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Supporting Information for

Tetradentate halogen bonding macrocyclic anion receptor inspired by the "Texas-sized" molecular box

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1.General information

All solvents and chemicals used were purchased from Aldrich, TCI, and Acros and used without further purification. TLC analyses were carried out using Sorbent Technologies silica gel (200 mm) sheets. Column chromatography was performed on Sorbent silica gel 60 (40–63 mm). NMR spectra were recorded on a Agilent MR 400 instrument. The NMR spectra were referenced to solvent residue peaks and the spectroscopic solvents were purchased from Cambridge Isotope Laboratories and Aldrich. Electrospray ionization (ESI) mass spectra were recorded on a VG AutoSpec apparatus. X-ray crystallographic analyses were carried out on either a Rigaku AFC12 diffractometer equipped with a Saturn 724+ CCD or an Agilent Technologies SuperNova Dual Source diffractometer using a μ -focus Cu K α radiation source ($\lambda = 1.5418$ Å) with collimating mirror monochromators. A graphite monochromator and MoK α radiation were used. Further details of the structures and their refinement is given in a later section.

Note: Many of the intermediates and products contain azide or triazole moieties, or both. Due to their potential thermal instability all manipulations involving such species were carried out at or below room temperature.

2.Synthesis and characterization

2.1 Synthesis of 1



Precursor **1** was synthesized using a modification of a literature report.^{S1} Briefly, 2,6dibromopyridine-4-carboxylic acid (5.6 g, 20 mmol), EDC·HCl (7.67 g, 40 mmol) and dimethylaminopyridine (DMAP) (0.48 g, 4 mmol) were dissolved in dry dichloromethane (DCM) (70 ml). *tert*-Butyl alcohol (3.80 ml, 40 mmol) was then added to the mixture. The reaction was stirred at rt for 24 h and then washed with water and brine. The organic layer was separated, dried over MgSO₄ and evaporated under vacuum. The residue was purified by column chromatography (1:2 CH₂Cl₂/hexanes) to give **1** (5.97 g, 90%) as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 7.90 (s, 2H), 1.58 (s, 9H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 143.48, 141.31, 126.66, 83.91, 27.93. HRMS (ESI⁺) *m*/*z* [M+H]⁺: Calcd for C₁₀H₁₂Br₂NO₂ 335.9229, found: 335.9233. 2.2 Synthesis of 2



In accord with a literature report,^{S2} α, α' -dibromo-*p*-xylene (6.72 g, 25.4 mmol) and sodium azide (1.61 g, 25.4 mmol) were dissolved in dimethylformamide (DMF) (30 mL). The reaction was stirred at room temperature overnight. Water (100 mL) was then added to quench the reaction and the mixture was extracted with DCM (3 × 100 mL). The combined organic layer was then washed with water several times to remove residual DMF. After evaporation of the volatiles, the crude product was purified by column chromatography (6:1 hexanes/CH₂Cl₂) to give **2** (2.30 g, 10.2 mmol, 45%) as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 7.42 (d, *J* = 8.1 Hz, 2H), 7.30 (d, *J* = 8.2 Hz, 2H), 4.50 (s, 2H), 4.34 (s, 2H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 137.9, 135.7, 129.5, 128.6, 54.4, 32.9. HRMS (CI⁺) *m*/*z* [M]⁺: Calcd for C₈H₈BrN₃ 224.9902, found: 224.9900.

2.3 Synthesis of 3



Precursor 1 (6.12 g, 18.16 mmol), CuI (340 mg, 1.82 mmol) and Pd(PPh₃)₄ (2.10 g, 1.82 mmol) were dissolved in 120 ml of a 1:1 (v/v) mixture of tetrahydrofuran (THF) and diisopropylamine. The solution was purged with N₂ gas for 20 minutes. Trimethylsilylacetylene (6.5 ml, 45.4 mmol) was then added using a syringe. The reaction was stirred at 50 °C for 8 h. After the volatiles were removed under reduced pressure, the residue was purified by column chromatography (20:1 hexanes/ethyl acetate) to give **3** (5.69 g, 84%) as a bright yellow solid. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 7.82 (s, 2H), 1.59 (s, 9H), 0.25 (s, 18H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 162.90, 144.00, 139.95, 125.82, 102.60, 96.54, 83.07, 28.00, -0.40. HRMS (ESI⁺) *m*/z [M+H]⁺: Calcd for C₂₀H₃₀NO₂Si₂ 372.1810, found: 372.1824.

