Supporting Information

A Transtion-Metal Free Three-Component Coupling Approach to

Quinap Derivates

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1. General Information

Commercial reagents were purchased from commercial suppliers and used without further purification. All dry solvents were treated according to standard procedures prior to use unless otherwise noted. Thin-layer chromatography (TLC) was performed using 60 mesh silica gel plates visualized with short-wavelength UV light (254 nm). Column chromatography was performed using silica gel (200-300 mesh). ¹H and ¹³C NMR spectra were recorded on a Bruker instrument 400(400 MHz for ¹H,100 MHz for ¹³C and 162 MHz for ³¹P) in CDCl₃ or Bruker instrument 300 (300 MHz for ¹H, 75 MHz for ¹³C and and 121 MHz for ³¹P) using CDCl₃ as the solvent and tetramethyl silane (TMS) as internal reference. TMS Chemical shifts (δ) were measured in ppm relative to TMS $\delta = 0$ for ¹H, or to chloroform $\delta = 77.0$ for ¹³C as internal standard. The following abbreviations (or combinations there of) were used to explain multiplicities: s = singlet, d =doublet, t = triplet, q = quartet, m = multiplet, br = broad signals. The ¹H NMR spectra are reported as follows: chemical shift δ in ppm relative to TMS ($\delta = 0$ ppm), multiplicity, coupling constant J, are reported in hertz, number of protons. 13 C NMR spectra are reported as follows: chemical shift δ in ppm relative to CDCl₃ (δ = 77.0 ppm), multiplicity, coupling constant J, are reported in hertz. Mass spectroscopy data of the products were collected on a Bruker esquire 6000 instrument using ESI ionization.

2. Preparation of starting materials

Substituted aniline **1a-1k**, were purchased from commercial source, and used without further purification.

Synthesis of substituted Alkyne

Substituted alkyne 2a-2e and 2i-2m were purchased from commercial source, and used without further purification. $2f^{[1][2]}, 2g^{[3]}$ was prepared according to corresponding literatures. Procedure to prepare $2f^{[1][2]}$:



To a solution of the veratraldehyde (0.83 g, 5.0 mmol) in 10 ml dry DCM, carbon tetrabromide (1.74 g, 5.25 mmol) was added. The mixture was cooled to 0 °C, and a solution of triphenylphosphine (2.62 g, 10.0 mmol) in 10 mL dry DCM was added dropwise. The reaction mixture was allowed to gradually warm up to room temperature and stirred overnight at room temperature. The reaction mixture was quenched with water, and the organic layer was separated and washed with brine. The crude mixture concentrated under vacuum, and purified by flash column chromatography get the dibromoethane derivative S1, yellow oil (1.45g, 90%).

To a stirred solution of dibromoethane derivative S1 (1.45 g, 4.5 mmol) in anhydrous CH_3CN (10 mL) was added DBU (2.8 mL, 18.0 mmol) dropwise at 0 °C. The reaction mixture was warm to ambient temperature (25-30 °C) stir at for one day. After completion of reaction (monitored by TLC), reaction mixture quenched by dropwise addition of aqueous HCl then continued stirring for 5 mins. The reaction mixture was extracted three times with DCM; organic layers were washed with brine, solvent was evaporated under vacuum, and resulting residues were purified by flash

column chromatography to afford the **2f** as a yellow solid (0.47 g, 64%). ¹**H** NMR (400 MHz, CDCl₃) δ 7.11 (dd, J = 8.6, 1.8 Hz, 1H), 6.99 (d, J = 1.6 Hz, 1 H), 6.81 (d, J = 8.4 Hz, 1H), 3.89 (s, 3 H), 3.88 (s, 3 H), 3.01 (s, 1 H); ¹³C NMR (100 MHz, CDCl₃) δ 149.8, 148.5, 125.4, 114.6, 114.1, 110.8, 83.7, 75.7, 55.8. MS (ESI): found [M+H]⁺ 162.1. Spectral data for this compound is consistent with that previously reported.^[2]

Procedure to prepare 2g^[3]:



To a solution of o-ethynyl aniline (0.227 mL, 2.0 mmol), Et₃N (0.42 mL, 3.2 mmol) in THF (4 mL) was added acetyl chloride (0.21 mL,3.0 mmol) dropwise at 0 °C. After the addition was complete, the reaction mixture was stirred at room temperature. The reaction was monitored by TLC, after completion of the reaction, the reaction was quenched by water and the resulting mixture was extracted three times with DCM. The combined extracts were washed with brine and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel to afford *N*-acetyl-o-ethynyl aniline **2g** as a white solid (0.30g, 95%). ¹H NMR (400 MHz, CDCl₃) δ 8.40 (d, *J* = 8.4 Hz, 1H), 7.92 (br, 1H), 7.46 (dd, *J* = 7.6 , 1.2 Hz, 1H), 7.36 (t, *J* = 8.0 Hz, 1H), 7.04 (t, *J* = 7.6 Hz, 1H), 3.51 (s, 1H), 2.23 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 168.3, 139.6, 132.1, 130.2, 123.3, 119.2, 110.4, 84.3, 79.2, 24.9. MS (ESI): found [M+H]⁺ 159.9. Spectral data for this compound is consistent with that previously reported.^[3]

Synthesis of substituted aldehyde

General Procedure A to prepare 3a-3d,3h^{[4][5]}:



To a solution of 2-bromobenzonitrile (1.0 equiv) in dry THF under Ar at -78 °C, nbutyllithium (2.4M in hexane, 1.0 equiv) was added dropwise, the mixture stirs at -78 °C for 1 hours. Then the solution of Ph₂PCl or Cy₂PCl (1.0 equiv) in dry THF was added dropwise to the solution, the mixture was stirred for further 30 min at -78 °C, and the reaction mixture allowed to warm to room temperature stir over night, and quenched by water. The resulting mixture was extracted three times with DCM. The combined extracts concentrated in vacuo, and the residue washed by methanol to give the rough product, use in next step without further purification.

A mixture of S2 in concerned hydrochloric acid (1 mmol S2~2 mL) was refluxed at 120 °C with vigorous stirring for overnight. During this period, after cooling down to room temperature, water was added and the mixture stirred for 30 min, and the resulting mixture was extracted three times with DCM. The combined extracts were washed with brine and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel to afford product.

2-(diphenylphosphoryl)benzaldehyde (3a)

2-bromobenzonitrile (9.1 g, 50.0 mmol) was following the General Procedure A. The crude mixture was purified by column chromatography (petroleum ether /ethyl acetate = 1:1) to afford a white solid (8.9 g, 60%). ¹H NMR (400 MHz, CDCl₃): δ 10.75 (s, 1H), 8.21-8.10 (m, 1H), 7.74-7.46 (m, 12 H), 7.23 (dd, J = 13.6, 7.6Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 191.2 (d, $J_{CP} =$

5.6 Hz), 139.4 (d, $J_{C-P} = 6.5$ Hz), 135.1 (d, $J_{C-P} = 95.4$ Hz), 133.6 (d, $J_{C-P} = 11.0$ Hz), 132.8, 132.7, 132.3 (d, $J_{C-P} = 2.8$ Hz), 131.8 (d, $J_{C-P} = 10.0$ Hz), 131.6 (d, $J_{C-P} = 12.8$ Hz), 129.0 (d, $J_{C-P} = 8.7$ Hz), 128.8 (d, $J_{C-P} = 12.3$ Hz); ³¹P NMR (162 MHz, CDCl₃): δ 31.05. MS (ESI): found [M+H]⁺ 307.0. Spectral data for this compound is consistent with that previously reported.^[5]

2-(diphenylphosphoryl)-6-methylbenzaldehyde (3b)

2-bromo-6-methylbenzonitrile (3.6 g, 18.5 mmol) was following the General Procedure A. The crude mixture was purified by column chromatography (petroleum ether /ethyl acetate = 1:1) to afford a white solid (2.87 g, 48%). ¹**H NMR** (400 MHz, CDCl₃) δ 10.84 (s, 1H), 7.65 (dd, *J* = 12.2, 7.4 Hz, 4H), 7.58 (t, *J* = 7.4 Hz, 2H), 7.53-7.44 (m, 5H), 7.37 (td, *J* = 7.6, 2.4Hz, 1H), 7.10 (dd, *J* = 14.0, 7.6 Hz, 1H), 2.62 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 193.4 (d, *J*_{C-P} = 6.7 Hz), 141.5 (d, *J*_{C-P} = 8.7 Hz), 137.9 (d, *J*_{C-P} = 6.4 Hz), 136.1 (d, *J*_{C-P} = 2.5 Hz), 136.0, 135.1, 133.0, 132.2 (d, *J*_{C-P} = 2.8 Hz), 131.9 (d, *J*_{C-P} = 9.8 Hz), 131.1 (d, *J*_{C-P} = 13.2 Hz), 128.7 (d, *J*_{C-P} = 12.3 Hz), 21.7; ³¹**P NMR** (162 MHz, CDCl₃) δ 32.38. **MS (ESI):** found [M+H]⁺ 321.2. Spectral data for this compound is consistent with that previously reported.^[5]

2-(diphenylphosphoryl)-4-methylbenzaldehyde (3c)

