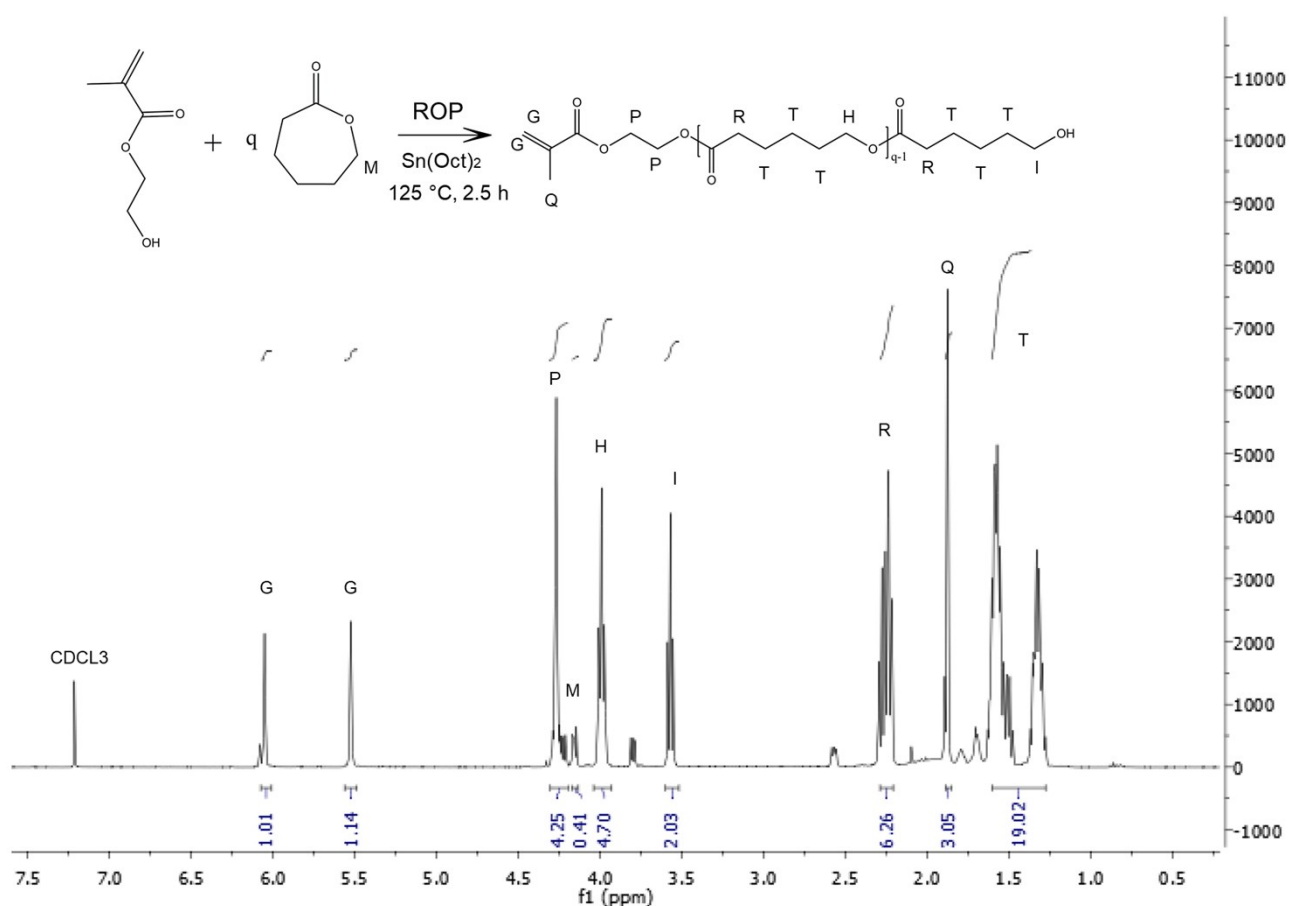


Supporting Information for

# Limonene-in-Water Pickering Emulsion and On-Demand Separation Using Thermo-Responsive Biodegradable Nanoparticles

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**Figure S1:** <sup>1</sup>H-NMR characterization and proton assignment for HEMACL3 ( $q=3$ ). The analysis was performed on a Bruker Ultrashield 400 MHz spectrometer using deuterated chloroform ( $\text{CDCl}_3$ ) as solvent.

