

[Supplementary Information]

***Spider Silk-inspired Peptide Multiblock Hybrid Copolymers for Self-healable Thin Film Materials***

*Tomoyuki Koga<sup>1,\*</sup>, Tomotaka Morishita<sup>1</sup>, Yushi Harumoto<sup>1</sup>,*

*Shin-nosuke Nishimura<sup>2</sup>, Nobuyuki Higashi<sup>1\*</sup>*

<sup>1</sup>Department of Molecular Chemistry and Biochemistry, Faculty of Science and Engineering, Doshisha University, Kyotanabe, Kyoto 610-0321, JAPAN

<sup>2</sup>Institute for Materials Chemistry and Engineering, Kyushu University, Fukuoka 819-0395, JAPAN

**Corresponding Author**

\*E-mail: tkoga@mail.doshisha.ac.jp (T. K.)

\*E-mail: nhigashi@mail.doshisha.ac.jp (N. H.)

## Synthesis of amine-terminated self-assembling oligopeptides

Amine-terminated oligopeptides with different chain lengths and sequences ((Ala)<sub>6</sub>, (Ala)<sub>8</sub>, (Ala)<sub>10</sub>, (Gly)<sub>8</sub>, (Leu)<sub>8</sub>, and (Val)<sub>8</sub>) were synthesized *via* SPPS using 9-fluorenylmethoxycarbonyl (Fmoc) chemistry. The oligo(Ala) blocks were first synthesized on a H<sub>2</sub>N-(CH<sub>2</sub>)<sub>2</sub>-NH-Trt-resin using Fmoc-L-Ala-OH, Fmoc-Gly-OH, Fmoc-L-Val-OH, Fmoc-L-Leu-OH, Fmoc-β-Ala-OH, Fmoc-*deg*-COOH (3 equiv), HOBt (3 equiv), and DIPC (3 equiv) in DMF for coupling and piperidine (25%)/DMF for Fmoc removal. Fmoc-*deg*-COOH was introduced at the C- and N-termini of the peptide as a spacer. To cleave the peptide from the resin, the resin was treated with TFA/TIPS/DCM (10/5/85 [v/v/v]). The resultant amine-terminated oligopeptides were purified *via* a reprecipitation method using diethyl ether and identified *via* <sup>1</sup>H NMR and MALDI-TOF MS analyses (Fig. S1).

H<sub>2</sub>N-*deg*-(Ala)<sub>*n*</sub>-*deg*-NH<sub>2</sub>

MALDI-TOFMS: *n*=6; *m/z* 849.1 [M+H]<sup>+</sup>, 871.1 [M+Na]<sup>+</sup> (849.0 [M+H]<sup>+</sup><sub>Theory</sub>, 871.0 [M+ Na]<sup>+</sup><sub>Theory</sub>), *n*=8; *m/z* 991.7 [M+H]<sup>+</sup>, 1013.5 [M+Na]<sup>+</sup> (991.1 [M+H]<sup>+</sup><sub>Theory</sub>, 1013.1 [M+ Na]<sup>+</sup><sub>Theory</sub>), *n*=10; *m/z* 1133.3 [M+H]<sup>+</sup>, 1155.2 [M+Na]<sup>+</sup>, 1171.2 [M+K]<sup>+</sup> (1133.3 [M+H]<sup>+</sup><sub>Theory</sub>, 1155.3 [M+ Na]<sup>+</sup><sub>Theory</sub>, 1171.4 [M+ K]<sup>+</sup><sub>Theory</sub>). <sup>1</sup>H NMR (D<sub>2</sub>O, DSS): 1.4 ppm (-CH<sub>3</sub> : side chain of Ala), 2.7 ppm (H<sub>2</sub>N-CH<sub>2</sub>CH<sub>2</sub>-CO-), 3.2-3.3 ppm (-CH<sub>2</sub>NH<sub>2</sub>), 3.4-3.5 ppm (-NH-CH<sub>2</sub>CH<sub>2</sub>-O-), 3.6 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>NH-), 3.65 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>O-), 3.75 ppm (-OCH<sub>2</sub>CH<sub>2</sub>O-), 4.1-4.4 ppm (α-CH : main chain of Ala, -OCH<sub>2</sub>CO-).