2-bromo-4-methylbenzonitrile (2.8 g, 14.4 mmol) was following the General Procedure A. The crude mixture was purified by column chromatography (petroleum ether/ethyl acetate = 1:1) to afford a white solid (2.1 g, 46 %). ¹H NMR (400 MHz, CDCl₃) δ 10.62 (s, 1 H), 8.07 (qt, *J* = 4.0, 1.6 Hz, 1H), 7.71-7.62 (m, 4 H), 7.61-7.55 (m, 2H), 7.54-7.45 (m, 5 H), 7.08 (d, *J* = 14.0 Hz, 1 H), 2.35 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 190.9 (d, *J*_{C-P} = 5.3 Hz), 143.9 (d, *J*_{C-P} = 11.8 Hz), 137.1 (d, *J*_{C-P} = 6.6 Hz), 135.4, 134.3 (d, *J*_{C-P} = 10.5 Hz), 133.0 (d, *J*_{C-P} = 2.3 Hz),132.3(d, *J*_{C-P} = 2.7 Hz), 131.9 (d, *J*_{C-P} = 9.9 Hz), 131.8, 129.4 (d, *J*_{C-P} = 9.3 Hz), 128.8 (d, *J*_{C-P} = 12.2 Hz), 21.9; ³¹P NMR (162MHz, CDCl₃) δ 30.86. MS (ESI): found [M+H]⁺ 321.2. Spectral data for this compound is consistent with that previously reported.^[5]



2-(diphenylphosphoryl)-1-naphthaldehyde (3d)

To obtain the substrate S3, n-BuLi (2.4 M in hexane, 23 mL, 1.2 equiv.) was slowly added to the 2,2,6,6-tetramethylpiperidine in THF solution at -5 °C - 0 °C. After the reaction for 1 h, the temperature was reduced to -78 °C, then add 1-naphthalenenitrile (45.7 mmol, 1.0 equiv.) in THF slowly to the solution, continue the reaction for 1h. Then add I₂ (45.7 mmol, 1.0 equiv.) solution in THF into reaction systems, continue stir at -78 °C for 2h, slowly raise to room temperature and stir overnight, the reaction was quenched with water, the reaction solution was washed with NaHSO₃ solution, and extracted three times with DCM. The organic phases were combined and the solvent was evaporated. 2-Iodo-1-naphthalenecarbonitrile was isolated by column chromatography.

1-iodo-2-naphthonitrile (9.7 g, 34.8 mmol) was following the General Procedure A. The residue was purified by column chromatography (petroleum ether/ethyl acetate = 1:2) to provide a white solid **3d** (4.3 g, 35 %), ¹**H NMR** (400 MHz, CDCl₃) δ 11.22 (s, 1H), 8.91(d, J = 8.2 Hz, 1H), 7.96 (dd, J = 8.4, 2.2 Hz, 1H), 7.90-7.87 (m, 1H), 7.74-7.56 (m, 8H), 7.50 (td, J = 7.4, 3.0

Hz, 4H), 7.28 (dd, J = 12.3, 8.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 194.2 (d, $J_{C-P} = 7.5$ Hz), 135.1 (d, $J_{C-P} = 2.2$ Hz), 135.0, 134.0, 133.0, 132.4 (d, $J_{C-P} = 2.7$ Hz), 132.3, 132.0 (d, $J_{C-P} = 10.0$ Hz), 130.5 (d, $J_{C-P} = 10.2$ Hz), 129.4, 128.7 (d, $J_{C-P} = 12.3$ Hz), 128.6, 128.3, 127.9 (d, $J_{C-P} = 11.5$ Hz), 126.6; ³¹P NMR (162 MHz, CDCl₃) δ 32.49. MS (ESI): found [M+H]⁺ 357.0. Spectral data for this compound is consistent with that previously reported.^[5]



2-(dicyclohexylphosphoryl)benzaldehyde (3h)

2-bromobenzonitrile (4.0 g, 21.9 mmol) was following the General Procedure A. The crude mixture was purified by column chromatography (petroleum ether /ethyl acetate = 1:1) to afford a white solid **3h** (2.5 g, 36 %), ¹**H NMR** (400 MHz, CDCl₃) δ 10.88 (br, 1H), 8.07 (s, 1H), 7.87-7.56 (m, 3H), 2.34-2.04 (m, 4H), 1.86 (d, J = 12.0 Hz, 2H) ,1.78-1.58 (m, 4H), 1.56-1.06 (m, 12H); ¹³**C NMR** (100 MHz, CDCl₃) δ 193.9, 132.7, 132.6, 132.4, 131.6, 131.4, 131.4, 36.0 (d, $J_{C-P} = 66.6 \text{ Hz}$), 26.3 (dd, $J_{C-P} = 12.8$, 10.3 Hz), 25.7 (d, $J_{C-P} = 2.7 \text{ Hz}$), 25.6 (d, $J_{C-P} = 0.8 \text{ Hz}$), 25.1 (d, $J_{C-P} = 1.6 \text{ Hz}$); ³¹**P NMR** (162MHz, CDCl₃) δ 51.28. **MS (ESI):** found [M+H]⁺ 319.2. Spectral data for this compound is consistent with that previously reported.^[5]

Procedure for prepare 3e-3g^{[5][6][7]}:

General Procedure B to prepare Cyclic acetal

A solution of the aldehyde (1.0 equiv), Ethylene glycol (5.0 equiv), and *p*-Toluenesulfonic acid (5.0 mol %) in toluene (10 mL/g) was heated at reflux for 24 h. After cooling to room temperature, the reaction was quenched by the addition of saturated aqueous NaHCO₃. The mixture was extracted with DCM and the combined organic was washed with water and brine, filtered and concentrated in vacuo, the crude residue was then purified by column chromatography.

General Procedure C to prepare Cycloacetal phosphoxide

For the Synthesis of 1,3-dioxolanes diphenyl phosphine oxide, to a solution of cyclic acetal (1.0 eq) in anhydrous THF (10 mL/g), kept in an oven-dried Schlenk flask under an atmosphere of dry argon, was added dropwise a solution of n-BuLi in hexane (1.0 eq) at -78 °C. After stirring at -78 °C for 1 h, Ph₂PCl (1.0 eq) in anhydrous THF was added dropwise. Then the solution was stirred for 1 h at -78 °C, allowed to warm to room temperature and stirred for overnight. The reaction was quenched with a solution of NH₄Cl (aq.) and the product was extracted with DCM. The combined organic layer was washed with water and brine. Filtered and the residue was purified by column chromatography to afford the product.

General Procedure D to prepare Aldehyde phosphoxide^[7]

p-Toluenesulfonic acid (5 mol %) was added to a solution of 1,3-dioxolanes diphenyl phosphine oxide (1.0 eq) in a mixture of acetone/water. The mixture was refluxed until completion. After the completion of the reaction, the mixture was cooled to room temperature, and the acetone was removed under vacuum. The residue was dissolved in DCM, washed with saturated aqueous NaHCO₃, water and brine, and concentrated under vacuum. The crude product was purified by silica gel column chromatography to afford the product.

Procedure for synthesis of 3e^[7]:



2-(3-bromophenyl)-1,3-dioxolane (85)

3-bromobenzaldehyde (3.7 g, 20 mmol) was following the General Procedure B. The residue was purified by column chromatography (petroleum ether/ethyl acetate = 50:1) to afford a Colorless oily compound (3.04g, 67%).

(3-(1,3-dioxolan-2-yl)phenyl)diphenylphosphine oxide (S6)

2-(3-bromophenyl)-1,3-dioxolane (3.04 g, 13.3 mmol) was following the General Procedure C. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:3) to afford a white solid (1.42g, 31%).

3-(diphenylphosphoryl) benzaldehyde (3e)

(3-(1,3-dioxolan-2-yl)phenyl)diphenylphosphine oxide (1.42 g, 4.1 mmol) was following the General Procedure D. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:2) to provide a white viscous substance **3e** (1.07g, 85%). ¹**H NMR** (400 MHz, CDCl₃) δ 10.03 (s, 1H), 8.21-8.15 (m, 1H), 8.11-8.05 (m, 1H), 8.02-7.94 (m, 1H), 7.73-7.64 (m, 5H), 7.63-7.56 (m, 2H), 7.54-7.46 (m, 4H); ¹³**C NMR** (100 MHz, CDCl₃) δ 191.4, 137.5 (d, *J*_{C-P} = 9.9 Hz), 136.2 (d, *J*_{C-P} = 10.9 Hz), 134.3 (d, *J*_{C-P} = 101.5 Hz), 133.6 (d, *J*_{C-P} = 10.1 Hz), 132.3 (d, *J*_{C-P} = 2.8 Hz), 132.2 (d, *J*_{C-P} = 2.5 Hz), 132.0 (d, *J*_{C-P} = 10.0 Hz), 131.0, 129.4 (d, *J*_{C-P} = 11.5 Hz), 128.7 (d, *J*_{C-P} = 12.2 Hz); ³¹**P NMR** (162 MHz, CDCl₃) δ 28.28. **MS (ESI):** found [M+H]⁺ 307.0. Spectral data for this compound is consistent with that previously reported.^[7]

Procedure for synthesis of 3f-3g^[5]:



5-bromo-6-(1,3-dioxolan-2-yl)benzo[d][1,3]dioxole (S7a)

6-bromobenzo[d][1,3]dioxole-5-carbaldehyde (3.43 g, 14.9 mmol) was following the General Procedure B. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 30:1) to afford a white solid (3.63 g, 89 %).

(6-(1,3-dioxolan-2-yl)benzo[d][1,3]dioxol-5-yl)diphenylphosphine oxide (S8a)

5-bromo-6-(1,3-dioxolan-2-yl)benzo[d][1,3]dioxole (3.5 g, 12.9 mmol) was following the General Procedure C. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:3) to afford a white solid (4.0 g, 79 %).