2.4 Synthesis of 4



According to a modified literature procedure,^{S3} intermediate **3** (1.5 g, 4.04 mmol), AgNO₃ (100 mg, 0.6 mmol) and N-iodosuccinimide (NIS) (1.98 g, 8.8 mmol) were dissolved in dry DMF (30

ml). The mixture was stirred in darkness at room temperature overnight. Subsequently, the solution was partitioned with water (100 ml) and DCM (100 ml). The organic phase was separated off, washed with water a few more times and dried over MgSO₄. This organic solution was then concentrated under vacuum and the resulting crude material purified by column chromatography over silica gel (4:1 hexanes/ethyl acetate) to give **4** as a dark red powder (1.73 g, 89%). ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 7.79 (s, 2H), 1.57 (s, 9H). ¹³C NMR (CDCl₃, 100 MHz) δ (ppm): 143.83, 126.31, 92.97, 83.29, 27.99, 12.25. HRMS (CI⁺) *m/z* [M]⁺: Calcd for C₁₄H₁₁NO₂I₂ 478.8879, found: 478.8880.

2.5 Synthesis of 5



Intermediates **2** (742.5 mg, 3.3 mmol) and **4** (632.4 mg, 1.32 mmol) were added to dry THF (40 mL). The mixture was purged with N₂ gas. Cu[(MeCN)₄]PF₆ (50 mg, 0.132 mmol) and tris[(1-benzyl-1*H*-1,2,3-triazol-4-yl)methyl]amine (TBTA) (70 mg, 0.132 mmol) were then added to the mixture. The reaction was stirred at room temperature overnight protected from light. Subsequently, the reaction mixture was concentrated under vacuum and partitioned between a Na₂EDTA solution (100 ml) and DCM (100 ml). The organic phase was collected and concentrated under vacuum. The residue was purified by column chromatography (2:1 hexanes/ethyl acetate) over silica gel to give **5** as a pale yellow solid (880 mg, 72%). ¹H NMR (DMSO-*d*₆, 400 MHz) δ (ppm): 8.35 (s, 2H), 7.12-7.55 (m, 8H), 5.77 (s, 4H), 4.69 (s, 4H), 1.61 (s, 9H). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ (ppm):138.37, 135.73, 130.20, 128.22, 34.35, 28.07. HRMS (ESI⁺) *m*/z [M+H]⁺: Calcd for C₃₀H₂₈Br₂I₂N₇O₂ 929.8755, found: 929.8732.

2.6 Synthesis of 6 & Ibox



Intermediate 5 (880 mg, 0.947 mmol) and NaN₃ (154 mg, 2.37 mmol) were dissolved in DMF (60 ml). The mixture was stirred at room temperature protected from the light for 24 h. Then, water (300 mL) was added to quench the reaction, which was extracted with DCM (100 ml).

The organic layer was washed with water a few more times and dried over Na₂SO₄ before the volatiles were removed under vacuum to give crude **6** in essentially quantitative yield. This material was used without further purification. Here, Cu[(MeCN)₄]PF₆ (7.9 mg, 0.021mmol) and TBTA (11 mg, 0.021 mmol) were dissolved in dry THF (100 ml) under an N₂ atmosphere (referred to as the catalyst solution). Bis-azide **6** (178 mg, 0.209 mmol) and diiodide **4** (100 mg, 0.209 mmol) were dissolved in 10 ml dry THF and added to the catalyst solution over the course of 10 h by means of a syringe pump. The reaction mixture was stirred for another 48 h in the dark. Subsequently, the reaction mixture was concentrated under vacuum and partitioned between a Na₂EDTA solution (100 ml) and DCM (100 ml). The organic phase was separated and concentrated under reduced pressure. The residue was purified by column chromatography over silica gel (1:1 hexane/ethyl acetate) to give the Ibox in 60% isolated yield. ¹H NMR (CDCl₃, 400 MHz) δ (ppm): 8.56 (s, 4H), 7.18 (s, 8H), 5.68 (s, 8H), 1.62 (s, 18H). Due to solubility issues, a clean ¹³C NMR spectrum could not be obtained. HRMS (ESI⁺) *m/z* [M+2H]²⁺: Calcd for C₄₄H₄₀I₄N₁₄O₄ 667.9762, found: 667.9778. This product was also characterized by means of a single crystal X-ray diffraction analysis.

2.7 Characterization data



Fig. S1. ¹H NMR spectrum of compound 1 recorded in CDCl₃.



Fig. S2. ¹³C NMR spectrum of compound 1 recorded in CDCl₃.



Fig. S3. ¹H NMR spectrum of compound 2 recorded in CDCl₃.



Fig. S4. ¹³C NMR spectrum of compound 2 recorded in CDCl₃.



Fig. S5. ¹H NMR spectrum of compound 3 recorded in CDCl₃.



Fig. S6. ¹³C NMR spectrum of compound 3 recorded in CDCl₃.



Fig. S7. ¹H NMR spectrum of compound 4 recorded in CDCl₃.



Fig. S8. ¹³C NMR spectrum of compound 4 recorded in CDCl₃.



Fig. S9. ¹H NMR spectrum of compound 5 recorded in DMSO-*d*₆.



