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Electronic Supplementary Information

Cation-Chloride Cotransport Mediated by An Ion Pair

Transporter

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1. General methods

Reagents for synthesis and analysis were purchased from J&K or Sigma-Aldrich. A Mini-Extruder used for vesicle preparation, egg yolk phosphatidylcholine (EYPC). ¹H and ¹³C NMR spectra were recorded on Bruker 400 or 500 MHz NMR spectrometer. Chemical shifts are reported in ppm and referenced to tetramethylsilane (TMS) or the residual solvent resonance. X-ray crystallography were performed at the Analytical Laboratory of the Institute. All solvents were dried according to standard procedures prior to use. All other major chemicals were obtained from commercial sources and used without further purification.

2. ¹H NMR titration and data analysis

2.1 ¹H NMR titration and data analysis for binding of 1 with cations

The stock solution of $\mathbf{1}$ (1.05 × 10⁻² M) in deuterated acetonitrile was prepared, then an initial spectrum was recorded and additional spectra were obtained after aliquots of salt (LiClO₄, NaClO₄, KPF₆, Mg(ClO₄)₂, Ca(ClO₄)₂·4H₂O) solution (salt solutions were prepared by using host stock solution as solvent to avoid dilution). As the proton *a*, *b*, *e* are the most sensitive protons towards cation binding, these three protons were applied to the data fitting program to obtain the cation binding constants. The binding constants were calculated by *Bindfit v0.5* program^[1] using a 1:1 binding stoichiometry for [$\mathbf{1}$ +M²⁺] complex.



Figure S1. (a) A schematic binding mode of $[1 \cdot Li^+]$ complex, (b)The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of 1 (1.05 × 10⁻² M) with increasing equivalents of LiClO₄ in CD₃CN.



Figure S2. (a) A schematic binding mode of $[1 \cdot Na^+]$ complex, (b)The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of $1 (1.05 \times 10^{-2} \text{ M})$ with increasing equivalents of NaClO₄ in CD₃CN.



Figure S3. (a) A schematic binding mode of $[1 \cdot K^+]$ complex, (b)The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of **1** (1.05 × 10⁻² M) with increasing equivalents of KPF₆ in CD₃CN.



Figure S4. (a) A schematic binding mode of $[1 \cdot Mg^{2+}]$ complex, (b)The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of $1 (1.05 \times 10^{-2} \text{ M})$ with increasing equivalents of Mg(ClO₄)₂ in CD₃CN.



Figure S5. (a) A schematic binding mode of $[1 \cdot Ca^{2+}]$ complex, (b)The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of **1** (1.05 × 10⁻² M) with increasing equivalents of Ca(ClO₄)₂ in CD₃CN.

Table S1. Association constants for 1 binding alkaline earth metal cation.^[a]

Trai	nsporter	Li ⁺	Na ⁺	\mathbf{K}^+	Mg^{2+}	Ca ²⁺
1	<i>K</i> (M ⁻¹)	21	20	32	65	9383

[a] association constants were determined by ¹H NMR titration in CD₃CN, and fitted according to 1:1 binding model of *Bindfit v0.5* program.

2.2 ¹H NMR titration and data analysis for binding of 1 with anions

The stock solution of **1** $(1.05 \times 10^{-2} \text{ M})$ in deuterated acetonitrile was prepared, then an initial spectrum was recorded and additional spectra were obtained after aliquots of salt (TBACl, TBABr, TBAI, TBAClO₄) solution (salt solutions were prepared by using host stock solution as solvent to avoid dilution).



Figure S6. ¹H NMR titration of **1** (1.05×10^{-2} M) with increasing equivalents of TBACl in CD₃CN.



Figure S7. ¹H NMR titration of 1 (1.05×10^{-2} M) with increasing equivalents of TBABr in CD₃CN.



Figure S8. ¹H NMR titration of **1** (1.05×10^{-2} M) with increasing equivalents of TBAI in CD₃CN.