6-(diphenylphosphoryl)benzo[d][1,3]dioxole-5-carbaldehyde (3f)

(6-(1,3-dioxolan-2-yl)benzo[d][1,3]dioxol-5-yl)diphenylphosphine oxide (2.0 g, 5.07 mmol) was following the General Procedure D. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:2) to provide a white solid **3f** (1.6 g, 90 %), ¹**H NMR** (400 MHz, CDCl₃) δ 10.63 (s, 1H), 7.77-7.56 (m, 7H), 7.50 (td, *J* = 7.4, 3.1 Hz, 4H), 6.59 (d, *J* = 13.2

Hz, 1H), 6.11 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 189.5 (d, $J_{C-P} = 6.1$ Hz), 151.2 (d, $J_{C-P} = 17.8$ Hz), 151.0 (d, $J_{C-P} = 2.5$ Hz), 136.1 (d, $J_{C-P} = 7.1$ Hz), 132.8, 132.4 (d, $J_{C-P} = 2.7$ Hz), 131.8 (d, $J_{C-P} = 10.0$ Hz), 130.8, 128.9 (d, $J_{C-P} = 12.3$ Hz), 113.1 (d, $J_{C-P} = 13.7$ Hz), 108.7 (d, $J_{C-P} = 11.2$ Hz), 102.6; ³¹P NMR (162 MHz, CDCl₃) δ 31.01. MS (ESI): found [M+H]⁺ 351.0. Spectral data for this compound is consistent with that previously reported.^[5]

2-(2-bromo-5-fluorophenyl)-1,3-dioxolane (S7b)

2-bromo-5-fluorobenzaldehyde (3.0 g, 14.8 mmol) was following the General Procedure B. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 30:1) to afford a Colorless oily compound (3.4 g, 93 %).

(2-(1,3-dioxolan-2-yl)-4-fluorophenyl)diphenylphosphine oxide (S8b)

2-(2-bromo-5-fluorophenyl)-1,3-dioxolane (3.4 g, 13.8 mmol) was following the General Procedure C. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:4) to afford a white solid (3.2 g, 78 %).

2-(diphenylphosphoryl)-5-fluorobenzaldehyde (3g)

(2-(1,3-dioxolan-2-yl)-4-fluorophenyl) diphenylphosphine oxide (1.1 g, 2.99 mmol) was following the General Procedure D. The residue was purified by column chromatography (petroleum ether /ethyl acetate = 1:4) to provide a white solid **3g** (0.86 g, 89 %), ¹**H NMR** (400 MHz, CDCl₃) δ 10.73 (d, *J* = 3.1 Hz, 1 H), 7.88-7.80 (m, 1 H), 7.70-7.58 (m, 6 H), 7.55-7.48 (m, 4 H), 7.26-7.17 (m, 2 H); ¹³**C NMR** (100 MHz, CDCl₃) δ 189.8 (dd, *J*_{C-F} = 1.0 Hz, *J*_{C-P} = 5.2 Hz), 166.2 (d, *J*_{C-P} = 2.9 Hz), 163.7 (d, *J*_{C-P} = 2.8 Hz), 142.4 (t, *J*_{C-P} = 7.2 Hz), 136.2 (dd, *J*_{C-F} = 8.0 Hz, *J*_{C-P} = 12.4 Hz), 132.6 (d, *J*_{C-P} = 2.8 Hz), 132.5, 131.8 (d, *J*_{C-P} = 10.0 Hz), 131.4, 128.9 (d, *J*_{C-P} = 12.3 Hz), 119.5 (dd, *J*_{C-F} = 13.0 Hz, *J*_{C-P} = 21.2 Hz), 115.9 (dd, *J*_{C-F} = 9.8 Hz, *J*_{C-P} = 22.4 Hz); ³¹**P NMR** (162 MHz, CDCl₃) δ 30.11. **MS (ESI):** found [M+H]⁺ 325.1. Spectral data for this compound is consistent with that previously reported.^[5]

3. Optimization of Reaction Condition

Table S1. Screening of different solvents ^[a] .				
	IH ₂ +	+ CHO PPh ₂ Al(OT 12	F) ₃ (20 mol %) plvent, air 0 °C, 24 h	^N N
1a	2a	3a		4a
-	Entry	solvent	Yield[%] ^b	-
	1	PhMe	45	-
	2	xylene	47	
	3	PhCl	61	
	4	1,4-dioxane	trace	
	5	DCE	41	
	6	H_2O	trace	
	7	DMF	N.R.	
	8	DMSO	N.R.	
	9	^t BuOH	34	
	10	Ethylene glycol	trace	
	11	neat	48	
-	12	HOAc	69	_

^a Reaction Conditions: Al(OTf)₃ (0.04 mmol,20 mol %), **1a** (0.22 mmol), **2a** (0.30 mmol), **3a**(0.20 mmol), solvent (2.0 mL), 120 °C

under Air condition for 24 h. ^b Isolated yield.

Table S2. Screening of different LA catalyst ^[a] .				
l) 1a	NH ₂ +	+ CHO LA cata PPh ₂ HI 120 3a	alyst (20 mol %) DAc, air I °C, 24 h	Ph ₂ P Ph Ph 4a
	Entry	LA catalyst	yield [%] ^b	
	1	AgOTf	55	
	2	Cu(OTf) ₂	42	
	3	Fe(OTf) ₂	58	
	4	Mg(OTf) ₂	52	
	5	Zn(OTf) ₂	64	
	6	Sc(OTf) ₂	58	
	7	Dy(OTf) ₂	56	
	8	AlCl3	63	
	9	FeCl ₃	57	
	10	FeCl ₂	55	
	11	HOTf	47	

^a Reaction Conditions: LA catalyst (20 mol %), **1a** (0.22 mmol), **2a** (0.30 mmol), **3a**(0.20 mmol), HOAc (2.0 mL), 120 °C under Air

condition for 24 h. ^b Isolated yield. ^c 0.40 mmol of **2a** were used.

Table S3. Screening of different temperature ^[a] .				
NH2	2 + + +	PPh ₂ Al(DTf) ₃ (20 mol %) HOAc, air T °C, 24 h	Ph ₂ P N
1a	2a	3a		Ph 4a
	Entry	T [°C]	Yield [%] ^b	
	1	rt	10	
	2	60	46	
	3	80	54	
	4	100	62	
	5	120	69	
	6	130	55	

^a Reaction Conditions: Al(OTf)₃ (0.04 mmol,20 mol %), **1a** (0.22 mmol), **2a** (0.30 mmol), **3a**(0.20 mmol), HOAc (2.0 mL), t °C under

Air condition for 24 h. ^b Isolated yield.

Table S4. Screening of different catalyst loading [a].



Entry	Al(OTf) ₃ (mol %)	Yield [%] ^b
1	20	69
2	10	55
3	5	50
4	0	34

^a Reaction Conditions: Al(OTf)₃ (x mol %), **1a** (0.22 mmol), **2a** (0.30 mmol), **3a**(0.20 mmol), HOAc (2.0 mL), 120 °C under Air

condition for 24 h. ^b Isolated yield. ^c 0.40 mmol of **2a** were used.

Table S5 . Screening of different oxidant ^[a] .			
	H ₂ + +	PPh ₂ Al(OTf) ₃ (20 oxidant (1 HOAc, 120	0 mol %) 5 eq.) °C, 24 h
1a	2a	3a	Ph 4a
	Entry	Oxidant	Yield [%] ^b
	1	$Cu(OAc)_2 \cdot H_2O$	40
	2	AgOAc	64
	3	1,4-BQ	48
	4	PhI(OAc) ₂	24
	5	$Ce(SO_4)_2$	45
	6	Mn(OAc) ₃	26

^aReaction Conditions: Al(OTf)₃ (0.04 mmol,20 mol %), **1a** (0.22 mmol), **2a** (0.30 mmol), **3a**(0.20 mmol), HOAc (2.0 mL), oxidant (0.30 mmol), 120 °C under Ar condition for 24 h. ^bIsolated yield.

4. The General Procedures

General procedure for reaction:



The mixture of amine **1a-1k** (1.1 equiv, 0.22 mmol), alkyne **2a-2l** (1.5 equiv, 0.30 mmol), aldehyde **3a-3h** (1.0 equiv, 0.20 mmol) and aluminium trifluromethanesulfonate (20 mol %, 0.04 mmol) was dissolved in acetic acid (2 mL). The reaction mixture was stirred at 120 °C in an oil bath for 24 hours under air atmosphere. After the starting material was consumed as indicated by TLC, acetic acid was evaporated concentrated. The crude product was extracted with dichloromethane, washed with saturated Na₂CO₃, and the organic solvent was evaporated concentrated in vacuo, purified by silica gel column chromatography (eluent: Petroleum ether: EtOAc = 1:1-1:3) affording the desired product.

General procedure for gram-scale reaction:



The mixture of amine 1a (1.1 equiv, 5.5 mmol), alkyne 2a (1.5 equiv, 7.5 mmol), aldehyde 3b or

3d (1.0 equiv, 5.0 mmol) and aluminium trifluromethanesulfonate (20 mol %, 1.0 mmol) was dissolved in acetic acid (50 mL) in a 100 mL flask. The reaction mixture was stirred at 120 °C in an oil bath for 24 hours under air atmosphere. After the starting material was consumed as indicated by TLC, acetic acid was evaporated concentrated. The crude product was extracted with dichloromethane, washed with saturated Na₂CO₃, and the organic solvent was evaporated concentrated in vacuo, purified by silica gel column chromatography (DCM: EtOAc = 10:1-2:1) affording the desired product.

