Binaphthyl-Prolinol chiral ligands: Design and Their Application in Enantioselective Arylation of Aromatic Aldehydes

Chao Yao,^a Yaoqi Chen,^a Ruize Sun,^a Yue Huang,^a Lin Li^a and Yue-Ming Li^{a,b*}

^a State Key Laboratory of Medicinal Chemical Biology, College of Pharmacy and Tianjin Key Laboratory of Molecular Drug Research, Nankai University, Tianjin, People's Republic of China Email: ymli@nankai.edu.cn

^b CAS Key Laboratory of Synthetic Chemistry of Natural Substances, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 345 Lingling Road, Shanghai 200032, People's Republic of China

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1. Optimization of Reaction Conditions for Enantioselective Arylation-Lactonization of Methyl 2-Formylbenzoate

The general procedure for aryl transfer was adopted. Further study showed that the reaction temperature played a crucial role on the enantioselectivity and reactivity (Table S1, entries 1-2). The yield could be improved when the reaction time was prolonged from 24 h to 36 h. (Table S1, entries 2-3). Further increasing the reaction time didn't improve the yield of the reaction (Table S1, entries 3-4). The amount of chiral ligand was also studied, and 10 mol% of the chiral ligand was still the best choice (Table S1, entries 3, 5 and 6). Therefore, the final reaction condition was fixed as follows: reactions were performed on 0.25 mmol scale with PhB(OH)₂ (2.4 equiv), Et₂Zn (7.2 equiv) in toluene (stirring at 60 °C for 12 h), then addition of **3f** (10 mol%), DiMPEG 2000 (10 mol%) and aldehyde at 0 °C. The reaction mixture was stirred at 0 °C for 36 h under in an argon atmosphere.

Table S1. Optimization of enantioselective arylation-lactonization of methyl 2-formylbenzoate^a

3f. DiMPEG 2000

OH

ZnEt₂, 60 °C

		l ───► - toluene, 12 h	CH	о СН3		
entry	Ligand (mol%)	$T(^{\circ}C)$	<i>t</i> (h)	yield (%) ^b	ee (%) ^c	config. ^d
1	10	25	24	83	89	R
2	10	0	24	67	94	R
3	10	0	36	76	95	R
4	10	0	48	75	95	R
5	5	0	36	73	93	R
6	20	0	36	71	90	R

^a Reactions were performed on a 0.25 mmol scale with PhB(OH)₂ (2.4 equiv), Et₂Zn (7.2 equiv)

in toluene (stirring at 60 °C for 12 h), then addition of **3f** (10 mol%), DiMPEG 2000 (10 mol%) and aldehyde at 0 °C, with stirring for 36 h under an atmosphere of argon. ^b Isolated yields. ^c The ee values were determined by HPLC (chiralcel OD-H column). ^d Absolute configuration was assigned by comparison to reported value and sign of specific rotation of the product.¹

2. Preparation and Characterization of Chiral Ligands

The method for preparation of intermediate 4, ligands 1 and 2 can be found in our previously report.²

3. Characterization and Enantiomeric Excesses of Products



(*R*)-phenyl(p-tolyl)methanol $(15a)^3$

90% yield, 98% ee, Colorless oil; HPLC (Chiralcel OD-H, 2% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 27.3$ min for enantiomer (S), $t_R =$

30.7 min for enantiomer (R). [*a*]_D²⁰ = +17.8 (*c* = 1.00, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.24 (m, 4H), 7.24 – 7.15 (m, 3H), 7.09 (d, *J* = 7.9 Hz, 2H), 5.69 (s, 1H), 2.62 (s, 1H), 2.29 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 144.0, 141.0, 137.3, 129.6, 129.2, 128.5, 127.5, 126.7, 126.6, 120.4, 115.5, 76.1, 21.2.

(S)-phenyl (p-tolyl) methanol

93% yield, 88% ee, Colorless oil; HPLC (Chiralcel OD-H, 2% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 28.0$ min for enantiomer (S), $t_R = 32.3$ min for enantiomer (R). $[\alpha]_D^{20} = -8.9$ (c = 0.60, CHCl₃)



HPLC Chromatograms - Enantiomer ${\mathcal S}$

Me

OH





89% yield, 95% ee, Colorless oil; HPLC (Chiralcel OB-H, 25% IPA in

hexane, 1.0 mL/min, UV 220 nm): $t_R = 7.4$ min for enantiomer (R), $t_R = 12.2$ min for enantiomer (S). $[a]_D^{20} = +4.8$ (c = 0.40, CHCl₃).

¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.27 (m, 4H), 7.22 – 7.09 (m, 4H), 7.06 – 6.99 (m, 1H), 5.67 (s, 1H), 2.67 (s, 1H), 2.28 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 143.9, 143.8, 138.2, 128.5, 127.5, 127.3, 126.6, 123.8, 120.4, 115.5, 76.3, 21.5.

(S)-phenyl(m-tolyl)methanol⁴

78% yield, 81% ee, Colorless oil; HPLC (Chiralcel OB-H, 25% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 7.8$ min for enantiomer (R), $t_R = 12.0$ min for enantiomer (S). $[a]_D^{20} = -2.6$ (c = 0.40, CHCl₃).



