Electronic Supplementary Material (ESI) for RSC Medicinal Chemistry. This journal is © The Royal Society of Chemistry 2022

## **Supporting Information**

# Stereoisomeric Pam<sub>2</sub>CS Based TLR2 Agonists: Synthesis, Structural Modelling and Activity as Vaccine Adjuvants

Arshpreet Kaur,<sup>1</sup> Sakshi Piplani,<sup>2,3</sup> Deepender Kaushik,<sup>1</sup> Johnson Fung,<sup>2,3</sup> Isaac G. Sakala,<sup>2,3</sup> Yoshikazu Honda-Okubo,<sup>2,3</sup> Surinder K. Mehta,<sup>1</sup> Nikolai Petrovsky<sup>2,3,\*</sup> and Deepak B. Salunke<sup>1,4,\*</sup>

<sup>1</sup>Department of Chemistry and Centre for Advanced Studies, Panjab University, Chandigarh, India

<sup>2</sup>Vaxine Pty Ltd, Warradale, Australia

<sup>3</sup>College of Medicine and Public Health, Flinders University, Adelaide, Australia

<sup>4</sup>National Interdisciplinary Centre of Vaccines, Immunotherapeutics and Antimicrobials, Panjab University, Chandigarh, India

Email: <u>nikolai.petrovsky@flinders.edu.au;</u> <u>salunke@pu.ac.in</u>

\*Corresponding authors

### **Table of Contents**

S.No.	Content	Page No.
1	<sup>1</sup> H NMR of Compound <b>2</b>	<b>S</b> 3
2	<sup>1</sup> H NMR and <sup>13</sup> C NMR of Compound <b>3</b>	S4-S5
3	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>5a</b>	S6-S8
4	<sup>1</sup> H NMR, <sup>13</sup> C NMR and MS of Compound <b>5b</b>	S9-S11
5	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>5</b> c	S12-14
6	<sup>1</sup> H NMR, <sup>13</sup> C NMR and MS of Compound <b>6a</b>	S15-S17
7	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>6b</b>	S18-S20
8	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>6c</b>	S21-S23
9	<sup>1</sup> H NMR, <sup>13</sup> C NMR and MS of Compound <b>7a</b>	S24-S26
10	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>7b</b>	\$27-\$29
11	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>7</b> c	\$30-\$32
12	<sup>1</sup> H NMR, <sup>13</sup> C NMR, MS and HRMS of Compound <b>8a</b>	\$33-\$36
13	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>8b</b>	\$37-\$39
14	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>8c</b>	S40-S42
15	<sup>1</sup> H NMR, <sup>13</sup> C NMR, MS and HRMS of Compound <b>9a</b>	S43-S46
16	<sup>1</sup> H NMR, <sup>13</sup> C NMR, MS and HRMS of Compound <b>9b</b>	S47-S50
17	<sup>1</sup> H NMR, <sup>13</sup> C NMR and HRMS of Compound <b>9c</b>	\$51-\$53
18	Table S1 of Interacting residues of TLR2/6 with each ligand and H-bond distances	S54-S55
19	Surface representation of hTLR2/6 with the enantiopure compounds ( <b>7b</b> , <b>7b</b> , <b>8b</b> , <b>8c</b> , <b>PamCS</b> , <i>N</i> - <b>Ac-PamCS</b> ) and their interaction residues	S54-S63
20	Modellar Scripts for Docking Studies.	S64-S66
21	Table S2 to correlate all the available data, including computation, <i>in vitro</i> and <i>in vivo</i> data	S67



<sup>1</sup>**H NMR** of Compound **3** (in CDCl<sub>3</sub>, 400 MHz)



**S**4

ESI

# <sup>13</sup>C NMR of Compound **3** (in CDCl<sub>3</sub>, 100 MHz)



<sup>1</sup>**H NMR** of Compound **5a** (in CDCl<sub>3</sub>, 400 MHz)







## HRMS of Compound 5a



# <sup>1</sup>**H NMR** of Compound **5b** (in CDCl<sub>3</sub>, 400 MHz)



<sup>13</sup>C NMR of Compound **5b** (in CDCl<sub>3</sub>, 100 MHz)



## MS of Compound 5b



<sup>1</sup>**H NMR** of Compound **5c** (in CDCl<sub>3</sub>, 500 MHz)



