Supporting Information

Two novel tetraphenylethylene-skeleton salamo-type fluorescent probes: Specific recognition of cyanide through

different response patterns

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2.1. Materials and Measurements

All raw materials are obtained from suppliers and used directly. The melting points were determined using SGW X-4A Shanghai Jingke Micro Melting Point Apparatus (Shanghai, China). ¹H NMR/¹C NMR spectra were measured using a Bruker 500/126 MHz spectrometer with CDCl₃/DMSO-d₆ as the solvent (Bruck, Germany). The IR spectrum of the target compound was recorded on a Vertex 70 FT-IR spectrophotometer, and KBr was used as a tablet (Bruck, Germany). The absorption and emission spectra were measured on UV-3900 and F-7000 FL spectrophotometers (Tokyo, Japan), respectively.

Preparation of anhydrous tetrahydrofuran (THF) solvent: Under the protection of nitrogen, benzophenone was selected as the indicator and sodium metal was used to remove water and dry. When the solvent system turns dark blue, it was collected and used after distillation (now steamed and used now).

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Probe	LOD (µM)	Response method	Response time	solvent	Ref
	0.027	Ratiometric fluorescence	30 s	Tris-HCl, 10%DMSO	22(a)
Соон	0.0928	Fluorescence turn-on	-	HEPES	22(b)
N C C	0.066	Fluorescence turn-on	100 s	MeOH:HEPE, 1:1	22(c)
	0.017	Fluorescence turn-off	< 30s	DMF:H ₂ O 7:3	22(d)
	0.0772	Fluorescence turn-on	< 60 s	DMSO:H ₂ O, 8:2	22(e)
N O O OH N O O O Br	0.00032	fluorescence turn-off	12 min	DMSO:PBS (4:1, pH = 7.4)	22(f)
No. Co.o	1.0	Fluorescence turn-on	3 min	H ₂ O:CH ₃ CN, 7:3,	22(g)
	0.014	Fluorescence turn-on	240 s	THF:H ₂ O, 9:1	22(h)
HO HO HO HO HO HO HO HO HO HO HO HO HO H	0.0856	Ratiometric fluorescence turn-on	< 4s	DMSO-H ₂ O, 9:1	This Work
	0.0573	Fluorescence turn-on	< 5s	DMSO-H ₂ O, 9:1	This Work

Table S1. Comparison of reported fluorescent probes for CN⁻ with TPES1 and TPES2.



Fig. S1 ¹H NMR spectrum of compound TPE-CHO.



Fig. S2 ¹H NMR spectrum of compound TPE-2CHO.



Fig. S3 ¹H NMR spectrum of probe TPES1.



Fig. S4 ¹³C NMR of **TPES1** in CDCl₃. ¹³C NMR (500 MHz, Chloroform-d) δ 157.50, 156.00, 151.76, 148.43, 143.68, 143.48, 140.63, 139.64, 135.27, 134.48, 133.67, 132.47, 131.92, 131.35, 131.28, 131.23, 128.99, 128.32, 127.85, 127.71, 127.63, 127.48, 126.55, 126.50, 126.37, 123.53, 120.23, 118.81, 116.09, 115.70, 106.85, 77.21, 71.13, 71.08, 28.40.



Fig. S5. Mass spectrum of the probe TPES1.



Fig. S6 ¹H NMR spectrum of probe TPES2.



Fig. S7 ¹³C NMR of **TPES2** in CDCl₃. (500 MHz, CDCl₃) δ, 157.50, 154.18, 148.30, 147.82, 143.50, 143.45, 142.98, 141.21, 139.14, 135.27, 133.17, 132.42, 132.34, 131.92, 131.34, 131.25, 131.21, 131.15, 128.99, 128.31, 127.95, 127.89, 127.77, 127.65, 127.46, 126.65, 126.63, 126.44, 123.51, 120.21, 118.82, 117.90, 106.88, 77.27, 77.22, 77.02, 76.76, 71.36, 70.81, 28.57.



Fig. S8. Mass spectrum of the probe TPES2.



Fig. S9 (a) Response time and (b) fluorescence stability of probe TPES1 towards CN⁻.



Fig. S10 UV-Vis and fluorescence spectra of initial addition and titration end points.



Fig. S11 (a) **TPES2** (b) **TPES2** + 10 eq. CN-, (c) **TPES2** + 160 eq. CN⁻ dynamic light scattering experiments.



Fig. S12 (a) The relationship between the fluorescence intensity (473 nm) and the CN⁻ ion concentration (0-16.0 \times 10⁻⁴ M); (b) anti-interference experiment of probe PETS1 (473 nm).



Fig. S13 The binding constant curve after CN⁻ is added to TPES1



Fig. S14 The binding constant curve after CN⁻ is added to TPES2



Fig. S15 The fluorescence intensity of the probe (a)**TPSES1** and (b) **TPES2** changed with pH after adding CN⁻.



Fig. S16 (a) **TPES1** and **TPES1**+CN⁻, (b) **TPES2** and **TPES2**+CN- fitting curve of fluorescence lifetime



Fig. S17 The mass spectrum of probe TPES1 and CN⁻ after mixing for 15 minutes.



Fig. S18 The mass spectra (a) measured immediately; (b) after mixing for 15 minutes after probe TPES2 and CN^- are mixed.