

Supporting information for

Simultaneous single-cell phenotype analysis of hepatocellular carcinoma CTCs using SERS-aptamer based microfluidic chip

Rongke Gao,^{*, ab} Changbiao Zhan,^a Chunyu Wu,^a Yang Lu,^b Baoqiang Cao,^c Jing Huang,^d Feng Wang,^e Liandong Yu^{*, b}

^a State Key Laboratory of Advanced Display Technology, School of Instrument Science and Optoelectronic Engineering, Hefei University of Technology, Hefei 230009, China

^b College of Control Science and Engineering, China University of Petroleum (East China), Qingdao 266580, China

^c Department of Hepatobiliary Pancreatic Surgery, Anhui No.2 Provincial People's Hospital, Hefei 230041, China

^d Hefei University of Technology Hospital, Hefei 230009, China

^e School of Food and Biological Engineering, Hefei University of Technology, Hefei, 230009, China

*** Address for correspondence:**

Rongke Gao

Telephone: +86-13023091659, E-mail address: rkgao@upc.edu.cn

Liandong Yu

E-mail address: liandongyu@upc.edu.cn

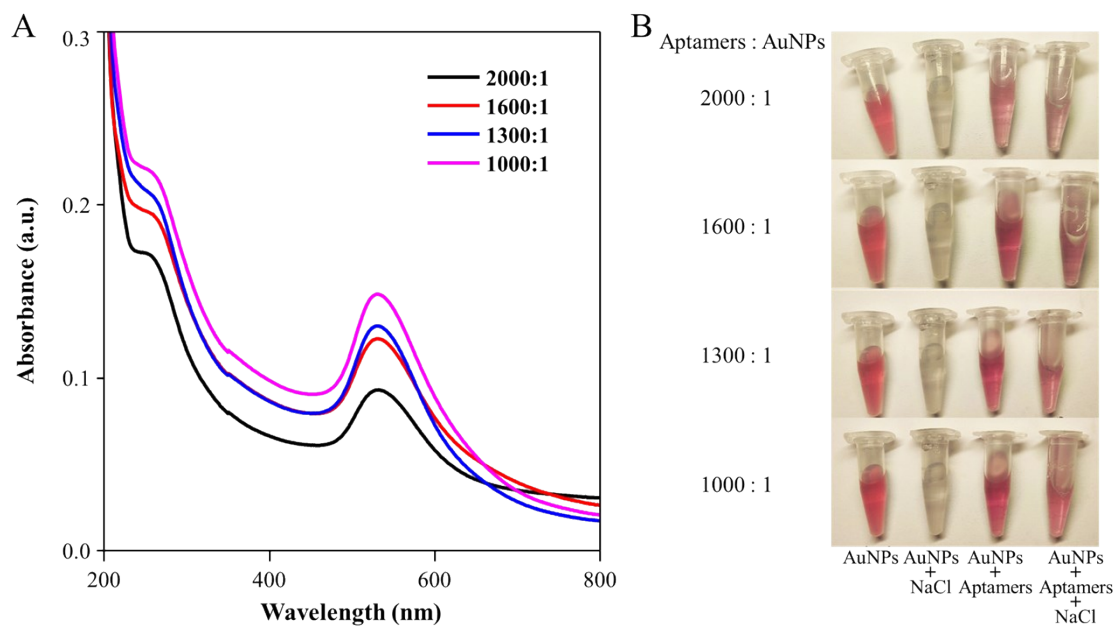


Figure S1. The UV-Vis spectra (A) and photographs (B) of various mixing ratios of aptamers and AuNPs.

