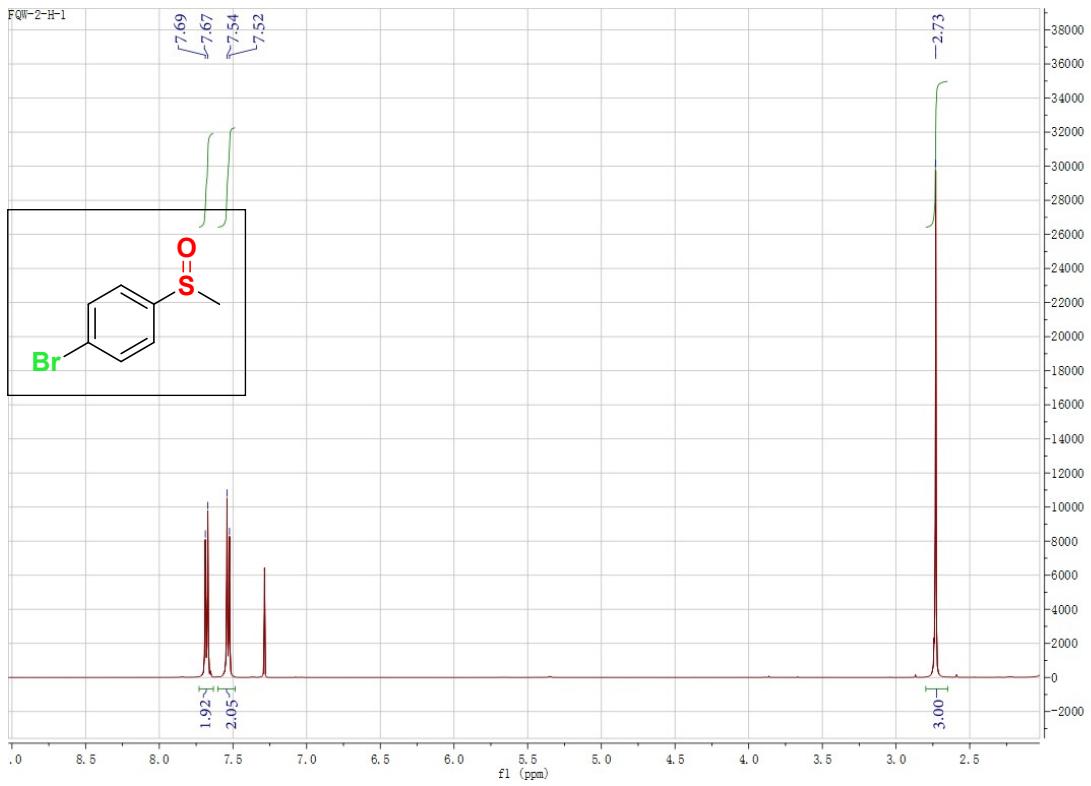


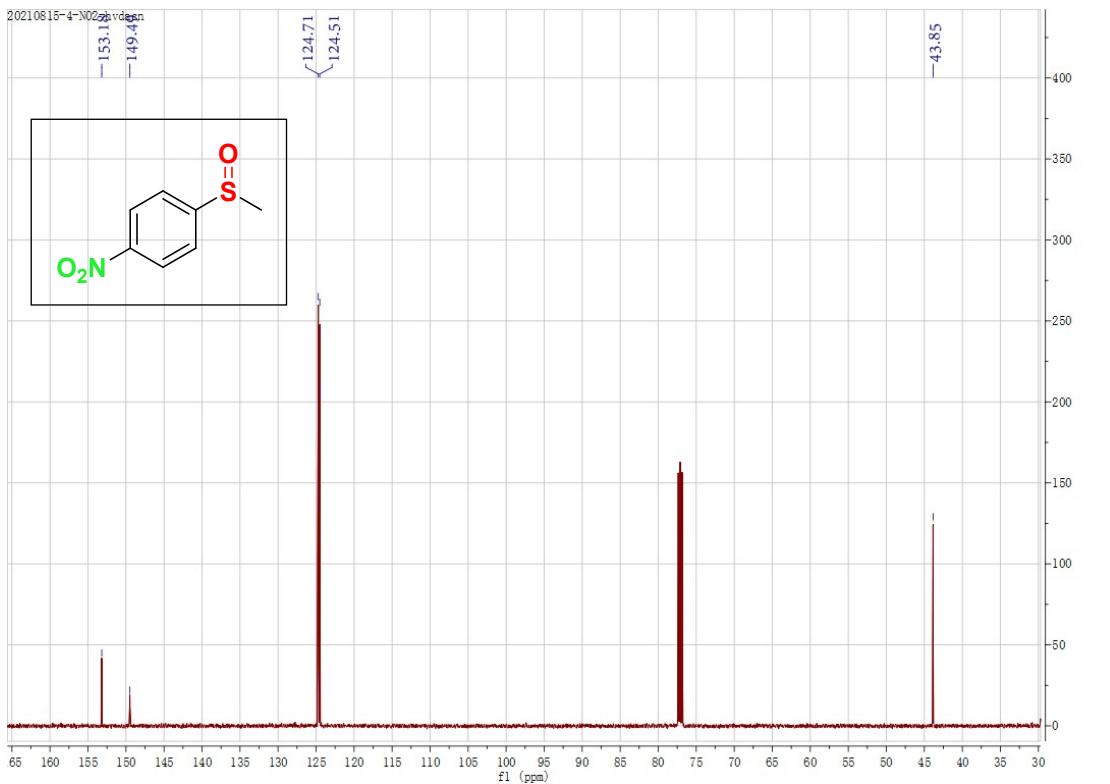
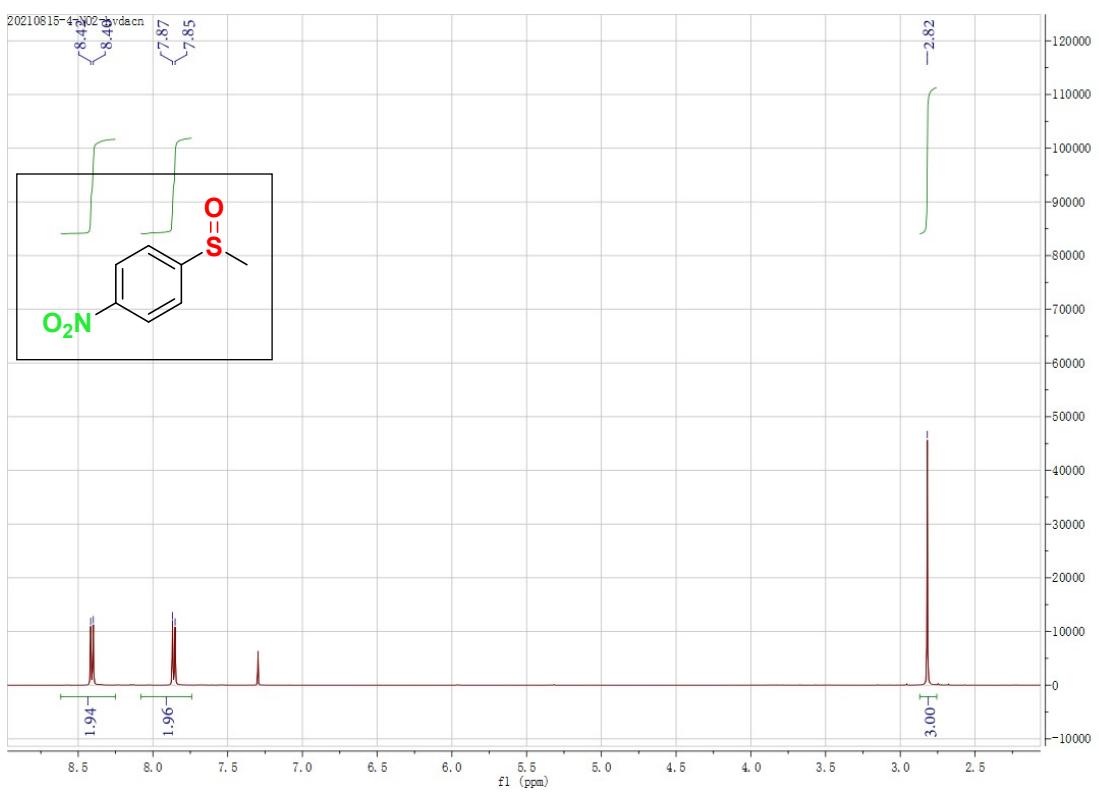


