

## Supporting Information

### Self-cascade MoS<sub>2</sub> nanozymes for efficient intracellular antioxidation and hepatic fibrosis therapy

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## Other Methods

### Electron spin resonance measurements

All electron spin resonance (ESR) measurements were carried out at ambient temperature using a Bruker E500 ESR spectrometer.

*Measurement of CAT-like activity.* Electron spin resonance (ESR) spin label oximetry was employed to investigate the oxygen generation from  $\text{H}_2\text{O}_2$  catalyzed by  $\text{MoS}_2$  nanosheets at two pH levels. Sample solutions including 0.1 mM spin label  $^{15}\text{N}$ -PDT (Cambridge Isotope Labs, USA), 100  $\mu\text{g}/\text{mL}$   $\text{MoS}_2$  nanosheets, and buffer solutions (pH 4.5 or 7.4) were deoxygenated with nitrogen before the initiation of reactions by addition of 1.0 mM  $\text{H}_2\text{O}_2$ . ESR spectral measurements were obtained using the following settings: 0.04 G field modulation, 3 G scan range, and 1 mW microwave power.

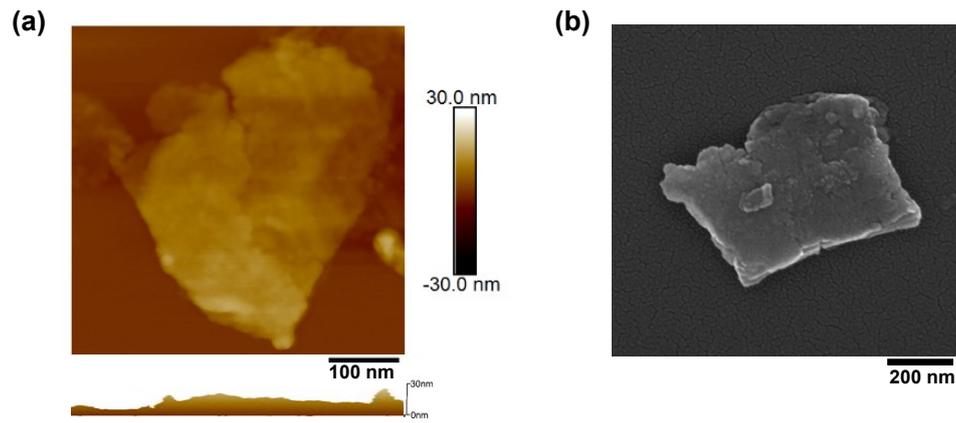
*Measurement of SOD-like activity.* To investigate the  $\text{O}_2^{\cdot-}$  scavenging ability of  $\text{MoS}_2$  nanosheets, two  $\text{O}_2^{\cdot-}$  sources, xanthine-xanthine oxidase (XAN-XOD) and  $\text{KO}_2/18$ -crown-6-ether systems, were used to generate  $\text{O}_2^{\cdot-}$ . BMPO (Dojindo Laboratories, Japan) was used to trap the  $\text{O}_2^{\cdot-}$  in the form of spin adduct  $\text{BMPO}/\cdot\text{OOH}$ . ESR spectral measurements were obtained using the following settings: 1 G field modulation, 100 G scan range, and 20 mW microwave power.

*Measurement of antioxidative effects of  $\text{MoS}_2$  nanosheets through interaction with cytochrome *c*/ $\text{H}_2\text{O}_2$ .* Oxidation of the spin trap DMPO (Dojindo Laboratories, Japan), to form 5,5-dimethyl-1-pyrrolidone-N-oxyl (DMPOX), was monitored by ESR to determine the effects of  $\text{MoS}_2$  nanosheets on cytochrome *c* (Cyt *c*)'s ability to catalyze