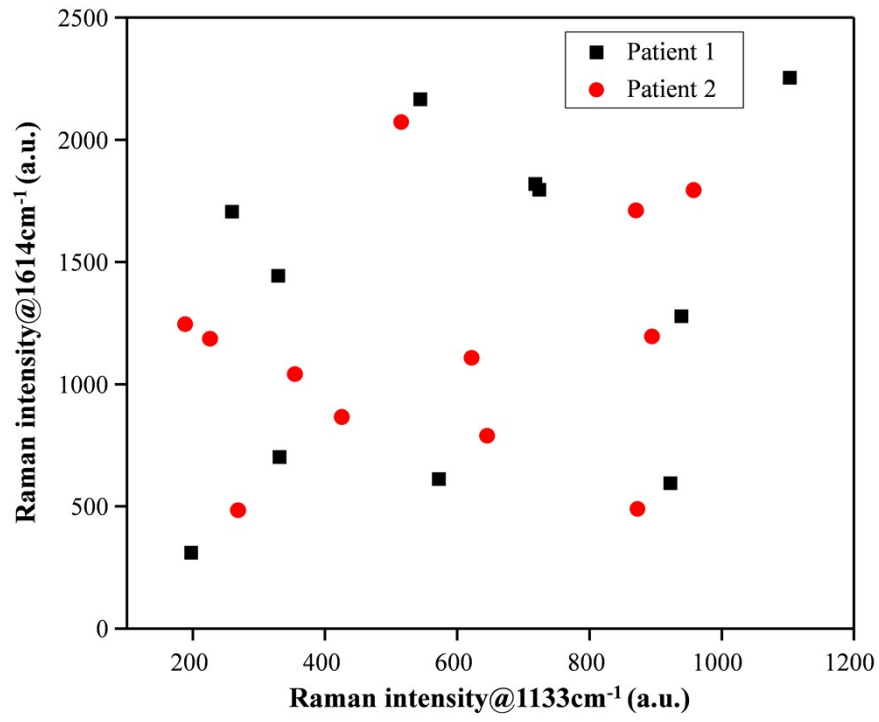


Figure S2. The single-cell phenotype analysis of two clinical specimens from HCC patients.

Table S1. Micro/nanofluidic technologies for the isolation of CTCs

Approach	Target cell line	Efficiency	Throughput	Ref.
Size filtration	MDA-MB-231, MCF7, SKBR3	$87 \pm 8\%$	$1 \mu\text{L min}^{-1}$	1
	A549, SK-MES-1, and H446 cells	90%	0.4 mL h^{-1}	2
	SK-HEP-1, HepG2	84%	$20 \mu\text{L min}^{-1}$	Our work
Aptamer-based isolation	CEM	$\sim 95\%$	600 nL s^{-1}	3
	SW480	86.7%-89.4%	1 mL h^{-1}	4
Antibody-based isolation	SK-HEP-1, HuH-7	85%, 89%	0.6 mL h^{-1}	5
Dean flow fractionation	MCF-7, T24, MDA-MB-231	$>80\%$	0.9 mL min^{-1}	6
	MCF-7	$91.5\% \pm 0.9\%$	2.5 mL min^{-1}	7
Immunomagnetic	MCF-7, SKBR3, MDA-MB-231, PC3	$\sim 90\%$	$>3 \text{ mL h}^{-1}$	8
Deterministic lateral displacement	SW480	$92.2 \pm 6.4\%$	1 mL h^{-1}	9

References

1. Y. Zhang, Z. Wang, L. Wu, S. Zong, B. Yun and Y. Cui, *Small*, 2018, **14**, e1704433.
2. T. Huang, C. P. Jia, Y. Jun, W. J. Sun, W. T. Wang, H. L. Zhang, H. Cong, F. X. Jing, H. J. Mao, Q. H. Jin, Z. Zhang, Y. J. Chen, G. Li, G. X. Mao and J. L. Zhao, *Biosens Bioelectron*, 2014, **51**, 213-218.
3. W. Sheng, T. Chen, R. Kamath, X. Xiong, W. Tan and Z. H. Fan, *Anal Chem*, 2012, **84**, 4199-4206.
4. Y. Song, Y. Shi, M. Huang, W. Wang, Y. Wang, J. Cheng, Z. Lei, Z. Zhu and C. Yang, *Angew Chem Int Ed Engl*, 2019, **58**, 2236-2240.
5. L. Zhu, H. Lin, S. Wan, X. Chen, L. Wu, Z. Zhu, Y. Song, B. Hu and C. Yang, *Anal Chem*, 2020, **92**, 15229-15235.
6. M. E. Warkiani, G. Guan, K. B. Luan, W. C. Lee, A. A. Bhagat, P. K. Chaudhuri, D. S. Tan, W. T. Lim, S. C. Lee, P. C. Chen, C. T. Lim and J. Han, *Lab Chip*, 2014, **14**, 128-137.
7. E. Lin, L. Rivera-Baez, S. Fouladdel, H. J. Yoon, S. Guthrie, J. Wiegner, Y. Deol, E. Keller, V. Sahai, D. M. Simeone, M. L. Burness, E. Azizi, M. S. Wicha and S. Nagrath, *Cell Syst*, 2017, **5**, 295-304 e294.
8. J. Autebert, B. Coudert, J. Champ, L. Saias, E. T. Guneri, R. Lebofsky, F. C. Bidard, J. Y. Pierga, F. Farace, S. Descroix, L. Malaquin and J. L. Viovy, *Lab Chip*, 2015, **15**, 2090-2101.
9. M. G. Ahmed, M. F. Abate, Y. Song, Z. Zhu, F. Yan, Y. Xu, X. Wang, Q. Li and C. Yang, *Angew Chem Int Ed Engl*, 2017, **56**, 10681-10685.