Magnetoresponsive biocomposite hydrogels comprising of gelatin and valine based magnetic ionic liquid surfactant as controlled release nanocarrier for drug delivery

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1. Structural scheme of synthesized [ValC₁₆][FeCl₄].



Fig. S1. Synthetic procedure of $[ValC_{16}][FeCl_4]$.

2. ¹H NMR of the synthesis of [ValC₁₆][FeCl₄]



Fig. S2: ¹H NMR of [ValC₁₆][Cl]

¹H NMR chemical shift values of [ValC₁₆][Cl] CDCl₃, 500MH_z: δ_H (ppm) 8.43 (s, 3H), 4.16(m, 3H), 3.98(t,2H), 2.16(s,1H), 1.60 (m,2H), 1.25-1.29(m, 26H,CH₂), 0.96(dd, 6H), 0.85(t,3H).

3. LCMS of the synthesis of [ValC₁₆][FeCl₄]

LCMS of [ValC₁₆][FeCl₄]: ESI+ 342.3, ESI – 197.8



Fig. S3 LCMS of of [ValC₁₆][FeCl₄].

4. Raman spectra of [ValC₁₆][FeCl₄]



Fig. S4. Raman spectra of [ValC₁₆][FeCl₄]

5. UV spectra of [ValC₁₆][FeCl₄]¹



Fig. S5. UV spectra of [ValC₁₆][FeCl₄]

6. EPR spectra of [ValC₁₆][FeCl₄]



Fig. S6. EPR spectra of [ValC₁₆][FeCl₄]

7. DSC thermogram of [ValC₁₆][FeCl₄] & [ValC₁₆][Cl].



Fig. S7 DSC thermogram of [ValC₁₆][FeCl₄] & [ValC₁₆][Cl].

8. Thermogravimetric analysis and parameters



Fig. S8 TGA analysis of [ValC₁₆][FeCl₄] & [ValC₁₆][Cl].

9. Thermal parameters of [ValC₁₆][FeCl₄] & [ValC₁₆][Cl].

Table S1: T_d, T_{start} and T_{onset} of [ValC₁₆][FeCl₄] & [ValC₁₆]Cl.

| SAILs | T _{start} (°C) | T _d (°C) | T _{onset} (^o C) |
|---|-------------------------|---------------------|--------------------------------------|
| [ValC ₁₆][Cl] | 174 | 242 | 193 |
| [ValC ₁₆][FeCl ₄] | 180 | 251 | 207 |

10. Wavelength vs I_1/I_3 graph



Fig. S9 Wavelength vs I_1/I_3 on increasing concentration of $[ValC_{16}][FeCl_4]$.

11. Concentration dependent morphological transitions in [ValC₁₆][FeCl₄].



Fig. S10 Concentration dependent morphological transitions in [ValC₁₆][FeCl₄].

12. DLS plot of micelle formation using [ValC₁₆][FeCl₄]



Fig. S11 DLS plot of micelle using [ValC₁₆][FeCl₄]

13. Autocorrelation function of [ValC₁₆][FeCl₄].



Fig. S12. Autocorrelation function of [ValC₁₆][FeCl₄].

14. Zeta Potential of Micelles and vesicles of [ValC₁₆][FeCl₄]



Fig. S13 Zeta Potential of Micelles and vesicles of [ValC₁₆][FeCl₄]

15. EDX elemental mapping of Gelatin- [ValC₁₆][FeCl₄] biocomposite gel



Fig. S14 EDX elemental mapping of Gelatin-[ValC₁₆][FeCl₄] biocomposite gel
16. Stability of gelatin- [ValC₁₆][FeCl₄] biocomposite gel



Fig. S15 Stability of gelatin-[ValC₁₆][FeCl₄] biocomposite gel.

17. Swelling behavior of gelatin-[ValC₁₆][FeCl₄] biocomposite gel



Fig. S16 Swelling behavior of gelatin-[ValC $_{16}$][FeCl₄] biocomposite gel

18. Comparison of previously reported ornidazole drug loading efficiency in various systems.

Table S2 Comparison of previously reported ornidazole drug loading efficiency in various systems.

| S.No | Materials | Loading efficiency of Oridazole(%) | Author | Year |
|------|---|--|-----------------------------------|------|
| 1. | Present work (magnetic biocomposite gel) | 69.3% | - | - |
| 2. | Beta-cyclodextrin polymer microspheres (βCDPM) | 8.86% | Y. Liu et al. ² | 2020 |
| 3. | PVP electrospun fibers | - | S. Tort et al. ³ | 2018 |
| 4. | Polyethylene glycol-based micron- level particle | 15-20% | S. Tamilvanan et al. ⁴ | 2019 |
| 5. | Ornidazole-Loaded Graphene Paper | 10-17.3% | W. Quian et al. ⁵ | 2018 |
| 6. | Biopolymer-dextrin and poly(methyl methacrylate) microgel | 30.54 % | D. Das et al. ⁶ | 2016 |
| 7. | Biocompatible hydrogel derived from glycogen and poly(N- isopropylacrylamide) | 19-31 % | Priyapratim Patra et al.7 | 2016 |