# <sup>13</sup>C NMR of Compound **5**c (in CDCl<sub>3</sub>, 125 MHz)







x10 5 +ESI Scan (# 17-61, 45 Scans) Frag=118.0V KD-13.d Subtract (3)

# <sup>1</sup>**H NMR** of Compound **6a** (in CDCl<sub>3</sub>, 400 MHz)



# <sup>13</sup>C NMR of Compound **6a** (in CDCl<sub>3</sub>, 100 MHz)



## MS of Compound 6a



ESI

# <sup>1</sup>**H NMR** of Compound **6b** (in CDCl<sub>3</sub>, 400 MHz)



ESI

# <sup>13</sup>C NMR of Compound **6b** (in CDCl<sub>3</sub>, 100 MHz)



## HRMS of Compound 6b

Data:AK-2-78 Comment: Description: Ionization Mode:ESI+ History:Average(MS[1] 0.38..0.41) Acquired:4/26/2019 3:12:27 PM Operator:AccuTOF m/z Calibration File:20190422-TFANa\_... Created:4/29/2019 2:06:28 PM Created by:AccuTOF

Charge number:1 Tolerance:200.00[ppm], 250.00 ... 250.... Unsaturation Number:-100.5 ... 200.0 (... Element:<sup>12</sup>C:19 .. 19, <sup>1</sup>H:36 ... 37, <sup>14</sup>N:2 ... 2, <sup>23</sup>Na:0 ... 1, <sup>16</sup>O:8 ... 8, <sup>32</sup>S:1 ... 1



Mass	Intensity	Calc. Mass	Mass Difference [ppm]	Possible Formula			
453.22688	4645.38	453.22706	-0.40	12C <sub>19</sub> 1H <sub>37</sub> 14N <sub>2</sub> 16O <sub>8</sub> 32S <sub>1</sub>			
475.20929	156003.54	475.20901	0.60	$^{12}C_{19}^{1}H_{36}^{14}N_{2}^{23}Na_{1}^{16}O_{8}^{32}S_{1}$			

## <sup>1</sup>**H NMR** of Compound **6c** (in CDCl<sub>3</sub>, 500 MHz)



ESI

# <sup>13</sup>C NMR of Compound 6c (in CDCl<sub>3</sub>, 125 MHz)





## <sup>1</sup>**H NMR** of Compound **7a** (in CDCl<sub>3</sub>, 400 MHz)





## MS of Compound 7a



## <sup>1</sup>**H NMR** of Compound **7b** (in CDCl<sub>3</sub>, 400 MHz)



# <sup>13</sup>C NMR of Compound 7b (in CDCl<sub>3</sub>, 100 MHz)



## HRMS of Compound 7b

Data:AK-2-66 Comment: Description: Ionization Mode:ESI+ History:Average(MS[1] 0.40..0.42) Acquired:4/26/2019 3:24:45 PM Operator:AccuTOF m/z Calibration File:20190422-TFANa\_... Created:4/29/2019 2:05:13 PM Created by:AccuTOF

 Charge number:1
 Tolerance:200.00[ppm], 250.00 .. 250....
 Unsaturation Number:-100.5 .. 200.0 (...

 Element:<sup>12</sup>C:51 .. 51, <sup>1</sup>H:96 .. 97, <sup>14</sup>N:2 .. 2, <sup>23</sup>Na:0 .. 1, <sup>16</sup>O:10 .. 10, <sup>32</sup>S:1 .. 1
 1

Relative Intensity 100 -951.66656 NHBoc C<sub>15</sub>H<sub>31</sub> 0 C<sub>15</sub>H<sub>31</sub>、 OMe e ö O<sup>t</sup>Bu 50 Exact Mass: 928.6786 [M + H]<sup>+</sup>: 929.6858 [M + Na]<sup>+</sup> : 951.6678 967.65183 946.70129 0 920.0 940.0 960.0 980.0 m/z

Mass	Intensity	Calc. Mass	Mass Difference [ppm]	Possible Formula
951.66656	72038.76	951.66833	-1.86	${}^{12}C_{51}H_{96}H_{2}^{23}Na_{1}H_{010}^{32}S_{1}$