HPLC Chromatograms - Enantiomer R



OH Me (*R*)-phenyl(o-tolyl)methanol (15c)⁵
88% yield, 98% ee, Colorless oil; HPLC (Chiralcel OJ-H, 10% IPA in hexane, 0.4 mL/min, UV 210 nm): t_R = 25.4 min for enantiomer (R), t_R = 27.9 min for enantiomer (S). [*a*]_D²⁰ = -7.8 (*c* = 1.25, CHCl₃).
¹H NMR (400 MHz, CDCl₃) ¹H NMR (400 MHz, CDCl₃) δ 7.51 - 7.49 (m, 1H), 7.34 - 7.28 (m, 4H), 7.27 - 7.19 (m, 3H), 7.14 - 7.12 (m, 1H), 5.99 (s, 1H), 2.24 (s, 3H), 1.94 (s, 1H).¹³C NMR (100 MHz, CDCl₃) δ 143.1, 141.6, 135.5, 130.7, 128.6, 127.7, 127.7, 127.2, 126.5, 126.3, 73.6, 19.5.
(*S*)-phenyl(o-tolyl)methanol⁵

81% yield, 92% ee, Colorless oil; HPLC (Chiralcel OJ-H, 10% IPA in hexane, 0.4 mL/min, UV 210 nm): $t_R = 29.9$ min for enantiomer (R), $t_R = 31.7$ min for enantiomer (S). $[a]_D^{20} = +11.8$ (c = 0.70, CHCl₃)









OH Me Me

(*R*)-(3,4-dimethylphenyl)(phenyl)methanol (15d)⁶
95% yield, 94% ee, Colorless oil; HPLC (Chiralcel OD-H, 2% IPA in

Me hexane, 1.0 mL/min, UV 220 nm): $t_R = 28.9$ min for enantiomer (S), $t_R =$

36.4 min for enantiomer (R). $[a]_{D^{20}} = -7.7$ (*c* = 0.30, CHCl₃).

¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.35 (m, 2H), 7.23 (d, *J* = 2.4 Hz, 2H), 7.16-7.13 (m, 1H), 7.08 (d, *J* = 1.3 Hz, 3H), 5.77 (s, 1H), 2.23 (s, 6H), 2.21 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 144.2, 141.6, 136.9, 136.1, 129.9, 128.6, 128.0, 127.5, 127.3, 126.6, 124.2, 76.3, 19.9, 19.5.



OH

(R)-(4-methoxyphenyl)(phenyl)methanol (15e)⁴

91% yield, 98% ee, Colorless oil; HPLC (Chiralcel AD-H, 3% IPA in OMe hexane, 1.0 mL/min, UV 215 nm): $t_R = 26.6$ min for enantiomer (R), $t_R =$ 29.2 min for enantiomer (S). $[a]_{D}^{20} = +19.8 (c = 0.60, CHCl_{3}).$

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.28 (m, 4H), 7.27 – 7.20 (m, 3H), 6.87 – 6.75 (m, 2H), 5.75 (s, 1H), 3.75 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 159.2, 144.2, 136.4, 128.5, 128.0, 127.5, 126.5, 114.0, 75.9, 55.4.

HPLC Chromatograms - Enantiomer R



ΟН OMe (*R*)-(3-methoxyphenyl)(phenyl)methanol $(15f)^7$

96% yield, 99% ee, Colorless oil; HPLC (Chiralcel IB, 5% IPA in hexane, 0.8 mL/min, UV 220 nm): t_R = 14.8 min for enantiomer (R), t_R = 19.0 min for enantiomer (S). $[\alpha]_{D}^{20} = -24.6$ (*c* = 1.0, CHCl₃).

¹H NMR (400 MHz, CDCl₃) δ 7.29 – 7.21 (m, 4H), 7.19 – 7.13 (m, 3H), 6.83 – 6.71 (m, 2H), 5.68 (s, 1H), 3.68 (s, 3H), 2.29 (d, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 159.1, 144.2, 136.3, 128.5, 128.5, 128.0, 127.5, 126.5, 114.0, 75.9, 55.4.

(S)-(3-methoxyphenyl)(phenyl)methanol

90% yield, 99% ee, Colorless oil; HPLC (Chiralcel IB, 5% IPA in hexane, 0.8 mL/min, UV 220 nm): $t_{R} = 14.7$ min for enantiomer (R), $t_{R} = 18.8$ min for enantiomer (S). $[a]_{D}^{20} = +22.9$ (c = 1.00, $CHCl_3$).



HPLC Chromatograms - Enantiomer ${\mathcal S}$



Peak	Ret. Time.	Area	Area%
1	14.700	237547	0.562
2	18.853	42034180	99.438

QH OMe (R)-(2-methoxyphenyl)(phenyl)methanol (15g)⁸

92% yield, 94% ee, Colorless oil; HPLC (Chiralcel OD-H, 2% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 20.8$ min for enantiomer (S), $t_R = 22.8$ min for enantiomer (R). $[a]_D^{20} = +22.9$ (c = 0.60, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.36 (m, 2H), 7.33 – 7.28 (m, 2H), 7.26 – 7.19 (m, 3H), 6.99 – 6.82 (m, 2H), 6.05 (d, J = 4.8 Hz, 1H), 3.79 (s, 3H), 3.01 (d, J = 5.3 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 156.9, 143.5, 132.2, 128.8, 128.3, 128.0, 127.3, 126.7, 121.0, 110.9, 72.4, 55.5. HPLC Chromatograms - Enantiomer *R*



(*R*)-(4-chlorophenyl)(phenyl)methanol $(15h)^4$

ОН

CI

93% yield, 94% ee, Colorless oil; HPLC (Chiralcel AD-H, 5% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 11.7$ min for enantiomer (R), $t_R = 13.2$ min

for enantiomer (S). [*a*]_D²⁰ = -12.7 (*c* = 0.80, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 6.90 (m, 9H), 5.59 (d, *J* = 1.9 Hz, 1H), 2.62 (d, *J* = 2.6 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 143.5, 142.3, 133.3, 128.7, 128.7, 128.0, 127.9, 127.9, 126.6, 75.6.