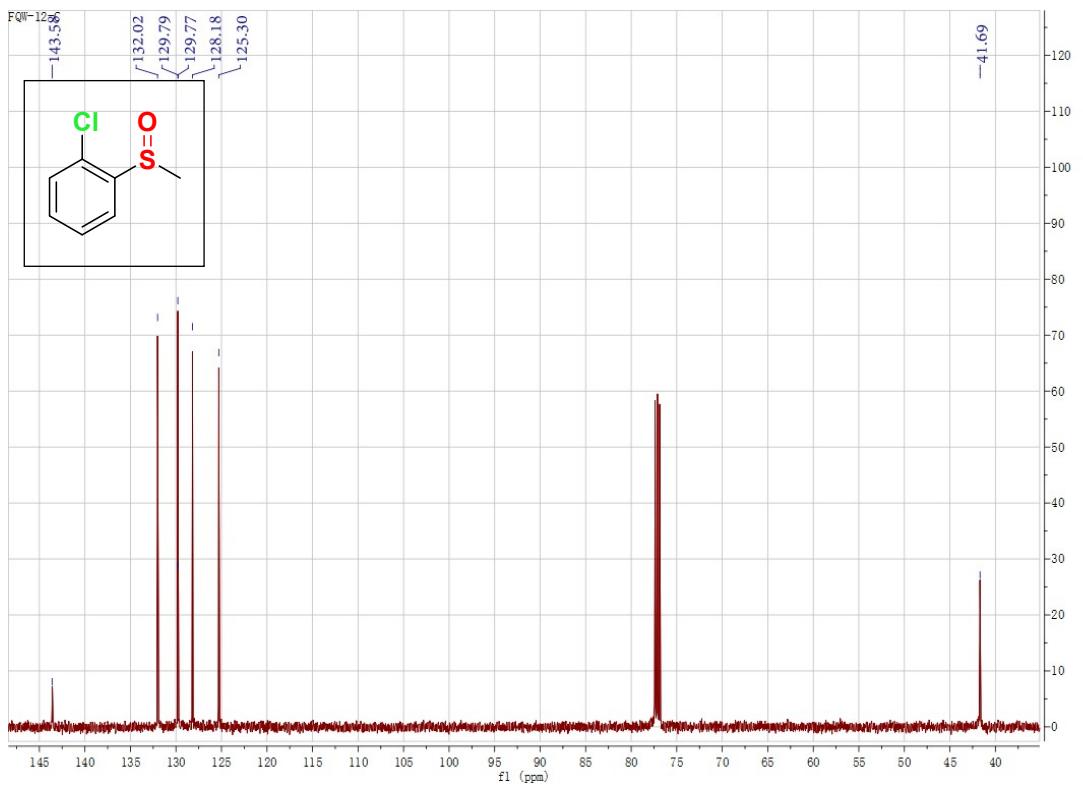
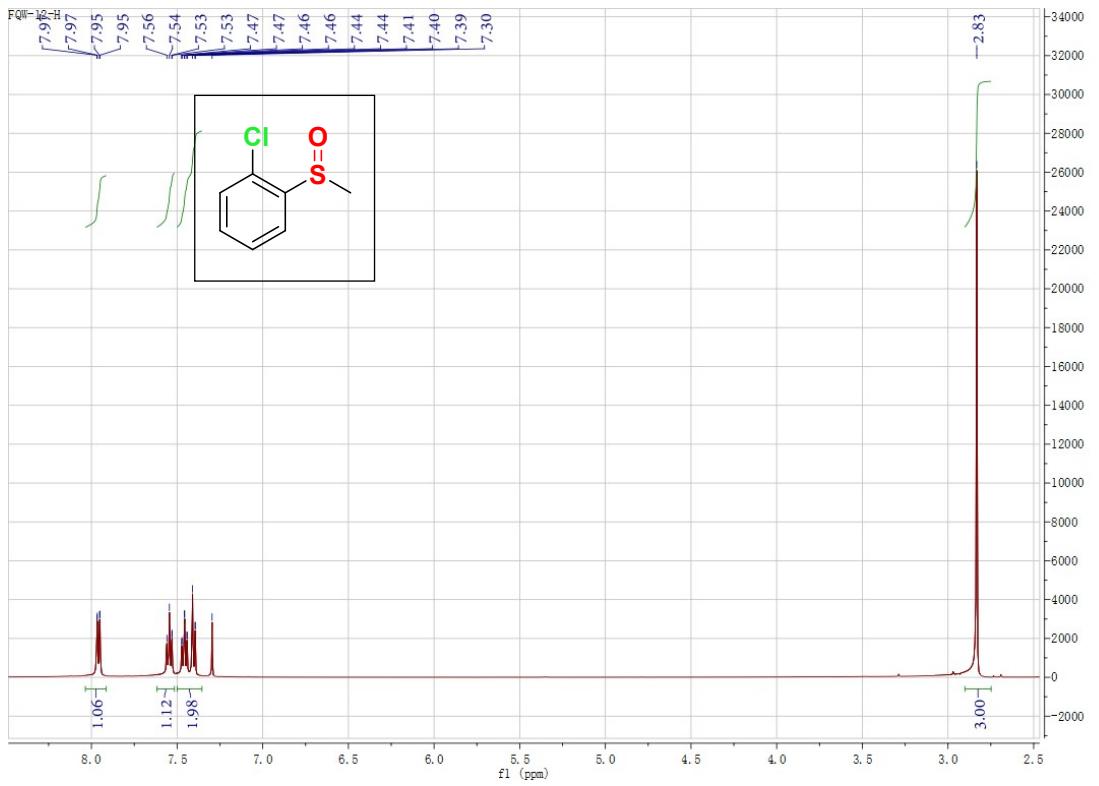


