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Supporting Information

CeO₂-nanocubes as efficient and selective catalysts for the hydroboration of carbonyl groups

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I. General Information

All reagents were purchased from TCI, SDFCL or Aldrich and were used as received. The ceria precursor Ce(NO₃)₃·6H₂O was purchased from TCI. C₆D₆ and CDCl₃ were purchased from Cambridge isotope laboratories and were dried using molecular sieves and deoxygenated using the freeze-pump-thaw method. HBpin for bulk reactions was prepared from B₂pin₂ according to literature procedures.^{1a} CeO₂ nanoparticles were prepared according to the literature procedure.^{1b} Commercially available, pre-coated TLC-sheets ALUGRAM[®] Xtra Sil G/UV₂₅₄ was purchased from MACHEREY-NAGEL GmbH & Co. KG. The removal of solvent was performed on a rotary evaporator *in vacuo* at a maximum temperature of 40 °C.

All NMR spectra were recorded at ambient temperature using a Bruker Advance 400 NMR spectrometer (¹H, 400 MHz; ¹³C, 100 MHz; ¹¹B, 128 MHz). ¹H NMR chemical shifts are reported relative to TMS and were referenced *via* residual proton resonances of the corresponding deuterated solvent (CDCl₃: 7.26 ppm, C₆D₆: 7.16 ppm) whereas ¹³C NMR spectra are reported relative to TMS using the carbon signals of the deuterated solvent (CDCl₃: 77.16 ppm, C₆D₆: 128.06 ppm). ¹¹B NMR signals were quoted relative to BF₃·Et₂O and ¹⁹F NMR signals were quoted using FCCl₃ as an internal standard. All ¹¹B and ¹³C NMR spectra were broad-band ¹H decoupled. The IR spectra were obtained with a BRUKER ALPHA spectrometer in the range of 400 to 4000 cm⁻¹ using KBr windows. GC-MS data were acquired using SHIMADZU GCMS QP 2010SE system.

The microstructure of the CeO₂ NPs was studied by Rigaku Ultima IV powder X-ray diffractometer using Cu K α radiation (scan rate of 3° min⁻¹). Scanning electron microscopy (SEM) and Energy-dispersive X-ray spectroscopic (EDX) spectra were performed on a JSM 7100F JEOL FESEM with EDS. TEM was carried out using FEI Tecnai T20 transmission electron microscope operating at 200 kV after casting a drop of nanoparticle dispersion in isopropyl alcohol over Cu grid.

II. Experimental Details

Synthesis of CeO₂ nanoparticles.^{1b} To obtain CeO₂ nano-cubes, a hydrothermal process was employed. Ce(NO₃)₃·6H₂O (4 mmol) and NaOH (480 mmol) were dissolved in 10 mL and 70 mL of distilled water, respectively. Then both the solution was mixed and stirred for 30 min at room temperature to get purple slurry. Subsequently, the slurry was transferred into a 100 mL Teflon-lined stainless autoclave and heated at 180 °C for 24 h to obtain nano-cubes. After the hydrothermal treatment, the precipitates were separated by filtration, washed with distilled water and ethanol several times. After drying at 80 °C for 1 day, the products were calcined at 600 °C for 5 h to get a yellow product. The CeO₂ nanoparticles were characterised by using field emission scanning electron microscope (FE-SEM), transmission electron microscopy (TEM), energy dispersive X-ray spectroscopy (EDX) and powder XRD (X-ray diffraction).



Figure S1. FESEM image of CeO₂ nanoparticles.





Figure S2. EDX spectrum and overlay map of CeO_2 nanoparticles.

TEM images and particle size distributions of CeO2 NPs

10 5

> 0 L 0

20

40

60

80



c)

Particle siz (nm)

100 120 140

160

180

200

Figure S3. a) TEM images of CeO_2 nanoparticles. b) Particle size distribution chart of CeO_2 nanoparticles.

The TEM analysis of CeO₂ nanoparticles (Figure S3) shows that the nanoparticles were having particle size in the range of 20-200 nm and an average particle size of \sim 55 nm.



Figure S4. Powder XRD pattern of CeO₂ nanoparticles.



Figure S5. BET analysis of CeO₂ nanoparticles.

BET analysis results based on the N_2 adsorption desorption isotherms, depicted in Figure S5. It was seen that of CeO₂ nanoparticles possesses a BET surface area of 23 m² g⁻¹.

Experimental procedures for examples described in Table 1.

In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, CeO₂-catalyst (0.025 mmol), HBpin (0.33 mmol), benzaldehyde (0.25 mmol) and solvent (1 mL) were added. The reaction mixture was stirred at room temperature for the indicated amount of time, then diluted with Et₂O (2 mL) and filtered through a plug of celite (\emptyset 3 mm × 8 mm). The solvents were removed *in vacuo*, and nitromethane was added as an internal standard.

	O H + HBpin R H 1a	<u>CeO₂ NPs (</u> MTBE (1ml RT, 24 h	(10 mol%) -)		3pin `H b
Entry	Catalyst (mol %)	Time (h)	HBpin (equiv)	Solvent (1 mL)	Yield $(\%)^b$
1	CeO ₂ NPs (10)	24	1.0	MTBE	72
2	CeO ₂ NPs (10)	24	1.1	MTBE	86
3	CeO ₂ NPs (10)	24	1.2	MTBE	97
4	CeO ₂ NPs (10)	24	1.3	MTBE	>99

Table S1: Effect of different equivalents of HBpin on the CeO₂ nanoparticles catalysed hydroboration of benzaldehyde (1a).^{*a*}

^{*a*} Reaction conditions: benzaldehyde (0.25 mmol), CeO₂ NPs (10 mol %, 4 mg), HBpin, MTBE (1 mL), at room temperature for 24 h. ^{*b*} Yields were determined by ¹H NMR, using nitromethane as an internal standard.

General procedure of hydroboration of aldehydes (examples described in Table 2).

In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.0250 mmol), HBpin (1.3 equiv, 47 μ L, 0.33 mmol), MTBE (1 mL) and aldehyde (0.25 mmol) were added and the reaction was stirred vigorously at room temperature for 24 h. The reaction mixture was then diluted with Et₂O (2 mL) and filtered

through a plug of celite (\emptyset 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard.

General procedure of hydroboration of ketones (examples described in Table 3).

In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.0250 mmol), HBpin (1.3 equiv, 47 μ L, 0.33 mmol), MTBE (1 mL) and ketone (0.25 mmol) were added and the reaction mixture was stirred vigorously at 50 °C for 24 h. The reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (Ø 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard.

Experimental procedures for catalytic conversion of aldehydes/ketones to $1^{\circ}/2^{\circ}$ alcohols: General procedure A. In a 50 mL round bottom flask equipped with a magnetic stirring bar, CeO₂ (10 mol %, 0.1 mmol, 17 mg), HBpin (1.3 mmol, 188 µL), MTBE (4 mL) and aldehyde (1.0 mmol) were added. The resulting reaction mixture was stirred vigorously at room temperature for 24 h (for ketone 50 °C, 24 h). After completion of the reaction, it was diluted with diethyl ether (10 mL) and filtered through a plug of celite (\emptyset 3 mm × 8 mm). The solvents were removed *in vacuo*, and then methanol (5 mL) and 1N HCl (1 mL) were added to the reaction mixture and was refluxed at 50 °C for 1 h (for ketone 3 h). The organic layer was extracted with dichloromethane (3 x 10 mL) and was removed by vacuum to get the crude product. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate as eluent and characterized by ¹H and ¹³C NMR spectroscopy.