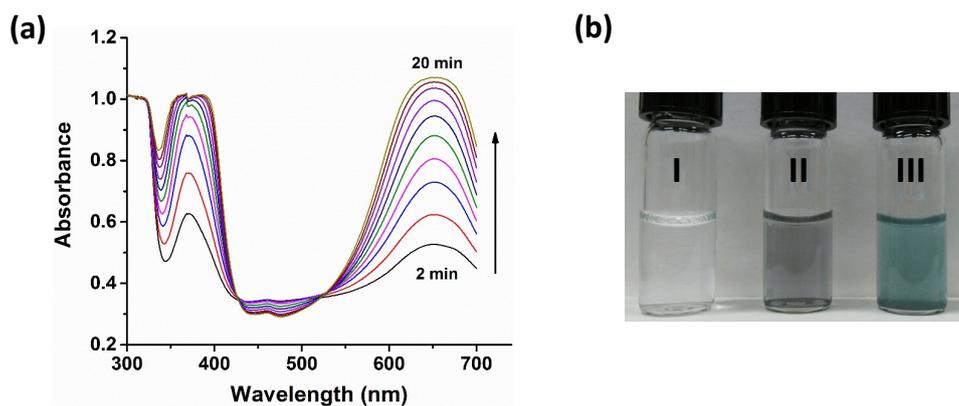
the oxidation of substrates by  $\text{H}_2\text{O}_2$ . Time dependence of the ESR signal for the Cyt *c*/ $\text{H}_2\text{O}_2$  system with  $\text{MoS}_2$  nanosheets or PBS (control) was measured. ESR spectral measurements were obtained using the following settings: 1 G field modulation, 100 G scan range, and 10 mW microwave power.

**Table S1** Michaelis-Menten parameters of MoS<sub>2</sub> nanosheets.

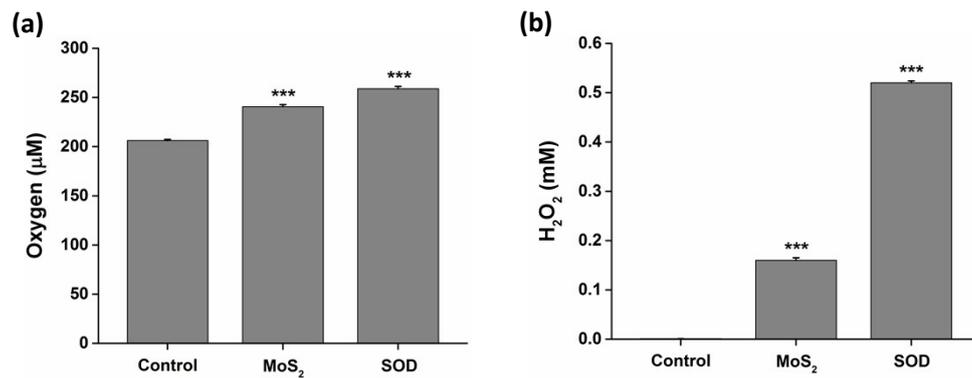
Substrate	$K_m$ (mM)	$V_{max}$ ( $10^{-6}$ M s <sup>-1</sup> )	$K_{cat}$ ( $10^{-3}$ s <sup>-1</sup> )	$K_{cat}/K_m$ ( $10^{-3}$ mM <sup>-1</sup> s <sup>-1</sup> )
H <sub>2</sub> O <sub>2</sub>	0.04	0.66	1.05	26.25
GSH	1.68	0.94	1.50	0.89



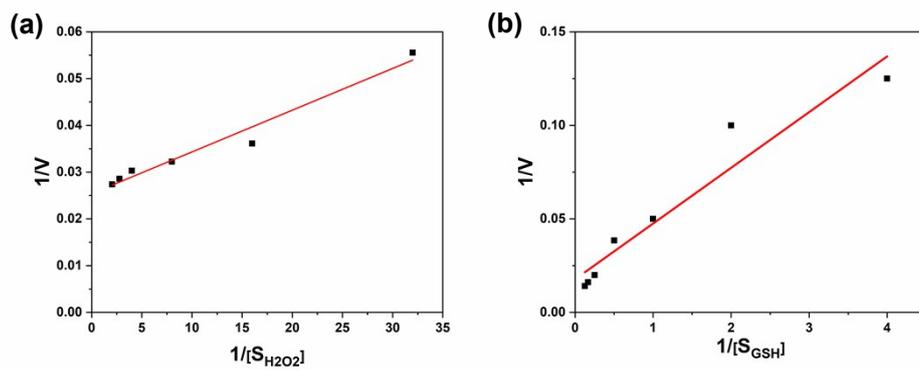
**Fig. S1** (a) AFM and (b) SEM images of MoS<sub>2</sub> nanosheets.