Figure S9. ¹H NMR titration of 1 (1.05×10^{-2} M) with increasing equivalents of TBAClO₄ in CD₃CN.

2.3 ¹H NMR titration and data analysis for binding of 1 with ion-pair

Based on calculation, when 2.0 eq. M^+/M^{2+} was added into the stock solution of **1**. The titration method is similar to above. An initial spectrum was recorded and additional spectra were obtained after aliquots of salt (TBAX, X = Cl⁻, Br⁻, I⁻).



Figure S10. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and LiClO₄ (2.10×10^{-2} M) with increasing equivalents of TBAI in CD₃CN.



Figure S11. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and LiClO₄ (2.10×10^{-2} M) with increasing equivalents of TBACl in CD₃CN.



Figure S12. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and LiClO₄ (2.10×10^{-2} M) with increasing equivalents of TBABr in CD₃CN.



Figure S13. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and NaClO₄ (2.10×10^{-2} M) with increasing equivalents of TBAI in CD₃CN.



Figure S14. ¹H NMR titration of a mixture of 1 (1.05×10^{-2} M) and NaClO₄ (2.10×10^{-2} M) with increasing equivalents of TBACl in CD₃CN.



Figure S15. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and NaClO₄ (2.10×10^{-2} M) with increasing equivalents of TBABr in CD₃CN.



Figure S16. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and KPF₆ (2.10×10^{-2} M) with increasing equivalents of TBAI in CD₃CN.



Figure S17. ¹H NMR titration of a mixture of 1 (1.05×10^{-2} M) and KPF₆ (2.10×10^{-2} M) with increasing equivalents of TBACl in CD₃CN.



Figure S18. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and KPF₆ (2.10×10^{-2} M) with increasing equivalents of TBABr in CD₃CN.



Figure S19. ¹H NMR titration of a mixture of 1 (1.05×10^{-2} M) and Mg(ClO₄)₂ (2.10 $\times 10^{-2}$ M) with increasing equivalents of TBAI in CD₃CN.



Figure S20. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and Mg(ClO₄)₂ (2.10 $\times 10^{-2}$ M) with increasing equivalents of TBAC1 in CD₃CN.



Figure S21. ¹H NMR titration of a mixture of 1 (1.05×10^{-2} M) and Mg(ClO₄)₂ (2.10 $\times 10^{-2}$ M) with increasing equivalents of TBABr in CD₃CN.



Figure S22. (a) A schematic binding mode of $[1 \cdot Ca^{2+} \cdot 2Br^{-}]$ complex, (b) The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of a mixture of **1** $(1.05 \times 10^{-2} \text{ M})$ and Ca(ClO₄)₂ $(2.10 \times 10^{-2} \text{ M})$ with increasing equivalents of Bu₄NBr in CD₃CN.



Figure S23. (a) A schematic binding mode of $[1 \cdot Ca^{2+} \cdot 2I^{-}]$ complex, (b) The fitting of the NMR titration data by *Bindfit v0.5* program, (c) ¹H NMR titration of a mixture of 1 $(1.05 \times 10^{-2} \text{ M})$ and Ca(ClO₄)₂ $(2.10 \times 10^{-2} \text{ M})$ with increasing equivalents of Bu₄NI in CD₃CN.



Figure S24. ¹H NMR titration of a mixture of **1** (1.05×10^{-2} M) and Ca(ClO₄)₂ (2.10×10^{-2} M) with increasing equivalents of Bu₄NCl in CD₃CN.

Table S2. Accumulative association constants determined by stepwise titrations. ^[a]



[a] accumulative association constants were determined by ¹H NMR titration in CD₃CN (298K, 400 MHz), and fitted according to 1:2 binding model of *Bindfit v0.5* program.