General procedure B. The resulted borate ester residue was hydrolysed with silica gel at 50 °C for 2-4 h. Then the organic layer was extracted with dichloromethane (3 x 10 mL) and the

volatiles were removed under *vacuo* and the residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate as eluent.

General procedure C. The resulted borate ester reaction mixture was exposed to the air and was stirred at room temperature for 14-16 h, for the quantitative conversion to corresponding alcohol. The organic layer was extracted with dichloromethane (3 x 10 mL) and was removed by vacuum to get the crude product. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate as eluent and characterized by ¹H and ¹³C NMR spectroscopy.

General procedure for preparative scale reactions.

In a 100 mL round bottom flask equipped with a magnetic stirring bar, CeO₂ (10 mol %, 0.5 mmol, 86 mg), HBpin (1.3 equiv, 6.5 mmol, 0.9 mL), MTBE (10 mL) and aldehyde (5.0 mmol) were added. The resulting reaction mixture was stirred vigorously at room temperature for 24 h. After completion of the reaction, it was diluted with diethyl ether (50 mL) and filtered through a plug of celite. The solvents were removed *in vacuo*, and then methanol (10 mL) and 1N HCl (2 mL) were added to the reaction mixture and was refluxed at 50 °C for 1 h. The organic layer was extracted with dichloromethane (3 x 20 mL) and was removed by vacuum to get the crude compound. The crude residue was purified by silica gel column chromatography using a mixture of hexane and ethyl acetate as eluent and characterized by ¹H and ¹³C NMR spectroscopy.

The hydroboration of 4-bromobenzaldehyde (9a, 0.925 gm, 5 mmol), 2methylbenzaldehyde (13a, 0.6 gm, 5 mmol) and 1-naphthaldehyde (17a, 0.781 gm, 5 mmol) gave the desired alcohols in good yields (9c: 0.795 g, 86%; 13c: 0.452 g, 75% and 17c: 0.492 g, 63%).

R	`H ⁺ HBpin <mark>CeO₂(1</mark> MTBE.	0 mol %)	O ^{BPin} R H	Method A, OH B or C \rightarrow H
а	(1.3 equiv)	·	L J b	с
Entry	Aldehyde		Borate ester	Yield (%) ^b (1 ^o alcohol (%)) ^c
1	O H		OBPin H	1b: >99 ^b 1c: (89) ^c
2	O H	, C	OBPin H	2b : >99 ^b 2c: (81) ^c
3	MeO H	МеО	OBPin H	3b : >99 ^b 3c : 96 ^b (90) ^c
4	O H		OBPin H	4b : 93 ^b 4c: 89 ^b (79) ^c
5	OMe O HO HO	но	Me OBPin H	5b : 84 ^b
6	Me ₂ N	Me ₂ N	H	6b : 96 ^b
7	F H	F	OBPin	7b: >99 ^b 7c: (85) ^c
8	CI	cı	OBPin H	8b : >99 ^b 8c: 98 ^b (87) ^c
9	Br	Br	OBPin H	9b : >99 ^b 9c: (82) ^c
10	O H Br	Br	OBPin H	10b : >99 ^b 10c: 96 ^b (78) ^c
11	O O ₂ N H	O ₂ N	OBPin H	11b : 86 ^b 11c: 84 ^b (70) ^c
12	NC H	NC	OBPin H	12b : >99 ^b 12c: (80) ^c
13	O H		OBPin H	13b : >99 ^b 13c: (84) ^c

Table S2a. Hydroboration of aldehydes catalysed by CeO₂ NPs.^a

^{*a*} Reaction conditions: Aldehyde (1.0 mmol), CeO₂ NPs (10 mol %, 0.1 mmol, 17 mg), HBpin (1.3 mmol, 188 μ L) and MTBE (2 mL), at room temperature for 24 h unless otherwise stated. ^{*b*} The yields were determined by ¹H NMR analysis using nitromethane as an internal standard. ^{*c*} Isolated yield after chromatographic workup.

R H	+ HBpin <mark>CeO₂(10 mol %</mark> MTBE, RT, 24	(⁶) h R H Metho B or C	d A, OH → R H
а	(1.3 equiv)	b	с
Entry	Aldehyde	Borate ester	Yield (%) ^b ° alcohol (%)) ^c
14	O CI	OBPin H Cl	14b : >99 ^b
15	O H Br	OBPin H Br	15b : >99 ^b
16	CI CI	OBPin H CI	16b : >99 ^b
17	H_O	H_OBPin	17b: >99 ^b 17c: (88) ^c
18	O H	OBPin H	18b : 74 ^b 18c: 72 ^b (59) ^c
19	И О О О	H OBPin	19b : 73 ^b
20	H H	OBPin OBPin H	20b : ^{<i>d</i>} 88 ^{<i>b</i>} 20c: (73) ^{<i>c</i>}
21	ОЦН	OBPin H	21b : >99 ^b
22	O H	OBPin H	22b : 85 ^b 22c: (80) ^c

Table S2b. Hydroboration of aldehydes catalysed by CeO₂ NPs.^a

^{*a*} Reaction conditions: Aldehyde (1.0 mmol), CeO₂ NPs (10 mol %, 0.1 mmol, 17.2 mg), HBpin (1.3 mmol, 188μL) and MTBE (2 mL), at room temperature for 24 h unless otherwise stated. ^b The yields were determined by ¹H NMR analysis using nitromethane as an internal standard. ^{*c*} Isolated yield after chromatographic workup. ^{*d*} Reaction was performed using 2.6. equiv of HBpin.

o	+ HPDin CeO ₂	(10 mol %) O ^{-BPir}	Method A, OH B or C I
R		E, 50 °C R R R ¹	R^{1}
а	(1.3 equiv)24 h	b	c
Entry	Ketone	Borate ester (2 ^c	Yield (%) ^b alcohol yield (%)) ^c
	Q	QBPin	
23			23b: >98 ^b 23c: (80) ^c
	° °	OBPin	()
24			24b :85 ^b 24c : 83 ^b (72) ^c
	Q Q	OBPin	240.03 (72)
25		MeO	25b :87 ^b 25c : (78) ^c
, in the second s	0	OBPin	
26			26b: >99⁵ 26c: (89) ^c
		OBPin	
27			27b :>99 ^b
F	ç v	F OBPin	
28	\square		28b :>99 ^b
(CI	L.
	Ň		29b :97 ^{<i>b</i>} 29c : 94 ^{<i>b</i>} (75) ^{<i>c</i>}
29	F	F CPBin	
			30b : 93 ^b
30	Br	Br OBPin	30C: 87° (71)°
			31b: 53 ^b 31c: (41) ^c
31	NO ₂	NO ₂	010.(41)
		OBPin	32b: 72 ^b
32			32c : (58) ^c
	0	OBPin	33b : 61 ^b
33	Br	Br	33c : 60 ^b (37) ^c
	0	OBPin	
34	Ň		34b :72 ^b 34c : 59 ^b (56) ^c
	OMe	OMe	
35	(S) O	OBPin	35b: 58 ^b
26	n n f f o	OBPin	36b: 84 ^b
30	\mathbb{M}		27h.64 ^h
37	O		3/0:01~
38	S S O	SOBPin	38b :86 ^b

Table S3. Hydroboration of ketones catalysed by CeO₂ NPs.^{*a*}

^{*a*} Reaction conditions: ketone (1.0 mmol), CeO₂ NPs (10 mol %, 0.01 mmol, 17.2 mg), HBpin (1.3 mmol, 188 μ L) and MTBE (2 mL), at 50 °C for 24 h unless otherwise stated. ^{*b*} The yields were determined by ¹H NMR analysis using nitromethane as an internal standard. ^{*c*} Isolated yield after chromatographic workup.