Fig. S10. ¹³C NMR spectrum of compound 5 recorded in DMSO-*d*₆.



Fig. S11. ¹H NMR spectrum of compound 6 recorded in CDCl₃.



Fig. S12. ¹H NMR spectrum of the Ibox recorded in CDCl₃. The asterisks denotes residual solvent peaks.



Fig. S13. HR-ESI mass spectrum of the Ibox.

3.¹H NMR titration data



Fig. S14. Selected region of the ¹H NMR spectra (CDCl₃, 298 K) acquired during the titration of Ibox with increasing quantities of TBACI: 0, 0.20, 0.39, 0.58, 0.77, 0.95, 1.13, 1.48, 1.82, 2.14, 2.46, 2.76, 3.33, 3.87, 4.38, 4.85, 5.29, 6.30, 7.18, 7.95, 8.64, 9.24, 10.29, 11.15, 11.87, 12.48, 13.00 and 13.46 equiv. from bottom to top.



Fig. S15. Nonlinear least-square analysis of the ¹H NMR binding data corresponding to the formation of $[Ibox \cdot 2Cl]^{2-}$ using the BindFit web applet.^{S4} The data were fitted to a 1:2 binding model to give $K_{11} = (173 \pm 5.3)$ and $K_{12} = (140 \pm 1.7)$. The residual distribution is shown below the binding isotherm.



9.4 9.2 9.0 8.8 8.6 8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 Chemical shift (ppm)

Fig. S16. Selected region of the ¹H NMR spectra (CDCl₃, 298 K) acquired during the titration of Ibox with increasing quantities of TBABr: 0, 0.20, 0.39, 0.58, 0.77, 0.95, 1.13, 1.48, 1.82, 2.14, 2.46, 2.76, 3.33, 3.87, 4.38, 4.85, 5.29, 6.30, 7.18, 7.95, 8.64, 9.24, 10.29, 11.15, 11.87, 12.48, 13.00 and 13.46 equiv. from bottom to top.



Fig. S17. Nonlinear least-square analysis of the ¹H NMR binding data corresponding to the formation of $[Ibox \cdot 2Br]^{2-}$ using the BindFit web applet. The data were fitted to a 1:2 binding model to give $K_{11} = (238 \pm 7.2)$ and $K_{12} = (283 \pm 3.1)$. The residual distribution is shown below the binding isotherm.



.4 9.2 9.0 8.8 8.6 8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 Chemical shift (ppm)

Fig. S18. Selected region of ¹H NMR spectra (CDCl₃, 298 K) acquired during the titration of Ibox with increasing quantities of TBAI: 0, 0.20, 0.39, 0.58, 0.77, 0.95, 1.13, 1.48, 1.82, 2.14, 2.46, 2.76, 3.33, 3.87, 4.38, 4.85, 5.29, 6.30, 7.18, 7.95, 8.64, 9.24, 10.29, 11.15, 11.87, 12.48, 13.00 and 13.46 equiv. from bottom to top.



Fig. S19. Nonlinear least-square analysis of the ¹H NMR binding data corresponding to the formation of $[Ibox \cdot 2I]^{2-}$ using the BindFit web applet. The data were fitted to a 1:2 binding model to give $K_{11} = (187 \pm 9.7)$ and $K_{12} = (516 \pm 14)$. The residual distribution is shown below the binding isotherm.

4. Supporting X-ray experimental details

X-ray Experimental for 2C₄₄H₃₈N₁₄O₄I₄ - CHCl₃: Crystals grew as colorless plates by vapor diffusion of hexane into a chloroform solution. The data crystal was cut from a larger crystal and had approximate dimensions; 0.16 x 0.047 x 0.044 mm. The data were collected on an Rigaku Oxford Diffraction SuperNova Dual Source diffractometer using a µ-focus Cu Ka radiation source ($\lambda = 1.5418$ Å) with collimating mirror monochromators. A total of 3146 frames of data were collected using ω -scans with a scan range of 1° and a counting time of 6 seconds per frame for frames collected with a detector offset of -41.6° and 24 seconds per frame with frames collected with a detector offset of 107.1°. The data were collected at 100 K using an Oxford Cryostream low temperature device. Details of crystal data, data collection and structure refinement are listed in Table 1. Data collection, unit cell refinement and data reduction were performed using Rigaku Oxford Diffraction's CrysAlisPro V 1.171.41.120a.^{S5} The structure was solved by direct methods using SHELXT^{S6} and refined by full-matrix least-squares on F² with anisotropic displacement parameters for the non-H atoms using SHELXL-2018/3.^{S7} Structure analysis was aided by use of the programs PLATON^{S8}, OLEX2^{S9}. The hydrogen atoms were calculated in ideal positions with isotropic displacement parameters set to 1.2xUeq of the attached atom (1.5xUeq for methyl hydrogen atoms).

