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Supporting information

Tuning compatibility and water uptake by protein charge modification in meltpolymerizable protein-based thermosets

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Figure S1. Representative raw mass spectra and deconvoluted molar masses for whey, wheyAc, and their methacrylated counterparts. Deconvoluted molar masses for three LC-MS replicates are shown.



Figure S2. Representative raw mass spectra and deconvoluted molar masses for wheyEt, wheySA, and wheySA5MA. Deconvoluted molar masses for three LC-MS replicates are shown.



Figure S3. Representative raw mass spectra for wheyEt5MA, wheyAcEt, and wheyAcEt5MA. Deconvolution was not performed due to the low signal to noise ratio

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			Whey		
		1:1	1:2	1:3	
Ecosurf	Nonionic, fatty acid based	у	у	у	
BG-10	Nonionic, alkyl polyglucoside	У	у	у	
Triton X-100	Nonionic, octiylphenol ethoxylate	У	у	у	
Glycerol	Nonionic polyol	У	у		

Table S1. Blending results for whey- non-ionic surfactant- n-BA. Miscible mixtures are noted with a 'Y'.

Table S2. Theoretical bovine β -lactoglobulin net charge at pH = 7, based on amino acid sequence and number of modification from LCMS results. Insoluble wheyEt5MA had 2 estimated methacrylations per bolvine β -lactoglobulin while wheyAcEt5MA was assumed to have same degree of acetylation and esterification as wheyAc and wheyEt, respectively, and 2 methacrylate groups.



Figure S4. ATR-FTIR performed on protein-surfactant-poly(n-butyl acrylate) blends prepared using non-methacrylated proteins. Differences in compatibility highlighted using the peak area ratio of the protein amide I (centered at 1647 cm⁻¹) to the poly(n-butyl acrylate) carbonyl (centered at 1737 cm⁻¹).



Figure S5. Representative FTIR plot showing the amide I baseline corrected peak (solid line), deconvoluted peaks (dotted line), and the fit (dash line). This measurement was taken on wheySA5MA protein powder (a) and copolymer (b).

Table S3. Deconvoluted ATR-FTIR peak positions (in cm⁻¹) for modified protein powder and protein-surfactant-poly(n-butyl acrylate) copolymer. Peak fitting was performed on amide I peaks, with peak positions, heights, and widths as fitting parameters. The two peaks at 1618-1628 cm⁻¹ and 1647-1657 cm⁻¹ were assigned to β -sheet, and α -helix and unordered structures respectively.¹ N=1 for protein powder, N=5 for copolymer.

Unmodified whey protein								
	Whey							
Peak 1 (cm ⁻¹)	1628							
Peak 2 (cm ⁻¹)	1653							
Methacrylated protein powder								
	Whey5MA	WheyAc5MA	WheyEt5MA	WheySA5MA	WheyAcEt5MA			
Peak 1 (cm ⁻¹)	1626	1624	1623	1625	1623			
Peak 2 (cm ⁻¹)	1650	1648	1647	1647	1647			
Copolymer								
	Whey5MA	WheyAc5MA	WheyEt5MA	WheySA5MA	WheyAcEt5MA			
Peak 1 (cm ⁻¹)	1624	1626	1619	1626	1626			
Peak 2 (cm ⁻¹)	1657	1655	1657	1655	1657			



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Figure S6. SEM-EDX of WheySA5MA with Na) sodium and Cl) chloride mapping, brighter is higher count rate.



Figure S7. SEM-EDX mapping of nitrogen (N) of protein copolymers at 3000x magnification.

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Figure S8. Representative SAXS and WAXS curves for whey5MA copolymer, with and without charge modification. WAXS curve offset to match the overlapping region with SAXS.



Figure S9. Relative-humidity water uptake of A) protein and B) protein:surfactant 1:1 complex.

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Figure S10. Main effect estimates of protein modification and complexation on water uptake of proteins. The 2⁴ factorial analysis was performed with methacrylation (5MA), esterification (Et), acetylation (Ac) and surfactant complexation (surf) as factors.

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Figure S11. Two-way interaction estimates of protein modification and complexation on water uptake of proteins. The 2⁴ factorial analysis was performed with methacrylation (5MA), esterification (Et), acetylation (Ac) and surfactant complexation (surf) as factors.



Figure S12. Three-way interaction estimates of protein modification and complexation on water uptake of proteins. The 2⁴ factorial analysis was performed with methacrylation (5MA), esterification (Et), acetylation (Ac) and surfactant complexation (surf) as factors.

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