Legends to supplementary figures

Figure S1: The average number of molecules of PO-PEG-COOH per nanocrystal was measured by Fourier Transform Infrared Spectroscopy. Infrared spectra in KBr pellets of various proportions of PO-PEG-COOH mixed with a constant amount of USPIONs were recorded. The normalized 1110 cm⁻¹ band was used for the establishment of the calibration curve and the average number of PO-PEG-COOH per nanoparticle was deduced from this curve and evaluated to 176 ± 16 .

Figure S2: Calibration curve using gastrin peptide by OPA assay.

Figure S3: a) CCK2R expression in CAF cell lines. Two immortalized human cancerassociated fibroblasts (CAF) cell lines, CAF1 and CAF2, were permanently transfected with an expression vector encoding the human CCK2R. 2 clones expressing the CCK2R were selected and termed respectively CAF1-CCK2 and CAF2-CCK2. Cells were incubated with 10 nM DY647-CCK9 agonist in presence or in absence of 10 μ M GV 150013X, a specific CCK2R antagonist. Cell-associated fluorescence was determined by flow cytometry, revealing the expression of CCK2R on CAF cell lines. b) USPION@Gastrin uptake depends on CCK2R expression. Cells were incubated with increased concentrations of USPION@Gastrin in presence or in absence of 10 μ M Gastrin-17, a specific CCK2R agonist, for 24 h. Cell-associated fluorescence was measured by flow cytometry. Results are expressed as fluorescence associated with the cells and are the mean \pm SEM of at least three separate experiments. **Figure S4:** Analysis of CAFs death upon 40 mT RMF exposure. CAF1-CCK2 cells were incubated for 24 h with increased concentration of USPION@Gastrin (4, 8 and 16 μ g magnetic Fe / ml), washed and exposed to RMF for 2 h. Different frequencies (0.2, 1, 10, 20 Hz) were assayed. Dead cells were labeled with AnnV/PI, 4h after RMF exposure, and counted by flow cytometry. Results are expressed as mean \pm SEM of at least three separate experiments. Statistical analysis was performed using two-way ANOVA test.

Figure S5: Analysis of CAFs death upon 20 mT RMF exposure. CAF1-CCK2 cells were incubated for 24 h with increased concentration of USPION@Gastrin (4, 8 and 16 μ g magnetic Fe / ml), washed and exposed to RMF for 2 h. Different frequencies (1, 10, 20 Hz) were assayed. Dead cells were labeled with AnnV/PI, 4h after RMF exposure, and counted by flow cytometry. Results are expressed as mean \pm SEM of at least three separate experiments. Statistical analysis was performed using two-way ANOVA test.

Figure S6: Analysis of CAFs death upon 60 mT / 1 Hz RMF exposure. CAF1-CCK2 cells were incubated for 24 h with increased concentration of USPION@Gastrin (4, 8 and 16 μ g magnetic Fe / ml), washed and exposed to RMF for 2 h. Dead cells were labeled with AnnV/PI, 4h after RMF exposure, and counted by flow cytometry. Results are expressed as mean ± SEM of at least three separate experiments. Statistical analysis was performed using two-way ANOVA test.

Figure S7: Analysis of CAFs death upon 200 mT RMF exposure. CAF1-CCK2 cells were incubated for 24 h with increased concentration of USPION@Gastrin (4, 8 and 16 µg magnetic Fe

/ ml), washed and exposed to RMF for 2 h. Different frequencies (1, 10, 20 Hz) were assayed. Dead cells were labeled with AnnV/PI, 4h after RMF exposure, and counted by flow cytometry. Results are expressed as mean \pm SEM of at least three separate experiments. Statistical analysis was performed using two-way ANOVA test.

Figure S8: Analysis of CAFs death upon 380 mT RMF exposure. CAF1-CCK2 cells were incubated for 24 h with increased concentration of USPION@Gastrin (4, 8 and 16 μ g magnetic Fe / ml), washed and exposed to RMF for 2 h. Different frequencies (1, 10, 20 Hz) were assayed. Dead cells were labeled with AnnV/PI, 4h after RMF exposure, and counted by flow cytometry. Results are expressed as mean \pm SEM of at least three separate experiments. Statistical analysis was performed using two-way ANOVA test.



Figure S1



Figure S2







Figure S4



Figure S5



Figure S6



Figure S7



Figure S8