## <sup>1</sup>**H NMR** of Compound **7c** (in CDCl<sub>3</sub>, 500 MHz)



# <sup>13</sup>C NMR of Compound 7c (in CDCl<sub>3</sub>, 125 MHz)





## <sup>1</sup>**H NMR** of Compound **8a** (in CDCl<sub>3</sub>, 400 MHz)



# <sup>13</sup>C NMR of Compound 8a (in CDCl<sub>3</sub>, 100 MHz)



## MS of Compound 8a



ESI



Mass	Intensity	Calc. Mass	Mass Difference [ppm]	Possible Formula		
773.57052	33839.88	773.57136	-1.09	12C <sub>42</sub> 1H <sub>81</sub> 14N <sub>2</sub> 16O <sub>8</sub> 32S <sub>1</sub>		

<sup>1</sup>**H NMR** of Compound **8b** (in  $CDCl_3 + CD_3OD$ , 400 MHz)



# <sup>13</sup>C NMR of Compound **8b** (in $CDCl_3 + CD_3OD$ , 100 MHz)



## HRMS of Compound 8b



<sup>1</sup>**H NMR** of Compound **8c** (in CDCl<sub>3</sub>, 500 MHz)





## HRMS of Compound 8c



## <sup>1</sup>H NMR of Compound 9a (in CDCl<sub>3</sub>, 400 MHz)



ESI

<sup>13</sup>C NMR of Compound 9a (in CDCl<sub>3</sub>, 100 MHz)



## MS of Compound 9a



ESI

## HRMS of Compound 9a

Data:AK-2-45 Comment: Description: Ionization Mode:ESI+ History:Average(MS[1] 0.35..0.38) Acquired:4/26/2019 4:13:52 PM Operator:AccuTOF m/z Calibration File:20190422-TFANa\_... Created:4/29/2019 2:14:28 PM Created by:AccuTOF

Charge number:1 Tolerance:200.00[ppm], 250.00 .. 250.... Unsaturation Number:-100.5 .. 200.0 (... Element:<sup>12</sup>C:44 .. 44, <sup>1</sup>H:0 .. 83, <sup>14</sup>N:2 .. 2, <sup>23</sup>Na:0 .. 1, <sup>16</sup>O:9 .. 9, <sup>32</sup>S:1 .. 1



S46
-----

# <sup>1</sup>**H NMR** of Compound **9b** (in CDCl<sub>3</sub>, 400 MHz)



ESI

# <sup>13</sup>C NMR of Compound 9b (in CDCl<sub>3</sub>, 100 MHz)



## MS of Compound 9b



## HRMS of Compound 9b

Data:AK-2-73 Comment: Description: Ionization Mode:ESI+ History:Average(MS[1] 0.36..0.38) Acquired:4/26/2019 3:37:12 PM Operator:AccuTOF m/z Calibration File:20190422-TFANa\_... Created:4/29/2019 2:12:09 PM Created by:AccuTOF

Charge number:1 Tolerance:200.00[ppm], 250.00 .. 250.... Unsaturation Number:-100.5 .. 200.0 (... Element:<sup>12</sup>C:44 .. 44, <sup>1</sup>H:0 .. 83, <sup>14</sup>N:2 .. 2, <sup>23</sup>Na:0 .. 1, <sup>16</sup>O:9 .. 9, <sup>32</sup>S:1 .. 1



<sup>1</sup>H NMR of Compound 9c (in CDCl<sub>3</sub>, 500 MHz)



