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## Supporting information

## Carboxamide carbonyl-ruthenium(II) complexes: Detailed structural and mechanistic

## studies in the transfer hydrogenation of ketones

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### 4.3 Synthesis of the carboxamide ligands

### 4.3.1 N-(benzo[d]thiazol-2-yl)pyrazine-2-carboxamide (HL1)

Pyrazine-2-dicarboxylic acid $(1.00 \mathrm{~g}, 5.02 \mathrm{mmol})$ and 2-aminobenzothiazole $(0.75 \mathrm{~g}$, 5.02 mmol ) were dissolved in 20 mL pyridine and then heated with stirring for 15 minutes at $110{ }^{\circ} \mathrm{C}$. Triphenylphosphite $\mathrm{P}(\mathrm{OPh})_{3}(1.55 \mathrm{~g}, 5.00 \mathrm{mmol})$ was introduced drop-wise to the resulting solution and then allowed to stir at $90^{\circ} \mathrm{C}$ for 12 h . The crude product was poured into ice-cold water, filtered, and washed with cold water and cold methanol. The yellow crude powder was recrystallized from methanol and toluene. Yield: 1.54 g (74\%). ${ }^{1} \mathrm{H}$ NMR (400 $\mathrm{MHz}, d_{6}$-DMSO): $9.37\left(\mathrm{~s}, 1 \mathrm{H}_{\text {amidate }}\right), 8.98\left(\mathrm{~d},{ }^{3} J_{\mathrm{HH}}=2.4 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{pyz}}\right), 8.90-8.85\left(\mathrm{~m}, 1 \mathrm{H}_{\mathrm{pyz}}\right)$, $8.07\left(\mathrm{~d},{ }^{3} J_{\mathrm{HH}}=7.8 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{pyz}}\right), 7.84\left(\mathrm{~d},{ }^{3} J_{\mathrm{HH}}=8.0 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{bz}}\right), 7.50\left(\mathrm{t},{ }^{3} J_{\mathrm{HH}}=7.6 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{bz}}\right), 7.38$ $\left(\mathrm{t},{ }^{3} J_{\mathrm{HH}}=7.6 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{bz}}\right) .{ }^{13} \mathrm{C}$ NMR ( $\left.101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) \delta: 161.3(\mathrm{C}$-carbonyl), 156.8(C-pyrazine), 148.6(C-pyrazine), 145.0(C-pyrazine), 143.1(C-pyrazine), 142.5(C-benzothiole), 132.5(Cbenzothiole), 126.5(C-benzothiole), 124.4(C-benzothiole), 121.5(C-benzothiole), 121.4(Cbenzothiole). FT-IR spectrum (Zn-Se ATR, cm ${ }^{-1}$ ): $3324(\mathrm{~N}-\mathrm{H}), 1691$ (C=O), 1533 (C=N). MS spectrum (m/z): Calcd. 510.04; Found $511.08\left(2 \mathrm{M}^{+}-\mathrm{H}\right)$. Anal. Cald. for: $\mathrm{C}_{12} \mathrm{H}_{8} \mathrm{~N}_{4} \mathrm{OS}$ : C, 56.24; H, 3.15; N, 21.86; S, 12.51\%. Found: C, 56.11; H, 3.52; N, 21.64; S, 12.11\%.