Table S4. Solvent effect on the CeO₂ nanoparticle catalysed hydroboration of 4-bromobenzaldehyde (7a).^{*a*}

	Br	H + HBpin	CeO ₂ NPs (10 RT	Br	OBpin H 9b	
Entry	Catalyst (mol%)	Temperature (°C)	Time (h)	HBpin (equiv)	Solvent (1 mL)	Product Yield (%) ^b
1	$CeO_2(10)$	RT	24	1.3	MTBE	>99
2	$CeO_2(10)$	RT	24	1.3	-	83
3	-	RT	24	1.3	-	15

^{*a*} Reactions were carried out using 0.25 mmol of **9a** (1 equiv), 1.3 equiv of HBpin (0.33 mmol) in 1 mL of MTBE at room temperature unless otherwise stated. ^{*b*} Yield was determined by ¹H NMR analysis, using nitromethane as an internal standard.

Table S5. Solvent effect on the CeO₂ nanoparticle catalysed hydroboration of acetophenone (**23a**).^{*a*} $_{OBpin}^{OBpin}$

		+ HBpir	CeO ₂ NPs (10 50 °C		OBpin 23b	
Entry	Catalyst	Temperature	Time	HBpin	Solvent	Product Yield
	(mol%)	(°C)	(h)	(equiv)	(1 mL)	$(\%)^{b}$
1	$CeO_2 NPs (10)$	50 °C	24	1.3	MTBE	98
2	CeO ₂ NPs (10)	50 °C	24	1.3	-	74

^{*a*} Reactions were carried out using 0.25 mmol of **23a** (1 equiv), 1.3 equiv of HBpin (0.33 mmol) in 1 mL of solvent at room temperature unless otherwise stated. ^{*b*} Yields were determined by ¹H NMR analysis, using nitromethane as an internal standard.

III. Spectroscopic Data of Hydroboration Products and Related Alcohols

2-(Benzyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 1b).^{2,3}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.41-7.28 (m, 5H), 4.97 (s, 2H), 1.31 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.2.

Phenylmethanol (1c).^{2,4}



Following General procedure A. Colourless oil. Yield: 89% (96 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.26-7.19 (m, 5H), 4.46 (s, 2H), 3.55 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 140.8, 128.3, 127.4, 127.0, 79.37, 64.6.

4,4,5,5-Tetramethyl-2-((4-methylbenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 2b).^{2,3}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.28 (d, *J* = 8 Hz, 2H), 7.18 (d, *J* = 8 Hz, 2H), 4.92 (s, 2H), 2.37 (s, 3H), 1.30 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1.

p-Tolylmethanol (2c).^{2,4}



Following General procedure A. Colourless oil. Yield: 81% (99 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.27 (d, *J* = 8 Hz, 2H), 7.19 (d, *J* = 8 Hz, 2H), 4.65 (s, 2H), 2.38 (s, 3H), 1.89 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 138.0, 137.4, 129.3, 127.2, 65.3, 21.2.

2-((4-Methoxybenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 3b).^{2,3}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.30 (d, *J* = 8 Hz, 2H), 6.89 (d, *J* = 8 Hz, 2H), 4.87 (s, 2H), 3.82 (s, 3H), 1.29 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

(4-Methoxyphenyl)methanol (3c).^{2,5}



Following General procedure A. Colourless oil. Yield: 90% (125 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.24 (d, *J* = 8 Hz, 2H), 6.87 (d, *J* = 8 Hz, 2H), 4.52 (s, 2H), 3.78 (s, 3H), 3.47 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 158.8, 133.1, 128.5, 113.7, 64.3, 55.1.

2-((3-Methoxybenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 4b).^{2,6}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.26 (br, 1H), 6.95–6.84 (m, 3H), 4.94 (s, 2H), 3.82 (s, 3H), 1.30 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

(3-Methoxyphenyl)methanol (4c).^{2,7}



Following General procedure A. A colorless oil. Yield: 79% (108 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.29-7.25 (m, 1H), 6.93-6.91 (m, 2H), 6.85-6.82 (m, 1H), 4.59 (s, 2H), 3.79 (s, 3H), 3.37 (s, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 159.6, 142.6, 129.4, 119.0, 112.9, 112.1, 64.6, 55.0.

4-(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)phenol (Table 2, 5b).^{2,6}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.20 (d, *J* = 8 Hz, 2H), 6.81 (d, *J* = 8 Hz, 2H), 4.84 (s, 2H), 1.28 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.2.





(**Table 2, 6b**).⁸ ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.25 (d, *J* = 8 Hz, 2H), 6.74 (d, *J* = 8 Hz, 2H), 4.85 (s, 2H), 2.97 (s, 6H), 1.30 (s, 12H). ¹¹B (128 MHz, CDCl₃, ppm): δ 22.3.

2-((4-Fluorobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 7b).^{2,7} ¹H



(4–Fluorophenyl)methanol (7c).^{2,5} Following General procedure A. Colourless oil. Yield: OH H B5% (107 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.21-7.19 (m, 2H), 6.95-6.91 (m, 2H), 4.50 (s, 2H), 2.42 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 162.4 (d, $J_{C-F} = 244$ Hz), 136.5 (d, $J_{C-F} = 3$ Hz), 128.8 (d, $J_{C-F} = 8$ Hz), 115.5 (d, $J_{C-F} = 20$ Hz), 64.5.

2-((4-Chlorobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 8b).^{2,3} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.30-7.29 (m, 4H), 4.89 (s, 2H), 1.27 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3. (4–Chlorophenyl)methanol (8c).^{2,7} Following General procedure A. White solid. Yield: OH 87% (126 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.29-7.24 (m, 4H), H 4.59 (s, 2H), 2.42 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 139.3, 133.3, 128.7, 128.3, 64.4.

2-((4-Bromobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 9b).^{2,6} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.46 (d, J = 8 Hz, 2H), 7.23 (d, J = 8Hz, 2H), 4.87 (s, 2H), 1.27 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

(4–Bromophenyl)methanol (9c).^{2,5} Following General procedure A. White solid. Yield: OH 82% (154 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.44 (d, *J* = 8 Hz, 2H), H 7.15 (m, 2H), 4.55 (s, 2H), 2.60 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 139.7, 131.6, 128.6, 121.4, 64.4.