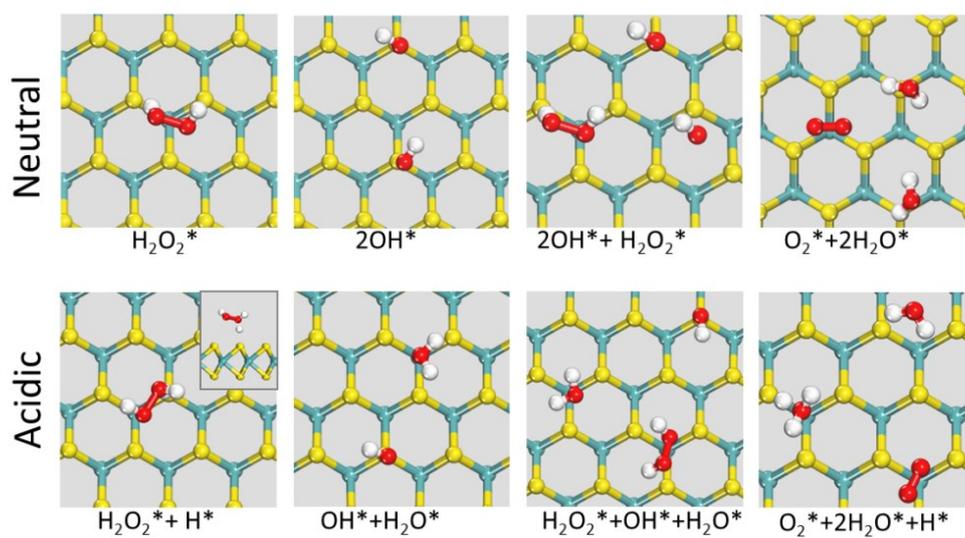
**Fig. S2** (a) The absorbance spectra of TMB oxidized by MoS<sub>2</sub> nanosheets. (b) Typical photographs of TMB reaction solutions oxidized by MoS<sub>2</sub> nanosheets in the presence of H<sub>2</sub>O<sub>2</sub> under pH 4.5. (I) 1 mM H<sub>2</sub>O<sub>2</sub> and 1 mM TMB; (II) 100 μg/mL MoS<sub>2</sub> nanosheets and 1 mM TMB; (III) 1 mM H<sub>2</sub>O<sub>2</sub>, 1 mM TMB and 100 μg/mL MoS<sub>2</sub> nanosheets.



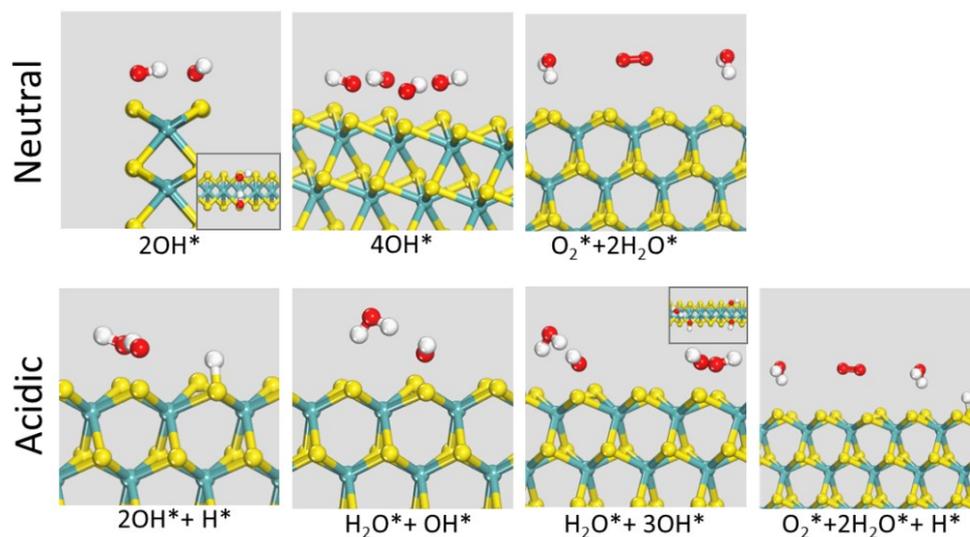
**Fig. S3** The quantification of (a) oxygen and (b) H<sub>2</sub>O<sub>2</sub> production from superoxide turnover by MoS<sub>2</sub> nanosheets vs 1 U/mL SOD in KO<sub>2</sub>/18-crown-6-ether system. \*\*\* $p < 0.001$  when compared with control group.