### 4.3.2 N-(1H-benzo[d]imidazol-2-yl)pyrazine-2-carboxamide (HL2)

Pyrazine-2-dicarboxylic acid ( $1.00 \mathrm{~g}, 5.00 \mathrm{mmol}$ ), 2-aminobenzothiazole ( $0.75 \mathrm{~g}, 5.00$ $\mathrm{mmol})$, and $\mathrm{P}(\mathrm{OPh}) 3(1.55 \mathrm{~g}, 5.00 \mathrm{mmol})$. Recrystallization was achieved from methanol to obtain a pale-yellow solid. Yield: $1.04 \mathrm{~g}(72 \%) .{ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{DMSO}$ ) $\delta 12.20$ (s, $\left.1 \mathrm{H}_{\text {amidate }}\right), 9.37\left(\mathrm{~d},{ }^{3} \mathrm{~J}_{\mathrm{HH}}=1.2 \mathrm{~Hz}, 1 \mathrm{H}_{\mathrm{pyz}}\right), 8.86\left(\mathrm{~d},{ }^{3} J_{\mathrm{HH}}=35.5,1.9 \mathrm{~Hz}, 2 \mathrm{H}_{\mathrm{pyz}}\right), 7.51\left(\mathrm{~d},{ }^{3} J_{\mathrm{HH}}=\right.$ $5.9,3.2 \mathrm{~Hz}, 2 \mathrm{H}_{\mathrm{bz}}$ ), $7.18\left(\mathrm{dd},{ }^{3} \mathrm{~J}_{\mathrm{HH}}=5.9,3.2 \mathrm{~Hz}, 2 \mathrm{H}_{\mathrm{bz}}\right) .{ }^{13} \mathrm{C}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $\delta$ : $163.2\left(\mathrm{C}_{\text {carbonyl }}\right), \quad 149.9\left(\mathrm{C}_{\text {pyrazine }}\right), \quad 149.0\left(\mathrm{C}_{\text {pyrazine }}\right), \quad 138.6\left(\mathrm{C}_{\text {pyrazine }}\right), \quad 131.5\left(\mathrm{C}_{\text {pyrazine }}\right)$,
127.6( $\left.\mathrm{C}_{\text {benzoimidazole }}\right), \quad 126.1\left(\mathrm{C}_{\text {benzoimidazole }}\right), \quad 126.5\left(\mathrm{C}_{\text {benzoimidazole }}\right), \quad 127.6\left(\mathrm{C}_{\text {benzoimidazole }}\right)$, 126.1 ( $\mathrm{C}_{\text {benzoimidazole }}$ ), 122.9( $\left.\mathrm{C}_{\text {benzoimidazole }}\right)$. FT-IR spectrum ((Zn-Se ATR, $\left.\mathrm{cm}^{-1}\right)$ : $3251(\mathrm{~N}-\mathrm{H})$, $1684(\mathrm{C}=\mathrm{O}), 1546(\mathrm{C}=\mathrm{N})$. MS spectrum, m/z: calcd. 239.08; found $240.07\left(\mathrm{M}^{+}+\mathrm{H}\right)$. Anal. Cald. for: $\mathrm{C}_{12} \mathrm{H}_{9} \mathrm{~N}_{5} \mathrm{O}: \mathrm{C}, 60.21$; H, 3.79; N, 29.28; O, 6.69; Found: C, 60.11; H, 3.52; N, 29.64.