(R)-(4-bromophenyl)(phenyl)methanol (15i)⁷

92% yield, 94% ee, Colorless oil; HPLC (Chiralcel AD-H, 5% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 12.5$ min for enantiomer (R), $t_R = 14.2$ min

for enantiomer (S). [*a*]_D²⁰ = -10.8 (*c* = 0.50, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.27 (m, 2H), 7.26 – 7.06 (m, 7H), 5.61 (s, 1H), 2.50 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 143.4, 142.8, 131.6, 131.6, 128.7, 128.3, 127.9, 126.6, 121.5, 75.7.

(S)-(4-bromophenyl)(phenyl)methanol

93% yield, 88% ee, Colorless oil; HPLC (Chiralcel AD-H, 5% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 14.1$ min for enantiomer (R), $t_R = 15.7$ min for enantiomer (S). $[a]_D^{20} = +15.6$ (c = 0.50, CHCl₃)



HPLC Chromatograms - Enantiomer ${\mathcal S}$



OH Br

(R)-(2-bromophenyl)(phenyl)methanol (15j)⁹

95% yield, 96% ee, Colorless oil; HPLC (Chiralcel OD-H, 10% IPA in hexane, 1.0 mL/min, UV 210 nm): t_R = 8.6 min for enantiomer (R), t_R = 11.3 min for

enantiomer (S). [*a*]_D²⁰ = +37.8 (*c* = 1.50, CHCl₃).¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.35 (m, 2H), 7.26 – 7.22 (m, 2H), 7.21 – 7.09 (m, 4H), 7.03 – 6.90 (m, 1H), 6.00 (s, 1H), 2.68 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 142.6, 142.2, 132.9, 129.1, 128.5, 128.5, 127.8, 127.8, 127.1, 122.8, 74.8. HPLC Chromatograms - Enantiomer *R*



OH

(*R*)-phenyl(4-(trifluoromethyl)phenyl)methanol $(15k)^4$

74% yield, 92% ee, Colorless oil; HPLC (Chiralcel AD-H, 10% IPA in CF_3 hexane, 1.0 mL/min, UV 220 nm): $t_R = 6.2$ min for enantiomer (R), $t_R =$

7.3 min for enantiomer (S). $[a]_D^{20} = -27.8$ (c = 1.30, CHCl₃).

¹H NMR (400 MHz, CDCl₃) δ 7.45 (d, J = 8.2 Hz, 2H), 7.34 (d, J = 8.1 Hz, 2H), 7.27 – 7.12 (m, 5H), 5.67 (s, 1H), 2.70 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 147.6, 143.2, 129.8 (q, ²*J*_{C-F} = 32.4 Hz), 128.9, 128.2, 126.8, 126.8, 125.5 (q, ³*J*_{C-F} = 3.9 Hz), 124.29 (q, ¹*J*_{C-F} = 271.9 Hz).75.8.





(R)-naphthalen-1-yl(phenyl)methanol $(15l)^4$

90% yield, 86% ee, Colorless oil; HPLC (Chiralcel OD-H, 10% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 15.3$ min for enantiomer (S), $t_R = 33.5$ min for enantiomer (R). $[\alpha]_{D}^{20} = +67.8 (c = 1.00, CHCl_3).$

¹H NMR (400 MHz, CDCl₃) δ 7.95 – 7.86 (m, 1H), 7.79 – 7.66 (m, 2H), 7.55 – 7.47 (m, 1H), 7.41 - 7.25 (m, 5H), 7.25 - 7.11 (m, 3H), 6.38 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 143.2, 138.9, 134.1, 130.8, 128.9, 128.6, 128.6, 127.8, 127.2, 126.3, 125.7, 125.4, 124.7, 124.1, 73.7.





(*R*)-naphthalen-2-yl(phenyl)methanol $(15m)^4$

86% yield, 97% ee, Colorless oil; HPLC (Chiralcel OD-H, 10% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 15.5$ min for enantiomer (R), $t_R = r(S) [r^2]^{20} = +22.6$ (r = 1.00 CHCl.)

19.0 min for enantiomer (S). $[a]_D^{20} = +22.6$ (*c* = 1.00, CHCl₃).

¹H NMR (400 MHz, CDCl₃) δ 7.79 – 7.55 (m, 4H), 7.38 – 7.29 (m, 2H), 7.29 – 7.22 (m, 3H), 7.22 – 7.09 (m, 3H), 5.92 – 5.68 (s, 1H), 2.55 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 143.7, 141.2, 133.3, 133.0, 129.7, 128.6, 128.4, 128.2, 127.8, 127.7, 126.8, 126.3, 126.1, 125.1, 124.9, 120.5, 115.4, 76.4.





