### **Supporting Information**

# Synthesis and biological evaluation of selective phosphonate-bearing 1,2,3-triazole-linked sialyltransferase inhibitors

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#### **Additional Synthesis**

#### **Uridine Synthons:**

#### 5'-O-Propargyl-2',3'-O-isopropylidenyluridine (7)

The acetonide protected 5'-O-propargyluridine compound (7) was synthesised by the method described by Sun *et al.*, and spectral data matched those reported.<sup>1</sup>

#### 5'-O-Propargyluridine (8)

Protected propargyl uridine (**7**, 540 mg, 1.68 mmol) was dissolved in 10 mL 9:1 ACN/H<sub>2</sub>O, with indium triflate (5% mol equiv.), for 4 hours at reflux. After reaction, the mixture was evaporated under reduced pressure and the crude product purified by column chromatography (DCM:MeOH, 9:1), to give a white foam (436 mg, 92 %).  $R_f$  0.46 (Silica, DCM:MeOH, 9:1). Spectral data matched those previously reported.<sup>1</sup>

#### Synthesis of *a*-hydroxyphosphonates:

All  $\alpha$ -hydroxyphosphonates (compounds **10a-g**) were synthesised as per the method of Montgomery *et al.*,<sup>2</sup> and **10a-d** and **10g** are characterised in that work. <sup>1</sup>H, <sup>13</sup>C, <sup>31</sup>P, and <sup>19</sup>F NMR spectra for **10e** and **10f** is provided in the supplementary information.

#### Dibenzyl a-hydroxy(3-trifluoromethyl)benzylphosphonate (10e)

From 3-trifluoromethylbenzaldehyde (300 mg, 1.72 mmol), and purified by column chromatography with a 9:1 Toluene/Acetone eluent to yield a white solid (752 mg, 78 %). R<sub>f</sub> 0.26 (Silica, Toluene/Acetone, 9:1). <sup>1</sup>**H NMR (400 MHz, CDCl<sub>3</sub>):** 7.71 (s, 1H), 7.59 (d, 1H, J = 7.8 Hz), 7.54 (d, 1H, J = 7.6 Hz), 7.42-7.36 (m, 1H), 7.32-7.20 (m, 10H), 5.10 (d, 1H,  ${}^{2}J_{(H,P)} = 10.1$  Hz), 5.04-4.91 (m, 4H), 4.59-4.38 (bs, 1H). <sup>13</sup>**C NMR (100 MHz, CDCl<sub>3</sub>):** 137.4, 135.7 (d,  ${}^{2}J_{(C,P)} = 5.8$  Hz), 130.4 (d,  ${}^{3}J_{(C,F)} = 5.6$  Hz), 128.7, 128.6, 128.0, 127.0, 124.9, 124.0 (q,  ${}^{1}J_{(C,F)} = 270.6$  Hz), 123.9, 70.5 (2 x d,  ${}^{1}J_{(C,P)} = 157.6$  Hz), 69.0-68.5 (m). <sup>31</sup>**P NMR (162 MHz, CDCl<sub>3</sub>):** 21.1 <sup>19</sup>**F NMR (376 MHz, CDCl<sub>3</sub>):** -62.6 **ESI-HRMS:** m/z calculated for C<sub>22</sub>H<sub>20</sub>F<sub>3</sub>O<sub>4</sub>PNa [M + Na]<sup>+</sup> 459.0949, found 459.0967.

#### Dibenzyl α-hydroxy(3-[1,1,2,2-tetrafluoroethoxy])benzyl phosphonate (10f)

From 3-(1,1,2,2-tetrafluoroethoxy) benzaldehyde (861 mg, 4.34 mmol), and purified by column chromatography with a 9:1 Toluene/Acetone eluent to yield a white solid (1750 mg, 70 %). R<sub>f</sub> 0.85 (Silica, Toluene/Acetone, 9:1). <sup>1</sup>**H NMR (400 MHz, CDCl<sub>3</sub>):** 7.37-7.14 (m, 14H), 5.87 (tt, 1H,  ${}^{2}J_{(H,F)}$  = 53.0 Hz,  ${}^{3}J_{(H,F)}$  = 2.5 Hz), 5.05 (d, 1H,  ${}^{2}J_{(H,P)}$  = 10.5 Hz), 5.01-4.90 (m, 4H), 2.70-2.36 (bs, 1H).<sup>13</sup>**C NMR (125 MHz, CDCl<sub>3</sub>):** 148.2, 139.2, 135.9 (t,  ${}^{3}J_{(C,P)}$  = 5.6 Hz), 129.4, 128.5-127.9 (m), 125.3 (d,  ${}^{3}J_{(C,P)}$  = 5.6 Hz), 121.0 (d,  ${}^{4}J_{(C,F)}$  = 1.8 Hz), 120.5 (d,  ${}^{3}J_{(C,P)}$  = 5.6 Hz), 116.5 (tt,  ${}^{1}J_{(C,F)}$  = 270.3 Hz,  ${}^{2}J_{(C,F)}$  = 27.8 Hz), 107.7 (tt,  ${}^{1}J_{(C,F)}$  = 249.9 Hz,  ${}^{2}J_{(C,F)}$  = 41.6 Hz), 70.3 (d,  ${}^{1}J_{(C,P)}$  = 160.1 Hz), 68.9 (d,  ${}^{2}J_{(C,P)}$  =

7.4 Hz), 68.5 (d,  ${}^{2}J_{(C,P)} = 7.5$  Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>): 21.2 <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): -88.0 (t, J = 5.4 Hz), -136.7 (t, J = 5.4 Hz). ESI-HRMS: m/z calculated for C<sub>22</sub>H<sub>20</sub>F<sub>3</sub>O<sub>4</sub>PNa [M + H]<sup>+</sup> 485.1141, found 485.1143.

#### Synthesis of α-azidophosphonates:

Dibenzyl  $\alpha$ -hydroxyphosphonate (**10a-g**, 1 equiv.) and triphenylphosphine (3 equiv.), were dissolved in dry THF under an inert atmosphere at 0 °C. Freshly prepared HN<sub>3</sub> (30 mL) was added, along with diisopropylazodicarboxyate (DIAD, 3 equiv.) dropwise, and the reaction was allowed to warm to room temperature. Upon completion (judged by TLC), the reaction mixture was taken up in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), washed with saturated NaHCO<sub>3</sub> solution (3 x 5 mL) and brine (3 x 5 mL). The organic phase was separated and dried with anhydrous MgSO<sub>4</sub> and evaporated, with the resultant product purified by column chromatography. The  $\alpha$ -azidophosphonates proved difficult to purify, as the hydrazine byproduct of DIAD seemed to 'stick' to the desired product during column chromatography, and would not readily precipitate when the crude product was taken up in a non-polar solvent such as hexane. This was not deemed an issue, as the hydrazine did not impact the proceeding click reaction and so it was not necessary for the  $\alpha$ -azidophosphonate to be completely purified. <sup>1</sup>H, <sup>13</sup>C, <sup>31</sup>P, and <sup>19</sup>F NMR spectra for these compounds is provided.

#### Dibenzyl a-azido-3-phenoxybenzylphosphonate (11a)

From **10a** (1.25 g, 2.71 mmol): purified by column chromatography using a Toluene/Acetone (1:1) eluent, to yield a white solid (1.095 g, 83.7%). R<sub>f</sub> 0.48 (Silica, Toluene/Acetone, 1:1). <sup>1</sup>**H NMR (400 MHz, CDCl<sub>3</sub>):** 7.23-7.18 (m, 12H), 7.18-7.13 (m, 2H), 7.08 (s, 1H), 7.00 (t, J = 7.3 Hz, 1H), 6.92 (d, J = 7.7 Hz, 3H), 4.97-4.82 (m, 4H), 4.67 (d,  ${}^{2}J_{(H,P)} = 16.0$  Hz, 1H). <sup>13</sup>**C NMR (100 MHz, CDCl<sub>3</sub>):** 157.6, 156.8, 135.9, 135.8, 134.3, 130.2, 129.9, 128.6, 128.2, 123.7, 123.2, 119.2, 118.7, 68.8, 61.3 (d,  ${}^{2}J_{(C,P)} = 157.2$  Hz). <sup>31</sup>**P NMR (162 MHz, CDCl<sub>3</sub>):** 19.2. **ESI-HRMS:** *m*/*z* calculated for C<sub>27</sub>H<sub>24</sub>N<sub>3</sub>O<sub>4</sub>PNa [M + Na]<sup>+</sup>: 508.1406, found 508.1402.