3 Ion transport

3.1 General preparation for MCl ⊂ EYPC-LUVs^[2-5]

The solution was produced during the EYPC (50 mg) was dissolved in the solution of MeOH/CHCl₃ (4 mL, v/v =1:1). The thin lipid film was obtained by evaporating this solution under reduced pressure on a rotary evaporator (40 °C) and then dried in vacuo for 48h. After hydration with 2.0 mL buffers (10 mM phosphate buffer, 500 mM NaCl or 500 mM KCl or 250 mM NaCl and 250 mM KCl or 166 mM CaCl₂, pH = 7.2) for 20 mins under the room temperature, the resulting suspension was subjected to 9 freeze-thaw cycles (with liquid N₂, 40 °C water bath) and 23 times extruded through a polycarbonate membrane (pore size 100 nm). Nonencapsulated salts were removed by dialysis.

3.2 General preparation for CF \subset EYPC-LUVs^{[2-4]}

The solution was produced during the EYPC (25 mg) was dissolved in the solution of MeOH/CHCl₃ (2 mL, v/v =1:1). The thin lipid film was obtained by evaporating this solution under reduced pressure on a rotary evaporator (40 °C) and then dried in vacuo for 48h. After hydration with 1.0 mL buffers (10 mM phosphate buffer, 100 mM NaNO₃, 50.0 mM CF, pH = 7.4) for 20 mins under the room temperature, the resulting suspension was subjected to 5 freeze-thaw cycles (with liquid N₂, 50 °C water bath) and 21 times extruded through a polycarbonate membrane (pore size 100 nm). Extravesicular components were removed by size exclusion chromatography (Sephadex G-50). The corresponding buffer as mobile phase.

3.3 ISE assay^[5]

A series of vesicles were suspended in an external solution (500 Mm NaNO₃ or 500 Mm KNO₃ or166 mM Na₂SO₄, buffered to pH 7.2 with 10 mM phosphate salts). The lipid concentration for each sample was 1 mM. An aliquot of the DMF solution of

a symporter, and the ion release from vesicles was monitored using an ion-selective electrode. The initial reading was considered as 0% release of an ion and the reading after the addition of 10% Triton X-100 to the lipid solution at 600 s was considered as 100% release. Transport experiments were performed on a ISE meter, and the chloride or potassium ion efflux was measured using a chloride or potassium ion selective electrode.

3.4 CF fluorescent assay^[2-4]

CF \subset EYPC-LUVs (50 µL) were added to gently stirred buffer (1950 µL, 100 mM NaNO₃, 50.0 mM CF, pH = 7.4) in a fluorescence cuvette (t = 0 s) at 25 °C. The timedependent change in fluorescence intensity (*I*_t) was monitored at excitation wavelengths simultaneously ($\lambda_{ex} = 369$ nm, $\lambda_{em} = 505$ nm), during addition of acetone or carrier **1** in acetone (25 µL) at t = 50 s (*I*₀), and triton X-100 (25 µL, 10% in water) at t = 300 s (*I*_∞). The fluorescence intensity *I*_t was normalized to fractional intensity *I*_f according to equation:

$$I_{\rm f} = (I_{\rm t} - I_0) / (I_{\infty} - I_0) \qquad (1)$$

where I_0 is I_t at t = 50 s before addition of carrier, I_∞ is I_t at t = 300 s after addition of triton X-100.

Ion transport



Figure S25. Chloride efflux facilitated by **1**, **2** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM NaCl in 10 mM sodium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ in 10 mM sodium phosphate buffer at pH = 7.2). Triton-100 was added to lyse the vesicles after 600 s, and chloride efflux was measured using a chloride selective electrode



Figure S26. (A) Chloride efflux facilitated by **1** (in varied concentration) from EYPC vesicles (500 mM in 10 mM sodium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ in 10 mM sodium phosphate buffer at pH = 7.2). Triton-100 was added to lyse the vesicles after 600 s, and chloride efflux was measured using a chloride selective electrode (B) Hill plot of the normalized intensities at 500 s. The EC₅₀ is 119 μ M.



Figure S27. Chloride efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM NaCl in 10 mM sodium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ or 166 mM Na₂SO₄ in 10 mM phosphate buffer at pH = 7.2).