General procedure for reduction 4 to 5:



In a 25 mL of reaction tube under Ar atmosphere, 4 (1.0 equiv,0.6 mmol) was added to dry toluene(6 mL), trichlorosilane(5.0 equiv, 3.0 mmol)was add to solutions dropwise, then triethylamine (3.3 mmol, 5.5 equiv) was added dropwise to the solution and refluxed at 100 °C overnight, after which the reaction mixture was cooled, carefully quenching the reaction with saturated aqueous NaHCO₃ solution, and then extracting the solution three times with DCM, The organic phase are combined, and removed solvent, and then the flash column chromatography(eluent: Petroleum ether : EtOAc = 20:1-5:1) method is used to obtain the phosphorus-containing quinoline compound 5.

5. Mechanism explaining the regioselectivity

As show in the following figure, a tentative mechanism was proposed. Initially, Intermediate B was formed by coordination of imine A, which was generated in situ, and alkyne to $Al^{(III)}$, and then addition of alkyne to imine A forms the propargylamine intermediate C, which then undergoes an intramolecular hydroarylation of alkyne to give dihydroquinoline intermediate D. Subsequently, a oxidation of D by O_2 in air affords the quinoline product 4a.

For unsymmetrical aniline derivatives as substrate, the regioselectivity of the reaction mainly depends on the electronic and steric effects of the ortho position of amino on the propargylamine intermediate C. The greater the steric hindrance effect of the ortho position of amino on the propargylamine intermediate C, the worse the regioselectivity, and the higher the electron cloud density of the ortho position of amino on the propargylamine intermediate C, the better the regioselectivity.





Scheme S1 Mechanism explaining the regioselectivity

6. Characterization of product



4a, Light yellow solid, 69 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.77-7.44 (m, 16H), 7.40 (dd, J = 8.6, 1.8 Hz, 1H), 7.28-7.15 (m, 6 H), 2.42 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 157.0 (d, $J_{C-P} = 3.8$ Hz), 146.5 (d, $J_{C-P} = 41.8$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 41.8$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 41.8$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 8.1$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 8.1$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 8.1$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 8.1$ Hz), 145.8 (d, $J_$

10.9 Hz), 133.2 (d, $J_{C-P} = 105.1$ Hz), 132.3, 132.0 (d, $J_{C-P} = 2.4$ Hz), 131.7 (d, $J_{C-P} = 9.7$ Hz), 131.3, 131.3, 131.0, 130.9 (d, $J_{C-P} = 2.5$ Hz), 129.6, 129.2, 128.4, 128.1, 127.9 (d, $J_{C-P} = 11.9$ Hz), 127.7 (d, $J_{C-P} = 12.2$ Hz), 124.9, 124.0, 123.0, 21.8; ³¹P NMR (162 MHz, CDCl₃) δ 29.79; HRMS (ESI, m/z): calculated for C₃₄H₂₇NOP⁺: 496.1825, found :496.1827 [M+H]⁺.



4b, Light yellow solid, 63 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.74-7.42 (m, 15H), 7.28-7.16 (m, 7H), 7.10 (d, *J*=2.8 Hz. 1H), 3.77 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 157.8, 155.5 (d, *J*_{*C-P*} = 3.8 Hz), 146.0, 145.8 (d, *J*_{*C-P*} = 8.1 Hz), 143.8, 138.1, 134.6 (d, *J*_{*C-P*} = 10.9 Hz), 133.2 (d, *J*_{*C-P*} = 105.0 Hz), 132.3, 132.0 (d, *J*_{*C-P*} = 2.5 Hz), 131.6 (d, *J*_{*C-P*} = 9.6 Hz), 131.3, 130.9 (d, *J*_{*C-P*} = 3.0 Hz), 130.8 (d, *J*_{*C-P*} = 2.6 Hz), 128.9 (d, *J*_{*C-P*} = 79.1 Hz), 128.2, 127.8 (d, *J*_{*C-P*} = 12.1 Hz), 127.7 (d, *J*_{*C-P*} = 12.2 Hz), 127.7 (d, *J*_{*C-P*} = 3.0 Hz), 125.9, 123.3, 121.4, 103.2, 55.3; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.72; **HRMS (ESI, m/z)**: calculated for C₃₄H₂₇NO₂P⁺: 512.1774, found: 512.1775 [M+H]⁺.



4c, Light yellow solid, 47 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 8.59 (d, J = 1.7 Hz, 1H), 8.18 (dd, J = 8.8, 1.9 Hz, 1H), 7.75-7.59 (m, 9H), 7.57-7.47 (m, 6H), 7.27-7.16 (m, 6H), 3.92 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 166.8, 160.1 (d, $J_{C-P} = 3.9$ Hz), 149.3(d, $J_{C-P} = 81.3$ Hz), 145.3 (d, $J_{C-P} = 7.7$ Hz), 137.2, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.6, 132.5, 132.5, 132.1 (d, $J_{C-P} = 2.4$ Hz), 131.7 (d, $J_{C-P} = 9.6$ Hz), 131.5, 131.1 (d, $J_{C-P} = 2.7$ Hz), 130.9 (d, $J_{C-P} = 9.2$ Hz), 129.8, 129.7, 128.8, 128.7 (d, $J_{C-P} = 0.8$ Hz), 128.6, 128.3 (d, $J_{C-P} = 12.1$ Hz), 127.9, 127.8, 124.3, 123.9, 52.3; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.53; HRMS (ESI, m/z): calculated for C₃₅H₂₇NO₃P⁺: 540.1723, found: 540.1722 [M+H]⁺.



4d, Light yellow solid, 61 % yield, ¹**H NMR** (400 MHz, CDCl₃) δ 7.96 (d, J = 2.1 Hz, 1H), 7.76-7.57 (m, 9H), 7.57-7.44 (m, 7H), 7.32-7.18 (m, 6H); ¹³**C NMR** (100 MHz, CDCl₃) δ 158.3(d, $J_{C-P} = 3.8$ Hz), 146.6, 146.3, 145.3 (d, $J_{C-P} = 7.8$ Hz), 137.1, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.6, 132.5, 132.0 (d, $J_{C-P} = 2.4$ Hz), 132.4, 131.6 (d, $J_{C-P} = 9.6$ Hz), 131.4, 131.2, 131.0 (d, $J_{C-P} = 2.6$ Hz), 130.8 (d, $J_{C-P} = 9.3$ Hz), 129.1 (d, $J_{C-P} = 79.2$ Hz), 128.6, 128.2 (d, $J_{C-P} = 12.1$ Hz), 127.8 (d, $J_{C-P} = 12.2$ Hz), 127.5, 126.2, 123.7, 120.5; ³¹**P NMR** (162 MHz, CDCl₃) δ 29.62; **HRMS (ESI, m/z)**: calculated for C₃₃H₂₃BrNOPNa⁺: 582.0593, found: 582.0591 [M+Na]⁺.



4e1, 4e2, Light yellow solid, 67 % yield (1:1.04),¹**H** NMR (400 MHz, CDCl₃) δ 7.77-7.57 (m, 14H), 7.55-7.34 (m, 16H), 7.31-7.14 (m, 14H), 2.39 (s, 3H), 2.35 (s, 3H), 2.33 (s, 3H), 1.82 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.8 (d, $J_{C-P} = 3.7$ Hz), 155.7 (d, $J_{C-P} = 3.8$ Hz), 147.6, 147.2, 146.9, 146.5, 146.0 (d, $J_{C-P} = 8.0$ Hz), 145.4 (d, $J_{C-P} = 8.1$ Hz), 142.8, 139.2, 138.2, 136.3, 135.9, 134.7 (d, $J_{C-P} = 10.9$ Hz), 133.8 (d, $J_{C-P} = 7.5$ Hz), 132.8 (d, $J_{C-P} = 7.4$ Hz), 132.3 (d, $J_{C-P} = 4.4$ Hz), 132.0, 132.0, 131.9 (d, $J_{C-P} = 7.1$ Hz), 131.9 (d, $J_{C-P} = 2.0$ Hz), 131.7 (d, $J_{C-P} = 3.7$ Hz), 131.6 (d, $J_{C-P} = 3.8$ Hz), 131.3 (d, $J_{C-P} = 4.6$ Hz), 130.9, 130.8, 130.8, 130.8, 130.8, 130.7, 129.5, 128.9, 128.5 (d, $J_{C-P} = 17.9$ Hz), 128.1, 128.0, 128.0, 127.9. 127.7 (d, $J_{C-P} = 1.6$ Hz), 127.6 (d, $J_{C-P} = 1.5$ Hz), 127.4, 127.2, 125.2, 125.0, 124.4, 123.5, 122.1, 21.0, 20.2, 20.2, 20.2; ³¹P NMR (162 MHz, CDCl₃) δ 29.90,29.77; **HRMS (ESI, m/z):** calculated for C₃₅H₂₈NOPNa⁺: 532.1801, found: 532.1802 [M+Na]⁺.