#### Dibenzyl a-azido-3-cyclopentoxybenzylphosphonate (11b)

From **10b** (232 mg, 0.512 mmol): purified by column chromatography using a DCM/EtOAc (99:1) eluent, to give a white solid (195 mg, 80%).  $R_f$  0.68 (Silica, DCM:EtOAc, 99:1). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 7.35-7.18 (m, 11H), 6.95-6.92 (m, 2H), 6.87-6.84 (m, 1H, H4), 5.04-4.82 (m, 4H), 4.68 (d, 1H, *J* = 16.8 Hz), 4.65 (m, 1H), 1.89-1.70 (m, 6H), 1.63-1.53 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): 158.3 (d, *J* = 2.4 Hz), 135.8 (d, *J* = 5.8 Hz), 135.7 (d, *J* = 5.9 Hz), 133.1 (d, *J* = 3.6 Hz), 129.7 (d, *J* = 1.5 Hz), 128.6, 128.5, 128.09, 128.05, 120.3 (d, *J* = 6.7 Hz), 116.5 (d, *J* = 2.6 Hz), 115.0 (d, *J* = 6.0 Hz), 79.2, 68.83 (d, *J* = 6.8 Hz), 68.79 (d, *J* = 7.2 Hz), 61.8 (d, *J* = 158.7 Hz), 32.7, 24.0. <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>): 19.3. ESI-HRMS: *m*/*z* calculated for C<sub>26</sub>H<sub>28</sub>N<sub>3</sub>O<sub>4</sub>PNa [M + Na]<sup>+</sup>: 500.1716, found 500.1709.

#### Dibenzyl α-azido-3-phenoxy-4-fluorobenzylphosphonate (11c)

From **10c** (179 mg, 0.375 mmol): purified by column chromatography using a DCM/EtOAc (99:1) eluent, to give a white solid (141 mg, 75%).  $R_f 0.83$  (Silica, DCM:EtOAc, 99:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 7.31-7.18 (m, 12H), 7.12-7.05 (m, 4H), 6.92-6.90 (m, 2H), 5.01-4.88 (m, 4H), 4.61 (d, 1H,  $J_{(H,P)} = 16.3$  Hz). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 156.8, 154.3 (d, J = 251.1 Hz), 144.0 (dd, J = 12.1, 2.8 Hz), 135.5 (2 x d, J = 5.8 Hz), 129.8, 128.9 (d, J = 3.9 Hz), 128.7, 128.6 (d, J = 3.2 Hz), 128.13, 128.08, 126.4 (d, J = 6.1 Hz), 124.5 (2 x d, J = 6.4 Hz), 123.5, 121.4 (d, J = 4.5 Hz), 117.5, 117.3 (dd, J = 19.2, 2.2 Hz), 68.9 (2 x d, J = 6.8 Hz), 60.9 (d, J = 159.2 Hz). <sup>31</sup>P NMR (202 MHz, CDCl<sub>3</sub>): 18.8. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): -131.7. ESI-HRMS: m/z calculated for C<sub>27</sub>H<sub>23</sub>FN<sub>3</sub>O<sub>4</sub>PNa [M + Na]<sup>+</sup>: 526.1308, found 526.1321.

#### Dibenzyl α-azido-4-fluorobenzylphosphonate (11d)

From **10d** (400 mg, 1.04 mmol): purified by column chromatography using a DCM/EtOAc (99:1) eluent, to give a white solid (379 mg, 89%).  $R_f 0.68$  (Silica, DCM:EtOAc, 99:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 7.36-7.27 (m, 10H), 7.21-7.19 (m, 2H), 7.00 (dd, <sup>3</sup>*J*<sub>(*H,F*)</sub> = 8.5 Hz, 2H), 5.04-4.86 (m, 4H), 4.69 (d, 1H, *J*<sub>(*H,P*)</sub> = 16.3 Hz). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 162.9 (dd, <sup>1</sup>*J*<sub>(*C,F*)</sub> = 248.0 Hz, <sup>5</sup>*J*<sub>(*C,P*)</sub> = 3.0 Hz), 135.6 (2 x d, <sup>2</sup>*J*<sub>(*C,P*)</sub> = 18.4 Hz), 130.2 (dd, <sup>3</sup>*J*<sub>(*C,P*)</sub> = 8.3 Hz, <sup>3</sup>*J*<sub>(*C,F*)</sub> = 6.4 Hz), 128.6 (m), 128.2, 128.1, 127.8 (t, <sup>3</sup>*J*<sub>(*C,P*)</sub> = 3.4 Hz), 115.8 (dd, <sup>2</sup>*J*<sub>(*C,F*)</sub> = 22.0 Hz), 68.9 (m), 61.1 (d, <sup>1</sup>*J*<sub>(*C,P*)</sub> = 160.9 Hz).<sup>31</sup>P NMR (202 MHz, CDCl<sub>3</sub>): 19.2. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): -112.3 (2 x s). ESI-HRMS: *m*/*z* calculated for C<sub>21</sub>H<sub>19</sub>FN<sub>3</sub>O<sub>3</sub>PNa [M + Na]<sup>+</sup>: 434.1046, found 434.1066.

#### Dibenzyl α-azido-3-trifluoromethylbenzylphosphonate (11e)

From **10e** (600 mg, 1.38 mmol): purified by column chromatography using a DCM/EtOAc (99:1) eluent, to give a white solid (608 mg, 96%).  $R_f 0.77$  (Silica, DCM:EtOAc, 99:1). <sup>1</sup>H NMR (**400 MHz**, **CDCl**<sub>3</sub>): 7.61 (bs, 1H), 7.57 (bs, 1H), 7.55 (bs, 1H), 7.41 (t, 1H, J = 7.8 Hz), 7.20-7.33 (m, 10H), 4.94-5.03 (m, 4H), 4.79 (d, 1H, J = 16.5 Hz). <sup>13</sup>C NMR (**100 MHz**, **CDCl**<sub>3</sub>): 135.4-135.6 (m), 133.5 (d, J = 3.7 Hz), 131.6 (d, J = 4.4 Hz), 131.0 (dq, J = 33.3, 2.4 Hz), 129.1 (d, J = 2.2 Hz), 128.8, 128.69, 128.67, 128.2, 125.0 (quint, J = 3.5 Hz), 125.0 (sext, J = 3.5 Hz), 123.8 (q, J = 272.7 Hz), 69.1, 69.0, 61.4 (d, J = 157.7 Hz). <sup>31</sup>P NMR (**162 MHz**, **CDCl**<sub>3</sub>): 18.5. <sup>19</sup>F NMR (**376 MHz**, **CDCl**<sub>3</sub>): -62.6. **ESI-HRMS**: m/z calculated for C<sub>22</sub>H<sub>19</sub>F<sub>3</sub>N<sub>3</sub>O<sub>3</sub>PNa [M + Na]<sup>+</sup>: 484.1014, found 484.1037.