2-((3-Bromobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 10b).^{2,6} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.54 (br, 1H), 7.42-7.27 (m, 3H), 4.91 (s, 2H), 1.29 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

(**3-Bromophenyl)methanol** (**10c**).^{2,7} Following General procedure A. Colourless oil. Yield: 78% (146 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.45 (br, 1H), 7.38-7.37 (m, 1H), 7.20-7.06 (m, 2H), 4.55 (s, 2H), 2.94 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 143.1, 130.7, 130.5, 129.9, 125.4, 122.6, 64.2.

4,4,5,5-Tetramethyl-2-((**4-nitrobenzyl**)**oxy**)-**1,3,2-dioxaborolane** (**Table 2, 11b**).^{2,3 1}H NMR (400 MHz, CDCl₃, ppm): δ 8.20 (d, J = 8 Hz, 2H), 7.51 (d, J = 8 Hz, 2H), 5.03 (s, 2H), 1.28 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.0.

(4-Nitrophenyl)methanol (11c).^{2,9} Following General procedure B. Brown crystalline solid. OH Yield: 70% (107 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 8.19 (d, J = 8Hz, 2H), 7.51 (d, J = 8 Hz, 2H), 4.82 (s, 2H), 2.36 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 148.3, 147.3, 127.1, 123.8, 64.0.

4-(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)benzonitrile (Table 2, 12b).^{2,6} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.62 (d, J = 8 Hz, 2H), 7.44 (d, J = 8 Hz, 2H), 4.97 (s, 2H), 1.26 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

4-(Hydroxymethyl)benzonitrile (12c).^{2,7} Following General procedure B. White crystalline



solid. Yield: 80% (105 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.56 (d, *J* = 8 Hz, 2H), 7.42 (d, *J* = 8 Hz, 2H), 4.69 (s, 2H), 3.58 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 146.6, 132.2, 127.0, 118.9, 110.6, 63.8.



o-Tolylmethanol (13c).^{2,11} Following General procedure A. Colourless oil. Yield: 84% (103 OH mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.31-7.30 (m, 1H), 7.19-7.15 (m, H 3H), 4.58 (s, 2H), 2.58 (br, 1H), 2.30 (s, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 138.8, 136.0, 130.3, 127.7, 127.5, 126.0, 63.2, 18.6.

4,4,5,5-Tetramethyl-2-((2-Chlorobenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 14b).^{2,10}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.52 (br, 1H), 7.37-7.21 (m, 3H), 5.03 (s, 2H), 1.28 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

4,4,5,5-Tetramethyl-2-((2-Bromobenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 15b).¹²



¹H NMR 400 MHz, CDCl₃, ppm): δ 7.51 (br, 1H), 7.34-7.12 (m, 3H), 4.98 (s, 2H), 1.28 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4.



Cl ¹H NMR 400 MHz, CDCl₃, ppm): δ 7.47-7.35 (br, 1H), 7.35-7.34 (m, 1H), 7.28-7.24 (m, 1H), 4.97 (s, 2H), 1.28 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3. ¹³C NMR (100 MHz, CDCl₃, ppm): δ 137.0, 135.5, 133.6, 132.9, 128.9, 83.3, 63.6, 24.8.

4,4,5,5-Tetramethyl-2-(naphthalen-1-ylmethoxy)-1,3,2-dioxaborolane (Table 2, 17b).^{2,3}



¹H NMR (400 MHz, CDCl₃, ppm): δ 8.12-8.10 (m, 1H), 7.93-7.90 (m, 1H), 7.86-7.64 (m, 1H), 7.57-7.49 (m, 4H), 5.47 (s, 2H), 1.34 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4.

 Naphthalen-2-ylmethanol (17c).^{2,14} Following General procedure A. Colourless oil. Yield:

 HO
 HO

 HO
 HD

 NBW
 (140 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 8.12-8.10 (m, 1H), 7.91

 7.89 (m, 1H), 7.84-7.82 (m, 1H), 7.57-7.44 (m, 4H), 5.11 (s, 2H), 2.19 (br,

 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 136.3, 133.8, 131.2, 128.7, 128.6,

 126.4, 125.9, 125.4, 125.3, 123.7, 63.6.

2-(Cinnamyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 18b).^{2,6 1}H NMR (400 MHz, CDCl₃, ppm): δ 7.46-7.13 (m, 5H), 6.64 (d, *J* = 16 Hz, 1H), 6.31 (dt, *J* = 16, 6 Hz, 1H), 4.56 (d, *J* = 6 Hz, 2H), 1.30 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.2.



2-(Furan-2-ylmethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 19b).^{2,6 1}H



NMR (400 MHz, CDCl₃, ppm): δ 7.37 (br, 1H), 6.31 (br, 2H), 4.82 (s, 2H), 1.27 (s, 12H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 152.7, 142.5, 110.6, 108.4, 83.2, 59.2, 24.4. ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

1,3-*Bis*(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)benzene (Table2,



20b).^{2,3} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.36-7.30 (m, 4H), 4.95 (s, 4H), 1.29 (s, 24H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.0.

1,3-Phenylenedimethanol (Table 2, 20c).^{2,14}



Following General procedure A. A colorless oil. Yield: 73% (100 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.38-7.36 (m, 2H), 7.31-7.28 (m, 2H), 4.68 (s, 4H), 2.53 (br, 2H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 141.2, 128.7, 126.2, 125.6, 64.9.

2-(Cyclohexyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 21b).¹²



¹H NMR (400 MHz, CDCl₃, ppm): δ 3.63 (d, J = 7 Hz, 2H), 1.73-1.47 (m, 6H), 1.24 (s, 12H), 1.18-1.08 (m, 3H), 0.97-0.89 (m, 2H) ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

2-(Decyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 22b).^{2,4} ¹H NMR (400 MHz, CDCl₃, ppm): δ 3.74 (t, *J* = 7 Hz, 2H), 1.62-1.53 (m, 2H), 1.26 (br, 26H), 0.87 (t, *J* = 7 Hz, 3H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.2.

Decan-1-ol (22c).^{2,4} Following General procedure B. Colorless oil. Yield: 80% (127 mg). ¹H OH NMR (400 MHz, CDCl₃, ppm): δ 3.59 (t, J = 6 Hz, 2H), 2.73 (br, 1H), 1.55-1.52 (m, 2H), 1.26-1.24 (m, 14H), 0.87 (t, J = 6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 62.8, 32.7, 31.9, 29.7, 29.6, 29.5, 29.4, 25.8,

22.7, 14.1.

4,4,5,5-Tetramethyl-2-(1-phenylethoxy)-1,3,2-dioxaborolane (Table 3, 23b).^{2,3} ¹H NMR



(400 MHz, CDCl₃, ppm): δ 7.41-7.27 (m, 5H), 5.28 (q, J = 6 Hz, 1H), 1.53 (d, J = 8 Hz, 3H), 1.28 (s, 6H), 1.25 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1.

1-Phenylethanol (23c).^{2,4} Following General procedure A. Colourless oil. Yield: 80% (91
Mg) ¹H NMR (400 MHz, CDCl3): δ 7.40-7.35 (m, 4H), 7.32-7.28 (m, 1H),
4.90 (q, J = 6.5 Hz, 1H), 2.13 (br, 1H), 1.51 (d, J = 4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 145.9, 128.5, 127.5, 125.5, 70.4, 25.2.

