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Horeau Amplification in the Sequential Acylative Kinetic Resolution of (±)-1,2-Diols and (±)-1,3-Diols in Flow

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Experimental Procedures

Reactions involving moisture sensitive reagents were carried out in flame-dried glassware under an argon or nitrogen atmosphere using standard vacuum line techniques and using anhydrous solvents. Anhydrous solvents were obtained from an anhydrous solvent system (purified using an alumina column, Mbraun SPS-800). All other reactions were performed in standard glassware with no precautions to exclude air or moisture. Solvents and commercial reagents were used as supplied without further purification unless otherwise stated.

'in vacuo' refers to the use of either a Büchi Rotavapor R-200 with a Büchi V-491 heating bath and Büchi V-800 vacuum controller, a Büchi Rotavapor R-210 with a Büchi V-491 heating bath and Büchi V-850 vacuum controller, a Heidolph Laborota 4001 with vacuum controller, an IKA RV10 rotary evaporator with a IKA HB10 heating bath and ILMVAC vacuum controller, or an IKA RV10 rotary evaporator with a IKA HB10 heating bath and Vacuubrand CVC3000 vacuum controller. Rotary evaporator condensers are fitted to Julabo FL601 Recirculating Coolers filled with ethylene glycol and set to -5 °C.

Analytical thin layer chromatography was performed on pre-coated aluminium plates (Kieselgel 60 F_{254} silica). TLC visualisation was carried out with ultraviolet light (254 nm), followed by staining with a 1% aqueous KMnO₄ solution. Manual column chromatography was performed in glass columns fitted with porosity 3 sintered discs over Kieselgel 60 silica using the solvent system stated. HPLC analyses were obtained using either a Shimadzu HPLC consisting of a DGU-20A5 degassing unit, LC-20AT liquid chromatography pump, SIL-20AHT autosampler, CMB-20A communications bus module, SPD-M20A diode array detector and a CTO-20A column oven; or a Shimadzu HPLC consisting of a DGU-20A5R degassing unit, LC-20AD liquid chromatography pump, SIL-20AHT autosampler, SPD-20A UV/Vis detector and a CTO-20A column oven. Separation was achieved using DAICEL CHIRALCEL OD-H and OJ-H columns or DAICEL CHIRALPAK ID, IC and AS-H columns. All HPLC traces of enantiomerically-enriched compounds were compared with authentic racemic spectra.

¹H, ¹³C, and ¹⁹F nuclear magnetic resonance (NMR) spectra were acquired on either a Bruker Avance II 400 (¹H 400 MHz; ¹³C 101 MHz, ¹⁹F 377 MHz) or a Bruker Avance II 500 (¹H 500 MHz; ¹³C 126 MHz, ¹⁹F 470 MHz) spectrometer at ambient temperature in the deuterated solvent stated. All chemical shifts are quoted in parts per million (ppm) and referenced to the residual solvent peak. All coupling constants, *J*, are quoted in Hz. Multiplicities are indicated by: s (singlet), brs (broad singlet), d (doublet), t (triplet), q (quartet), hept (heptet), dd (doublet of doublets), dt (doublet of triplets), qd (quartet of doublets), td (triplet of doublets), ddd (doublet of doublet of doublets), dp (doublet of pentets) and m (multiplet). The abbreviation Ar is used to denote aromatic, Ph to denote phenyl, Et

to denote ethyl and *i*-Pr to denote isopropyl. NMR peak assignments of monoester **3** were confirmed using 2D 1 H $^{-13}$ C heteronuclear single quantum coherence (HSQC) and 2D 1 H $^{-13}$ C heteronuclear multiple-bond correlation spectroscopy (HMBC).

Absolute configuration of monoester **3** were determined following hydrolysis to the corresponding diol and through CSP-HPLC analysis.

Melting points were recorded on an Electrothermal 9100 melting point apparatus and are uncorrected. Optical rotations were measured on a Perkin Elmer Precisly/Model-341 polarimeter operating at the sodium D line with a 100 mm path cell at 20 °C.

Infrared spectra were recorded on a Shimadzu IRAffinity-1 Fourier transform IR spectrophotometer fitted with a Specac Quest ATR accessory (diamond puck). Spectra were recorded of thin films, with characteristic absorption wave numbers (vmax) reported in cm^{-1} .

Continuous flow experiments were performed using the same polystyrene-supported (2R,3S)-HyperBTM (8) catalyst batch used in previous experiments.¹ The catalyst resin was packed into an Omnifit column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL]. A Gilson 305 HPLC pump was used to pump solvent for column equilibration and regeneration. A Legato 200 series syringe pump (World Precision Instruments) was used to deliver solutions of reagents.

For the kinetic resolution of monoesters, selectivity factors (*s*) were calculated according to Kagan's equation: $s = \ln((1-c)(1-ee_{rsm}))/\ln((1-c)(1+ee_{rsm})) = \ln(1-c(1+ee_{prod}))/\ln(1-c(1-ee_{prod}))$, wherein c is conversion of the reaction, ee_{prod} is the enantiomeric excess of diester product and ee_{rsm} is the enantiomeric excess of the recovered monoester. Conversions were calculated by the following equation: $c = ee_{rsm}/(ee_{prod}+ee_{rsm})$.²

Additional results of the optimization study for the Sequential Acylative Kinetic Resolution of

(±)-5a (Table S1)

Table S1 Full optimization study for the sequential acylative kinetic resolution of (±)-5a in flow-mode conditions.^a



| Entry | (±)- 5a [M] | (<i>i</i> -PrCO) ₂ O [M] | <i>i</i> -Pr ₂ NEt [M] | Product Ratio (5a:6a:7a) ^b | 5a er (<i>S</i> , <i>S</i>):(<i>R</i> , <i>R</i>) ^c | 6a er $(S,S):(R,R)^{c}$ | 7a er (<i>R</i> , <i>R</i>):(<i>S</i> , <i>S</i>) ^c |
|-------------------|-----------------------|---|--------------------------------------|--|--|--------------------------------|--|
| 1 | 0.2 | 0.3 | 0.32 | 17:39:44 | 99:1 | 85:15 | 95:5 |
| 2^d | 0.2 | 0.3 | 0.32 | 12:39:49 | >99:1 | 92:8 | 97:3 |
| 3 ^e | 0.2 | 0.3 | 0.32 | 4:43:53 | - | 98:2 | 93:7 |
| 4 | 0.1 | 0.15 | 0.16 | 10:44:46 | >99:1 | 75:25 | 94:6 |
| 5 | 0.2 | 0.4 | 0.44 | 1:40:59 | - | >99:1 | 75:25 |
| 6 | 0.2 | 0.35 | 0.35 | 7:41:52 | 99:1 | 97:3 | 90:10 |
| $7^{\rm f}$ | 0.2 | 0.3 | 0.32 | 14:39:47 | 99:1 | 88:12 | 96:4 |
| 8 ^g | 0.2 | 0.3 | 0.32 | 9:39:52 | >99:1 | 96:4 | 94:6 |
| 9 ^h | 0.2 | 0.3 | 0.32 | 10:45:45 | 99:1 | 89:11 | 94:6 |
| 10^{i} | 0.2 | 0.35 | 0.35 | 7:41:52 | >99:1 (3%) ^j | 98:2 (38%) ^j | 94:6 (47%) ^j |
| 11 ^{i,f} | 0.2 | 0.35 | 0.35 | 7:40:53 | >99:1 (3%) ^j | 98:2 (35%) ^j | 94:6 (48%) ^j |

^aConditions: (\pm)-**5a** (0.5 mmol), r.t., flow rate 0.1 mL min⁻¹. ^bDetermined by ¹H NMR spectroscopic analysis of the crude reaction product. ^cDetermined by CSP-HPLC analysis. ^dFlow rate 0.05 mL min⁻¹. ^eFlow rate 0.04 mL min⁻¹. ^fFlow set-up: diol and anhydride in one syringe and base in the other one. ^gFlow set-up: all the reagents together in one syringe. ^bFlow set-up: diol and base in one syringe and anhydride in the other one. ⁱConditions: (\pm)-**5a** (2.0 mmol), r.t., flow rate 0.1 mL min⁻¹. ^jIsolated yield.

Additional results of the optimization study for the Sequential Acylative Kinetic Resolution of

(±)-2a in homogeneous conditions (Table S2)

Table S2 Optimization study for the sequential acylative kinetic resolution of (±)-2a with homogeneous HyperBTM.^a



^aConditions: (±)-**2a** (0.2 M), (EtCO)₂O (1.05 equiv.), *i*-Pr₂NEt (1.05 equiv), r.t. 2 h. ^bIsolated yield. ^cDetermined by CSP-HPLC analysis.

Optimization study for the Sequential Acylative Kinetic Resolution of (±)-2a (Table S3)

Table S3. Optimization study for the sequential acylative kinetic resolution of (\pm) -2a in flow-mode conditions.^a



^aConditions: (\pm)-**2a** (0.5 mmol), r.t., flow rate 0.1 ml min⁻¹. ^bDetermined by ¹H NMR spectroscopic analysis of the crude reaction product. ^cDetermined by CSP-HPLC analysis. ^dIsolated yield. ^eFlow rate 0.2 ml min⁻¹. ^fFlow rate 0.16 ml min⁻¹. ^gFlow set-up: diol and base in one syringe and anhydride in the other one.

Synthesis of (\pm) -syn-1,2-diols 5, monoesters (\pm) -6, and diesters (\pm) -7

(±)-*syn*-1,2-Diols **5a-5g**, monoesters (±)-**6** and diesters (±)-**7** were synthesized following a literature procedure.³ Spectral data were in accordance with the literature.^{4,5}

Synthesis of (±)-anti-1,3-diols 2 and monoesters (±)-3

(\pm)-*anti*-1,3-Diols **2** and monoester (\pm)-**3** were synthesised following a literature procedure.⁶ Spectral data were in accordance with the literature.⁶

<u>General procedure for the Sequential Acylative Kinetic Resolution of (±)-syn-1,2-diols 5 in flow-</u> mode conditions

A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin 8 (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping CHCl₃ at 1 mL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of the appropriate diol (±)-5 (2.0 mmol, 1.0 equiv.) and (*i*-PrCO)₂O (3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) and the second syringe with *i*-Pr₂NEt (3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume). Both solutions were injected at 50 µL min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 100 µL min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 100 µL min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in CHCl₃ was then pumped at 200 µL min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping CHCl₃ at 200 µL min⁻¹ for 30 min. The mixture was diluted with CH₂Cl₂ and washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the crude products which were purified by column chromatography.

Hydrolysis of diesters and monoesters

In some instances it was difficult to find conditions to separate the enantiomers of diesters or monoesters by HPLC using a chiral support, and therefore these products were hydrolysed to the diol prior to HPLC analysis: LiOH•H₂O (3 equiv.) was added to a solution of the diester or monoester (1 equiv.) in MeOH (0.3 M) and allowed to stir at 50 °C until completion, based on TLC analysis. The

mixture was diluted with EtOAc and washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the diol product.

Kinetic Resolution of (±)-1,2-diphenylethane-1,2-diol ((±)-5a)

According to the **General Procedure**, (\pm)-1,2-diphenylethane-1,2-diol (428 mg, 2.0 mmol) and (*i*-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume), and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*S*,2*S*)-1,2-Diphenylethane-1,2-diol (5a)



(13 mg, 3%) as a colourless solid with spectroscopic data in accordance with the literature.^{3,4} **mp** 118–119 °C {Lit.³ 121 °C}; $[\alpha]_D^{20}$ –87.1 (*c* 0.55 in CHCl₃) {Lit.³ (*ent*, >99:1 er) $[\alpha]_D^{20}$ +88.0 (*c* 0.15 in CHCl₃)}; **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(1*R*,2*R*): 11.6 min, t_R(1*S*,2*S*): 14.9 min, 0.27:99.73 er; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 2.74 – 2.97 (brs, 2H, OH), 4.74 (s, 2H, CHOH), 7.10 – 7.20 (m, 4H, ArH), 7.20 – 7.28 (m, 6H, ArH). (**1S**,2*S*)-2-Hydroxy-1,2-diphenylethyl isobutyrate (6a)



(200 mg, 35%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 80–82 °C; $[\alpha]_D^{20}$ –6.2 (*c* 0.50 in CHCl₃) {Lit.³ (*ent*, >99:1 er) $[\alpha]_D^{20}$ +2.6 (*c* 0.50 in CHCl₃)}; **Chiral HPLC analysis** Chiralcel OD-H (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*S*,2*S*): 11.9 min, t_R (1*R*,2*R*): 17.5 min, 97.90:2.10 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.18 (d, *J* 7.0 Hz, 3H, CH₃), 1.19 (d, *J* 7.0 Hz, 3H, CH₃), 2.53 (d, *J* 3.6 Hz, 1H, OH), 2.66 (hept, *J* 7.0 Hz, 1H, CHCH₃), 4.96 (dd, *J* 7.1, 3.3 Hz, 1H, CHOH), 5.88 (d, *J* 7.1 Hz, 1H, CHOC(O)*i*-Pr), 7.07 – 7.22 (m, 4H, ArH), 7.22 – 7.28 (m, 6H, ArH).