#### Dibenzyl a-azido-3-(1,1,2,2-tetrafluoroethoxy)benzyl phosphonate (11f)

From **10f** (331 mg, 0.840 mmol): purified by column chromatography using a DCM/EtOAc (4:1) eluent, to give a white solid (289 mg, 83%).  $R_f$  0.95 (Silica, DCM:EtOAc, 4:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 7.36-7.20 (m, 14H), 5.89 (tt, 1H, J = 53.1, 2.8 Hz), 5.02-4.90 (m, 4H), 4.73 (d, 1H, J = 16.5 Hz). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 148.9, 135.6 (d, J = 6.1 Hz), 135.5 (d, J = 6.2 Hz), 134.3 (d, J = 4.6 Hz), 130.0 (d, J = 1.8 Hz), 128.7, 128.64, 128.62, 128.19, 128.17, 126.3 (d, J = 6.3 Hz), 121.9 (d, J = 2.4 Hz), 121.5 (d, J = 6.2 Hz), 116.5 (tt, J = 272.5, 28.7 Hz), 107.6 (tt, J = 252.0, 41.2 Hz), 69.1 (m), 61.3

(d, J = 158.7 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>): 18.6. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>): -88.1 (t, J = 6.8 Hz), -136.7 (dt, J = 53.0, 5.7 Hz). ESI-HRMS: m/z calculated for C<sub>23</sub>H<sub>20</sub>F<sub>4</sub>N<sub>3</sub>O<sub>4</sub>PNa [M + Na]<sup>+</sup>: 532.1025, found 532.1035.

#### Dibenzyl a-azido-(3-benzothiophene)methylphosphonate (11g)

From **10g** (460 mg, 1.08 mmol): purified by column chromatography, with a Hexane/EtOAc (1:1) eluent. This afforded a clear oil (390 mg, 80 %).  $R_f 0.73$  (Silica, Hexane/EtOAc, 1:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 7.86-7.75 (m, 3H), 7.38-7.35 (m, 2H), 7.34-7.29 (m, 5H), 7.27-7.21 (m, 3H), 7.14-7.11 (m, 2H), 5.01 (d, 1H, *J* = 16.8 Hz), 5.13-4.79 (m, 4H) <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 140.0, 137.3 (2 x s), 135.6 (2 x s), 135.4 (2 x s), 128.56, 128.54, 128.48, 128.43, 128.1, 128.0, 127.5 (d, *J* = 7.5 Hz), 126.1, 124.9, 124.5, 122.7, 121.9, 68.9-68.8 (m), 55.4 (d, *J* = 162.7 Hz) <sup>31</sup>P NMR (202 MHz, CDCl<sub>3</sub>): 19.1. ESI-HRMS: *m/z* calculated for C<sub>23</sub>H<sub>20</sub>N<sub>3</sub>O<sub>3</sub>PSNa [M + Na]<sup>+</sup>: 472.08552, found 472.08550.

#### **CuAAC** 'click' coupling to form 1,2,3-triazoles:

An  $\alpha$ -azidophosphonate (**11a-g**, 1 equiv.), **8** (1.2 equiv.), Cu(OAc)<sub>2</sub> (0.25 equiv.), and sodium ascorbate (0.5 equiv.) were suspended in a mixture of THF and water (1:1) and stirred at room temperature until starting material disappeared (4-12 hours). Upon completion (judged by TLC), the reaction mixture was concentrated under reduced pressure and extracted with EtOAc. The organic layer washed with brine, dried with anhydrous MgSO<sub>4</sub>, filtered and evaporated, with the resultant product purified by column chromatography.

#### 5'-O-[1-(Dibenzoxyphosphoryl-3-phenoxyphenylmethyl)-1,2,3-triazol-4-yl]methyluridine (12a)

From **11a** (150 mg, 0.309 mmol): purified by column chromatography to give white solid (100 mg, 51%).  $R_f$  0.64 (DCM/MeOH, 9:1). <sup>1</sup>H NMR (**400** MHz, CDCl<sub>3</sub>): 9.18 (2 x bs, 1H), 8.04 (2 x s, 1H), 7.73 (2 x d, J = 8.2 Hz), 7.33-7.21 (m, 10H), 7.16-7.09 (m, 6H), 7.00-6.92 (m, 3H), 6.13 (2 x d, 1H, J = 21.6 Hz), 5.80 (2 x d, 1H, J = 2.3 Hz), 5.63 (2 x d, 1H, J = 8.1 Hz), 4.97-4.80 (m, 4H), 4.69-4.59 (m, 2H), 4.39 (bs, 1H), 4.20-4.15 (m, 3H), 3.86-3.51 (m, 3H). <sup>13</sup>C NMR (**100** MHz, CDCl<sub>3</sub>): 163.3, 158.0 (2 x s), 156.3 (2 x s), 151.0, 144.6, 140.4 (2 x s), 135.1 (m), 133.6 (2 x s), 130.6 (2 x s), 129.9 (2 x s), 128.8, 128.7, 128.1 (m), 123.9 (2 x s), 123.0 (2 x s), 119.3 (2 x s), 118.8 (2 x s), 102.3 (2 x s), 90.7 (2 x s), 83.8 (2 x s), 75.4 (2 x s), 70.5 (2 x s), 69.7 (2 x s), 69.2 (2 x s), 69.0 (2 x s), 64.4 (2 x s), 61.7 (d, J = 159.1 Hz). <sup>31</sup>P NMR (**162** MHz, CDCl<sub>3</sub>): 16.4. ESI-HRMS: m/z calculated for C<sub>39</sub>H<sub>38</sub>N<sub>5</sub>O<sub>10</sub>PNa [M + Na]<sup>+</sup>: 790.2271, found 790.2254.

5'-O-[1-(Dibenzoxyphosphoryl-3-cyclopentoxyphenylmethyl)-1,2,3-triazol-4-yl]methyluridine (12b) From 11b (156 mg, 0.326 mmol): purified by column chromatography with a 9:1 DCM/MeOH eluent to afford a white foam (129 mg, 52 %).  $R_f$  0.68 (DCM/MeOH, 9:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 8.80 (2 x s, 1H), 8.02 (2 x s, 1H), 7.72 (2 x d, 1H, J = 8.2 Hz), 7.31-7.23 (m, 7H), 7.19-7.17 (m, 2H), 7.117.03 (m, 4H), 6.89-6.86 (m, 1H), 6.11 (2 x d, 1H, J = 21.3 Hz), 5.79 (2 x d, 1H, J = 3.4 Hz), 4.96-4.60 (m, 4H), 4.70-4.60 (m, 3H), 4. 5.53 (2 x dd, 1H, J = 8.1, 2.0 Hz), 24-4.10 (m, 4H), 3.86-3.45 (m, 3H), 1.91-1.72 (m, 6H), 1.67-1.57 (m, 2H). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 163.0, 158.6 (2 x s), 150.8, 144.5, 140.3 (2 x s), 135.2-135.0 (m), 132.9 (2 x s), 130.3 (2 x s), 128.8-128.6 (m), 128.1 (m), 123.0, 120.4 (m), 116.6 (2 x s), 116.0 (2 x d, J = 7.1 Hz), 102.2, 91.0, 84.0 (2 x s), 79.4, 75.6, 70.7 (2 x s), 69.7 (2 x s), 69.1 (2 x s), 68.9 (2 x s), 63.4 (2 x s), 62.0 (2 x d, J = 155.4 Hz), 32.8, 24.1. <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>): 16.8. ESI-HRMS: m/z calculated for C<sub>39</sub>H<sub>38</sub>N<sub>5</sub>O<sub>10</sub>PNa [M + Na]<sup>+</sup>: 782.2567, found 782.2598.

## 5'-O-[1-(Dibenzoxyphosphoryl-3-phenoxy4-fluorophenylmethyl)-1,2,3-triazol-4-yl]methyluridine (12c)

From **11c** (59.9 mg, 0.119 mmol): purified by column chromatography using a DCM/MeOH (9:1) eluent, to give a white solid (51.1 mg, 66%).  $R_f 0.67$  (Silica, DCM:MeOH, 9:1). <sup>1</sup>H NMR (400 MHz, MeOD): 8.16 (s, 1H), 7.86 (2 x d, 1H, J = 8.2 Hz), 7.34-7.10 (m, 19H), 6.90-6.88 (m, 2H), 6.54 (2 x d, 1H, J = 22.2 Hz), 5.88 (d, 1H, J = 4.5 Hz), 5.53 (2 x d, 1H, J = 8.3 Hz), 5.03-4.94 (m, 4H), 4.68 (m, 2H), 4.12 (m, 3H), 3.79 (m, 2H). <sup>13</sup>C NMR (100 MHz, MeOD): 164.6, 151.0, 149.0, 144.5, 140.8, 135.4, 129.6, 128.4, 127.8, 126.9, 124.3, 123.3, 121.9, 121.6 (d, J = 17.2 Hz), 117.2, 101.4, 89.0, 83.4, 74.5, 70.2, 69.4-69.2 (m), 63.4, 60.6 (d, J = 155.1 Hz). <sup>31</sup>P NMR (162 MHz, MeOD): 16.3. <sup>19</sup>F NMR (376 MHz, MeOD): -131.6 (2 x s). ESI-HRMS: m/z calculated for C<sub>55</sub>H<sub>49</sub>FN<sub>5</sub>O<sub>14</sub>PNa [M + Na]<sup>+</sup>: 808.2158, found 808.2160.

