

An integrated microfluidic electrochemical assay for cervical cancer detection at point-of-care testing

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Table 1. The sequence of nucleic acids used in this paper.

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Table S1. The sequence of nucleic acids used in this paper

| Nucleic acid strand | Sequence |
|--------------------------|--------------------------------------|
| hr-HPV 16 cssDNA (Probe) | GGGCTCTGTCCGGTTCTGCTTGTCCA/3AmMO/ |
| hr-HPV 16 cDNA (Target) | TGGACAAGCAGAACCGGACAGAGCCC |
| Cy3-rDNA | /5Cy3/GGTGGTGGGGGGGGTTGGTAGGGTGTCTTC |

Table 2. Recent studies of hr-HPV DNA detection

| Reference | Working electrode | Electrochemical technique | Medium | Target | LOD |
|------------------|---|--|---|--------|---------|
| Ref ¹ | Polypyrrole (PPy) films and gold nanoparticles (AuNPs) | Cyclic voltammetry | Cervical specimen | HPV16 | 13.4 pM |
| Ref ² | Gold electrode | Differential pulse voltammetry (reduction of MB) | Extracted DNA from cervical swabs | HPV16 | 18.1 nM |
| Ref ³ | Carbon | Anthraquinone square wave voltammetry | Human cancer cell line (PCR amplified) | HPV16 | 4 nM |
| Ref ⁴ | Nanocomposite of reduced graphene oxide(rGO) and multiwalled carbon nanotubes (MWCNTs) + Au nanoparticles | Differential pulse voltammetry | Extracted and PCR amplified DNA from cervix | HPV18 | 0.05 fM |
| IMEAC | Graphene oxide functionalized with cssDNA | Differential pulse voltammetry | Plasma | HPV16 | 0.48 μM |

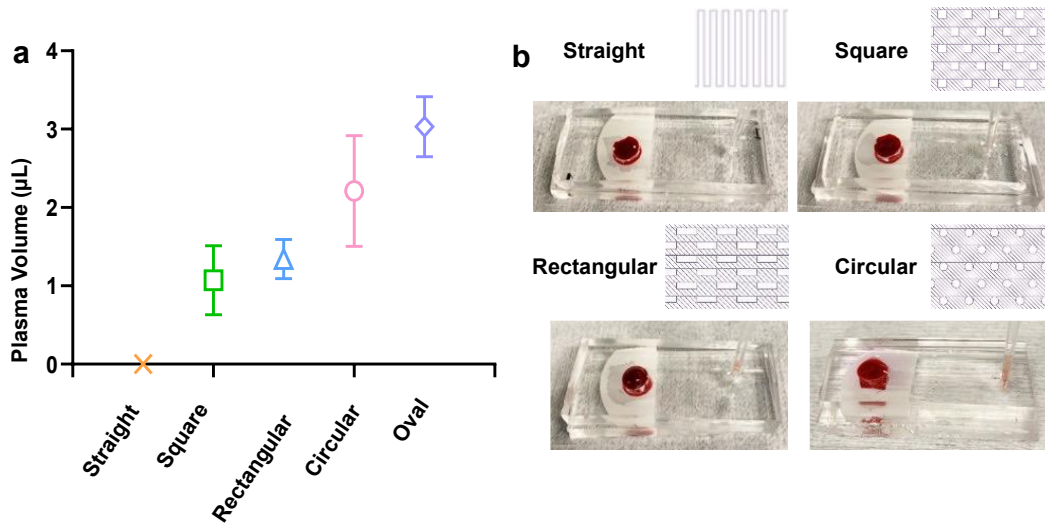


Fig. S1. Optimizing the shape of structures incorporated inside the micropump design based on the collected plasma volume.