(1R,2R)-1,2-Diphenylethane-1,2-diyl bis(2-methylpropanoate) (7a)



(340 mg, 48%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 71–72 °C {Lit.⁵ 73–74 °C}; $[\alpha]_D^{20}$ –22.9 (*c* 1.0 in CHCl₃) {Lit.³ (*ent*, 97:3 er) $[\alpha]_D^{20}$ +20.2 (*c* 1.0 in CHCl₃)}; following hydrolysis to 1,2-diphenylethane-1,2-diol: **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(1*R*,2*R*): 12.7 min, t_R(1*S*,2*S*): 15.4

min, 94.45:5.55 er; ¹H NMR (400 MHz, CDCl₃) δ_H: 1.16 (d, *J* 7.0 Hz, 6H, CH₃), 1.17 (d, *J* 7.0 Hz, 6H, CH₃), 2.60 (hept, *J* 7.0 Hz, 2H, CHCH₃), 6.07 (s, 2H, CHOC(O)*i*-Pr), 7.07 – 7.18 (m, 4H, ArH), 7.17 – 7.27 (m, 6H, ArH).

Kinetic Resolution of (±)-1,2-bis(4-chlorophenyl)ethane-1,2-diol ((±)-5b)

According to the **General Procedure**, (\pm) -1,2-bis(4-chlorophenyl)ethane-1,2-diol (567 mg, 2.0 mmol) and (i-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*S*,2*S*)-1,2-bis(4-chlorophenyl)ethane-1,2-diol (5b)



(45 mg, 8%) as a colourless solid with spectroscopic data in accordance with the literature.⁷ mp 131–133 °C {Lit.⁸ 126–128 °C}; $[\alpha]_D^{20}$ –183.0 (*c* 0.25 in CHCl₃) {Lit.³ (*ent*, >99:1 er) $[\alpha]_D^{20}$ +112 (*c* 0.25 in CHCl₃)}; Chiral HPLC analysis Chiralpak AS-H (98:2 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 41.5 min, 0:100 er; ¹H NMR (500 MHz, CDCl₃) $\delta_{\rm H}$: 2.91 (q, *J* 1.2 Hz, 2H, OH), 4.65 (t, *J* 1.2 Hz, 2H, CHOH), 7.00 – 7.09 (m, 4H, ArH), 7.18 – 7.27 (m, 4H, ArH).

(1*S*,2*S*)-1,2-bis(4-chlorophenyl)-2-hydroxyethyl isobutyrate (6b)



(240 mg, 34%) as a colourless solid with spectroscopic data in accordance with the literature.³ **mp** 141–142 °C {Lit.³ 143 °C}; $[\alpha]_D^{20}$ –34.5 (*c* 1.0 in CHCl₃) {Lit.³ (*ent*, 99:1 er) $[\alpha]_D^{20}$ +35.0 (*c* 1.0 in CHCl₃)}; **Chiral HPLC analysis** Chiralpak AS-H (99.5:0.5 Hexane:IPA, flow rate 0.7 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 35.9 min, t_R (1*R*,2*R*): 46.7 min, 95.53:4.47 er; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.18 (d, *J* 7.0 Hz, 3H, CH₃), 1.20 (d, *J* 7.0 Hz, 3H, CH₃), 2.54 (d, *J* 3.4 Hz, 1H, OH), 2.65 (hept, *J* 7.0 Hz, 1H, CHCH₃), 4.91 (dd, *J* 7.3, 3.3 Hz, 1H, CHOH), 5.76 (d, *J* 7.3 Hz, 1H, CHOC(O)*i*-Pr), 6.98 – 7.13 (m, 4H, ArH), 7.19 – 7.28 (m, 4H, ArH).

(1R,2R)-1,2-bis(4-chlorophenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7b)



(381 mg, 45%) as a colourless oil with spectroscopic data in accordance with the literature.³ $[\alpha]_D^{20}$ -5.3 (*c* 1.0 in CHCl₃) {Lit.³ (*ent*, 95:5 er) $[\alpha]_D^{20}$ +4.8 (*c* 0.5 in CHCl₃)}; following hydrolysis to 1,2bis(4-chlorophenyl)ethane-1,2-diol: **Chiral HPLC analysis** Chiralpak AS-H (98:2 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*R*,2*R*): 31.0 min, t_R (1*S*,2*S*): 43.8 min, 93.62:6.38 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.15 (d, *J* 7.0 Hz, 6H, CH₃), 1.17 (d, *J* 7.0 Hz, 6H, CH₃), 2.59 (hept, *J* 7.0 Hz, 2H, CHCH₃), 5.99 (s, 2H, CHOC(O)*i*-Pr), 7.02 – 7.12 (m, 4H, ArH), 7.17 – 7.27 (m, 4H, ArH).

Kinetic Resolution of (±)-1,2-bis(4-chlorophenyl)ethane-1,2-diol ((±)-5c)

According to the **General Procedure**, (\pm) -1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diol (700 mg, 2.0 mmol) and (*i*-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(15,2S)-1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diol (5c)



(8 mg, 1%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 135–137 °C {Lit.⁹ 128–130 °C}; $[\alpha]_D^{20}$ –10.7 (*c* 0.7 in CHCl₃) {Lit.³ (*ent*, >99:1 er) $[\alpha]_D^{20}$ +41.1 (*c* 1.0 in CHCl₃)}; **Chiral HPLC analysis** Chiralpak AS-H (97:3 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*R*,2*R*): 11.3 min, t_R (1*S*,2*S*): 15.6 min, 0.27:99.73 er; ¹**H** NMR (500 MHz, CDCl₃) $\delta_{\rm H}$: 2.98 (t, *J* 1.4 Hz, 2H, OH), 4.73 – 4.81 (m, 2H, CHOH), 7.26 (d, *J* 8.0 Hz, 4H, ArH), 7.54 (d, *J* 8.1 Hz, 4H, ArH); ¹⁹**F** NMR (377 MHz, CDCl₃) $\delta_{\rm F}$: –62.58.

(15,2S)-2-hydroxy-1,2-bis(4-(trifluoromethyl)phenyl)ethyl isobutyrate (6c)



(285 mg, 34%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 147–148 °C {Lit.⁵ 148 °C}; $[\alpha]_D^{20}$ –4.4 (*c* 1.0 in CHCl₃) {Lit.⁵ (96:4 er) $[\alpha]_D^{23}$ –1.06 (*c* 1.01 in

CHCl₃)}; **Chiral HPLC analysis** Chiralcel OJ-H (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 7.7 min, 100:0 er; ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.19 (d, *J* 7.1, 6H, CH₃), 2.58 (d, *J* 3.7 Hz, 1H, OH), 2.67 (hept, *J* 7.0 Hz, 1H, CHCH₃), 5.05 (dd, *J* 6.7, 3.7 Hz, 1H, CHOH), 5.91 (d, *J* 6.6 Hz, 1H, CHOC(O)*i*-Pr), 7.23 – 7.34 (m, 4H, ArH), 7.55 (dd, *J* 8.3, 2.1 Hz, 4H, ArH); ¹⁹F NMR (377 MHz, CDCl₃) $\delta_{\rm F}$: -62.61, -62.68.

(1R,2R)-1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7c)



(520 mg, 53%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 109–110 °C {Lit.⁴ 108 °C}; $[\alpha]_D^{20}$ –9.0 (*c* 1.0 in CHCl₃) {Lit.³ (*ent*, 97:3 er) $[\alpha]_D^{20}$ +11.0 (*c* 0.10 in CHCl₃)}; following hydrolysis to 1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diol: **Chiral HPLC analysis** Chiralpak AS-H (97:3 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*R*,2*R*): 11.5 min, t_R (1*S*,2*S*): 15.8 min, 81.67:18.33 er; ¹**H** NMR (400 MHz, CDCl₃) δ_{H} : 1.16 (d, *J* 7.0 Hz, 12H, CH₃), 2.55 – 2.69 (m, *J* 6.9 Hz, 2H, CHCH₃), 6.12 (s, 2H, CHOC(O)*i*-Pr), 7.34 – 7.22 (m, 4H, ArH), 7.46 – 7.62 (m, 4H, ArH); ¹⁹**F** NMR (377 MHz, CDCl₃) δ_{F} : –62.72.

Kinetic Resolution of (±)-1,2-bis(4-methoxyphenyl)ethane-1,2-diol ((±)-5d)

According to the **General Procedure**, (\pm)-1,2-diphenylethane-1,2-diol (548 mg, 2.0 mmol) and (*i*-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.), in CHCl₃ (10 mL total volume), and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*S*,2*S*)-1,2-bis(4-methoxyphenyl)ethane-1,2-diol (5d)



(33 mg, 6%) as a colourless solid with spectroscopic data in accordance with the literature.¹⁰ **mp** 106–107 °C {Lit.⁸ 118–119 °C}; $[\alpha]_D^{20}$ –102 (*c* 0.25 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, >99:1 er) $[\alpha]_D^{20}$ +126 (*c* 0.25 in CHCl₃)}; **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*S*,2*S*): 24.09 min, 0:100 er; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 3.79 (s, 6H, OCH₃), 4.66 (s, 2H, CHOH), 6.74 – 6.84 (m, 4H, ArH), 7.02 – 7.11 (m, 4H, ArH).

(15,2S)-2-hydroxy-1,2-bis(4-methoxyphenyl)ethyl isobutyrate (6d)



(254 mg, 37%) as a colourless solid with spectroscopic data in accordance with the literature.³ **mp** 88–89 °C {Lit.³ 88 °C}; $[\alpha]_D^{20}$ –26.7 (*c* 0.5 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 99:1 er) +23.6 (*c* 0.5 in CHCl₃)}; following hydrolysis to 1,2-bis(4-methoxyphenyl)ethane-1,2-diol: **Chiral HPLC analysis** Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*R*,2*R*): 21.91 min, t_R (1*S*,2*S*): 36.53 min, 1.56:98.44 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.18 (d, *J* 7.0 Hz, 3H, CH₃), 1.20 (d, *J* 7.0 Hz, 3H, CH₃), 2.65 (hept, *J* 7.0 Hz, 1H, CHCH₃), 3.78 (d, *J* 3.8 Hz, 6H, OCH₃), 4.89 (d, *J* 7.8 Hz, 1H, CHOH), 5.78 (d, *J* 7.7 Hz, 1H, CHOC(O)*i*-Pr), 6.71 – 6.84 (m, 4H, ArH), 6.98 – 7.13 (m, 4H, ArH).

(1R,2R)-1,2-bis(4-methoxyphenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7d)



(380 mg, 45%) as a colourless oil with spectroscopic data in accordance with the literature.⁵ mp 122–124 °C {Lit.³ 126–128 °C}; $[\alpha]_D^{20}$ –25.0 (*c* 1.0 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 98:2 er) +19.3 (*c* 1.0 in CHCl₃)}; following hydrolysis to 1,2-bis(4-methoxyphenyl)ethane-1,2-diol: Chiral HPLC analysis Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*R*,2*R*): 21.81 min, t_R (1*S*,2*S*): 38.61 min, 94.66:5.34 er; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 1.15 (d, *J* 7.0 Hz, 6H, CH₃), 1.18 (d, *J* 7.0 Hz, 6H, CH₃), 2.58 (hept, *J* 7.0 Hz, 2H, CHCH₃), 3.76 (s, 6H, OCH₃), 6.00 (s, 2H, CHOC(O)*i*-Pr), 6.68 – 6.83 (m, 4H, ArH), 7.00 – 7.11 (m, 4H, ArH).

Kinetic Resolution of (±)-1,2-di(naphthalen-1-yl)ethane-1,2-diol ((±)-5e)

According to the **General Procedure**, (\pm) -1,2-di(naphthalen-1-yl)ethane-1,2-diol (628 mg, 2.0 mmol) and (i-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.), in 1:1 CHCl₃:THF mixture (10 mL total volume), and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in 1:1 CHCl₃:THF mixture (10 mL total volume) gave crude products that by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*S*,2*S*)-1,2-di(naphthalen-1-yl)ethane-1,2-diol (5e)



(125 mg, 20%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 128–130 °C {Lit.¹¹ 122–124 °C}; $[\alpha]_D^{20}$ –28.6 (*c* 0.25 in CHCl₃) {Lit.⁵ $[\alpha]_D^{24}$ (*ent*, 88:12 er) +40.7 (*c* 0.99 in THF)}; **Chiral HPLC** Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 22.6 min, t_R (1*R*,2*R*): 33.7 min, 97.49:2.51 er; ¹**H NMR** (500 MHz, CDCl₃) $\delta_{\rm H}$: 3.00 (dd, *J* 2.1, 1.1 Hz, 2H, OH), 5.76 – 5.86 (m, 2H, CHOH), 7.31 (ddd, *J* 8.3, 6.8, 1.4 Hz, 2H, ArH), 7.35 – 7.47 (m, 4H, ArH), 7.65 – 7.87 (m, 6H, ArH), 7.90 (dd, *J* 8.6, 1.2 Hz, 2H, ArH).