#### 5'-O-[1-(Dibenzoxyphosphoryl-4-fluorophenylmethyl)-1,2,3-triazol-4-yl]methyluridine (12d)

From **11d** (109 mg, 0.265 mmol): purified by column chromatography with a 9:1 DCM/MeOH eluent to afford a white foam (106 mg, 57 %).  $R_f 0.62$  (DCM/MeOH, 9:1). <sup>1</sup>H NMR (**400 MHz, CDCl<sub>3</sub>**): 9.52 (2 x s, 1H), 8.01 (2 x s, 1H), 7.74 (m, 1H, J = 8.1 Hz), 7.50-7.46 (m, 2H), 7.30-7.24 (m, 6H), 7.17-7.15 (m, 2H), 7.11-7.08 (m, 2H), 7.01 (2 x t, 2H, J = 8.6 Hz), 6.15 (2 x d, 1H, J = 22.2 Hz), 5.82 (2 x d, 1H, J = 3.0 Hz), 5.59 (2 x d, 1H, J = 8.4 Hz), 4.96-4.80 (m, 4H), 4.59-4.67 (m, 3H), 4.22-4.16 (m, 3H), 3.86-3.49 (m, 3H). <sup>13</sup>C NMR (**100 MHz, CDCl<sub>3</sub>**): 163.5 (2 x s), 163.4 (2 x d, J = 274.4 Hz), 151.0, 144.7 (2 x s), 140.5 (2 x s), 134.9-135.1 (m), 130.8 (m), 128.8, 128.68, 128.65, 128.15, 128.12, 128.09, 122.9 (2 x s), 116.3 (2 x d, J = 21.3 Hz), 103.0 (2 x s), 102.3 (2 x s), 90.7 (2 x s), 83.8, 75.4 (2 x s), 70.5 (2 x s), 69.8 (2 x s), 69.3 (2 x s), 69.1 (2 x s), 64.4, 61.2 (d, J = 155.8 Hz). <sup>31</sup>P NMR (**162 MHz, CDCl<sub>3</sub>**): 16.5. <sup>19</sup>F NMR (**376 MHz, CDCl<sub>3</sub>**): -111.0 (2 x m). **ESI-HRMS:** *m*/*z* calculated for C<sub>39</sub>H<sub>38</sub>N<sub>5</sub>O<sub>10</sub>PNa [M + Na]<sup>+</sup>: 716.1898, found 716.1899.

## 5'-O-[1-(Dibenzoxyphosphoryl-3-trifluoromethylphenylmethyl)-1,2,3-triazol-4-yl]methyluridine (12e)

From **11e** (159 mg, 0.344 mmol): purified by column chromatography with a 9:1 DCM/MeOH eluent to afford a white foam (156 mg, 60 %).  $R_f$  0.56 (DCM/MeOH, 9:1). <sup>1</sup>H NMR (**400 MHz, MeOD**): 8.85-8.34 (2 x s, 1H), 8.31-8.11 (2 x s, 1H), 7.71 (2 x d, 1H, J = 8.2 Hz), 7.41-7.34 (m, 7H), 7.25-7.21

(m, 2H), 7.18-7.10 (m, 4H), 6.93-6.90 (m, 1H), 6.18-6.12 (2 x d, 1H, J = 22.0 Hz), 5.80 (2 x d, 1H, J = 2.3 Hz), 5.71-5.66 (m, 1H), 5.13-4.79 (m, 4H), 4.68-4.59 (m, 2H), 4.49 (bs, 1H), 4.18-4.14 (m, 3H), 3.86-3.50 (m, 3H). <sup>13</sup>C NMR (100 MHz, MeOD): 162.6 (2 x s), 150.0, 144.7 (2 x s), 140.6 (2 x s), 135.0, 134.8 (2 x s), 134.7 (2 x s), 132.7 (2 x s), 131.9 (m), 129.8 (2 x s), 128.9-128.2 (m), 126.3 (m), 125.6 (m), 123.5 (q, J = 272.7 Hz), 123.0 (2 x s), 103.2 (2 x s), 90.2 (2 x s), 83.6 (2 x s), 75.3 (2 x s), 70.9 (2 x s), 69.7 (m), 69.5 (m), 68.9 (2 x s), 65.0 (2 x s), 61.6 (d, J = 155.3 Hz). <sup>31</sup>P NMR (162 MHz, MeOD): 15.6. <sup>19</sup>F NMR (376 MHz, MeOD): -62.7 (2 x s). ESI-HRMS: m/z calculated for  $C_{34}H_{33}F_{3}N_5O_9PNa$  [M + Na]<sup>+</sup>: 765.2158, found 765.2182.

#### 5'-O-[1-(Dibenzoxyphosphoryl-3-(1,1,2,2,-tetrafluoroethoxy)phenylmethyl)-1,2,3-triazol-4yl]methyluridine (12f)

From **11f** (18.5 mg, 0.0363 mmol): purified by column chromatography with a 9:1 DCM/MeOH eluent to afford a white foam (12.1 mg, 52 %). R<sub>f</sub> 0.61 (Silica, DCM:MeOH, 9:1). <sup>1</sup>**H** NMR (500 MHz, MeOD): 8.21 (s, 1H), 7.86 (2 x d, 1H, J = 8.2 Hz), 7.50-7.43 (m, 3H), 7.32-7.29 (m, 7H), 7.21-7.19 (m, 4H), 6.54 (2 x d, 1H, J = 22.4 Hz), 6.32 (tt, 1H, J = 52.5, 3.0 Hz), 5.88 (d, 1H, J = 4.5 Hz), 5.55 (2 x d, 1H, J = 8.0 Hz), 5.07-4.94 (m, 4H), 4.68 (m, 2H), 4.12 (m, 3H), 3.79 (m, 2H). <sup>13</sup>C NMR (125 MHz, MeOD): 164.6, 151.0, 149.0, 144.6, 140.9, 135.3, 134.4, 130.2, 128.6-127.8 (m), 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 127.5, 127.4 (2 x s), 126.9, 124.4, 122.3, 121.9, 121.6 (d, J = 17.2 Hz), 116.6 (t, J = 271.0 Hz), 108.0 (t, J = 250.6 Hz), 101.5, 89.1, 83.4, 74.5, 70.2, 69.5-69.2 (m), 65.5, 63.5, 60.6 (d, J = 155.1 Hz). <sup>31</sup>P NMR (162 MHz, MeOD): 16.1. <sup>19</sup>F NMR (376 MHz, MeOD): -89.8, -139.2. ESI-HRMS: m/z calculated for C<sub>35</sub>H<sub>33</sub>F<sub>4</sub>N<sub>5</sub>O<sub>10</sub>P [M - H]<sup>-</sup>: 790.1901, found 790.1902.