4f, Light yellow solid, 63 % yield, ¹H NMR (400 MHz, CDCl₃) δ 7.79 (*d*, J = 2.2 Hz, 1H), 7.73-7.57 (m, 9H), 7.56-7.45 (m, 7H), 7.31-7.17 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 158.2(d, *J*_{*C-P*} = 3.9 Hz), 146.7, 146.1, 145.4 (d, *J*_{*C-P*} = 7.8 Hz), 137.2, 134.7 (d, *J*_{*C-P*} = 11.0 Hz), 133.6, 132.5, 132.4, 132.3, 132.1 (d, *J*_{*C-P*} = 2.3 Hz), 131.6 (d, *J*_{*C-P*} = 9.6 Hz), 131.4, 131.1, 131.0 (d, *J*_{*C-P*} = 2.6 Hz), 130.8 (d, *J*_{*C-P*} = 9.3 Hz), 130.0, 129.1 (d, *J*_{*C-P*} = 79.3 Hz), 128.6, 128.2 (d, *J*_{*C-P*} = 12.0 Hz), 127.8 (d, *J*_{*C-P*} = 12.1 Hz), 125.7, 124.2, 123.8; ³¹P NMR (162 MHz, CDCl₃) δ 29.62; HRMS (ESI, m/z): calculated for C₃₃H₂₃CINOPNa⁺: 538.1098, found: 538.1099 [M+Na]⁺.



4g, Light yellow solid, 44 % yield, ¹**H NMR** (400 MHz, CDCl₃) δ 7.80 (dd, J = 13.6, 7.7 Hz, 1H), 7.68-7.62 (m, 2H), 7.62-7.54 (m, 4H), 7.53-7.41 (m, 6H), 7.37 (s, 1H) 7.26-7.09 (m, 7H), 6.91(d, J = 2.7 Hz ,1H), 3.74 (s, 3H), 2.51 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 157.2, 154.7 (d, $J_{C-P} = 3.8$ Hz), 146.3 (d, $J_{C-P} = 8.7$ Hz), 146.1, 143.3, 139.6, 138.6, 134.6 (d, $J_{C-P} = 10.6$ Hz), 133.7, 132.7, 131.9 (d, $J_{C-P} = 2.5$ Hz), 131.6 (d, $J_{C-P} = 9.7$ Hz), 131.1 (d, $J_{C-P} = 9.6$ Hz), 131.1, 130.7 (d, $J_{C-P} = 2.7$ Hz), 129.4, 128.5, 128.0, 127.7 (d, $J_{C-P} = 11.8$ Hz), 127.7 (d, $J_{C-P} = 12.1$ Hz), 126.0, 123.4, 121.4, 55.2, 18.5; ³¹**P NMR** (162 MHz, CDCl₃) δ 29.34; **HRMS (ESI, m/z):** calculated for C₃₅H₂₉NO₂P⁺: 526.1930, found: 526.1930 [M+H]⁺.



4h, yellow solid, 25 % yield, ¹**H NMR** (400 MHz, CDCl₃) δ 7.82 (d, J = 7.8 Hz, 1H), 7.76-7.55 (m, 10H), 7.53-7.37 (m, 7H), 7.25-7.15 (m, 5H); ¹³**C NMR** (100 MHz, CDCl₃) δ 157.9 (d, $J_{C-P} = 4.2$ Hz), 147.6 (d, $J_{C-P} = 29.4$ Hz), 137.8, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.6, 132.5, 132.1 (d, $J_{C-P} = 2.5$ Hz), 131.9 (d, $J_{C-P} = 100.9$ Hz), 131.7 (d, $J_{C-P} = 9.6$ Hz), 131.4, 130.9, 130.9, 130.8, 129.7, 129.3 (d, $J_{C-P} = 43.4$ Hz), 128.5, 128.3, 128.1 (d, $J_{C-P} = 11.9$ Hz), 127.7 (d, $J_{C-P} = 12.2$ Hz), 126.4, 125.4, 125.0, 123.1; ³¹**P NMR** (162 MHz, CDCl₃) δ 29.69; **HRMS (ESI, m/z)**: calculated for C₃₃H₂₄N₂O₃P⁺; 527.1519, found: 527.1516 [M+H]⁺.



4i, Light yellow solid, 12 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 8.81 (d, *J*= 2.5 Hz, 1H), 8.35 (dd, *J* = 9.2,2.5 Hz, 1H), 7.86 (s, 1H), 7.78-7.41 (m, 16H), 7.36-7.28 (m, 2H), 7.27-7.20 (m, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 161.5(d, *J*_{C-P} = 3.8 Hz), 149.7 (d, *J*_{C-P} = 23.0 Hz), 145.4, 144.9 (d, *J*_{C-P} = 7.5 Hz), 136.4, 134.8 (d, *J*_{C-P} = 11.3 Hz), 133.5, 132.6, 132.5, 132.2 (d, *J*_{C-P} = 2.4 Hz), 131.6 (d, *J*_{C-P} = 9.6 Hz), 131.2, 131.2, 130.9 (d, *J*_{C-P} = 9.2 Hz), 129.3 (d, *J*_{C-P} = 55.1 Hz), 129.2, 128.7, 128.6 (d, *J*_{C-P} = 12.1 Hz), 128.4, 128.0 (d, *J*_{C-P} = 12.2 Hz), 124.7, 124.1, 122.6 (d, *J*_{C-P} = 18.7 Hz); ³¹**P** NMR (162 MHz, CDCl₃) δ 29.74; HRMS (ESI, m/z): calculated for C₃₃H₂₅NOPH⁺: 482.1668, found: 482.1666 [M+H]⁺.



4j, Light yellow solid, 67 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.74-7.56 (m, 9H), 7.55-7.41 (m, 7H), 7.39-7.32 (td, J = 9.0, 2.8 Hz, 1H), 7.31-7.18 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 161.9, 159.4, 157.4 (t, $J_{C-P} = 3.3$ Hz), 147.0 (d, $J_{C-P} = 5.5$ Hz), 145.5 (d, $J_{C-P} = 8.0$ Hz), 144.8, 137.4, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.2 (d, $J_{C-F} = 104.9$ Hz), 132.1 (d, $J_{C-P} = 2.5$ Hz), 132.0 (d, $J_{C-P} = 9.1$ Hz), 131.9 (d, $J_{C-P} = 100.7$ Hz), 131.7 (d, $J_{C-P} = 9.6$ Hz), 131.0 (d, $J_{C-P} = 2.8$ Hz), 130.9 (d, $J_{C-P} = 9.4$ Hz), 129.1 (d, $J_{C-F} = 73.3$ Hz), 128.5, 128.1 (d, $J_{C-F} = 12.0$ Hz), 127.8 (d, $J_{C-F} = 12.1$ Hz), 125.9 (d, $J_{C-P} = 9.6$ Hz), 123.6, 119.3 (d, $J_{C-F} = 25.5$ Hz), 108.8 (d, $J_{C-P} = 23.0$ Hz); ³¹P NMR (162 MHz, CDCl₃) δ 29.59; ¹⁹F NMR (376 MHz, CDCl₃) δ -112.66. HRMS (ESI, m/z): calculated for C₃₃H₂₃FNOPNa⁺: 522.1394, found: 522.1395 [M+Na]⁺.



4k, Light yellow solid, 48 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.89-7.76 (m, 3H), 7.75-7.57 (m, 8H), 7.55 (d, J = 9.0 Hz, 1H), 7.52-7.37 (m, 7H), 7.23-7.04 (m, 7H); ¹³C NMR (100 MHz, CDCl₃) δ 156.4 (d, $J_{C-P} = 3.8$ Hz), 148.6, 147.4, 145.2 (d, $J_{C-P} = 8.1$ Hz), 142.2, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.6, 132.8, 132.6, 132.5, 132.1 (d, $J_{C-P} = 2.3$ Hz), 131.7, 131.6, 131.5, 131.1, 131.0 (d, $J_{C-P} = 9.4$ Hz), 130.9 (d, $J_{C-P} = 2.8$ Hz), 129.4, 129.1, 128.4, 128.2 (d, $J_{C-P} = 12.5$ Hz), 128.1 (d, $J_{C-P} = 12.6$ Hz), 127.8 (d, $J_{C-P} = 12.2$ Hz), 126.4, 125.7, 125.2, 122.3; ³¹P NMR (162 MHz, CDCl₃) δ 29.67; HRMS (ESI, m/z): calculated for C₃₇H₂₆NOPNa⁺: 554.1644, found: 554.1646 [M+Na]⁺.



41, Light yellow solid, 69 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.93 (t, J = 9.2 Hz, 2H), 7.81 (dd, J = 13.7, 7.7 Hz, 1H), 7.76-7.69 (m, 3H), 7.67-7.44 (m, 8H), 7.42-7.26 (m, 7H), 7.23-7.15 (m, 3H), 7.01 (s, 1H), 2.24 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.8 (d, $J_{C-P} = 3.4$ Hz), 145.9 (d, $J_{C-P} = 38.3$ Hz), 145.5 (d, $J_{C-P} = 7.6$ Hz), 136.0 (d, $J_{C-P} = 53.9$ Hz), 135.1 (d, $J_{C-P} = 10.8$ Hz), 134.2 (d, $J_{C-P} = 3.5$ Hz), 133.3, 133.2 (d, $J_{C-P} = 3.8$ Hz), 132.1, 131.9, 131.9, 131.8 (d, $J_{C-P} = 2.6$ Hz), 131.7, 131.6 (d, $J_{C-P} = 5.8$ Hz), 131.1, 131.0, 131.0 (d, $J_{C-P} = 2.6$ Hz), 130.7 (d, $J_{C-P} = 2.6$ Hz), 128.8 (d, $J_{C-P} = 58.8$ Hz), 128.1, 128.0, 127.9, 127.7 (d, $J_{C-P} = 6.0$ Hz), 127.4 (d, $J_{C-P} = 30.6$ Hz), 126.3, 126.1 (d, $J_{C-P} = 10.3$ Hz), 125.6 (d, $J_{C-P} = 69.4$ Hz), 124.5, 123.3, 21.6; ³¹P NMR (162 MHz, CDCl₃) δ 30.53; **HRMS (ESI, m/z):** calculated for C₃₈H₂₉NOP⁺**:** 546.1981, found: 546.1918 [M+H]⁺.