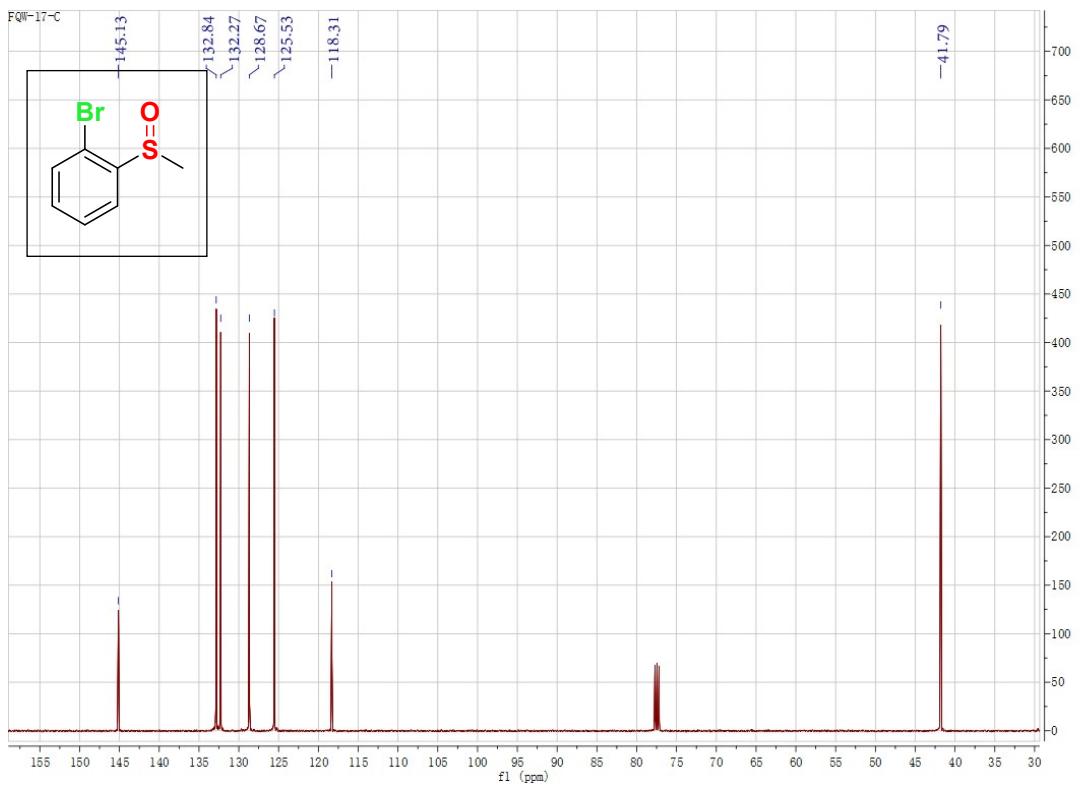
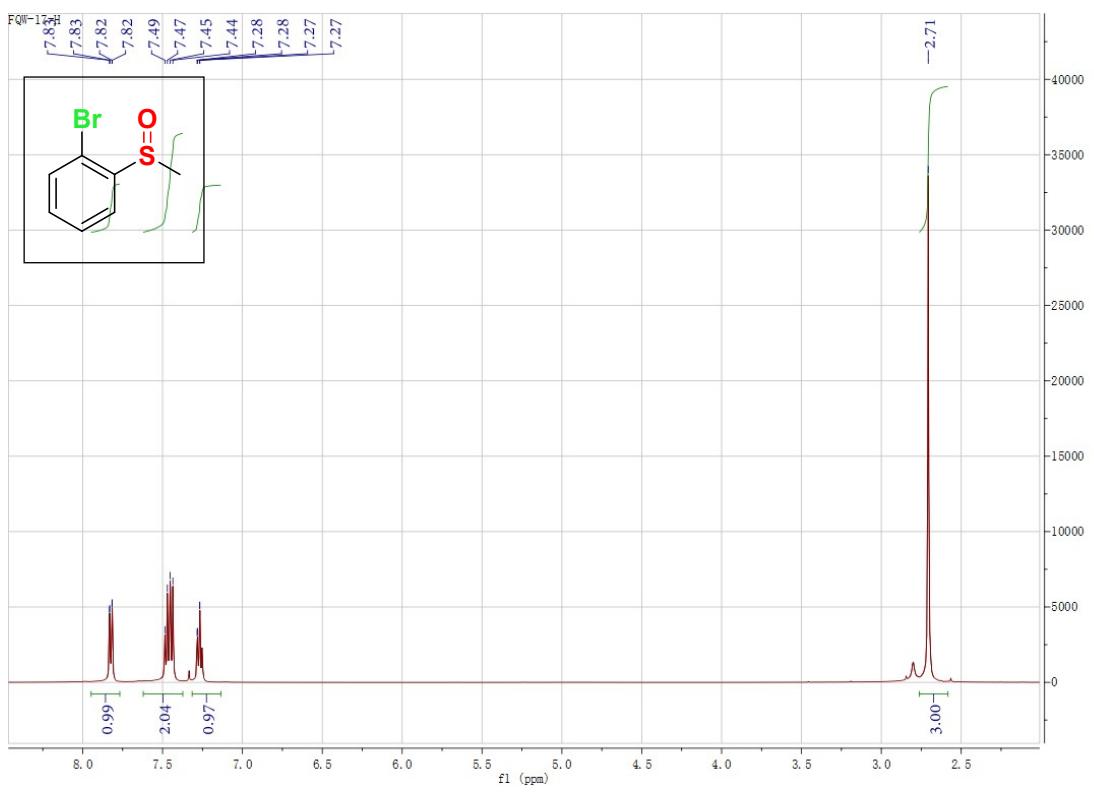


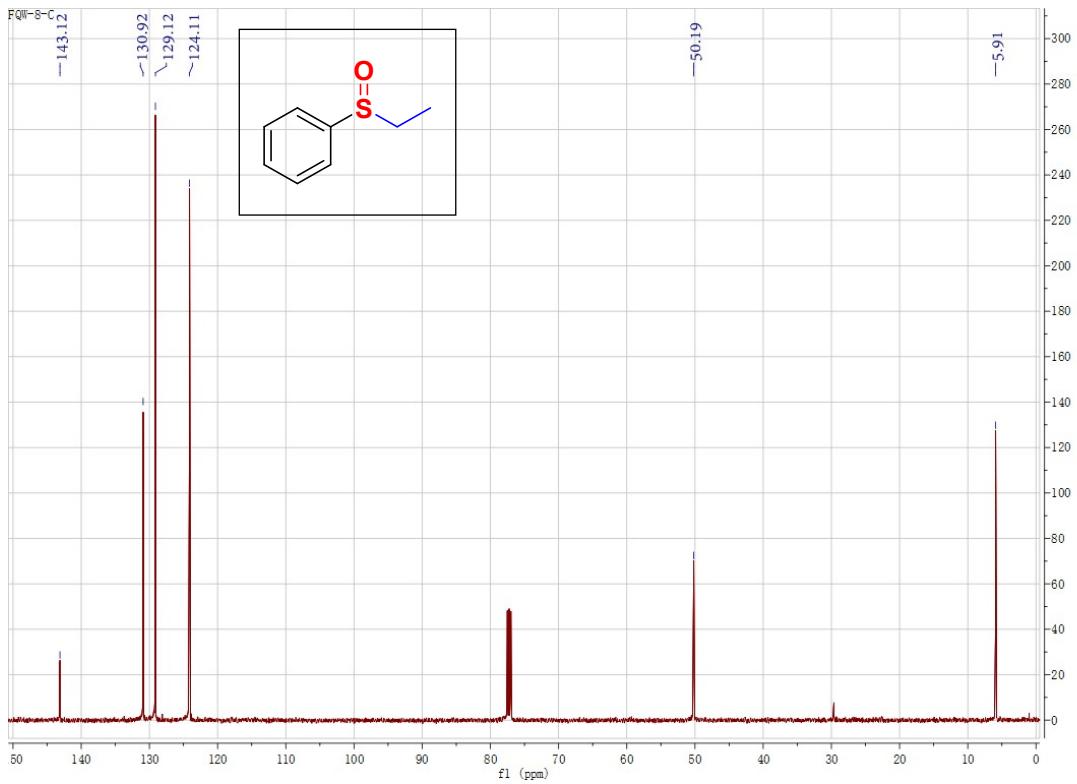
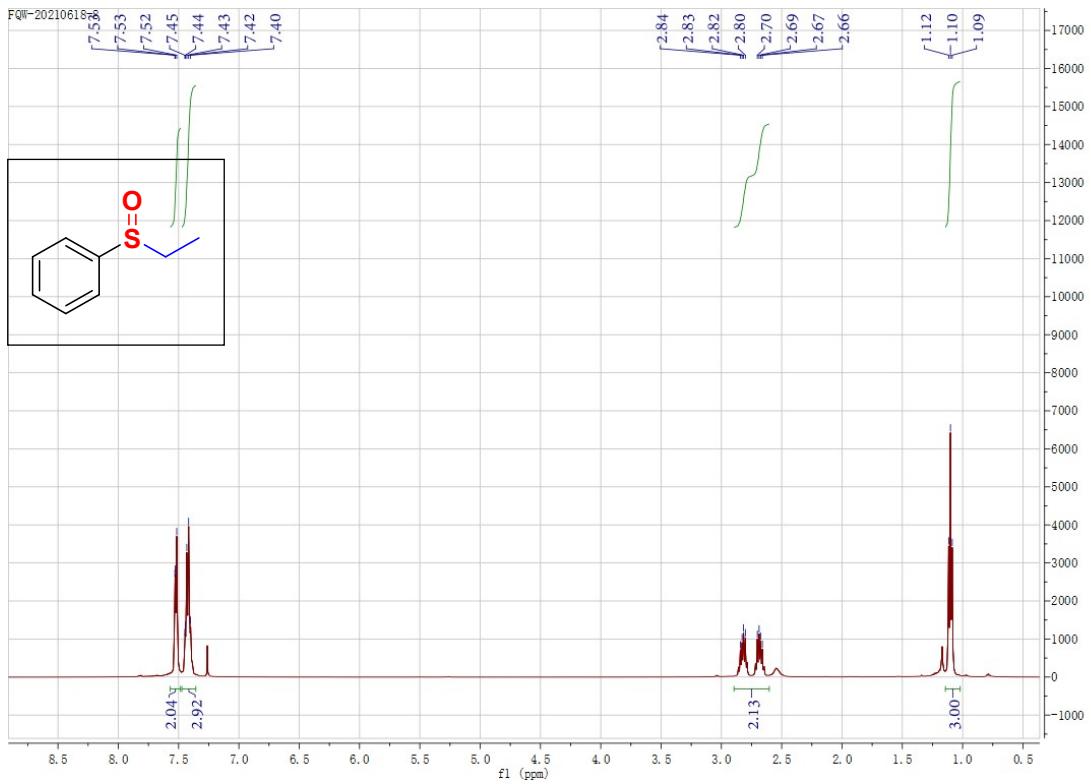