The monomer conversion ( $X_{CL}$ ) was calculated according to eq. S1:

$$X_{CL} = \left( \frac{H + I}{H + I + M} \right) * 100 \quad (\text{S1})$$

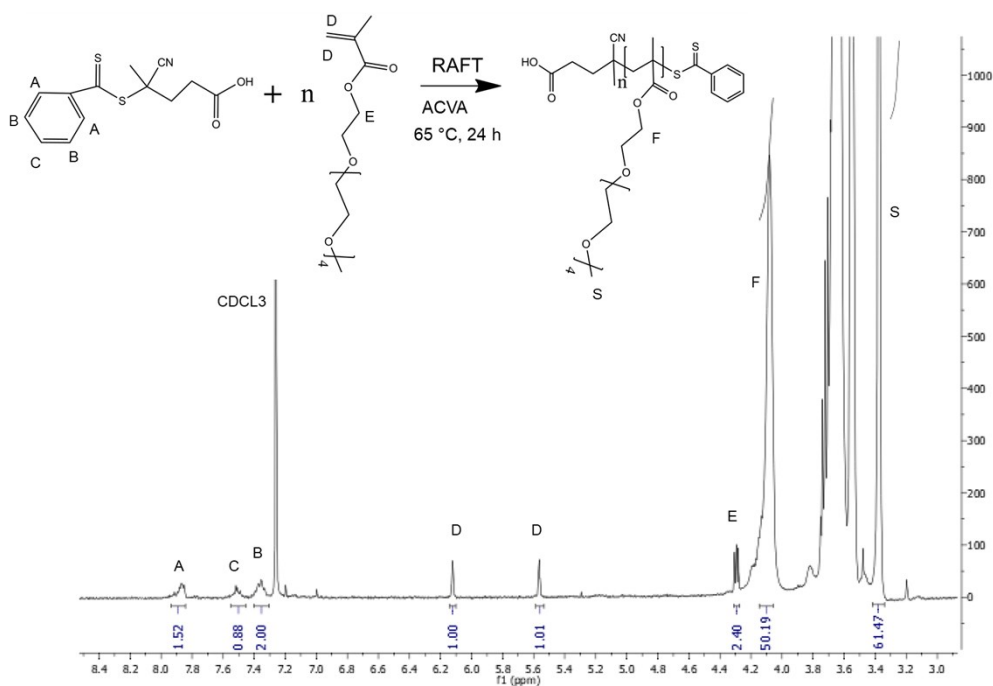
Where M represents the area under the signal of the two hydrogens adjacent to the ester group in the CL monomer, H indicates the peaks equivalent to the two hydrogen atoms adjacent to the ester group and I corresponds to hydrogens of the carbon near the chain-end hydroxyl group.

The degree of polymerization (q) was obtained using the area of the same signals according to **eq. S2**:

$$q = \frac{H}{I} + 1 \quad (\text{S2})$$

**Table S1:** *Properties of the oligoesters synthesized. The  $\epsilon$ -caprolactone conversion ( $X_{CL}$ ) and average macromonomer chain length ( $q$ ) were determined via NMR according to eqs. S1 and S2. The number-average molecular weight ( $M_n$ ) and dispersity ( $\mathcal{D}$ ) were obtained via GPC.*

<b>Sample</b>	$X_{CL}$ [%]	$q$ [-]	<b><math>M_n</math></b> [Da]	<b><math>\mathcal{D}</math></b> [-]
HEMACL3	94.25	3.31	577	1.25
HEMACL5	98.91	5.24	858	1.22



**Figure S2:**  $^1\text{H-NMR}$  characterization and proton assignment for  $20\text{EG}_4$  ( $n=20$ ). The analysis was performed on a Bruker Ultrashield 400 MHz spectrometer using deuterated chloroform ( $\text{CDCl}_3$ ) as solvent.

The monomer conversion ( $X_{EG_4}$ ) was calculated according to **eq. S3**:

$$X_{EG_4} = \left( \frac{F}{E + F} \right) * 100 \quad (\text{S3})$$

Where E and F are the area of the peaks attributed to the two hydrogens close to the oxygen in the unreacted monomer and polymer, respectively.