H<sub>2</sub>N-*deg*-(Gly)<sub>8</sub>-*deg*-NH<sub>2</sub>

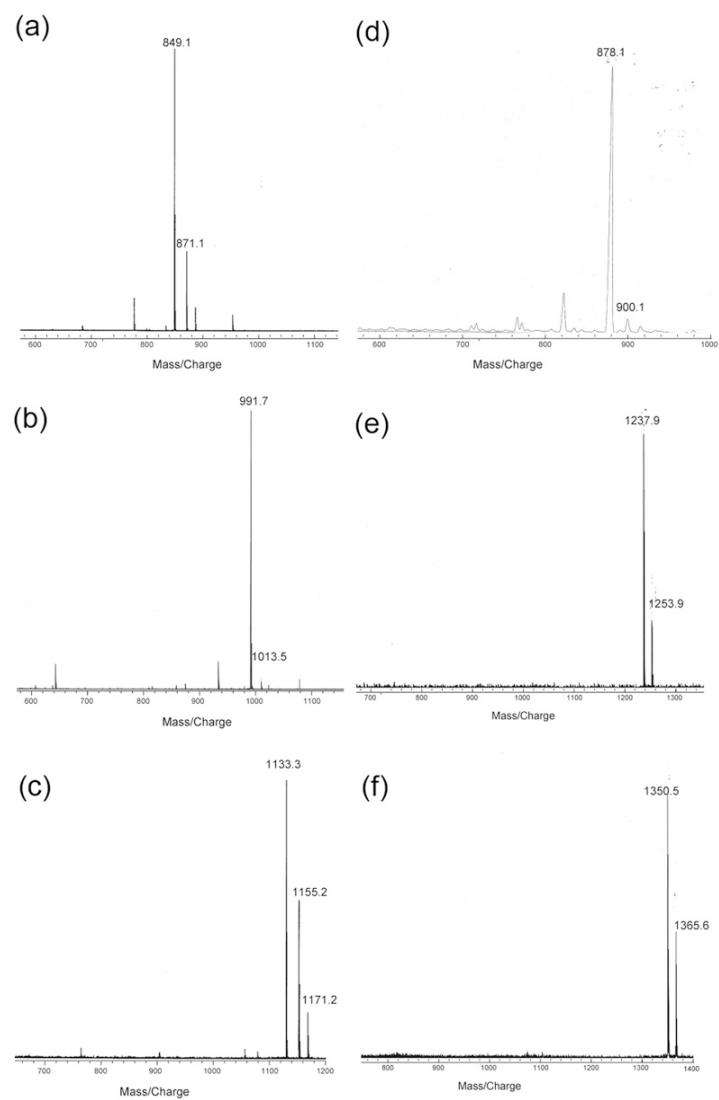
MALDI-TOFMS: *m/z* 878.1 [M+H]<sup>+</sup>, 900.1 [M+Na]<sup>+</sup> (877.9 [M+H]<sup>+</sup><sub>Theory</sub>, 899.9 [M+ Na]<sup>+</sup><sub>Theory</sub>). <sup>1</sup>H NMR (D<sub>2</sub>O, DSS): 2.7 ppm (H<sub>2</sub>N-CH<sub>2</sub>CH<sub>2</sub>-CO-), 3.2-3.3 ppm (-CH<sub>2</sub>NH<sub>2</sub>), 3.4-3.5 ppm (-NH-CH<sub>2</sub>CH<sub>2</sub>-O-), 3.6 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>), 3.65 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>O-), 3.7-3.8 ppm (-OCH<sub>2</sub>CH<sub>2</sub>O-), 4.0-4.3 ppm (-CH<sub>2</sub>- : main chain of Gly, -OCH<sub>2</sub>CO-).