4,4,5,5-Tetramethyl-2-(1-(p-tolyl)ethoxy)-1,3,2-dioxaborolane (Table 3, 24b).^{2,3 1}H NMR (400 MHz, CDCl₃, ppm): δ 7.27 (d, J = 8 Hz, 2H), 7.16 (d, J = 8 Hz, 2H), 5.25 (q, J = 6 Hz, 1H), 2.36 (s, 3H), 1.51 (d, J = 8 Hz, 3H), 1.29 (s, 6H), 1.28 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1.

1-(*p***-Tolyl)ethanol (24c).^{2,4}** Following General procedure B. Colourless oil. Yield: 72% (96 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.29 (d, J = 6.4 Hz, 2H), 7.18 (d, J = 6.5 Hz, 2H), 4.88 (q, J = 6 Hz, 1H), 2.37 (s, 3H), 1.93 (s, 1H), 1.50 (d, J = 8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 143.0, 137.2, 129.2, 125.4, 70.3, 25.1, 21.2.

2-(1-(4-Methoxyphenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3,



25b).^{2,3} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.32 (d, *J* = 8 Hz, 2H), 6.88 (d, *J* = 8 Hz, 2H), 5.24 (q, *J* = 6 Hz, 1H), 3.81 (s, 3H), 1.50 (d, *J* = 4 Hz, 3H), 1.26 (s, 6H), 1.23 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

1-(4-Methoxyphenyl)ethanol (Table 3, 25c).² Following General procedure B. Colorless oil.



Yield: 78% (118 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.33-7.31 (m, 2H), 6.93-6.90 (m, 2H), 4.85 (q, *J* = 6 Hz, 1H), 3.83 (s, 3H), 2.56 (br, 1H), 1.50 (d, *J* = 4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 158.9, 138.1, 126.7, 113.8, 69.8, 55.2, 25.0.

2-(Benzhydryloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**Table 3, 26b**).^{2,6 1}H NMR (400 MHz, CDCl₃, ppm): δ 7.51-7.50 (m, 4H), 7.42-7.39 (m, 4H), 7.35-7.32 (m, 2H), 6.31 (s, 1H), 1.31(s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4. Diphenylmethanol (Table 3, 26c).² Following General procedure B. White solid. Yield: OH 89% (162 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.17-7.07 (m, 10H), 5.48 (s, 1H), 3.20 (br, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 143.7, 128.3, 127.3, 126.5, 75.8.

2-((4-Fluorophenyl)(phenyl)methoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3,



27b). ^{2,6} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.45-7.07 (m, 9H), 6.26 (s, 1H), 1.29 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4.

2-((4-Chlorophenyl)(phenyl)methoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3,



28b).^{2,6} ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.42-7.35 (m, 9H), 6.22 (s, 1H), 1.27 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4. ¹³C NMR (100 MHz, CDCl₃, ppm): δ 142.6, 141.7, 133.1, 128.4, 128.4, 128.0, 127.5, 126.4, 62.2, 83.1, 24.6, 24.4.

2-(1-(4-Fluorophenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 29b).^{2,14}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.35 (m, 2H), 7.03-6.99 (m, 2H), 5.24 (q, *J* = 6 Hz, 1H), 1.48 (d, *J* = 8 Hz, 3H), 1.26 (s, 6H), 1.23 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1.

1-(4-Fluorophenyl)ethanol (29c).^{2,14} Following General procedure A. Colorless oil. Yield: OH 75% (105 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.35-7.32 (m, 2H), 7.06-7.02 (m, 2H), 4.86 (q, J = 6 Hz, 1H), 2.35 (s, 1H), 1.48 (d, J = 6.5 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 162.2 (d, J = 245 Hz), 141.6, 127.1 (d, J = 8 Hz), 115.3 (d, J = 21 Hz), 69.8, 25.3.

2-(1-(4-Bromophenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 30b).^{2,3}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.46 (m, 2H), 7.27 (m, 2H), 5.22 (br, 1H), 1.49 (br, 3H), 1.26 (s, 6H), 1.24 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1.

1-(4-Bromophenyl)ethanol (30c).^{2,10} Following General procedure C. The 87% yield of 30c was determined by ¹H NMR spectroscopy, using nitromethane as an internal standard. Colourless oil. Yield: 71% (142 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.42 (d, *J* = 8 Hz, 2H), 7.17 (d, *J* = 8 Hz, 2H), 4.77 (q, *J* = 6 Hz, 1H), 2.70 (br, 1H), 1.40 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (100

MHz, CDCl₃, ppm): δ 144.8, 131.5, 127.2, 121.13, 69.7, 25.2.

4,4,5,5-Tetramethyl-2-(1-(3-nitrophenyl)ethoxy)-1,3,2-dioxaborolane (Table 3, 31b).² ¹H NMR (400 MHz, CDCl₃, ppm): δ 8.22 (s, 1H), 8.10-8.08 (m, 1H), 7.70-7.68 (m, 1H), 7.51-7.49 (m, 1H), 5.32 (q, *J* = 6.4 Hz, 1H), 1.52 (d, *J* = 6.5 Hz, 3H), 1.24 (s, 6H), 1.21 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4.