4m, Light yellow solid, 64 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.73-7.57 (m, 11H), 7.51-7.38 (m, 5H), 7.27-7.15 (m, 6H), 2.44 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 156.9 (d, $J_{C-P} = 3.8$ Hz), 145.8 (d, $J_{C-P} = 100.1$ Hz), 145.7 (d, $J_{C-P} = 8.0$ Hz), 136.9, 136.5, 134.6 (d, $J_{C-P} = 11.1$ Hz), 133.0 (d, $J_{C-P} = 104.8$ Hz), 132.2, 132.1 (d, $J_{C-P} = 2.4$ Hz), 131.6, 131.6, 131.5, 131.4, 131.2, 130.9 (d, $J_{C-P} = 9.2$ Hz), 130.9 (d, $J_{C-P} = 2.6$ Hz), 129.3, 128.0 (d, $J_{C-P} = 12.0$ Hz), 127.7 (d, $J_{C-P} =$ 12.1 Hz), 124.6, 123.6, 123.1, 122.5, 21.8; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.74; HRMS (ESI, m/z): calculated for C₃₄H₂₅BrNOPNa⁺: 596.0749, found: 596.0750 [M+Na]⁺.



4n, Light yellow solid, 42 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 8.51-8.28 (m, 2H), 7.84-7.37 (m, 15H), 7.31-7.20 (m, 5H), 2.46 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.9 (d, $J_{C-P} =$ 4.0 Hz), 147.6, 146.4, 145.5 (d, $J_{C-P} =$ 8.0 Hz), 144.3 (d, $J_{C-P} =$ 97.8 Hz), 137.1, 134.5 (d, $J_{C-P} =$ 11.4 Hz), 133.3, 132.2 (d, $J_{C-P} =$ 4.3 Hz), 132.2 (d, $J_{C-P} =$ 2.3 Hz), 131.7, 131.5 (d, $J_{C-P} =$ 9.6 Hz), 131.2, 131.1, 131.1, 131.0, 130.6, 129.5, 128.1 (d, $J_{C-P} =$ 12.1 Hz), 127.8 (d, $J_{C-P} =$ 12.1 Hz), 124.1, 123.7, 123.3 (d, $J_{C-P} =$ 31.3 Hz), 21.8; ³¹P NMR (162 MHz, CDCl₃) δ 29.75; HRMS (ESI, m/z): calculated for C₃₄H₂₆N₂O₃P⁺: 541.1676, found: 541.1674 [M+H]⁺.



40, Light yellow solid, 65 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.75-7.53 (m, 10H), 7.51-7.44 (m, 3H), 7.40 (dd, J = 8.6, 1.7 Hz, 1H), 7.26-7.14 (m, 6H), 7.05 (d, J = 8.7 Hz, 2H), 3.89(s, 3H), 2.43(s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 159.6, 156.9 (d, $J_{C-P} = 4.0$ Hz), 146.4 (d, $J_{C-P} = 2.8$ Hz), 145.9 (d, $J_{C-P} = 8.2$ Hz), 136.1, 134.6 (d, $J_{C-P} = 10.9$ Hz), 133.1 (d, $J_{C-P} = 105.1$ Hz), 132.3, 132.3, 132.0 (d, $J_{C-P} = 2.3$ Hz), 131.6 (d, $J_{C-P} = 9.6$ Hz), 131.3, 131.2, 130.9, 130.9, 130.8 (d, $J_{C-P} = 2.9$ Hz), 130.3, 129.2, 127.9 (d, $J_{C-P} = 12.0$ Hz), 127.7 (d, $J_{C-P} = 12.1$ Hz), 125.1, 123.5 (d, $J_{C-P} = 110.9$ Hz), 113.9, 55.3, 21.8; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.73; HRMS (ESI, m/z): calculated for C₃₅H₂₉NO₂P⁺: 526.1930, found: 526.1930 [M+H]⁺.



4p, Light yellow solid, 50 % yield, ¹**H NMR** (400 MHz, CDCl₃) δ 7.74-7.53 (m, 10H), 7.51-7.45 (m, 1H), 7.41 (dd, J = 8.6, 1.7 Hz, 1H), 7.27-7.15 (m, 6H), 7.12-7.00 (m, 3H), 3.98 (s, 3H), 3.96 (s, 3H), 2.44 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 157.0 (d, $J_{C-P} = 3.9$ Hz), 148.9 (d, $J_{C-P} =$ 25.1 Hz), 146.5 (d, $J_{C-P} = 11.8$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 136.1, 134.7 (d, $J_{C-P} = 10.9$ Hz), 133.2 (d, $J_{C-P} = 105.0$ Hz), 132.3, 132.0 (d, $J_{C-P} = 2.3$ Hz), 131.6 (d, $J_{C-P} = 9.7$ Hz), 131.3, 131.2, 130.9, 130.8, 130.8, 130.6, 129.2, 127.9 (d, $J_{C-P} = 11.9$ Hz), 127.7 (d, $J_{C-P} = 12.1$ Hz), 125.1, 124.1, 122.9, 122.2, 112.9, 111.0, 56.0, 55.9, 21.8; ³¹**P NMR** (162 MHz, CDCl₃) δ 29.71; **HRMS** (**ESI, m/z):** calculated for C₃₆H₃₁NO₃P⁺**:** 556.2036, found: 556.2037 [M+H]⁺.



4q, Light yellow solid, 58 % yield, ¹H NMR (400 MHz, CDCl₃) & 8.98 (s, 1H), 8.23 (d,

J=8.3 Hz, 1H), 7.75 (dd, *J* = 7.2,3.3 Hz,1H), 7.71-7.64 (m, 3H), 7.61 (d, *J* = 8.6 Hz, 1H), 7.54-7.22 (m, 14H), 7.21-7.13 (m, 2H), 2.40 (s, 3H), 2.07 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 156.5 (d, *J*_{*C-P*} = 4.1 Hz), 146.6 (d, *J*_{*C-P*} = 7.3 Hz), 146.1, 142.3, 136.5 (d, *J*_{*C-P*} = 49.3 Hz), 133.4 (d, *J*_{*C-P*} = 2.2 Hz), 133.3, 132.6 (d, *J*_{*C-P*} = 41.2 Hz), 132.2 (d, *J*_{*C-P*} = 2.4 Hz), 131.9, 131.8, 131.7, 131.6, 131.5 (d, *J*_{*C-P*} = 2.8 Hz), 131.3 (d, *J*_{*C-P*} = 10.0 Hz), 130.9 (d, *J*_{*C-P*} = 9.4 Hz), 130.0, 129.1, 128.9 (d, *J*_{*C-P*} = 46.1 Hz), 128.5 (d, *J*_{*C-P*} = 12.0 Hz), 128.1 (d, *J*_{*C-P*} = 12.3 Hz), 127.6 (d, *J*_{*C-P*} = 12.5 Hz), 125.9, 124.9 (d, *J*_{*C-P*} = 93.8 Hz), 123.9 (d, *J*_{*C-P*} = 49.3 Hz), 24.0, 21.6; ³¹P NMR (162 MHz, CDCl₃) δ 30.87; HRMS (ESI, m/z): calculated for C₃₆H₂₉N₂O₂PNa⁺: 575.1859, found: 575.1860 [M+Na]⁺.



4r, Light yellow solid, 69 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.98 (d, J = 8.4 Hz, 1H), 7.95-7.89 (m, 3H), 7.81-7.69 (m, 2H), 7.69-7.39 (m, 13H), 7.30-7.17 (m, 6H), 2.40 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 157.0(d, $J_{C-P} = 3.8$ Hz), 146.5 (d, $J_{C-P} = 47.3$ Hz), 145.7 (d, $J_{C-P} = 8.1$ Hz), 136.4, 135.5, 134.7 (d, $J_{C-P} = 10.8$ Hz), 133.8, 133.1, 132.8 (d, $J_{C-P} = 12.1$ Hz), 132.3, 132.0 (d, $J_{C-P} = 2.3$ Hz), 131.7 (d, $J_{C-P} = 9.7$ Hz), 131.4, 131.3, 130.9, 130.9 (d, $J_{C-P} = 2.7$ Hz), 129.2, 128.7, 128.2, 128.0, 127.9, 127.8, 127.7, 127.6, 127.4, 126.4 (d, $J_{C-P} = 6.6$ Hz), 125.1, 124.1, 123.1, 21.8 ; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.93; HRMS (ESI, m/z): calculated for C₃₈H₂₉NOP ⁺: 546.1981, found: 546.1981 [M+H]⁺.