H<sub>2</sub>N-*deg*-(Val)<sub>8</sub>-*deg*-NH<sub>2</sub>

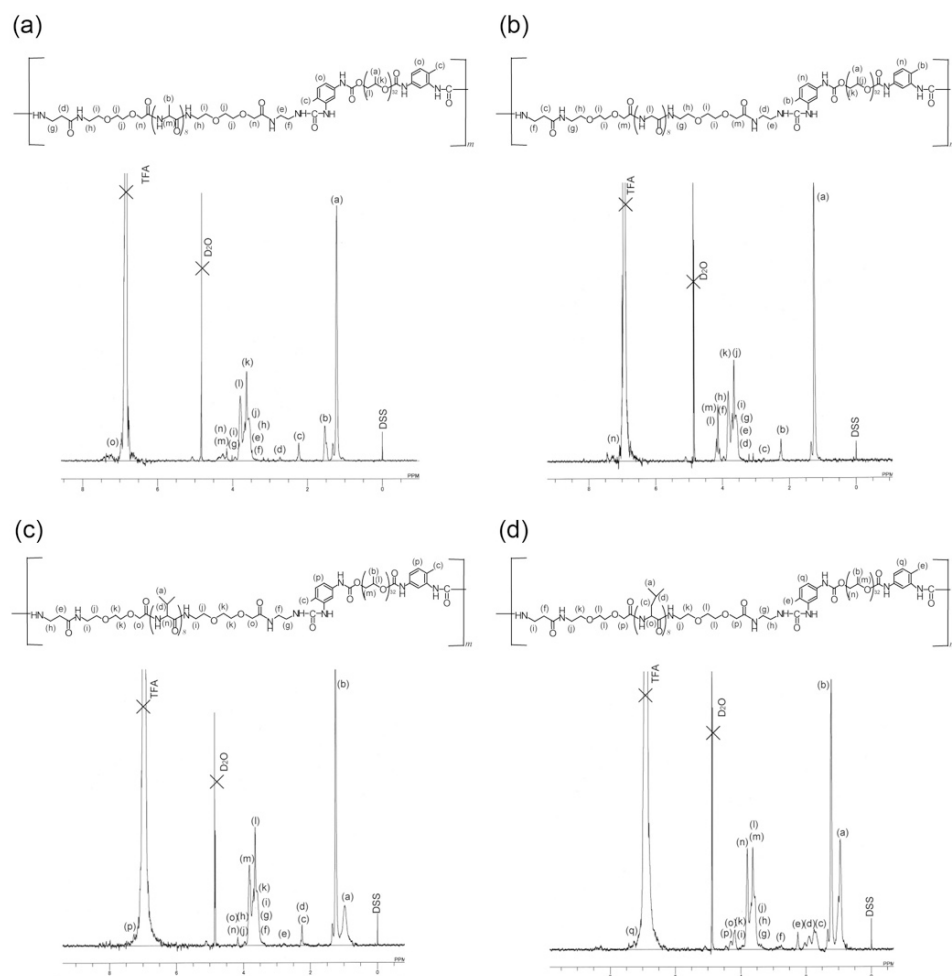
MALDI-TOFMS: *m/z* 1237.9 [M+Na]<sup>+</sup>, 1253.9 [M+K]<sup>+</sup> (1237.8 [M+ Na]<sup>+</sup><sub>Theory</sub>, 1253.9 [M+ K]<sup>+</sup><sub>Theory</sub>). <sup>1</sup>H NMR (D<sub>2</sub>O/TFA, DSS): 1.0-1.2 ppm (-CH(CH<sub>3</sub>)<sub>2</sub> : side chain of Val), 2.2 ppm (-CH(CH<sub>3</sub>)<sub>2</sub> : side chain of Val), 2.9 ppm (H<sub>2</sub>N-CH<sub>2</sub>CH<sub>2</sub>-CO-), 3.4-3.6 ppm (-CH<sub>2</sub>NH<sub>2</sub>, -NH-CH<sub>2</sub>CH<sub>2</sub>-O-), 3.7-3.8 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>, -NHCH<sub>2</sub>CH<sub>2</sub>O-), 3.9 ppm (-OCH<sub>2</sub>CH<sub>2</sub>O-), 4.2-4.6 ppm (α-CH : main chain of Val, -OCH<sub>2</sub>CO-).