19. Comparison of previously reported 5-Fluoro uracil drug loading efficiency in various systems.

Table S3 Comparison of previously reported 5-Fluoro uracil drug loading efficiency in various systems.

| S.No | Materials | Loading efficiency of 5-FU (%) | Author | Year | |
|------|--|---|---------------------------------|------|--|
| 1. | Present work (magnetic biocomposite gel) | 78.3% | - | - | |
| 2. | Poly(ϵ -caprolactone) with 6-(chloromethyl)uracil | 42% | S. Zhu et al. ⁸ | 2020 | |
| 3. | Gelatin–rosin gum complex nanoparticle | 50% | S. Joshi ⁹ | 2020 | |
| 4. | polyurethane-based hydrogels | 45-54 % | M. Kamaci et al. ¹⁰ | 2020 | |
| 5. | PEGylated Ag2S QDs functionalized with Cetuximab (Cet) antibody | 7.34% | F. D. Dumanet al. ¹¹ | 2019 | |
| 6. | zinc nanoMOFs functionalized with folate | 24% | B.Yang et al. ¹² | 2017 | |
| 7. | Self-Assembling Monomeric Nucleoside Molecular Nanoparticle | 53% | H. Zhao et al. ¹³ | 2015 | |

20. Kinetics study of drug release pattern using mathematical models

| Guest Molecule | Release condition | *Zero Order | | *First order | | *Higuchi Model | | *KorsMeyar Peppas | | *Hixon Crowell | |
|-------------------|-------------------|----------------|-------|-----------------|-------|-------------------|-------|----------------------|-------|-------------------|-------|
| | | R ² | Slope | R ² | Slope | R ² | Slope | R ² | Slope | R ² | Slope |
| Ornidazole | pH 7.4 | 0.83 | 0.75 | 0.89 | 0.006 | 0.93 | 8.3 | 0.98 | 0.62 | 0.87 | 0.018 |
| 5-Fluoro Uracil | pH 7.4 | 0.72 | 0.42 | 0.85 | 0.005 | 0.86 | 6.88 | 0.97 | 0.58 | 0.85 | 0.012 |

Table S4: Kinetics study of release of guest drugs by various mathematical models.

21. Drug release pattern in different electrolyte solutions



Fig. S17 Drug release pattern in different electrolyte solution

Annexure I

The Surface parameter equations are as follow:

1. The Adsorption efficiency(pC_{20}) and Effectiveness of Surface tension reduction(Π_{CAC}) of surfactant at air-water interface is estimated by using the relation (1)

$$pC_{20} = -\log C_{20}, \qquad \pi_{CMC} = \gamma_{H,0} - \gamma_{CAC}$$
.....(1)

where, C_{20} is the concentration reduce by 20mNm⁻¹ from the surface tension of the solvent (water)¹, γ_{H_2O} stands for the surface tension of the pure water and γ_{CMC} stands for the surface tension of the solvent medium at CAC.

2. The amount of surfactants adsorbed at the interface is estimated from relative surface excess concentration (Γ_{max}). The values Γ_{max} of at the CMC have been calculated using Gibbs adsorption Eq. 2.

$$\Gamma_{\max} = -\frac{1}{nRT} \frac{\partial \gamma}{\partial \ln C}....(2)$$

where " $\partial \gamma / \partial \ln C$ " is the slope of $\gamma - \ln C$ plot in the pre *CMC* region and n is Gibbs adsorption coefficient.

3. Minimum area occupied by monomers at the interface was calculated using equation 4.

$$A_{\min} = \frac{10^{10}}{\Gamma_{\max}.N_A}....(3)$$

where N_A is Avogadro number and the Unit of A_{min} is Å².

4. The β value is obtained from the formula ($\beta = 1-\alpha$) where the α is the degree of counterion dissociation which is obtained by ratio of the slope post micellar region and the pre micellar region (S_2/S_1) then the β value is further used to derive the value of standard free energy of micellization from the equation as follow:

$$\Delta G_{mic}^{o} = (1+\beta)RT\ln X_{cmc} \quad(4)$$

Annexure-II

Various mathematical models and their equations

1. Zero order mathematical model:

$$C_o - C_t = K_o t$$

where $C_o =$ initial concentration of the drug at time, t = 0, $C_t =$ amount of drug released at time t, $K_o =$ zero order constant

2. First Order mathematical model:

 $\ln C = \ln C_0 - K_1 t$

where C_o = initial concentration of the drug at time K_1 = first order rate constant, C = percent of drug remaining at time

3. Higuchi model:

$$Q = A_{\sqrt{D(2Co - Cs)Cst}}$$

Q=Cumulative amount of drug released at time per unit area, C_S is the drug solubility in the matrix and D is the diffusion coefficient of the drug molecule in the matrix, C_S =drug solubility in the matrix.