4s, Light yellow solid, 37 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.77-7.59 (m, 7H), 7.58-7.43 (m, 9H), 7.41 (dd, J = 8.5, 1.9 Hz, 1H), 7.26-7.16 (m, 5H) , 2.43 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.0 (d, $J_{C-P} = 3.8$ Hz), 146.5 (d, $J_{C-P} = 41.5$ Hz), 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.1, 136.3, 134.7 (d, $J_{C-P} = 11.0$ Hz), 133.7, 132.7, 132.3, 132.0 (d, $J_{C-P} = 2.5$ Hz), 131.7 (d, $J_{C-P} = 9.7$ Hz), 131.1 (d, $J_{C-P} = 42.0$ Hz), 131.0, 130.9, 129.6, 129.2, 128.4, 128.1, 127.9 (d, $J_{C-P} = 12.0$ Hz), 127.7 (d, $J_{C-P} = 12.2$ Hz), 124.9, 124.1, 123.0, 21.8; ³¹P NMR (162 MHz, CDCl₃) δ 29.80; HRMS (ESI, m/z): calculated for C₃₄H₂₈BNO₃P⁺: 540.1894, found: 540.1897 [M+H]⁺.



4t, Light yellow solid, 68 % yield, ¹H NMR (400 MHz, CDCl₃) δ 7.78 (dd, J = 13.9,7.4 Hz, 1H), 7.74-7.63 (m, 4H), 7.62-7.55 (m, 2H), 7.52-7.43 (m, 3H), 7.40-7.27 (m, 7H), 7.27-7.22 (m, 1H), 7.20-7.11 (m, 3H), 7.07(s, 1H), 2.36(s, 3H), 1.97(s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.8 (d, $J_{C-P} = 3.5$ Hz), 146.5 (d, $J_{C-P} = 157.3$ Hz), 145.6 (d, $J_{C-P} = 7.5$ Hz), 137.6, 136.2 (d, $J_{C-P} = 29.8$ Hz), 135.0 (d, $J_{C-P} = 10.8$ Hz), 134.2 (d, $J_{C-P} = 7.7$ Hz), 133.1 (d, $J_{C-P} = 7.7$ Hz), 131.9 (d, $J_{C-P} = 9.6$ Hz), 131.9 (d, $J_{C-P} = 2.5$ Hz), 131.6 (d, $J_{C-P} = 110.0$ Hz), 131.6 (d, $J_{C-P} = 9.7$ Hz), 131.4, 131.1 (d, $J_{C-P} = 13.6$ Hz), 131.0 (d, $J_{C-P} = 0.9$ Hz), 130.8 (d, $J_{C-P} = 2.7$ Hz), 129.7 (d, $J_{C-P} = 48.1$ Hz), 128.6 (d, $J_{C-P} = 100.4$ Hz), 128.0, 127.9, 127.7 (d, $J_{C-P} = 3.8$ Hz), 127.6, 125.6, 124.9 (d, $J_{C-P} = 133.9$ Hz), 124.2, 21.7, 20.1; ³¹P NMR (162 MHz, CDCl₃) δ 30.51; HRMS (ESI, m/z): calculated for C₃₅H₂₉NOP⁺: 510.1981, found: 510.1982 [M+H]⁺.



4u, Light yellow solid, 68 % yield, ¹H NMR (400 MHz, CDCl₃) δ 7.77 (dd, J = 13.5, 8.0 Hz, 1H), 7.70-7.57 (m, 6H), 7.56-7.46 (m, 4H), 7.43-7.35 (m, 2H), 7.29-7.23 (m, 5H), 7.22-7.15 (m, 4H), 2.45 (s, 3H), 2.43(s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.0 (d, $J_{C-P} = 3.8$ Hz), 146.6 (d, $J_{C-P} = 79.4$ Hz), 145.7 (d, $J_{C-P} = 8.2$ Hz), 138.1, 138.0, 136.2, 134.7 (d, $J_{C-P} = 10.7$ Hz), 133.8, 132.8, 132.3, 132.0 (d, $J_{C-P} = 2.3$ Hz), 131.7 (d, $J_{C-P} = 9.7$ Hz), 131.3, 131.1 (d, $J_{C-P} = 42.2$ Hz), 130.9, 130.8, 130.1, 129.0 (d, $J_{C-P} = 27.6$ Hz), 128.3, 127.9 (d, $J_{C-P} = 12.0$ Hz), 127.7 (d, $J_{C-P} = 12.2$ Hz), 126.7, 125.0, 124.1, 122.7, 21.8, 21.5; ³¹P NMR (162 MHz, CDCl₃) δ 29.88; HRMS (ESI, m/z): calculated for C₃₅H₂₉NOP⁺: 510.1981, found: 510.1982 [M+H]⁺.



4v, Light yellow solid, 69 % yield, ¹**H NMR** (400 MHz, CDCl₃) δ 7.75 (dd, J = 13.7, 7.6 Hz, 1H), 7.70-7.53 (m, 8H), 7.52-7.45 (m, 2H), 7.41-7.36 (m, 3H), 7.32 (d, J=8.0 Hz, 2H), 7.27-7.14 (m, 6H), 2.45(s, 3H), 2.42 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ 156.9 (d, $J_{C-P} = 3.8$ Hz), 146.8, 146.3, 145.8 (d, $J_{C-P} = 8.1$ Hz), 138.0, 135.6 (d, $J_{C-P} = 102.5$ Hz), 134.7 (d, $J_{C-P} = 10.7$ Hz), 133.7, 132.7, 132.3, 132.0 (d, $J_{C-P} = 2.4$ Hz), 131.7 (d, $J_{C-P} = 9.6$ Hz), 131.3, 131.2, 130.9, 130.8, 130.8, 129.3 (d, $J_{C-P} = 35.3$ Hz), 129.2, 127.9 (d, $J_{C-P} = 11.9$ Hz), 127.6 (d, $J_{C-P} = 12.1$ Hz), 125.0, 123.5 (d, $J_{C-P} = 126.2$ Hz), 21.8, 21.2; ³¹**P NMR** (162 MHz, CDCl₃) δ 29.78; **HRMS (ESI, m/z):** calculated for C₃₅H₂₉NOP⁺: 510.1981, found: 510.1982 [M+H]⁺.



4w, Light yellow solid, 53 % yield, ¹H NMR (400 MHz, CDCl₃) δ 7.93-7.27 (m, 18H), 7.25-

6.84 (br, 4H), 2.46 (s, 3H), 2.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.1 (d, $J_{C-P} = 4.8$ Hz), 146.2 (d, $J_{C-P} = 3.1$ Hz), 144.5 (d, $J_{C-P} = 8.9$ Hz), 137.9, 137.8 (d, $J_{C-P} = 9.7$ Hz), 136.3, 134.4 (d, $J_{C-P} = 2.5$ Hz), 133.1, 132.5, 132.1, 131.8 (d, $J_{C-P} = 9.8$ Hz), 131.5 (d, $J_{C-P} = 11.3$ Hz), 131.5, 130.9, 129.8, 129.2, 128.4, 128.1, 127.8, 127.7, 124.8, 124.5, 124.0, 21.8, 20.4; ³¹P NMR (162 MHz, CDCl₃) δ 28.91; HRMS (ESI, m/z): calculated for C₃₅H₂₉NOP⁺: 510.1981, found: 510.1982 [M+H]⁺.



4x, Light yellow solid, 65 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.70 (d, J = 13.9 Hz, 1H), 7.67-7.59 (m, 4H), 7.56 (dd, J = 7.8, 4.6Hz, 1H), 7.53-7.36 (m, 10H), 7.27-7.21 (m, 2H), 7.20-7.12 (m, 4H), 2.42 (s, 3H), 2.40 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.1 (d, $J_{C-P} = 3.6$ Hz), 146.5 (d, $J_{C-P} = 8.2$ Hz), 142.7 (d, $J_{C-P} = 8.2$ Hz), 138.0, 137.9, 136.1, 135.2 (d, $J_{C-P} = 10.2$ Hz), 133.8, 132.8, 132.6 (d, $J_{C-P} = 2.5$ Hz),132.0, 131.8 (d, $J_{C-P} = 9.8$ Hz), 131.3, 131.0, 130.8, 130.8, 130.7, 129.0 (d, $J_{C-P} = 110.8$ Hz), 128.6 (d, $J_{C-P} = 97.4$ Hz), 127.6 (d, $J_{C-P} = 12.2$ Hz), 124.8, 124.0, 122.8, 21.8, 21.3; ³¹P NMR (162 MHz, CDCl₃) δ 30.48; HRMS (ESI, m/z): calculated for C₃₅H₂₉NOP⁺: 510.1981, found: 510.1982 [M+H]⁺.



4y, Light brown solid, 50 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 8.01-7.86 (m, 4H), 7.81(dd, J = 11.7, 8.6 Hz, 1H), 7.71-7.64 (m, 2H), 7.60-7.49 (m, 6H), 7.49-7.31 (m, 9H), 7.02-6.74 (m, 3H), 2.48 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 155.3 (d, $J_{C-P} = 5.5$ Hz), 146.2 (d, $J_{C-P} = 16.5$ Hz), 144.3 (d, $J_{C-P} = 8.9$ Hz), 137.8, 136.6, 135.0 (d, $J_{C-P} = 2.1$ Hz), 132.3, 132.2, 131.4, 131.2, 131.1 (d, $J_{C-P} = 8.6$ Hz), 130.4, 129.8, 129.4, 128.8, 128.4, 128.3 (d, $J_{C-P} = 5.7$ Hz), 128.2, 128.1, 128.1, 128.0, 127.9, 127.2 (d, $J_{C-P} = 12.8$ Hz), 127.0 (d, $J_{C-P} = 3.9$ Hz), 125.3, 125.1, 124.1, 21.8; ³¹**P** NMR (162 MHz, CDCl₃) δ 29.26; HRMS (ESI, m/z):calculated for C₃₈H₂₈NOP Na⁺: 568.1801, found: 568.1801 [M+Na]⁺.