H<sub>2</sub>N-*deg*-(Leu)<sub>8</sub>-*deg*-NH<sub>2</sub>

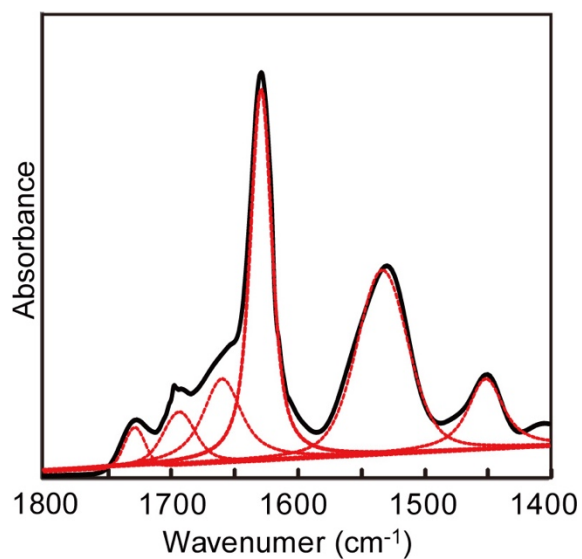
MALDI-TOFMS: *m/z* 1350.5 [M+Na]<sup>+</sup>, 1365.6 [M+K]<sup>+</sup> (1349.5 [M+ Na]<sup>+</sup><sub>Theory</sub>, 1365.6 [M+ K]<sup>+</sup><sub>Theory</sub>). <sup>1</sup>H NMR (D<sub>2</sub>O/TFA, DSS): 0.9-1.1 ppm (-CH(CH<sub>3</sub>)<sub>2</sub> :side chain of Leu), 1.6-2.0 ppm (-CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>, -CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub> : side chain of Leu), 2.8 ppm (H<sub>2</sub>N-CH<sub>2</sub>CH<sub>2</sub>-CO-), 3.3-3.5 ppm (-CH<sub>2</sub>NH<sub>2</sub>), 3.5-3.7 ppm (-NH-CH<sub>2</sub>CH<sub>2</sub>-O-, -NHCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>), 3.7-3.8 ppm (-NHCH<sub>2</sub>CH<sub>2</sub>O-), 3.8 ppm (-OCH<sub>2</sub>CH<sub>2</sub>O-), 4.1-4.4 ppm (α-CH : main chain of Leu, -OCH<sub>2</sub>CO-).



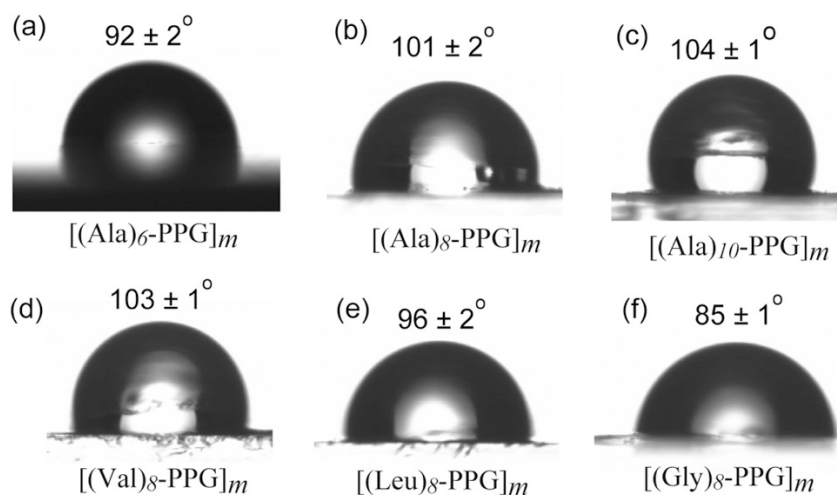
**Figure S1.** MALDI TOF MS spectra of  $H_2N\text{-deg-(Ala)}_6\text{-deg-NH}_2$  (a),  $H_2N\text{-deg-(Ala)}_8\text{-deg-NH}_2$  (b),  $H_2N\text{-deg-(Ala)}_{10}\text{-deg-NH}_2$  (c),  $H_2N\text{-deg-(Gly)}_8\text{-deg-NH}_2$  (d),  $H_2N\text{-deg-(Val)}_8\text{-deg-NH}_2$  (e), and  $H_2N\text{-deg-(Leu)}_8\text{-deg-NH}_2$  (f). Matrix: CHCA.