# <sup>13</sup>C NMR of Compound 9c (in CDCl<sub>3</sub>, 125MHz)





### Table S1. List of interacting residues of TLR2/6 heterodimer with compounds and H-bond

### distances.

Compound	MMPBSA_Score (Free energy binding)	Interacting Residue	H-bond residue	H-bond distance	
7b	-20.42	ChainA: Leu240, Phe258, Leu263, Phe269, Leu286, Ile288, Leu291, Ile293, Phe299, Tyr300, Asp301, Leu302, Leu305, Tyr306, Thr309, Ile315, Val317, Ser320, Lys321, Val322, Phe323, Leu324, Pro326, Leu329 ChainB: Phe317, Phe319, Ser320, Gln321	Gln321(O6- NE2)	3.16Å	
7c	-25.78	ChainA: Leu263, Phe269, Leu286, Ile288, Leu291, Ile293, Phe296, Tyr297, Phe299, Tyr300, Asp301, Leu302, Leu305, Tyr306, Thr309, Ile315, Val317, Ser320, Lys321, Val322, Phe323, Leu324, Val325, Pro326, Leu329 ChainB: Phe317, Leu318, Phe319, Ser320, Gln321	Phe323(O8- N) Leu324(O8- N)	3.21Å 3.25Å	
8b	-36.52	ChainA: Ile235, Leu240, Phe258, Leu263, Phe269, Leu286, Ile288, Leu291, Ile293, Phe296, Tyr297, Phe299, Leu305, Thr309, Ile315, Val317, Ser320, Lys321, Val322, Phe323, Leu324, Pro326, Leu329 ChainB: Phe317, Phe319, Ser320, Gln321	Phe296(N2- O) Tyr297(O5-O) Phe299(N2- N) Phe323(O4- N) Ser320(O5- OG) Gln321(O5- N) Gln(O5-NE2)	3.09Å 2.97Å 3.07Å 3.23Å 3.29Å 3.01Å 2.89Å	
8c	-32.10	ChainA: Phe258, Leu286, Ile288, Ile293, Phe296, Tyr297, Leu298, Phe299, Asp301 Leu302, Leu305, Tyr306, Thr308, Ile315, Val317, Lys321, Val322, Phe323, Leu324, Val325, Pro326, Leu329, Leu341 ChainB: Val316, Gln321	Phe296(O8- O) Phe299(O8- O) Phe323(O4- N) Leu324(O4- N)	3.28Å 3.05Å 3.00Å 3.25Å	

9b	-34.22	ChainA: Leu240, Phe258, Phe269, Leu286, Ile288, Leu291, Ile293, Tyr297, Phe299, Asp301, Leu302, Leu305, Tyr306, Leu308, Thr309, Ile315, Val317, Ser320, Val322, Phe323, Leu324, Val325, Pro326, Leu329, Leu341 ChainB: Phe317, Gln321	Phe317(O5- O) Gln321(O4- NE2)	3.09Å 2.96Å
9c	-37.91	ChainA: Ile235, Leu240, Leu263, Phe269, Leu286, Ile288, Leu291, Tyr297, Leu302, Leu305, Thr309, Ile315, Val317, Lys321, Val322, Phe323, Leu324, Val325, Pro326, Leu329 ChainB: Val316, Phe317, Ser320, Thr322, Gln321	Tyr297(O8-O) Phe323(O7- N) Leu324(O7- N) Ser320(O8- OG) Gln321(O8- N) Thr322(O8-N) Thr322(O8- OG1)	3.02Å 3.10Å 3.03Å 3.12Å 2.99Å 3.34Å 3.14Å

Figure S1. Ramachandran Plot of human TLR2/6.





Figure S2. Surface representation of hTLR2/6 with compound 7b

Figure S2a. Interacting amino acid residues of hTLR2/6 with compounds 7b





### Figure S3. Surface representation of hTLR2/6 with compound 7c

Figure S3a. Interacting amino acid residues of hTLR2/6 with compounds 7c





Figure S4. Surface representation of hTLR2/6 with compound 8b

Figure S4a. Interacting amino acid residues of hTLR2/6 with compounds 8b





Figure S5. Surface representation of hTLR2/6 with compound 8c

Figure S5a. Interacting amino acid residues of hTLR2/6 with compounds 8c





Figure S6. Surface representation of hTLR2/6 with compound 9b

Figure S6a. Interacting amino acid residues of hTLR2/6 with compounds 9b





### Figure S7. Surface representation of hTLR2/6 with compound 9c

Figure S7a. Interacting amino acid residues of hTLR2/6 with compounds 9c



Figure S8. Modelled structure of human TLR2/6 showing the position of the ligand binding pocket



Figure S9. Superimposition of the structures of compounds 8b and 8c in hTLR2/6



### **Modeller Scripts for Docking Studies:**

### Script1:

from modeller import \*

log.verbose()
env = environ()
#-- Prepare the input files

#-- Read in the target sequence/alignment aln = alignment(env) aln.append(file='tlr2-6.ali', alignment\_format='PIR', align\_codes='ALL')

#-- Convert the input sequence/alignment into
# profile format
prf = aln.to\_profile()

#-- Scan sequence database to pick up homologous sequences
prf.build(sdb, matrix\_offset=-450, rr\_file='\${LIB}/blosum62.sim.mat',
 gap\_penalties\_1d=(-500, -50), n\_prof\_iterations=1,
 check\_profile=False, max\_aln\_evalue=0.01)