(*R*)-phenyl(thiophen-2-yl)methanol $(15n)^7$

92% yield, 97% ee, Colorless oil; HPLC (Chiralcel OD-H, 5% IPA in hexane, 0.8 mL/min, UV 220 nm): t_R = 21.3 min for enantiomer (S), t_R = 22.5 min for

enantiomer (R). [*a*]_D²⁰ = -15.7 (*c* = 1.25, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.33 (m, 2H), 7.31 – 7.21 (m, 3H), 7.19 – 7.15 (m, 1H), 6.90 – 6.82 (m, 1H), 6.82 – 6.73 (m, 1H), 5.95 (d, *J* = 2.8 Hz, 1H), 2.43 (d, *J* = 3.6 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 148.3, 143.3, 128.7, 128.1, 126.8, 126.4, 125.5, 125.0, 72.5.





(*R*)-furan-2-yl(phenyl)methanol $(15o)^7$

96% yield, 95% ee, Yellow oil; HPLC (Chiralcel OD-H, 3% IPA in hexane, 0.25 mL/min, UV 220 nm): $t_R = 78.0$ min for enantiomer (S), $t_R = 90.7$ min for

enantiomer (R). [*a*]_D²⁰ = +19.1(*c* = 1.30, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.35 (m, 2H), 7.35 – 7.23 (m, 4H), 6.34 – 6.19 (m, 1H), 6.06 (d, *J* = 3.2 Hz, 1H), 5.72 (s, 1H), 2.86 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 156.1, 142.5, 141.0, 128.5, 128.0, 126.7, 110.3, 107.4, 70.1.



OH

(*R,E*)-1,3-diphenylprop-2-en-1-ol (**15p**)¹⁰

78% yield, 91% ee, Colorless oil; HPLC (Chiralcel OD-H, 10% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 14.5$ min for enantiomer (R), $t_R =$

18.9 min for enantiomer (S). [*a*]_D²⁰ = +29.8 (*c* = 0.90, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.29 (m, 2H), 7.29 – 7.23 (m, 4H), 7.22 – 7.17 (m, 3H), 7.15 – 7.11 (m, 1H), 6.56 (d, *J* = 15.9 Hz, 1H), 6.38 – 6.09 (m, 1H), 5.24 (d, *J* = 6.5 Hz, 1H), 2.18 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 142.9, 136.7, 131.7, 130.6, 128.7, 128.7, 128.6, 127.9, 127.2, 126.7, 126.5, 75.2.



 $\begin{array}{l} (R)-(4-\text{chlorophenyl})(\text{p-tolyl})\text{ methanol }(15\text{q})^8 \\ \text{Me} \\ (R)-(4-\text{chlorophenyl})(\text{p-tolyl})\text{ methanol }(15\text{q})^8 \\ \text{83\% yield, }97\% \text{ ee, Colorless oil; HPLC (Chiralcel OD-H, 5\% IPA in hexane, 1.0 mL/min, UV 220 nm): } t_R = 10.8 \text{ min for enantiomer }(R), t_R \\ = 11.6 \text{ min for enantiomer }(S). [a]_D^{20} = +8.9 (c = 0.90, \text{CHCl}_3). ^1\text{H NMR }(400 \text{ MHz, CDCl}_3) \delta 7.31 \\ - 7.25 (m, 4\text{H}), 7.23 - 7.18 (m, 2\text{H}), 7.17 - 7.10 (m, 2\text{H}), 5.76 (s, 1\text{H}), 2.32 (s, 3\text{H}). ^{13}\text{C NMR }(100 \\ \end{array}$

MHz, CDCl₃) δ 142.5, 140.7, 137.8, 133.3, 129.5, 128.7, 127.9, 126.6, 75.6, 21.2



OH Me

(R)-(4-chlorophenyl)(o-tolyl)methanol (15r)⁸

89% yield, 93% ee, Colorless oil; HPLC (Chiralcel AD-H, 5% IPA in hexane, 1.0 mL/min, UV 220 nm): $t_R = 11.3$ min for enantiomer (R), $t_R = 12.1$ min

for enantiomer (S). [*a*]_D²⁰ = +49.8 (*c* = 1.20, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.40 (m, 1H), 7.30 – 7.19 (m, 6H), 7.16 – 7.11 (m, 1H), 5.95 (s, 1H), 2.23 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 141.5, 141.2, 135.5, 133.4, 130.8, 128.7, 128.6, 127.9, 126.5, 126.4, 72.9, 19.5.



(R)-N,N-dimethyl-2-(phenyl(o-tolyl)methoxy)ethan-1-amine-Orphenadrine¹¹71% yield, 98% ee, Colorless oil; HPLC (Chiralcel IB, 95:5*n*-Heptane /(Ethanol /*iso*-propanol 50:50 + 0.5% diethylamine), 1.0 mL/min, UV 220 nm): $t_R = 6.9 min for enantiomer (R), t_R = 8.5 min for enantiomer (S). <math>[a]_D^{20} = +4.8$ (*c* = 0.90, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.46 - 7.41 (m, 1H), 7.32 - 7.25 (m, 4H), 7.24 -7.13 (m, 3H), 7.13 - 7.08 (m, 1H), 5.53 (s, 1H), 3.64 - 3.49 (m, 2H), 2.69 - 2.55 (m, 2H), 2.28 (s, 6H), 2.25 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 141.1, 139.8, 135.9, 130.6, 128.3, 127.6, 127.4, 127.1, 126.0, 81.4, 67.3, 58.9, 45.8, 19.5.