Table S1. Crystal data and structure refinement for complexes Ru1, Ru2 and Ru4

|  | Ru1 | Ru2 | Ru4 |
| :---: | :---: | :---: | :---: |
| Empirical formula | $\mathrm{C}_{99} \mathrm{H}_{75} \mathrm{Cl}_{4} \mathrm{~N}_{8} \mathrm{O}_{4} \mathrm{P}_{4} \mathrm{Ru}_{2} \mathrm{~S}_{2}$ | $\mathrm{C}_{50} \mathrm{H}_{40} \mathrm{Cl}_{2} \mathrm{~N}_{4} \mathrm{O}_{2} \mathrm{P}_{2} \mathrm{RuS}$ | $\mathrm{C}_{49} \mathrm{H}_{39} \mathrm{Cl}_{2} \mathrm{~N}_{5} \mathrm{O}_{2} \mathrm{P}_{2} \mathrm{Ru}$ |
| Formula weight | 1972.61 | 994.83 | 963.76 |
| Temperature | 100(2) K | 100(2) K | 100(2) K |
| Wavelength | 0.71073 Å | 0.71073 Å | $0.71073 \AA$ |
| Crystal <br> system | Monoclinic | Monoclinic | Monoclinic |
| Space group | P 21/c | P 21/n | P $21 / \mathrm{n}$ |
| Unit cell | $\mathrm{a}=21.3269(17) \AA$ | $\mathrm{a}=12.1017(6) \AA$ | $\mathrm{a}=9.8144(6) \AA$ |
| dimensions | $\mathrm{b}=12.2526(10) \AA$ | $\mathrm{b}=13.8083(7) \AA$ | $\mathrm{b}=18.5959(12) \AA$ |
|  | $\mathrm{c}=37.414(3) \AA$ | $c=27.2988(14) \AA$ | $c=23.1600(15) \AA$ |
|  | $\alpha=90^{\circ}$ | $\alpha=90^{\circ}$ |  |
|  | $\beta=101.929(2)^{\circ}$ | $\beta=101.929(2)^{\circ}$ | $\beta=94.309(3)^{\circ}$ |
|  | $\gamma=90^{\circ}$ | $\gamma=90^{\circ}$ | $\gamma=90^{\circ}$ |
| Volume | $9685.9(14) \AA^{3}$ | 4463.2(4) $\AA^{3}$ | 4214.9(5) $\AA^{3}$ |
| Z | 4 | 4 | 4 |
| Density (calculated) | $1.353 \mathrm{Mg} / \mathrm{m}^{3}$ | $1.480 \mathrm{Mg} / \mathrm{m}^{3}$ | $1.519 \mathrm{Mg} / \mathrm{m}^{3}$ |
| Absorption coefficient | $0.585 \mathrm{~mm}^{-1}$ | $0.636 \mathrm{~mm}^{-1}$ | $0.623 \mathrm{~mm}^{-1}$ |
| F(000) | 4020 | 2032 | 1968 |
| $\begin{aligned} & \text { Crystal size } \\ & \left(\mathrm{mm}^{3}\right) \end{aligned}$ | $0.260 \times 0.200 \times 0.140$ | $0.23 \times 0.14 \times 0.08$ | $0.640 \times 0.440 \times 0.240$ |
| Theta range for data collection | 1.099 to $28.300^{\circ}$ | 1.525 to $28.165^{\circ}$ | 1.406 to $28.291^{\circ}$ |
| Index ranges | $-28<=\mathrm{h}<=28$, | $-16<=h<=15$ | $-13<=\mathrm{h}<=13$, |
|  | $\begin{aligned} & -16<=k<=16 \\ & -49<=1<=49 \end{aligned}$ | $\begin{aligned} & -18<=\mathrm{k}<=16, \\ & -36<=1<=36 \end{aligned}$ | $\begin{aligned} & -24<=k<=22, \\ & -30<=1<=30 \end{aligned}$ |
| Reflections collected | 176176 | 78016 | 76052 |
| Independent reflections | $\begin{aligned} & 24094[\mathrm{R}(\text { int })= \\ & 0.0515] \end{aligned}$ | $10946[\mathrm{R}(\mathrm{int})=0.0335]$ | $\begin{aligned} & 10451[\mathrm{R}(\mathrm{int})= \\ & 0.0332] \end{aligned}$ |
| Completeness to theta | 100.0 \% | 100.0 \% | 99.9 \% |
| Data restraints parameters | 23990 / 0 / 1108 | 10882 / 0 / 563 | 10403 / 0 / 554 |
| Goodness-offit on $\mathrm{F}^{2}$ | 1.061 | 1.602 | 1.047 |
| Final indices [I>2sigma(I)] | $\begin{aligned} & \mathrm{R} 1=0.0638, \mathrm{wR} 2= \\ & 0.1935 \end{aligned}$ | $\begin{aligned} & \mathrm{R} 1=0.0269 \\ & \mathrm{wR} 2=0.0627 \end{aligned}$ | $\begin{aligned} & \mathrm{R} 1=0.0410, \\ & \mathrm{wR} 2=0.1157 \end{aligned}$ |
| R indices (all data) | $\begin{aligned} & \mathrm{R} 1=0.0779 \\ & \mathrm{wR} 2=0.2063 \end{aligned}$ | $\begin{aligned} & \mathrm{R} 1=0.0324, \\ & \mathrm{wR} 2=0.0691 \end{aligned}$ | $\begin{aligned} & \mathrm{R} 1=0.0445, \\ & \mathrm{wR} 2=0.1186 \end{aligned}$ |


| Largest diff. | 4.897 and -2.779 | 0.645 and -0.664 e..$\AA^{-3}$ | 1.803 and -1.997 |
| :--- | :--- | :--- | :--- |
| peak and hole | e. $\AA^{-3}$ |  | e. $\AA^{-3}$ |

Table S2. Selected bond lengths $(\AA)$ and bond angles $\left({ }^{\circ}\right)$ for Ru1, Ru2 and Ru4