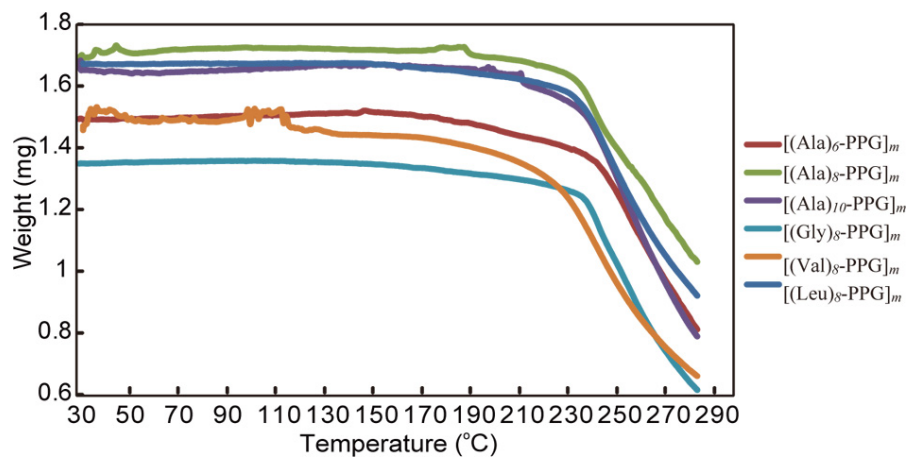
**Figure S2.**  $^1\text{H-NMR}$  spectra of  $[(\text{Ala})_8\text{-PPG}]_m$  (a),  $[(\text{Gly})_8\text{-PPG}]_m$  (b),  $[(\text{Val})_8\text{-PPG}]_m$  (c), and  $[(\text{Leu})_8\text{-PPG}]_m$  (d) in  $\text{D}_2\text{O}$  containing TFA at  $20^\circ\text{C}$ .



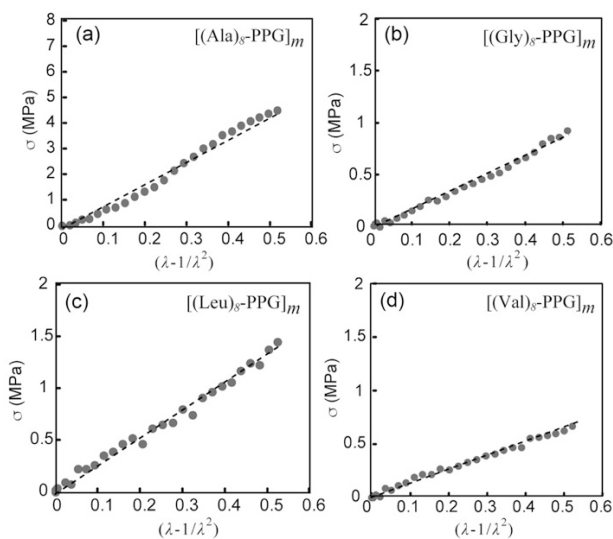
**Figure S3.** Peak deconvolution of FTIR spectrum of  $[(\text{Ala})_{10}\text{-PPG}]_m$  microfilm ( $\sim 40 \mu\text{m}$  thick).



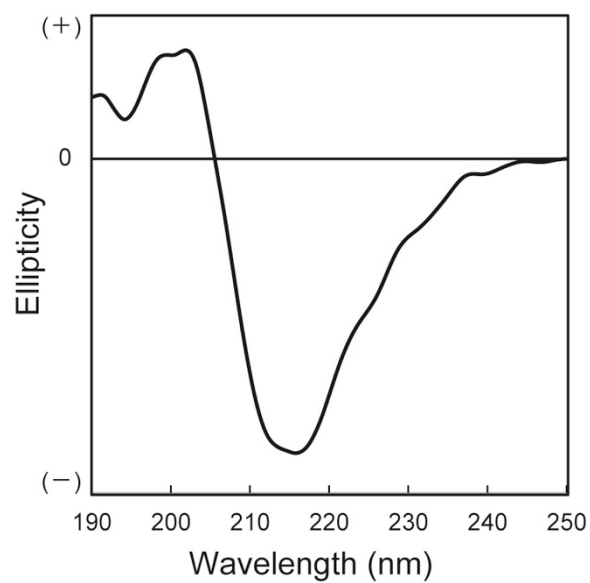
**Figure S4.** Water contact angles for various multiblock polymer films. (a)  $[(\text{Ala})_6\text{-PPG}]_m$ , (b)  $[(\text{Ala})_8\text{-PPG}]_m$ , (c)  $[(\text{Ala})_{10}\text{-PPG}]_m$ , (d)  $[(\text{Val})_8\text{-PPG}]_m$ , (e)  $[(\text{Leu})_8\text{-PPG}]_m$ , and (f)  $[(\text{Gly})_8\text{-PPG}]_m$  films.