Figure S28. Chloride efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM KCl in 10 mM potassium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ or 166 mM Na₂SO₄ in 10 mM sodium phosphate buffer at pH = 7.2).



Figure S29. Chloride efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM KCl, 500 mM NaCl or 166 mM CaCl₂ in 10 mM sodium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ in 10 mM sodium phosphate buffer at pH = 7.2). Triton X-100 was added to lyse the vesicles after 600 s, and chloride efflux was measured using a chloride selective electrode



Figure S30. Chloride or Potassium efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM KCl in 10 mM potassium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ in 10 mM sodium phosphate buffer at pH = 7.2). Triton X-100 was added to lyse the vesicles after 600 s, and chloride efflux was measured using a chloride or Potassium selective electrode



Figure S31. Chloride or Potassium efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (250 mM KCl and 250 mM NaCl in 5 mM sodium phosphate and 5mM potassium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ in 10 mM sodium phosphate buffer at pH = 7.2). Triton X-100 was added to lyse the vesicles after 1200 s, and chloride efflux was measured using a chloride or Potassium selective electrode



Figure S32. Chloride or Calcium efflux facilitated by **1** and solvent (10 mol % of each compound to lipid) from EYPC vesicles (500 mM KCl or 166 mM CaCl₂ in 10 mM sodium phosphate buffer at pH = 7.2) to a phosphate buffer (500 mM NaNO₃ or 166mM Na₂SO₄ in 10 mM sodium phosphate buffer at pH = 7.2). Triton X-100 was added to lyse the vesicles after 600 s, and chloride efflux was measured using a chloride or Calcium selective electrode

CF release experiment



Figure S33. Normalized fluorescence intensity of CF (10 mM HEPES, 100 mM NaNO₃, pH = 7.4) \subset EYPC-LUVs with the addition of the transporter **1** (70 μ M) or acetone, (A) the extravesicular salt is CaCl₂. (B) the extravesicular salt is NaCl₂ $\lambda_{ex} = 492$ nm, $\lambda_{em} = 517$ nm.

4. X-Ray diffraction data

CCDC number	2103643
Empirical formula	C31 H53 Ca Cl3 I2 N8 O12
Formula weight	1130.04
Temperature	173.15 K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	P 1 21/n 1
Unit cell dimensions	$a = 15.2822(5) \text{ Å} \alpha = 90^{\circ}$
	b = 20.3116(5) Å β = 117.084(5)°.
	$c = 16.7995(6) \text{ Å} \gamma = 90^{\circ}.$
Volume	4642.8(3) Å ³
Z	4
Density (calculated)	1.617 Mg/m ³
Absorption coefficient	1.699 m ⁻¹
F(000)	2272
Crystal size	0.338 x 0.217 x 0.056 mm ³
Theta range for data collection	1.691 to 27.538°.
Index ranges	-19<=h<=17, -26<=k<=26, -20<=l<=21
Reflections collected	10593
Independent reflections	10593 [R(int) = ?]
Completeness to theta = 25.242°	99.7 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.00000 and 0.85477
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	10593 / 0 / 521
Goodness-of-fit on F^2	1.058
Final R indices [I>2sigma(I)]	R1 = 0.0338, $wR2 = 0.0808$
R indices (all data)	R1 = 0.0383, $wR2 = 0.0828$
Extinction coefficient	n/a
Largest diff. peak and hole	1.053 and -0.660 e.Å ⁻³

Table S3. Crystal data and structure refinement for $[1 \cdot Ca^{2+} \cdot 2I^{-} \cdot 4H_2O \cdot CHCl_3]$

5. DFT optimization

For all free host molecules, free guest anions, and complexes, geometrical optimizations and energy calculations were carried out with the Gaussian16 suite of program using the X3LYP/6-31 G+(d) functional. The binding energy was calculated with BSSE correction.



Figure S34. The DFT optimized structure of $[1 \cdot Na^+ \cdot Cl^-]$ complex.



Figure S35. The DFT optimized structure of $[1 \cdot K^+ \cdot Cl^-]$ complex.