4. Korsmeyer- Peppas model

$$M_t/M_\infty = K_{kp} t^n$$

Mt = amount of drug released in time t, $M\infty$ = amount of drug released after time ∞ , n = diffusional exponent or drug release exponent, and K_{kp} = Korsmeyer release rate constant.

5. Hixson–Crowell model

$$C_{0}^{1/3} - C_{t}^{1/3} = K_{HC}t$$

 K_{HC} = Hixson–Crowell constant

References

1. Vander Hoogerstraete, T.; Wellens, S.; Verachtert, K.; Binnemans, K., Removal of transition metals from rare earths by solvent extraction with an undiluted phosphonium ionic liquid: separations relevant to rare-earth magnet recycling. *Green Chemistry* **2013**, *15* (4), 919-927.

2. Liu, Y.; Tang, P.; Pu, H.; Qian, H.; Sun, Q.; Zhao, L.; Li, M.; Li, H., Study on the synthesis and drug-loading optimization of beta-cyclodextrin polymer microspheres containing ornidazole. *Journal of Drug Delivery Science and Technology* **2020**, *58*, 101836.

3. Tort, S.; Yıldız, A.; Tuğcu-Demiröz, F.; Akca, G.; Kuzukıran, Ö.; Acartürk, F., Development and characterization of rapid dissolving ornidazole loaded PVP electrospun fibers. *Pharmaceutical Development and Technology* **2019**, *24* (7), 864-873.

4. Tamilvanan, S.; Chanda, P., Ornidazole-loaded polyethylene glycol-based micron-level particles: influence of eutectic liquid on reservoir-type particle formation, drug entrapment efficiency and drug dissolution or release behavior. *Polymer Bulletin* **2019**, *76* (9), 4389-4398.

5. Qian, W.; Wang, Z.; He, D.; Huang, X.; Su, J., Ornidazole-loaded graphene paper for combined antibacterial materials. *Journal of Saudi Chemical Society* **2018**, *22* (5), 581-587.

6. Das, D.; Rameshbabu, A. P.; Patra, P.; Ghosh, P.; Dhara, S.; Pal, S., Biocompatible amphiphilic microgel derived from dextrin and poly(methyl methacrylate) for dual drugs carrier. *Polymer* **2016**, *107*, 282-291.

7. Patra, P.; Rameshbabu, A. P.; Das, D.; Dhara, S.; Panda, A. B.; Pal, S., Stimuli-responsive, biocompatible hydrogel derived from glycogen and poly(N-isopropylacrylamide) for colon targeted delivery of ornidazole and 5-amino salicylic acid. *Polymer Chemistry* **2016**, *7* (34), 5426-5435.

8. Zhu, S.; Wen, L.; Xiao, Y.; Lang, M., Poly(ε-caprolactone) with pH and UCST responsiveness as a 5-fluorouracil carrier. *Polymer Chemistry* **2020**, *11* (32), 5173-5180.

9. Joshi, S.; Singh, V., Gelatin–rosin gum complex nanoparticles: preparation, characterization and colon targeted delivery of 5-fluorouracil. *Chemical Papers* **2020**, *74* (12), 4241-4252.

10. Kamaci, M., Polyurethane-based hydrogels for controlled drug delivery applications. *European Polymer Journal* **2020**, *123*, 109444.

11. Duman, F. D.; Akkoc, Y.; Demirci, G.; Bavili, N.; Kiraz, A.; Gozuacik, D.; Acar, H. Y., Bypassing pro-survival and resistance mechanisms of autophagy in EGFR-positive lung cancer cells by targeted delivery of 5FU using theranostic Ag2S quantum dots. *Journal of Materials Chemistry B* **2019**, *7* (46), 7363-7376.

12. Yang, B.; Shen, M.; Liu, J.; Ren, F., Post-Synthetic Modification Nanoscale Metal-Organic Frameworks for Targeted Drug Delivery in Cancer Cells. *Pharmaceutical Research* **2017**, *34* (11), 2440-2450.

13. Zhao, H.; Feng, H.; Liu, D.; Liu, J.; Ji, N.; Chen, F.; Luo, X.; Zhou, Y.; Dan, H.; Zeng, X.; Li, J.; Sun, C.; Meng, J.; Ju, X.; Zhou, M.; Yang, H.; Li, L.; Liang, X.; Chu, L.; Jiang, L.; He, Y.; Chen, Q., Self-Assembling Monomeric Nucleoside Molecular Nanoparticles Loaded with 5-FU Enhancing Therapeutic Efficacy against Oral Cancer. *ACS Nano* **2015**, *9* (10), 9638-9651.