

SUPPORTING INFORMATION

Identification of the first structurally validated covalent ligands of the small GTPase RAB27A

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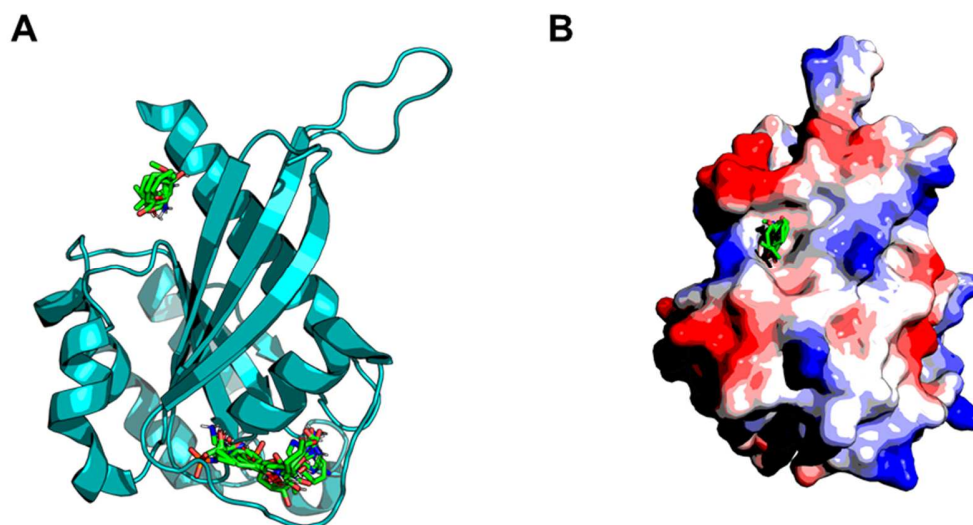


Figure S1. Evaluating Rab27A (PDB: 3BC1, chain A) hotspots using FTMap server.¹ This server identifies binding pockets within a protein surface by evaluating binding energy of molecules with different physicochemical properties. A) and B) demonstrate that organic molecule clusters predominantly occupy the nucleotide binding site and the WF pocket.

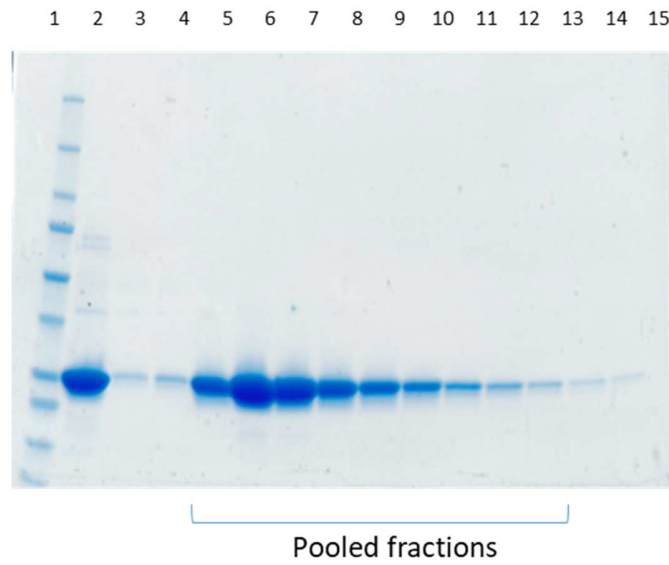


Figure S2. Last purification step for fRab27A-C188. Size-exclusion chromatography (SEC) fractions A10 to B8 were pooled to obtain pure recombinant protein. Gel lanes: 1. ladder; 2. SEC input; SEC fractions: 3. A8; 4. A9; 5. A10; 6. A11; 7. B1; 8. B3; 9. B4; 10. B5; 11. B6; 12. B7; 13. B8; 14. B9; 15. B10.

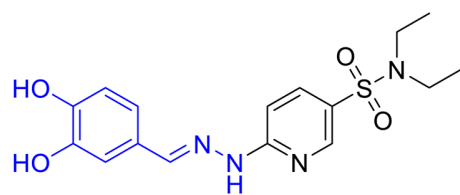
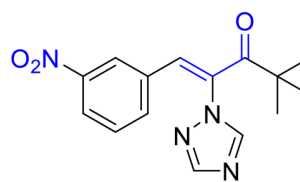
A**BMD-11****B****Nexinhib 20**

Figure S3. Structure of reported Rab27A non-covalent inhibitors: BMD-11² and Nexinhib 20³. Motifs commonly associated with PAINS are highlighted in blue, such as catechols and vinyl ketones.⁴

RAB27A	-----	0
RAB27B	-----	0
RAB24	-----	0
RAB6C	-----	0
RAB6D	-----	0
RAB6A	-----	0
RAB6B	-----	0
RAB17	-----MAQ-----	3
RAB21	-----MAA-----	3
RAB5B	-----MT-----	2
RAB5A	-----MA-----	2
RAB5C	-----MAG-----	3
RAB22A	-----	0
RAB22B(31)	-----	0
RAB20	-----	0
RAB28	-----	0
RAB29	-----	0
RAB32	-----MAGGGAGDPG-----LG	12
RAB38	-----	0
RAB34	-----	0
RAB36	MVIAGASWMLGRAAA--SPTQTPPTTSTIRVARRSRVALVAMVIAAAGSGGPGRAPQLS	58
RAB42	-----	0
RAB39A	-----	0
RAB39B	-----	0
RAB7B	-----	0
RAB7A	-----	0
RAB9A	-----	0
RAB9B	-----	0
RAB23	-----	0
RAB40C	-----	0
RAB40B	-----	0
RAB40A	-----	0
RAB40AL	-----	0
RAB33A	-----MAQPILGHGSLQPA----S	15
RAB33B	-----MAEEMES--SLE-A----S	12
RAB30	-----	0
RAB19	-----	0
RAB43	-----	0
RAB18	-----	0
RAB2A	-----	0
RAB2B	-----	0
RAB14	-----	0
RAB4A	-----	0
RAB4B	-----	0
RAB25	-----	0
RAB11A	-----	0
RAB11B	-----	0
RAB12	-----MDPG----AALQRRAGGGGGLGAGSPALS	25
RAB26	-----MSRKKTPKSKGASTPAASTLPTANG----ARPARS--GTALSGPDAPPNG	44
RAB37	-----MTGTPG----AVATRD--GE-----APERS	19
RAB3D	-----M-----	1
RAB3B	-----M-----	1
RAB3A	-----M-----	1
RAB3C	-----MRHEAPMQM-----	9
RAB15	-----	0
RAB13	-----	0
RAB10	-----	0
RAB8A	-----	0
RAB8B	-----	0
RAB35	-----	0
RAB1A	-----	0
RAB1B	-----	0

RAB27A	-----MSDG	4
RAB27B	-----MTDG	4
RAB24	-----MS	2
RAB6C	-----MSAGGDFG	8
RAB6D	-----MSAGGDFG	8
RAB6A	-----MSTGGDFG	8
RAB6B	-----MSAGGDFG	8
RAB17	-----AHR--TPQPRAAP	14
RAB21	-----AGG--GGGAAAA	14
RAB5B	-----SRSTARPNQPPQA	15
RAB5A	-----SRGATRPNGPNTG	15
RAB5C	-----RGGAARPNPAAAG	16
RAB22A	-----	0
RAB22B(31)	-----	0
RAB20	-----	0
RAB28	-----MSDSEE	7
RAB29	-----MG	2
RAB32	-----AAAAPAPE	20
RAB38	-----MQAP	4
RAB34	-----MNILAPVRRDRVLAELPQCLRKEAALHGKDFHPRVTCACQEHTGT	47
RAB36	QPSLDCGRMRSSLTPLGPPVSRDRVIASF PKWYTP EACLQLREHFHGQVSAACQRRNTGT	118
RAB42	-----MEAE	4
RAB39A	-----MET	3
RAB39B	-----MEA	3
RAB7B	-----MNP	3
RAB7A	-----MTS	3
RAB9A	-----MA	2
RAB9B	-----MS	2
RAB23	-----MLEE	4
RAB40C	-----MG-----SQGSPVK	9
RAB40B	-----MS-----ALGSPVR	9
RAB40A	-----MS-----APGSPDQ	9
RAB40AL	-----MS-----APGSPDQ	9
RAB33A	AAGLASLEL-----DSSLDQY	31
RAB33B	FSSSGAVSG-----ASGFLPP	28
RAB30	-----MSME	4
RAB19	----MHFSS-----SARAAD	12
RAB43	---MAGPGP-----GPGDPDE	13
RAB18	-----MDE	3
RAB2A	-----M	1
RAB2B	-----M	1
RAB14	-----MATAPY	6
RAB4A	-----M-----SQTAMSE	8
RAB4B	-----MAE	3
RAB25	-----MGNGTEE	7
RAB11A	-----MGTRDD	6
RAB11B	-----MGTRDD	6
RAB12	--GGQGR-----RRKQPPR	37
RAB26	--PLQPGRP-----SLGGGVD	58
RAB37	-----P-----PCSP	24
RAB3D	ASAGDTQAG-----PRDAADQ	17
RAB3B	ASVTDGKTG-----VKDASDQ	17
RAB3A	ASATDSRYG-----QKSSDQ	17
RAB3C	ASAQDARYG-----QKDSSDQ	25
RAB15	-----MAK	3
RAB13	-----MAK	3
RAB10	-----MAKK	4
RAB8A	-----MAK	3
RAB8B	-----MAK	3
RAB35	-----MAR	3
RAB1A	-----MSSMNP	6
RAB1B	-----MNP	3

RAB27A	DYDYLKFLALGDSGVGKTSVLVYQYTDGKFN-S----	KFITTVGIDFREKRVVYRASGPD	59
RAB27B	DYDYLKLLALGDSGVGKTTFLRYTDNKNF-P----	KFITTVGIDFREKRVVYNAQGP	59
RAB24	GQRVDVKKVMLGKEYVGKTSLVERYVHDRFLVG----	PYQNTIGAAFAVKVMCV-----	52
RAB6C	NPLRKFKLVFLGEQSVAKTSLITRFRYDSFD-N----	TYQAIIGIDFLSKTMYL-----	57
RAB6D	NPLRKFKLVFLGEQSVAKTSLITRFRYDSFD-N----	TYQAIIGIDFLSKTMYL-----	57
RAB6A	NPLRKFKLVFLGEQSVGKTSLITRFMYDSFD-N----	TYQATIGIDFLSKTMYL-----	57
RAB6B	NPLRKFKLVFLGEQSVGKTSLITRFMYDSFD-N----	TYQATIGIDFLSKTMYL-----	57
RAB17	SQPRVFKLVLLGSGSVGKSSLALRYVKNDFK-S----	ILPTVGCAFFTQVVDV-----	62
RAB21	GRAYSFKVVLLGEGCVGKTSVLRLYCENKFN-D----	KHITTLQASFLTCKLNI-----	63
RAB5B	SKICQFKLVLLGESAVGKSSLVLRVFKGQFH-E----	YQESTIGAAFLTQSVCL-----	64
RAB5A	NKICQFKLVLLGESAVGKSSLVLRVFKGQFH-E----	FQESTIGAAFLTQTVCL-----	64
RAB5C	NKICQFKLVLLGESAVGKSSLVLRVFKGQFH-E----	YQESTIGAAFLTQTVCL-----	65
RAB22A	MALRELKVCLLGDTGVGKSSIWRVFVEDSFD-P----	NINPTIGASFMTKTQVY-----	49
RAB22B(31)	MAIRELKVCLLGDTGVGKSSIWRVFVEDSFD-P----	NINPTIGASFMTKTQVY-----	49
RAB20	MRKPDVSKIVLLGDMNVGKTSLLQRYMERRFP-----	DTVSTVGGAFLKQWRS-----	48
RAB28	SQDRQLKIVVLGDGASGKTSLLTTCFAQETFG-K----	QYKQTIGLDFLRRITL-----	56
RAB29	SRDHLFKVLVVGDAAVGKTSLVQYRSQDSFS-K----	HYKSTVGVDFAKVLQW-----	51
RAB32	TREHLFKVLVIGELGVGKTSIIKRYVHQLFS-Q----	HYRATIGVDFALKVLNW-----	69
RAB38	HKEHLYKLLVIGDLGVGKTSIIKRYVHQNFS-S----	HYRATIGVDFALKVLHW-----	53
RAB34	VGFKISKVIVVGDLSVGKTCILNRFCKDTFD-K----	NYKATIGVDFEMERFEV-----	96
RAB36	VGLKLSKVVVVDLYVGKTSIIHRFCNVFD-R----	DYKATIGVDFEIERFEI-----	167
RAB42	GCRYQFRVALLGDAAVGKTSLLRSYVAGAPGAPEPEPEPTVGAECYRRALQL-----		58
RAB39A	IWIYQFRLIVIGDSTVGKSCLLHRFTQGRFPG-LRSPACDPTVGVDFFSRLLLEI-----		56
RAB39B	IWLQYFRLIVIGDSTVGKSCLLHRFTQGRFAQ-V----	SDPTVGVDFFSRLLVEI-----	52
RAB7B	RKKVDLKLIIVGAIGVGKTSLLHQYVHKTFY-E----	EYQTTLGASILSKIIIL-----	52
RAB7A	RKKVLLKKVILGDSGVGKTSMLNQYVNNKFS-N----	QYKATIGADFLTKEVMV-----	52
RAB9A	GKSSLFKVLLGDDGVGKSSLMNRYVTNKFD-T----	QLFHTIGVEFLNKDLEV-----	51
RAB9B	GKSSLKVLVLLGDDGVGKSSLMNRYVTNKFD-S----	QAFHTIGVEFLNRDLEV-----	51
RAB23	DMEVAIKMVVVGNGAVGKSSMIQRYCKGIFT-K----	DYKKTIGVDFLERQIQV-----	53
RAB40C	SYDYLLKFLLVGDSVVGKGEILESQDGAAE-S----	PYAYSNGIDYKTTTILL-----	58
RAB40B	AYDFLLKFLLVGDSVVGKGEILASLQDGAAE-S----	PYGHYPAGIDYKTTTILL-----	58
RAB40A	AYDFLLKFLLVGDRDVGKSEILESQDGAAE-S----	PYSHLGGIDYKTTTILL-----	58
RAB40AL	AYDFLLKFLLVGDRDVGKSEILESQDGTAE-S----	PYSHLGGIDYKTTTILL-----	58
RAB33A	VQIRIFKIIIVIGDSNVGKTCLTRFCGGTFP-D----	KTEATIGVDFREKTVEI-----	80
RAB33B	ARSRIKFIIIVIGDSNVGKTCLTRYFCAGRFP-D----	RTEATIGVDFRERAVEI-----	77
RAB30	DYDFLFKIVLIGNAGVGKTCVLRFRFTQGLFP-P----	GQGATIGVDFMIKTVEI-----	53
RAB19	NFDYLFKIIIVIGDSNVGKTCVQVHFQSGVYT-E----	TQNTIGVDFTVRSLDI-----	61
RAB43	QYDFLFKLVVGDASVGKTCVQVRFKTAFA-S-E----	RQGSTIGVDFMTKLTLEI-----	62
RAB18	DVLTTLKILIIIGESGVGKSSLLRFTDDTFD-P----	ELAATIGVDFKVKTISV-----	52
RAB2A	AYAYLFKYIIIGDTGVGKSCLLLQFTDKRFQ-P----	VHDLTIGVEFGARMITI-----	50
RAB2B	TYAYLFKYIIIGDTGVGKSCLLLQFTDKRFQ-P----	VHDLTIGVEFGARMVNI-----	50
RAB14	NYSYIFKYIIIGDMGVGKSCLLHQFTEKKFM-A----	DCPHTIGVEFGTRIIEV-----	55
RAB4A	TYDFLFKFLVIGNAGTGKSCLLHQFIEKKFK-D----	DSNHTIGVEFGSKIINV-----	57
RAB4B	TYDFLFKFLVIGSAGTGKSCLLHQFIENKFK-Q----	DSNHTIGVEFGSRVNV-----	52
RAB25	DYNFVFKVVLIGESGVGKTNLLSRFTRNEFS-H----	DSRTTIGVEFSTRVML-----	56
RAB11A	EYDYLFKVVLLIGDSGVGKSNLLSRFTRNEFN-L----	ESKSTIGVEFATRSIQV-----	55
RAB11B	EYDYLFKVVLLIGDSGVGKSNLLSRFTRNEFN-L----	ESKSTIGVEFATRSIQV-----	55
RAB12	PADFKLQVIIIGSRGVGKTSLMRFTDDTFC-E----	ACKSTVGVDFKIKTVEL-----	86
RAB26	FYDVAFKVMLVGDSGVGKTCVLRFRKDGAFLAG----	TFISTVGIDFRNKVLDV-----	108
RAB37	SYDLTGKVMLLGDTGVGKTCFLIQFKDGAFLSG----	TFIATVGIDFRNKVVTV-----	74
RAB3D	NFDYMFKLLLIIGNSSVGKTSFLFRYADDSFT-P----	AFVSTVGIDFKVKTYYR-----	66
RAB3B	NFDYMFKLLLIIGNSSVGKTSFLFRYADDTFT-P----	AFVSTVGIDFKVKTYYR-----	66
RAB3A	NFDYMFKLLLIIGNSSVGKTSFLFRYADDSFT-P----	AFVSTVGIDFKVKTYYR-----	66
RAB3C	NFDYMFKLLLIIGNSSVGKTSFLFRYADDSFT-S----	AFVSTVGIDFKVKTVFK-----	74
RAB15	QYDVLFRLLLIIGDSGVGKTCVLRFRSDDAFN-T----	SHISTIGVDFKMKTIEV-----	52
RAB13	AYDHLFKLLLIIGDSGVGKTCVLRFRFAEDNFN-N----	TYISTIGIDFKIRTVDI-----	52
RAB10	TYDLLFKLLLIIGDSGVGKTCVLRFRSDDAFN-T----	TFISTIGIDFKIKTVEL-----	53
RAB8A	TYDYLFKLLLIIGDSGVGKTCVLRFRSDDAFN-S----	TFISTIGIDFKIRTIEL-----	52
RAB8B	TYDYLFKLLLIIGDSGVGKTCVLRFRSDDAFN-T----	TFISTIGIDFKIRTIEL-----	52
RAB35	DYDHLFKLLLIIGDSGVGKSSLLRFADNTFS-G----	SYITTIGVDFKIRTVEI-----	52
RAB1A	EYDYLFKLLLIIGDSGVGKSCVLRFRADDTYT-E----	SYISTIGVDFKIRTIEL-----	55
RAB1B	EYDYLFKLLLIIGDSGVGKSCVLRFRADDTYT-E----	SYISTIGVDFKIRTIEL-----	52