5'-O-[1-(Dibenzoxyphosphorylbenzothiophen-3-ylmethyl)-1,2,3-triazol-4-yl]methyluridine (12g) From 11g (105 mg, 0.234 mmol): purified by column chromatography with a 9:1 DCM/MeOH eluent to afford a white foam (97.7 mg, 57 %). R<sub>f</sub> 0.65 (DCM/MeOH, 9:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 8.95 (2 x s, 1H), 8.22 (2 x s, 1H), 7.88-7.80 (m, 2H), 7.67-7.64 (m, 2H), 7.38-7.35 (m, 2H), 7.32-7.29 (m, 3H), 7.08-7.07 (m, 2H), 7.24-7.17 (m, 5H), 6.59 (2 x d, 1H, J = 20.9 Hz), 5.80 (2 x d, 1H, J = 3.1Hz), 5.62 (2 x d, 1H, J = 8.1 Hz), 5.01-4.75 (m, 4H), 4.63-4.53 (m, 2H), 4.20-4.13 (m, 4H), 3.82-3.56 (m, 3H).<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): 163.0, 150.9 (2 x s), 144.8 (2 x s), 140.3 (2 x s), 139.8 (2 x s), 137.2 (d, J = 10.7 Hz), 135.03, 134.99, 134.8 (2 x d, J = 5.1 Hz), 129.0-128.6 (m), 128.14, 128.08 (2 x s), 125.5 (2 x s), 125.3 (2 x s), 125.0 (2 x s), 123.0 (2 x s), 122.8 (2 x s), 121.2, 102.2 (2 x s), 90.9 (2 x s), 83.9, 75.5 (2 x s), 70.6 (2 x s), 69.8-69.7 (m), 69.4-69.3 (m), 69.0 (2 x s), 64.5 (2 x s), 54.9 (d, J =159.1 Hz). <sup>31</sup>P NMR (162 MHz, CDCl<sub>3</sub>): 16.6 (2 x s). ESI-HRMS: m/z calculated for C<sub>35</sub>H<sub>34</sub>N<sub>5</sub>O<sub>9</sub>PSNa [M + Na]<sup>+</sup>: 754.1717, found 754.1713.

#### **CMP-Glo based Sialyltransferase Inhibition Assay**

Recombinant human ST3Gal I and ST6Gal I were obtained from R&D Systems, CMP-Neu5Ac (purified prior to use by size exclusion chromatography), Gal- $\beta$ 1,3-GalNAc and LacNAc were from Carbosynth. Assays were performed in a sodium cacodylate buffer (5.0 mM sodium cacodylate, 15.0  $\mu$ M NaCl, 0.05 % Triton X-100). Assays were performed in a solid white 96-well plate, in a 25  $\mu$ L volume for one hour, incubated at room temperature. CMP detection reagent was prepared as per Promega's guidelines, and 25  $\mu$ L was added, with luminescence measured after a further hour of incubation. In each assay a CMP standard curve was established in duplicate, with concentrations ranging from 0-100 $\mu$ M.

#### **Enzyme activity curve**

To determine the amount of enzyme to use in each assay, an assay was performed using amounts of enzyme ranging from 0-500 ng/well. The amount of enzyme was added in 15  $\mu$ L of assay buffer, to which was added 10  $\mu$ L of a mixture containing 25  $\mu$ M CMP-Neu5Ac and 2.5 mM acceptor (Gal- $\beta$ 1,3-GalNAc and LacNAc for ST3Gal I and ST6Gal I respectively). The resultant sigmoidal activity curve of luminescence vs quantity of enzyme showed a linear region of response, which gave a guideline as to the amount of enzyme that should be used for subsequent reactions.



Internolation	
morpolation	Luminescence (RLU)
I	Y
Sigmoidal, 4PL, X is log(concentration)	
Best-fit values	
Тор	193637
Bottom	3751
LogIC50	68.26
HillSlope	0.01169
IC50	1.824e+068
Span	189886
95% CI (asymptotic)	
Тор	178245 to 209028
Bottom	-30104 to 37606
LogIC50	50.31 to 86.21
HillSlope	0.006578 to 0.01679
IC50	2.038e+050 to 1.632e+086
Span	147449 to 232323
Goodness of Fit	
Degrees of Freedom	20
R square	0.9756
Adjusted R square	0.972
Absolute Sum of Squares	1724622611

Figure S1. Enzyme-activity curve for ST6Gal I in the CMP-Glo<sup>TM</sup> assay. The assay gave a  $R^2 = 0.9756$  for the sigmoidal response curve.



**Figure S2.** Enzyme-activity curve for ST3Gal I in the CMP-Glo<sup>TM</sup> assay. The assay gave a  $R^2 = 0.9942$  for the sigmoidal response curve.

#### Determination of CMP-Neu5Ac Km against hST6Gal I

CMP-Neu5Ac was diluted to 1250, 625, 312.5, 156.3, 78.1, 39.1, 19.5, 9.8, 4.9, 2.4, 1.2, and 0  $\mu$ M, while the enzyme was diluted to 60 ng/5  $\mu$ L. In duplicate on a solid white 96-well plate, 10  $\mu$ L of donor, 10  $\mu$ L of 2.5 mM acceptor, and 5  $\mu$ L of enzyme were added. The assay was then performed as per the general procedure detailed above. The  $K_m$  was calculated using non-linear regression analysis with GraphPad Prism 7.



**Figure S3.** Non-linear regression analysis in Michaelis-Menten equation of CMP-Neu5Ac with recombinant hST6Gal I. This is the same data as from our previous work.<sup>2</sup>

#### Single point inhibition at 100 and 10 $\mu$ M

Enzyme was diluted to 20 ng/5  $\mu$ L and 60 ng/5  $\mu$ L for ST3Gal I and ST6Gal I respectively. To a solid white 96 well plate, 10  $\mu$ L of a mixture of 2.5 mM acceptor and 250  $\mu$ M donor in assay buffer, 10  $\mu$ L

of a 250 or 25  $\mu$ M solution of inhibitor in assay buffer (for inhibition at 100 or 10  $\mu$ M respectively), and 5  $\mu$ L of enzyme solution were added to each well in duplicate. A positive control with no inhibitor was also prepared, as well as a negative control where no enzyme was present. The assay was then performed as per the general procedure detailed above. Percentage inhibition was calculated relative to the positive control, with the negative control used as a blank.

#### Determination of inhibitor K<sub>i</sub> against hST6Gal I

CMP-Neu5Ac was diluted to 2500, 625, 156.3, 39.1, and 9.8, while the enzyme was diluted to 60 ng/5  $\mu$ L. Inhibitors were diluted to three concentrations, usually between 0.5-62.5  $\mu$ M. To a solid white 96 well plate, 5  $\mu$ L of CMP-Neu5Ac solution, 5  $\mu$ L of 5 mM acceptor in assay buffer, 10  $\mu$ L of inhibitor solution, and 5  $\mu$ L of enzyme solution were added to each well in duplicate. The assay was then performed as per the general procedure detailed above. The *K*<sub>i</sub>'s were calculated using non-linear regression analysis with GraphPad Prism 8.



Figure S4. Non-linear regression analysis in of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three 13a-s concentrations.

		Velocity vs. CMP	-Neu5Ac			1/V-1/[S]	
2×10 <sup>-15</sup> .				<ul> <li>■ 25000 nM</li> <li>▲ 5000 nM</li> <li>▼ 1000 nM</li> </ul>	1300-16	7 9 Mc 19	137-72-14 0.9553 7 = 72-13 + 72-14 8 <sup>2</sup> = 0.9937
2 1×10 <sup>-16.</sup> 5×10 <sup>-16.</sup>		200		-200	50043 6-04 1005-04 5005-00 5006-00	100-64 2007-04 1007-04	4.00E+04 5.00E+04
		[CMP-Neu5Ac]	μ <b>Μ</b>	B 5000 nM	C 1000 nM	D Global (shared)	
		4					
	1	Noncompetitive inhibition					-
	2	Best-fit values					
	3	Vmax	2.200e-015	2.200e-015	2.200e-015	2.200e-015	
	4	1	= 25000	= 5000	= 1000		
	5	Ki	53686	53686	53686	53686	
	6	KM	201.9	201.9	201.9	201.9	
	7	Std. Error					
	8	Vmax	4.707e-017	4.707e-017	4.707e-017	4.707e-017	
	9	Ki	10220	10220	10220	10220	
	10	KM	29.35	29.35	29.35	29.35	
	11	95% CI (asymptotic)					
	12	Vmax	2.101e-015 to 2.299e-015	2.101e-015 to 2.299e-015	2.101e-015 to 2.299e-015	2.101e-015 to 2.299e-015	
	13	Ki	32214 to 75159	32214 to 75159	32214 to 75159	32214 to 75159	
	14	KM	140.3 to 263.6	140.3 to 263.6	140.3 to 263.6	140.3 to 263.6	
	15	Goodness of Fit					
	16	Degrees of Freedom				18	
	17	R squared	0.9635	0.9868	0.9805	0.9797	

**Figure S5.** Non-linear regression analysis of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three **13a-***l* concentrations.