HPLC Chromatograms - Enantiomer R

Ω

Me



(*R*)-N,N-dimethyl-2-(phenyl(p-tolyl)methoxy)ethan-1-amine-Neobenodi ne¹²

76% yield, 98% ee, Colorless oil; HPLC (Chiralcel OD-H, 5% IPA in hexane, 1.0 mL/min, UV 220 nm, 1.0 mL/min, UV 220 nm): $t_R = 27.4$ min

for enantiomer (S), t_R = 29.2 min for enantiomer (R). [*a*]_D²⁰ = -6.6 (*c* = 0.90, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.25 (m, 4H), 7.25 – 7.16 (m, 3H), 7.10 (d, *J* = 7.9 Hz, 2H), 5.33 (s, 1H), 3.57 (t, *J* = 5.9 Hz, 2H), 2.64 (t, *J* = 5.9 Hz, 2H), 2.30 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 142.4, 139.3, 137.1, 129.1, 128.4, 127.4, 127.0, 126.9, 83.9, 67.2, 58.8, 45.8, 21.2.



(R)-3-phenylisobenzofuran-1(3H)-one (16a)¹³ 83% yield, 95% ee; after recrystallization: 98.5% ee; White solid. $[a]_D^{20} = -52.98$ (c = 1.20 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 9.2 min for enantiomer (S), t_R = 11.5 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 7.93 – 7.85 (m, 1H), 7.62 – 7.52 (m, 1H), 7.51 – 7.43 (m, 1H), 7.31 – 7.23 (m, 4H), 7.22 – 7.17 (m, 2H), 6.33 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 149.8, 136.5, 134.4, 129.5, 129.4, 129.1, 127.1, 125.8, 125.7, 123.0, 82.8.



After recrystallization



Me (R)-3-(p-tolyl)isobenzofuran-1(3H)-one $(16b)^{13}$



88% yield, 97% ee; after recrystallization: 99.1% ee; White solid. $[\alpha]_D^{20} = -23.88$ (*c* = 1.00 in CHCl₃). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 7.0 min for enantiomer (S), t_R = 8.9 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 7.99 – 7.92 (m, 1H), 7.68 – 7.61 (m, 1H), 7.58 – 7.52

(m, 1H), 7.34 – 7.29 (m, 1H), 7.20 – 7.13 (m, 4H), 6.38 (s, 1H), 2.35 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 150.0, 139.5, 134.4, 133.5, 129.8, 129.7, 129.4, 127.2, 125.8, 125.7, 123.0, 82.9, 21.4.





Me (R)-3-(m-tolyl)isobenzofuran-1(3H)-one (16c)¹⁴ 90% yield, 98% ee; after recrystallization: 99.9% ee; White solid. $[\alpha]_D^{20} = -2.60$ (c = 0.15 in CHCl₃). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 6.2 min for enantiomer (S), t_R = 7.6 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 7.96 (dt, J = 7.6, 1.0 Hz, 1H), 7.65 (td, J = 7.5, 1.2 Hz, 1H), 7.55 (tt, J

= 7.5, 0.8 Hz, 1H), 7.35-7.32 (m, 1H), 7.28 – 7.23 (m, 1H), 7.22 – 7.14 (m, 1H), 7.08 (dd, J = 9.3, 1.9 Hz, 2H), 6.37 (s, 1H), 2.33 (s, 3H).¹³C NMR (100 MHz, CDCl₃) δ 170.7, 149.9, 139.0, 136.4, 134.4, 130.2, 129.4, 129.0, 127.6, 125.7, 125.7, 124.2, 123.0, 82.9, 21.5.





After recrystallization

'n



(R)-3-(o-tolyl)isobenzofuran-1(3H)-one $(16d)^{13}$

Me 84% yield, 94% ee; after recrystallization: 99.9% ee; White solid. $[\alpha]_D^{20} = +13.78$ (c = 0.50 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 8.1 min for enantiomer (S), t_R = 10.5 min for enantiomer (R).

¹H NMR (400 MHz, CDCl₃) δ 7.98 (dt, J = 7.6, 1.0 Hz, 1H), 7.67 (td, J = 7.5, 1.2 Hz, 1H), 7.59-7.55 (m, 1H), 7.36-7.33 (m, 1H), 7.28 – 7.25 (m, 2H), 7.15-7.11 (m, 1H), 6.97 – 6.88 (m, 1H), 6.68 (s, 1H), 2.50 (s, 3H).¹³C NMR (100 MHz, CDCl₃) δ 170.8, 149.4, 137.3, 134.3, 134.2, 131.3, 129.5, 129.4, 127.4, 126.5, 125.9, 123.1, 80.7, 19.5.