| Ru1 | Ru2 |  | Ru4 |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- |
| $\mathrm{Ru}(1)-\mathrm{C}(49)$ | $1.984(7)$ | $\mathrm{Ru}(1)-\mathrm{C}(13)$ | $1.8519(18)$ | $\mathrm{Ru}(1)-\mathrm{C}(13)$ | $1.984(7)$ |
| $\mathrm{Ru}(1)-\mathrm{N}(1)$ | $2.113(3)$ | $\mathrm{Ru}(1)-\mathrm{N}(1)$ | $2.1180(14)$ | $\mathrm{Ru}(1)-\mathrm{N}(1)$ | $2.113(3)$ |
| $\mathrm{Ru}(1)-\mathrm{N}(2)$ | $2.115(3)$ | $\mathrm{Ru}(1)-\mathrm{N}(2)$ | $2.116(14)$ | $\mathrm{Ru}(1)-\mathrm{N}(2)$ | $2.115(3)$ |
| $\mathrm{Ru}(1)-\mathrm{P}(1)$ | $2.4000(11)$ | $\mathrm{Ru}(1)-\mathrm{P}(1)$ | $2.3636(4)$ | $\mathrm{Ru}(1)-\mathrm{P}(1)$ | $2.4000(11)$ |
| $\mathrm{Ru}(1)-\mathrm{Cl}(1)$ | $2.4258(10)$ | $\mathrm{Ru}(1)-\mathrm{H}(1 \mathrm{~A})$ | $1.55(2)$ | $\mathrm{Ru}(1)-\mathrm{H}(1 \mathrm{~A})$ | $1.9672(10)$ |
| Ru 1 | Ru 2 | Ru |  |  |  |
| $\mathrm{C}(49)-\mathrm{Ru}(1)-\mathrm{N}(1)$ | $176.32(16)$ | $\mathrm{C}(13)-\mathrm{Ru}(1)-\mathrm{N}(1)$ | $177.73(7)$ | $\mathrm{C}(13)-\mathrm{Ru}(1)-\mathrm{N}(1)$ | $100.81(10)$ |
| $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{N}(2)$ | $77.84(13)$ | $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{N}(2)$ | $76.13(5)$ | $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{N}(3)$ | $76.24(8)$ |
| $\mathrm{C}(49)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $89.04(13)$ | $\mathrm{C}(13)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $90.76(5)$ | $\mathrm{C}(13)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $89.05(8)$ |
| $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $91.12(9)$ | $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $88.31(4)$ | $\mathrm{N}(1)-\mathrm{Ru}(1)-\mathrm{P}(2)$ | $95.06(6)$ |
| $\mathrm{P}(2)-\mathrm{Ru}(1)-\mathrm{P}(1)$ | $176.88(4)$ | $\mathrm{P}(2)-\mathrm{Ru}(1)-\mathrm{P}(1)$ | $165.302(16)$ | $\mathrm{P}(2)-\mathrm{Ru}(1)-\mathrm{P}(1)$ | $168.60(2)$ |



Figure S1. ${ }^{1} \mathrm{H}$ NMR spectrum ( 400 MHz , 298K $d_{6}$-DMSO) of 1 N -(benzo[d]thiazol-2-yl)pyrazine-2-carboxamide ligand, HL1 showing the diagnostic signal at $\delta_{\mathrm{H}}: 12.68 \mathrm{ppm}$ and all key signals between $\delta_{\mathrm{H}}$ : 7.37-9.37 ppm.


Figure S2. ${ }^{1} \mathrm{H}$ NMR spectrum ( 400 MHz , $298 \mathrm{~K} d_{6}$-DMSO) of N -( 1 H -benzo[d]imidazol-2-yl)pyrazine-2-carboxamide ligand, HL2 showing the signature peak at $\delta_{\mathrm{H}}: 12.20 \mathrm{ppm}$ and all key peaks between $\delta_{\mathrm{H}}: 7 \cdot 17-9.37 \mathrm{ppm}$.




Figure $\mathrm{S} 3 .{ }^{1} \mathrm{H}$ NMR spectrum ( $400 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of complex, Ru1 showing the signature peak $\left(\mathrm{H}_{\mathrm{pyrazine}}\right)$ shifting from $\delta_{\mathrm{H}}: 9.37$ to $\delta 9.15 \mathrm{ppm}$ upfield.


Figure $\mathrm{S} 4 .{ }^{1} \mathrm{H}$ NMR spectrum $\left(400 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}\right)$ of complex $\mathbf{R u} 2$ showing triplet peak of co-ligand $\left(\mathrm{H}^{-}\right)$ranging between $\delta_{\mathrm{H}}:-13.08-13.16 \mathrm{ppm}$.




Figure S5. ${ }^{1} \mathrm{H}$ NMR spectrum ( $400 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of complex Ru3 indicating shift of signals corresponding to $\mathrm{H}_{\mathrm{N}-\mathrm{H}}\left(\delta_{\mathrm{H}}: 12.20\right.$ to 11.89 ppm ) and $\mathrm{H}_{\mathrm{pyrazine}}\left(\delta_{\mathrm{H}}: 9.37\right.$ to 9.08 ppm$)$ which confirm successful coordination.


Figure S6. ${ }^{1} \mathrm{H}$ NMR spectrum ( $400 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of complex Ru4 indicating triplet peak of co-ligand $\left(\mathrm{H}^{-}\right)$ranging between $\delta_{\mathrm{H}}$ : -13.08-13.16 ppm and shifts in $\mathrm{H}_{\text {pyrazine }}$ (from $\delta_{\mathrm{H}}$ : 9.20 to 9.24 ppm ) and $\mathrm{H}_{\mathrm{N}-\mathrm{H}}\left(\right.$ from $\delta_{\mathrm{H}}: 12.08$ to 12.02 ppm ) proton signals confirming formation the compound.