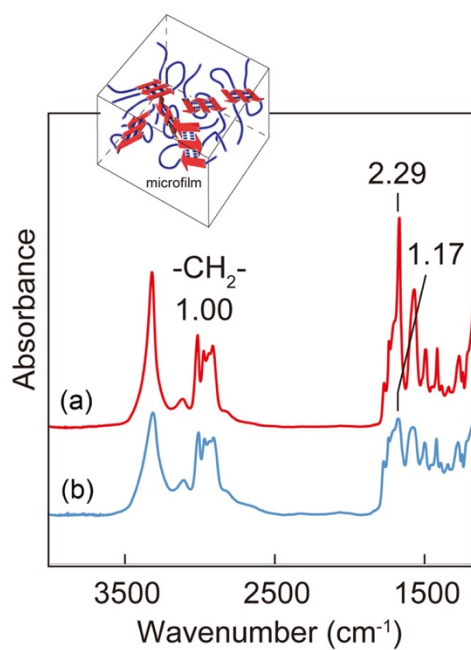
**Figure S5.** Thermogravimetric analyses of various  $[(\text{peptide})_n\text{-PPG}]_m$  films.



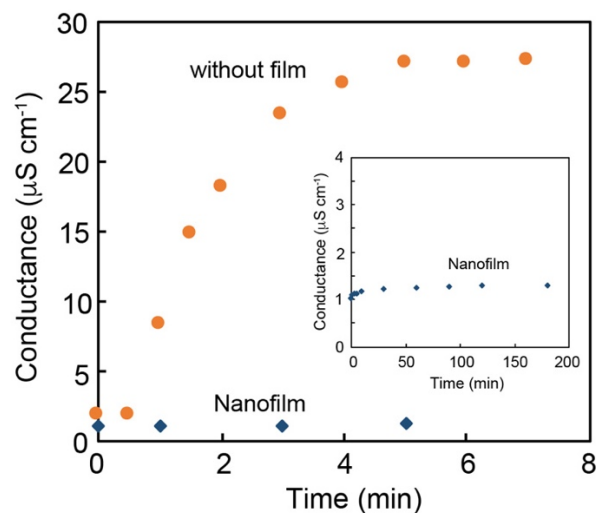
**Figure S6.** Stress vs.  $(\lambda-1/\lambda^2)$  for various  $[(\text{peptide})_8\text{-PPG}]_m$  microfilms, where slopes indicate shear modulus  $G$ .



**Figure S7.** CD spectrum of  $[(\text{Ala})_{10}\text{-PPG}]_m$  nanofilm (300 nm thickness) on quartz plate.



**Figure S8.** (a) Transmission– and (b) reflection absorption–FTIR spectra of  $[(\text{Ala})_{10}\text{-PPG}]_m$  microfilm ( $\sim 40 \mu\text{m}$  thick) on  $\text{CaF}_2$  and Au plates, respectively.



**Figure S9.** Time courses of ion permeation (*p*TSNa) through [(Ala)<sub>10</sub>-PPG]<sub>*m*</sub> nanofilm (300 nm thick). Inset shows time course of electrical conductance over a long duration (180 min).

**Table S1.** Film thickness of [(Ala)<sub>*n*</sub>-PPG]<sub>*m*</sub>-nanofilms estimated *via* AFM and SEM analyses.

Polymer	Polymer Concentration			
		0.5 wt%	1.0 wt%	2.0 wt%
[(Ala) <sub>6</sub> -PPG] <sub><i>m</i></sub>	AFM	50 nm	100 nm	290 nm
	SEM	60 nm	110 nm	300 nm
[(Ala) <sub>8</sub> -PPG] <sub><i>m</i></sub>	AFM	50 nm	170 nm	300 nm
	SEM	70 nm	150 nm	300 nm
[(Ala) <sub>10</sub> -PPG] <sub><i>m</i></sub>	AFM	140 nm	180 nm	290 nm
	SEM	150 nm	200 nm	300 nm