RAB27A	GATGRGQRIHLQLWDTAGQERFR-SLTTAFFRDAMGFLLLFDLTNEQSFLNVRNWISQLQ	118
RAB27B	GSSGKAFKVHLQLWDTAGQERFR-SLTTAFFRDAMGFLLMFDLTSQQSFLNVRNWMSQLQ	118
RAB24	----GDRTVTLG IWDTAGSERYE-AMSR IYYRGAKAAIVCYDLT DSSSFERAKFWVKELR	107
RAB6C	----EDGTIGRLRLWDTAGQERLR-SLIPRYIRDSAAAVVVYDITNVNSFQQTTKWIDDVR	112
RAB6D	----EDGTIGRLRLWDTAGQERLR-SLIPRYIRDSAAAVVVYDITNVNSFQQTTKWIDDVR	112
RAB6A	----EDRTVRLQLWDTAGQERFR-SLIPSYIRDSTVAVVVYDITNVNSFQQTTKWIDDVR	112
RAB6B	----EDRTVRLQLWDTAGQERFR-SLIPSYIRDSTVAVVVYDITNLNSFQQTTSKWIDDVR	112
RAB17	----GATSLKLEIWDTAGQEKYH-SVCHLYFRGANAAALLVYDITRKDSFLKAQQWLKDLE	117
RAB21	----GGKRVNLAIWDTAGQERFH-ALGPIYYRDSNGAILVYDITDEDSFQKVKNWVKELR	118
RAB5B	----DDTTVKFEIWDTAGQERYH-SLAPMYRGAQAAIVVYDITNQETFARAKTWVKELQ	119
RAB5A	----DDTTVKFEIWDTAGQERYH-SLAPMYRGAQAAIVVYDITNEESFARAKNWVKELQ	119
RAB5C	----DDTTVKFEIWDTAGQERYH-SLAPMYRGAQAAIVVYDITNTDTFARAKNWVKELQ	120
RAB22A	----QNELHKFLIWDTAGQERFR-ALAPMYRGSAAAIIVYDITKEETFSTLKNWVKELR	104
RAB22B(31)	----GNELHKFLIWDTAGQERFH-SLAPMYRGSAAAVIVYDITKQDSFYTLKKWVKELK	104
RAB20	-----YNISIWDTAGREQFH-GLGSMYCRGAAAIILTVDNHRQSLVELEDRFLGLT	99
RAB28	---PGNLNVTLQIWDIGGQTIGG-KMLDKYIYGAQGVLLVYDITNYQSFENLEDWYTVVK	112
RAB29	---SDYEIVRLQLWDIAGQERFT-SMTRLYYRDASACVIMFDVTNATTFSNSQRWKQDLD	107
RAB32	---DSRTLVRQLWDIAGQERFG-NMTRVYYKEAVGAFVVDISRSTFEAVLKWKSDLD	125
RAB38	---DPETVVRQLWDIAGQERFG-NMTRVYYREAMGAFIVFDVTRPATFEAVAKWKNLD	109
RAB34	----LGIPFSLQLWDTAGQERFK-CIASTYYRGAQAIIVFNLDVASLEHTKQWLADAL	151
RAB36	----AGIPVSLQIWDTAGQEKFK-CIASAYYRGAQVIITAFDLTDVQTLLEHTRQWLEDAL	222
RAB42	---RAGPRVKLQLWDTAGHERFR-CITRSFYRNVVGVLLVFDVTNRKSFEHIQDWHQEV	114
RAB39A	---EPGKRIKLQLWDTAGQERFR-SITRSYYRNSVGGFLVFDITNRRSFEHVKDWEAK	112
RAB39B	---EPGKRIKLQLWDTAGQERFR-SITRAYRNSVGGLLFDITNRRSFQNVHEWLEETK	108
RAB7B	----GD TTLKLQIWD TGQERFR-SMVSTFYKSGDGCILAFDVTDL ESFEALDIWRGDVL	107
RAB7A	----DDR LVTMQIWD TAGQERFQ-SLGVAFYRGADCCVLVFDVTAPNTFKTLD SWRDEFL	107
RAB9A	----DGHFVTMQIWD TAGQERFR-SLRTPFYRGSDCCLLTF SVDDSQSFQNL SNWKKEFI	106
RAB9B	----DGRFVTLQIWD TAGQERFK-SLRTPFYRGADCCLLTF SVDDRQSFENLGNWKKEFI	106
RAB23	----NDEDVRLMLWDTAGQEEFD-AITKAYYRGAQACVLVFSTTDRESFEAVSSWREKVV	108
RAB40C	----DGRRVKLELWDTSGQGRFC-TIFRSYSRGAQGILLVYDITNRWSFDGIDRWIKEID	113
RAB40B	----DGRRVKLELWDTSGQGRFC-TIFRSYSRGAQGVILVYDIANRWSFDGIDRWIKEID	113
RAB40A	----DGQRVKLELWDTSGQGRFC-TIFRSYSRGAQGVILVYDIANRWSFEGMDRWIKKIE	113
RAB40AL	----DGQRVKLELWDTSGQGRFC-TIFRSYSRGAQGVILVYDIANRWSFEGMDRWIKKIE	113
RAB33A	----EGEKIKVQVWD TAGQERFRKSMVEHYRNHVAVVFVYDVTKMTSFTNLKMWIQECN	136
RAB33B	----DGERIKIQLWDTAGQERFRKSMVQHYRNHVAVVFYDMTNMASFHSPLPSWIEECK	133
RAB30	----NGEKVKLQIWD TAGQERFR-SITQSYYSANALILTYDITCEESFRCLPEWLREIE	108
RAB19	----DGKKVKMQVWD TAGQERFR-TITQSYYSAHAAIIAYDLTRRSTFESIPHWIHEIE	116
RAB43	----QGKRVKLQIWD TAGQERFR-TITQSYYSANGAILAYDITKRSSFLSVPHWIEDVR	117
RAB18	----DGNKAKLAIWD TAGQERFR-TLTPSYRGAQGVILVYDVTRRDTFVKLDNWLNELE	107
RAB2A	----DGKQIKLQIWD TAGQESFR-SITRSYYRGAAGALLVYDITRRDTFNHLT TWLEDAR	105
RAB2B	----DGKQIKLQIWD TAGQESFR-SITRSYYRGAAGALLVYDITRRET FNHLT SWLEDAR	105
RAB14	----SGQKIKLQIWD TAGQERFR-AVTRSYRGAAGALMVYDITRRSTYNHLSSWLT DAR	110
RAB4A	----GGKYVKLQIWD TAGQERFR-SVTRSYRGAAGALLVYDITSRETYNALT NWLT DAR	112
RAB4B	----GGKTVKLQIWD TAGQERFR-SVTRSYRGAAGALLVYDITSRETYNSLAAWLT DAR	107
RAB25	----GTA AVKAQIWD TAGLERYR-AITSAYYRGAVGALLVFDLT KHQTYAVVERWLKELY	111
RAB11A	----DGKTIKAQIWD TAGQERYR-AITSAYYRGAVGALLVYDIAKHLTYENVERWLKELR	110
RAB11B	----DGKTIKAQIWD TAGQERYR-AITSAYYRGAVGALLVYDIAKHLTYENVERWLKELR	110
RAB12	----RGKKIRLQIWD TAGQERFN-SITSAYYRSAGIILVYDITKKETFDDL PKWMMKID	141
RAB26	----DGVKVKLQIWD TAGQERFR-SVTHAYYRDAHALLLYDVTNKASF DNIQAWL TEIH	163
RAB37	----DGVRVKLQIWD TAGQERFR-SVTHAYYRDAQALLLYDITNKSSF DNIRAWL TEIH	129
RAB3D	----HDKRIKLQIWD TAGQERYR-TITTAYYRGAMGFLMYDIANQESFAAVQDWATQIK	121
RAB3B	----HEKRVKLQIWD TAGQERYR-TITTAYYRGAMGFLMYDITNEESFNAVQDWATQIK	121
RAB3A	----NDKRIKLQIWD TAGQERYR-TITTAYYRGAMGFLMYDITNEESFNAVQDWSTQIK	121
RAB3C	----NEKRIKLQIWD TAGQERYR-TITTAYYRGAMGFLMYDITNEESFNAVQDWSTQIK	129
RAB15	----DGIKVRIQIWD TAGQERYQ-TITKQYYRRAQGIFLVYDISSERSYQHIMKWVSDVD	107
RAB13	----EGKKIKLQVWD TAGQERFK-TITTAYYRGAMGIILVYDITDEKSFENIQNMKSIK	107
RAB10	----QGKKIKLQIWD TAGQERFH-TITTSYYRGAMGIMLVYDITNGKSFENISKWLRNID	108
RAB8A	----DGKRIKLQIWD TAGQERFR-TITTAYYRGAMGIMLVYDITNEKSF DNIKNWIRNIE	107
RAB8B	----DGKKIKLQIWD TAGQERFR-TITTAYYRGAMGIMLVYDITNEKSF DNIKNWIRNIE	107
RAB35	----NGEKVKLQIWD TAGQERFR-TITSTYYRGTHGVIVVYDV TSAESFNVVKRWLHEIN	107
RAB1A	----DGKTIKLQIWD TAGQERFR-TITSSYYRGAHGIIVVYDVT DQESFNVKQWLQEID	110
RAB1B	----DGKTIKLQIWD TAGQERFR-TITSSYYRGAHGIIVVYDVT DQESYANVKQWLQEID	107

