Supplementary Information for

Multivalent non-covalent interactions lead to strongest polymer adhesion

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1. AFM cantilever tip functionalization

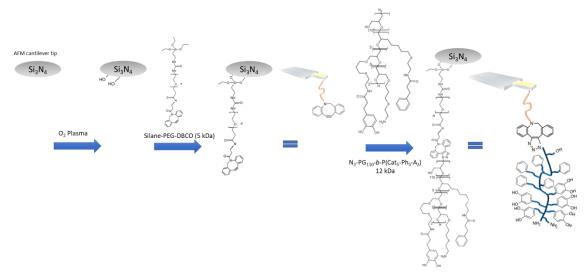


Fig. S1. Functionalization of an AFM cantilever tip. To obtain a covalently bound PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor, oxygen plasma treatment, silane-PEG-DBCO (5 kDa) and N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) (12 kDa) were used.

2. Contact angle measurements

To further characterize the presence of N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layers on a TiO₂ substrate, contact angle measurements were performed using a homebuilt goniometer. The control samples were TiO₂ incubated in the pure solvent (MOPS buffer pH 6.0, 0.1 M) instead of the polymer solution. Ten droplets of H₂O (1 µL) were put on each layer. For each droplet, the contact angle was determined using the distance tool of the software *Gwyddion 2.47*.¹ Finally, the average value and standard deviation were calculated (Fig. S2). The presence of N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layers increased the hydrophilicity and therefore the contact angle became smaller.

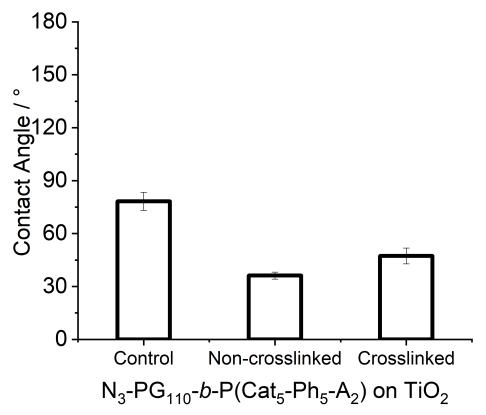
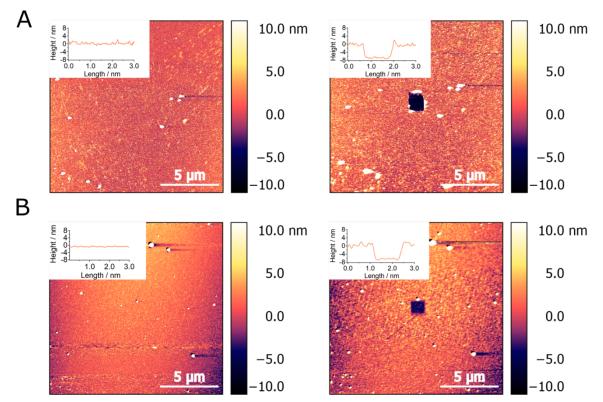


Fig. S2. Contact angle measurements of a non-crosslinked N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂, a crosslinked N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ and bare TiO₂ as a control. The control samples on TiO₂ were incubated in the pure solvent (MOPS buffer pH 6.0, 0.1 M) instead of the polymer solution. The contact angles amounted to (78 ± 5) ° for the control, (36 ± 2) ° for the non-crosslinked layer and (47 ± 5) ° for the crosslinked layer.



3. Thickness determination of a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ by AFM-based scratching

Fig. S3. Scratching of a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂. Imaging of the layer with a scan size of 15•15 μ m² (left). Imaging of the layer after the scratch with a scan size of 15•15 μ m² (right). A scan rate of 2 Hz was used for all images. **A.** The thickness of a non-crosslinked N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ in buffer (pH 6.0) amounted to (8.5 ± 0.7) nm. **B.** A crosslinked layer on TiO₂ in buffer (pH 6.0) showed a thickness of (6.8 ± 0.9) nm.

4. Force spectroscopy

4a. PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on SiO₂

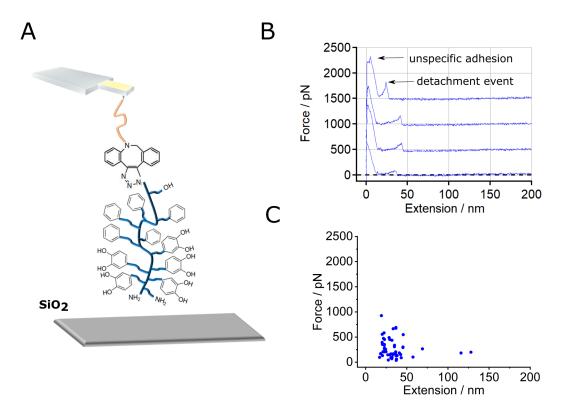


Fig. S4. PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on SiO₂. A. Scheme of the experiment. An AFM cantilever tip was functionalized with silane-PEG-DBCO (linker) and N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) and measured on SiO₂ in H₂O with a contact force of 500 pN and a contact time of 1 s. **B**. Representative force-extension curves. Out of 200 force-extension curves, 40 showed convex stretch events (20 %), 6 curves showed double convex stretches (3 %) and 5 curves showed multiple stretches (3 %). There were no concave stretches or plateaus. The curves were offset by 500 pN for graphical presentation. **C.** Scatter plot showing the detachment force distribution (including all peaks except unspecific adhesion) vs the detachment position. The average detachment force showed a value of (276 ± 192) pN and the average detachment position a value of (34 ± 20) nm.