Figure S6. Non-linear regression analysis of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three 13c-s concentrations.



**Figure S7.** Non-linear regression analysis of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three **13c-***l* concentrations.



Figure S8. Non-linear regression analysis of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three 13f-s concentrations.



**Figure S9.** Non-linear regression analysis of velocity vs [CMP-NeuAc] with recombinant hST6Gal I at three **13f-***l* concentrations.

#### **References:**

- 1. J. Sun, R. Liu, Q. Fu, J. Zang, Q. Tao, J. Wu and H. Zhu, *Helv. Chim. Acta*, 2014, **97**, 733-743.
- 2. A. P. Montgomery, C. Dobie, R. Szabo, L. Hallam, M. Ranson, H. Yu and D. Skropeta, *Bioorg. Med. Chem.*, 2020, **28**, 115561.





		21.059		Current NAME EXPNO PROCNO	Data Parameters CD36 9 1
10e ( <sup>31</sup> P NMR)				F2 - Acq Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 D11 TD0 SFO1 NUC1 P1 PLW1 SFO2 NUC2 CPDPRG[2 PCP2 PLW2 PLW12 PLW13	uisition Parameters 20171020 1.25 h spect 2108618_0921 ( zgpg30 65536 CDC13 16 4 64102.563 Hz 1.956255 Hz 0.5111808 sec 196.38 7.800 use 6.50 use 297.6 K 2.00000000 sec 0.03000000 sec 161.9796378 MHz 31P 15.00 use 11.77099991 W 400.1616006 MHz 1H waltz16 90.00 use 11.52400017 W 0.27886000 W 0.14026000 W
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			<b>I</b>					NAM EXP PRO	2 10 2N0	CD9 2 1	
10f ( <sup>31</sup> P NN	MR)							F2 Date Timm INS: PRO: PUL: TD SOL' NS DS SWH FID: AQ RG DW DE TE D1 D11 D11 D11 D11 D11 D11 TD0 SF00 NUC: CPDD PCPI PLW SF02 SI SSB LB GB PC	- Acqu - Acqu - ROG PROG PROG RES 1 1 2 PRG[2 2 12 13 - Proc	isition Parameters 20190205 10.57 h spect 2108618_0921 ( 2gpg30 65536 CDC13 16 4 64102.563 Hz 1.956255 Hz 0.5111808 sec 196.38 7.800 use 299.9 K 2.00000000 sec 0.03000000 sec 161.9796378 MHz 31P 15.00 use 11.7709991 W 400.1616006 MHz 11.7709991 W 400.1616006 MHz 11.52400017 W 0.27886000 W 0.14026000 W tessing parameters 32768 161.9877372 MHz EM 0 1.00 Hz 0 1.00 Hz	
	100	50	0	-50	-100	-150	-200	ppm			







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11a ( <sup>31</sup> P NMR)									Current NAME EXPNO PROCNO	Data Parameters LH14 (C and P) 2 1
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	00	50	0	-50	-100	-150	-200	ppm		





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0

-40

**-60** 

**-80** 

-100

-140

-160

-180

-200 ppm

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-120














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							PLW2 PLW12 PLW13	11.52400017 W 0.27886000 W 0.14026000 W
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100	50	0	-50	-100	-150	-200	ppm	

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		62.571								BF	UKER
F <sub>3</sub> C N <sub>3</sub> O P O.		I								Current Da NAME EXPNO PROCNO	ata Parameters CD77 7 1
11e ( <sup>19</sup> F NMR	)									F2 - Acqu. Date_ Time INSTRUM PROBHD D SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 D12 TD0 SF01 NUC1 P1 PLW1 SF02 NUC2 CPDPRG[2 PCPD2 PLW2 PLW12	Lsition Parameters 20180706 1.16 h spect 2108618_0921 ( 2gfhigqn.2 131072 CDC13 16 4 89285.711 Hz 1.362392 Hz 0.7340032 sec 196.38 5.600 usec 6.50 usec 298.8 K 1.0000000 sec 0.0300000 sec 10000200 sec 1376.4889418 MHz 19F 15.00 usec 17.75399971 W 400.1616006 MHz 1H waltz16 90.00 usec 1.52400017 W 0.27886000 W
										F2 - Proce SI SF WDW SSB ()	essing parameters 65536 376.5265944 MHz EM 0
										LB GB PC	0.30 Hz 0 1.00
0 -2	20 -40	-60	-80	-100	-120	-140	-160	-180	-200 p	pm	





$F \xrightarrow{F} 0 \xrightarrow{N_3} 0 \xrightarrow{P} 0$ $F \xrightarrow{F} 0 \xrightarrow{I} 0 \xrightarrow{I} 0$ $F \xrightarrow{I} 0 \xrightarrow{I} 0$ $I 11f (^{31}P NMR)$		18.58					Current NAME EXPNO FROCNO FROCNO FROBHD FULPRO TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1	Data Parameters CD54 B3P39 P+F 2 1 cquisition Parameters 20190801 10.03 h spect 2108618_0921 ( 3 zg30 65536 C CDC13 4 0 64102.563 Hz 1.956255 Hz 0.5111808 sec 196.38 7.800 usec 298.8 K 2.0000000 sec
n ma di kan bahan pangangan kan bahan kan bahan kan bahan bahan bahan bahan bahan bahan bahan bahan bahan bahan Mangangan pertemangkan gan mangan pertemangan pertemangan bahan bahan bahan bahan bahan bahan bahan bahan bahan Mangangan pertemangkan gan mangan bahan	de fra de de fille de se de de la deserve na forma de se de se na forma de se d	ulaid isili dia va fala budi ni dia pia aska bi 1914 isili dia ya fala budi ni dia pia pia pia pia pia pia pia pia pia p	a 11 an 1 19 19 19 19 19 19 19 19 19 19 19 19 19 1	tering meneral products and a second s	dalan ya da ka sa ka Na ka sa ka sa Na ka sa	d dad beskil statistister versierer Ar an generger versierer	P1 PLW1 F2 - P1 SF WDW SSB LB GB PC	15.00 usec 11.77099991 W cocessing parameters 32768 161.9877372 MHz EM 0 1.00 Hz 0 1.40
100	<b>50</b>	0	-50	-100	-150	-200	ppm	























	Current Da NAME EXPNO PROCNO	ata Parameters CD1071 5 1	
ŇH ↓ O	F2 - Acqui Date_ Time INSTRUM C PROBHD 2 PULPROG TD SOLVENT NS DS SWH FIDRES AQ DW FIDRES AQ DW DE TE D1 D1 D1 D1 D1 TE D1 D1 TD0 SF01 NUC1 F1 PLW1 SF02 PLW2 PLW12 PLW12	isition Paramet 20191214 18.13 2AB AV4 500 MH 2150364_0005 ( 2gpg30 65536 CDC13 16 4 81967.211 2.501441 0.3997696 101 6.100 2.8000 0.03000000 0.03000000 0.03000000 1202.2899643 31P 12.00 45.76100159 499.7459990 1H waltz16 80.00 15.53100014 0.34944999 0.17549001	ers h Z BASIC Hz Hz sec usec usec K sec sec MHz usec W MHz usec W W W W W
	F2 - Proce SI SF WDW SSB LB GB PC	essing paramete 32768 202.3000793 EM 0 1.00 0	rs MHz Hz

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1	100	50	0	-50	-100	-150	-200	ppm

2Na<sup>+</sup>

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 $N \equiv N$ 

13a-(*l*) (<sup>31</sup>P NMR)