Figure $\mathrm{S} 7 .{ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum ( 100 MHz , $298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of N -(benzo[d]thiazol-2-yl)pyrazine-2-carboxamide ligand, HL1 showing the $\mathrm{C}=\mathrm{O}$ signal at $\delta$ : 161.3 ppm confirming the successful formation of the ligands.



Figure S8. ${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum of N -( 1 H -benzo[d]imidazol-2-yl)pyrazine-2-carboxamide ligand, HL2 showing the $\mathrm{C}=\mathrm{O}$ signal at $\delta: 161.3 \mathrm{ppm}$ confirming the successful formation of the ligands.



Figure $\mathrm{S} 9 .{ }^{13} \mathrm{C}\{1 \mathrm{H}\}$ NMR spectrum ( 100 MHz , $298 \mathrm{~K} \mathrm{~d}_{-} \mathrm{CDCl}_{3}$ ) of complex Ru1 showing signal at $\delta=166.88 \mathrm{ppm}$ corresponding to the $\mathrm{C}=\mathrm{O}$ signal confirming the successful coordination.




Figure S10. ${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum ( $100 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of complex $\mathbf{R u 2}$ indicating a shift in the chemical shift of $\mathrm{C}=\mathrm{O}(\delta: 166.81 \mathrm{ppm})$ downfield upon coordination.


Figure $\mathrm{S} 11 .{ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum $\left(100 \mathrm{MHz}, 298 \mathrm{~K} d-\mathrm{CDCl}_{3}\right)$ of complex $\mathbf{R u} 3$ indicating a shift in the chemical shift of $\mathrm{C}=\mathrm{O}(\delta: 166.81 \mathrm{ppm})$ downfield upon coordination.


Figure $\mathrm{S} 12 .{ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum $\left(100 \mathrm{MHz}\right.$, $298 \mathrm{~K} d-\mathrm{CDCl}_{3}$ ) of complex Ru4 indicating a shift in the chemical shift of $\mathrm{C}=\mathrm{O}(\delta: 166.81 \mathrm{ppm})$ downfield upon coordination.


Figure $\mathrm{S} 13 .{ }^{31} \mathrm{P}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum ( 100 MHz , $298 \mathrm{~K} d_{6} \mathrm{CDCl}_{3}$ ) of complex Ru1 indicating a signal corresponding to two equivalent $\mathrm{PPh}_{3}$ trans to each other.
Ru-2 -31P



Figure S14. ${ }^{31} \mathrm{P}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum of $\left(100 \mathrm{MHz}, 298 \mathrm{~K} d_{6} \mathrm{CDCl}_{3}\right)$ complex $\mathbf{R u 2}$ indicating a signal corresponding to two equivalent $\mathrm{PPh}_{3}$ trans to each other.



Figure $\mathrm{S} 15 .{ }^{31} \mathrm{P}\left\{{ }^{1} \mathrm{H}\right\}$ NMR spectrum ( $100 \mathrm{MHz}, 298 \mathrm{~K} d_{6} \mathrm{CDCl}_{3}$ ) of complex Ru3 showing a signal at $\delta: 42.74 \mathrm{ppm}$ confirming to two equivalent $\mathrm{PPh}_{3}$ trans to each other.

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31P NMR of Ru4 in CDCl3


Figure S16. \({ }^{31} \mathrm{P}\left\{{ }^{1} \mathrm{H}\right\}\) NMR spectrum ( 122 MHz , \(298 \mathrm{~K} d-\mathrm{CDCl}_{3}\) ) of complex Ru4 showing a signal at \(\delta: 48.33 \mathrm{ppm}\) implicating presence of two equivalent \(\mathrm{PPh}_{3}\) trans to each other.


Figure S17. FT-IR spectrum of N -(benzo[d]thiazol-2-yl)pyrazine-2-carboxamide ligand, HL1 showing its characteristic signals of \(\mathrm{v}(\mathrm{N}-\mathrm{H}): 3324 \mathrm{~cm}^{-1}, 1691\); \((\mathrm{C}=\mathrm{O}): 1691 \mathrm{~cm}^{-1} ; 1543\) \(v(C=N)\) :


Figure S18. FT-IR spectrum of \(N\)-(benzo[d]imidazol-2-yl)pyrazine-2-carboxamide ligand, HL2 depicting the characteristic signals of \(v(N-H): 3251 \mathrm{~cm}^{-1}, v(\mathrm{C}=\mathrm{O}): 1684 \mathrm{~cm}^{-1}, 1546 \mathrm{~cm}^{-}\) \({ }^{1} v(C=N)\).