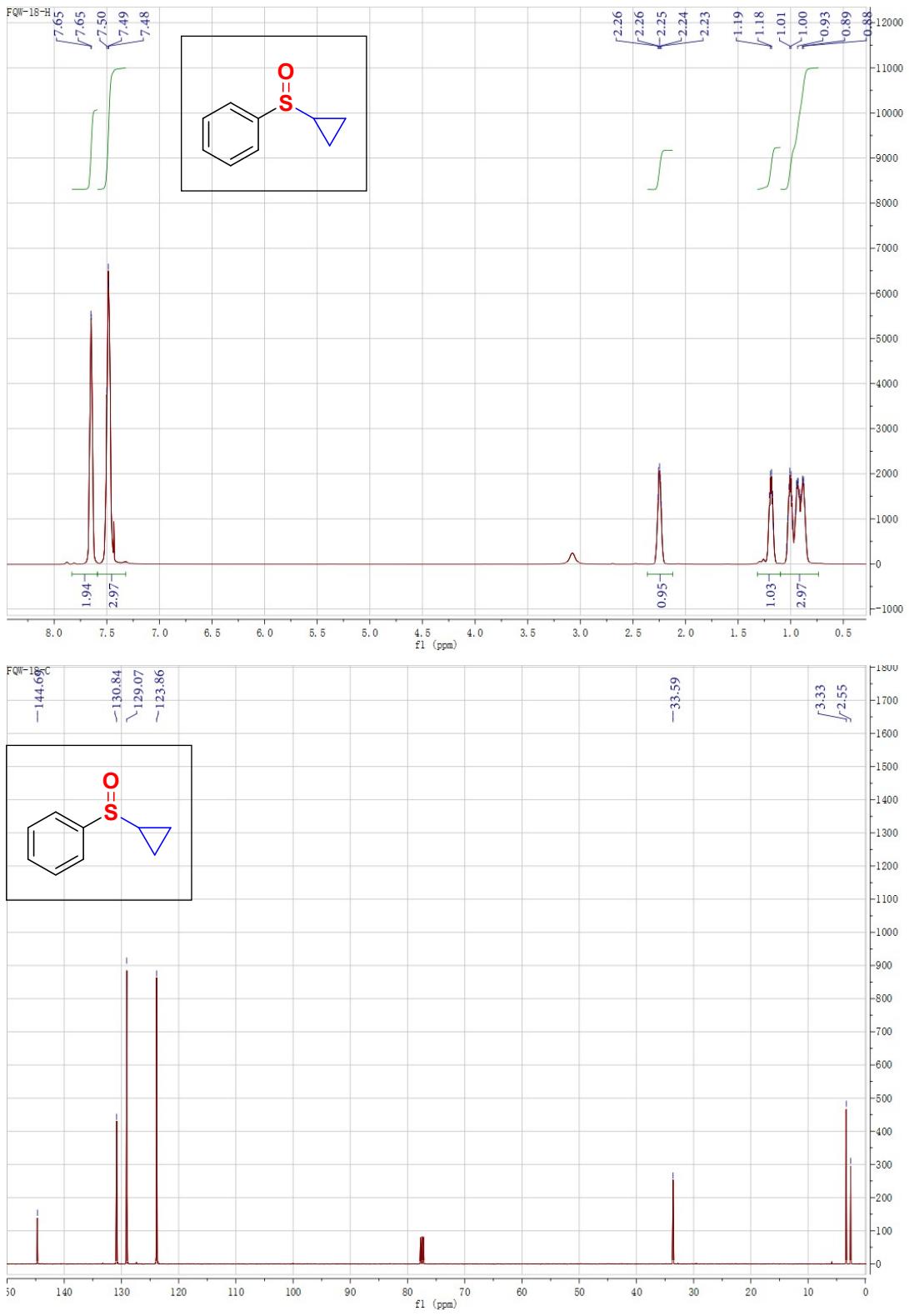


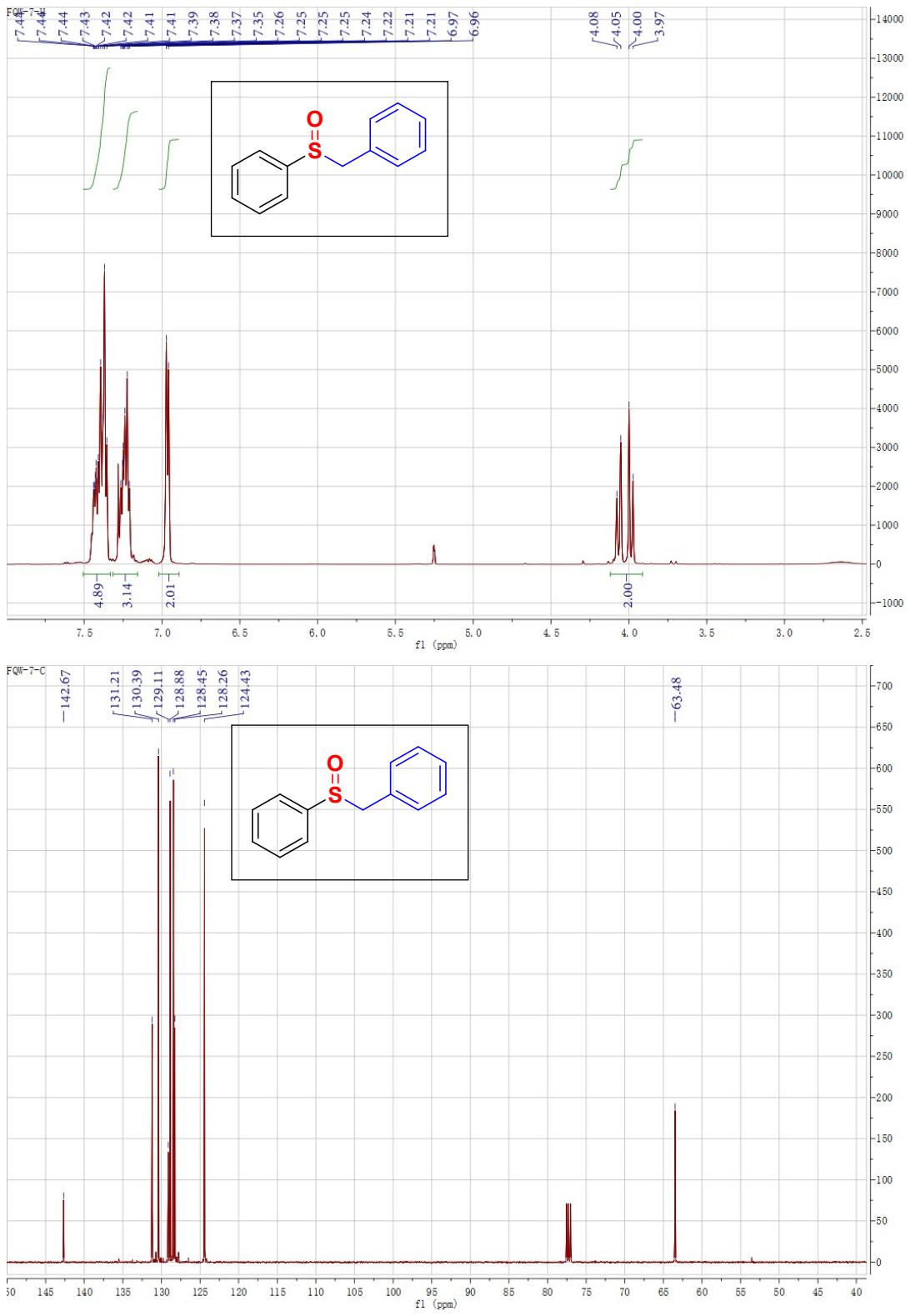


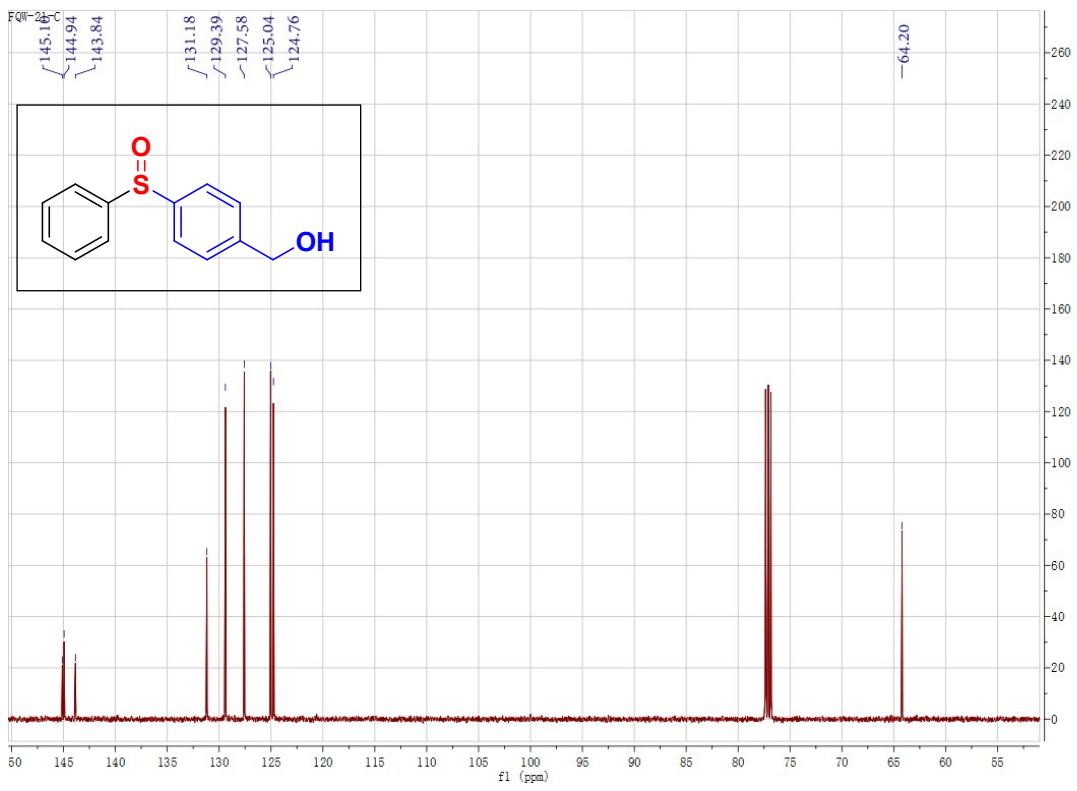
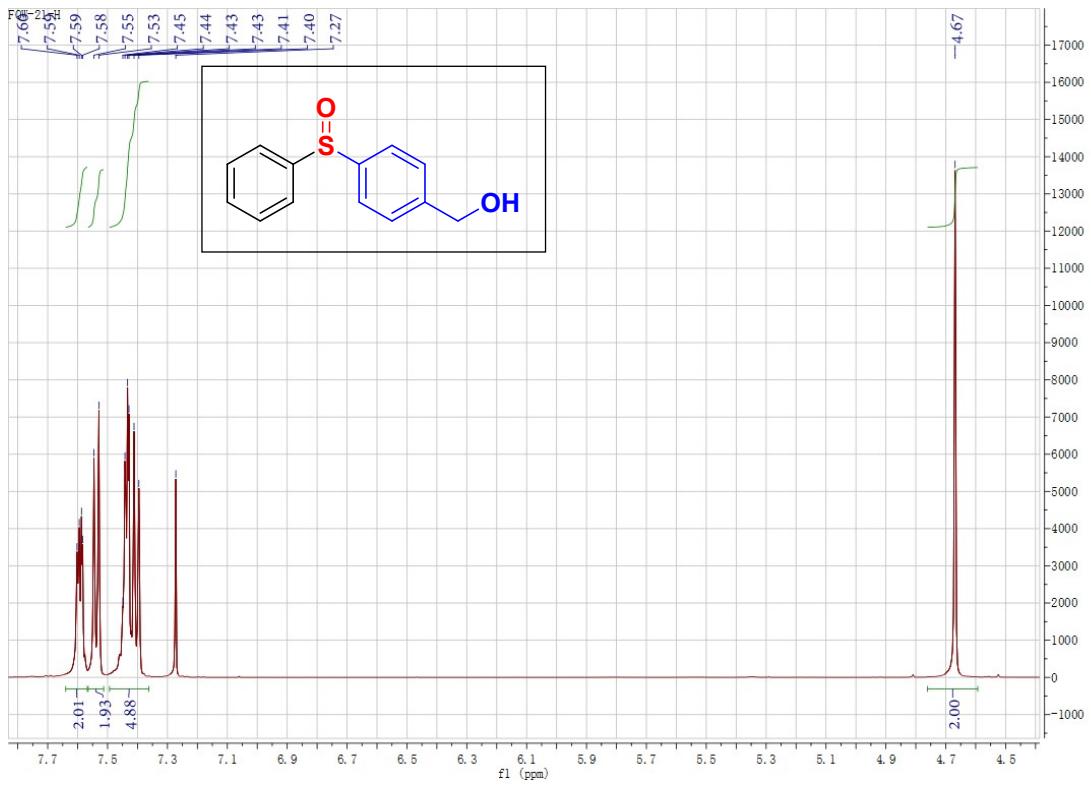