4z, Light yellow solid, 63 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 8.61 (d, J = 12.8 Hz, 1H), 8.51-8.42 (m, 1H), 8.09 (d, J = 8.6 Hz, 1H), 7.79-7.68 (m, 5H), 7.64 (s, 1H), 7.62-7.57 (m, 2H), 7.56-7.43 (m, 12H), 2.45 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.6, 148.7, 147.2, 140.2 (d, $J_{C-P} = 11.7$ Hz), 138.2, 136.6, 133.4, 132.9, 132.5, 132.1 (d, $J_{C-P} = 49.7$ Hz), 132.1 (d, $J_{C-P} = 9.9$ Hz), 131.9 (d, $J_{C-P} = 2.7$ Hz), 131.8, 131.0 (d, $J_{C-P} = 2.6$ Hz), 130.9 (d, $J_{C-P} = 9.8$ Hz), 129.8, 129.0 (d, $J_{C-P} = 89.7$ Hz), 128.8 (d, $J_{C-P} = 12.5$ Hz), 128.4 (d, $J_{C-P} = 6.7$ Hz), 128.3, 125.8, 124.3, 119.2,

21.8; ³¹P NMR (162 MHz, CDCl₃) δ 29.46; HRMS (ESI, m/z): calculated for C₃₄H₂₇NOP⁺: 496.1825, found: 496.1827 [M+H]⁺.



4aa, Light yellow solid, 58 % yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.66-7.55 (m, 5H), 7.55-7.42 (m, 7H),7.40 (dd, J = 8.6, 1.7 Hz, 1H), 7.27-7.09 (m, 8H), 6.05 (s, 2H), 2.42 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 156.5 (d, $J_{C-P} = 4.1$ Hz), 150.4 (d, $J_{C-P} = 2.6$ Hz), 147.4 (d, $J_{C-P} = 17.7$ Hz), 146.5 (d, $J_{C-P} = 38.2$ Hz), 141.7 (d, $J_{C-P} = 9.0$ Hz), 138.0, 136.2, 133.2 (d, $J_{C-P} = 105.8$ Hz), 131.6 (d, $J_{C-P} = 9.6$ Hz), 131.3, 130.8 (d, $J_{C-P} = 2.7$ Hz), 129.6, 129.1, 128.4, 128.1, 127.7 (d, $J_{C-P} = 12.2$ Hz), 125.7, 124.9, 124.7, 124.0, 123.1, 114.0 (d, $J_{C-P} = 13.3$ Hz), 111.5 (d, $J_{C-P} = 12.1$ Hz), 101.9, 21.7; ³¹P NMR (162 MHz, CDCl₃) δ 29.45; HRMS (ESI, m/z): calculated for C₃₅H₂₇NO₃P +: 540.1723, found: 540.1726 [M+H]⁺.



4ab, light yellow solid, 63% yield, ¹**H** NMR (400 MHz, CDCl₃) δ 7.78-7.68 (m, 1H), 7.66-7.37 (m, 14H), 7.29-7.14 (m, 7H), 2.43 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ 165.8 (d, $J_{C-P} =$ 2.8 Hz), 163.3 (d, $J_{C-P} =$ 3.0 Hz), 155.6 (dd, $J_{C-F} =$ 1.7 Hz, $J_{C-P} =$ 3.4 Hz), 148.6 (dd, $J_{C-F} =$ 7.9 Hz, $J_{C-P} =$ 9.6 Hz), 146.9, 146.2, 137.8, 137.3 (dd, $J_{C-F} =$ 8.9 Hz, $J_{C-P} =$ 12.3 Hz), 136.6, 133.3, 132.2, 131.6, 131.5, 131.0 (d, $J_{C-P} =$ 2.6 Hz), 129.0 (d, $J_{C-P} =$ 105.9 Hz), 128.7 (d, $J_{C-P} =$ 89.1 Hz), 127.8 (d, $J_{C-P} =$ 12.2 Hz), 127.4 (d, $J_{C-P} =$ 3.3 Hz), 125.0, 124.0, 122.7, 118.3 (dd, $J_{C-F} =$ 10.6 Hz, $J_{C-P} =$ 21.9 Hz), 115.0 (dd, $J_{C-F} =$ 13.0 Hz, $J_{C-P} =$ 20.4 Hz), 21.8; ³¹P NMR (162 MHz, CDCl₃) δ 28.92; ¹⁹F NMR (376 MHz, CDCl₃) δ -106.54; HRMS (ESI, m/z): calculated for C₃₄H₂₆FNOP⁺: 514.1731, found: 514.1731 [M+H]⁺.



4ac, yellow solid, 37% yield, ¹**H NMR** (400 MHz, CDCl₃) δ 8.31 (ddd, J = 10.7, 5.6, 3.5 Hz, 1H), 8.06 (d, J = 8.5 Hz, 1H), 7.75 (s, 1H), 7.65 (dd, J = 8.5, 1.4 Hz, 1H), 7.61-7.50 (m, 8H), 7.47(s, 1H), 2.53 (s, 3H), 1.89-1.53 (m, 13H), 1.35-1.23 (m, 3H), 1.19-1.09 (m, 2H), 0.99-0.90 (m, 4H); ¹³C **NMR** (100 MHz, CDCl₃) δ 158.5 (d, $J_{C-P} = 2.0$ Hz), 148.6, 146.1, 142.8 (d, $J_{C-P} = 8.0$ Hz), 137.9, 137.0, 132.4, 131.4, 130.7 (d, $J_{C-P} = 2.3$ Hz), 130.6, 130.2 (d, $J_{C-P} = 8.8$ Hz), 129.4, 129.0, 128.6, 128.5, 128.2 (d, $J_{C-P} = 9.4$ Hz), 125.3, 124.7, 122.4, 38.7, 38.0, 26.7 (dd, $J_{C-P} = 12.9$, 9.3 Hz), 26.4 (dd, $J_{C-P} = 11.7, 3.5$ Hz), 25.7, 21.9; ³¹P **NMR** (162 MHz, CDCl₃) δ 50.17; **HRMS** (**ESI, m/z**): calculated for C₃₄H₃₉NOP⁺: 508.2764, found: 508.2763 [M+H]⁺.



5w, White solid, 73 % yield, ¹H NMR (300 MHz, CDCl₃) δ 7.87 (d, J = 8.6 Hz, 1H), 7.61 (s, 1H), 7.44 (d, J = 8.5 Hz, 1H), 7.36 – 7.27 (m, 3H), 7.24 –7.08 (m, 14H), 6.92 – 6.83 (m, 2H), 2.38 (s, 3H), 2.10 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 158.3 (d, $J_{C\cdot P} = 5.4$ Hz), 146.9, 146.6, 146.4, 146.0, 138.1, 137.7, 137.5, 136.5, 136.4, 136.3, 134.1, 133.8, 131.5 (d, $J_{C\cdot P} = 5.5$ Hz), 131.0, 129.7, 129.6, 128.3, 128.2, 128.0, 125.1, 124.2, 123.9 (d, $J_{C\cdot P} = 5.5$ Hz), 21.8, 20.4; ³¹P NMR (121 MHz, CDCl₃) δ -12.15; HRMS (ESI, m/z): calculated for C₃₅H₂₉NP⁺: 494.2032, found: 494.2032 [M+H]⁺.



5y, Light yellow solid, 80 % yield, ¹H NMR (300 MHz, CDCl₃) δ 7.98–7.92 (m, 1H), 7.73 (dd, J = 16.2, 7.5 Hz, 3H), 7.49 (t, J = 7.6 Hz, 2H), 7.42 – 7.10 (m, 19H), 2.41 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 157.5 (d, $J_{C-P} = 7.5$ Hz), 147.0, 146.7, 145.6, 145.2, 138.1, 136.6, 133.9, 133.7, 133.6, 132.2, 132.1, 131.6, 129.9, 129.8, 129.6, 128.4, 128.3, 128.1, 127.9, 126.6, 126.5, 126.5, 126.5, 125.3, 124.8, 124.8, 21.8; ³¹P NMR (121 MHz, CDCl₃) δ -11.92; HRMS (ESI, m/z): calculated for C₃₈H₂₉NP⁺: 530.2032, found: 530.2031 [M+H]⁺.

7. References

[1] M. L. Abrams, F. Foarta and C. R. Landis, J. Am. Chem. Soc., 2014, 136, 14583.

[2] A. K. Morri, Y. Thummala and V. R. Doddi, Org. Lett., 2015, 17, 4640.

[3] X. Qi, H. Zhang, Z. Pan, R.-B. Liang, C. Zhu, J. Li, Q. Tong, X. Gao, L. Wu and J. Zhong, *Chem. Commun.*, 2019, **55**, 10848.

[4] V. Ravindar, H. Hemling, H. Schumann and J. Blum, Synth. Commun., 1992, 22, 1453.

[5] W. Ren, Q.-M. Zuo and S.-D. Yang, Synlett, 2019, 30, 1719.

[6] G. London, M. Von, W. Rakowski, O. Dumile, W. B. Schweizer, J.-P. Gisselbrecht, C. Boudon and F. Diederich, *Chem. Sci.*, 2014, **5**, 965.

[7] F. Puls, N. Richter, O. Kataeva and H. J. Knolker, Chem. Eur. J., 2017, 23, 17576.

7. Copies of NMR spectra
































































51

210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)




























































---21.84



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)