Supporting Information for "Adoptive CD8+T-cell grafted with liposomal

immunotherapy drugs counteract immune suppressive tumor microenvironment

to elicit enhanced therapy on melanoma"

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Figure S1 Stability of LP-CpG and LP-BMS-202

(a) Change of particle size and PDI of LP-CpG and LP-BMS-202 during storage at 4 °C, blue line: mean; (b) Change of entrapment efficiency (EE) of LP-CpG and LP-BMS-202 over storage time at 4 °C;

Samples	Loading efficiency	Encapsulate efficiency	Particle Size	PDI	Zeta potential
	(mg/mg)	(EE)	(nm)		(mV)
Anionic LP			85.5±2.3	0.29±0.05	-38.7±4.2
Cationic LP			124.7±3.4	0.231±0.04	+39.0±5.4
MPB-LP			95.5±2.3	0.19±0.03	-40.7±2.2
LP-CpG	10%	90±4%	205.0±4.2	0.33±0.23	-43.6±3.2
LP-BMS-202	10%	95±4%	122.3±3.5	0.173±0.12	-43.0±3.4

Table S1. Characteristics of the optimized liposome formulations.



Figure S2. UV-vis spectra of free BMS-202 and LP-BMS-202.



Figure S3. Photographs of hemolysis rate for A-LPs with various lipid concentrations. The degree of hemolysis was determined by measuring hemoglobin concentration, and the hemolysis rate less than 5% relative to positive control indicates good hemocompatibility of the liposomes.



Figure S4. mRNA expression of IL-6(a) and TNF-α(b) in DC2.4 after treatment of

various samples including PBS as negative control (NT), LPS as positive control (LPS), blank liposomes (A-LP/C-LP), free CpG (CpG), A-LP-CpG and C-LP-CpG; Data are expressed as mean \pm SD of each group (n = 3). * p < 0.05.



Figure S5. Surface marker of activated CD8⁺T cell after culture for 48h with CD3/ CD28 antibody.



Figure S6.T-cell conjugated with DiD labeled anionic liposomes cultured for various time points including 1h (a), 12h (b) and 24h (c).



Figure S7. Distribution of LP and CD8-T-LP in the main organs of mice at 24-hour after administration.



Figure S8. Representative histological HE staining photographs of the main organs after the treatment of the following groups including PBS, LP-CpG, LP-BMS-202, CD8-T, CD8-T+LP-CpG, CD8-T+LP-BMS-202, CD8-T-LP-CpG, CD8-T-LP-BMS-202. The scale bar represents 100 μm.



Figure S9. Flow cytometry gating strategy for the subgroups of cells including M-MDSC (CD45+ CD11b+ Ly6C^{high} Ly6G-), PMN-MDSC (CD45+ CD11b+ Ly6C^{low} Ly6G+), M1-like TAMs (CD45+CD11b+F4/80+CD86+CD206-) and M2-like TAMs (CD45+ CD11b+ F4/80+ CD86-CD206+), and Treg(CD45+CD4+CD25+FoxP3+).



Figure S10. MDSCs fraction gated from CD45+ cells in TIL after the treatment of the following groups including PBS, LP-CpG, CD8-T, CD8-T+LP-CpG, CD8-T-LP-CpG.



Figure S11. Flow cytometry quantification of the absolute number of lymphocyte subtypes in TIL including MDSCs, M2s, Marophages from tumor at 72h following the last treatment (n=3). *P < 0.05, **P < 0.01 by t-test.



Figure S12. T Antitumor effect of OT-1 CD8-T-LP-BMS-202 in the subcutaneous B16-OVA tumor model (n=5). (a) Body weight change curves of mice; (b) Tumor weight of each group.



Figure S13. Immune regulation of OT-1 CD8-T-LP-BMS-202 in the subcutaneous B16-OVA tumor model. (a) Statistic results of M-MDSC and PMN-MDSC frequency in TIL; (b) Flow cytometry quantification of the absolute number of lymphocyte subtypes in TIL including MDSCs, M2-TMAs, Macrophages from tumor at 72h following the last treatment (n = 3). *P<0.05,**P< 0.01 by t-test.



Figure S14. Immune memory establishment after treatment of all the groups including PBS, LP-BMS-202, CD8-T, CD8T+LP-BMS-202, CD8-T-LP-BMS-202 in the subcutaneous B16-OVA tumor model (n=5). (a) Treatment schedule for this assay; (b) Tumor growth curves of the 1st tumor; (c) Tumor weight of the 1st tumor; (d) Tumor weight of the 2nd tumor; (e) Determination of effector memory

T cells (CD44^{high} CD62L^{low}) and central memory T cells (CD44^{high} CD62L^{high}) of splenic CD8⁺T cells by flow cytometry; (f) Statistic analysis of effector memory T cells (n = 3).



Figure S15. Inhibition of tumor angiogenesis and proliferation by CD8-T-LP-CpG(a) and CD8-T-LP-BMS-202 (b). Immunohistochemical analysis of CD31 (pan-endothelial marker) and Ki67 (proliferation marker) stained with anti-CD31 antibody (200×) and anti-KI67 antibody (200×), respectively. Blue: DAPI, Green:CD31 or KI67. Statistical results given by ImageJ (c-f) are presented as the mean \pm SD, n = 3, ** p < 0.01, *** p<0.001.