Cartesian Coordinates (Angstrom) of the Optimized Complexes

 $[\mathbf{1} \cdot \mathbf{Ca}^{2+} \cdot 2\mathbf{Br}^{-}]$

Atom	Х	Y	Z
Br	-0.20560000	0.19795100	1.25544500
Br	-2.98484300	-3.58792700	-1.87583800
Ca	-1.58905800	-1.81369000	-0.13375200
0	4.83300100	2.36820200	-0.77899400
0	3.35505300	-1.97352000	-0.55257000
0	0.52429200	-3.18095600	-0.58075200
0	-0.91833300	-3.44460900	1.69066100
0	-3.20319100	-1.95825300	1.86753900
0	-3.68671600	-0.36353800	-0.30150000
0	-3.13870200	1.98261000	-1.92789700
0	0.44713200	3.93900600	0.08448800
0	-1.13925600	-1.05068900	-2.40433100
Н	-1.64227800	-1.74293500	-2.88410500
Ν	6.04865200	0.84615500	0.33926000
Ν	5.26619700	-1.41203700	0.48554300
Ν	4.01472300	0.21670700	-0.72234100
Ν	-3.40592500	2.88383200	0.18175700
Ν	-1.50533300	3.95705000	1.14248200
Ν	-1.34244000	2.94907100	-1.00849600
С	4.95988700	1.08760500	-0.36893100
С	6.16296900	-0.43810500	0.75107100
С	4.23852500	-1.01271100	-0.25114600
С	2.24149200	-1.59754800	-1.38802000
Н	2.61998200	-1.19647700	-2.33425200
Н	1.65083400	-0.82275100	-0.89072200
С	1.41353800	-2.83988900	-1.64872700
Н	0.76272100	-2.65496400	-2.50414700
Н	2.06289700	-3.69615200	-1.87345200
С	1.11729700	-3.81582000	0.55227800
Н	1.63596100	-3.07821800	1.17413800
Н	1.83864200	-4.57554700	0.22436800
С	-0.00145900	-4.47557800	1.33222200
Н	-0.50927900	-5.23632000	0.72411500
Н	0.40672700	-4.94692300	2.23737200
С	-1.91603600	-3.79289700	2.64141300
Н	-1.45412700	-4.25300500	3.52637700
Н	-2.63293100	-4.50154900	2.20297900
С	-2.60683100	-2.50552000	3.04134600

Н	-3.37998800	-2.71206700	3.79548300
Н	-1.87424600	-1.79920500	3.45293100
С	-3.78709100	-0.66584700	2.06184100
Н	-2.99071700	0.06922100	2.23460500
Н	-4.46971200	-0.68676100	2.92343800
С	-4.57294400	-0.32891800	0.81416000
Н	-5.37605400	-1.06341200	0.66061100
Н	-5.00757100	0.67123600	0.92890200
С	-4.31112500	-0.09849700	-1.55931900
Н	-5.29759800	-0.58025200	-1.59911900
Н	-3.67834900	-0.57245300	-2.30908800
С	-4.44007300	1.39101800	-1.84146900
Н	-4.89799400	1.52744500	-2.82643400
Н	-5.03840700	1.91148400	-1.09290000
С	-2.62770100	2.62178400	-0.85962200
С	-2.79265000	3.58153900	1.16834600
С	-0.84907200	3.58212000	0.05313800
С	1.37673400	3.58783000	-0.88448500
С	1.22327100	3.88456200	-2.23908600
Н	0.29515000	4.30657600	-2.60176500
С	2.28599800	3.62311200	-3.10383700
Н	2.18087200	3.85271400	-4.16010300
С	3.48537000	3.08554100	-2.63323200
Н	4.32152100	2.89217700	-3.29632900
С	3.60317700	2.80321400	-1.27733600
С	2.56024500	3.04781400	-0.39105500
Н	2.65542400	2.83264900	0.66631900
С	-0.76765900	0.04690700	-3.24569700
Н	-0.02061300	-0.26714300	-3.98483800
Н	-0.34894800	0.81691800	-2.59749700
Н	-1.64015100	0.46436200	-3.75740200
Ν	-3.53354900	3.93502900	2.25314900
Ν	7.25611400	-0.76619900	1.48729800
С	-4.93997300	3.60378200	2.36957900
Н	-5.33245500	3.31461100	1.39696400
Н	-5.09948800	2.78087200	3.08244300
Н	-5.49111900	4.47790700	2.73656700
С	-2.93095400	4.57987100	3.40878200
Н	-3.43030300	5.53611400	3.61052500
Н	-3.03677900	3.94186400	4.29647800
Н	-1.87554500	4.75332500	3.21253500
С	7.46750300	-2.11208900	1.98982700
Н	7.50674700	-2.10528300	3.08730600
Н	6.65312700	-2.75309800	1.66005900

