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Supplementary Material

## Iron and nitrogen co-doped graphene quantum dots as highly active peroxidase for sensitive detection of L-cysteine

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Fig. S1 The photographs of Fe,N-GQDs solution diluted by a factors 10 after storage at room temperature for one month.



Fig. S2 The absorbance change ratio of Fe,N-GQDs+H<sub>2</sub>O<sub>2</sub>+TMB after different kinds of amino acids (50  $\mu$ M) was added and reacted for 10 min.



Fig. S3 Linear calibration plot for L-Cys detection in amino acid tablets solution based on the peroxidase of Fe,N-GQDs.

	Substrate TMB		Substrate H2O2		Poforonco
Catalyst _					
	Km	V <sub>max</sub>	Km	V <sub>max</sub>	Kelelence
	( <b>mM</b> )	(10 <sup>-8</sup> M/s)	( <b>mM</b> )	(10 <sup>-8</sup> M/s)	
HRP	0.434	10	2.39	4.36	[1]
SWNT-NH <sub>2</sub> @hemin	0.528	10.08	26.012	22.558	[2]
PEI-rGO-Hemin-BSA	1.8	8.451	2.13	2.339	[3]
MA-Hem/Au-Ag	2.39	1.42	2.7	14.1	[4]
Hemin-porous g-C <sub>3</sub> N <sub>4</sub> hybrid nanosheets	0.119	11.6	0.682	2	[5]
His-GQD/hemin	0.133	9.7	3.8	10.55	[6]
Fe,N-GQDs	0.1351	15.99	1.905	12.09	This work

**Table S1** The apparent Michaelis-Menten constants ( $K_m$ ) and maximum reaction rates ( $V_{max}$ ) of different catalysts

Songing mothod	Motorial	<b>Detection limit</b>	Linear range	Reference	
Sensing method	Wateria	(µM)	(µM)	Kelerence	
Colorimetry	Ag <sup>+</sup> -CDs	0.82	1-60	[7]	
Colorimetry	$g-C_3N_4$	0.2	1-20	[8]	
Colorimetry	FeCo carbon nanofibers	0.15	1-20	[9]	
Electrochemistry	N-doped rGO modified	0.8	1.3-720	[10]	
	with Y <sub>2</sub> O <sub>3</sub>				
Electrochemistry	Carbon electrode	0.09	0.3-3.6	[11]	
	modified with carbon		3.9-7.2		
	dots				
Chemiluminescence	Lucigenin-carbon dots	8.8	10-100	[12]	
Fluorometry	N-CDs	0.35	0-100	[13]	
Fluorometry	N-acetyl-L-cysteine-	0.16	1.0-110	[14]	
	capped AuNPs-CDs				
Fluorometry	AuNCs-CDs	0.96	1-100	[15]	
Fluorometry	N,S-CQDs	0.54	10-200	[16]	
Colorimetry	Fe,N-GQDs	0.14	0.5-50	This work	

Table S2 Comparison of the performance for detection of L-Cys using different carbon-based materials

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