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1-(3-Nitrophenyl)ethanol (31c).<sup>2</sup> Following General procedure B. Brown solid. Yield: 41%
OH (67 mg). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm): \delta 8.22 (s, 1H), 8.09 (d, J = 8
Hz, 1H), 7.70 (d, J = 8 Hz, 1H), 7.51 (t, J = 8 Hz, 1H), 5.00 (q, J = 6.4 Hz,
1H), 2.80 (s, 1H), 1.52 (d, J = 6.5 Hz, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>,
ppm): \delta 148.3, 148.0, 131.7, 129.4, 122.3, 120.4, 69.3, 25.4.
```

2-(2-Chloro-1-phenylethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 32b).^{2,15}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.37-7.34 (m, 5H), 5.26 (m, 1H), 3.63 (m, 2H), 1.25 (s, 6H), 1.22 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

2-Chloro-1-phenylethan-1-ol (32c).^{2,16} Following General procedure A. Colorless oil. Yield:
OH 58% (89 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.39-7.34 (m, 5H), 4.91CH₂Cl 4.88 (m, 1H), 3.74 (dd, J = 11.3, 3.6 Hz, 1H), 3.65 (dd, J = 11.3, 8.6 Hz, 1H), 2.82 (s, 1H).¹³C NMR (100 MHz, CDCl₃, ppm): δ 140.0, 128.7, 128.5, 126.1, 74.1, 50.8.

2-(2-Bromo-1-phenylethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 33b).^{2,15}



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.40-7.32 (m, 5H), 5.33 (m, 1H), 3.56 (m, 2H), 1.28 (s, 6H), 1.25 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.4.

2-Bromo-1-phenylethan-1-ol (33c).^{2,16}



Following General procedure C. The 60% yield of **33c** was determined by ¹H NMR spectroscopy, using nitromethane as an internal standard. Colorless oil. Yield: 37% (57 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.39-7.34 (m, 5H), 4.94-4.90 (m, 1H), 3.64 (dd, *J* = 11.3, 3.6 Hz, 1H), 3.55 (dd, *J* = 11.3, 8.6 Hz, 1H), 2.78 (s, 1H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 140.4, 128.6, 128.4, 126.04, 73.7, 39.9.

2-(1-(2-Methoxyphenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 34b).²



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.56 (d, *J* = 8 Hz, 1H), 7.26-7.22 (m, 1H), 7.00-6.97 (m, 1H), 6.86 (d, *J* = 8 Hz, 1H), 5.61 (q, *J* = 6.3 Hz, 1H), 3.84 (s, 3H), 1.48 (d, *J* = 6 Hz, 3H), 1.28 (s, 6H), 1.26 (s, 6H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.3.

1-(2-Methoxyphenyl)ethanol (34c).^{2,17} Following General procedure C. The 59% yield of



34c was determined by ¹H NMR spectroscopy, using nitromethane as an internal standard. Colorless oil. Yield: 56% (84 mg). ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.43-7.41 (m, 1H), 7.33-7.29 (m, 1H), 7.05-7.01 (m, 1H), 7.01-6.92 (m, 1H), 5.17 (q, *J* = 6 Hz, 1H), 3.90 (s, 3H), 3.10 (s, 1H), 1.56 (d,

J = 6.5 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 156.4, 133.5, 128.1, 126.02, 120.7, 110.3, 66.1, 55.2, 22.9.

3-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolane-2-yloxy)quinuclidine (Table 3, 35b).¹⁸ ¹H $\bigwedge_{0} \xrightarrow{0} \xrightarrow{B} \xrightarrow{0} \xrightarrow{0} \xrightarrow{11} B$ NMR: δ 3.72 (br, 1H), 2.91-2.62 (m, 6H), 1.96-1.30 (m, 5H), 1.23 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.0.

2-(Adamantan-2-yloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**Table 3, 36b**).¹⁸ ¹H NMR (400 MHz, CDCl₃, ppm): δ 4.19 (m, 1H), 2.13-1.45 (m, 14H), 1.25 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 21.9. ¹³C NMR (100 MHz, CDCl₃, ppm): δ 81.2, 71.7, 48.1, 36.5, 35.2, 25.7, 23.5.

2-(Cyclohexyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 37b).¹⁸ ¹H NMR



(400 MHz, CDCl₃, ppm): δ 3.97 (m, 1H), 1.88-1.31 (m, 10H), 1.24 (s, 12H). ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 21.8. ¹³C NMR (100 MHz, CDCl₃): δ 82.4, 72.6, 34.2, 25.4, 24.5, 23.8.

2-(Thiophene-2-yloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 38b).¹²



¹H NMR (400 MHz, CDCl₃, ppm): δ 7.37 (br, 1H), 7.22-6.98 (br, 2H), 5.51 (br, 1H), 1.62 (br, 3H), 1.27 (s, 12H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ 152.7, 142.5, 110.6, 108.4, 83.2, 59.2, 24.4. ¹¹B NMR (128 MHz, CDCl₃, ppm): δ 22.1. ¹³C NMR (100MHz, CDCl₃, ppm): 148.2, 126.5, 124.2, 123.3, 82.9, 68.7, 25.2, 24.5.

IV. NMR Spectra of Hydroboration Products and Related Alcohols

Note: The borate esters spectra are shown along with internal standard (IS) nitromethane. Resonances are denoted as follows: unreacted aldehydic or ketonic compound (*), and solvent/grease (#). In the ¹¹B NMR spectra of few substrates, aside from the signal for borate ester ($\delta = 22$ ppm), a broad signal corresponded to B₂pin₃, pinBOBpin, and/or HOBpin, was observed at $\delta = 21$ ppm (overlapped with borate ester signal).^{19,20}





¹¹B NMR of **1b** (128 MHz, CDCl₃)



¹³C NMR of **1c** (100 MHz, CDCl₃)

4,4,5,5-Tetramethyl-2-((4-methylbenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 2b).^{2,3}



¹¹B NMR of **2b** (128 MHz, CDCl₃)

p-Tolylmethanol (2c).^{2,4}





¹¹B NMR of **3b** (128 MHz, CDCl₃)



¹³C NMR of **3c** (100 MHz, CDCl₃)




¹¹B NMR of **4b** (128 MHz, CDCl₃)

(3-Methoxyphenyl)methanol (4c).²



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4-(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)phenol (Table 2, 5b).²



¹¹B NMR of **5b** (128 MHz, CDCl₃)

N,N-Dimethyl-4-(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl) aniline (Table 2, 6b). 8



¹¹B NMR of **6b** (128 MHz, CDCl₃)

2-((4-Fluorobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 7b).²



¹¹B NMR of **7b** (128 MHz, CDCl₃)

(4–Fluorophenyl)methanol (7c).^{2,5}



¹³C NMR of **7c** (100 MHz, CDCl₃)

2-((4-Chlorobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 8b).^{2,3}



¹¹B NMR of **8b** (128 MHz, CDCl₃)

(4–Chlorophenyl)methanol (8c).²



2-((4-Bromobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 9b).^{2,6}



¹¹B NMR of **9b** (128 MHz, CDCl₃)



¹³ C NMR of **9c** (100 MHz, CDCl₃)

2-((3-Bromobenzyl)oxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 10b).^{2,6}



¹¹B NMR of **10b** (128 MHz, CDCl₃)

(3-Bromophenyl)methanol (10c).²



¹³ C NMR of **10c** (100 MHz, CDCl₃)



¹¹B NMR of **11b** (128 MHz, CDCl₃)





¹³C NMR of **11c** (100 MHz, CDCl₃)

4-(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)benzonitrile (Table 2, 12b).^{2,6}



¹¹B NMR of **12b** (128 MHz, CDCl₃)

4-(Hydroxymethyl)benzonitrile (12c).²



¹³C NMR of **12c** (100 MHz, CDCl₃)



¹¹B NMR of **13b** (128 MHz, CDCl₃)









¹¹B NMR of **14b** (128 MHz, CDCl₃)

4,4,5,5-Tetramethyl-2-((2-Bromobenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 15b).¹²



¹¹B NMR of **15b** (128 MHz, CDCl₃)

4,4,5,5-Tetramethyl-2-((2,4-Dichlorobenzyl)oxy)-1,3,2-dioxaborolane (Table 2, 16b).¹³



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¹³C NMR of **16b** (100 MHz, CDCl₃)

4,4,5,5-Tetramethyl-1-(naphthalen-1-ylmethoxy)-1,3,2-dioxaborolane (Table 2, 17b).^{2,3}



¹¹B NMR of **17b** (128 MHz, CDCl₃)

Naphthalen-1-ylmethanol (17c).²