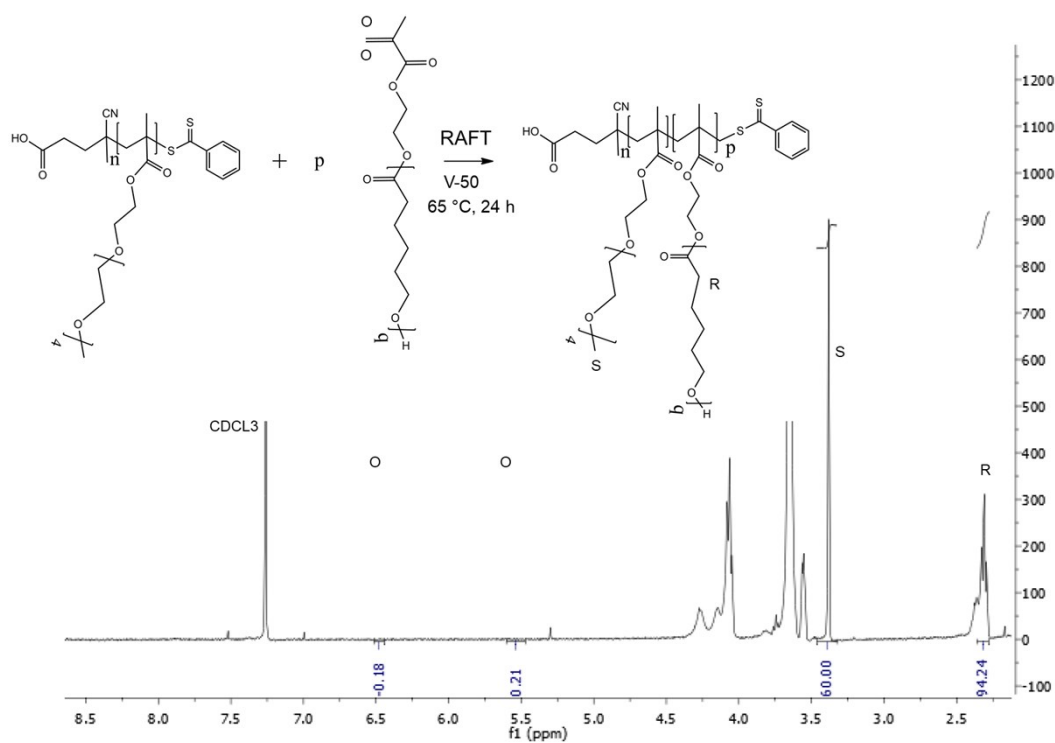
The degree of polymerization ( $n$ ) was calculated with **eq. S4**:

$$n = \frac{F}{B} \quad (\text{S4})$$

Where B is associated to the two hydrogens of the aromatic ring in the chain transfer agent.

**Table S2:** Properties of the macro CTAs synthesized. The monomer conversion ( $X_{EG_4}$ ) and average chain length ( $n$ ) were determined via NMR according to eqs. S3 and S4. The number-average molecular weight ( $M_n$ ) and dispersity ( $\mathcal{D}$ ) were obtained via GPC.

Sample	$X_{EG_4}$ [%]	$n$ [-]	$M_n$ [Da]	$\mathcal{D}$ [-]
20EG <sub>4</sub>	95.43	25.09	8456	1.12
40EG <sub>4</sub>	96.23	43.41	12013	1.07



**Figure S3:**  $^1\text{H-NMR}$  characterization and proton assignment for 20EG<sub>4</sub>-15CL3 ( $p=15$ ). The analysis was performed on a Bruker Ultrashield 400 MHz spectrometer using deuterated chloroform ( $\text{CDCl}_3$ ) as solvent.

The monomer conversion ( $X$ ) according to eq. S5:

$$X = \left(1 - \frac{2qO}{R}\right) * 100 \quad (\text{S5})$$

R is the area of the peak attributed to the two vinyl hydrogens in the unreacted monomer, O is the area of the peak associated to the hydrogens near the carbonyl group either in the monomer or in the polymer and  $q$  the average chain length of the macromonomer.

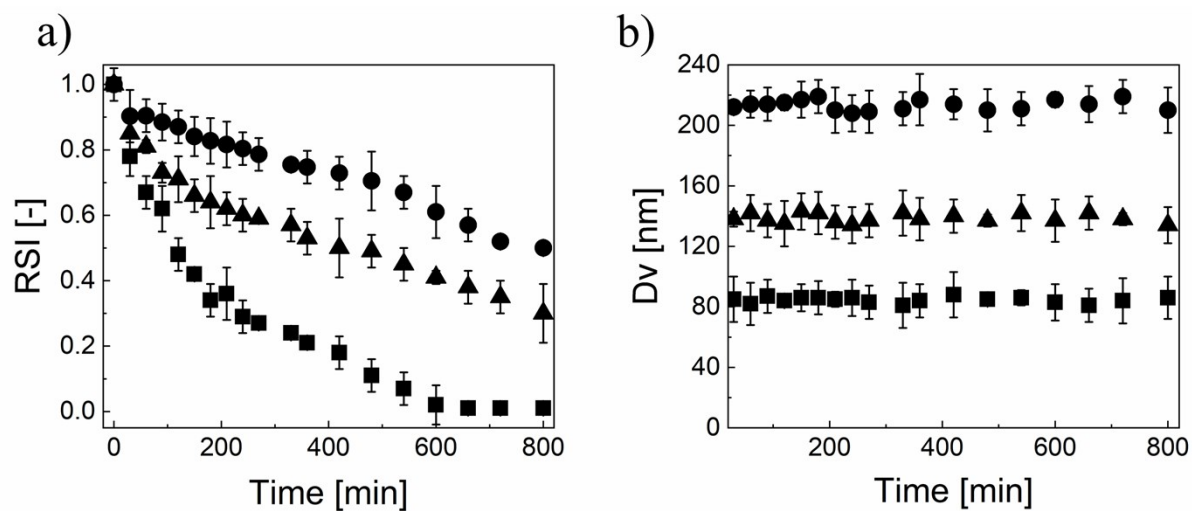
The degree of polymerization ( $p$ ) was calculated according to **eq. S6**:

$$p = \frac{\frac{R}{2q}}{\frac{S}{3n}} = \frac{3nR}{2qS} \quad (\text{S6})$$

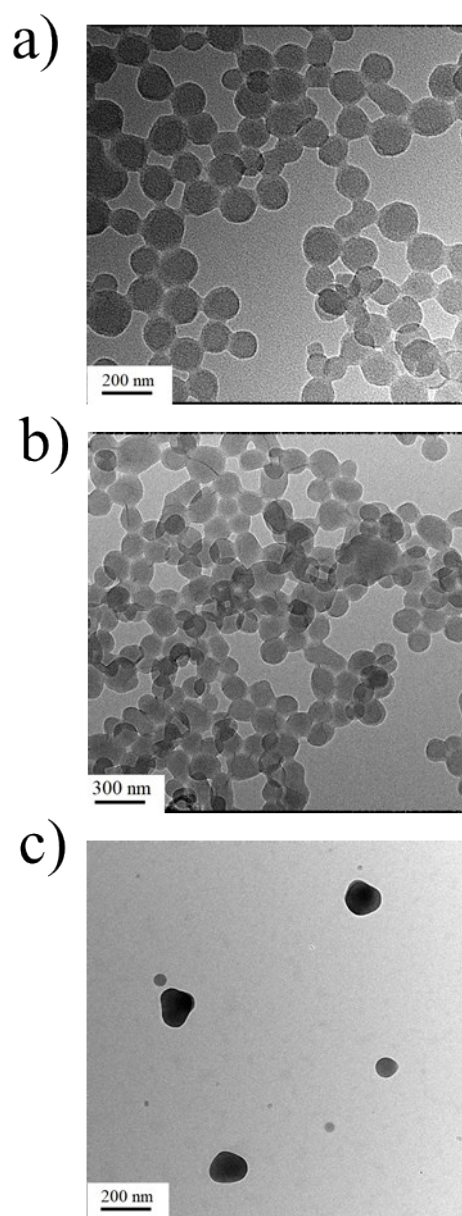
Where R and S are the areas of the corresponding peaks shown in **Figure S3**, while  $q$  and  $n$  are the average chain lengths of the macromonomer and the macro CTA adopted using the synthesis and reported in **Table S1** and **Table S2**, respectively.

**Table S3:** *Properties of the synthesized NPs in terms of monomer conversion ( $X$ ) and average degree of polymerization ( $p$ ) determined from  $^1\text{H}$  NMR and number-average molecular weight ( $M_n$ ) and dispersity ( $\mathcal{D}$ ) measured via GPC.*

Sample	$X$ [%]	$p$ [-]	$M_n$ [Da]	$\mathcal{D}$ [-]
20EG <sub>4</sub> -15CL3	98.90	15.42	23145	1.11
20EG <sub>4</sub> -25CL3	98.65	23.27	26432	1.11
20EG <sub>4</sub> -50CL3	98.90	52.59	42145	1.21
40EG <sub>4</sub> -15CL3	98.21	17.30	28121	1.11
40EG <sub>4</sub> -25CL3	99.62	24.80	29573	1.17
40EG <sub>4</sub> -50CL3	99.16	53.32	52890	1.21
20EG <sub>4</sub> -25CL5	98.72	25.17	32113	1.26
40EG <sub>4</sub> -15CL5	99.52	13.51	31065	1.19
40EG <sub>4</sub> -25CL5	99.26	27.43	43125	1.21
40EG <sub>4</sub> -50CL5	97.94	48.15	56859	1.12
20EG <sub>4</sub> -25CL3_Rh	99.33	26.78	28819	1.32



**Figure S4:** (a) Relative scattering intensity (RSI) as a function of time in the case of 40EG<sub>4</sub>-15CL5 (■) 40EG<sub>4</sub>-25CL5 (▲) and 40EG<sub>4</sub>-50CL5 (●), when the samples are diluted to 0.5% w/w in a solution of NaOH 0.1 M. (b) NP size as function of time in the case of 40EG<sub>4</sub>-15CL5 (■) 40EG<sub>4</sub>-25CL5 (▲) and 40EG<sub>4</sub>-50CL5 (●), when the samples are diluted to 0.5% w/w in a solution of NaOH 0.1 M.



**Figure S5:** (a) TEM image of 40EG<sub>4</sub>-50CL5, scale bar = 200 nm. (b) TEM image of 20EG<sub>4</sub>-50CL3, scale bar = 300 nm. (c) TEM image of 20EG<sub>4</sub>-25CL3\_Rh, scale bar = 200 nm