RAB27A	MHAYC---ENPDIVLCGNKSDLED---QRVV-KEEE-----	147
RAB27B	ANAYC---ENPDIVLIGNKADLPD---QREV-NERQ-----	147
RAB24	SLEEG-----CQIYLCGTSKDLLEEDRRRRRV-DFHD-----	138
RAB6C	TERGS----DVIITLVGNRTDLAD----KRQV-SVEE-----	140
RAB6D	TEGGS----DVIITLVGNKTDLAD----KRQV-SIEE-----	140
RAB6A	TERGS----DVIIMLVGNKTDLAD----KRQV-SIEE-----	140
RAB6B	TERGS----DVIIMLVGNKTDLAD----KRQI-TIEE-----	140
RAB17	EELHP---GEVLVMLVGNKTDLSQ---EREV-TFQE-----	146
RAB21	KMLGN---EICLCIVGNKIDLEK---ERHV-SIQE-----	146
RAB5B	RQASP---SIVIALAGNKADLAN---KRMV-EYEE-----	147
RAB5A	RQASP---NIVIALSGNKADLAN---KRAV-DFQE-----	147
RAB5C	RQASP---NIVIALAGNKADLAS---KRAV-EFQE-----	148
RAB22A	QHGGP---NIVVAIAGNKCDLID---VREV-MERD-----	132
RAB22B(31)	EHGPE---NIVMAIAGNKCDLSD---IREV-PLKD-----	132
RAB20	DTASK---DCLFAIVGNKVDLTE---EGAL-AGQEKEECSPNMDAGDRVSPRAPKQVQ	150
RAB28	KVSEE-SETQPLVALVGNKIDLEH---MRTI-KPEK-----	143
RAB29	SKLTLPNGEPVPCLLANKCDLSP---WAV--SRDQ-----	138
RAB32	SKVHLPNGSPIPAVLLANKCDQNK---DSSQ-SPSQ-----	157
RAB38	SKLSLPNGKPVSVVLLANKCDQKG---DVLMNGLK-----	142
RAB34	KENDP---SSVLLFLVGSKKDLST---PAQY-ALMEKD-----	182
RAB36	RENEA---GSCFIFLVGTTKDLLS---GAAC-EQAEAD-----	253
RAB42	ATQGP---DKVIFLLVGHKSDLQS---TRCV-SAQE-----	143
RAB39A	MYVQP---FRIVFLLVGHKCDLAS---QRQV-TREE-----	141
RAB39B	VHVQP---YQIVFVLVGHKCDLDT---QRQV-TRHE-----	137
RAB7B	AKIVP-MEQSYPMVLLGNKIDLA---DRKV-PQEV-----	137
RAB7A	IQASPRDPENFPFVVLGNKIDLE---NRQV-ATKR-----	138
RAB9A	YYADVKEPESFPFVILGNKIDIS---ERQV-STEE-----	137
RAB9B	YYADVKDPEHFPFVVLGNKVDKE---DRQV-TTEE-----	137
RAB23	AEVG-----DIPTVLVQNKIDLLD---DSCI-KNEE-----	135
RAB40C	EHAP-----GVPRIILVGNRLHLAF---KRQV-PTEQ-----	140
RAB40B	EHAP-----GVPKILVGNRLHLAF---KRQV-PTEQ-----	140
RAB40A	EHAP-----GVPKILVGNRLHLAF---KRQV-PREQ-----	140
RAB40AL	EHAP-----GVPKILVGNRLHLAF---KRQV-PREQ-----	140
RAB33A	GHAVP---PLVPKVLVGNKCDLRE---QIQV-PSNL-----	165
RAB33B	QHLLA---NDIPRIILVGNKCDLRS---AIQV-PTDL-----	162
RAB30	QYASN---KVITVLVGNKIDLAE---RREV-SQQR-----	136
RAB19	KYGAA---NVVIMLIGNKCDLWE---KRHV-LFED-----	144
RAB43	KYAGS---NIVQLLIGNKSDLSE---LREV-SLAE-----	145
RAB18	TYCTR---NDIVNMLVGNKIDKE---NREV-DRNE-----	135
RAB2A	QHSNS---NMVIMLIGNKSDLES---RREV-KKEE-----	133
RAB2B	QHSSS---NMVIMLIGNKSDLES---RRDV-KREE-----	133
RAB14	NLTNP---NTVILIGNKADLEA---QRDV-TYEE-----	138
RAB4A	MLASQ---NIVILCGNKKDLDA---DREV-TFLE-----	140
RAB4B	TLASP---NIVILCGNKKDLDP---EREV-TFLE-----	135
RAB25	DHAEA---TIVVMLVGNKSDLSQ---AREV-PTDE-----	139
RAB11A	DHADS---NIVIMLVGNKSDLRH---LRAV-PTDE-----	138
RAB11B	DHADS---NIVIMLVGNKSDLRH---LRAV-PTDE-----	138
RAB12	KYASE---DAELLLVGNKLDCE---DREI-TRQQ-----	169
RAB26	EYAQH---DVALMLLGNKVDSAH---ERVV-KRED-----	191
RAB37	EYAGR---DVVIMLLGNKADMSS---ERVI-RSED-----	157
RAB3D	TYSWD---NAQVILVGNKCDLED---ERVV-PAED-----	149
RAB3B	TYSWD---NAQVILVGNKCDMEE---ERVV-PTEK-----	149
RAB3A	TYSWD---NAQVLLVGNKCDMED---ERVV-SSER-----	149
RAB3C	TYSWD---NAQVILVGNKCDMED---ERVI-STER-----	157
RAB15	EYAPE---GVQKILIGNKADEEQ---KRQV-GREQ-----	135
RAB13	ENASA---GVERLLLGNKCDMEA---KRKV-QKEQ-----	135
RAB10	EHANE---DVERMLLGNKCDMDD---KRVV-PKGK-----	136
RAB8A	EHASA---DVEKMILGNKCDVND---KRQV-SKER-----	135
RAB8B	EHASS---DVERMILGNKCDMND---KRQV-SKER-----	135
RAB35	QNC-D---DVCRIILVGNKNDDEPE---RKVV-ETED-----	134
RAB1A	RYASE---NVNKLLVGNKCDLTT---KKVV-DYTT-----	138
RAB1B	RYASE---NVNKLLVGNKSDLTT---KKVV-DNTT-----	135

RAB27A	---AIALAEK-Y-----	GI-PYFETSAAN---	GTNISQAIEMLLDLIMKRMER	187
RAB27B	---ARELADK-Y-----	GI-PYFETSAAT---	GQNVEKAVETLLDLIMKRMEQ	187
RAB24	---VQDYADN-I-----	KA-QLFETSSKT---	GQSVDELQKVAEDYVSVAAF	178
RAB6C	---GERKAKG-L-----	NV-TFIETRAKA---	GYNVKQLFRRVAAALPGMEST	180
RAB6D	---GERKAKG-L-----	NV-TFIETRAKA---	GYNVKQLFRRVAAALPGMEST	180
RAB6A	---GERKAKE-L-----	NV-MFIETSAKA---	GYNVKQLFRRVAAALPGMEST	180
RAB6B	---GEQRAKE-L-----	SV-MFIETSAKT---	GYNVKQLFRRVASALPGMENV	180
RAB17	---GKEFADS-Q-----	KL-LFMETSAKL---	NHQVSEVFNTVAQELLQRSDE	186
RAB21	---AESYAES-V-----	GA-KHYHTSAKQ---	NKGIEELFLDLCKRMIETAQV	186
RAB5B	---AQAYADD-N-----	SL-LFMETSAKT---	AMNVNDLFLAIKKLPKSEPQ	187
RAB5A	---AQSYADD-N-----	SL-LFMETSAKT---	SMNVNEIFMAIAKKLPKNEPQ	187
RAB5C	---AQAYADD-N-----	SL-LFMETSAKT---	AMNVNEIFMAIAKKLPKNEPQ	188
RAB22A	---AKDYADS-I-----	HA-IFVETSAKN---	AININELFIEISRRIPSTDAN	172
RAB22B(31)	---AKEYAES-I-----	GA-IVVETSAKN---	AINIEELFQGISRQIPPLDPH	172
RAB20	LEDAAVALYKKILKYKMLDEQDVPAAEQMCFETSAKT---	GYNVDLLFETLFDLVVPMILQ	207	
RAB28	---HLRFCQE-N-----	GF-SSHVFSAKT---	GDSVFLCFQKVAEEILGIKLN	183
RAB29	---IDRFSKE-N-----	GFTGWTETSVKE---	NKNINEAMRVLIEKMMRNSTE	179
RAB32	---VDQFCKE-H-----	GFAGWFETSAKD---	NINIEEAARFLVEKILVNHQS	198
RAB38	---MDQFCKE-H-----	GFVGWFETSAKE---	NINIDEASRCLVKHILANEC	183
RAB34	---ALQVAQE-M-----	KA-EYWAVSSLT---	GENVREFFFRVAALTFEANVL	222
RAB36	---AVHLARE-M-----	QA-EYWSVSAKT---	GENVKAFFSRVAALAFEQSVL	293
RAB42	---AEELAAS-L-----	GM-AFVETSVKN---	NCNVDLAFDTLADAIQQALQQ	183
RAB39A	---AEKLSAD-C-----	GM-KYIETSAKD---	ATNVEESFTILTRDIYELIKK	181
RAB39B	---AEKLAAS-Y-----	GM-KYIETSARD---	AINVEKAFTDLTRDIYELVKR	177
RAB7B	---AQGWCRE-K-----	DIPYFEVSAKN---	DINVVQAFEMLASRALSRYQS	177
RAB7A	---AQAWCYS-K-----	NNIPYFETSAKE---	AINVEQAFQTIARNALKQETE	179
RAB9A	---AQAWCRD-N-----	GDYPYFETSAKD---	ATNVAAAFEEAVRRVLATEDR	178
RAB9B	---AQTWCME-N-----	GDYPYLETSAKD---	DTNVTVAFEEAVRQVLAVEEQ	178
RAB23	---AEALAKR-L-----	KL-RFYRTSVKE---	DLNVNEVFKYLAEKYLQKLKQ	175
RAB40C	---ARAYAER-L-----	CM-TFFEVSPLC---	NFNVTVAFEEAVRQVLAVEEQ	180
RAB40B	---AQAYAER-L-----	GV-TFFEVSPLC---	NFNVTVAFEEAVRQVLAVEEQ	180
RAB40A	---AQAYAER-L-----	GV-TFFEVSPLC---	NFNVTVAFEEAVRQVLAVEEQ	180
RAB40AL	---AQAYAER-L-----	GV-TFFEVSPLC---	NFNVTVAFEEAVRQVLAVEEQ	180
RAB33A	---ALKFADA-H-----	NM-LLFETSAKDPKESQNVESIFMCLACRLKAQKSL	208	
RAB33B	---AQKFADT-H-----	SM-PLFETSAKNPNDNDHVEAIFMTLAHKLKSHKPL	205	
RAB30	---AEFSEA-Q-----	DM-YYLETSAKE---	SDNVEKLFLDLACRLISEARQ	176
RAB19	---ACTLAKE-Y-----	GLLAVLETSAKE---	SKNIEEVFVLMAKELIARNSL	185
RAB43	---AQSLAEH-Y-----	DILCAIETSAKD---	SSNVEEAFLRVATELIMRHGG	186
RAB18	---GLKFARK-H-----	SM-LFIEASAKT---	CDGVQCAFEELVEKIIQTPGL	175
RAB2A	---GEAFARE-H-----	GL-IFMETSAKT---	ASNVEEAFINTAKEIYEKIQE	173
RAB2B	---GEAFARE-H-----	GL-IFMETSAKT---	ACNVEEAFINTAKEIYRKIQE	173
RAB14	---AKQFAEE-N-----	GL-LFLEASAKT---	GENVEDAFLEAAKKIYQNIQD	178
RAB4A	---ASRFAQE-N-----	EL-MFLETSAKT---	GENVEEAFVQCARKILNKIES	180
RAB4B	---ASRFAQE-N-----	EL-MFLETSAKT---	GENVEEAFVQCARKILNKIES	175
RAB25	---ARMFAEN-N-----	GL-LFLETSAKD---	STNVELAFETVLKEIFAKVSK	179
RAB11A	---ARAFAEK-N-----	GL-SFIETSALD---	STNVEEAFQITLTIYRIVSQ	178
RAB11B	---ARAFAEK-N-----	NL-SFIETSALD---	STNVEEAFKNILTEIYRIVSQ	178
RAB12	---GEKFQQIT-----	GM-RFCEASAKD---	NFNVDLAFETVLKEIFAKVSK	210
RAB26	---GEKLAKY-Y-----	GL-PFMETSAKT---	GLNVDLAFTAIKELKQSRMK	231
RAB37	---GETLARE-Y-----	GV-PFLETSAKT---	GMNVELAFLAIAKELKYRAGH	197
RAB3D	---GRRLLAD-L-----	GF-EFFEASAKE---	NINVKQVFERLVDVICEKMNE	189
RAB3B	---GQLLAQ-L-----	GF-DFFEASAKE---	NISVRQAFERLVDICDKMSD	189
RAB3A	---GRQLADH-L-----	GF-EFFEASAKD---	NINVKQVFERLVDVICEKMSE	189
RAB3C	---GQHLGEQ-L-----	GF-EFFETSAKD---	NINVKQVFERLVDIICDKMSE	197
RAB15	---GQQLAKE-Y-----	GM-DFYETSACT---	NLNIKESFTRLTELVLQAHRK	175
RAB13	---ADKLARE-H-----	GI-RFFETSAKS---	SMNVDEAFSSSLARDILLKSGG	175
RAB10	---GEQIARE-H-----	GI-RFFETSAKA---	NINIEKAFTLAEDILRKTPV	176
RAB8A	---GEKLALD-Y-----	GI-KFMETSAKA---	NINVENAFTLARDIAKAKMDK	175
RAB8B	---GEKLALD-Y-----	GI-KFLETSAKS---	SANVEEAFFTLARDIMTKLNR	175
RAB35	---AYKFAGQ-M-----	GI-QLFETSAKE---	NVNVEEMFNCITELVLRRAKD	174
RAB1A	---AKEFADS-L-----	GI-PFLETSAKN---	ATNVEQSFMTMAAEIKKRMGP	178
RAB1B	---AKEFADS-L-----	GI-PFLETSAKN---	ATNVEQAFMTMAAEIKKRMGP	175