Η	8.42113400	-2.50854600	1.61700000
С	8.27103700	0.21264700	1.83714000
Н	8.35028700	0.30095500	2.92878500
Η	9.24847800	-0.10159700	1.44804900
Η	8.00465300	1.17834900	1.41379100

 $[\mathbf{2} \cdot \mathbf{Ca}^{2+} \cdot \mathbf{2Br}^{-}]$

Atom	Х	Y	Z
Br	0.03786600	0.27792200	-1.20205800
Br	3.30364700	-3.07898300	1.93001100
Ca	1.71011600	-1.54625100	0.13016400
Cl	-7.44359800	-1.80807800	-1.77133400
Cl	3.21380000	4.06618200	-2.88137700
0	-5.15323700	1.89122000	0.39493600
0	-3.19196900	-2.25749200	0.53967900
0	-0.23323400	-3.12327200	0.65111900
0	1.22138000	-3.30337700	-1.61916900
0	3.29345700	-1.54786900	-1.87617700
0	3.60339700	0.16370600	0.22459700
0	2.76635400	2.47748700	1.75463600
0	-0.95653300	3.93258500	-0.43210300
0	1.18748000	-0.78101600	2.39860400
Н	1.78070800	-1.40729700	2.86977400
Ν	-6.17955400	0.14552900	-0.58485000
Ν	-5.14519200	-2.00103500	-0.54663200
Ν	-4.09661300	-0.14979800	0.52724200
Ν	2.95955300	3.18748900	-0.43306500
Ν	1.00088200	4.00840000	-1.49003900
Ν	0.89587600	3.20496100	0.75288700
С	-5.12401600	0.58291300	0.10686100
С	-6.10501000	-1.14857900	-0.86459100
С	-4.16195500	-1.43137500	0.16528900
С	-2.12594700	-1.70218400	1.34681600
Н	-2.55801400	-1.28131700	2.26058000
Н	-1.62501600	-0.90617200	0.78725300
С	-1.16813800	-2.82520500	1.69050200
Н	-0.55558500	-2.51143600	2.53655000
Н	-1.71805400	-3.73417800	1.96635400
С	-0.71959400	-3.93420700	-0.41972000
Н	-1.37180100	-3.35028700	-1.07819700
Н	-1.28973100	-4.78300500	-0.01981800
С	0.48650700	-4.44580900	-1.18004500
Н	1.11867900	-5.07347400	-0.53768500

Н	0.15237300	-5.03360000	-2.04607200
С	2.25093500	-3.55964900	-2.56853000
Н	1.84527800	-4.11023600	-3.42856700
Н	3.05360000	-4.15367200	-2.10940500
С	2.77268100	-2.21346900	-3.02524500
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Н	1.95899400	-1.62235300	-3.46550800
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Η	-2.73264200	3.75918900	0.13484100
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 $[\mathbf{1} \cdot \mathbf{K}^+ \cdot \mathbf{Cl}^-]$

Н	3.30919500	5.17864500	2.34195000
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Н	4.39612300	-5.15449600	3.73610600
Κ	1.57721500	1.89869100	-0.21682500

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