(1S,2S)-2-hydroxy-1,2-di(naphthalen-1-yl)ethyl isobutyrate (6e)



(177 mg, 23%) as a colourless oil with spectroscopic data in accordance with the literature.⁵ **mp** 102–104 °C {Lit.⁵ 110–111 °C}; $[\alpha]_D^{20}$ –49.4 (*c* 1.2 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, >99:1 er) +57.2 (*c* 1.0 in CHCl₃)}; following hydrolysis to 1,2-di(naphthalen-1-yl)ethane-1,2-diol **Chiral HPLC** Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 23.1 min, t_R (1*R*,2*R*): 33.5 min, 70.30:29.70 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} :. 1.18 (d, *J* 7.0 Hz, 3H, CH₃), 1.20 (d, *J* 7.0 Hz, 3H, CH₃), 2.66 (dp, *J* 31.8, 7.0 Hz, 1H, CHCH₃), 5.95 (d, *J* 6.5 Hz, 1H, CHOH), 6.94 (d, *J* 6.5 Hz, 1H, CHOC(O)*i*-Pr), 7.31 – 7.47 (m, 6H, ArH), 7.48 – 7.63 (m, 2H, ArH), 7.66 – 7.80 (m, 4H, ArH), 8.01 – 8.14 (m, 2H, ArH).

(1R,2R)-1,2-di(naphthalen-1-yl)ethane-1,2-diyl bis(2-methylpropanoate) (7e)



(372 mg, 41%) as a colourless solid with spectroscopic data in accordance with the literature.⁵ **mp** 115–117 °C {Lit.⁵ 113–114 °C}; $[\alpha]_D^{20}$ +71.2 (*c* 1.0 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 95:5 er) –67.5 (*c* 1.0 in CHCl₃)}; following hydrolysis to 1,2-di(naphthalen-1-yl)ethane-1,2-diol **Chiral HPLC analysis** Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 23.9 min, t_R (1*R*,2*R*): 32.4 min, 6.41:93.59 er; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.17 (d, *J* 7.0 Hz, 6H, CH₃), 1.22 (d, *J* 7.0 Hz, 6H, CH₃), 2.55 – 2.75 (m, 2H, CHCH₃), 7.11 (s, 2H, CHOC(O)*i*-Pr), 7.24 (t, *J* 7.7 Hz, 2H, ArH), 7.39 (td, *J* 6.8, 5.9, 2.9 Hz, 6H, ArH), 7.60 – 7.77 (m, 4H, ArH), 8.11 – 8.31 (m, 2H, ArH).

Kinetic Resolution of (±)-(1E,5E)-1,6-diphenylhexa-1,5-diene-3,4-diol ((±)-5f)

According to the **General Procedure**, (\pm)-(1*E*,5*E*)-1,6-diphenylhexa-1,5-diene-3,4-diol (532 mg, 2.0 mmol) and (*i*-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.), in CHCl₃ (10 mL total volume), and *i*-Pr₂NEt (610 µL, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1E,3S,4S,5E)-4-hydroxy-1,6-diphenylhexa-1,5-dien-3-yl isobutyrate (6f)



(208 mg, 31%) as a colourless solid with spectroscopic data in accordance with the literature.³ **mp** 102–103 °C {Lit.³ 95–96 °C}; $[\alpha]_D^{20}$ –0.6 (*c* 0.4 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 77:23 er) +0.9 (*c* 1.0 in CHCl₃)}; **Chiral HPLC analysis** Chiralpak IC (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 254 nm, 30 °C) t_R (3*S*,4*S*): 18.0 min, t_R (3*R*,4*R*): 22.7 min, 80.96:19.04 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.21 (d, *J* 7.1 Hz, 3H, CH₃), 1.23 (d, *J* 7.1 Hz, 3H, CH₃), 2.66 (hept, *J* 6.8 Hz, 1H, CHCH₃), 4.53 (td, *J* 5.7, 1.5 Hz, 1H, CHOH), 5.54 (ddd, *J* 7.0, 5.6, 1.1 Hz, 1H, CHOC(O)*i*-Pr), 6.26 (ddd, *J* 16.0, 6.5, 5.2 Hz, 2H, PhCH=CH), 6.75 (dt, *J* 16.0, 1.5 Hz, 2H, PhCH=CH), 7.32 – 7.53 (m, 10H, ArH).

(1E,3R,4R,5E)-1,6-diphenylhexa-1,5-diene-3,4-diyl bis(2-methylpropanoate) (7f)



(422 mg, 52%) as a colourless solid with spectroscopic data in accordance with the literature.³ **mp** 78–80 °C {Lit.³ 84–85 °C}; $[\alpha]_D^{20}$ –4.57 (*c* 0.35 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 79:21 er) +10.8 (*c* 1.0 in CHCl₃)}; following hydrolysis to (1*E*,5*E*)-1,6-diphenylhexa-1,5-diene-3,4-diol: **Chiral HPLC analysis** Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 254 nm, 30 °C) t_R (1*S*,2*S*): 13.0 min, t_R (1*R*,2*R*): 14.7 min, 35.00:65.00 er; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.17 (d, *J* 7.1 Hz, 12H, CH₃), 2.56 – 2.71 (m, *J* 7.0 Hz, 2H, CHCH₃), 5.62 – 5.75 (m, 2H, CHOC(O)*i*-Pr), 6.08 – 6.23 (m, 2H, PhCH=CH), 6.72 (d, *J* 15.9 Hz, 2H, PhCH=CH), 7.23 – 7.43 (m, 10H, ArH).

Kinetic Resolution of (±)-1,6-diphenylhexa-1,5-diyne-3,4-diol ((±)-5g)

According to the **General Procedure**, (\pm)-1,6-diphenylhexa-1,5-diyne-3,4-diol (525 mg, 2.0 mmol) and (*i*-PrCO)₂O (580 µL, 3.5 mmol, 1.75 equiv.), in CHCl₃ (10 mL total volume), and *i*-Pr₂NEt (610

 μ L, 3.5 mmol, 1.75 equiv.) in CHCl₃ (10 mL total volume) gave crude products that were purified by by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(3S,4S)-1,6-diphenylhexa-1,5-diyne-3,4-diol (5g)



(10 mg, 2%) as a colourless solid with spectroscopic data in accordance with the literature.³ mp 74–77 °C {Lit.³ 78–79 °C}; $[\alpha]_D^{20}$ –101 (*c* 0.1 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, >99:1 er) +97.2 (*c* 0.25 in CHCl₃)}; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 13.6 min, t_R (3*R*,4*R*): 19.1 min, 77.37:22.63 er; ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 4.77 (s, 2H, CHOH), 7.31 – 7.40 (m, 6H, ArH), 7.46 – 7.53 (m, 4H, ArH).

(3S,4S)-4-hydroxy-1,6-diphenylhexa-1,5-diyn-3-yl isobutyrate (6g)



(99 mg, 15%) as a colourless solid with spectroscopic data in accordance with the literature.³ mp 71–72 °C {Lit.³ 71 °C}; $[\alpha]_D^{20}$ +12.8 (*c* 1.0 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 85:15 er) –12.2 (*c* 1.0 in CHCl₃)}; following hydrolysis to 1,6-diphenylhexa-1,5-diyne-3,4-diol: Chiral HPLC analysis Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 12.6 min, t_R (3*R*,4*R*): 18.1 min, 88.34:11.66 er; ¹H NMR (500 MHz, CDCl₃) δ_{H} : 1.26 (d, *J* 7.0 Hz, 3H, CH₃), 1.27 (d, *J* 7.0 Hz, 3H, CH₃), 2.72 (hept, *J* 7.0 Hz, 1H, CHCH₃), 4.87 (d, *J* 5.7 Hz, 1H, CHOH), 5.86 (d, *J* 5.7 Hz, 1H, CHOC(O)*i*-Pr), 7.30 – 7.40 (m, 6H, ArH), 7.43 – 7.55 (m, 4H, ArH).

(3R,4R)-1,6-diphenylhexa-1,5-diyne-3,4-diyl bis(2-methylpropanoate) (7g)



(569 mg, 70%) as a colourless oil with spectroscopic data in accordance with the literature.³ $[\alpha]_D^{20}$ -10.4 (*c* 1.0 in CHCl₃) {Lit.³ $[\alpha]_D^{20}$ (*ent*, 83:17 er) +39.8 (*c* 0.5 in CHCl₃)}; following hydrolysis to 1,6-diphenylhexa-1,5-diyne-3,4-diol: **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 12.6 min, t_R (3*R*,4*R*): 18.1 min, 41.64:58.36 er; ¹**H NMR** (500 MHz, CDCl₃) δ_{H} : 1.24 (d, *J* 7.0 Hz, 6H, CH₃), 1.25 (d, *J* 7.0 Hz, 6H, CH₃), 2.68 (hept, *J* 7.0 Hz, 2H, CHCH₃), 6.00 (s, 2H, CHOC(O)*i*-Pr), 7.30 – 7.40 (m, 6H, ArH), 7.43 – 7.50 (m, 4H, ArH).

Procedure for the Kinetic Resolution of (±)-5a in flow-mode conditions (to lower conversion)

A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin 8 (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping CHCl₃ at 1 mL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of (±)-1,2-diphenylethane-1,2-diol 5a (2 mmol, 1.0 equiv.) and (i-PrCO)₂O (2 mmol, 1.0 equiv.) in CHCl₃ (10 mL total volume) and the second syringe with *i*-Pr₂NEt (2 mmol, 1.0 equiv.) in CHCl₃ (10 mL total volume). Both solutions were injected at 50 µL min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 100 µL min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 100 µL min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in CHCl₃ was then pumped at 200 µL min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping CHCl₃ at 200 μ L min⁻¹ for 30 min. The mixture was diluted with CH₂Cl₂ and washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the crude products. The corresponding diol and monoester were then isolated by flash chromatography (60:40 Hexane:EtOAc).

(15,2S)-1,2-Diphenylethane-1,2-diol (5a): Chiral HPLC analysis Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(1R,2R)$: 12.8 min, $t_R(1S,2S)$: 15.3 min, 0.11:99.89 er.



| Detector A Channel 1 211nm | | | |
|----------------------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 12.783 | 0.106 | |
| 2 | 15.291 | 99.894 | |
| Total | | 100.000 | |

Procedure for the Kinetic Resolution of (±)-6a in flow-mode conditions

A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin 8 (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping CHCl₃ at 1 mL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of 2-hydroxy-1,2-diphenylethyl isobutyrate 6a (0.3 mmol, 1.0 equiv.) and (i-PrCO)₂O (0.26 mmol, 0.85 equiv.) in CHCl₃ (1.5 mL total volume) and the second syringe with *i*-Pr₂NEt (0.26 mmol, 0.85 equiv.) in CHCl₃ (1.5 mL total volume). Both solutions were injected at 50 μ L min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 100 μ L min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 100 µL min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in CHCl₃ was then pumped at 200 µL min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping CHCl₃ at 200 µL min⁻¹ for 30 min. The mixture was diluted with CH₂Cl₂ and washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the crude products. The corresponding monoester and diester were then isolated by flash chromatography (80:20 Hexane:EtOAc).

(15,25)-2-Hydroxy-1,2-diphenylethyl isobutyrate (6a): Chiral HPLC analysis Chiralcel OD-H (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*S*,2*S*): 11.9 min, t_R (1*R*,2*R*): 17.5 min, 82.19:17.82 er.



| Detect | Detector A Channel 1 211nm | | | |
|--------|----------------------------|---------|--|--|
| Peak# | Ret. Time | Area% | | |
| 1 | 11.810 | 82.186 | | |
| 2 | 17.284 | 17.814 | | |
| Total | | 100.000 | | |

(1*R*,2*R*)-1,2-Diphenylethane-1,2-diyl bis(2-methylpropanoate) (7a): Chiral HPLC analysis Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(1R,2R)$: 12.7 min, $t_R(1S,2S)$: 15.4 min, 95.20:4.80 er.



| Detector A Channel 1 211nm | | | |
|----------------------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 12.719 | 95.196 | |
| 2 | 15.044 | 4.804 | |
| Total | | 100.000 | |

HPLC chromatograms for the Sequential Acylative Kinetic Resolution of (±)-syn-1,2-diols 5 (15,2S)-1,2-Diphenylethane-1,2-diol (5a)



Chiral HPLC analysis Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(1R,2R)$: 11.6 min, $t_R(1S,2S)$: 14.9 min, 0.27:99.73 er.



<Peak Table>

| Detect | Detector A Channel 1 211nm | | | | |
|--------|----------------------------|---------|--|--|--|
| Peak# | Ret. Time | Area% | | | |
| 1 | 12.853 | 49.934 | | | |
| 2 | 15.129 | 50.066 | | | |
| Total | | 100.000 | | | |



| Detector A Channel 1 211nm | | | | |
|----------------------------|-----------|---------|--|--|
| Peak# | Ret. Time | Area% | | |
| 1 | 12.692 | 0.275 | | |
| 2 | 14.940 | 99.725 | | |
| Total | | 100.000 | | |

(1S,2S)-2-Hydroxy-1,2-diphenylethyl isobutyrate (6a)



Chiral HPLC analysis Chiralcel OD-H (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*S*,2*S*): 11.9 min, t_R (1*R*,2*R*): 17.5 min, 97.90:2.10 er.