RAB27A	C	VDKSWIPEGV-VRSNGHA-----S-TD-----QLSEEKEKGA	218
RAB27B	C	VEKTQIPDTV-NGGNSGN-----LDGEKPPPEKK	215
RAB24		QVMTE-----DKGV---DLS-----QK-ANPYFYS	199
RAB6C		QDGSRE-----DMSDIKLE-----KPQEQTVSEG	204
RAB6D		QDGSRE-----DMSDIKLE-----KPQEQTVSEG	204
RAB6A		QDRSRE-----DMIDIKLE-----KPQEQPVSEG	204
RAB6B		QEKSKE-----GMIDIKLD-----KPQEPPASEG	204
RAB17		EGQA-----LRGDAVALN-----K-GPARQAK	208
RAB21		DERAKGNGSSQ-----PGTARRGVQIIDDE-----PQ-AQTSGGG	220
RAB5B		NLG---G---A-----A-GRSRGV---DLH-----EQ-SQQNKSQ	211
RAB5A		NPG---A---N-----S-ARGRGV---DLT-----EP-TQPTRNQ	211
RAB5C		NAT---G---A-----P-GRNRGV---DLQ-----EN-NPASRSQ	212
RAB22A		LPS---G-----GKGF---KLR-----RQ-PSEPKRS	192
RAB22B(31)		ENG---N-----NGTI---KVE-----KP-TMQASRR	192
RAB20		QRAERPSHTVD-----ISS-----HKPPKRTMSG	231
RAB28		KAEIEQ-SQRVVKA-----D-IVNYNQEPMS---RTVNPPRS	215
RAB29		DIMSLSTQG-----D-YINLQTK-----S---SSWS	201
RAB32		FPNE-ENDV-----D-KIKLDQE-----TLRAENKSQ	223
RAB38		LMESIEPDV-----V-KPHLT-----STKVASCSSG	207
RAB34		AELEKSGARRI-----GD-VVRINSDDSN-LYLTASKKKP	255
RAB36		QDLERQSSARL-----QV-----GNGD-LIQMEGSPPE-TQESKRPSL	330
RAB42		GDIKLEEGWGGVRLI-----H-KTQIPRS---P-SRKQHSQP	215
RAB39A		GEICIQDQWEGVKSG-----F-VPNTVHS---SEEAVKPRKE	214
RAB39B		GEITIQEGWEGVKSG-----F-VPNVVHS---SEEVVKSERR	210
RAB7B		ILE-N---HLTE-----SIKL-----S-P-DQSRSR	197
RAB7A		VELYNEFPEPI-----KLDK-----NDR-AKASAE	203
RAB9A		SDHLI---QTD-----TVNL-----HRK-PKPSSS	199
RAB9B		LEHCM---LGH-----TIDL-----NSG-SKAGSS	199
RAB23		QIAEDPELTHSSSNKIGVFNTSGGSHSGQNSGTLNGGD-VINLRPN-KQRTKKNRNPFSS	233
RAB40C		EKIWRPNRV-----FSLQDL	195
RAB40B		DRLWRPSKV-----LSLQDL	195
RAB40A		NWLGRPSKV-----LSLQDL	195
RAB40AL		NWLGRPSKV-----LSLQDL	195
RAB33A		LYRDAERQQGK---V-----Q-KLEF-----PQEANSKTS	234
RAB33B		MLSQPPDN-G-----IIL-----KPEPKPAMT	226
RAB30		NTLVNNV-----SSPL-----PGEKGSISYL	197
RAB19		HLYGESALN-G---LPLD-----S-SPVL-----MAQGPSEKTH	214
RAB43		PLFSEKSPD-H---IQLN-----S-KDI-----GEGWG	209
RAB18		WESENQNK---G---VKLS-----H-REE-----GQG-GGACGG	201
RAB2A		GVFDINNEANGIKIGPQHA-----A-TNATHAG---NQGGQQAGGG	210
RAB2B		GLFDVHNEANGIKIGPQQS-----I-STSVGPSASQRNSRDIGSNSG	214
RAB14		GSLDLNAAESGVQHKPSAP-----Q-GGRL-TS---EPQPQREGCG	214
RAB4A		GELDPERMGSGIQYGDAAL-----R-QLRSPRR---AQAPNAQECG	217
RAB4B		GELDPERMGSGIQYGDASL-----R-QLRQPRS---AQAVAPQPCG	212
RAB25		QRQNSIRTNAITLGSQA-----AG---QEPGPGEKRA	208
RAB11A		KQMSDRRENDMSPSNNVV-----PIHVPPT---TEN---KPKVQ	211
RAB11B		KQIADRAAHDESPGNNVV-----DISVPPT---TDGQKPNKLQ	213
RAB12		DILRNELS---NSI---LSLQ-----P-EPEIPPE---LPP-PRPHVR	242
RAB26		APSEPRFR-----LH-----D-YV-----KR-EGRGAS	252
RAB37		QADEPSFQ-----IR-----D-YV-----ES-QKKRSS	218
RAB3D		SLEPSSSS-GSNGKGPVAVG-----D-AP-----APQPS	216
RAB3B		SLDT-DPSMLGSSKNTRLS-----D-TP-----PLLQQN	216
RAB3A		SLDTADPAVTGAKQGPQLS-----D-QQ-----VPPHQD	217
RAB3C		SLET-DPAITAANKQNTLRLK-----E-TP-----PPPQPN	224
RAB15		ELEGLMRASNELALAELE-----E-EEG-KPE---GP---ANSSKT	209
RAB13		RRSGNGNKPPSTD---LK-----TCD-KKNTNK	199
RAB10		KEPNSENVDISSGGGVT-----GW---KS-----K	198
RAB8A		KLEGNSPQ---GSNQGVKIT-----PD---QK-RSSFRR	203
RAB8B		KMNDNSNA---GAGGPVKIT-----EN---RSK-KTSFFR	203
RAB35		NLAKQQQQ---QQNDVVKLT-----KN-----SKRKKR	199
RAB1A		GATAGGAE---KSNVKIQST-----PV-----KQSGGG	203
RAB1B		GAASGG-E---RPNLKIDST-----PV-----KPAGGG	199

RAB27A	CGC-----	221
RAB27B	CIC-----	218
RAB24	CCHH-----	203
RAB6C	GCSCYSPMSSSTLPQKPPYSFIDCSVNI GLNLFPSLITFCNSSLLPVSWR-----	254
RAB6D	GCSCYSPMSSSTLPQKPPYSFIDCSVNI GLNLFPSLITFCNSSLLPVSWR-----	254
RAB6A	GCSC-----	208
RAB6B	GCSC-----	208
RAB17	CCAH-----	212
RAB21	CCSSG-----	225
RAB5B	CCSN-----	215
RAB5A	CCSN-----	215
RAB5C	CCSN-----	216
RAB22A	CC-----	194
RAB22B(31)	CC-----	194
RAB20	CCA-----	234
RAB28	SMCAVQ-----	221
RAB29	CC-----	203
RAB32	CC-----	225
RAB38	CAKS-----	211
RAB34	TCCP-----	259
RAB36	GCC-----	333
RAB42	CQC-----	218
RAB39A	CFC-----	217
RAB39B	CLC-----	213
RAB7B	CC-----	199
RAB7A	SCSC-----	207
RAB9A	CC-----	201
RAB9B	CC-----	201
RAB23	CSIP-----	237
RAB40C	CCRAIVSCTPVHLIDKLPLPVTIKS---HLKSFSMANGMNAVMMHGRSYSLASGAGGGGS	252
RAB40B	CCRAVVSCTPVHLVDKLPLPIALRS---HLKSFSMANGLNARMMHGGSYSLTTSST---H	249
RAB40A	CCRTIVSCTPVHLVDKLPLPSTLRS---HLKSFSMAKGLNARMMRGLSYSLTTSST---H	249
RAB40AL	CCRTIVSCTPVHLVDKLPLPIALRS---HLKSFSMAKGLNARMMRGLSYSLTTSST---H	249
RAB33A	CPC-----	237
RAB33B	CWC-----	229
RAB30	TCCNFN-----	203
RAB19	CTC-----	217
RAB43	CGC-----	212
RAB18	YCSVL-----	206
RAB2A	CC-----	212
RAB2B	CC-----	216
RAB14	C-----	215
RAB4A	C-----	218
RAB4B	C-----	213
RAB25	CCISL-----	213
RAB11A	CCQNI-----	216
RAB11B	CCQNL-----	218
RAB12	CC-----	244
RAB26	CCRP-----	256
RAB37	CCSFM-----	223
RAB3D	CSC-----	219
RAB3B	CSC-----	219
RAB3A	CAC-----	220
RAB3C	CAC-----	227
RAB15	CWC-----	212
RAB13	CSLG-----	203
RAB10	CC-----	200
RAB8A	CVLL-----	207
RAB8B	CSLL-----	207
RAB35	CC-----	201
RAB1A	CC-----	205
RAB1B	CC-----	201

RAB27A	-----	221
RAB27B	-----	218
RAB24	-----	203
RAB6C	-----	254
RAB6D	-----	254
RAB6A	-----	208
RAB6B	-----	208
RAB17	-----	212
RAB21	-----	225
RAB5B	-----	215
RAB5A	-----	215
RAB5C	-----	216
RAB22A	-----	194
RAB22B(31)	-----	194
RAB20	-----	234
RAB28	-----	221
RAB29	-----	203
RAB32	-----	225
RAB38	-----	211
RAB34	-----	259
RAB36	-----	333
RAB42	-----	218
RAB39A	-----	217
RAB39B	-----	213
RAB7B	-----	199
RAB7A	-----	207
RAB9A	-----	201
RAB9B	-----	201
RAB23	-----	237
RAB40C	KGNSLKRSKSIRPPQSPPQNCRSNCKIS	281
RAB40B	KRSSLRKVKLVPPQSPPKNCTRNSCKIS	278
RAB40A	K-SSLCKVEIVCPPQSPPKNCTRNSCKIS	277
RAB40AL	KRSSLCKVKIVCPPQSPPKNCTRNSCKIS	278
RAB33A	-----	237
RAB33B	-----	229
RAB30	-----	203
RAB19	-----	217
RAB43	-----	212
RAB18	-----	206
RAB2A	-----	212
RAB2B	-----	216
RAB14	-----	215
RAB4A	-----	218
RAB4B	-----	213
RAB25	-----	213
RAB11A	-----	216
RAB11B	-----	218
RAB12	-----	244
RAB26	-----	256
RAB37	-----	223
RAB3D	-----	219
RAB3B	-----	219
RAB3A	-----	220
RAB3C	-----	227
RAB15	-----	212
RAB13	-----	203
RAB10	-----	200
RAB8A	-----	207
RAB8B	-----	207
RAB35	-----	201
RAB1A	-----	205
RAB1B	-----	201

Figure S4. Full Sequence alignment of Rab proteins in phylogenetic order compared to Rab27A and B (top). Unique cysteines C123 and C188 are highlighted in red.

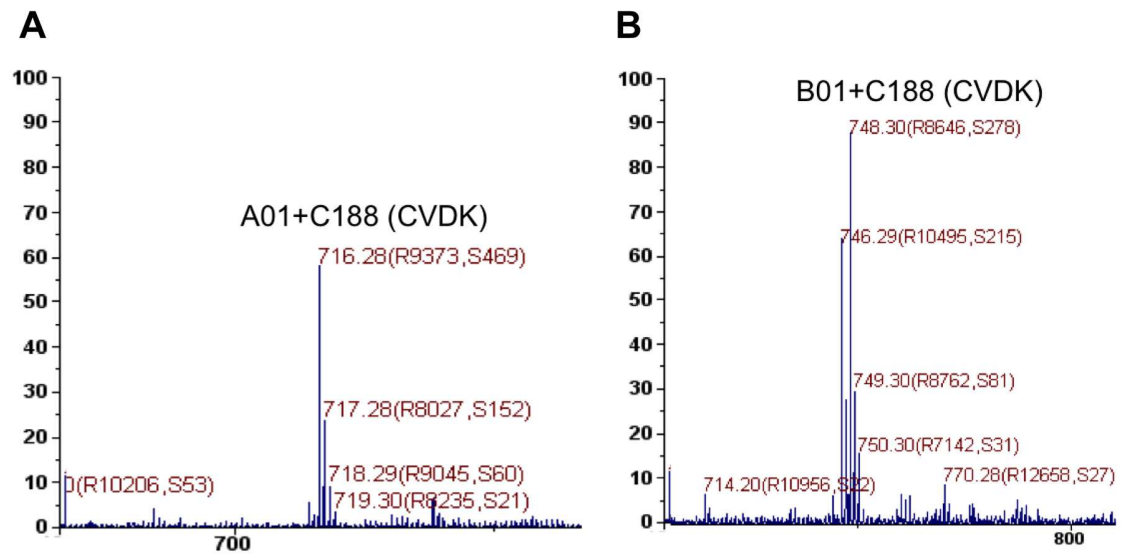


Figure S5. Tryptic digestion and peptide mass fingerprinting for labelling site-ID of A) **A01**-fRab27A-C188 and B) **B01**-fRab27A-C188.

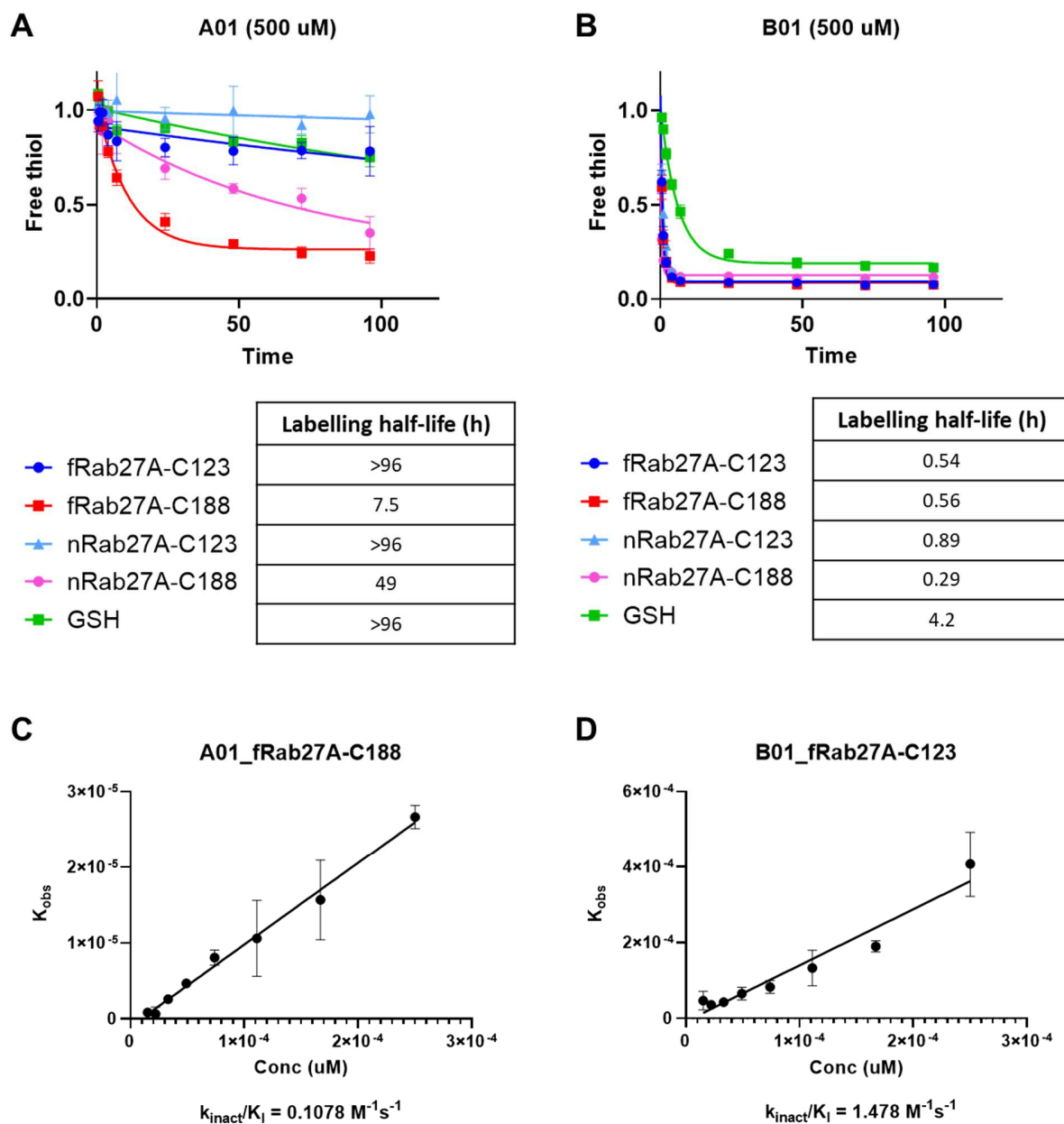


Figure S6. Biochemical characterisation of hits. A–B) qIT data against fRab27A-C123, fRab27A-C188, nRab27A-C123, nRab27A-C188 and GSH including labelling half-lives for resynthesised hits **A01** (A) and **B01** (B). C–D) $k_{obs}/[I]$ graphs for **A01** against fRab27A-C188 (C), and for **B01** against fRab27A-C123 (D)

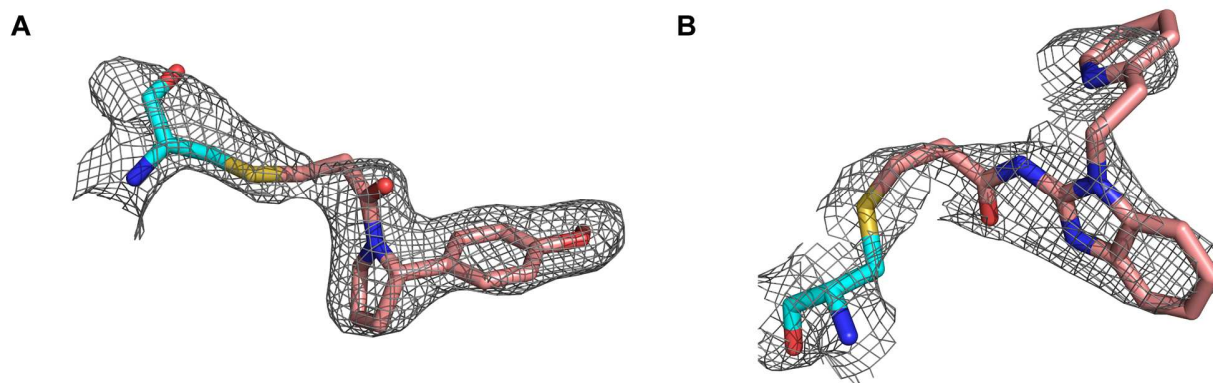


Figure S7. Electron density maps for **A01** and **B01**. $2F_o - F_c$ electron density maps (grey) contoured at 1.0σ for ligands (pink) covalently bound to a cysteine residue (cyan). A) **A01** bound to fRab27A-C188 (full structure shown in **Fig. 3B** and **3C**) and B) **B01** bound to fRab27A-C123 (full structure shown in **Fig. 3E** and **3F**) superimposed on the final model of the respective ligands.