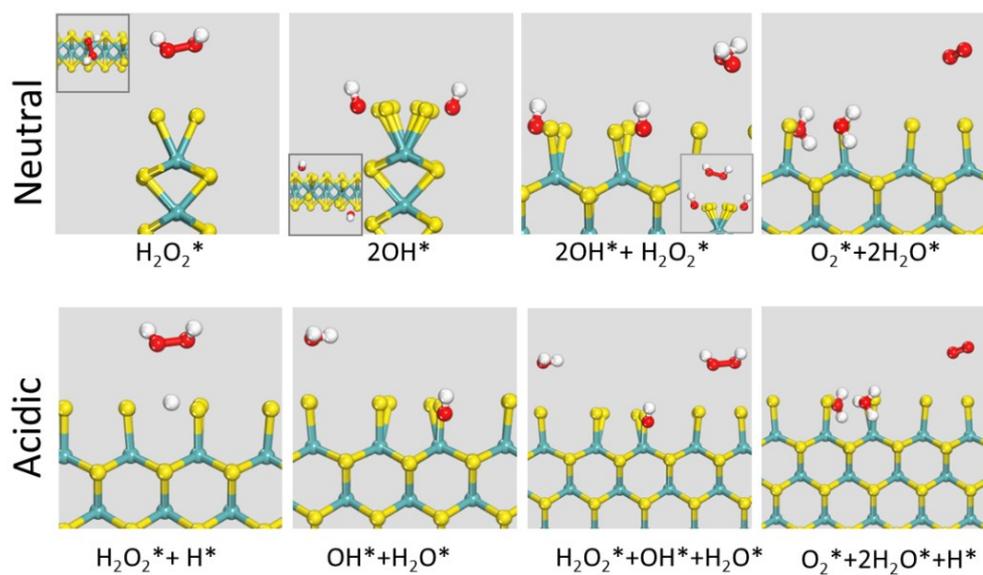
**Fig. S4** Corresponding double-reciprocal plots of MoS<sub>2</sub> nanosheets at a fixed concentration of one substrate *versus* varying the concentration of another for (a) H<sub>2</sub>O<sub>2</sub> and (b) GSH.



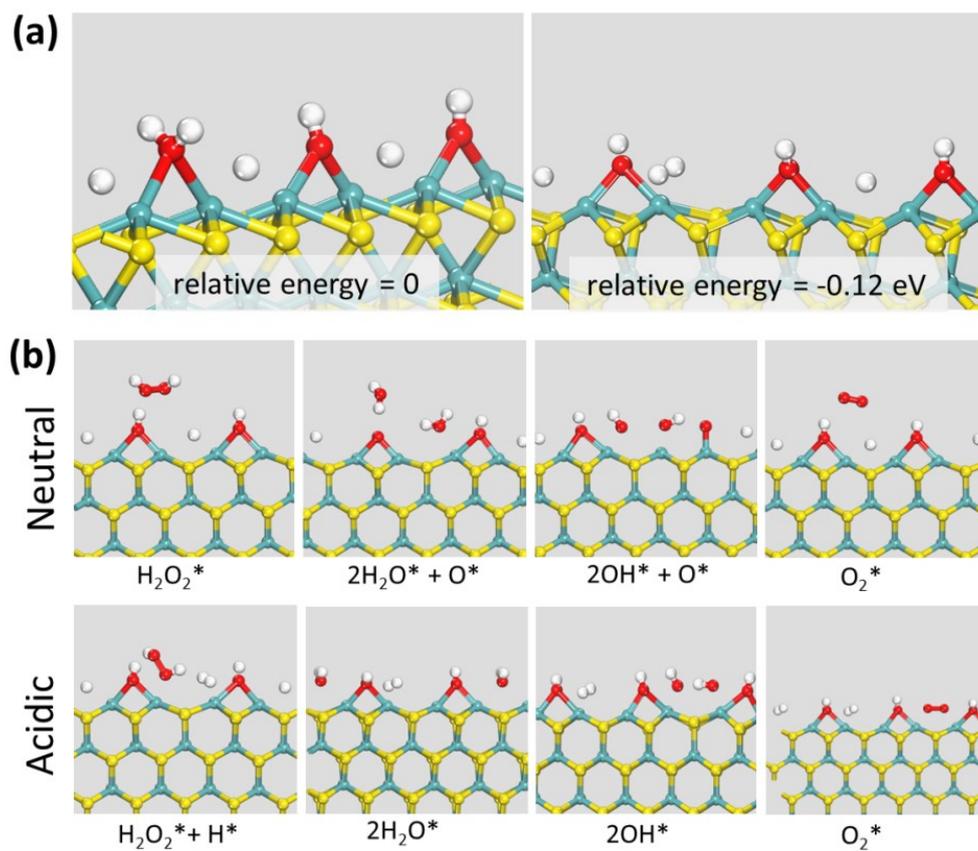
**Fig. S5** Lowest-energy adsorption structures of the intermediate species during the  $\text{H}_2\text{O}_2$  decomposition process on the basal plane of the  $\text{MoS}_2$  nanosheets under neutral (top panel) and acidic (bottom panel) conditions (white, H; red, O; yellow, S; cyan, Mo).