#-- Write out the profile in text format
prf.write(file='build\_profile.prf', profile\_format='TEXT')

#-- Convert the profile back to alignment format
aln = prf.to\_alignment()

#-- Write out the alignment file
aln.write(file='build\_profile.ali', alignment\_format='PIR')

### Script2:

from modeller import \*

```
env = environ()
aln = alignment(env)
for (pdb, chain) in (('2z7x', 'A'), ('3a79', 'A')):
    m = model(env, file=pdb, model_segment=('FIRST:'+chain, 'LAST:'+chain))
    aln.append_model(m, atom_files=pdb, align_codes=pdb+chain)
aln.malign()
aln.malign3d()
aln.compare_structures()
aln.id_table(matrix_file='family.mat')
env.dendrogram(matrix_file='family.mat', cluster_cut=-1.0)
```

### Script3:

from modeller import \*

```
env = environ()
aln = alignment(env)
mdl = model(env, file='3a79', model_segment=('FIRST:A','LAST:B'))
aln.append_model(mdl, align_codes='3a79', atom_files='3a79.pdb')
aln.append(file='tlr2-6.ali', align_codes='tlr2-6')
aln.align2d()
aln.write(file='tlr-1.ali', alignment_format='PIR')
aln.write(file='tlr-1.pap', alignment_format='PAP')
```

### Script4:

from modeller import \*
from modeller.automodel import \*
#from modeller import soap\_protein\_od

```
env = environ()

a = automodel(env, alnfile='tlr2-6.ali',

knowns='3a79', sequence='tlr2-6',

assess_methods=(assess.DOPE,

#soap_protein_od.Scorer(),

assess.GA341))

a.starting_model = 1

a.ending_model = 10

a.make()
```

### **Autodock Vina Scripts:**

### Conf.txt:

receptor = tlr2\_6model.pdbqt

center\_x = 138.161 center\_y = 109.587 center\_z = 33.389 size\_x = 40 size\_y = 80 size\_z = 60 num\_modes = 10

### Vina\_screen\_local.sh:

#! /bin/bash

```
for f in *.pdbqt; do
    b=`basename $f .pdbqt`
    echo Processing ligand $b
    mkdir -p $b
    vina --config conf.txt --ligand $f --out ${b}/out.pdbqt --log ${b}/log.txt
done
```

Table S2 to correlate all the available data, including computation, in vitro and in vivo data

Compound No.	Docking Score (MMPBSA)	TLR2/6 activity	Cytokine Levels		MHC-II+ CD19+ cells (B cells) †		MHC-II+ CD11c+ cells (DCs)†		MHC-II <sup>+</sup> CD11b <sup>+</sup> cells (Monocytes) <sup>†</sup>		Protection against influenza challenge <sup>‡</sup>	
	Free energy binding	ЕС <sub>50</sub> (µМ)	IL-6	IL-10	TNF-α	CD40	CD86	CD40	CD86	CD40	CD86	
Pam <sub>2</sub> CSK <sub>4</sub>		0.0003	+	++	+	+++	+++	+++	+++	+++	+++	-
Pam <sub>3</sub> CSK <sub>4</sub>		0.0193	++	+++	++	++	++	++	++	++	++	-
7a		5.63	-	-	-	vvv	+++	vvv	+++	vvv	+++	NT
7b	-20.42	5.86	-	-	-	vvv	+++	vvv	++	vvv	+++	NT
7c	-25.78	4.27	-	-	-	+vv	+++	+vv	+++	+vv	vvv	NT
8a		0.0010	+	+	+	vvv	+++	vvv	+++	vvv	++V	++
8b	-36.52	0.004	+	+	+	+++	+++	+++	+++	+++	++V	+++
8c	-32.10	0.1337	-	-	-	+++	+++	+++	+++	+++	V+V	-
9a		0.0050	+	++	+	+++	V++	+++	V++	+++	VV+	+
9b	-34.22	0.0008	++	+++	++	vvv	+++	+vv	+++	++V	+vv	+++
9c	-37.91	0.2518	+	+	+	V++	+++	V++	+++	+++	vv+	-
<sup>†</sup> CD expressions seen for three doses; 10.0, 1.0, 0.1 μg/mL; + for upregulation; ∨ for down regulation;												

<sup>‡</sup>Cumulative weight loss; +++ stands for no weight loss; - stands for weight loss equivalent to control group (saline); NT is not tested