Supporting information

A Schiff base sensor for relay monitoring $\mathrm{In^{3+}}$ and $\mathrm{Fe^{3+}}$ through an

"off-on-off" fluorescence signals

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Experimental section

Materials

All metal salts such as MgCl₂, CaCl₂, CuCl₂·2H₂O, NiCl₂·6H₂O, FeCl₂·3H₂O, AlCl₃·6H₂O, AgCl, CdCl₂·2.5H₂O, FeCl₃·6H₂O, CoCl₂·6H₂O, CrCl₃·6H₂O, InCl₃, MnCl₂·4H₂O, KCl, ZnCl₂, HgCl₂, LiCl and Ga(NO₃)₃ were analytical grade and used without further purification. All other organic reagents were purchased and used as received.

Instruments

UV-vis spectra were recorded through a Shimadzu 3100 spectrometer. Fluorescence measurements were carried out using an Edinburgh Instruments Ltd-FLS920 fluorescence spectrophotometer Fluorescence measurements were performed when excitation at 410 nm. The slits of excitation and emission were 10 nm and 20 nm, respectively. ¹H NMR spectra were executed with a Bruker AV III 400 MHz NMR spectrometer with tetramethysilane (TMS) as an internal standard. ¹³C NMR spectral data were obtained using a Bruker AV III 100 MHz NMR spectrometer with tetramethylsilane (TMS) as the internal standard and DMSO as the solvent. Infrared spectral data were obtained using a Bruker obtained using a Bruker Vertex 70 FT-IR spectrometer and samples as KBr pellets.

Sample preparation

All tests involved in this work were carried out with distilled water at room temperature (25 °C). In the experiments of titration with various metal ions, the sensor was dissolved in Tris DMF – H₂O (9 : 1, v/v) buffer solution to afford the test solution (1 × 10⁻⁵ M). Stock solutions (1 × 10⁻⁵ M) of KCl, AlCl₃, MgCl₂, CaCl₂, LiCl, FeCl₂, HgCl₂, NiCl₂, CuCl₂, ZnCl₂, AgCl, MnCl₂, FeCl₃, CoCl₂, CrCl₃, CdCl₂, InCl₃, Ga(NO₃)₃ were prepared at a concentration of 0.03 M with distilled water.

Calculation of quantum yield

The quantum yield of **LB2**, [**LB2**+In³⁺] and [**LB2**+Fe³⁺] were determined according to the following equation:

$$\phi_u = \phi_s \frac{F_u A_s {n_u}^2}{F_s A_u {n_s}^2}$$

where ϕ is quantum yield; F is integrated area under the corrected emission

spectra; *A* is the absorbance at the excitation wavelength; *n* is the refractive index of the solution; the subscripts *u* and *s* refer to the unknown and the standard, fluorescence respectively. Rhodamine B in ethanol solution was used as the standard, which has a quantum yield of 0.97.⁵⁹

Theoretical calculation

Density functional theory (DFT) structural optimizations were performed on the Gaussian 09 program. In all conditions, the structures were optimized via the B3LYP functional and the mixed basis sets 6-31G(d) and LANL2DZ. Each structure was subsequently subjected to TD-DFT calculation using the B3LYP functional soon afterwards .The absence of imaginary frequencies for all the optimized structures were confirmed through executing the frequency calculations. The molecular orbitals were visualized and plotted with the GaussView 09 program.⁶⁰

Synthesis of LB2

Compoundmethyl4,5-dihydronaphtho[1,2-b]thiophene-2-carboxylate(1)and4,5-dihydronaphtho[1,2-b]thiophene-2-carbohydrazide(2)was synthesized according to the previous work. 61-62SynthesizeofN'-(3-ethoxy-2-hydroxybenzylidene)-4,5-dihydronaphtho[1,2-b]thiophene-2-carbohydrazide (LB2)

Compound **2** (25 mg, 0.1 mmol) and 3-ethoxy-2-hydroxybenzaldehyde (20 mg, 0.12 mmol) was mixed in ethanol (2 ml). The mixture was then stirred at room temperature overnight. The precipitate was filtered and washed

with ethanol to afford the target product (white solid). Yield: 25 mg, 64.9%. ESI-MS: $m/z = 393.1388 [M + H]^+$. FTIR (KBr, cm⁻¹):3128(N-H), 1400(C=N). 1H NMR (400 MHz, DMSO) δ 11.93 (d, J = 142.8 Hz, 0H), 9.93 (d, J = 655.0 Hz, 0H), 8.57 (d, J = 51.2 Hz, 0H), 7.87 (d, J = 40.6 Hz, 1H), 7.47 (d, J = 5.9 Hz, 1H), 7.30 (dd, J = 13.8, 6.3 Hz, 1H), 7.17 (d, J = 7.3 Hz, 0H), 7.03 (d, J = 7.8 Hz, 0H), 6.86 (d, J = 7.6 Hz, 1H), 4.08 (d, J = 6.6 Hz, 1H), 2.95 (d, J = 6.8 Hz, 1H), 2.85 (t, J = 7.3 Hz, 2H), 1.37 (t, J = 6.7 Hz, 1H).



Fig.S1 ESI mass spectrum of probe LB2.



Fig.S3 ¹H NMR spectrum of compound LB2.



Fig.S4 ¹³C NMR spectrum of compound LB2.