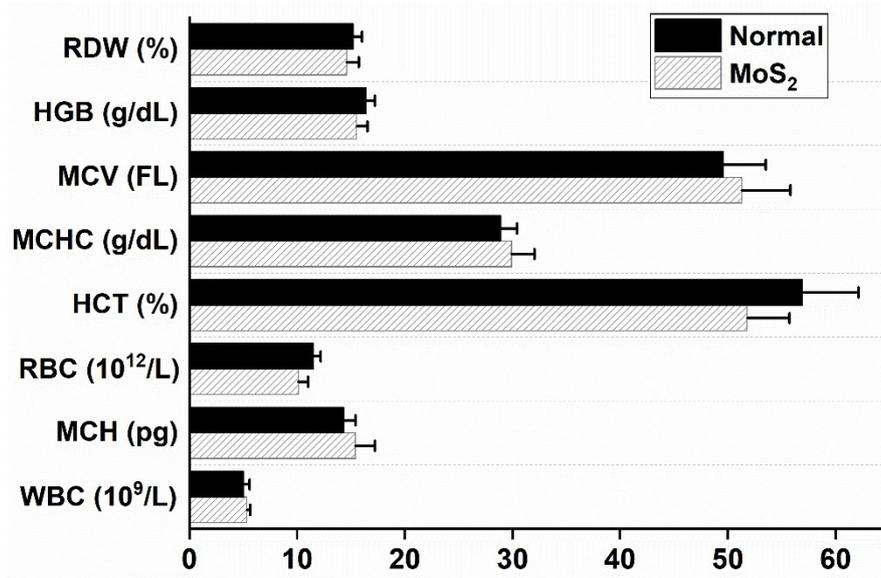
**Fig. S6** Lowest-energy adsorption structures of the intermediate species during the  $\text{H}_2\text{O}_2$  decomposition process on the Mo-S-edge of the  $\text{MoS}_2$  nanosheets under neutral (top panel) and acidic (bottom panel) conditions (white, H; red, O; yellow, S; cyan, Mo). Here, it is noteworthy that the adsorption of  $\text{H}_2\text{O}_2$  on Mo-S-edge is a chemical dissociative adsorption.



**Fig. S7** Lowest-energy adsorption structures of the intermediate species during the  $\text{H}_2\text{O}_2$  decomposition process on the S-edge of the  $\text{MoS}_2$  nanosheets under neutral (top panel) and acidic (bottom panel) conditions (white, H; red, O; yellow, S; cyan, Mo).



**Fig. S8** (a) Comparison between two stable adsorptions of H<sup>+</sup> on the Mo-edge; the adsorption at the H-site is more energetically stable. (b) Lowest-energy adsorption structures of the intermediate species during the H<sub>2</sub>O<sub>2</sub> decomposition process on the Mo-edge of the MoS<sub>2</sub> nanosheets under neutral (top panel) and acidic (bottom panel) conditions (white, H; red, O; yellow, S; cyan, Mo).



**Fig. S9** Blood hematology data of mice injected with saline (normal group) or MoS<sub>2</sub> nanosheets (0.5 mg/kg.bw) once a week for four weeks.