Figure S19. FT-IR spectrum of complex Ru1 depicting signature signals at: \(1935 \mathrm{~cm}^{-1}, \nu \mathrm{C} \equiv \mathrm{O}\) : and \(1629 \mathrm{~cm}^{-1}, 1565 \mathrm{~cm}^{-1} v(\mathrm{C}=\mathrm{N})\).


Figure S20. FT-IR spectrum of complex Ru2 depicting signature signals: \(1935 \mathrm{~cm}^{-1} \nu \mathrm{C} \equiv \mathrm{O}\) : \(1626 \mathrm{~cm}^{-1}, v(\mathrm{C}=\mathrm{O}) ; 1567 \mathrm{v}(\mathrm{C}=\mathrm{N})\).


Figure S21. FT-IR spectrum of complex Ru3 depicting signature signals at: \(1947 \mathrm{~cm}^{-1} v(\mathrm{C} \equiv \mathrm{O})\); \(1622 \mathrm{~cm}^{-1} v(\mathrm{C}=\mathrm{O})\); and \(1562 \mathrm{~cm}^{-1} v(\mathrm{C}=\mathrm{N})\).


Figure S22. FT-IR spectrum of complex Ru4 depicting signature signals at \(1946 \mathrm{~cm}^{-1} v(\mathrm{C} \equiv \mathrm{O})\); \(1612 \mathrm{~cm}^{-1} v(\mathrm{C}=\mathrm{O})\); and \(1569 \mathrm{~cm}^{-1} v(\mathrm{C}=\mathrm{N})\).


Figure S23. LR MS spectrum (positive ion mode) of HL1 indicating dimeric form of the ligand at \(\mathrm{m} / \mathrm{z}=511.07\). The theoretical isotopic mass distribution plot (inset).


Figure S24. LR MS spectrum (positive ion mode) of HL2 depicting a peak at \(\mathrm{m} / \mathrm{z}=242\) corresponding to \([\mathrm{M}+\mathrm{H}]\). The theorical isotopic mass distribution plot (inset).


Figure S25. LC-MS spectrum of Ru1. The simulated theoretical mass distribution plot (inset).


Figure S26. LC-MS spectrum of Ru2. The simulated theoretical mass distribution plot (inset).


Figure S27. LC-MS spectrum of Ru3. The simulated theoretical mass distribution plot (inset).


Figure S28. LC-MS spectrum of Ru4 indicating the molecular mass of the fragment [M-H] at \(\mathrm{m} / \mathrm{z}=892.12\). Inset is the simulated theoretical mass distribution plot.


Figure S29. \({ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of acetophenone reaction mixture (Ru3 was used as catalyst). Aliquot withdrawn and analysed after 4 h of reaction. The integral values of methyl peaks of acetophenone and 1-phenylethanol corresponding to percentage conversions of \(50 \%\) and yields of \(50 \%\).


Figure S30. \({ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of TH of acetophenone. (Ru4 was used as catalyst). Aliquot was taken and analysed after 6 h of reaction. The integral values of methyl protons of acetophenone and 1-phenylethanol corresponding to conversion of \(97 \%\) and Yield of \(97 \%\).


Figure \(\mathrm{S} 31 .{ }^{1} \mathrm{H}\) NMR spectrum \(\left(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\right)\) of the reaction of transfer hydrogenation of acetophenone (Ru4 used as catalyst). Aliquot was taken and analysed after 6 h of reaction. The integral values of methyl protons of acetophenone and 1-phenylethanol correspond to conversion of \(91 \%\) and yield of \(90 \%\).


Figure \(\mathrm{S} 32 .{ }^{1} \mathrm{H}\) NMR spectrum \(\left(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\right)\) of the reaction of transfer hydrogenation of acetophenone (Ru3:catalyst loading of \(0.25 \mathrm{~mol} \%\) ). Aliquot sampled and analysed after 4 \(h\) of reaction. The integral values of the methyl protons of acetophenone and 1-phenylethanol correspond to percentage conversion of \(46 \%\) and yield of \(41 \%\).


Figure S33. \({ }^{1} \mathrm{H}\) NMR spectrum \(\left(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\right)\) of the reaction of transfer hydrogenation of acetophenone (Ru1: catalyst loading of \(0.25 \mathrm{~mol} \%\) ). Aliquot was sampled and analysed after 1 h of reaction. The integral values of the methyl protons of acetophenone and 1phenylethanol correspond to the percentage conversion of \(27 \%\) and yield of \(27 \%\) (Figure S47).