17.520

17.5

20.0

min



| Detect | Detector A Channel 1 211nm | | | |
|--------|----------------------------|---------|--|--|
| Peak# | Ret. Time | Area% | | |
| 1 | 11.859 | 97.904 | | |
| 2 | 17.520 | 2.096 | | |
| Total | | 100.000 | | |

(1*R*,2*R*)-1,2-Diphenylethane-1,2-diol (7a)



1,2-Diphenylethane-1,2-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,2-diphenylethane-1,2-diol; **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(1R,2R)$: 12.7 min, $t_R(1S,2S)$: 15.4 min, 94.45:5.55 er.



| Detector A Channel 1 211nm | | | |
|----------------------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 12.859 | 94.449 | |
| 2 | 15.338 | 5.551 | |
| Total | | 100.000 | |

(1*S*,2*S*)-1,2-bis(4-chlorophenyl)ethane-1,2-diol (5b)



Chiral HPLC analysis Chiralpak AS-H (98:2 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 41.5 min, 0:100 er.



<Peak Table>

| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 31.137 | 50.572 |
| 2 | 41.198 | 49.428 |
| Total | | 100.000 |





| PDA C | h2 220nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 41.535 | 100.000 |
| Total | | 100.000 |

(1*S*,2*S*)-1,2-bis(4-chlorophenyl)-2-hydroxyethyl isobutyrate (6b)



Chiral HPLC analysis Chiralpak AS-H (99.5:0.5 Hexane:IPA, flow rate 0.7 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 35.9 min, t_R (1*R*,2*R*): 46.7 min, 95.53:4.47 er.



| PDA C | h2 220nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 34.477 | 95.529 |
| 2 | 45.048 | 4.471 |
| Total | | 100.000 |

(1R,2R)-1,2-bis(4-chlorophenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7b)



1,2-bis(4-chlorophenyl)ethane-1,2-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,2-bis(4-chlorophenyl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak AS-H (98:2 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*R*,2*R*): 31.0 min, t_R (1*S*,2*S*): 43.8 min, 93.62:6.38 er;



<Peak Table>

| PDA C | PDA Ch2 220nm | | |
|-------|---------------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 30.078 | 93.621 | |
| 2 | 41.982 | 6.379 | |
| Total | | 100.000 | |

(1*S*,2*S*)-1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diol (5c)



Chiral HPLC analysis Chiralpak AS-H (97:3 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*R*,2*R*): 11.3 min, t_R (1*S*,2*S*): 15.6 min, 0.27:99.73 er.



| PDA Ch2 220nm | | |
|---------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.347 | 0.274 |
| 2 | 15.641 | 99.726 |
| Total | | 100.000 |

(15,25)-2-hydroxy-1,2-bis(4-(trifluoromethyl)phenyl)ethyl isobutyrate (6c)



Chiral HPLC analysis Chiralcel OJ-H (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 7.7 min, 100:0 er.



<Peak Table>

| PDA | PDA Ch2 220nm | | |
|-----|---------------|-----------|---------|
| Pea | ık# | Ret. Time | Area% |
| | 1 | 7.704 | 50.553 |
| | 2 | 19.403 | 49.447 |
| To | otal | | 100.000 |





| PDA Ch2 220nm | | | |
|---------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 7.706 | 100.000 | |
| Total | | 100.000 | |

(1*R*,2*R*)-1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7c)



1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak AS-H (97:3 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*R*,2*R*): 11.5 min, t_R (1*S*,2*S*): 15.8 min, 81.67:18.33 er.



<Peak Table>

| PDA Ch2 220nm | | | |
|---------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 11.680 | 49.936 | |
| 2 | 15.945 | 50.064 | |
| Total | | 100.000 | |
| | | | |



| PDA C | h2 220nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.475 | 81.670 |
| 2 | 15.837 | 18.330 |
| Total | | 100.000 |

(1S,2S)-1,2-bis(4-methoxyphenyl)ethane-1,2-diol (5d)



Chiral HPLC analysis Chiralpak ID (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*S*,2*S*): 24.09 min, 0:100 er.



<Peak Table>

| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 15.484 | 49.915 |
| 2 | 25.341 | 50.085 |
| Total | | 100.000 |





<Peak Table>

| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 24.093 | 100.000 |
| Total | | 100.000 |

(1*S*,2*S*)-2-hydroxy-1,2-bis(4-methoxyphenyl)ethyl isobutyrate (6d)



2-hydroxy-1,2-bis(4-methoxyphenyl)ethyl isobutyrate could not be separated by HPLC so it was hydrolysed to give 1,2-bis(4-methoxyphenyl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*R*,2*R*): 21.91 min, t_R (1*S*,2*S*): 36.53 min, 1.56:98.44 er.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 21.911 | 1.557 |
| 2 | 36.527 | 98.443 |
| Total | | 100.000 |

(1R,2R)-1,2-bis(4-methoxyphenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7d)



1,2-bis(4-methoxyphenyl)ethane-1,2-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,2-bis(4-methoxyphenyl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (1*R*,2*R*): 21.81 min, t_R (1*S*,2*S*): 38.61 min, 94.66:5.34 er.



Detector & Channel 1 211

| Delector A Channel 1 2 mm | | |
|---------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 21.818 | 94.655 |
| 2 | 38.611 | 5.345 |
| Total | | 100.000 |

(1*S*,2*S*)-1,2-di(naphthalen-1-yl)ethane-1,2-diol (5e)



Chiral HPLC analysis Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 22.6 min, , t_R (1*R*,2*R*): 33.7 min, 97.49:2.51 er.



| PDA Ch2 220nm | | | |
|---------------|-----------|---------|--|
| Peak# | Ret. Time | Area% | |
| 1 | 22.550 | 97.485 | |
| 2 | 33.669 | 2.515 | |
| Total | | 100.000 | |

(1S,2S)-2-hydroxy-1,2-di(naphthalen-1-yl)ethyl isobutyrate (6e)



2-hydroxy-1,2-di(naphthalen-1-yl)ethyl isobutyrate could not be separated by HPLC so it was hydrolysed to give 1,2-di(naphthalen-1-yl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 23.1 min, t_R (1*R*,2*R*): 33.5 min, 70.30:29.70 er.



| 2 | 33.518 | 29.703 |
|-------|--------|---------|
| Total | | 100.000 |

(1R,2R)-1,2-di(naphthalen-1-yl)ethane-1,2-diyl bis(2-methylpropanoate) (7e)



1,2-di(naphthalen-1-yl)ethane-1,2-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,2-di(naphthalen-1-yl)ethane-1,2-diol; **Chiral HPLC analysis** Chiralpak OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 220 nm, 30 °C) t_R (1*S*,2*S*): 23.9 min, t_R (1*R*,2*R*): 32.4 min, 6.41:93.59 er.



(1E,3S,4S,5E)-4-hydroxy-1,6-diphenylhexa-1,5-dien-3-yl isobutyrate (6f)

Chiral HPLC analysis Chiralpak IC (95:5 Hexane:IPA, flow rate 1 mL min⁻¹, 254 nm, 30 °C) t_R (3*S*,4*S*): 18.0 min, t_R (3*R*,4*R*): 22.7 min, 80.96:19.04 er.



| Detector A Channel 2 254nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 18.047 | 80.962 |
| 2 | 22.687 | 19.038 |
| Total | | 100.000 |

(1E,3R,4R,5E)-1,6-diphenylhexa-1,5-diene-3,4-diyl bis(2-methylpropanoate) (7f)



(1*E*,5*E*)-1,6-diphenylhexa-1,5-diene-3,4-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give (1*E*,5*E*)-1,6-diphenylhexa-1,5-diene-3,4-diol; Chiral HPLC analysis Chiralpak ID (85:15 Hexane:IPA, flow rate 1 mL min⁻¹, 254 nm, 30 °C) t_R (1*S*,2*S*): 13.0 min, t_R (1*R*,2*R*): 14.7 min, 35.00:65.00 er.



| Detector A Channel 2 254nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 12.982 | 35.003 |
| 2 | 14.714 | 64.997 |
| Total | | 100.000 |
(3S,4S)-1,6-diphenylhexa-1,5-diyne-3,4-diol (5g)



Chiral HPLC analysis Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 13.6 min, t_R (3*R*,4*R*): 19.1 min, 77.37:22.63 er.





| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 12.859 | 49.746 |
| 2 | 18.158 | 50.254 |
| Total | | 100.000 |



| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 13.613 | 77.366 |
| 2 | 19.068 | 22.634 |
| Total | | 100.000 |

(3S,4S)-4-hydroxy-1,6-diphenylhexa-1,5-diyn-3-yl isobutyrate (6g)



4-hydroxy-1,6-diphenylhexa-1,5-diyn-3-yl isobutyrate could not be separated by HPLC so it was hydrolysed to give 1,6-diphenylhexa-1,5-diyne-3,4-diol; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 12.6 min, t_R (3*R*,4*R*): 18.1 min, 88.34:11.66 er.



| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 12.629 | 88.343 |
| 2 | 18.116 | 11.657 |
| Total | | 100.000 |

(3R,4R)-1,6-diphenylhexa-1,5-diyne-3,4-diyl bis(2-methylpropanoate) (7g)



1,6-diphenylhexa-1,5-diyne-3,4-diyl bis(2-methylpropanoate) could not be separated by HPLC so it was hydrolysed to give 1,6-diphenylhexa-1,5-diyne-3,4-diol; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (3*S*,4*S*): 12.6 min, t_R (3*R*,4*R*): 18.1 min, 41.64:58.36 er.



<Peak Table>

| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 12.932 | 41.637 |
| 2 | 18.353 | 58.363 |
| Total | | 100.000 |

<u>General procedure for the Sequential Acylative Kinetic Resolution (±)-anti-1,3-diols 2 in flow-</u> <u>mode conditions</u>

A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin 8 (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping 1:1 THF:CHCl₃ mixture at 1 mL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of the appropriate diol (\pm) -2 (0.75 mmol, 1.0 equiv.) and (EtCO)₂O (0.79 mmol, 1.05 equiv.) in 1:1 THF:CHCl₃ mixture (7.5 mL total volume) and the second syringe with *i*-Pr₂NEt (0.79 mmol, 1.05 equiv.) in 1:1 THF:CHCl₃ mixture (7.5 mL total volume). Both solutions were injected at 100 μ L min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 200 μ L min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 200 µL min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in 1:1 THF:CHCl₃ mixture was then pumped at 200 μ L min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping 1:1 THF:CHCl₃ mixture at 200 µL min⁻¹ for 30 min. The mixture was concentrated to remove THF and later diluted with CH₂Cl₂. So, it was washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the crude products which were purified by column chromatography.

Hydrolysis of diesters and monoesters

In some instances it was difficult to find conditions to separate the enantiomers of diesters or monoesters by HPLC using a chiral support, and therefore these products were hydrolysed to the diol prior to HPLC analysis: LiOH•H₂O (3 equiv.) was added to a solution of the diester or monoester (1 equiv.) in MeOH (0.3 M) and allowed to stir at 50 °C until completion, based on TLC analysis. The mixture was diluted with EtOAc and washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the diol product.

Kinetic Resolution of (±)-(1*E*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol ((±)-2a)

According to the **General Procedure**, (\pm)-(1*E*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol (210 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture

(7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*E*,3*R*,5*R*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol (2a)

(63 mg, 30%) as a colourless solid with spectroscopic data in accordance with the literature.⁶ **mp** 140–141 °C {Lit.⁶ 149 °C}; $[\alpha]_D^{20}$ +25.0 (*c* 1.0 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (>99:1 er) +16.2 (*c* 0.50, CHCl₃)}; **Chiral HPLC analysis** Chiralpak IC (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*S*,5*S*): 12.3 min, t_R(3*R*,5*R*): 14.0 min, 3.19:96.81 er; ¹**H NMR** (500 MHz, CDCl₃) δ_{H} : 2.01 (t, *J* 5.4 Hz, 2H, C(2)H₂), 2.57 (d, *J* 3.9 Hz, 2H, OH), 4.72 (s, 2H, CHOH), 6.33 (dd, *J* 15.7, 6.2 Hz, 2H, PhCH=CH), 6.68 (d, *J* 15.7 Hz, 2H, PhCH=CH), 7.23 – 7.27 (m, 2H, ArH), 7.35 (t, *J* 7.3 Hz, 4H, ArH), 7.42 (d, *J* 7.5 Hz, 4H, ArH).

(1E,3S,5S,6E)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate (3a)



As an inseparable mixture of combined *anti* and *syn* diastereomers (80:20 dr_{anti:syn}) (45 mg, 18%) as a colourless oil with spectroscopic data in accordance with the literature.^{6,12} $[\alpha]_D^{20}$ –21.5 (*c* 1.0 in CHCl₃).