After recrystallization





OMe (R)-3-(3-methoxyphenyl)isobenzofuran-1(3H)-one $(16e)^{13}$

84% yield, 90% ee; yellow oil. $[a]_D^{20} = -33.42$ (c = 0.70 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 8.9 min for enantiomer (S), t_R = 12.4 min for enantiomer (R). ¹H NMR (400 MHz,

CDCl₃) δ 7.87 (dd, J = 7.8, 1.0 Hz, 1H), 7.56 (td, J = 7.5, 1.1 Hz, 1H), 7.46 (t, J = 7.4 Hz, 1H), 7.29 – 7.24 (m, 1H), 7.24 – 7.18 (m, 1H), 6.85 – 6.76 (m, 2H), 6.71 (t, J = 2.1 Hz, 1H), 6.29 (s, 1H), 3.69 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 160.1, 149.7, 138.0, 134.5, 130.2, 129.5, 125.7, 125.5, 122.9, 119.2, 114.7, 112.5, 82.6, 55.4.



(R)-3-(naphthalen-1-yl)isobenzofuran-1(3H)-one (16f)¹³

65% yield, 86% ee; after recrystallization: 99.9% ee; White solid. $[\alpha]_D^{20} = +$ 9.81 (c = 0.90 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1

mL/min, UV 215 nm): $t_R = 14.1$ min for enantiomer (S), $t_R = 22.9$ min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 8.30 – 8.22 (m, 1H), 8.02 (dt, J = 7.4, 1.1 Hz, 1H), 7.98 – 7.91 (m, 1H), 7.91 – 7.84 (m, 1H), 7.72 – 7.52 (m, 4H), 7.48 – 7.35 (m, 2H), 7.28 – 7.25 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 149.5, 134.3, 134.2, 132.1, 131.5, 130.1, 129.6, 129.2, 127.2, 126.4, 126.3, 126.2, 125.4, 124.7, 123.4, 123.1, 79.8.



After recrystallization



Peak	Ret. Time.	Area	Area%
1	14.218	8740	0.003
2	22.625	299472667	99.997

 ${\mathsf B}^{\mathsf{r}}$ (R)-3-(4-bromophenyl)isobenzofuran-1(3H)-one (**16g**)¹³

91% yield, 97% ee; after recrystallization: 99.5% ee; White solid. $[\alpha]_D^{20} = -18.77$ (c = 1.00 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 8.3 min for enantiomer (S), t_R = 9.8 min for enantiomer (R).¹H NMR (400 MHz, CDCl₃) δ 7.96 (dt, J = 7.6, 1.0 Hz, 1H), 7.67 (td, J = 7.5, 1.2 Hz, 1H),

7.57 (tt, J = 7.6, 0.8 Hz, 1H), 7.54 – 7.48 (m, 2H), 7.32 (dq, J = 7.8, 0.9 Hz, 1H), 7.20 – 7.12 (m, 2H), 6.36 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 149.3, 135.6, 134.6, 132.3, 129.7, 128.7, 125.9, 125.6, 123.6, 122.9, 82.0.



Peak	Ret. Time.	Area	Area%
1	8.547	901606	1.225
2	10.075	72701751	98.775

After recrystallization



(R)-3-(4-chlorophenyl)isobenzofuran-1(3H)-one (16h)¹³



91% yield, 97% ee; after recrystallization: 99.9% ee; White solid. $[\alpha]_D^{20} = -29.88$ (*c* = 0.98 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 7.9 min for enantiomer (S), t_R = 9.5 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 7.97 (dt, J = 7.7, 1.0 Hz, 1H), 7.67 (td, J = 7.5, 1.2 Hz, 1H), 7.57 (tt, J = 7.5, 0.8 Hz, 1H), 7.40 – 7.29 (m, 3H), 7.25 – 7.19 (m, 2H), 6.38 (s, 1H). ¹³C NMR (100

MHz, CDCl₃) δ 170.4, 149.4, 135.5, 135.1, 134.6, 129.7, 129.4, 128.5, 126.0, 125.7, 122.9, 82.0.





After recrystallization



CI (R)-3-(3-chlorophenyl)isobenzofuran-1(3H)-one (16i)¹⁵ 93% yield, 94% ee; after recrystallization: 99.9% ee; White solid. $[\alpha]_D^{20} = -$ 46.31 (c = 1.20 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 7.9 min for enantiomer (S), t_R = 9.5 min for

enantiomer (R).¹H NMR (400 MHz, CDCl₃) δ 7.90 (dt, J = 7.6, 1.0 Hz, 1H), 7.60 (td, J = 7.5, 1.2

Hz, 1H), 7.52-7.48 (m, 1H), 7.31 – 7.21 (m, 3H), 7.19 (d, *J* = 1.8 Hz, 1H), 7.12 (dt, *J* = 6.8, 1.9 Hz, 1H), 6.29 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 149.2, 138.6, 135.1, 134.7, 130.4, 129.8, 129.6, 127.1, 126.0, 125.5, 125.2, 122.9, 81.8.



After recrystallization



Peak	Ret. Time.	Area	Area%
1	7.717	90132	0.007
2	8.918	84246720	99.993

(R)-3-(4-fluorophenyl)isobenzofuran-1(3H)-one (16j)¹⁵
95% yield, 96% ee; after recrystallization: 99.5% ee; White solid. [α]_D²⁰ = +23.28 (*c* = 1.00 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 7.6 min for enantiomer (S), t_R = 9.1 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) 8 7.96 (dt, *J* = 7.6, 1.0 Hz, 1H), 7.67 (td, *J* = 7.5, 1.2 Hz, 1H), 7.57 (tt, *J* = 7.5, 0.9 Hz, 1H), 7.32 (dt, *J* = 7.7, 1.0 Hz, 1H), 7.30 – 7.21 (m, 2H), 7.07 (ddt, *J* = 8.6, 6.5, 2.5 Hz, 2H), 6.40 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) 8 170.4, 164.5, 162.1, 149.5, 134.6, 132.4, 132.4, 129.6, 129.2, 129.1, 125.8, 125.7, 123.0, 116.2, 116.0, 82.1.