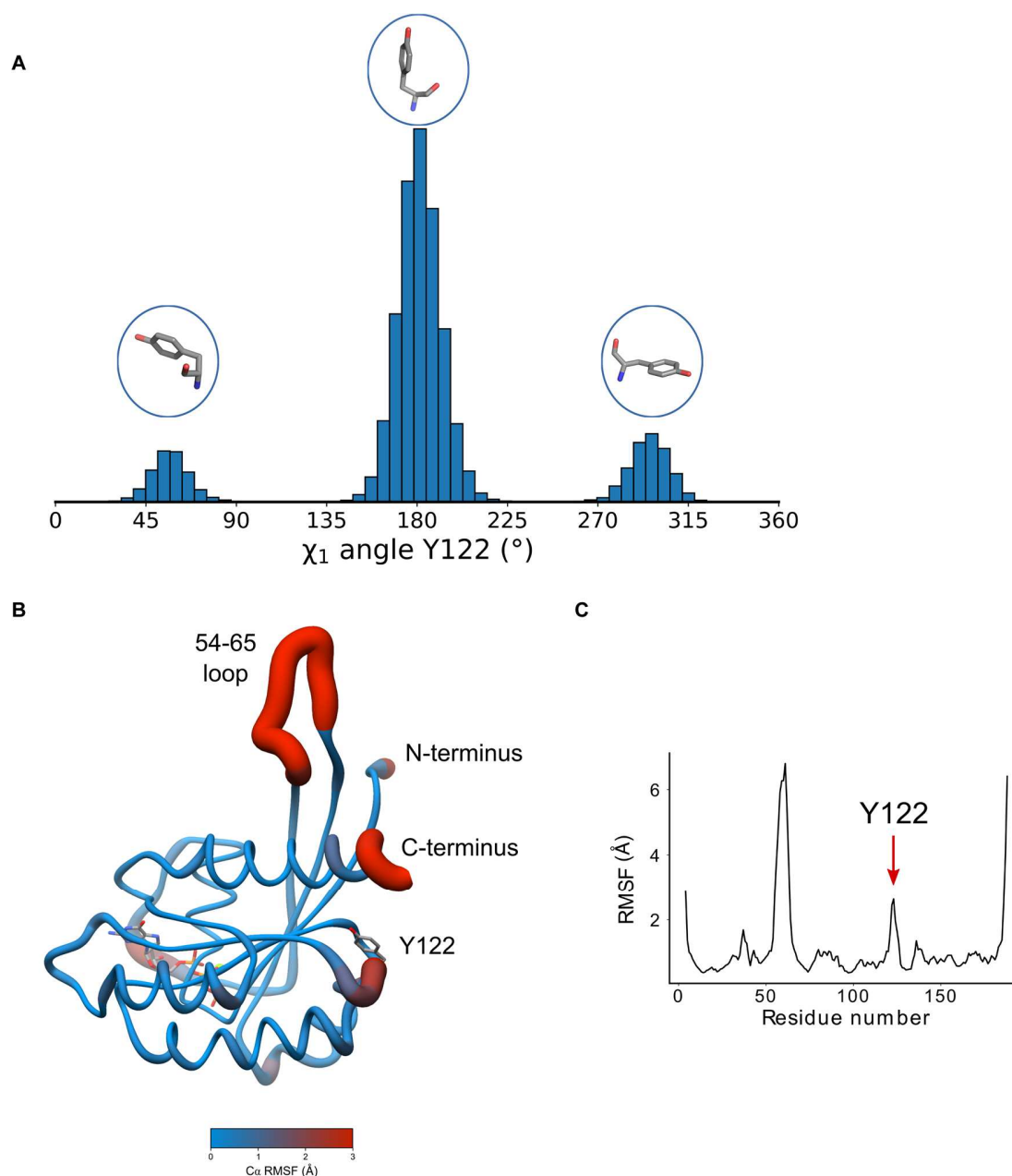


Figure S8. Molecular dynamics and rotameric properties of Y122 in Rab27A. (A) Distribution of chi-1 angles of Y122 during a 250 ns molecular dynamics simulation. (B) Structure of Rab27A, coloured according to the C-alpha Root Mean Squared Fluctuations. Red colours (thicker ribbon) correspond to more mobile regions, whereas blue regions are more rigid. (C) C-alpha Root Mean Squared Fluctuations of Rab27A plotted against its primary sequence.

Supplementary Tables

Table S1. Data Processing and Refinement Statistics for fRab27A

Data Collection	
Space group	<i>P</i> 3 ₁ 21
Unit cell parameters (Å)	<i>a</i> = 117.71, <i>b</i> = 117.71, <i>c</i> = 115.67
Wavelength (Å)	0.97949
Resolution (Å)	50.97 - 2.32 (2.40 - 2.32)
Total reflections	80969 (7952)
Unique reflections	40486 (3976)
Multiplicity	2.0 (2.0)
Completeness (%)	99.96 (99.95)
$\langle I \rangle / \langle \sigma(I) \rangle$	20.04 (3.29)
R_{merge}	0.027 (0.240)
R_{meas}	0.039 (0.340)
Wilson <i>B</i> factor	36.64
CC _{1/2}	0.999 (0.844)
Refinement	
Reflections used in refinement	40483 (3976)
Reflections used for R_{free}	1945 (229)
R_{work} (%)	0.159
R_{free} (%)	0.198
Rmsd bond lengths (Å)	0.008
Rmsd bond angles (°)	1.10
Average <i>B</i> factors (Å ²)/Number of atoms	
Macromolecules	42.13/ 3426
Water molecules	47.12/398
Ligand non-H atoms gppnhp-Mg ²⁺	28.3/66
Φ/Ψ angles (%)	
Ramachandran Most favored region (%)	97.79
Ramachandran allowed region (%)	2.1
Ramachandran outliers (%)	0.0
Rotamer outliers (%)	0.56

Statistics for the highest-resolution shell are shown in parentheses.

$$R_{\text{merge}} = \sum (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

$$R_{\text{meas}} = \sum \sqrt{(n_h/n_h - 1)} (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

Table S2. Data Processing and Refinement Statistics for fRab27A-C188 covalently bound to **A01**

Data Collection	
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁
Unit cell parameters (Å)	<i>a</i> = 61.71, <i>b</i> = 76.82, <i>c</i> = 117.82
Wavelength (Å)	0.97949
Resolution (Å)	36.52 - 2.23 (2.31 - 2.23)
Total reflections	54026 (3920)
Unique reflections	27128 (2005)
Multiplicity	2.0 (2.0)
Completeness (%)	96.86 (72.64)
$\langle I \rangle / \langle \sigma(I) \rangle$	13.67 (2.01)
R_{merge}	0.0340 (0.360)
R_{meas}	0.048 (0.509)
Wilson <i>B</i> factor	41.61
CC _{1/2}	1 (0.929)
Refinement	
Reflections used in refinement	27126 (2005)
Reflections used for R_{free}	1342 (98)
R_{work} (%)	0.177
R_{free} (%)	0.224
Rmsd bond lengths (Å)	0.255
Rmsd bond angles (°)	2.82
Average <i>B</i> factors (Å ²)/Number of atoms	
Macromolecules	46.28/3287
Water molecules	50.94/179
Ligand non-H atoms gppnhp-Mg ²⁺	51.70/160
Φ/Ψ angles (%)	
Ramachandran Most favored region (%)	98.02
Ramachandran allowed region (%)	1.98
Ramachandran outliers (%)	0.0
Rotamer outliers (%)	0.00

Statistics for the highest-resolution shell are shown in parentheses.

$$R_{\text{merge}} = \sum (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

$$R_{\text{meas}} = \sum \sqrt{(n_h/n_h - 1)} (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

Table S3. Data Processing and Refinement Statistics for fRab27A-C123 covalently bound to **B01**

Data Collection	
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁
Unit cell parameters (Å)	<i>a</i> = 61.38, <i>b</i> = 76.66, <i>c</i> = 118.24
Wavelength (Å)	0.97949
Resolution (Å)	64.33 - 2.32 (2.40 - 2.32)
Total reflections	49537 (4902)
Unique reflections	24831 (2455)
Multiplicity	2.0 (2.0)
Completeness (%)	99.82 (99.67)
$\langle I \rangle / \langle \sigma(I) \rangle$	9.77 (2.23)
R_{merge}	0.059 (0.325)
R_{meas}	0.083 (0.460)
Wilson <i>B</i> factor	32.90
CC _{1/2}	0.993 (0.832)
Refinement	
Reflections used in refinement	24809 (2448)
Reflections used for R_{free}	1209 (116)
R_{work} (%)	0.183
R_{free} (%)	0.254
Rmsd bond lengths (Å)	0.008
Rmsd bond angles (°)	1.02
Average <i>B</i> factors (Å ²)/Number of atoms	
Macromolecules	35.73/3449
Water molecules	38.33/249
Ligand non-H atoms gppnhp-Mg ²⁺	41.01/176
Φ/Ψ angles (%)	
Ramachandran Most favored region (%)	96.59
Ramachandran allowed region (%)	3.41
Ramachandran outliers (%)	0.0
Rotamer outliers (%)	0.84

Statistics for the highest-resolution shell are shown in parentheses.

$$R_{\text{merge}} = \sum (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

$$R_{\text{meas}} = \sum \sqrt{(n_h/n_h - 1)} (|I_h| - \langle I_h \rangle) / \sum \langle I_h \rangle$$

Table S4. Data from intact mass spectrometry and qIT screen used for hit validation against fRab27A-C123 (top) and fRab27A-C188 (bottom).

fRab27A-C123

	REF	qIT half-life	Mono-modification by intact protein MS	MS half-life	Validated
CA32/228	4.2	21.4 h	Yes: expect 250 Da, observed 252	34.3 h	No
CA84	2.3004E-12	-	Wrong mass: expect 255 Da, observed 326	-	No
CA144 (B01)	2.9	4.7 h	Yes: expect 292 Da, observed 293	6.6 h	Yes
CA92	4.8	-	Wrong mass: expect 268 Da, observed 352	-	No

fRab27A-C188

	REF	qIT half-life	Mono-modification by intact protein MS	MS half-life	Validated
CA32/228	7.6	-	Wrong mass: expect 250 Da, observed 359	-	No
CA84	2.3	-	Protein degraded	-	No
CA89	1.6	4.3 h	Yes: expect 284 Da, observed 282	7.4 h	Yes
CA144 (B01)	2.4	4.5 h	Yes: expect 292 Da, observed 291	11.8 h	No
EL1062 (A01)	2.5	26.2 h	Yes: expect 231 Da, observed 231	32.5 h	Yes
EL1064	2.2	12.8 h	Yes: expect 252 Da, observed 253	15.8 h	Yes
CA193	2	-	Wrong mass: expect 209 Da, observed 261	-	No
CA187	1.7	4.3 h	Yes: expect 252 Da, observed 250	13.2 h	No
CA53	5.7	-	No labelling	-	No

Materials and Methods

Protein expression and purification

All Rab27A constructs contain the sequence for human Rab27A (UniProt entry P51159, residues 1–192, mutations: Q78L and C123S or C188S or both as specified), which was cloned into a pET15b vector (Invitrogen) including a N-terminal His-tag followed by a Tobacco Etch Virus (TEV) recognition site (ENLYFQ↓G). Fusion constructs also contain the C-terminus of Slp2a SHD1 (SFLTEEEQEAIMKVLQRDAALKRAEEER (residues 5–32)) linked to the N-terminus of Rab27A via a flexible poly glycine-serine linker (GSGSGSG). For protein expression, plasmids were transformed to *E. coli* BL21 cells and spread on LB agar plates containing 100 mg/L Ampicillin for selection. Single colonies were picked for amplification and incubated overnight into LB media containing 100 mg/L Ampicillin at 37 °C, shaking. Big scale cultures were inoculated using these overnight cultures at 1% v/v, and grown at 37 °C until absorbance at 600 nm reached 0.7. Protein expression was induced using 0.5 mM isopropyl β-D-1-thiogalactopyranoside (IPTG) at 37 °C for 3 hours. Subsequently cells were pelleted at 4k rpm for 10 min, then re-suspended in lysis buffer containing 500 mM NaCl, 10 mM imidazole, 5 mM MgCl₂ and 50 mM Tris at pH 8.0. Cells were lysed with a cell disruptor at 25K psi and centrifuged at 15k rpm for 45 min. The supernatant was loaded on Ni²⁺-NTA resin equilibrated with lysis buffer, washed extensively and eluted using buffer containing 500 mM NaCl, 300 mM imidazole, 5 mM MgCl₂ and 50 mM Tris at pH 8.0. The protein was dialyzed for 6 h using 100 mM NaCl, 5 mM MgCl₂ and 50 mM Tris, pH 8.0. Afterwards TEV protease (obtained in-house as previously described⁵) was added to the protein solution at a molar ratio of 1/20 in the presence of 1 mM DTT, and the solution was incubated overnight at 4 °C, shaking. The solution was dialyzed using 100 mM NaCl, 5 mM MgCl₂ and 50 mM Tris, pH 8 and then loaded on Ni²⁺-NTA resin. The flowthrough was collected, concentrated to 5.5 mg/mL in 150 mM NaCl, 5 mM MgCl₂, 20 mM Tris pH 8 buffer. Then a 10x buffer containing 10 mM ZnCl₂ and 2 M (NH₄)₂SO₄, 4 molar excess of GppNHp and 25 units of Antarctic phosphatase (New England Biolabs) were added to the solution and incubated overnight at 4 °C. Finally the sample was loaded on a superdex S-75 gel filtration column at a flow rate of 1 mL/min. The column was pre-equilibrated with 150 mM NaCl, 5 mM MgCl₂, and 20 mM Tris at pH 8 for crystallography. The peaks corresponding to Rab27A constructs were analysed by SDS-PAGE, pooled, concentrated and flash frozen using liquid nitrogen. All Rab27A constructs containing exposed cysteines at C123 or C188 were purified in buffer containing an additional 0.1% β-mercaptoethanol (βME) during Ni²⁺-NTA steps.