*Data for major diastereomer anti-***3a** in accordance with the literature:⁶ following hydrolysis to (1*E*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol and the *anti*-diastereomer was purified through crystallization from hot Toluene:Hexane 1:1: **Chiral HPLC analysis** Chiralpak IC (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3R,5*R*): 12.2 min, t_R(3*R*,5*R*): 14.0 min, 64.99:35.01 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.11 – 1.24 (m, 3H, CH₃), 1.87 – 2.12 (m, 2H, C(2)H₂), 2.43 (q, *J* 7.5 Hz, 2H, CH₂CH₃), 2.62 (d, *J* 3.8 Hz, 1H, OH), 4.28 – 4.43 (m, 1H, CHOH), 5.71 – 5.82 (m, 1H, CHOC(O)Et), 6.15 – 6.34 (m, 2H, PhCH=C*H*), 6.66 (ddd, *J* 16.0, 6.0, 1.1 Hz, 2H, PhCH=CH), 7.25 – 7.31 (m, 2H, ArH), 7.32 – 7.38 (m, 4H, ArH), 7.38 – 7.44 (m, 4H, ArH). *Data for minor diastereomer syn-***3a** in accordance with the literature:¹² ¹**H NMR** (400 MHz, CDCl₃) (*selected*) δ_{H} : 4.41 – 4.51 (m, 1H, CHOH), 5.59 – 5.73 (m, 1H, CHOC(O)Et). (**1E,3S,5S,6E)-1,7-diphenylhepta-1,6-diene-3,5-diyl dipropionate (4a)**



(99 mg, 34%) as a colourless solid with spectroscopic data in accordance with the literature.⁶ mp 51–52 °C {Lit.⁶ 56 °C}; $[\alpha]_D^{20}$ –5.0 (*c* 0.3 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (98:2 er) –44.1 (*c* 1.00, CHCl₃)};

Chiral HPLC analysis Chiralcel OJ-H (85:15 Hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 9.5 min, $t_R(3S,5S)$: 13.4 min, 4.85:95.15 er.; ¹H NMR (500 MHz, CDCl₃) δ_H : 1.16 (t, *J* 7.6 Hz, 6H, CH₃), 2.16 (dd, *J* 7.3, 6.6 Hz, 2H, CH₂), 2.37 (qd, *J* 7.5, 4.5 Hz, 4H, CH₂CH₃), 5.59 (td, *J* 7.4, 0.9 Hz, 2H, CHOC(O)Et), 6.18 (dd, *J* 15.9, 7.3 Hz, 2H, PhCH=CH), 6.66 (d, *J* 15.9 Hz 2H, PhCH=CH), 7.25 – 7.30 (m, 2H, ArH), 7.31 – 7.37 (m, 4H, ArH), 7.37 – 7.42 (m, 4H, ArH).

Kinetic Resolution of (±)-1,3-diphenylpropane-1,3-diol ((±)-2b)

According to the **General Procedure**, (\pm)-1,3-diphenylpropane-1,3-diol (171 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture (7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*R*,3*R*)-1,3-diphenylpropane-1,3-diol (2b)



(60 mg, 35%) as a colourless solid with spectroscopic data in accordance with the literature.⁶ **mp** 141–143 °C {Lit.⁶ 153°C}; $[\alpha]_D^{20}$ +75.0 (*c* 0.5 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (99:1 er) +115.9 (*c* 1.0, CHCl₃)}; **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 11.4 min, 100:0 er.; ¹H NMR (500 MHz, CDCl₃) $\delta_{\rm H}$: 2.21 (dd, *J* 6.2, 5.4 Hz, 2H, C(2)H₂), 5.01 (dd, *J* 6.3, 5.4 Hz, 2H, CHOH), 7.29 – 7.33 (m, 2H, ArH), 7.36 – 7.41 (m, 8H, ArH). (**1S,3S)-3-hydroxy-1,3-diphenylpropyl propionate (3b)**



As an inseparable mixture of combined *anti* and *syn* diastereomers (90:10 dr_{*anti*:*syn*) (24 mg, 10%) as a colourless oil. $[\alpha]_D^{20}$ –21.8 (*c* 0.45 in CHCl₃); following hydrolysis to 1,3-diphenylpropane-1,3-diol **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 11.4 min, t_R(3*S*,5*S*): 14.6 min, 33.73:66.27 er; **IR** (neat) v_{max} cm⁻¹: 3447 (OH), 1754 (C=O), 1495, 1456, 1350, 1277, 1188, 1063, 1022; **HRMS** (ESI⁺) C₁₈H₂₀O₃Na [M+Na]⁺ found 307.1299, requires 307.1305 (–1.9 ppm).}

*Data for major diastereomer anti-***3b**: ¹**H NMR** (500 MHz, CDCl₃) δ_H: 1.19 (t, *J* 7.6 Hz, 3H, CH₃), 2.17 (ddd, *J* 14.5, 9.9, 3.4 Hz, 1H, C(2)H₂), 2.29 (ddd, *J* 14.6, 10.3, 3.4 Hz, 1H, C(2)H₂), 2.43 (qd, *J* 7.5, 4.4 Hz, 2H, CH₂CH₃), 2.77 (d, *J* 3.6 Hz, 1H, OH), 4.74 (dt, *J* 9.9, 3.5 Hz, 1H, CHOH), 6.11 (dd, *J* 10.3, 3.4 Hz, 1H, CHOC(O)Et), 7.29 – 7.34 (m, 2H, ArH), 7.34 – 7.42 (m, 8H, ArH); ¹³C NMR $(126 \text{ MHz}, \text{CDCl}_3) \delta_C: 9.2 (CH_3), 27.8 (CH_2CH_3), 46.6 (C(2)H_2), 70.4 (CHOH), 73.0 (CHOC(O)Et), 125.8 (2 × ArCH), 126.4 (2 × ArCH), 127.7 (ArCH), 128.0 (ArCH), 128.6 (4 × ArCH), 140.4 (ArC), 143.7 (ArC), 174.0 (CHOC(O)Et).$

*Data for minor diastereomer syn-***3b** in accordance with the literature:¹² ¹**H NMR** (500 MHz, CDCl₃) (*selected*) δ_{H} : 4.65 (dd, *J* 8.4, 4.8 Hz, 1H, CHOH), 5.90 (dd, *J* 7.8, 6.5 Hz, 1H, CHOC(O)Et).

(1*S*,3*S*)-1,3-diphenylpropane-1,3-diyl dipropionate (4b)



(105 mg, 39%) as a colourless oil with spectroscopic data in accordance with the literature.⁶ $[\alpha]_D^{20}$ –89.4 (*c* 0.4 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (98:2 er) –48.9 (*c* 1.0, CHCl₃)}; following hydrolysis to 1,3diphenylpropane-1,3-diol **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C), t_R(3*R*,5*R*): 11.4 min, t_R(3*S*,5*S*): 14.5 min, 3.45:96.55 er; ¹**H NMR** (500 MHz, CDCl₃) δ_{H} : 1.14 (t, *J* 7.5 Hz, 6H, CH₃), 2.24 – 2.46 (m, 6H, CH₂CH₃ and C(2)H₂), 5.90 (dd, *J* 7.7, 6.5 Hz, 2H, CHOC(O)Et), 7.29 – 7.39 (m, 10H, ArH).

Kinetic Resolution of (±)-1,3-bis(2-methoxyphenyl)propane-1,3-diol ((±)-2c)

According to the **General Procedure**, (\pm)-1,3-bis(2-methoxyphenyl)propane-1,3-diol (216 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture (7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*R*,3*R*)-1,3-bis(2-methoxyphenyl)propane-1,3-diol (2c)



(85 mg, 39%) as a colourless solid with spectroscopic data in accordance with the literature.⁶ **mp** 106–107 °C {Lit.⁶ 117 °C}; $[\alpha]_D^{20}$ +84.3 (*c* 1.0 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (>99:1 er) +121.8 (*c* 1.0, CHCl₃)}; **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 16.9 min, t_R(3*S*,5*S*): 21.1 min, 97.50:2.50 er; ¹**H NMR** (400 MHz, CDCl₃) δ_H : 2.31 (dd, *J* 6.3, 5.3 Hz, 2H, C(2)H₂), 3.83 (s, 6H, OCH₃), 5.23 (t, *J* 5.8 Hz, 2H, CHOH), 6.89 (dd, *J* 8.2, 1.1 Hz, 2H, ArH), 7.00 (td, *J* 7.5, 1.1 Hz, 2H, ArH), 7.23 – 7.30 (m, 2H, ArH), 7.41 – 7.48 (m, 2H, ArH).

(15,35)-3-hydroxy-1,3-bis(2-methoxyphenyl)propyl propionate (3c)



As an inseparable mixture of combined *anti* and *syn* diastereomers (94:6 dr_{*anti:syn*}) (56 mg, 22%) as a colourless oil. $[\alpha]_D^{20}$ –146.3 (*c* 0.6 in CHCl₃); following hydrolysis to 1,3-bis(2-methoxyphenyl)propane-1,3-diol **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 16.7 min, t_R(3*S*,5*S*): 20.2 min, 14.77:85.23 er; **IR** (neat) v_{max} cm⁻¹: 3450 (OH), 2940; 1732 (C=O), 1490, 1462; 1287; 1242; 1186; 1049; 1028; **HRMS** (ESI⁺) C₂₀H₂₄O₅Na [M+Na]⁺ found 367.1502, requires 367.1516 (–3.8 ppm).

Data for major diastereomer anti-**3c**: ¹**H NMR** (400 MHz, CDCl₃) δ_H: 1.20 (t, *J* 7.6 Hz, 3H, CH₃), 2.11 – 2.35 (m, 2H, C(2)H₂), 2.41 (qd, *J* 7.6, 0.9 Hz, 2H, CH₂CH₃), 3.20 (d, *J* 5.8 Hz, 1H, OH), 3.85 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 5.01 (dd, *J* 9.4, 3.5 Hz, 1H, CHOH), 6.50 (dd, *J* 9.9, 3.1 Hz, 1H, CHOC(O)Et), 6.88 (ddd, *J* 8.2, 4.4, 1.1 Hz, 2H, ArH), 6.92 – 7.01 (m, 2H, ArH), 7.20 – 7.28 (m, 2H, ArH), 7.35 (dd, *J* 7.6, 1.8 Hz, 1H, ArH), 7.38 – 7.44 (m, 1H, ArH); ¹³C NMR (126 MHz, CDCl₃) δ_C: 9.3 (CH₃), 27.9 (CH₂CH₃), 43.1 (C(2)H₂), 55.3 (OCH₃), 55.5 (OCH₃), 66.8 (CHOH), 68.1 (CHOC(O)Et), 110.3 (ArCH), 110.6 (ArCH), 120.5 (ArCH), 120.8 (ArCH), 126.2 (ArCH), 126.8 (ArCH), 128.2 (ArCH), 128.7 (ArCH), 129.4 (ArC), 131.9 (ArC), 156.0 (ArC), 156.2 (ArC), 174.3 (CHOC(O)Et).

*Data for minor diastereomer syn-***3c** in accordance with the literature:¹² ¹**H NMR** (400 MHz, CDCl₃) (*selected*) δ_{H} : 4.91 – 4.99 (m, 1H, CHOH), 6.16 – 6.30 (m, 1H, CHOC(O)Et).

(15,35)-1,3-bis(2-methoxyphenyl)propane-1,3-diyl dipropionate (4c)



(75 mg, 25%) as a colorless oil with spectroscopic data in accordance with the literature.⁶ $[\alpha]_D^{20}$ –48.0 (*c* 1.0 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (>99:1 er) –22.8 (*c* 1.0, CHCl₃)}; following hydrolysis to 1,3-bis(2-methoxyphenyl)propane-1,3-diol **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 17.1 min, t_R(3*S*,5*S*): 20.4 min, 0.46:99.54 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.18 (t, *J* 7.6 Hz, 6H, CH₃), 2.27 (dd, *J* 7.3, 5.9 Hz, 2H, C(2)H₂), 2.39 (qd, *J* 7.5, 3.2 Hz, 4H, CH₂CH₃), 3.85 (s, 6H, OCH₃), 6.33 (dd, *J* 7.3, 5.9 Hz, 2H, CHOC(O)Et), 6.85 (dd, *J* 8.2, 1.1 Hz, 2H, ArH), 6.94 (td, *J* 7.5, 1.1 Hz, 2, ArH), 7.23 (ddd, *J* 8.2, 7.4, 1.8 Hz, 2H, ArH), 7.31 (dd, *J* 7.6, 1.8 Hz, 2H, ArH).