Figure S34. \({ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of 4chloroacetophenone reaction mixture (Ru4 used as catalyst). Aliquot taken and analysed after at 4 h reaction. The integral values of the methyl protons of 4-chloroacetophenone and 4chlorophenylethanol correspond to the percentage conversion of \(98 \%\) and yield of \(98 \%\) (Table 4, entry 3 ).


Figure \(\mathrm{S} 35 .{ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of 2acetylpyridine reaction mixture (Ru4 used as catalyst). Aliquot taken and analysed after at 4h reaction. The integral values of the methyl protons of 2-acetylpyridine and product correspond to the percentage conversion of \(55 \%\) and yield of \(55 \%\) (Table 4, entry 11).


Figure \(\mathrm{S} 36 .{ }^{1} \mathrm{H}\) NMR spectrum \(\left(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\right)\) of transfer hydrogenation of 2-propanone reaction mixture (Ru4 used as catalyst). Aliquot taken and analysed after at 4 h reaction. The integral values of the methyl protons of 2-propanone and 2-propanol correspond to the percentage conversion of \(71 \%\) and yield of \(71 \%\) (Table 4, entry 11).


Figure S37. \({ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of 1acetylnaphthone reaction mixture (Ru4 used as catalyst). Aliquot taken and analysed after at 6 h reaction. The integral values of the methyl protons of 1-acetylnaphthone and product correspond to the percentage conversion of \(86 \%\) and yield of \(84 \%\) (Table 4 , entry 9).


Figure \(\mathrm{S} 38 .{ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of acetophenone reaction catalysed by Ru3 without a base. Aliquot taken and analysed after at 36 h of reaction. The integral values of the methyl protons of 1-acetylnaphthone and product correspond to the percentage conversion of \(99 \%\) and yield of \(99 \%\) (Table 2, entry 5).


Figure \(\mathrm{S} 39 .{ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of acetophenone reaction catalysed by Ru4 without a base. Aliquot taken and analysed after at 18 h of reaction. The integral values of the methyl protons of 1-acetylnaphthone and product correspond to the percentage conversion of \(93 \%\) and yield of \(93 \%\) (Table 2, entry 4).


Figure \(\mathrm{S} 40 .{ }^{1} \mathrm{H}\) NMR spectrum \(\left(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\right)\) of transfer hydrogenation of acetophenone reaction (without a catalyst). Aliquot taken and analysed after at 36 h of reaction. The integral values of the methyl protons of 1-acetylnaphthone and product correspond to the yield of \(42 \%\) (Table 2, entry 2 ).


Figure S41. \({ }^{1} \mathrm{H}\) NMR spectrum of isolated product (1-phenylethanol) from TH of acetophenone. Yield \(=0.11(79 \%){ }^{1} \mathrm{H}\) NMR spectrum \(\left(d-\mathrm{CDCl}_{3}\right): \delta(\mathrm{ppm}) 1.49\left(\mathrm{~d},{ }^{3} \mathrm{~J}_{\mathrm{H}-\mathrm{H}}=6.8\right.\) \(\left.\mathrm{Hz}, 3 \mathrm{H}\left(\mathrm{CH}_{3}\right)\right), 4.89\left(\mathrm{q},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=6.4 \mathrm{~Hz}, 1 \mathrm{H}(\mathrm{CH})\right), 4.85(\mathrm{~s}, 1 \mathrm{H}(\mathrm{OH})), 7.30-7.39(\mathrm{~m}\), cluster \(5 \mathrm{H}_{\text {arom }}\) ).


Figure S42. \({ }^{1} \mathrm{H}\) NMR spectrum of isolated product 1-(4-chloroPhenylethanol) of TH of 4chloroacetophemnone. 4-chloroacetophenone. Yield \(0.15 \mathrm{~g}(97 \%)={ }^{1} \mathrm{H}\) NMR \(\operatorname{spectrum}(d-\) \(\left.\mathrm{CDCl}_{3}\right): \delta(\mathrm{ppm}) 1.49\left(\mathrm{~d},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=6.8 \mathrm{~Hz}, 3 \mathrm{H}\left(\mathrm{CH}_{3}\right)\right), 4.87\left(\mathrm{q},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=6.8 \mathrm{~Hz}, 1 \mathrm{H}(\mathrm{CH})\right)\), and 7.32 (m, cluster \(4 \mathrm{H}_{\text {arom }}\) ).