Protein labelling and purification

To a 15 mL falcon tube were added 600 μL of desired construct (100 μM stock), 100 μL of 50% w/v TCEP-agarose beads (ThermoFisher), and 1.8 mL of 100 mM NaCl, 5 mM MgCl₂, and 20 mM HEPES pH 8.0. 50 μL of ligand (50 mM stock) were diluted with 2.45 mL of 100 mM NaCl, 5 mM MgCl₂, and 20 mM HEPES pH 8.0, followed by centrifugation (2400 rpm, 5 min). The supernatant was added to the protein mixture and incubated at 4 °C. The reaction was monitored as described in the QIT protocol (*v. infra*). When the labelling reached 90%, the labelled protein solution was concentrated to 0.5 mL by using a Vivaspin 20 filter (5000 MWCO). The protein was diluted with 4.5 mL of 100 mM NaCl, 5 mM MgCl₂, and 20 mM HEPES pH 8.0 and concentrated again to 0.5 mL (5x), to remove excess compound, then purified by superdex S-75 gel filtration at a flow rate of 1 mL/min in 150 mM NaCl, 5 mM MgCl₂, and 20 mM Tris at pH 8 for crystallography.

Protein crystallisation

Pure samples of fRab27A-C188 labelled with **A01** and fRab27A-C123 labelled with **B01** were concentrated to 15 mg/mL, in buffer containing 150 mM NaCl, 5 mM MgCl₂, and 20 mM Tris at pH 8.0. Crystals were grown at 4 °C using the sitting-drop vapor-diffusion method with a mother liquor containing 120 mM MgCl₂, 50 mM bis-Tris, and 15% 2-propanol, pH 6.8.

Crystal diffraction, data collection and data processing

Data collections were carried out at i02 beamline in Diamond Light Source (Oxford, UK) at 100K of temperature, wavelength 0.9795 Å, and using a Pilatus detector. Data were collected at 0.2°-0.5° oscillations per image and 200° total oscillation per crystal. Data was integrated, scaled and reduced using DIALS.⁶ Initial phases were calculated using the molecular replacement program Phaser.⁷ The coordinates of Rab27A from chain A of the Rab27A-Slp2a complex (PDB:3BC1) without the nucleotide and the magnesium ion were used as the search model. Subsequently, the initial model generated by phaser was refined through an iterative cycle using COOT⁹ and REFMAC5.¹⁰ Final model structures were validated using the Molprobit server¹¹ at <http://molprobit.biochem.duke.edu>. All structure images were prepared using Pymol (DeLano Scientific LLC, <http://pymol.sourceforge.net/>). X-ray data collection, processing and refinement statistics are given in Table S1.

Molecular dynamics simulations

The Rab27A structure (PDB: 3BC1) was simulated with bound GTP and Mg²⁺. The structure was parametrised using the latest CHARMM36 force field⁴, solvated with Tip3p waters¹² and neutralised with Na⁺ and Cl⁻ ions at a concentration of 150 mM. Temperature was coupled for 100 ps at 300 K with the V-rescale method,¹³ with positional restraints on the protein heavy atoms. Pressure was then coupled at 1 bar for another 100 ps with position restraints, using the Berendsen algorithm.¹⁴ The Particle mesh Ewald method¹⁵ was used for electrostatic interactions, and LINCS¹⁶ to define the constraints. The integration timestep was 2 fs. The final production simulation was extended for 250 ns. The simulation and data analysis were carried out using the GROMACS simulation package.¹⁷

qIT assay for screening and hit validation

126 electrophilic acrylamides (see supplementary excel file) were screened in the qIT assay adapted from Craven *et al*¹⁸. Briefly, the reaction buffer (20 mM HEPES pH 8.0, 100 mM NaCl, 5 mM MgCl₂) and quench buffer (20 mM HEPES pH 7.4, 100 mM NaCl, 5 mM MgCl₂) were prepared, filtered, de-gassed, and re-gassed with Ar for 15 min on ice. Reaction setup: To each well of a 96-well PCR plate (reaction plate), 8 µL of 50% w/v TCEP-agarose beads in reaction buffer was added, followed by the addition of 92 µL of 10.87 µM protein or glutathione (GSH). In a separate 96-well PCR plate (ligand plate), 3 µL of DMSO or 50 mM ligand in DMSO was added to 147 µL reaction buffer and centrifuged (1k rpm, 5 min, 4 °C). 100 µL of ligand solution or DMSO control from the ligand plate was added to the reaction plate (final concentration: 5 µM protein/GSH and 500 µM ligand). After mixing, the TCEP-agarose beads were pelleted by centrifugation (1k rpm, 5 min, 4 °C) and the plate was kept at 4 °C.

At a series of time points ($t = 0.25, 1, 2, 4, 7, 24, 48, 72$, and 96 h), a $3\ \mu\text{L}$ aliquot in duplicate from the reaction plate was quenched in a black 384-well plate, in which each well was pre-filled with $27\ \mu\text{L}$ of 7-Diethylamino-3-(4'-Maleimidylphenyl)-4-Methylcoumarin (CPM) solution ($1.4\ \mu\text{M}$ in quench buffer). The fluorescence plate was spun down (1 k rpm , 1 min) and incubated for 60 min at room temperature and then fluorescence intensity (excitation/emission: $384/470\text{ nm}$) was measured on an EnVision™ plate reader.

Data analysis: All analyses were conducted using Prism 9.0 software (Graphpad). Each fluorescence readout was normalized to the average of the DMSO controls. The normalized fluorescence was plotted against time. A one phase exponential decay was fitted to each plot (constraints: $Y(0) > 0.8$; $0 < \text{plateau} < 0.3$; $k > 0$). Data from at least three independent assay replicates were used to generate the graphs in Fig. S5.

qIT assay for k_{inact}/K_i determination

The k_{inact}/K_i values were determined from data obtained performing the qIT assay at different compound concentrations (eight 1:1.5 dilutions starting from $250\ \mu\text{M}$) at room temperature, quenching at different time-points ($t = 10, 20, 30, 60, 120, 180, 240, 360, 1440\text{ min}$). Kinetic curves of thiol labelling over time were used to estimate k_{obs} values, which were then plotted against inhibitor concentration. The resulting linear data were analysed by linear regression to obtain k_{inact}/K_i values.

Peptide mass fingerprint analysis

$5\ \mu\text{g}$ labelled or unlabelled recombinant Rab27A construct were run on a 12% SDS-PAGE gel and stained by Coomassie Blue. The expected band was excised and washed in $150\ \mu\text{L}$ of 50% v/v MeCN/ H_2O for 5 min at rt, shaking. The supernatant was discarded, and the solid was washed with $150\ \mu\text{L}$ of 50% v/v MeCN/ $50\text{ mM NH}_4\text{HCO}_3$ for 30 min at rt, shaking. The supernatant was discarded, and the solid was washed with $150\ \mu\text{L}$ of 50% v/v MeCN/ $10\text{ mM NH}_4\text{HCO}_3$ for 30 min at rt, shaking. The supernatant was dried *in vacuo* for 30 min at $45\text{ }^\circ\text{C}$, then $15\ \mu\text{L}$ of Trypsin ($20\ \mu\text{g}/100\ \mu\text{L}$ in $50\text{ mM NH}_4\text{HCO}_3$) were added. After 10 min at rt, the mixture was diluted with $15\ \mu\text{L}$ of $10\text{ mM NH}_4\text{HCO}_3$ and incubated overnight at $37\text{ }^\circ\text{C}$, shaking. The supernatant was diluted 1:1 with α -Cyano-4-hydroxycinnamic acid (10 mg/mL in 50% v/v MeCN/ H_2O with 0.1% TFA) and analysed by MALDI-QTOF.

Chemical Synthesis

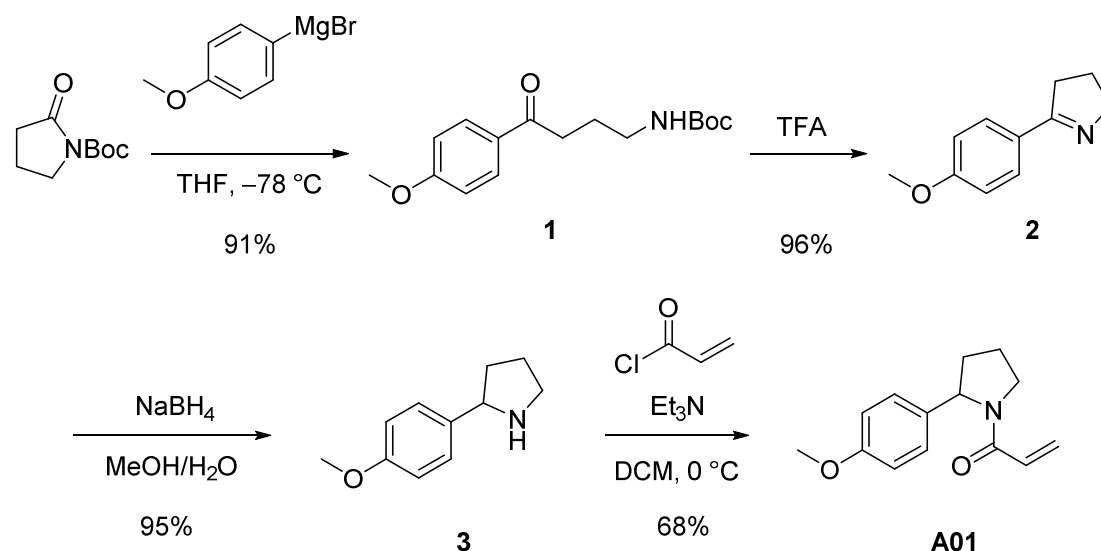
Abbreviations

DMF (dimethylformamide), EtOAc (ethyl acetate), FCC (flash column chromatography), rt (room temperature), TFA (trifluoroacetic acid), THF (tetrahydrofuran), TLC (thin layer chromatography)

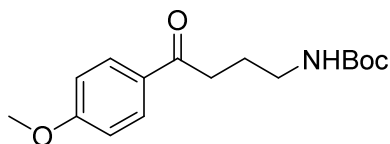
General Information

All chemicals were purchased from Sigma-Aldrich, Apollo Scientific, Acros Organics, Alfa Aesar and used without further purification unless otherwise indicated. All reactions were monitored by thin layer chromatography (TLC) using UV for visualisation unless otherwise stated. Compounds were purified using either an automated system using pre-packed silica cartridges with UV detection or by manual columns using an appropriate solvent mixture as detailed. ^1H and ^{13}C NMR spectra were recorded on 400 MHz and 101 MHz respectively Bruker AV instruments at room temperature unless specified otherwise and were referenced to residual solvent signals. Data are presented as follows: chemical shift in ppm, multiplicity (br = broad, app = apparent, s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, m = multiplet), coupling constants in Hz, integration, and rotameric conformation if applicable. High-resolution mass spectrometry (HRMS) and intact mass spectrometry data were obtained by the Imperial Mass Spectrometry facility. m/z values are reported in Daltons (Da) to the nearest 0.0001 Da.

Scheme S1. Synthesis of hit fragment **A01**



***tert*-butyl (4-(4-methoxyphenyl)-4-oxobutyl)carbamate (1)**

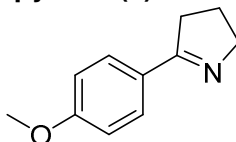


(4-Methoxyphenyl)magnesium bromide (0.5 M in THF, 24 mL, 12 mmol) was added dropwise over 30 min at -78°C to a stirred solution of *tert*-butyl 2-oxopyrrolidine-1-carboxylate (1.7 mL, 10 mmol) in THF (40 mL). The reaction was stirred for 1 h at -78°C , then slowly warmed to rt and stirred for 1 h before the pH was adjusted to 1–3 using 1 M HCl. The solution was concentrated *in vacuo*, then diluted with CH_2Cl_2 (70 mL) and NaHCO_3 (70 mL), and the aqueous layer extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic layers were dried over MgSO_4 , filtered, concentrated *in vacuo*. The crude product was purified by FCC (6%–60% EtOAc/hexane) to give the title compound **1** as a white amorphous solid (2.7 g, 91%).

^1H NMR (400 MHz, CDCl_3) δ 7.95–7.92 (2H, m), 6.94–6.91 (2H, m), 4.67 (br, 1H, NH), 3.87 (3H, s), 3.24–3.19 (2H, m), 2.97 (2H, t, $J = 7.2$ Hz), 1.92 (2H, p, $J = 7.0$ Hz), 1.42 (9H, s) ppm

^{13}C NMR (101 MHz, CDCl_3) δ 198.5, 163.6, 156.2, 130.4, 130.1, 113.9, 79.3, 55.6, 40.4, 35.5, 28.5, 24.8 ppm

5-(4-methoxyphenyl)-3,4-dihydro-2H-pyrrole (2)



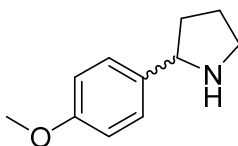
Ketone **1** (0.59 g, 2.0 mmol) was stirred in neat TFA (1.5 mL) at rt for 3.5 h. After the reaction was complete by TLC, the mixture was cooled to 0°C and 50% w/v NaOH solution was added to the mixture until pH 13–14. The aqueous layer was extracted with CH_2Cl_2 (3 \times 25 mL), then the combined organic layers were dried over MgSO_4 , filtered, and concentrated *in vacuo* to afford the title compound **2** as a white crystalline solid which was used without further purification (0.34 g, 96%).

^1H NMR (400 MHz, CDCl_3) δ 7.82–7.78 (2H, m), 6.94–6.90 (2H, m), 4.04 (2H, tt, $J = 7.3$, 1.9 Hz), 3.84 (3H, s), 2.95–2.90 (2H, m), 2.06–1.99 (2H, m) ppm

^{13}C NMR (101 MHz, CDCl_3) δ 172.9, 161.5, 129.4, 127.5, 113.9, 61.4, 55.5, 35.0, 22.8 ppm

HRMS (ES) m/z Calculated for $\text{C}_{11}\text{H}_{14}\text{NO}$ $[\text{M}+\text{H}]^+$ 176.1075, found 176.1080 (Δ 2.8 ppm)

2-(4-methoxyphenyl)pyrrolidine (3)



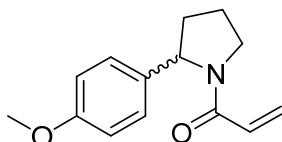
To a stirring solution of imide **2** (0.30 g, 1.7 mmol) in MeOH/H₂O 4:1 (2.0 mL) was added NaBH₄ (78 mg, 2.0 mmol) and the reaction was stirred for 20 h at rt. Additional NaBH₄ (20 mg, 0.8 mmol) was added and the reaction was stirred until completion as monitored by TLC. The reaction mixture was acidified with 1 M HCl to pH 1–3 and stirred for an additional 30 min, then 1 M NaOH was added until pH 13–15. The aqueous layer was extracted with CH₂Cl₂ (3x 10 mL). The combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo* to give the title compound **3** as a yellow oil (81 mg, 95%), which was used without further purification.