Peak	Ret. Time.	Area	Area%
1	7.748	1247790	1.969
2	9.217	62139491	98.031

After recrystallization



 $-CF_3$ (R)-3-(3-(trifluoromethyl)phenyl)isobenzofuran-1(3H)-one (**16k**)¹⁵ 84% yield, 98% ee; White solid. $[\alpha]_D^{20} = -53.21$ (c = 1.10 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 6.5 min for enantiomer (S), t_R = 7.5 min for enantiomer (R).¹H NMR (400 MHz,

CDCl₃) δ 8.00 (dt, *J* = 7.6, 1.0 Hz, 1H), 7.71 – 7.62 (m, 2H), 7.63 – 7.56 (m, 2H), 7.55 – 7.47 (m, 2H), 7.35 (dq, *J* = 7.7, 0.9 Hz, 1H), 6.46 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.2, 149.0, 137.7, 134.8, 131.60 (q, ²*J*_{C-F} = 32.7 Hz), 130.3, 129.9, 129.8, 126.3 (q, ³*J*_{C-F} = 3.8 Hz), 126.1, 125.5, 123.84 (q, ³*J*_{C-F} = 4.0 Hz), 123.82 (q, ¹*J*_{C-F} = 272.4 Hz), 122.9, 81.8.





(S)-3-(2,4-difluorophenyl)isobenzofuran-1(3H)-one (161)¹³

89% yield, 91% ee; after recrystallization: ee 99.5%; White solid. $[\alpha]_D^{20} = -14.88$ F (*c* = 1.00 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 7.6 min for enantiomer (S), t_R = 8.3 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ 7.97 (d, *J* = 7.6 Hz, 1H), 7.68 (td, *J* = 7.5, 1.1 Hz, 1H), 7.58 (t, *J* = 7.5 Hz, 1H), 7.41 (d, *J* = 7.7 Hz, 1H), 7.26 (s, 1H), 7.11 (td, *J* = 8.5, 6.1 Hz, 1H), 6.95 – 6.82 (m, 2H), 6.70 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 163.6 (dd, ¹*J*_{C-F} = 254.0 Hz, ³*J*_{C-F} =12.5 Hz), 160.94 (dd, ¹*J*_{C-F} = 251.5 Hz, ³*J*_{C-F} = 11.5 Hz), 149.0, 134.7, 129.8, 129.2 (dd, ³*J*_{C-F} = 10.1 Hz, ⁴*J*_{C-F} = 4.9 Hz), 126.0, 125.7, 122.9 (d, *J*_{C-F} = 1.9 Hz), 120.3 (d, ³*J*_{C-F} = 12.7 Hz), 112.2 (dd, ²*J*_{C-F} = 21.5 Hz, ¹*J*_{C-F} = 3.6 Hz), 104.7 (t, *J* = 25.4 Hz), 76.4 (d, ⁴*J*_{C-F} = 3.9 Hz).





After recrystallization





(R)-3-(thiophen-3-yl)isobenzofuran-1(3H)-one $(16m)^{15}$

77% yield, 87% ee; White solid. $[\alpha]_D{}^{20} = +18.19$ (*c* = 0.50 in DCM). HPLC (Chiralcel OD-H, 85% IPA in hexane, 1 mL/min, UV 215 nm): t_R = 10.4 min for enantiomer (S), t_R = 12.3 min for enantiomer (R). ¹H NMR (400 MHz, CDCl₃) δ

7.89 (d, J = 7.6 Hz, 1H), 7.62 (td, J = 7.5, 1.1 Hz, 1H), 7.51 (t, J = 7.5 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 7.31 – 7.22 (m, 2H), 6.88 (dd, J = 4.5, 1.9 Hz, 1H), 6.44 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 149.2, 137.4, 134.4, 129.6, 127.4, 126.1, 126.0, 125.9, 124.6, 123.0, 78.6.



4. Copies of NMR Spectra of Chiral Ligands.









20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -2 f1 (ppm)





= 149.0 = 149.0 = 139.1 = 132.6 = 132.6 = 132.6 = 132.6 = 132.6 = 132.6 = 122.3 = 122.3 = 122.3 = 122.4 = 12



-20.3







- 168.1 - 146.3 - 146.3 - 146.3 - 133.1 - 133.1 - 128.7 - 128.7 - 128.7 - 128.7 - 126.