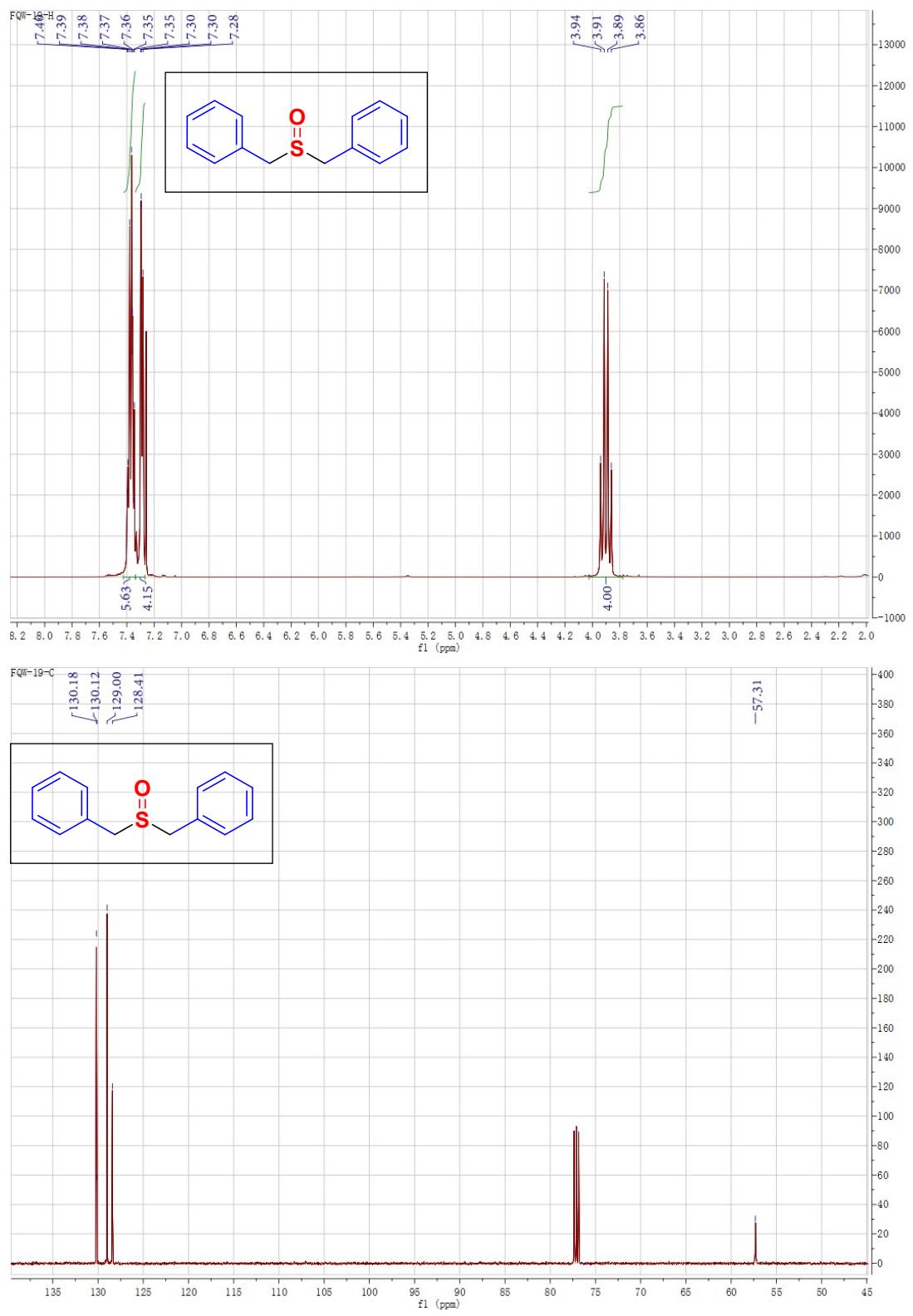


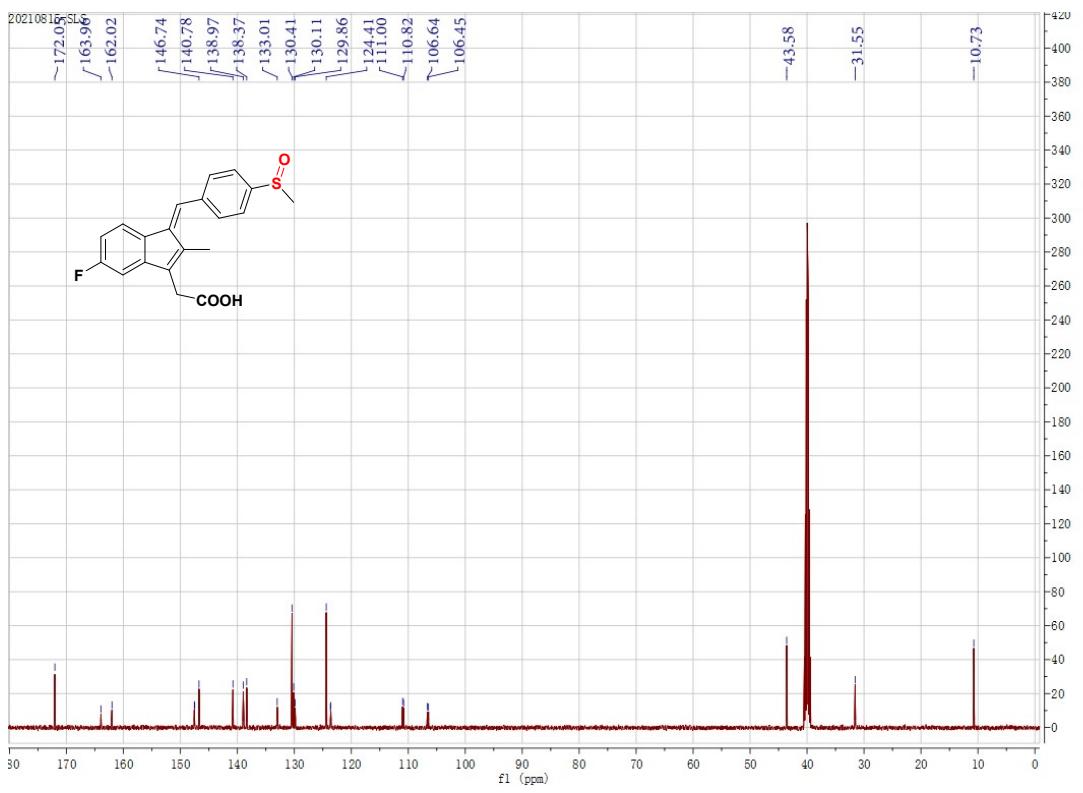
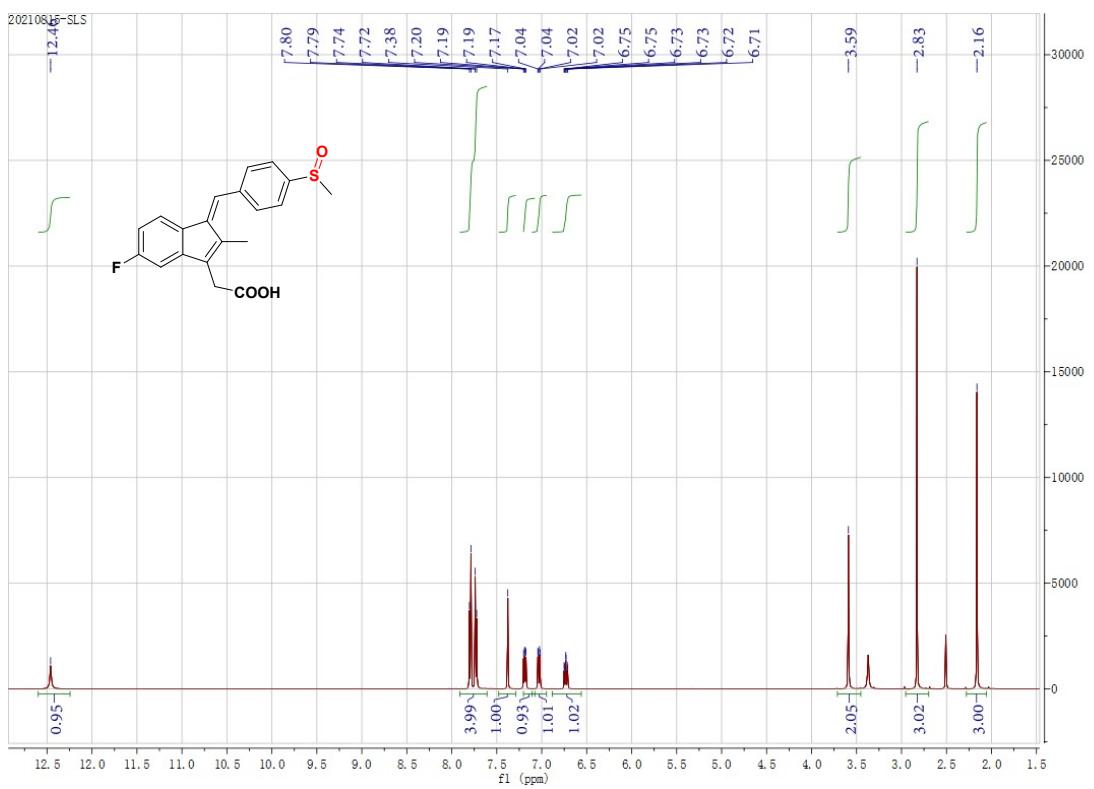