¹¹B NMR of **18b** (128 MHz, CDCl₃)



¹³C NMR of **18c** (100 MHz, CDCl₃)

2-(Furan-2-ylmethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 19b).^{2,6}



¹¹B NMR of **19b** (128 MHz, CDCl₃)



¹³C NMR of **19b** (100 MHz, CDCl₃)

1,3-bis(((4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)oxy)methyl)benzene (Table 2, 20b).^{2,3}



1,3-Phenylenedimethanol (20c)



¹³C NMR of **20b** (100 MHz, CDCl₃

2-(Cyclohexyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 21b).¹²



¹¹B NMR of **21b** (128 MHz, CDCl₃)

2-(Decyloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 2, 22b).^{2,4}



¹¹B NMR of **22b** (128 MHz, CDCl₃)



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4,4,5,5-Tetramethyl-2-(1-phenylethoxy)-1,3,2-dioxaborolane (Table 3, 23b).^{2,3}

¹¹B NMR of **23b** (128 MHz, CDCl₃)

1-Phenylethanol (23c).^{2,4}



¹³C NMR of **23c** (100 MHz, CDCl₃)




¹³C NMR of **24c** (100 MHz, CDCl₃)

2-(1-(4-Methoxyphenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 25b).^{2,3}



1-(4-Methoxyphenyl)ethanol (Table 3, 25c).²



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2-(Benzhydryloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 26b).^{2,6}

¹¹B NMR of **26b** (128 MHz, CDCl₃)



¹³C NMR of **26c** (100 MHz, CDCl₃)

 $\label{eq:2-((4-Fluorophenyl)(phenyl)methoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane~(Table 3, 27b).^2$



¹¹B NMR of **27b** (128 MHz, CDCl₃)



¹³C NMR of **27b** (100 MHz, CDCl₃)

 $\label{eq:2-((4-Chlorophenyl)(phenyl)methoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane~(Table 3, 28b).^2$



¹¹B NMR of **28b** (128 MHz, CDCl₃)



¹³C NMR of **28b** (100 MHz, CDCl₃)

2-(1-(4-Fluorophenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 29b).^{2,14}



¹¹B NMR of **29b** (128 MHz, CDCl₃)

1-(4-Fluorophenyl)ethanol (29c).^{2,14}



¹³C NMR of **29c** (100 MHz, CDCl₃)

2-(1-(4-Bromophenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 30b).^{2,3}



¹¹B NMR of **30b** (128 MHz, CDCl₃)

1-(4-Bromophenyl)ethanol (30c).²



¹³C NMR of **30c** (100 MHz, CDCl₃)



4,4,5,5-Tetramethyl-2-(1-(3-nitrophenyl)ethoxy)-1,3,2-dioxaborolane (Table 3, 31b).²

NMR of 31b (128 MHz, CDCl₃)

1-(3-Nitrophenyl)ethanol (31c).²







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2-(2-Bromo-1-phenylethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 33b).^{2,15}

¹¹B NMR of **33b** (128 MHz, CDCl₃)

2-Bromo-1-phenylethan-1-ol (33c).^{2,16}



¹³C NMR of **33c** (100 MHz, CDCl₃)



2-(1-(2-Methoxyphenyl)ethoxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 34b).²

¹¹B NMR of **34b** (128 MHz, CDCl₃)

1-(2-Methoxyphenyl)ethanol (34c).^{2,17}



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3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolane-2-yloxy)quinuclidine (Table 3, 35b).¹⁸





2-(Adamantan-2-yloxy)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (Table 3, 36b).¹⁸





¹³C NMR of **36b** (100 MHz, CDCl₃)



¹¹B NMR of **37b** (128 MHz, CDCl₃)





¹¹B NMR of **38b** (128MHz, CDCl₃)





V. Competitive Chemoselective Hydroboration Reactions

Experimental Procedure for the Examples Described in Scheme 1: In a 10 mL thickwalled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.0250 mmol), HBpin (1.0 equiv, 36 μ L, 0.25 mmol), MTBE (1 mL), aldehyde (0.25 mmol) and ketone (0.25 mmol) were added, and the reaction was stirred vigorously at room temperature for 24 h (for ketone at 50 °C). The reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (\emptyset 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate esters yields were determined by ¹H NMR spectroscopy using nitromethane as an internal standard.



Figure S6. ¹H NMR spectra of competitive hydroboration reaction between benzaldehyde (1a) and acetophenone (23a).



Figure S7. ¹H NMR spectra of competitive hydroboration reaction between 4methoxybenzaldehyde (**3a**) and 1-(4-methoxyphenyl)ethanone (**25a**).



Figure S8. ¹H NMR spectra of competitive hydroboration reaction between 4bromolbenzaldehyde (**9a**) and 1-(4-bromophenyl)ethenone (**30a**).



Figure S9. ¹H NMR spectra of competitive hydroboration reaction between benzaldehyde (1a) and styrene.



Figure S10. ¹H NMR spectra of competitive hydroboration reaction between acetophenone (**23a**) and styrene.



Figure S11. ¹H NMR spectra of competitive hydroboration reaction between benzaldehyde (1a) and phenylacetylene.



Figure S12. ¹H NMR spectra of competitive hydroboration reaction between acetophenone (**23a**) and phenylacetylene.
Experimental Procedure for the Examples Described in Scheme 2: In a 10 mL thickwalled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.0250 mmol), HBpin (1.0 equiv, 36 μ L, 0.25 mmol), MTBE (1 mL) and substrate (0.25 mmol) were added and the reaction was stirred vigorously at room temperature for 24 h (for ketone at 50 °C). The reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (Ø 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard.



Figure S13. ¹H NMR spectra of intramolecular competitive hydroboration reaction of 4-acetylbenzaldehyde (**39a**).



Figure S14. ¹H NMR spectra of intramolecular competitive hydroboration reaction of 4-formylphenyl acetate (**40a**).



Figure S15. ¹H NMR spectra of intramolecular competitive hydroboration reaction of N-(4-formylphenyl)acetamide (**41a**).



Figure S16. ¹H NMR spectra of intramolecular competitive hydroboration reaction of 4acetylbenzonitrile (**42a**).



Figure S17. ¹H NMR spectra of intramolecular competitive hydroboration reaction of (E)-chalcone (**43a**).



Figure S18. ¹H NMR spectra of intramolecular competitive hydroboration reaction of (E)-1- (4-nitrophenyl)-3-phenylprop-2-en-1-one (**44a**).

Experimental Procedure for BH₃ **Mediated Reduction:** In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, BH₃·THF (10 mol %, 0.0250 mmol), HBpin (1.0 equiv, 36 μ L, 0.25 mmol), MTBE (1 mL) and substrate (0.25 mmol) were added, and the reaction was stirred vigorously at room temperature for 24 h (for ketone at 50 °C). The reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (Ø 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard.

A). The BH₃ mediated carbonyl hydroboration of 4-nitrobenzaldehyde (**11a**) having both NO₂ and C=O groups gave reduction at the carbonyl part as well as nitro group.



Figure S19. ¹H NMR spectra of intramolecular competitive hydroboration reaction of 4nitrobenzaldehyde (**11a**) catalyse by BH₃·THF.

B). The intermolecular selectivity experiments were performed using BH_3 as a catalyst. The equimolar amounts of benzaldehyde (1a) and styrene with HBpin in the presence of BH_3 ·THF catalyst gave both aldehyde (1b: 48%) and styrene hydroboration (52%) products.



Figure S20. ¹H NMR spectra of intermolecular competitive hydroboration reaction between benzaldehyde (**1a**) and styrene catalyse by BH₃·THF.