1-(naphthalen-1-yI)ethanol(Isolated product)in CDCI 3


Figure S43. \({ }^{1} \mathrm{H}\) NMR spectrum of isolated product (1-naphthalen-1-yl)ethanol) from TH of 1acetylnaphthalenone. 1-(naphthalen-1-yl)ethan-1-ol. Yield 0.14 g (86\%) \(={ }^{1} \mathrm{H}\) NMR \(\operatorname{spectrum}\left(d-\mathrm{CDCl}_{3}\right): \delta(\mathrm{ppm}) 1.63\left(\mathrm{~d},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=6.4 \mathrm{~Hz}, 3 \mathrm{H}\left(\mathrm{CH}_{3}\right)\right), 4.98\left(\mathrm{q},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=6.4 \mathrm{~Hz}\right.\), \(1 \mathrm{H}(\mathrm{CH})\) ), 7.49-7.55 (m, cluster \(2 \mathrm{H}_{\text {arom }}\) ), \(7.84-7.88\left(\mathrm{~m}\right.\), cluster, \(\left.4 \mathrm{H}_{\text {arom }}\right)\).


Figure S44. \({ }^{1} \mathrm{H}\) NMR spectrum of isolated product (2-propanol) from TH of 2-propanone. 2propanol. Yield \(=0.06 \mathrm{~g}(67 \%) .{ }^{1} \mathrm{H}\) NMR spectrum \(\left(d-\mathrm{CDCl}_{3}\right): \delta(\mathrm{ppm}) 0.89\left(\mathrm{~d},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=1.6 \mathrm{~Hz}\right.\), \(3 \mathrm{H}(\mathbf{C H})), 1.23\left(\mathrm{t},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=1.6 \mathrm{~Hz}, 3 \mathrm{H}(\mathrm{CH})\right), 1.31\left(\mathrm{~m},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=1.6 \mathrm{~Hz}, 2 \mathrm{H}(\mathbf{C H})\right), 1.40\left(\mathrm{~d},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=\right.\) \(1.6 \mathrm{~Hz}, 2 \mathrm{H}(\mathrm{CH})), 4.03\left(\mathrm{~m},{ }^{3} J_{\mathrm{H}-\mathrm{H}}=1.6 \mathrm{~Hz}, 1 \mathrm{H}(\mathrm{CH})\right), 4.89(\mathrm{~s}, 1 \mathrm{H}(\mathrm{OH}))\).


Figure S 45 . Base free TH of acetophenone catalysed by Ru4 reaction, time, 6 h corresponding to percentage conversion of \(39 \%\) (Table 2 , entry \(3^{f}\) ).


Figure \(\mathrm{S} 46 .{ }^{1} \mathrm{H}\) NMR spectrum ( \(400 \mathrm{MHz}, d-\mathrm{CDCl}_{3}\) ) of transfer hydrogenation of acetophenone reaction catalysed by Ru3 with base loading of \(100 \mathrm{~mol} \%\). Aliquot taken and analysed after at 18 h of reaction. The integral values of the methyl protons of 1acetylnaphthone and product correspond to the percentage conversion of \(97 \%\) (Table 2, entry 10).


Figure S47. Plots of percentage of conversion vs time showing the effects of catalyst loading on the catalytic activity on transfer hydrogenation of acetophenone reaction using of Ru3 as a catalyst.


Figure S48. Plots of turnover number (TON) vs time showing the effects of catalyst loading on the catalytic activity of TH of acetophenone reaction using of \(\mathbf{R u 3}\) as a catalyst. Reaction conditions: acetophenone, \(1.00 \mathrm{mmol} ; \mathrm{K}^{\ell} \mathrm{BuO}, 0.100 \mathrm{mmol} ;{ }^{\mathrm{i}} \mathrm{PrOH}, 5 \mathrm{ml} ; 82^{\circ} \mathrm{C}\), time, 6 h . Percentage Conversion and yield determined by NMR spectroscopy (average of two independent runs), methoxybenzene was used as internal standard.


Figure S49. The (a) plot of In[acetophenone \(]_{t}[\text { acetophenone }]_{o}\) vs \(\operatorname{In}[t]\) for determination of the rate constants of each catalyst in TH of acetophenone reaction. Condition: acetophenone, 1.00 \(\mathrm{mmol} ;{ }^{\mathrm{i}} \mathrm{PrOH}, 5 \mathrm{ml} ; 82^{\circ} \mathrm{C}\), time, 6 h . Percentage conversions were determined by NMR spectroscopy (average of two independent runs), methoxybenzene was used as internal standard.```