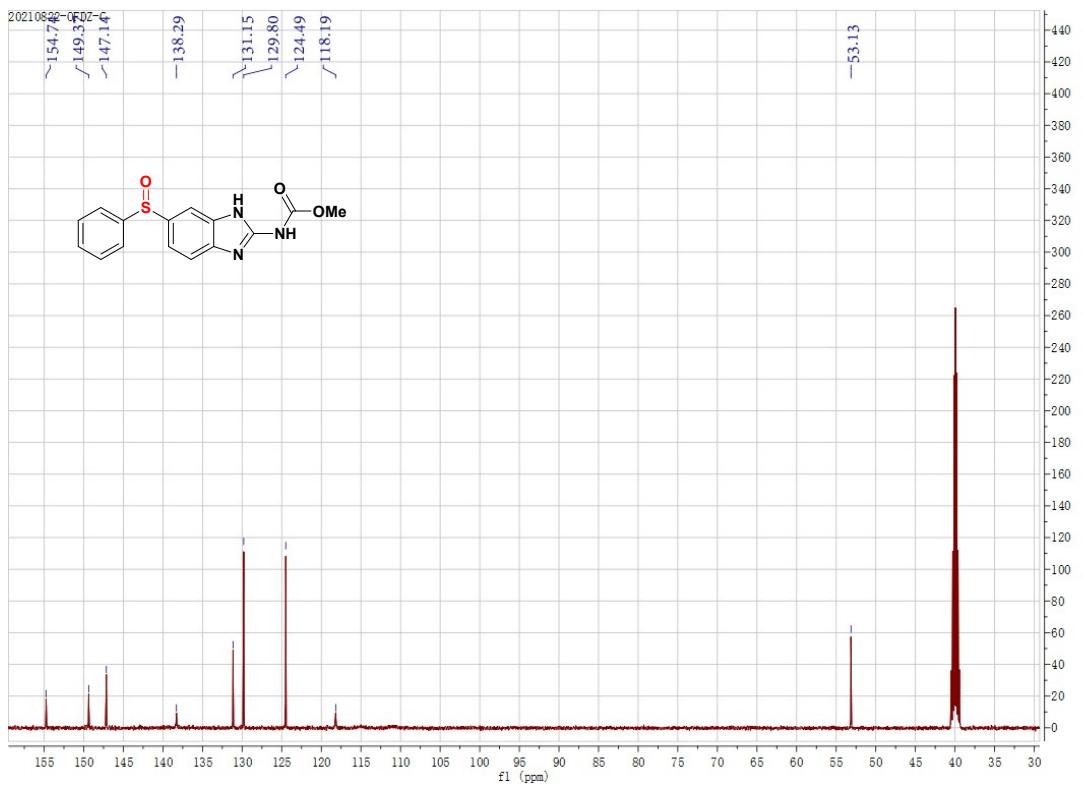
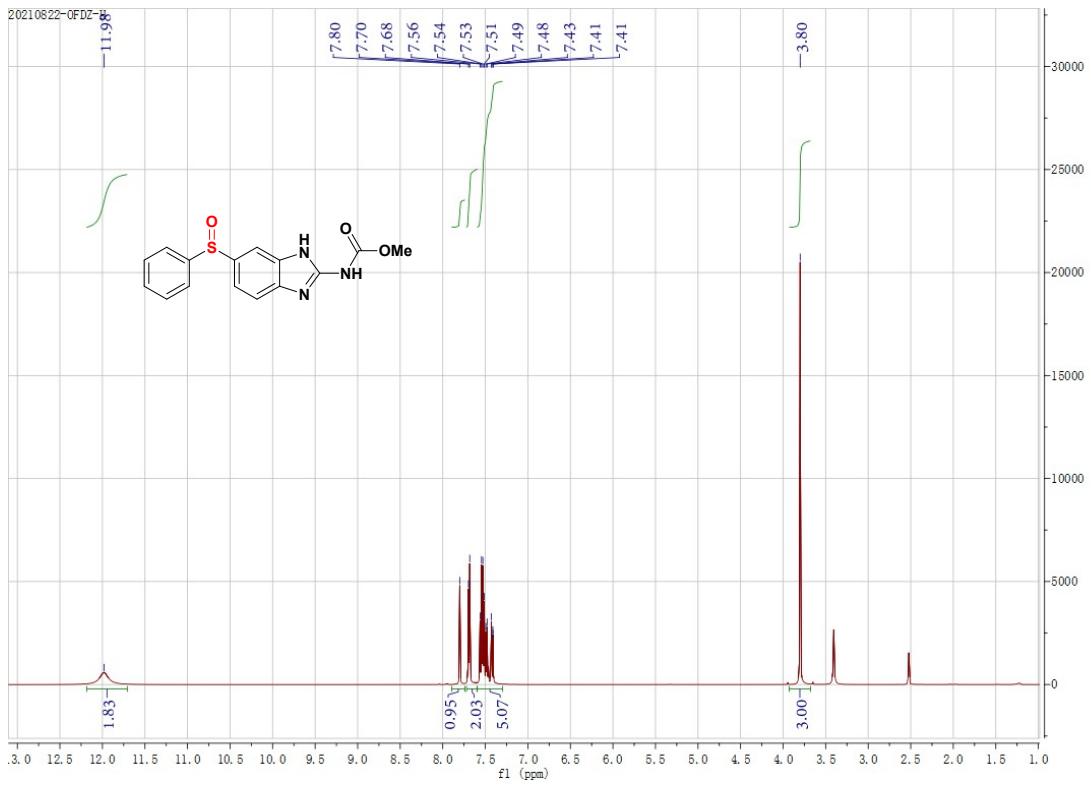