C). The intermolecular selectivity experiments were performed using BH₃ as a catalyst. The reaction between stoichiometric amounts of acetophenone, phenylacetylene and HBpin gave 8% ketone hydroboration and 92% alkyne hydroboration products.



Figure S21. ¹H NMR spectra of intermolecular competitive hydroboration reaction between acetophenone (**23a**) and phenylacetylene catalyse by BH₃·THF.

VI. Mechanistic Investigations

Mercury Poisoning Experiments.

a) In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.0250 mmol), HBpin (1.3equiv, 48 μ L, 0.33 mmol), methyl-*tert*-butyl ether (1 mL), benzaldehyde (**1a**, 0.25 mmol, 20 μ L) and Hg (300 equiv) were added, and the reaction was stirred vigorously at room temperature for 24 h. The reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (Ø 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard, no isolable product was obtained in this experiment.



b) In a 10 mL thick-walled reaction tube equipped with a magnetic stirring bar, CeO₂ NPs (10 mol %, 4 mg, 0.025 mmol), HBpin (1.3 equiv, 48 μ L, 0.33 mmol), methyl-*tert*-butyl ether (1 mL) and benzaldehyde (**1a**, 0.25 mmol, 20 μ L) were added, and the reaction was stirred vigorously at room temperature for 3 h. After 3 h reaction process, the reaction was interrupted and 300 equiv of Hg was added. After stirring for about 24 h at room temperature the reaction mixture was then diluted with Et₂O (2 mL) and filtered through a plug of celite (\emptyset 3 mm × 8 mm) with copious washing (Et₂O). The solvents were removed *in vacuo*, and the borate ester yield was determined by ¹H NMR spectroscopy using nitromethane as an internal standard.



Recyclability Experiment for Hydroboration of 4-bromobenzaldehyde (9a).



In a 50 mL round bottom flask equipped with a magnetic stirring bar, 10 mol% of CeO₂ (5 mol %, 0.1 mmol, 17 mg), HBpin (1.3 mmol, 192 μ L), methyl-tert-butyl ether (4 mL) and 4bromobenzaldehyde (**9a**, 1.0 mmol, 185 mg) were added. The resulting reaction mixture was stirred vigorously at room temperature for 24 h. After completion of the reaction, the solid was centrifuged out of suspension and extracted with diethyl ether (three times). The solvents were removed *in vacuo*, and the combined organic extracts were analyzed by ¹H NMR spectroscopy using nitromethane as an internal standard.

The recovered CeO₂ NPs was charged into 50 mL round bottom flask equipped with a magnetic stirring bar, to which additional HBpin (1.3 mmol, 192 μ L), methyl-*tert*-butyl ether (4 mL) and 4-bromobenzaldehyde (**9a**, 1.0 mmol, 185 mg) were added. The resulting reaction mixture was stirred vigorously at room temperature for 24 h. Then, NPs were centrifuged out of suspension and extracted with Et₂O. The combined organic product was extracted five times with Et₂O after each run and the reaction was continued with a fresh batch of substrate (4-bromobenzaldehyde, 1 mmol scale), HBpin (1.3 mmol, 192 μ L) and methyl-*tert*-butyl ether (4 mL) added to the separated CeO₂ NPs. The combined organic extracts were analyzed by ¹H NMR spectroscopy using nitromethane as an internal standard. The results of 10 recycling experiments are summarized in Figure S12.

Yields: 1^{st} cycle = >99%, 2^{nd} cycle = >99%, 3^{rd} cycle = 90%, 4^{th} cycle = 87 %, 5^{th} cycle = 85%, 6^{th} cycle = 84%, 8^{th} cycle = 83%, 9^{th} cycle = 81 %, 10^{th} cycle = 78%.



Figure S22.¹H NMR yields (%) of borate ester at different cycles in the recyclability experiment for CeO₂ NPs catalysed hydroboration of 4-bromobenzaldehyde.

Gratifyingly, when the recovered catalyst, after the 2^{nd} cycle of the reaction, was calcined at 600 °C for 2 h, and has been used as a catalyst for the hydroboration of 4-bromobenzaldehyde (**9a**) under standard reaction conditions gave the desire borate ester **9b** in 99% yield confirmed by ¹H NMR spectroscopy.

Furthermore, when the recovered catalyst, after the 10^{th} cycle of the reaction was calcined at 600 °C for 2 h, and has been used as a catalyst for the hydroboration of **9a** also gave the desire borate ester **9b** in 99% yield.



Figure S23. FE-SEM images of calcined CeO_2 nanoparticles, recovered after 10 catalytic cycles.

The calcined CeO₂ nanoparticles recovered after 10 catalytic cycles were characterized by field emission scanning electron microscopy (FESEM, Figure S13). The FESEM images demonstrated that the morphology of nanoparticles is intact.

Stoichiometric reaction of CeO2 NPs with benzaldehyde and HBpin.

Monitoring by FT-IR spectroscopy:

Several stoichiometric experiments were performed to gain insight into the hydroboration mechanism. In an argon-filled glovebox, the CeO₂ NPs (21 mg, 0.125 mmol) and benzaldehyde (13 mg, 0.125 mmol) were mixed, and the reaction mixture was analyzed by Fourier transform infra-red (FT-IR) spectroscopy (Figure S14). The FT-IR analysis of CeO₂ NPs shows strong absorption peak at 469 cm⁻¹, can be attributed to the Ce–O band vibrations (Figure S14a). The FT-IR analysis of benzaldehyde shows strong absorption peak at 1701 cm⁻¹ is due to the CO stretching frequency (Figure S14b). The FT-IR spectrum of the reaction mixture of CeO₂ and benzaldehyde shows shift in the absorption band correspond to the Ce–O vibration as well as in the CO stretching frequency towards lower wavenumber indicating interaction between CeO₂ NPs catalyst and benzaldehyde substrate (Figure S14c)



Figure S24. IR spectra of: (a) CeO_2 ; (b) C_6H_5CHO and (c) $CeO_2 + C_6H_5CHO$.

In an argon-filled glove box, the CeO₂ NPs (21 mg, 0.125 mmol) and excess HBpin (160 mg, 1.25 mmol) were mixed, and the reaction mixture was analyzed by FT-IR spectroscopy (Figure S15). The FT-IR spectrum of the reaction mixture does not show shift in the absorption band correspond to the Ce–O vibration, indicating no interaction between CeO₂ and HBpin (Figure S15c).



Figure S25. IR spectra of: (a) CeO₂; (b) HBpin and (c) CeO₂+ HBpin.

At this point we cannot speculate on the detailed mechanism of the catalytic reaction. However, based on the previous reports²¹ and our experimental results, a plausible mechanism for the hydroboration of aldehydes and ketones is proposed. In the first step, the CeO₂ NPs polarises the carbonyl oxygen moiety to give an active catalytic species.²² In the next step, a nucleophilic attack occurs on the electron-deficient carbonyl carbon atom by the incoming hydride of HBpin, leading to the formation of the four-membered transition state, which undergoes σ bond metathesis reaction, followed by reaction with another molecule of carbonyl substrate gives borate ester and regenerate the catalytic active species (Figure 16).



Figure 26. Possible schematic representation of carbonyl interaction with CeO₂ NPs with the formation of borate ester.

VII. References

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