4b. PEG-DBCO molecular force sensor on SiO₂

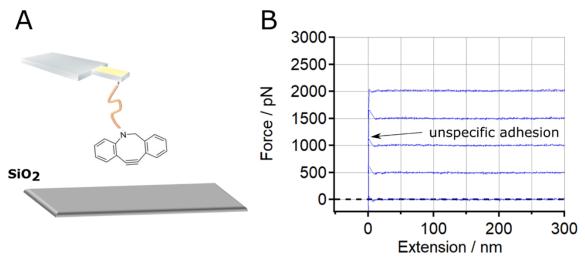


Fig. S5. PEG-DBCO molecular force sensor on SiO₂. **A.** Scheme of the experiment. An AFM cantilever was functionalized with silane-PEG-DBCO and measured on SiO₂ in H₂O with a contact force of 500 pN and a contact time of 1 s. **B.** Force-extension curves of silane-PEG-DBCO on SiO₂. Out of 200 curves, there were no curves with concave or convex stretches or plateaus. The curves were offset by 500 pN for graphical presentation.

4c. Contact force vs contact time for a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂

In order to understand the timescales involved in the formation of bonds between a $PG_{110} - b$ -P(Cat₅-Ph₅-A₂) molecular force sensor and a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂, we varied the contact time of the molecular force sensor on the layer and the contact force with which the AFM cantilever tip was pressed against this layer (Fig. S6).

As a control, we started with a contact force of 50 pN (below the breakthrough force) with contact times of 0 s, 1 s and 10 s. The real contact time (comprising dwell and contact during approach and retract of the AFM cantilever tip) for 0 s is 0.02 s for 50 pN, 0.05 s for 500 pN and 0.09 s for 3000 pN. For 1 s, the real contact times are 1.02 s for 50 pN, 1.04 s for 500 pN and 1.09 s for 3000 pN. Finally, a contact time of 10 s means a real contact time of 10.02 s for 50 pN, 10.04 s for 500 pN and 10.07 s for 3000 pN.

The experiments with a contact force of 50 pN only showed force-extension curves with low detachment forces in the range of 100 pN (Fig. S7). Only an increase of the contact force to 500 pN (above the breakthrough force) led to a noticeable increase in the detachment forces, reaching values greater than 2000 pN (Fig. S7). This is in accordance with the repelling properties assigned to N₃-PG₁₁₀-b-P(Cat₅-Ph₅-A₂) layers.²

Increasing the contact force further to a value of 3000 pN revealed diverse types of forceextension curves (see also Fig. 1d). We interpret this as a higher probability for the interactions between the functional groups of PG_{110} -*b*-P(Cat₅-Ph₅-A₂) on the AFM cantilever tip and the N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer. In that respect, we observed a significant amount of desorption plateaus as well as multiple plateaus. The highest number of convex stretches (35 %) was obtained for a contact time of 10 s and a contact force of 3000 pN. We believe that the increase of contact time results in a strengthening of the interactions between catechol groups of the PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor and the N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer. Indentation curves as the one given in Figure 1c, reveal that contact forces of at least 200 to 400 pN were necessary for a characteristic break-through of the N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer.

Based on these results, we chose a contact time of 1 s and a contact force of 500 pN as threshold values for further measurements, which is a good compromise between monitoring a great number of interactions and preventing a too high number of overlapping interactions (resulting in overlapping detachment events) rendering the force-extension curves uninterpretable.

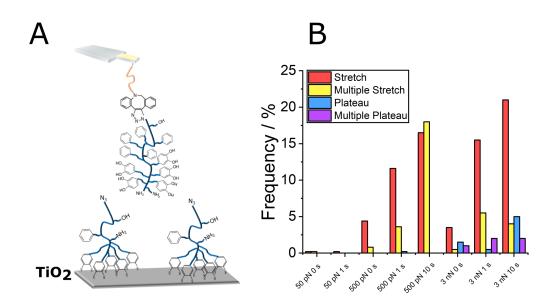


Fig. S6. Effect of contact force and contact time for a PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a non-crosslinked N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂. A. Scheme of a PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a non-crosslinked N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂. B. Frequency of the different curve types, showing stretches, multiple stretches, plateaus and multiple plateaus is given for different contact forces (50 pN, 500 pN, 3000 pN) and different contact times (0 s, 1 s, 10 s). The experiments were performed in H₂O. The frequency has been scaled to the total number of curves taken (200 for each condition).