 $< \frac{20.6}{20.4}$

168.3 143.0 134.8 134.8 134.8 132.6 131.2 132.9 131.2 132.9 132.9 132.9 132.9 132.9 132.9 132.6 132.9 123.8 123.9 122.9 122.9 122.9 122.9 122.9 122.9 122.9 122.9 122.9 122.0

 $< \frac{20.4}{20.0}$

















$\begin{array}{c} - 148.8 \\ 136.7 \\ 131.6 \\ 132.5 \\ 132.5 \\ 132.5 \\ 132.5 \\ 132.5 \\ 132.5 \\ 122.4 \\ 122.8 \\ 122.8 \\ 122.8 \\ 122.8 \\ 122.8 \\ 122.5 \\ 122.$



 $\begin{array}{c} -167.9 \\ 145.7 \\ 133.4.6 \\ 133.4.6 \\ 133.3.6 \\ 133.4.6 \\ 132.6 \\ 132.6 \\ 132.6 \\ 132.6 \\ 126.3 \\ 126.3 \\ 126.3 \\ 126.3 \\ 126.3 \\ 126.3 \\ 125.7 \\ 233.3 \\ 72.4 \\ 25.2 \\ -55.2 \\ -55.2 \\ -55.2 \\ -55.2 \\ 25.2 \\ 25.2 \\ 25.2 \\ 25.2 \\ 25.1$











S61







T.02 1.05 1.02 7.5 6.5 6.0 5.5 5.0 4.5 f1 (ppm) 7.0 2.5 1.0 0.0 9.0 8.5 4.0 2.0 0.5 8.0 3.0





20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -2 f1 (ppm)







188.1 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 137.8 132.5 132.6 132.6 132.6 132.6 132.8 132.8 132.5 122.8 122.8 122.8 122.3 122.4 122.5 122.4 122.5 122.5 122.5 122.5 122.5 122.5 122.5 122.6 122.7 122.7 122.6 122.7 122.6 122.7 122.7 122.7 122.7 122.1 122.3 122.4</



20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -2 f1 (ppm)





168.2 147.6 142.1 137.2 137.5 137.6 137.6 137.6 137.6 137.6 137.6 137.6 137.6 128.3 128.3 128.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4 127.4</



20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -2 f1 (ppm)



147.6 147.1 147.1 147.1 135.2 135.2 135.2 135.2 135.2 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 128.5 125.5 -- 29.4 -- 24.3






20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -2 f1 (ppm)





5. Copies of NMR Spectra of Products















































- 19.5































170.4 164.5 162.1 149.5 132.4 132.4 132.4 132.5 129.6 125.5 125.5 125.5 125.5 116.0 116.0



- 170.2 - 149.0 137.7 132.1 131.8 131.8 131.4 132.1 131.4 131.4 132.3 131.4 132.3 132.5 125.3 12



$\begin{bmatrix} 170.3 \\ 64.7 \\ 164.7 \\ 164.7 \\ 159.7 \\ 159.8 \\ 159.7 \\ 159.1 \\ 159.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 129.1 \\ 120.2 \\ 122.9 \\ 122.2 \\ 12$




6. References

- 1. A. M. M. Carlos, R. Stieler and D. S. Lüdtke, *Org. Biomol. Chem.*, 2019, **17**, 283-289.
- C. Yao, P. Wu, Y. Huang, Y. Chen, L. Li and Y.-M. Li, *Organic & Biomolecular Chemistry*, 2020, 18, 9712-9725.
- 3. M. Hatano, T. Miyamoto and K. Ishihara, *J Org Chem*, 2006, **71**, 6474-6484.
- 4. R. B. Jagt, P. Y. Toullec, J. G. de Vries, B. L. Feringa and A. J. Minnaard, *Org Biomol Chem*, 2006, **4**, 773-775.
- F. Ling, S. Nian, J. Chen, W. Luo, Z. Wang, Y. Lv and W. Zhong, *J Org Chem*, 2018, 83, 10749-10761.
- 6. H. Li, D. Zhu, L. Hua and E. R. Biehl, *Advanced Synthesis & Catalysis*, 2009, **351**, 583-588.
- Y.-X. Yang, Y. Liu, L. Zhang, Y.-E. Jia, P. Wang, F.-F. Zhuo, X.-T. An and C.-S. Da, *J Org Chem*, 2014, **79**, 10696-10702.
- M.-C. Wang, Q.-J. Zhang, W.-X. Zhao, X.-D. Wang, X. Ding, T.-T. Jing and M.-P. Song, J Org Chem, 2008, 73, 168-176.
- 9. H.-F. Duan, J.-H. Xie, W.-J. Shi, Q. Zhang and Q.-L. Zhou, *Org Lett*, 2006, **8**, 1479-1481.
- 10. Z. Lu, H. Zhang, Z. Yang, N. Ding, L. Meng and J. Wang, *ACS Catalysis*, 2019, **9**, 1457-1463.
- P. Chaumont-Olive, M. Rouen, G. Barozzino-Consiglio, A. Ben Abdeladhim, J. Maddaluno and A. Harrison-Marchand, *Angewandte Chemie International Edition*, 2019, 58, 3193-3197.
- 12. W. Liu, J. Guo, S. Xing and Z. Lu, *Org Lett*, 2020, **22**, 2532-2536.
- 13. A. M. M. Carlos, R. Stieler and D. S. Lüdtke, *Org Biomol Chem*, 2019, 17, 283-289.
- X. Song, Y. Z. Hua, J. G. Shi, P. P. Sun, M. C. Wang and J. Chang, *J Org Chem*, 2014, 79, 6087-6093.
- 15. M. Yohda and Y. Yamamoto, *Org Biomol Chem*, 2015, **13**, 10874-10880.