Fig.S5 Time dependent fluorescence response of LB2 (30 μ M) at λ =516nm in the presence of In³⁺ (3 eq.) in DMF-H₂O (9:1, v:v) 10 mM of Tris buffer solution at pH 7.4. λ ex = 410 nm



Fig.S6 Changes in absorption spectroscopy of LB2 (30 μ M) with different ions (30 μ M) in DMF-H₂O (9:1, v:v) 10 mM of Tris buffer solution at pH 7.4.



Fig.S7 Job's plot of the **LB2** in DMF–H₂O (9 : 1, Tris 0.01 M, pH=7.4) at 25 $^{\circ}$ C. The total concentration of **LB2** and In³⁺ was 0.05 mM. Excitation is at 410 nm, and emission is monitored at 516 nm.



Fig.S8 Fluorescence intensity of LB2 at 516 nm with the addition of In^{3+} (0 - 1 equiv.) in DMF-H₂O (9 : 1, Tris 0.01 M, pH =7.4).



Fig.S9 Fluorescence intensity of LB2 and LB2 in the presence of In^{3+} at various pH values in DMF– H_2O (9 : 1, v/v) 10 mM of Tris buffer solution.



Fig.S10 Fluorescence intensity of $[LB2+In^{3+}]$ at 516 nm with the addition of Fe³⁺ (0 - 1 equiv.) in DMF-H₂O (9 : 1, Tris 0.01 M, pH =7.4).



Fig.S11 Absorption spectra of $[LB2+In^{3+}]$ and $[LB2+Fe^{3+}]$ in DMF-H₂O buffer solution (v/v = 9:1, Tris = 10 mM, pH = 7.4).



Fig.S12 Changes in absorption spectroscopy of [LB2+In³⁺] (30 μ M) with incremental addition of In³⁺ (0–30 μ M) in DMF–H₂O (9 : 1, v/v) 10 mM of Tris buffer solution at pH 7.4.



Fig.S13 ESI mass spectrum of complex [LB2+In³⁺].



Fig.S14 ESI mass spectrum of complex [LB2+Fe³⁺].



Fig.S15 The optimized geometry of LB2, $[LB2+In^{3+}]$ and $[LB2+Fe^{3+}]$ at the B3LYP level of theory, where light-gray, red, blue, white, purple-gray, grayish-blue and green spheres denote C, O, N, H, In, Fe and Cl atoms, respectively.

sample	In ³⁺ added	In ³⁺ recovered	Recovery	RSD	
	(mol L ⁻¹)	(mol L ⁻¹)	(%)	(%)	
1	3×10^{-5}	3.2×10^{-5}	107.6	0.61	
2	4×10^{-5}	4.2×10^{-5}	106.5	0.86	
3	5 × 10 ⁻⁵	5.3×10^{-5}	105.4	0.97	

Table S1 Determination of the In³⁺ concentration in tap water samples

Table S2	2 Deter	mination	of the	Fe ³⁺	concentration	ı in	tap	water	sampl	les
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sample	Fe ³⁺ added	Fe ³⁺ recovered	Recovery	RSD
	(mol L ⁻¹)	$(mol L^{-1})$	(%)	(%)

1	1 × 10 ⁻⁵	1.1 × 10 ⁻⁵	113.1	2.08
2	2×10^{-5}	2.2×10^{-5}	110.2	2.43
3	4×10^{-5}	3.8×10^{-5}	95.3	2.29

Table S3 Determination of the In ³⁺	concentration in drink water samples
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sample	In ³⁺ added	In ³⁺ recovered	Recovery	RSD
_	(mol L ⁻¹)	$(mol L^{-1})$	(%)	(%)
1	2×10^{-5}	$1.9 imes 10^{-5}$	96.8	1.34
2	3 × 10 ⁻⁵	3.1 × 10 ⁻⁵	104.5	0.63
3	5×10^{-5}	5.2×10^{-5}	104.7	1.52

Table S4 Determination of the Fe³⁺ concentration in drink water samples

sample	Fe ³⁺ added	Fe ³⁺ recovered	Recovery	RSD
	(mol L-1)	(mol L-1)	(%)	(%)
1	1 × 10 ⁻⁵	1.0 × 10 ⁻⁵	109.3	1.38
2	2×10^{-5}	2.2×10^{-5}	111.8	1.69
3	3 × 10 ⁻⁵	2.8×10^{-5}	94.6	1.21

Table S5 Comparison of type of indium sensors and their detection limits

Solvent system	Detection limit	Response	Reference
DMF	$5.3\times10^{\text{-7}}M$	-	2
CH ₃ CN	$6.4\times10^{\text{-8}}M$	-	13
Ethanol	$6.1\times10^{\text{-7}}M$	turn-on	19
Methanol	$2 \times 10^{-6} M$	turn-on	28
DMF/H ₂ O(v/v,9:1)	$8.05\times 10^{\text{-9}}M$	off-on	this work

Table S6 Comparison of type of ferric ion sensors and their detection limits

Solvent system	Detection limit	Response	Reference
DMF	$4.4\times10^{\text{-7}}M$	-	2
DMSO/H ₂ O(v/v,9:1)	$6.7\times10^{\text{-8}}M$	off-on	30
Pure aqueous	$2.49\times10^{\text{-7}}M$	on-off	33
THF/H ₂ O(40:60)	$2.95\times10^{\text{-6}}M$	turn-on	34
DMF/H ₂ O(v/v,9:1)	$2.59\times10^{\text{-8}}M$	on-off	this work