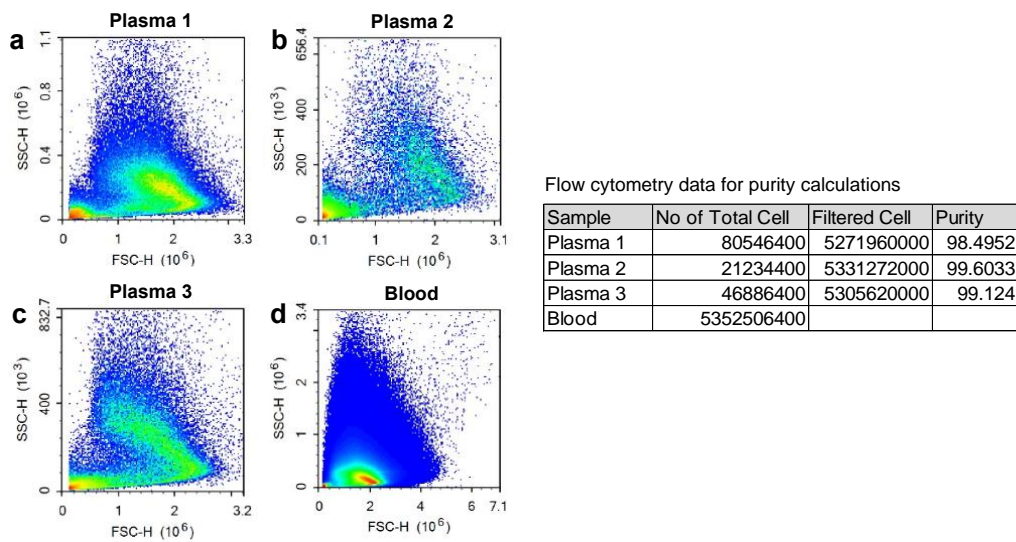


Fig. S2. Flow cytometry measurement for calculating the purity of plasma collected from the PPS device.

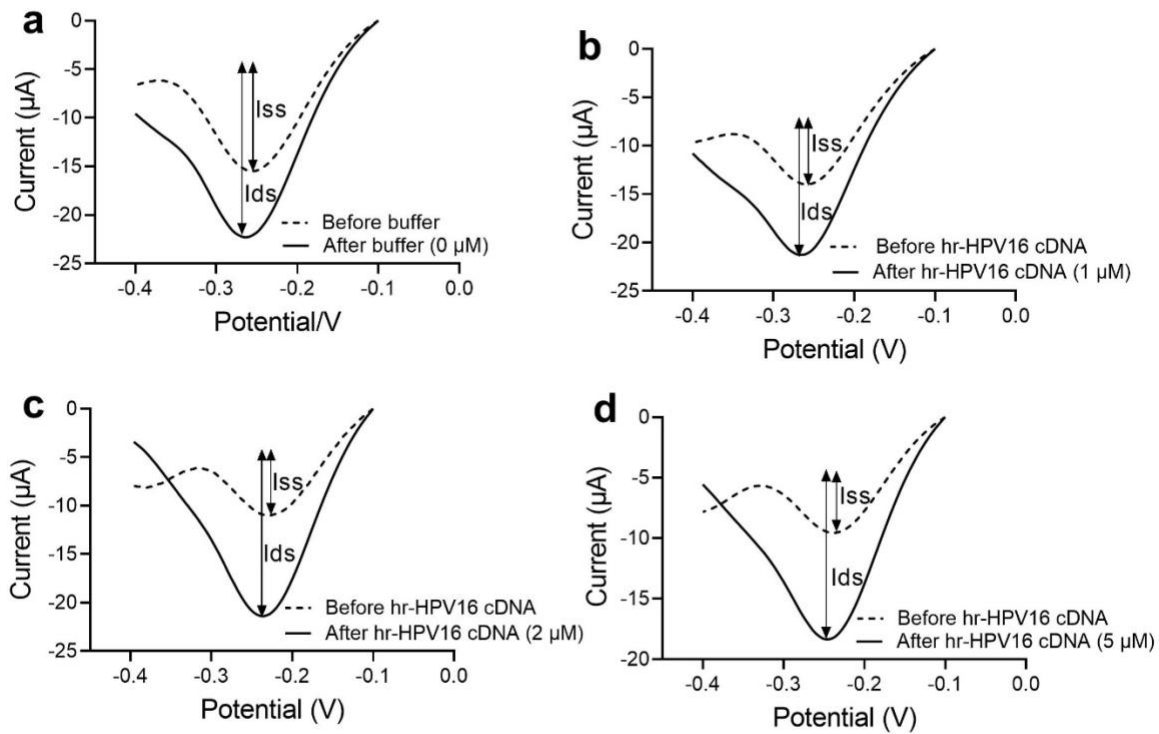


Fig. S3. The DPV scans before and after incubating GO-SPCE sensors with 0 μM (a), 1 μM (b), 2 μM (c), and 5 μM (d) of hr-HPV16 cDNA.

References

1. Avelino, K. Y. P. S., Oliveira, L. S., Lucena-Silva, N., Andrade, C. A. S. & Oliveira, M. D. L. Flexible sensor based on conducting polymer and gold nanoparticles for electrochemical screening of HPV families in cervical specimens. *Talanta* **226**, (2021).
2. Campos-Ferreira, D. S. *et al.* Electrochemical DNA biosensor for human papillomavirus 16 detection in real samples. *Anal. Chim. Acta* **804**, 258–263 (2013).
3. Jampasa, S. *et al.* Electrochemical detection of human papillomavirus DNA type 16 using a pyrrolidinyI peptide nucleic acid probe immobilized on screen-printed carbon electrodes. *Biosens. Bioelectron.* **54**, 428–434 (2014).
4. Mahmoodi, P. *et al.* Early-stage cervical cancer diagnosis based on an ultra-sensitive electrochemical DNA nanobiosensor for HPV-18 detection in real samples. *J. Nanobiotechnology* **18**, 1–12 (2020).