¹H NMR (400 MHz, CDCl₃) δ 7.30–7.26 (2H, m), 6.87–6.84 (2H, m), 4.05 (1H, t, *J* = 7.8 Hz), 3.79 (3H, s), 3.19 (1H, ddd, *J* = 10.3, 7.8, 5.3 Hz), 2.98 (1H, ddd, *J* = 10.3, 8.4, 6.6 Hz), 2.24 (1H, br s), 2.18–2.11 (1H, m), 1.99–1.78 (2H, m), 1.72–1.59 (1H, m) ppm

¹³C NMR (100 MHz, CDCl₃) δ 158.6, 136.6, 127.8, 113.9, 62.3, 55.4, 47.0, 34.3, 25.7 ppm

HRMS (ES) *m/z* Calculated for C₁₁H₁₆NO [M+H]⁺ 178.1232, found 178.1232 (Δ 0.0 ppm)

1-(2-(4-methoxyphenyl)pyrrolidin-1-yl)prop-2-en-1-one (A01)



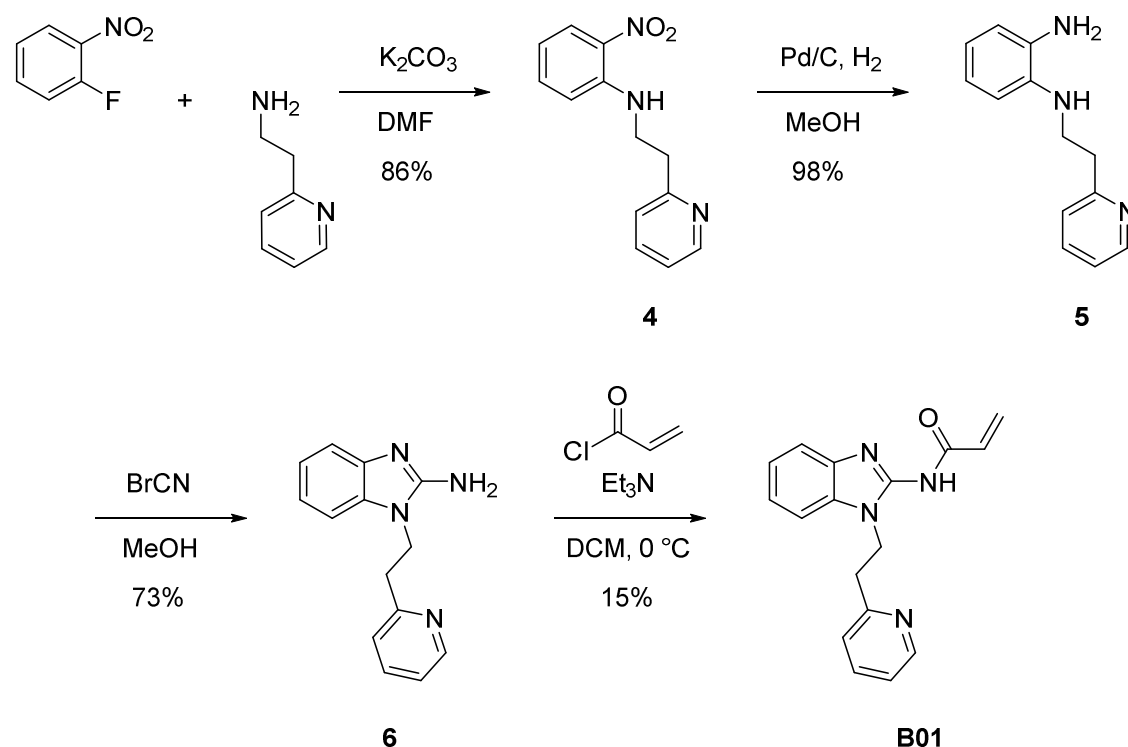
To a stirred solution of amine **3** (89 mg, 0.50 mmol) and Et₃N (0.10 mL, 0.75 mmol) in CH₂Cl₂ (1.5 mL) at 0 °C was added acryloyl chloride (49 μL, 0.60 mmol) in CH₂Cl₂ (0.50 mL) dropwise. The reaction was allowed to warm to rt, stirred for 2 h, diluted with CH₂Cl₂ (10 mL) and then quenched by slow addition of NaHCO₃ (10 mL). The aqueous layer was extracted with CH₂Cl₂ (2x 10 mL) then the combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by FCC (12%–100% EtOAc/hexanes) afforded the title compound **A01** as a clear, colourless oil (79 mg, 68%).

¹H NMR (400 MHz, CDCl₃) *Rotameric ratio maj:min 0.76:0.24*; δ 7.09–7.03 (2H, m), 6.86–6.79 (2H, m), 6.53 (1H, dd, *J* = 16.8, 10.3 Hz, min. rot.), 6.34 (1H, dd, *J* = 16.8, 2.1 Hz, maj. rot.), 6.29 (1H, dd, *J* = 16.8, 2.2 Hz, maj. rot.), 6.12 (1H, dd, *J* = 16.7, 10.3 Hz, maj. rot.), 5.66 (1H, dd, *J* = 10.3, 2.2 Hz, min. rot.), 5.44 (1H, dd, *J* = 10.2, 2.2 Hz, maj. rot.), 5.21 (1H, dd, *J* = 8.0, 3.2 Hz, min. rot.), 5.00 (1H, dd, *J* = 7.8, 2.0 Hz, maj. rot.), 3.83–3.78 (1H, m), 3.77 (3H, s, maj. rot.), 3.74 (3H, s, min. rot.), 3.72–3.65 (1H, m), 2.38–2.18 (1H, m), 2.01–1.80 (3H, m) ppm

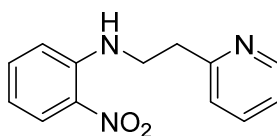
¹³C NMR (100 MHz, CDCl₃) δ 165.4 (maj. rot.), 164.4 (min. rot.), 158.8 (maj. rot.), 158.4 (min. rot.), 135.4 (maj. rot.), 135.0 (min. rot.), 129.0 (maj. rot.), 128.8 (min. rot.), 128.0 (min. rot.), 127.4 (maj. rot.), 126.7 (min. rot.), 126.6 (maj. rot.), 114.1 (maj. rot.), 113.9 (min. rot.), 60.8 (maj. rot.), 60.2 (min. rot.), 55.3, 47.6 (min. rot.), 47.1 (maj. rot.), 36.4 (maj. rot.), 34.0 (min. rot.), 23.9 (min. rot.), 21.6 (maj. rot.) ppm

HRMS (ES) *m/z* Calculated for C₁₄H₁₈NO₂ [M+H]⁺ 232.1338, found 232.1340 (Δ 0.9 ppm)

Scheme S2. Synthesis of hit fragment B01



2-nitro-N-(2-(pyridin-2-yl)ethyl)aniline (4)



1-Fluoro-2-nitrobenzene (0.42 mL, 4.0 mmol), 2-(2-aminoethyl)pyridine (0.48 mL, 4.0 mmol) and K₂CO₃ (1.1 g, 8.0 mmol) were dissolved in DMF (10 mL) and stirred at rt for 24 h. The reaction mixture was then diluted with EtOAc (50 mL) and NaHCO₃ (50 mL). The aqueous layer was extracted with EtOAc (3× 50 mL), then the combined organic layers were washed with 5% LiCl (30 mL) and brine (30 mL), dried over MgSO₄, filtered and concentrated *in vacuo*. Purification by

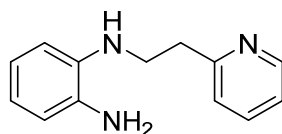
FCC (12%–100% EtOAc/hexane) afforded the title compound **4** as a bright orange oil (0.83 g, 86%).

¹H NMR (400 MHz, CDCl₃) δ 8.58 (1H, ddd, *J* = 4.9, 1.8, 0.9 Hz), 8.29 (1H, br s, NH), 8.12 (1H, dd, *J* = 8.7, 1.7 Hz), 7.61 (1H, dt, *J* = 7.7, 1.8 Hz), 7.40 (1H, ddd, *J* = 8.6, 7.0, 1.6 Hz), 7.20 (1H, d, *J* = 7.7 Hz), 7.15 (1H, ddd, *J* = 7.5, 4.9, 1.1 Hz), 6.90 (1H, d, *J* = 8.5 Hz), 6.60 (1H, ddd, *J* = 8.5, 6.9, 1.2 Hz), 3.73 (2H, dt, *J* = 6.8, 5.3 Hz), 3.17 (2H, t, *J* = 6.8 Hz) ppm

¹³C NMR (101 MHz, CDCl₃) δ 158.5, 149.8, 145.4, 136.8, 136.3, 132.0, 126.9, 123.5, 121.9, 115.3, 113.8, 42.7, 37.3 ppm

HRMS (ES) *m/z* Calculated for C₁₃H₁₄N₃O₂ [M+H]⁺ 244.1086, found 244.1090 (Δ 1.6 ppm)

***N'*-(2-(pyridin-2-yl)ethyl)benzene-1,2-diamine (5)**



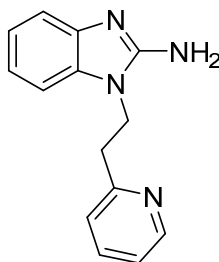
Pyridine **4** (0.65 mg, 2.7 mmol) was dissolved in MeOH (6.0 mL) and Pd/C (10%, 65.0 mg) was added. The reaction mixture was degassed and flushed with H₂ three times, then stirred at rt until reaction was complete by TLC. The reaction was filtered through celite to afford the title compound **5** as a dark brown oil (0.56 g, 98%), which was used without further purification.

¹H NMR (400 MHz, CDCl₃) δ 8.57 (1H, d, *J* = 4.6 Hz), 7.60 (1H, t, *J* = 7.7 Hz), 7.18 (1H, d, *J* = 7.8 Hz), 7.14 (1H, t, *J* = 6.2 Hz), 6.85–6.81 (1H, m), 6.73 (1H, d, *J* = 7.9 Hz), 6.71–6.66 (2H, m), 3.64 (3H, br), 3.53 (2H, t, *J* = 6.7 Hz), 3.14 (2H, t, *J* = 6.7 Hz) ppm

¹³C NMR (101 MHz, CDCl₃) δ 159.9, 149.3, 137.4, 136.5, 134.6, 123.3, 121.5, 120.4, 118.7, 116.2, 112.0, 44.0, 37.5 ppm

HRMS (ES) *m/z* Calculated for C₁₃H₁₆N₃ [M+H]⁺ 214.1344, found 214.1346 (Δ 0.2 ppm)

1-(2-(pyridin-2-yl)ethyl)-1*H*-benzo[d]imidazol-2-amine (6)



Aniline **5** (0.57 mg, 2.7 mmol) was dissolved in MeOH (20 mL), then cyanogen bromide (0.60 g, 4.0 mmol) was added to the solution. The reaction was stirred at rt for 2 h, and then concentrated *in vacuo*. The residue was diluted with EtOAc (60 mL) and 1 M NaOH (50 mL), and the aqueous layer extracted with EtOAc (3× 60 mL). The combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, filtered and concentrated *in vacuo*. Purification by FCC (5% MeOH/CH₂Cl₂, 0.5% NH₄OH) afforded the title compound **6** as a purple-grey powder (0.46 g, 73%).

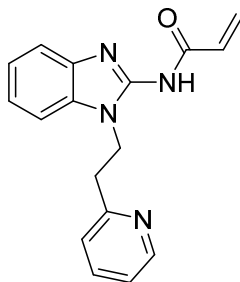
R_f 0.39 (10% MeOH/CH₂Cl₂, 0.5% NH₄OH)

¹H NMR (400 MHz, CDCl₃) δ 8.54 (1H, ddd, *J* = 4.9, 1.6, 0.7 Hz), 7.51 (1H, dt, *J* = 7.7, 1.8 Hz), 7.36 (1H, m), 7.16–7.03 (4H, m), 6.98 (1H, d, *J* = 7.9 Hz), 5.41 (2H, br), 4.48 (2H, t, *J* = 6.0 Hz), 3.32 (2H, t, *J* = 6.1 Hz) ppm

¹³C NMR (101 MHz, CDCl₃) δ 157.4, 154.5, 149.0, 141.8, 137.1, 134.1, 124.4, 122.3, 121.6, 119.7, 116.2, 107.5, 40.7, 36.6 ppm

HRMS (ES) *m/z* Calculated for C₁₄H₁₅N₄ [M+H]⁺ 239.1297, found 239.1297 (Δ 0.0 ppm)