Kinetic Resolution of (±)-1,3-bis(4-fluorophenyl)propane-1,3-diol ((±)-2d)

According to the **General Procedure**, (\pm)-1,3-bis(4-fluorophenyl)propane-1,3-diol (198 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture (7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1R,3R)-1,3-bis(4-fluorophenyl)propane-1,3-diol (2d)



(65 mg, 32%) as a colourless solid with spectroscopic data in accordance with the literature.⁶ **mp** 115–117 °C {Lit.⁶ 121.5 °C}; $[\alpha]_D^{20}$ +50.6 (*c* 1.0 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (>99:1 er) +73.5 (*c* 1.0, CHCl₃)}; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 6.4 min, t_R(3*S*,5*S*): 7.3 min, 99.49:0.51 er; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 2.14 (dd, *J* 6.3, 5.3 Hz, 2H, C(2)H₂), 4.99 (t, *J* 5.8 Hz, 2H, CHOH), 7.02 – 7.11 (m, 4H, ArH), 7.31 – 7.39 (m, 4H, ArH); ¹⁹**F NMR** (377 MHz, CDCl₃) $\delta_{\rm F}$: –115.02.

(1*S*,3*S*)-1,3-bis(4-fluorophenyl)-3-hydroxypropyl propionate (3d)



As an inseparable mixture of combined *anti* and *syn* diastereomers (92:8 dr_{*anti:syn*}) (34 mg, 14%) as a colourless oil. $[\alpha]_D^{20}$ –5.9 (*c* 0.45 in CHCl₃); following hydrolysis to 1,3-bis(4-fluorophenyl)propane-1,3-diol **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 6.4 min, t_R(3*S*,5*S*): 7.3 min, 40.40:59.60 er; **IR** (neat) v_{max} cm⁻¹: 3447 (OH), 1732 (C=O), 1605; 1510, 1223; 1186; 1157; **HRMS** (ESI⁺) C₁₈H₁₈O₃F₂Na [M+Na]⁺ found 343.1105, requires 343.1116 (–3.2 ppm).

Data for major diastereomer anti-**3d**: ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.18 (t, *J* 7.6 Hz, 3H, CH₃), 2.11 (ddd, *J* 14.5, 9.8, 3.4 Hz, 1H, C(2)H₂), 2.24 (ddd, *J* 14.5, 10.2, 3.5 Hz, 1H, C(2)H₂), 2.42 (qd, *J* 7.5, 3.9 Hz, 2H, CH₂CH₃), 2.78 (bs, 1H, OH), 4.71 (dd, *J* 9.8, 3.5 Hz, 1H, CHOH), 6.06 (dd, *J* 10.3, 3.4 Hz, 1H, CHOC(O)Et), 6.99 – 7.11 (m, 4H, ArH), 7.31 – 7.42 (m, 4H, ArH); ¹³C NMR (126 MHz, CDCl₃) δ_{C} : 9.1 (CH₃), 27.8 (CH₂CH₃), 46.6 (C(2)H₂), 69.7 (CHOH), 72.3 (CHOC(O)Et), 115.3 (dd, ²*J*_{C-F} 16.9 Hz, 2 × ArCH), 115.5 (dd, ²*J*_{C-F} 16.9 Hz, 2 × ArCH), 127.4 (d, ³*J*_{C-F} 8.1 Hz, 2 × ArCH), 128.2 (d, ³*J*_{C-F} 8.2 Hz, 2 × ArCH), 136.1 (d, ⁴*J*_{C-F} 3.1 Hz, ArC), 139.3 (d, ⁴*J*_{C-F} 3.0 Hz, ArC), 161.3 (d, ¹*J*_{C-F} 246.2 Hz, ArC), 163.4 (d, ¹*J*_{C-F} 246.2 Hz ArC), 174.5 (CHOC(O)Et); ¹⁹F NMR (377 MHz, CDCl₃) δ_{F} : –113.90, –114.81. *Data for minor diastereomer syn-***3d** in accordance with the literature:¹² ¹**H** NMR (400 MHz, CDCl₃) (*selected*) δ_{H} : 4.60 (dd, *J* 8.5, 4.8 Hz, 1H, CHOH), 5.87 (t, *J* 7.1 Hz, 1H, CHOC(O)Et); ¹⁹**F** NMR (377 MHz, CDCl₃) δ_{F} : -113.79, -114.46.

(15,35)-1,3-bis(4-fluorophenyl)propane-1,3-diyl dipropionate (4d)



(113 mg, 40%) as a colourless oil with spectroscopic data in accordance with the literature.⁶ $[\alpha]_D^{20}$ -66.1 (*c* 0.6 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (99:1 er) -68.4 (*c* 1.0, CHCl₃)}; following hydrolysis to 1,3-bis(4fluorophenyl)propane-1,3-diol **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 6.5 min, t_R(3*S*,5*S*): 7.4 min, 6.72:93.28 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.13 (t, *J* 7.5 Hz, 6H, CH₃), 2.25 – 2.43 (m, 6H, CH₂CH₃ and C(2)H₂), 5.79 – 5.88 (m, 2H, CHOC(O)Et), 7.00 – 7.08 (m, 4H, ArH), 7.30 – 7.37 (m, 4H, ArH); ¹⁹**F NMR** (377 MHz, CDCl₃) δ_{F} : –113.71.

Kinetic Resolution of (±)-1,5-diphenylpent-4-ene-1,3-diol ((±)-2e)

According to the **General Procedure**, (\pm)-1,5-diphenylpent-4-ene-1,3-diol (191 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture (7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1*R*,3*R*,*E*)-1,5-diphenylpent-4-ene-1,3-diol (2e)



(60 mg, 32%) as a white solid with spectroscopic data in accordance with the literature.⁶ **mp** 106–107 °C {Lit.⁶ 106 °C}; $[\alpha]_D^{20}$ –13.5 (*c* 1.0 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (>99:1 er) –11.8 (*c* 1.0, CHCl₃)}; **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R(3*S*,5*S*): 10.5 min, t_R(3*R*,5*R*): 17.5 min, 1.91:98.09 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 2.06 (ddd, *J* 14.5, 7.7, 3.3 Hz, 1H, C(2)H₂), 2.17 (ddd, *J* 14.6, 8.5, 3.5 Hz, 1H, C(2)H₂), 4.55 – 4.69 (m, 1H, C(3)HOH), 5.12 (dd, *J* 8.5, 3.4 Hz, 1H, C(1)HOH), 6.34 (dd, *J* 15.9, 6.1 Hz, 1H, PhCH=CH), 6.66 (dd, *J* 15.9, 1.4 Hz, 1H, PhCH=CH), 7.32 – 7.45 (m, 10H, ArH).

(3R,5R,E)-5-hydroxy-1,5-diphenylpent-1-en-3-yl propionate (3e')



(21 mg, 9%) as a pale yellow oil. $[\alpha]_D^{20}$ –5.6 (*c* 0.25 in CHCl₃); following hydrolysis to 1,5diphenylpent-4-ene-1,3-diol **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*S*,5*S*): 10.4 min, t_R(3*R*,5*R*): 18.1 min, 40.75:59.25 er; **IR** (neat) v_{max} cm⁻¹: 3447 (OH), 3028, 1732 (C=O), 1494, 1450, 1273, 1186, 1080, 1066; ¹**H NMR** (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.21 (t, *J* 7.6 Hz, 3H, CH₃), 2.05 – 2.25 (m, 2H, C(2)H₂), 2.43 (q, *J* 7.5 Hz, 2H, CH₂CH₃), 4.66 – 4.82 (m, 1H, CHOH), 5.76 (qd, *J* 6.8, 1.1 Hz, 1H, CHOC(O)Et), 6.21 (dd, *J* 15.9, 7.0 Hz, 1H, PhCH=CH), 6.66 (d, *J* 15.9 Hz, 1H, PhCH=CH), 7.29 – 7.44 (m, 10H, ArH); ¹³C NMR (126 MHz, CDCl₃) $\delta_{\rm C}$: 9.2 (CH₃), 27.9 (CH₂CH₃), 44.6 (C(2)H₂), 70.3 (CHOH), 71.9 (CHOC(O)Et), 125.8 (2 × ArCH), 126.6 (2 × ArCH), 127.3 (ArCH), 127.7 (ArCH), 128.1 (PhCH=CH), 128.6 (4 × ArCH), 132.4 (PhCH=CH), 136.2 (ArC), 143.7 (ArC), 174.7 (CHOC(O)Et); **HRMS** (ESI⁺) C₂₀H₂₂O₃Na [M+Na]⁺ found 333.1457, requires 333.1461 (-1.2 ppm).

(1S,3S,E)-3-hydroxy-1,5-diphenylpent-4-en-1-yl propionate (3e'')



As an inseparable mixture of combined *anti* and *syn* diastereomers (89:11 dr_{*anti:syn*}) (14 mg, 6%) as a pale yellow oil. $[\alpha]_D^{20}$ +23.6 (*c* 0.25 in CHCl₃); following hydrolysis to 1,5-diphenylpent-4-ene-1,3-diol **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*S*,5*S*): 10.5 min, t_R(3*R*,5*R*): 18.4 min, 56.02:43.98 er; **IR** (neat) v_{max} cm⁻¹: 3429 (OH), 1754 (C=O), 1494, 1450, 1275, 1186, 1080, 1068; **HRMS** (ESI⁺) C₂₀H₂₂O₃Na [M+Na]⁺ found 333.1459, requires 333.1461 (-0.6 ppm).

Data for major diastereomer anti-**3e''**: ¹**H NMR** (500 MHz, CDCl₃) δ_{H} : 1.17 (t, *J* 7.5 Hz, 3H, CH₃), 2.05 (ddd, *J* 14.4, 9.5, 3.6 Hz, 1H, C(2)H₂), 2.43 (qd, *J* 7.6, 4.9 Hz, 2H, CH₂CH₃), 2.17 – 2.26 (m, 1H, OH), 4.30 – 4.39 (m, 1H, CHOH), 6.08 (dd, *J* 10.2, 3.6 Hz, 1H, CHOC(O)Et), 6.21 – 6.30 (m, 1H, PhCH=CH), 6.60 – 6.71 (m, 1H, PhCH=CH), 7.30 – 7.36 (m, 2H, ArH), 7.36 – 7.43 (m, 8H, ArH); ¹³C NMR (126 MHz, CDCl₃) δ_{C} : 9.1 (CH₃), 27.8 (CH₂CH₃), 44.7 (C(2)H₂), 68.9 (CHOH), 72.8 (CHOC(O)Et), 126.4 (2 × ArCH), 126.5 (2 × ArCH), 127.7 (ArCH), 128.1 (ArCH), 128.6 (4 × ArCH), 130.4 (PhCH=CH), 131.1 (PhCH=CH), 136.6 (ArC), 140.4 (ArC), 174.6 (CHOC(O)Et).

*Data for minor diastereomer syn-***3e''**: ¹**H NMR** (500 MHz, CDCl₃) (*selected*) δ_{H} : 4.23 – 4.32 (m, 1H, CHOH), 6.00 (dd, *J* 7.7, 6.5 Hz, 1H, CHOC(O)Et), 6.56 (d, *J* 16.0 Hz, 1H, PhC*H*=CH); ¹³C **NMR** (126 MHz, CDCl₃) (*selected*) δ_{C} :70.1 (*C*HOH), 73.6 (*C*HOC(O)Et).

(1S,3S,E)-1,5-diphenylpent-4-ene-1,3-diyl dipropionate (4e)



(100 mg, 36%) as a pale yellow oil with spectroscopic data in accordance with the literature.⁶ $[\alpha]_D^{20}$ -53.2 (*c* 1.0 in CHCl₃) {Lit.⁵ $[\alpha]_D^{25}$ (98:2 er) -53.12 (*c* 1.0, CHCl₃)}; following hydrolysis to 1,5diphenylpent-4-ene-1,3-diol **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*S*,5*S*): 10.5 min, t_R(3*R*,5*R*): 18.0 min, 94.91:5.09 er; ¹**H NMR** (400 MHz, CDCl₃) δ_{H} : 1.15 (dt, *J* 15.2, 7.5 Hz, 6H, CH₃), 2.17 – 2.44 (m, 6H, CH₂CH₃ and C(2)H₂), 5.58 (td, *J* 8.8, 4.7 Hz, 1H, C(1)HOC(O)Et), 5.90 (dd, *J* 9.7, 4.5 Hz, 1H, C(3)HOC(O)Et), 6.15 (dd, *J* 15.9, 7.3 Hz, 1H, PhCH=CH), 6.55 – 6.72 (m, 1H, PhCH=CH), 7.30 – 7.41 (m, 10H, ArH).