There are two macrocycles in the asymmetric unit. Each lies around a different crystallographic inversion center. The phenyl rings were disordered in both. In one macrocycle, the *t*-butyl groups were disordered. The disorder was modeled using features available in OLEX2. A large region of disordered solvent was observed in the unit cell. The solvent mix could not be satisfactorily modeled. The contributions to the scattering factors due to these solvent molecules were removed by using SQUEEZE^{S10}.

The function, $\Sigma w(|F_0|^2 - |F_c|^2)^2$, was minimized, where $w = 1/[(\sigma (F_0))^2 + (0.2*P)^2]$ and $P = (|F_0|^2 + 2|F_c|^2)/3$. $R_w(F^2)$ refined to 0.225, with R(F) equal to 0.0720 and a goodness of fit, S, = 0.880. Definitions used for calculating R(F), $R_w(F^2)$ and the goodness of fit, S, are given below.^{S11} The data were checked for secondary extinction effects but no correction was

necessary. Neutral atom scattering factors and values used to calculate the linear absorption coefficient are from the International Tables for X-ray Crystallography (1992).^{S12} All figures were generated using SHELXTL/PC.^{S13} Tables of positional and thermal parameters, bond lengths and angles, torsion angles and figures are found in the cif files that may be obtained from the Cambridge Crystallographic Data Centre upon request by quoting no. 2126348.

Table S1. Crystal data and structure	refinement for 2Ibox·CHC	13.		
Empirical formula	C45 H39 C13 I4 N14	C45 H39 Cl3 I4 N14 O4		
Formula weight	1453.85	1453.85		
Temperature	100.02(14) K			
Wavelength	1.54184 Å	1.54184 Å		
Crystal system	triclinic			
Space group	P -1			
Unit cell dimensions	a = 13.4032(3) Å	$\alpha = 97.062(2)^{\circ}.$		
	b = 13.4811(6) Å	$\beta = 100.4278(18)^{\circ}.$		
	c = 19.0017(3) Å	$\gamma = 115.900(3)^{\circ}$.		
Volume	2957.28(17) Å ³			
Z	2			
Density (calculated)	1.633 Mg/m ³			
Absorption coefficient	18.208 mm ⁻¹			
F(000)	1404			
Crystal size	0.252 x 0.135 x 0.057	0.252 x 0.135 x 0.057 mm ³		
Theta range for data collection	2.428 to 73.796°.	2.428 to 73.796°.		
Index ranges	-16<=h<=14, -16<=k	-16<=h<=14, -16<=k<=16, -23<=l<=23		
Reflections collected	48255	48255		
Independent reflections	11564 [R(int) = 0.106	11564 [R(int) = 0.1067]		
Completeness to theta = 67.684°	99.5 %	99.5 %		
Absorption correction	Semi-empirical from	Semi-empirical from equivalents		
Max. and min. transmission	1.000 and 0.45264	1.000 and 0.45264		
Refinement method	Full-matrix least-squa	Full-matrix least-squares on F ²		
Data / restraints / parameters	11564 / 897 / 785	11564 / 897 / 785		
Goodness-of-fit on F ²	0.908	0.908		
Final R indices [I>2sigma(I)]	R1 = 0.0720, wR2 = 0	R1 = 0.0720, $wR2 = 0.2102$		

R indices (all data)	R1 = 0.0788, wR2 = 0.2250
Extinction coefficient	n/a
Largest diff. peak and hole	3.934 and -1.952 e.Å ⁻³
CCDC deposition number	2126348



Fig. S20. View of the macrocycle 1 in Ibox showing the atom labeling scheme. Displacement ellipsoids are scaled to the 50% probability level. The lower occupancy atoms of the disordered phenol ring groups were omitted. The macrocycle resides around a crystallographic inversion center at 1, $\frac{1}{2}$, 1. Atoms with labels appended by a ' are related by 2-x, 1-y, 2-z.



Fig. S21. View of the macrocycle 2 in 1 showing the atom labeling scheme. Displacement ellipsoids are scaled to the 50% probability level. The lower occupancy atoms of the disordered phenol ring groups were omitted. The macrocycle resides around a crystallographic inversion center at $\frac{1}{2}$, 0, 1. Atoms with labels appended by a ' are related by 1-x, -y, 2-z.

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- S11. $R_W(F^2) = \{\Sigma w(|F_0|^2 |F_c|^2)^2 / \Sigma w(|F_0|)^4\}^{1/2}$ where w is the weight given each reflection. $R(F) = \Sigma (|F_0| - |F_c|) / \Sigma |F_0|\}$ for reflections with $F_0 > 4(\sigma(F_0))$. $S = [\Sigma w(|F_0|^2 - |F_c|^2)^2 / (n - p)]^{1/2}$, where n is the number of reflections and p is the number of refined parameters.
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