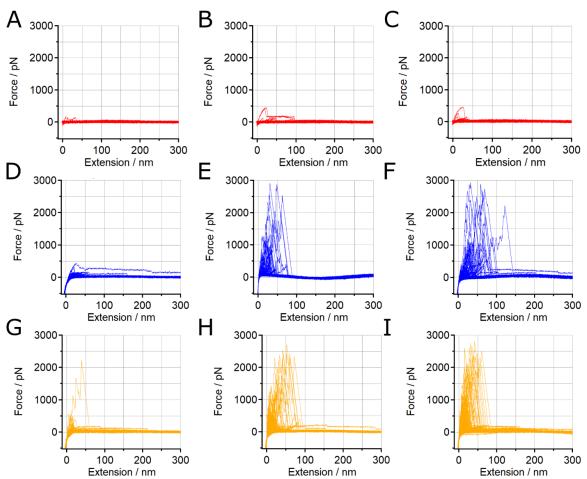
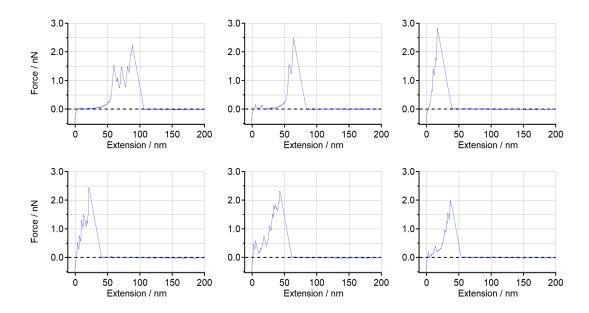


Fig. S7. PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ with different contact forces (50 pN, 500 pN, 3000 pN) and different contact times (0 s, 1 s, 10 s) measured in H₂O. A. 50 pN, 0 s B. 50 pN, 1 s C. 50 pN, 10 s D. 500 pN, 0 s, E. 500 pN, 1 s, F. 500 pN, 10 s, G. 3000 pN, 0 s, H. 3000 pN, 1 s, I. 3000 pN, 10 s.



4d. Force-extension curves with forces higher than 1500 pN

Fig. S8. Exemplary force-extension curves for a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ with detachment forces higher than 1500 pN. These force-extension curves were taken at the condition pH 8.6 crosslinked.

4e. Frequency of different force-extension motifs for a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂

The different types of force-extension curves for a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ are listed in Table S1. The numbers indicate the absolute number of a certain type of detachment event, while the numbers in the brackets indicate the respective frequency of occurrence in % with respect to the total number of force-extension curves taken.

Condition		pH 6.0	pH 8.6	pH 8.6
				crosslinked
Number of events	Concave stretch	5 (1 %)	0 (0 %)	0 (0%)
	Convex stretch	72 (14 %)	110 (21 %)	89 (17 %)
	Double convex stretch	11 (2 %)	39 (8 %)	23 (4 %)
	Multiple stretch	39 (8 %)	35 (7 %)	15 (3 %)
	Single plateau	3 (1 %)	0 (0 %)	1 (1 %)
	Multiple plateaus	1 (1 %)	0 (0 %)	0 (0 %)

Table S1. Number (frequency of occurrence) of the different force-extension curves for a PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂.

4f. Number of detachment events for a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂

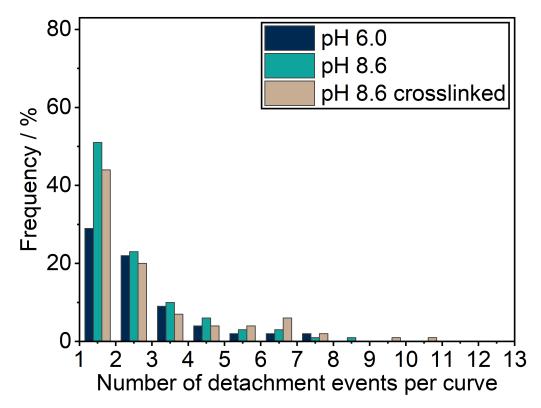


Fig. S9. Number of detachment events per curve. For a PG_{110} -*b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N_3 -PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ the following number of force-extension curves was obtained: 70 at pH 6.0, 98 at pH 8.6 and 89 at pH 8.6 crosslinked.

4g. Average energy per peak

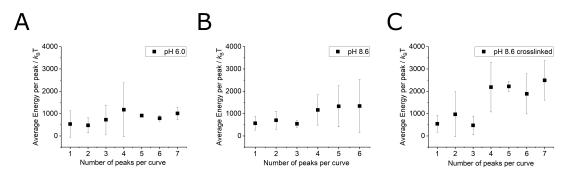


Fig. S10. Average energy per peak as a function of the number of peaks per curve. A PG₁₁₀ - *b*-P(Cat₅-Ph₅-A₂) molecular force sensor on a N₃-PG₁₁₀-*b*-P(Cat₅-Ph₅-A₂) layer on TiO₂ is shown for pH 6.0 (A), pH 8.6 (B) and pH 8.6 (C) crosslinked. The area between the force-extension curve and the baseline were determined by a trapezoidal integration.

SI References

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