Kinetic Resolution of (±)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol ((±)-2f)

According to the **General Procedure**, (\pm)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol (177 mg, 0.75 mmol) and propionic anhydride (101 µL, 0.787 mmol, 1.05 equiv.), in 1:1 CHCl₃:THF mixture (7.5 mL total volume), and *i*-Pr₂NEt (138 µL, 0.787 mmol, 1.05 equiv.) in 1:1 CHCl₃:THF mixture (7.5 mL total volume) gave crude products that were purified by column chromatography (80:20 to 60:40 Hexane:EtOAc) to give:

(1R,3R)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol (2f)



(77 mg, 44%) as a pale yellow solid with spectroscopic data in accordance with the literature.⁶ **mp** 141–144 °C {Lit.⁶ 139 °C}; $[\alpha]_D^{20}$ +29.7 (*c* 0.5 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (97:3 er) +7.8 (*c* 0.5, CHCl₃)}; **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 11.0 min, t_R(3*S*,5*S*): 12.8 min, 99.73:0.27 er.; ¹**H NMR** (500 MHz, CDCl₃) δ_{H} : 2.00 – 2.16 (m, 2H, OH), 2.17 – 2.36 (m, 2H, C(2)H₂), 4.96 – 5.09 (m, 2H, C(1)*H*OH and C(3)*H*OH), 6.30 (dt, *J* 3.3, 0.8 Hz, 1H, ArH), 6.37 (dd, *J* 3.3, 1.8 Hz, 1H, ArH), 7.04 – 7.09 (m, 2H, ArH), 7.34 – 7.39 (m, 2H, ArH), 7.41 (dd, *J* 1.8, 0.9 Hz, 1H, ArH); ¹⁹**F NMR** (471 MHz, CDCl₃) δ_{F} : –115.04. (**1***R*,**3***R*)-**3-(4-fluorophenyl)-1-(furan-2-yl)-3-hydroxypropyl propionate (3f')**



As an inseparable mixture of combined *anti* and *syn* diastereomers (82:18 dr_{anti:syn}) (9 mg, 4%) as a pale yellow oil. $[\alpha]_D^{20}$ +24.5 (*c* 0.1 in CHCl₃); following hydrolysis to 1-(4-fluorophenyl)-3-(furan-2-

yl)propane-1,3-diol **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.1 min, $t_R(3S,5S)$: 12.8 min, 72.97:27.03 er; **IR** (neat) v_{max} cm⁻¹: 3439 (OH), 1732 (C=O), 1606, 1510, 1223, 1182, 1157, 1080, 1065, 1011; **HRMS** (C₁₆H₁₇O₄FNa [M+Na]⁺ found 315.0995, requires 315.1009 (-4.4 ppm).

Data for major diastereomer anti-**3f**': ¹**H NMR** (500 MHz, CDCl₃) $\delta_{\rm H}$: 1.16 (t, *J* 7.5 Hz, 3H, CH₃), 2.26 (ddd, *J* 14.4, 9.7, 3.6 Hz, 1H, C(2)H₂), 2.34 – 2.47 (m, 3H, CH₂CH₃ and C(2)H₂), 2.70 (d, *J* 4.9 Hz, 1H, OH), 4.75 (dt, *J* 9.2, 4.2 Hz, 1H, C(1)*H*OH), 6.04 (dd, *J* 10.3, 3.6 Hz, 1H, C(3)*H*OC(O)Et), 6.28 (dt, *J* 3.2, 0.8 Hz, 1H, ArH), 6.33 – 6.38 (m, 1H, ArH), 7.02 – 7.09 (m, 2H, ArH), 7.33 – 7.38 (m, 2H, ArH), 7.40 (dd, *J* 1.8, 0.8 Hz, 1H, ArH); ¹³C **NMR** (126 MHz, CDCl₃) $\delta_{\rm C}$: 9.1 (CH₃), 27.7 (CH₂CH₃), 42.6 (C(2)H₂), 64.0 (C(1)HOH), 72.0 (C(3)HOC(O)Et), 106.2 (ArCH), 110.3 (ArCH), 115.5 (d, ²*J*_{C-F} 21.5 Hz, 2 ×ArCH), 128.2 (d, ³*J*_{C-F} 8.2 Hz 2 ×ArCH), 136.2 (d, ⁴*J*_{C-F} 3.2 Hz, ArC), 142.2 (ArCH), 155.6 (ArC), 162.4 (d, ¹*J*_{C-F} 246.6 Hz, ArC), 174.4 (CHOC(O)Et); ¹⁹F **NMR** (471 MHz, CDCl₃) $\delta_{\rm F}$: –113.95

*Data for minor diastereomer syn-***3f**': ¹**H NMR** (500 MHz, CDCl₃) (*selected*) δ_{H} : 4.61 (dd, *J* 8.3, 5.4 Hz, 1H, C(1)*H*OH), 5.89 (t, *J* 7.2 Hz, 1H, C(3)*H*OC(O)Et); ¹³C NMR (*selected*) (126 MHz, CDCl₃) δ_{C} : 64.9 (*C*(1)HOH), 72.8 (*C*(3)HOC(O)Et); ¹⁹F NMR (471 MHz, CDCl₃) δ_{F} : -113.85.

(15,35)-1-(4-fluorophenyl)-3-(furan-2-yl)-3-hydroxypropyl propionate (3f")



As an inseparable mixture of combined *anti* and *syn* diastereomers (71:29 dr_{*anti*:*syn*}) (14 mg, 6%) as a pale yellow oil. $[\alpha]_D^{20}$ –15.7 (*c* 0.07 in CHCl₃); following hydrolysis to 1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 11.1 min, t_R(3*S*,5*S*): 12.8 min, 45.34:54.66 er; **IR** (neat) v_{max} cm⁻¹: 3447 (OH), 2984, 1732 (C=O), 1605, 1510, 1221, 1182, 1157, 1067, 1013; **HRMS** (ESI⁺) C₁₆H₁₇O₄FNa [M+Na]⁺ found 315.0997, requires 315.1009 (–3.8 ppm).

Data for major diastereomer anti-**3f**":¹**H NMR** (500 MHz, CDCl₃) $\delta_{\rm H}$: 1.18 (t, *J* 7.6 Hz, 3H, CH₃), 2.25 (ddd, *J* 14.4, 9.9, 3.8 Hz, 1H, C(2)H₂), 2.34 – 2.49 (m, 3H, CH₂CH₃ and C(2)H₂), 2.83 (d, *J* 3.7 Hz, 1H, OH), 4.69 (dt, *J* 9.8, 3.6 Hz, 1H, C(1)*H*OH), 6.15 (dd, *J* 10.1, 3.7 Hz, 1H, C(3)*H*OC(O)Et), 6.24 – 6.46 (m, 2H, ArH), 6.96 – 7.11 (m, 2H, ArH), 7.32 – 7.38 (m, 2H, ArH), 7.41 (dd, *J* 1.8, 0.9 Hz, 1H, ArH); ¹³C NMR (126 MHz, CDCl₃) $\delta_{\rm C}$: 9.1 (*C*H₃), 27.6 (*C*H₂CH₃), 42.5 (*C*(2)H₂), 65.9 (*C*(3)HOC(O)Et), 69.4 (*C*(1)HOH), 108.5 (Ar*C*H), 110.3 (Ar*C*H), 115.3 (d, ²*J*_{*C*-*F*} 21.4 Hz, 2 × Ar*C*H), 127.4 (d, ³*J*_{*C*-*F*} 8.1 Hz, 2 × Ar*C*H), 139.2 (d, ⁴*J*_{*C*-*F*} 3.0 Hz, Ar*C*), 142.7 (Ar*C*H), 152.2 (Ar*C*), 161.2 (d, ¹*J*_{*C*-*F*} 245.6 Hz, Ar*C*), 174.7 (CHOC(O)Et); ¹⁹**F NMR** (471 MHz, CDCl₃) $\delta_{\rm F}$: –114.90. Data for minor diastereomer syn-**3**f'': ¹H NMR (500 MHz, CDCl₃) (selected) $\delta_{\rm H}$: 4.64 (dd, J 8.6, 4.7 Hz, 1H, C(1)HOH), 6.04 (t, J 7.2 Hz, 1H, C(3)HOC(O)Et); ¹³C NMR (126 MHz, CDCl₃) (selected) $\delta_{\rm C}$: 45.0 (*C*(2)H₂), 66.5 (*C*(3)HOC(O)Et), 71.0 (*C*(1)HOH); ¹⁹F NMR (471 MHz, CDCl₃) $\delta_{\rm F}$: -114.65.

(15,3S)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diyl dipropionate (4f)



(82 mg, 32%) as a yellow oil with spectroscopic data in accordance with the literature.⁶ $[\alpha]_D^{20}$ –88.8 (*c* 1.2 in CHCl₃) {Lit.⁶ $[\alpha]_D^{25}$ (98:2 er) –135.3 (*c* 1.0, CHCl₃)}; following hydrolysis to 1,5-diphenylpent-4-ene-1,3-diol **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 11.1 min, t_R(3*S*,5*S*): 12.8 min, 7.98:92.02 er; ¹**H NMR** (500 MHz, CDCl₃) $\delta_{\rm H}$: 1.13 (q, *J* 7.5 Hz, 6H, CH₃), 2.27 – 2.42 (m, 4H, CH₂CH₃), 2.44 – 2.57 (m, 2H, C(2)H₂), 5.79 – 5.86 (m, 1H, C(1)*H*OC(O)Et), 5.93 – 6.03 (m, 1H, C(3)*H*OC(O)Et), 6.35 (d, *J* 1.4 Hz, 2H, ArH), 7.01 – 7.09 (m, 2H, ArH), 7.30 – 7.37 (m, 2H, ArH), 7.40 (t, *J* 1.3 Hz, 1H, ArH); ¹⁹**F NMR** (471 MHz, CDCl₃) $\delta_{\rm F}$: –113.79.

Procedure for the Kinetic Resolution of (±)-3a in flow-mode conditions

A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin 8 (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping 1:1 THF:CHCl₃ mixture at 1 mL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of (\pm) -(1E,6E)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate **3a** (0.3 mmol, 1.0 equiv.) and (EtCO)₂O (0.17 mmol, 0.55 equiv.) in 1:1 THF:CHCl₃ mixture (3.0 mL total volume) and the second syringe with *i*-Pr₂NEt (0.17 mmol, 0.55 equiv.) in 1:1 THF:CHCl₃ mixture (3.0 mL total volume). Both solutions were injected at 100 µL min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 200 µL min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 200 μ L min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in 1:1 THF:CHCl₃ mixture was then pumped at 200 µL min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping 1:1 THF:CHCl₃ mixture at 200 µL min⁻¹ for 30 min. The mixture was concentrated to remove THF and later diluted with CH₂Cl₂. So, it was washed sequentially with HCl (1 M), saturated NaHCO₃ and brine. The organic layer was dried (Na₂SO₄), filtered and concentrated to give the crude products. The corresponding monoester and diester were then isolated by flash chromatography (90:10 Hexane:EtOAc).

(1*E*,3*R*,5*R*,6*E*)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate (3a): Chiral HPLC analysis Chiralpak AD-H (85:15 Hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(3R,5R)$: 12.2 min, $t_R(3S,5S)$: 14.0 min, 80.11:19.89 er.



| Detector A Channel 2 254nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.302 | 80.112 |
| 2 | 14.542 | 19.888 |
| Total | | 100.000 |

(1*E*,3*S*,5*S*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diyl dipropionate (4a): Chiral HPLC analysis Chiralcel OJ-H (85:15 Hexane:IPA, flow rate 1.5 mL min⁻¹, 254 nm, 30 °C) $t_R(3R,5R)$: 9.5 min, $t_R(3S,5S)$: 13.3 min, 19.38:80.62 er.



| Detector A Channel 2 254nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 9.491 | 19.376 |
| 2 | 13.286 | 80.624 |
| Total | | 100.000 |

HPLC chromatograms for the Sequential Acylative Kinetic Resolution of (±)-*anti*-1,3-diols 2 (1*E*,3*R*,5*R*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol (2a)

Chiral HPLC analysis Chiralpak IC (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3S,5S)$: 12.3 min, $t_R(3R,5R)$: 14.0 min, 3.19:96.81 er.





(1E,3S,5S,6E)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate (3a)



(1*E*,6*E*)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate was hydrolysed to (1*E*,6*E*)-1,7diphenylhepta-1,6-diene-3,5-diol and the *anti*-diastereomer was purified through crystallization form hot Toluene:Hexane 1:1. Chiral HPLC analysis Chiralpak IC (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*S*,5*S*): 12.2 min, t_R(3*R*,5*R*): 14.0 min, 64.99:35.01 er.