								C	
2012+		0 						Current NAME EXPNO PROCNO	Data Parameters cd9_200213_53 6 1
			Ö					F2 - Acc Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH	uisition Parameters 20200213 22.04 h CAB AV4 500 MHZ BASIC 2150364_0005 ( 2gpg30 65536 D20 16 4 81967.211 Hz
	13b-( <i>l</i> ) ( <sup>31</sup> P N	IMR)						FIDRES AQ RG DW DE TE D1	2.501441 Hz 0.3997696 sec 101 6.100 usec 18.00 usec 286.1 K 2.0000000 sec
								D11 TD0 SF01 NUC1 P1 PLW1 SF02 NUC2 CPDPRG[2 PCPD2 PLW2 PLW2 PLW12	0.03000000 sec 1 202.2899643 MHz 31P 12.00 usec 45.76100159 W 499.7459990 MHz 1H waltz16 80.00 usec 15.53100014 W 0.34944999 W
								PLW13 F2 - Pro SI SF WDW SSB LB GB PC	0.17549001 W cessing parameters 32768 202.3000793 MHz EM 0 1.00 Hz 0 1.40
	A A A SA A A A A A A A A A A A A A A A						an and a state of the state of		
1	100	50	0	-50	-100	-150	-200	ppm	

7.98



216.72		162.88 158.43	145.09 141.09 139.17 136.91 136.91 131.34 127.31 127.31	126.07 124.76 122.51 118.25 117.87 116.38 116.38 115.38	90.36	84.52 75.78 71.54 70.18	60.48	4 0 0	1		BR	UKER	
	$2Na^{\dagger}$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$ $(-) = 0$	<sup>3</sup> C NMR)									Current I NAME EXPNO PROCNO F2 - Acqu Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 D11 TD0 SF01 NUC1 P1 PLM1 SF02 NUC2 CEPDRG[2 PCPD2 PLW2 PLW13 F2 - Proc SI SF WDW SSB LB GB PC	)ata Parameters CD67_s 8 1 iisition Parameters 20181012 0.17 h CAB AV4 500 MHZ BAS Z150364_0005 ( 2gg30 65536 D20 2056 4 30120.482 Hz 0.919204 Hz 1.0878977 sec 101 16.600 usec 288.0 K 2.00000000 sec 0.03000000 sec 125.6732948 MHz 13C 10.00 usec 51.23600006 W 499.7459990 MHz 1H waltz16 80.00 usec 15.53100014 W 0.34944999 W 0.17549001 W cessing parameters 32768 125.6605389 MHz EM 0 1.00 Hz 0 1.40	IIC
1	200 180	160	140	<b>120</b> 1	00	80	60	40	20	0	ppm		







			143.63 141.78 138.97 136.77	131.29 130.51 127.31 126.54	116.81 1116.81		102.29	90.22		 70.20 66.46 65.48	64.68	49.49					BF	UKE	R
																	Current NAME EXPNO PROCNO	Data Parameter CD67_	s 1 4 1
	2Na <sup>+</sup> 0 0 - - - - - - - - - - - - - - - - -	3 <b>c</b> -( <i>l</i> ) (	$\sum_{n=1}^{N=N}$	о он IR)	D D D D D D D D D D D D D D	H >>O											$F2 - Act Date_Time INSTRUM PROBHD PULPROG TD SOLVENT NS SWH FIDRES AQ RG DW DE TE D1 D11 TD0 SF01 NUC1 P1 PLW1 SF02 NUC2 CPDPRG I$	cquisition Param 2018101 19.4 CAB AV4 5001 Z150364_0005 Zgpg3 6553 D2 200 30120.48 0.91920 1.087897 100 16.60 18.00 2.0000000 125.673294 133 10.0 51.2360000 499.745999 2 waltz1	eters 1 9 h MHZ BASIC ( 0 6 0 0 2 Hz 4 Hz 7 sec 1 0 usec 0 kc 0 sec 1 0 sec 1 8 MHz C 0 usec 6 W 0 MHz H 6
																	PCPD2 PLW2 PLW12 PLW13 F2 - Pr	80.0 15.5310001 0.3494499 0.1754900 rocessing parame 3276	0 usec 4 W 9 W 1 W ters 8
																	SF WDW SSB LB GB PC	125.660553 E 1.0 1.4	MHz M O Hz O
hallense konstanten an der		un an					h jerren an der dat het.	and in the second s	nias iškielis ir Vientiniaujus				la an	internet and the second	ngen ski generalijska sta	lagi pirati ka di si ka di	i st		
 170	160	150	<b>140</b>	130	<b>120</b>	110	<b>100</b>	90	80	 70	<b>60</b>	<b>50</b>	<b>40</b>	<b>30</b>	<b>20</b>	ק ומק	ריי <b>n</b>		









Current	Data Parameters
NAME	CD104s
EXPNO	5
PROCNO	1
F2 - Acq	uisition Parameters
Date_	20191220
Time	6.23 h
INSTRUM	CAB AV4 500 MHZ BASIC
FROBHD	Z150364_0005 (
PULPROG	zgpg30
TD	65536
SOLVENT	D20
NS	16
DS	4
SWH	81967.211 Hz
FIDRES	2.501441 Hz
AQ	0.3997696 sec
RG	101
DW	6.100 usec
DE	18.00 usec
TE	298.0 K
D1	2.00000000 sec
D11	0.03000000 sec
TD0	1
SF01	202.2899643 MHz
NUC1	31P
P1	12.00 usec
PLW1	45.76100159 W
SF02	499.7459990 MHz
NUC2	1H
CPDPRG[2	waltz16
PCPD2	80.00 usec
PLW2	15.53100014 W
PLW12	0.34944999 W
PLW13	0.17549001 W
F2 - Pro SI WDW SSB LB	cessing parameters 32768 202.3000793 MHz EM 0 1.00 Hz
GB	0
PC	1.40



-8.44



2Na <sup>+</sup> O O F	$\frac{0}{N} = N$ $\frac{13d}{(s)} (^{19}F NM)$	NH NH NH NH NH NH NH NH NH NH NH NH NH N	0			-112.85	Current NAME EXPNO PROCNO F2 - AC Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS SSWH FIDRES AQ RG DW DE TE D1 TD0 SSOL	Data Parameters 19F 1 1 1 1 1 1 1 1 1 1 1 1 1
-85		-95	-100	-105	-110	-115	F2 - Pr SI SF WDW SSB LB GB PC	rocessing parameters 65536 376.5265940 MHz EM 0 0.30 Hz 0 1.00

	8 8 		7.39	7.01		5.81 5.81 5.81	5.44	11	4.20 4.18 4.17 4.15	4.14 4.13 3.98 3.88 3.88 3.87 3.87 3.86	BF	UKER
											Current NAME EXPNO PROCNO	Data Parameters CD1041 1 1
		$2Na^+$ $0^-/P$ F 13c	<sup>N</sup> =N ○ 	NH NH OH OH	ò						F2 - Ac Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 TD0 SF01 NUC1 P1 PLW1	quisition Parameters 20191219 11.46 h CAB AV4 500 MHZ BASIC Z150364_0005 ( zg30 65536 D20 16 2 10000.000 Hz 0.305176 Hz 3.2767999 sec 92.3077 50.000 usec 16.23 usec 298.0 K 1.00000000 sec 1 499.7470859 MHz 1H 12.00 usec 15.53100014 W
											F2 - Pr SI SF WDW SSB LB GB GB PC	ocessing parameters 65536 499.7439587 MHz EM 0 0.30 Hz 0 1.00
	lul						M					
9.0	8.5	8.0	7.5	7.0	6.5	6.0	5.5	5.0	4.5	4.0 pr	) m	