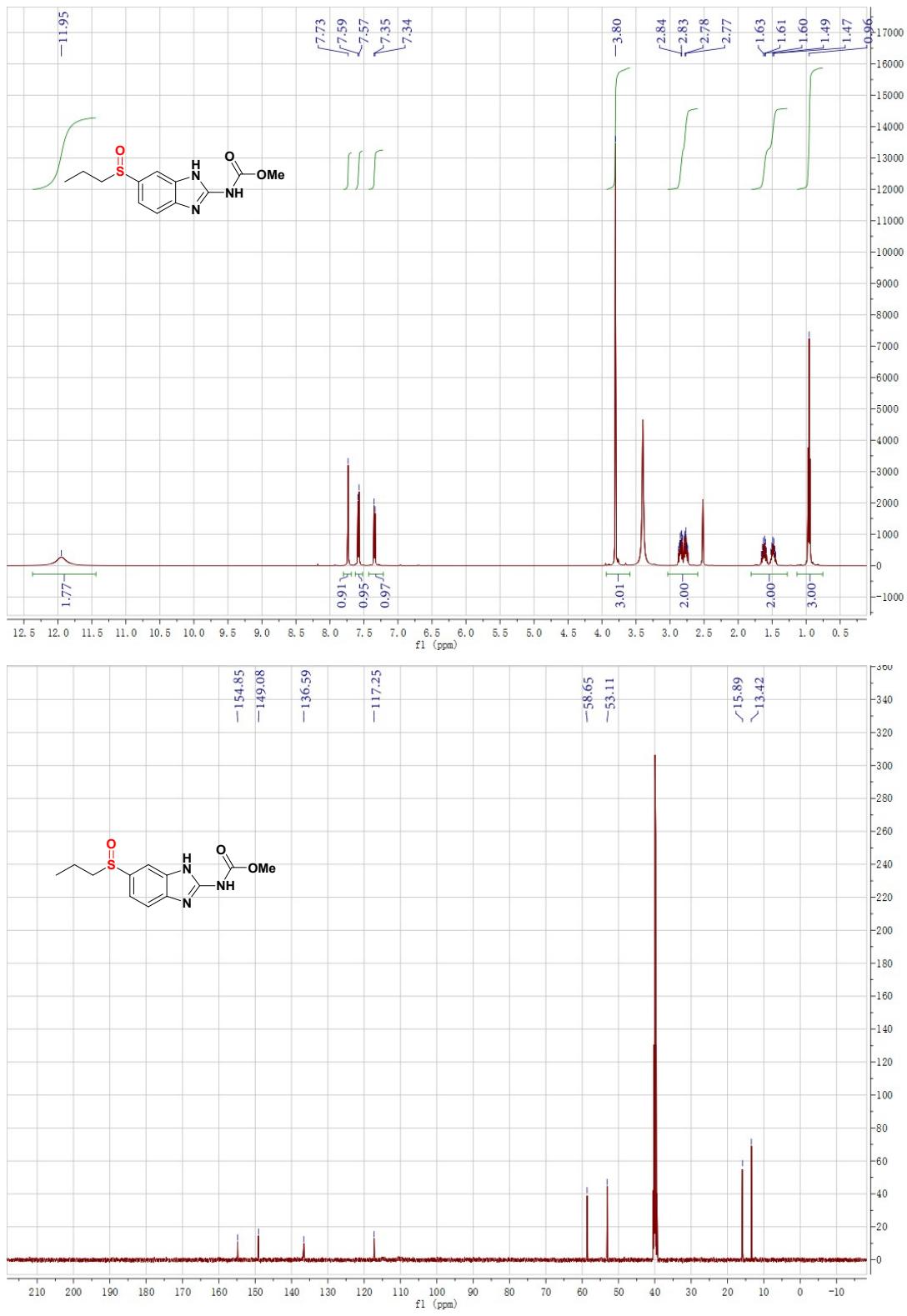


Fig.S3 ^1H and ^{13}C NMR spectra of sulfoxides

Cartesian coordination of stationary points

Cartesian coordination of 1a

C	-1.69988803	1.37808849	0.00006517
C	-0.33198345	1.09561001	0.00003500
C	0.10853065	-0.23319660	-0.00000516
C	-0.84006786	-1.26906326	-0.00001531
C	-2.20019740	-0.97623152	0.00001540
C	-2.63993717	0.35006426	0.00005609
H	-2.02496951	2.41364566	0.00009597
H	0.37260497	1.91739176	0.00004259
H	-0.51371495	-2.30479584	-0.00004827
H	-2.91934632	-1.78903314	0.00000677
H	-3.70079236	0.57603400	0.00008011
S	1.82641197	-0.72144940	-0.00004257
C	2.72923261	0.86198129	-0.00004446
H	2.51172980	1.44533775	0.89476634
H	3.78607220	0.59573645	-0.00007080
H	2.51168842	1.44535777	-0.89483199

Cartesian coordination of 1a*

C	1.71165000	1.41901100	-0.00005100
C	0.30214800	1.13430200	-0.00008300
C	-0.09179300	-0.20418000	-0.00002600
C	0.80173200	-1.30508200	0.00005700
C	2.20839700	-0.99662600	0.00007700
C	2.61656400	0.33532500	0.00002200
H	2.06516200	2.44161200	-0.00009800
H	-0.41478700	1.94400400	-0.00015700
H	0.44453700	-2.32721700	0.00011500
H	2.93927400	-1.79601100	0.00013900

H	3.68197000	0.55200000	0.00003600
S	-1.77911900	-0.70472300	-0.00009100
C	-2.76957700	0.81083300	0.00014500
H	-2.55555300	1.39891000	-0.89219200
H	-3.81394400	0.50208000	0.00014700
H	-2.55547400	1.39868000	0.89261400