<Peak Table>

| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 12.237 | 64.989 |
| 2 | 13.987 | 35.011 |
| Total | | 100.000 |

(1E,3S,5S,6E)-1,7-diphenylhepta-1,6-diene-3,5-diyl dipropionate (4a)



Chiral HPLC analysis Chiralcel OJ-H (85:15 Hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 9.5 min, $t_R(3S,5S)$: 13.4 min, 4.85:95.15 er.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 9.468 | 4.849 |
| 2 | 13.398 | 95.151 |
| Total | | 100.000 |

(1*R*,3*R*)-1,3-diphenylpropane-1,3-diol (2b)



Chiral HPLC analysis Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.4 min, 100:0 er. The additional peak at $t_R \approx 12.2$ min could be identified as the meso diastereomer.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.360 | 100.000 |
| Total | | 100.000 |

(1*S*,3*S*)-3-hydroxy-1,3-diphenylpropyl propionate (3b)



3-hydroxy-1,3-diphenylpropyl propionate was hydrolysed to give 1,3-diphenylpropane-1,3-diol; **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.4 min $t_R(3S,5S)$: 14.6 min, 33.73:66.27 er. The additional peak at $t_R \approx 12.2$ min could be identified as the meso diastereomer.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.395 | 33.733 |
| 2 | 14.609 | 66.267 |
| Total | | 100.000 |

(1*S*,3*S*)-1,3-diphenylpropane-1,3-diyl dipropionate (4b)



1,3-diphenylpropane-1,3-diyl dipropionate was hydrolysed to give 1,3-diphenylpropane-1,3-diol; **Chiral HPLC analysis** Chiralpak ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.4 min, $t_R(3S,5S)$: 14.5 min, 3.45:96.55 er. The additional peak at $t_R \approx 12.2$ min could be identified as the meso diastereomer.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.368 | 3.453 |
| 2 | 14.485 | 96.547 |
| Total | | 100.000 |

(1*R*,3*R*)-1,3-bis(2-methoxyphenyl)propane-1,3-diol (2c)



Chiral HPLC analysis Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 16.9 min, $t_R(3S,5S)$: 21.1 min, 97.50:2.50 er. The additional peak at $t_R \approx 18.25$ min could be identified as the meso diastereomer.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 16.944 | 97.496 |
| 2 | 21.148 | 2.504 |
| Total | | 100.000 |



3-hydroxy-1,3-bis(2-methoxyphenyl)propyl propionate was hydrolysed to give 1,3diphenylpropane-1,3-diol; **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 16.7 min, $t_R(3S,5S)$: 20.2 min, 14.77:85.23 er. The additional peak at $t_R \approx 18.25$ min could be identified as the meso diastereomer.



| E | Detect | or A Chann | el 1 211nm |
|---|--------|------------|------------|
| F | Peak# | Ret. Time | Area% |
| Γ | 1 | 16.726 | 14.774 |
| Γ | 2 | 20.230 | 85.226 |
| | Total | | 100.000 |

(15,3S)-1,3-bis(2-methoxyphenyl)propane-1,3-diyl dipropionate (4c)



1,3-bis(2-methoxyphenyl)propane-1,3-diyl dipropionate was hydrolysed to give 1,3-bis(2-methoxyphenyl)propane-1,3-diol; **Chiral HPLC analysis** Chiralpak ID (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 17.1 min, $t_R(3S,5S)$: 20.4 min, 0.46:99.54 er.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 16.916 | 50.110 |
| 2 | 20.960 | 49.890 |
| Total | | 100.000 |



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 17.085 | 0.460 |
| 2 | 20.441 | 99.540 |
| Total | | 100.000 |

(1*R*,3*R*)-1,3-bis(4-fluorophenyl)propane-1,3-diol (2d)



Chiral HPLC analysis Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 6.4 min, $t_R(3S,5S)$: 7.3 min, 99.49:0.51 er. The additional peak at $t_R \approx 7.9$ min could be identified as the meso diastereomer.



| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 6.389 | 99.492 |
| 2 | 7.308 | 0.508 |
| Total | | 100.000 |

(15,3S)-1,3-bis(4-fluorophenyl)-3-hydroxypropyl propionate (3d)



1,3-bis(4-fluorophenyl)-3-hydroxypropyl propionate was hydrolysed to give 1,3-bis(4-fluorophenyl)propane-1,3-diol; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 6.4 min, t_R(3*S*,5*S*): 7.3 min, 40.40:59.60 er. The additional peak at t_R \approx 7.9 min could be identified as the meso diastereomer.



| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 6.401 | 40.397 |
| 2 | 7.335 | 59.603 |
| Total | | 100.000 |

(15,3S)-1,3-bis(4-fluorophenyl)propane-1,3-diyl dipropionate (4d)



1,3-bis(4-fluorophenyl)propane-1,3-diyl dipropionate was hydrolysed to give 1,3-bis(4-fluorophenyl)propane-1,3-diol; **Chiral HPLC analysis** Chiralcel OJ-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R(3*R*,5*R*): 6.5 min, t_R(3*S*,5*S*): 7.4 min, 6.72:93.28 er. The additional peak at t_R \approx 7.9 min could be identified as the meso diastereomer.



<Peak Table>

| PDA C | h1 211nm | |
|-------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 6.494 | 6.719 |
| 2 | 7.410 | 93.281 |
| Total | | 100.000 |

(1*R*,3*R*,*E*)-1,5-diphenylpent-4-ene-1,3-diol (2e)



Chiral HPLC analysis Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(3S,5S)$: 10.5 min, $t_R(3R,5R)$: 17.5 min, 1.91:98.09 er.





| Detect | or A Channe | el 2 254nm |
|--------|-------------|------------|
| Peak# | Ret. Time | Area% |
| 1 | 10.502 | 50.290 |
| 2 | 17.943 | 49.710 |
| Total | | 100.000 |



| Detector A Channel 2 254nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 10.520 | 1.905 |
| 2 | 17.534 | 98.095 |
| Total | | 100.000 |

(3R,5R,E)-5-hydroxy-1,5-diphenylpent-1-en-3-yl propionate (3e')



5-hydroxy-1,5-diphenylpent-1-en-3-yl propionate was hydrolysed to give 1,5-diphenylpent-4-ene-1,3-diol; **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3S,5S)$: 10.4 min, $t_R(3R,5R)$: 18.1 min, 40.75:59.25 er.



<Peak Table>

| Detect | or A Channe | el 1 211nm |
|--------|-------------|------------|
| Peak# | Ret. Time | Area% |
| 1 | 10.500 | 50.099 |
| 2 | 17.942 | 49.901 |
| Total | | 100.000 |





| Detect | or A Chann | el 1 211nm |
|--------|------------|------------|
| Peak# | Ret. Time | Area% |
| 1 | 10.443 | 40.746 |
| 2 | 18.133 | 59.254 |
| Total | | 100.000 |

(1*S*,3*S*,*E*)-3-hydroxy-1,5-diphenylpent-4-en-1-yl propionate (3e'')



3-hydroxy-1,5-diphenylpent-4-en-1-yl propionate was hydrolysed to give 1,5-diphenylpent-4-ene-1,3-diol; **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3S,5S)$: 10.5 min, $t_R(3R,5R)$: 18.4 min, 56.02:43.98 er.



<Peak Table>

| Detect | or A Channe | el 1 211nm |
|--------|-------------|------------|
| Peak# | Ret. Time | Area% |
| 1 | 10.500 | 50.099 |
| 2 | 17.942 | 49.901 |
| Total | | 100.000 |



| Detect | or A Chann | el 1 211nm |
|--------|------------|------------|
| Peak# | Ret. Time | Area% |
| 1 | 10.538 | 56.020 |
| 2 | 18.374 | 43.980 |
| Total | | 100.000 |

(1S,3S,E)-1,5-diphenylpent-4-ene-1,3-diyl dipropionate (4e)



1,5-diphenylpent-4-ene-1,3-diyl dipropionate was hydrolysed to give 1,5-diphenylpent-4-ene-1,3-diol; **Chiral HPLC analysis** Chiralcel OD-H (80:20 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3S,5S)$: 10.5 min, $t_R(3R,5R)$: 18.0 min, 94.91:5.09 er.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 10.456 | 94.911 |
| 2 | 18.002 | 5.089 |
| Total | | 100.000 |

(1R,3R)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol (2f)



Chiral HPLC analysis Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.0 min, $t_R(3S,5S)$: 12.8 min, 99.73:0.27 er. The additional peaks at $t_R \approx 12.0$ min could be identified as the diastereomer.





| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.097 | 50.511 |
| 2 | 12.837 | 49.489 |
| Total | | 100.000 |



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.042 | 99.734 |
| 2 | 12.819 | 0.266 |
| Total | | 100.000 |

(1*R*,3*R*)-3-(4-fluorophenyl)-1-(furan-2-yl)-3-hydroxypropyl propionate (3f')



3-(4-fluorophenyl)-1-(furan-2-yl)-3-hydroxypropyl propionate was hydrolysed to give 1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol; **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.1 min, $t_R(3S,5S)$: 12.8 min, 72.97:27.03 er. The additional peaks at $t_R \approx 12.0$ min could be identified as the diastereomer.





| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.097 | 50.511 |
| 2 | 12.837 | 49.489 |
| Total | | 100.000 |
| | | |



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.065 | 72.974 |
| 2 | 12.831 | 27.026 |
| Total | | 100.000 |

(15,3S)-1-(4-fluorophenyl)-3-(furan-2-yl)-3-hydroxypropyl propionate (3f")



1-(4-fluorophenyl)-3-(furan-2-yl)-3-hydroxypropyl propionate was hydrolysed to give 1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol; **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.1 min, $t_R(3S,5S)$: 12.8 min, 45.34:54.66 er. The additional peaks at $t_R \approx 12.0$ min could be identified as the diastereomer.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.104 | 45.344 |
| 2 | 12.859 | 54.656 |
| Total | | 100.000 |
(15,3S)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diyl dipropionate (4f)



1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diyl dipropionate was hydrolysed to give 1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diol; **Chiral HPLC analysis** Chiralcel ID (90:10 Hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(3R,5R)$: 11.1 min, $t_R(3S,5S)$: 12.8 min, 7.98:92.02 er.



| Detector A Channel 1 211nm | | |
|----------------------------|-----------|---------|
| Peak# | Ret. Time | Area% |
| 1 | 11.109 | 7.976 |
| 2 | 12.826 | 92.024 |
| Total | | 100.000 |

NMR Spectra

(1*E*,3*R*,5*R*,6*E*)-1,7-diphenylhepta-1,6-diene-3,5-diol (2a)



(1E,3S,5S,6E)-5-hydroxy-1,7-diphenylhepta-1,6-dien-3-yl propionate (3a)





(1E,3S,5S,6E)-1,7-diphenylhepta-1,6-diene-3,5-diyl dipropionate (4a)

(1*R*,3*R*)-1,3-diphenylpropane-1,3-diol (2b)









(1*S*,3*S*)-1,3-diphenylpropane-1,3-diyl dipropionate (4b)







(15,35)-3-hydroxy-1,3-bis(2-methoxyphenyl)propyl propionate (3c)



(1*S*,3*S*)-1,3-bis(2-methoxyphenyl)propane-1,3-diyl dipropionate (4c)







(1*S*,3*S*)-1,3-bis(4-fluorophenyl)-3-hydroxypropyl propionate (3d)







(15,35)-1,3-bis(4-fluorophenyl)propane-1,3-diyl dipropionate (4d)

(1*R*,3*R*,*E*)-1,5-diphenylpent-4-ene-1,3-diol (2e)



(3R,5R,E)-5-hydroxy-1,5-diphenylpent-1-en-3-yl propionate (3e')





(1S,3S,E)-3-hydroxy-1,5-diphenylpent-4-en-1-yl propionate (3e'')





(1*S*,3*S*,*E*)-1,5-diphenylpent-4-ene-1,3-diyl dipropionate (4e)





(1R, 3R) - 1 - (4 - fluorophenyl) - 3 - (furan - 2 - yl) propane - 1, 3 - diol~(2f)









(15,35)-1-(4-fluorophenyl)-3-(furan-2-yl)propane-1,3-diyl dipropionate (4f)

(15,25)-1,2-Diphenylethane-1,2-diol (5a)



(15,25)-2-Hydroxy-1,2-diphenylethyl isobutyrate (6a)





(1*R*,2*R*)-1,2-Diphenylethane-1,2-diyl bis(2-methylpropanoate) (7a)

(1*S*,2*S*)-1,2-bis(4-chlorophenyl)ethane-1,2-diol (5b)





(1*S*,2*S*)-1,2-bis(4-chlorophenyl)-2-hydroxyethyl isobutyrate (6b)

(1R,2R)-1,2-bis(4-chlorophenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7b)









$(1S,\!2S)\mbox{-}2\mbox{-}hydroxy\mbox{-}1,\!2\mbox{-}bis(4\mbox{-}(trifluoromethyl)\mbox{phenyl})\mbox{ethyl}\mbox{ isobutyrate (6c)}$



(1*R*,2*R*)-1,2-bis(4-(trifluoromethyl)phenyl)ethane-1,2-diyl bis(2-methylpropanoate) (7c)



(1*S*,2*S*)-1,2-bis(4-methoxyphenyl)ethane-1,2-diol (5d)

(1*S*,2*S*)-2-hydroxy-1,2-bis(4-methoxyphenyl)ethyl isobutyrate (6d)













(1S,2S)-2-hydroxy-1,2-di(naphthalen-1-yl)ethyl isobutyrate (6e)

(1R,2R)-1,2-di(naphthalen-1-yl)ethane-1,2-diyl bis(2-methylpropanoate) (7e)



(1E,3S,4S,5E)-4-hydroxy-1,6-diphenylhexa-1,5-dien-3-yl isobutyrate (6f)



(1E,3R,4R,5E)-1,6-diphenylhexa-1,5-diene-3,4-diyl bis(2-methylpropanoate) (7f)



(3S,4S)-1,6-diphenylhexa-1,5-diyne-3,4-diol (5g)



(3S,4S)-4-hydroxy-1,6-diphenylhexa-1,5-diyn-3-yl isobutyrate (6g)



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(3R,4R)-1,6-diphenylhexa-1,5-diyne-3,4-diyl bis(2-methylpropanoate) (7g)

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