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C N E F	Current Data NAME EXPNO PROCNO	Parameters CD1041 5 1	
F C I I F F T S N	72 - Acquisit Date_ Cime INSTRUM CAB ROBHD Z150 PULPROG CD SOLVENT IS	ion Paramet 20191220 2.18 AV4 500 ME 364_0005 ( zgpg30 65536 D20 16	ers h IZ BASIC
L S F L L L L L L L L L L L L L L L L L	SSWH FIDRES AQ RG WW DE E D1	4 81967.211 2.501441 0.3997696 101 6.100 18.00 298.0 2.0000000	Hz Hz usec usec K sec
L S N F F S N N	D11 ED0 SFO1 2 UUC1 P1 PLW1 4 SFO2 4 UUC2	0.03000000 1 202.2899643 31P 12.00 5.76100159 199.7459990 1H	sec MHz usec W MHz
כ ד ד ד ד	CPDPRG[2 PCPD2 PLW2 1 PLW12 PLW13 PCW13 PCP2 - Processi	waltz16 80.00 5.53100014 0.34944999 0.17549001	usec W W W
S S M S S I C C E	SI SF 2 ADW SSB SB SB SB SC	32768 202.3000793 EM 0 1.00 0 1.40	MHz Hz



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					10	1.30	,
					GB	0	) )
					SSB J D	0	1
					SF	161.9877372	MHz
					F2 - Pro	cessing paramet	ers
					PLW1	11.77099991	W
					NUC1 Pl	31P 15.00	usec
					TD0 SF01	1 161.9796378	MHz
					D1	299.2	sec
130-(3)					DE	50.00	usec
13e-(s)	$(^{31}\mathbf{P}\mathbf{NMR})$				RG	196.38	11500
	он он				FIDRES AO	1.956255 0.5111808	Hz sec
					DS SWH	0 64102.563	) Hz
					NS	4	ł
	N ~ ~	С			TD	65536	5
					PROBHD PULPROG	Z108618_0921 ( zg30	
_ 0_	NH				INSTRUM	spect	
2Na <sup>+</sup>	Ű				Date_ Time	20191217 3.42	h
	0				F2 - Acq	uisition Parame	ters
					PROCNO	1	
					NAME	CD108s	
					Current	Data Parameters	

-8.01



									C	
									Current NAME EXPNO PROCNO	Data Parameters CD108s 1
$2Na^{+}$ $O_{-} O_{-} N_{-} N_{-} N_{-} O_{-} $	NH NH NH NH NH NH NH NH NH NH NH NH NH N								PROCNO F2 - Acc Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 TD0 SF01 NUC1 P1 PLW1	1 quisition Parameters 20191217 3.43 h spect 2108618_0921 ( zgflqn 65536 D20 4 0 89285.711 Hz 2.724784 Hz 0.3670016 sec 196.38 5.600 use 299.2 K 1.00000000 sec 1 376.4889413 MHz 19F 15.00 use 17.75399971 W
									F2 - Pro SI SF WDW SSB LB GB PC	ocessing parameters 65536 376.5265940 MHz EM 0 0.30 Hz 0 1.00
	-40 -6	0 -80	-100	-120	-140	-160	-180	-200	ppm	

--62.22









-100

-150

-200

ppm

-50

0

100

**50** 



	Current Da	ta Parameters
	NAME	CD1081
	EXPNO	6
	PROCNO	1
	1 ROCKO	±
	F2 - Acqui	sition Parameters.
	Date_	20191217
	Time	4.48 h
	INSTRUM	spect
	PROBHD 7	108618 0921 (
<b>N</b>	PULPROG	zaflan
NH	TD	65536
		00000
$\wedge$	SOLVENI	DZQ
<b>`</b> 0	NS	4
	DS	0
	SWH	89285.711 Hz
	FIDRES	2.724784 Hz
	AO	0.3670016 sec
	BG	196.38
	DW	5 600 11560
		100 00 usec
	DE	IZU.UU USEC
	ΤE	299.1 K
	D1	1.00000000 sec
	TDO	1
	SF01	376.4889413 MHz
	NUC1	19F
	P1	15.00 usec
	DIW1	17 75399971 W
	L TM T	17.73399971 W
	F2 - Proce	ssing parameters
	SI	65536
	SF	376.5265940 MHz
	WDW	EM
	SSB	
	I B	030 =-
		0.30 HZ
	GB	U
	PC	1.00



--62.14

-60

-40

-20

0

-80

-100

-140

-120

-160

-180

-200

ppm







	-88.31				$\underbrace{-137.61}_{-137.65}$		BR	UKER
$2Na^{\dagger}$ $F = F$ $F = F$ $I3f-(s) (^{19}F NM)$	O NH NH O O O O H O H O H O H O H O H O H O H O H O H O H O H O O H O O H O H O O H O H O H O H O H O H O O H O O H O O H O H O H O H O H O H O H O H O O H O O H O O H O O H O O H O O O O O H O O O O O O O O O O O O O						Current DA NAME EXPNO FROCNO F2 - Acqu. Date_ Time INSTRUM FROBHD FULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 D11 D12 TD0 SFO1 NUCL P1	ata Parameters CD66_s 6 1 isition Parameters 20180604 22.10 h spect 2108618_0921 ( 2gfhigqn.2 131072 D20 16 4 89285.711 Hz 1.362392 Hz 0.7340032 sec 196.38 5.600 usec 299.1 K 1.0000000 sec 0.0300000 sec 0.0300000 sec 1 376.4889418 MHz 19F 15.00 usec
							PLW1 SF02 CPDPRG[2 PCPD2 PLW2 PLW12	17.75399971 W 400.1616006 MHz HH waltz16 90.00 usec 11.52400017 W 0.27886000 W
Na an an airs an			nak da cara tang akaly sa panakaji kata			111 11 11 11 11 11 11 11 11 11 11 11 11	F2 - Proce SI SF WDW SSB LB GB PC	essing parameters 65536 376.5265944 MHz 0 0.30 Hz 0 1.00
-40 -50 -60 -	70 -80 -	90 -100	-110 -12	:0 -130	-140 ·	-150 -160	ppm	







	° L		Current NAME EXPNO PROCNO	Data Parameters CD66_1 6 1
			F2 - Acq Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS	uisition Parameters 20180605 2.24 h spect 2108618_0921 ( zgpg30 65536 D20 16
13f-( <i>l</i> ) ( <sup>31</sup> P NMR)	)		DS SWH FIDRES AQ RG DW DE TE D1 D11 TD0 SF01 NUC1 P1 P1W1 SF02 NUC2 CPDPRG[2 PCPD2 PLW2 PLW2 PLW12	4 64102.563 Hz 1.956255 Hz 0.5111808 sec 196.38 7.800 usec 299.1 K 2.00000000 sec 0.03000000 sec 1 161.9796378 MHz 31P 15.00 usec 11.77099991 W 400.1616006 MHz 1H waltz16 90.00 usec 11.52400017 W 0.27886000 W
te en		n has be added a to a she water to a	PLW13 F2 - Pro SI SF WDW SSB LB GB PC	0.14026000 W ocessing parameters 32768 161.9877372 MHz EM 0 1.00 Hz 0 1.40
-100	-150	-200	ppm	

- 7.60

100

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0

2Na<sup>+</sup>

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0 Ο

-50

-87.78 -88.17 -88.71	-137.59	BRUKER
$2Na^{+} \qquad \qquad$		Current Data Parameters NAME CD66_1 EXPNO 5 PROCNO 1 F2 - Acquisition Parameters Date_ 20180605 Time 2.21 h INSTRUM spect PROBHD Z108618_0921 ( PULPROG zgfhigqn.2 TD 131072 SOLVENT D20 NS 16 DS 4 SWH 89285.711 Hz FIDRES 1.362392 Hz AQ 0.7340032 sec RG 196.38 DW 5.600 usec DE 6.50 usec TE 298.9 K D1 1.0000000 sec D11 0.0300000 sec D12 0.000200 sec D12 0.000200 sec D11 1.0000000 sec D11 1.0000000 sec D12 0.000200 sec D12 0.000200 sec D12 0.000200 sec D12 0.000200 sec D14 1.0000000 sec D1500 1 SF01 376.4889418 MHz NUC1 19F P1 15.00 usec PLW1 17.7539971 W SF02 400.1616006 MHz NUC2 1H CPDPRG[2 waltz16 PCPD2 90.00 usec PLW2 11.52400017 W PLW12 0.27886000 W
		F2 - Processing parameters SI 65536 SF 376.5265944 MHz WDW EM SSB 0 LB 0.30 Hz GB 0 PC 1.00