***N*-(1-(2-(pyridin-2-yl)ethyl)-1*H*-benzo[*d*]imidazol-2-yl)acrylamide (B01)**



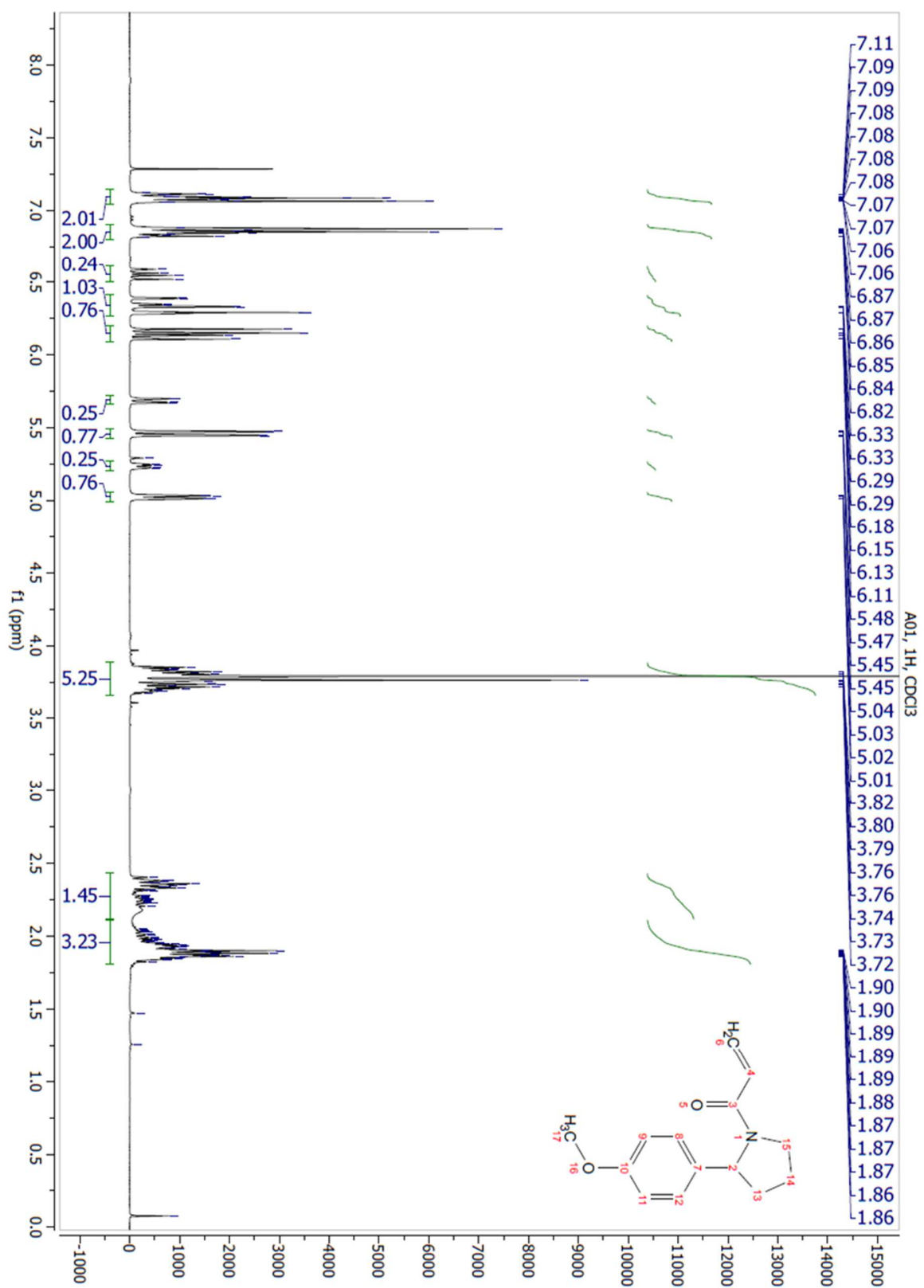
Acryloyl chloride (31 μL, 0.39 mmol) was dissolved in CH₂Cl₂ (1.0 mL) and the resulting solution was added dropwise to a stirring solution of benzimidazole **6** (92 mg, 0.39 mmol) and Et₃N (81 μL, 0.58 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. The reaction was then allowed to warm to rt, stirred for 2 h, diluted with CH₂Cl₂ (10 mL) and then quenched by addition of NaHCO₃ (10 mL). The aqueous layer was extracted with CH₂Cl₂ (3× 10 mL), then the combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by FCC (25%–100% EtOAc/hexane) afforded the title compound as a white solid (9.6 mg, 10%).

¹H NMR (400 MHz, CDCl₃) δ 12.30 (s, 1H), 8.58 (ddd, *J* = 4.9, 1.9, 0.9 Hz, 1H), 7.51 (td, *J* = 7.7, 1.9 Hz, 1H), 7.31–7.25 (m, 1H), 7.21–7.10 (m, 4H), 7.06 (dt, *J* = 7.8, 1.1 Hz, 1H), 6.47 (d, *J* = 6.0 Hz, 2H), 5.76–5.64 (m, 1H), 4.59 (dd, *J* = 7.6, 6.7 Hz, 2H), 3.33 (t, *J* = 7.1 Hz, 2H) ppm

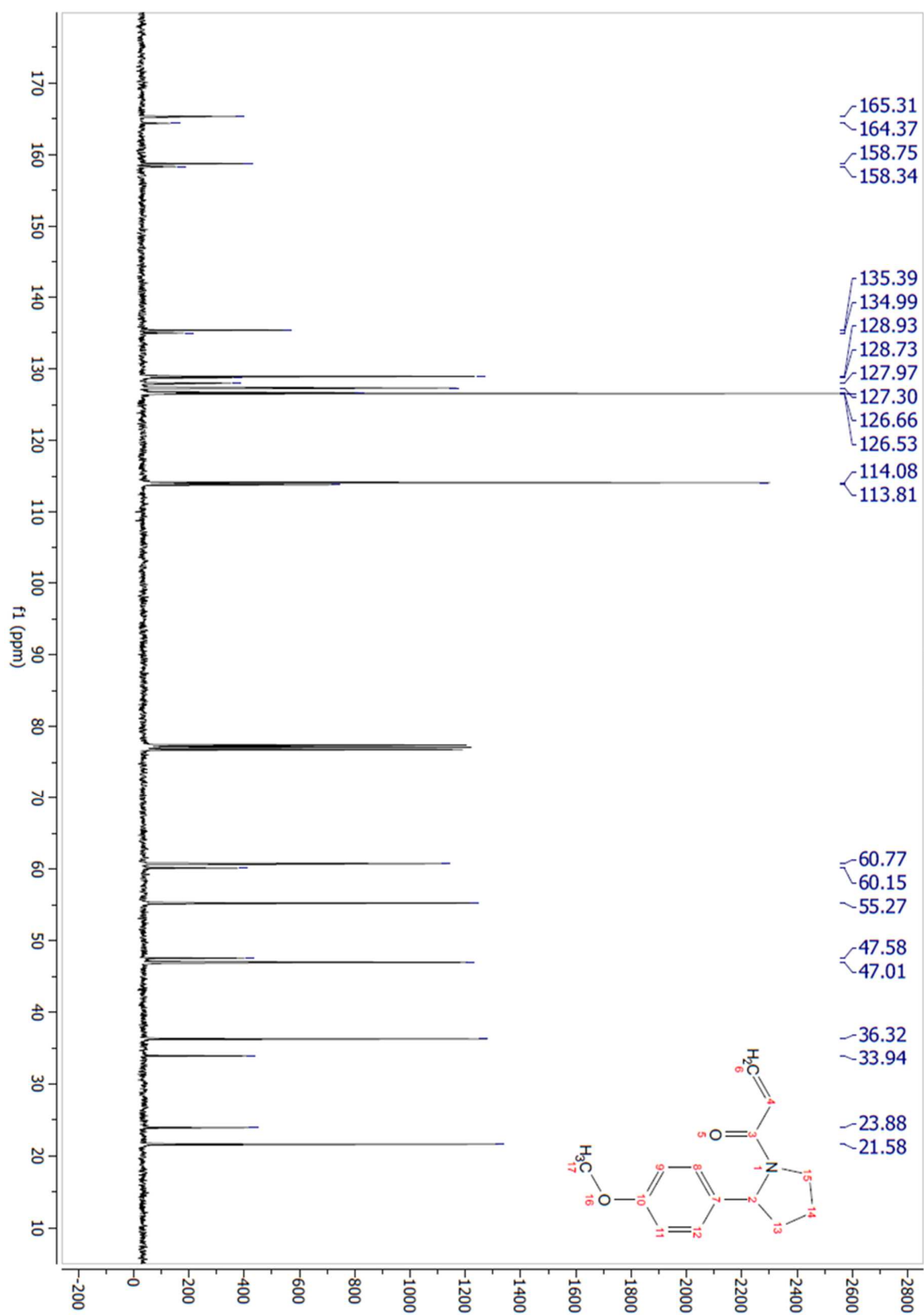
¹³C NMR (101 MHz, CDCl₃) δ 176.5, 158.0, 153.5, 149.5, 137.1, 136.5, 129.6, 126.0, 123.7, 122.9, 122.8, 121.9, 111.2, 109.4, 42.0, 36.7 ppm

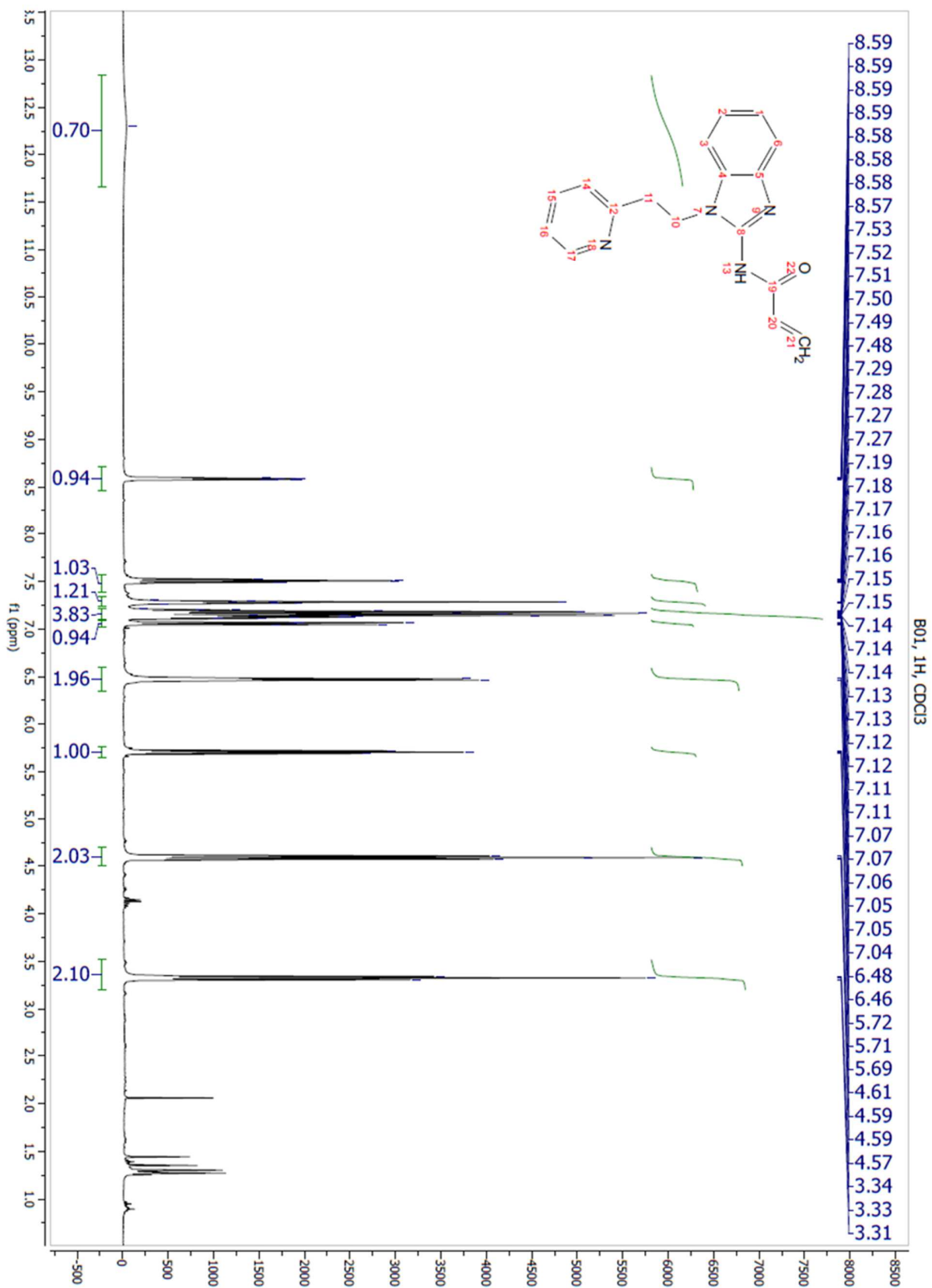
HRMS (ES) *m/z* Calculated for C₁₂H₁₅N₄ [M+H]⁺ 215.1297, found 215.1297 (Δ 0.0 ppm)

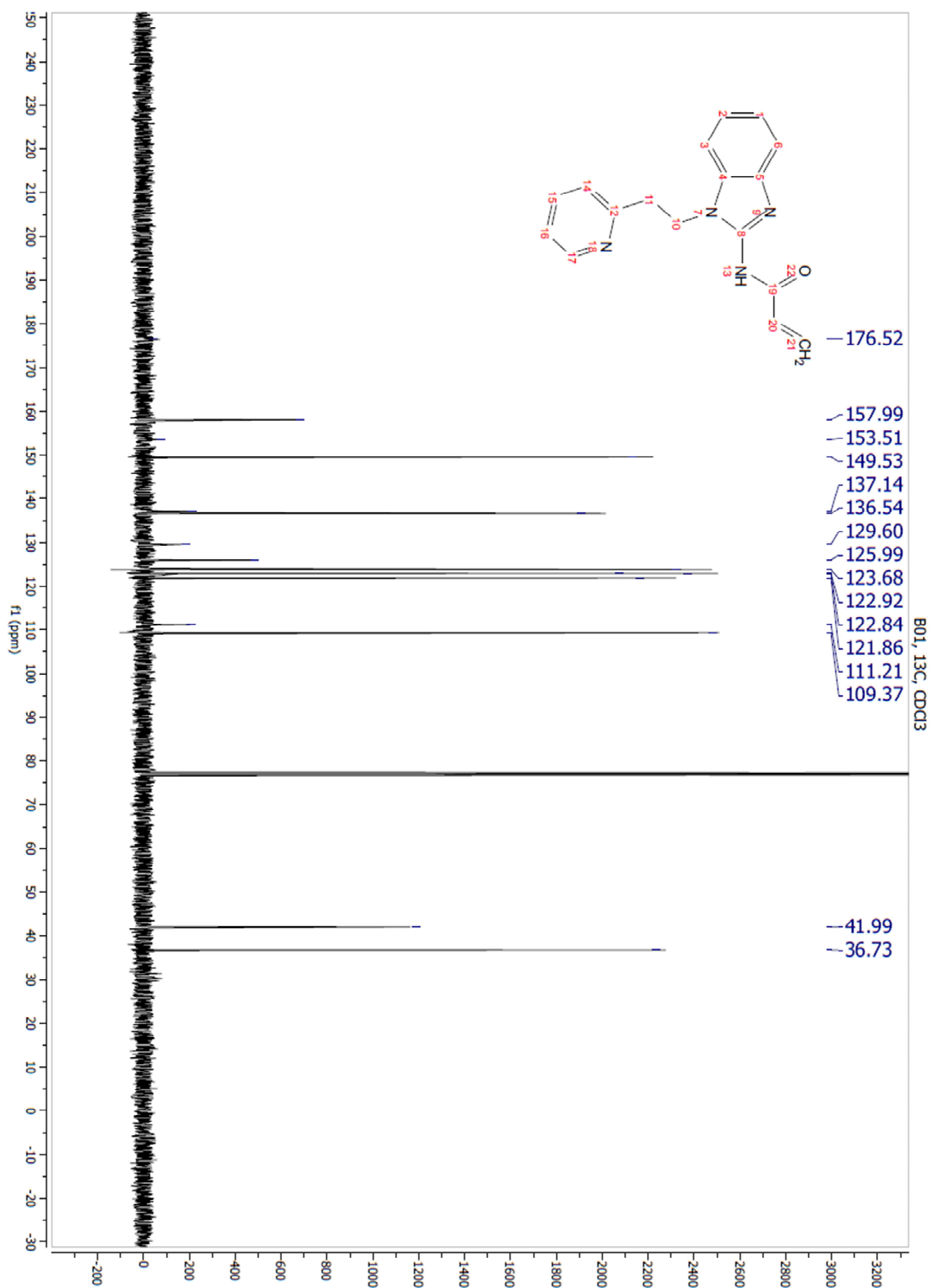
NMR spectra for A01 and B01



A01, ¹³C, CDCl₃







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