Cartesian coordination of 1a^{3,*}

C	1.65684700	1.44361900	-0.00007700
C	0.32114300	1.16093300	-0.00005100
C	-0.14212700	-0.18556400	0.00000000
C	0.86698300	-1.31105400	0.00002100
C	2.20805800	-0.99860800	-0.00000600
C	2.65270900	0.33160100	-0.00005300
H	2.00563500	2.46944700	-0.00011700
H	-0.39452800	1.97587200	-0.00007000
H	0.52220500	-2.33845700	0.00005700
H	2.93891000	-1.80273000	0.00001000
H	3.71130600	0.56113000	-0.00007400
S	-1.78279200	-0.72345000	0.00004000
C	-2.77756800	0.80045200	0.00006100
H	-2.57809600	1.38978700	-0.89454700
H	-3.81888900	0.48214900	0.00002600
H	-2.57814100	1.38973200	0.89471500

Cartesian coordination of ³O₂

O	0.00000000	0.00000000	0.60267408
O	0.00000000	0.00000000	-0.60267408

Cartesian coordination of TS1

C	2.26097993	1.24915010	-0.44524782
C	0.88175049	1.08932339	-0.33690231
C	0.37030559	-0.13911101	0.08171011
C	1.21293195	-1.20908016	0.37683175
C	2.59539297	-1.03569796	0.27532935
C	3.11747221	0.18927389	-0.13544482
H	2.66741136	2.20034018	-0.77169756
H	0.21882278	1.91084470	-0.58476896
H	0.80433850	-2.16454645	0.68905494
H	3.25803162	-1.86093811	0.51263916
H	4.19103665	0.31985676	-0.21852019
S	-1.43190201	-0.44967614	0.26054833
C	-1.67738327	0.84948108	1.54193761
H	-1.47104543	1.83830705	1.13565708
H	-2.71123060	0.78558799	1.87798565
H	-0.98502804	0.62643405	2.35504741
O	-2.80213643	0.12355023	-0.65145571
O	-1.65168904	-0.42618821	-1.51272606

Cartesian coordination of IM1

C	2.15396592	-0.61955044	-1.20255844
C	0.78838601	-0.41568061	-1.03446880
C	0.34435936	0.17766970	0.14741134
C	1.22025267	0.55391512	1.16382984
C	2.58780635	0.35469982	0.97435234
C	3.05077012	-0.22941669	-0.20371934
H	2.51890482	-1.08460301	-2.11135641
H	0.08222438	-0.72987849	-1.79412877
H	0.84996461	0.99372202	2.08331799
H	3.28498508	0.64692893	1.75123769

H	4.11410349	-0.38963711	-0.34375097
S	-1.40662230	0.47714556	0.41424695
C	-1.76625948	1.69377649	-0.88269425
H	-1.46812025	1.28598410	-1.84662545
H	-2.83462867	1.89846605	-0.84837792
H	-1.19676743	2.59177796	-0.64691856
O	-2.14695906	-0.81243831	-0.27061280
O	-1.99309031	-1.93000790	0.69007969

Cartesian coordination of 2a

C	2.28289200	1.10711100	0.22823300
C	0.90431700	1.18618300	0.41882400
C	0.12593400	0.04984200	0.19942600
C	0.69452400	-1.16036000	-0.18208000
C	2.07633500	-1.22964600	-0.36632900
C	2.86808500	-0.09870100	-0.16503600
H	2.89907200	1.98454200	0.39232100
H	0.44961900	2.11980300	0.73551000
H	0.06116000	-2.02867100	-0.32562700
H	2.53212500	-2.16753300	-0.66544000
H	3.94188600	-0.15695000	-0.30716300
S	-1.67676100	0.13607300	0.46674700
C	-2.08871300	0.97783500	-1.10592100
H	-1.75235400	0.35288200	-1.93325700
H	-3.17086500	1.10392200	-1.12854200
H	-1.59554700	1.94999800	-1.12477200
O	-2.21464500	-1.29109400	0.34078900

Cartesian coordination of MECP

C	1.72529000	1.41942400	-0.00008100
C	0.28960300	1.12683800	0.00004300
C	-0.07520300	-0.20204000	0.00025600
C	0.78706600	-1.30821100	0.00065400
C	2.22302900	-0.99224800	0.00008200
C	2.61458100	0.33178000	-0.00040100
H	2.07804500	2.44277600	-0.00065300
H	-0.43297600	1.93262200	-0.00023600
H	0.42365900	-2.32820100	0.00003800
H	2.95408500	-1.79267500	-0.00007100
H	3.68300500	0.55010400	-0.00112300
S	-1.78507600	-0.70311700	-0.00013000
C	-2.77146300	0.81271600	0.00007800
H	-2.54000000	1.39914100	-0.89240200
H	-3.82202800	0.51772800	-0.00001100
H	-2.53999500	1.39882600	0.89275200

Relax scanning curve of the S and O atoms distance

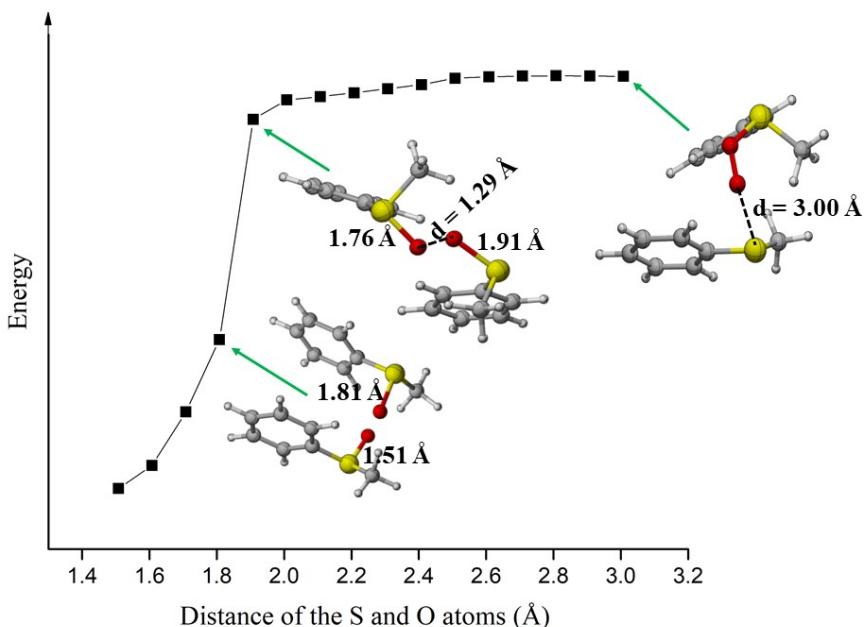


Fig S4 Relax scanning curve of the formation of S=O bond

References

1. C. Lee, W. Yang and R. G. Parr, *Phys. Rev. B*, 1988, **37**, 785-789.
2. A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 1372-1377.
3. S. Grimme, J. Antony, S. Ehrlich and H. Krieg, *J. Chem. Phys.*, 2010, **132**, 154104.
4. L. Goerigk and S. Grimme, *Phys. Chem. Chem. Phys.*, 2011, **13**, 6670-6688.
5. S. Grimme, *J. Comput. Chem.*, 2006, **27**, 1787-1799.
6. C. Gonzalez and H. B. Schlegel, *J. Chem. Phys.*, 1989, **90**, 2154-2161.
7. V. Barone and M. Cossi, *J. Chem. Phys. A*, 1998, **102**, 1995-2001.
8. B. Mennucci and J. Tomasi, *J. Chem. Phys.* 1997, **106**, 5151-5158.
9. J. J. Stewart, *J. Mol. Model.*, 2007, **13**, 1173-1213.
10. P. H. Krupenie, *J. Phys. Chem. Ref. Data*, 1972, **1**, 423-534.
11. M. L. McKee, *J. Am. Chem. Soc.*, 1998, **120**, 3963-3969.
12. D. Kashinski, G. Chase, R. Nelson, O. Di Nallo, A. Scales, D. VanderLey and E. Byrd, *J. Chem. Phys. A.*, 2017, **121**, 2265-2273.
13. M. Frisch, G. Trucks, H. Schlegel, G. Scuseria, M. Robb, J. Cheeseman, G. Scalmani, V. Barone, G. Petersson and H. Nakatsuji, *Journal*, 2016.
14. J. Harvey, M. Aschi, H Schwarz and W. Koch, *Theor. Chem. Acc.*, 1998, **99**, 95-99.
15. Tian Lu, sobMECP program, <http://sobereva.com/286> (accessed May 2, 2018).