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Supporting Information for:

Synthesis and Self-Aggregated Nanostructures of Hydrogen-Bonding Polydimethylsiloxane

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Experimental section

Materials

 α -Mono-carbinol functionalized PDMS (OH-PDMS1, 2 and 3) were purchased from Gelest, Inc. All the other chemicals were purchased from Alfa Aesar or Sigma-Aldrich. Unless otherwise indicated the other chemicals were used without further purification. HW-PDMS1,¹ 2^2 and 3^1 were synthesized according to previous works.

Characterization methods

¹H, ¹³C and ²⁹Si NMR spectra were recorded on Varian Gemini 2000 FT-NMR spectrometer (400 MHz) or Varian unity Inova 500 (500 MHz) NMR spectrometer, using CDCl₃ or DMSOd₆ as solvent. The ¹H T1 relaxation was determined using the inversion recovery method. Polymers were analyzed by size exclusion chromatography (SEC) running in THF at 35°C (flow rate: 1 mL·min⁻¹) and recorded on GPCmax VE 2001 from Viscotek[™], which equipped with a column set of a H_{HR}-H Guard-17369 column, a CLM30111 column and a G2500H_{HR}-17354 column. The average molar masse of polymers was derived from refractive index signal based on polystyrene calibration curve.

Matrix-assisted laser desorption/ionization time of flight mass spectrometry (MALDI-TOF MS) measurements were performed on Bruker Autoflex III system (Bruker Daltonics) operating in linear mode. Data evaluation was carried out on DataAnalysis software. Ions were formed by laser desorption (smart beam laser at 355, 532, 808, and 1064 \pm 5 nm; up to 50 Hz repetition rate), accelerated by a voltage of 20 kV and detected as positive ions. Samples were

prepared by mixing 50 μ L of 2,5-dihydroxybenzoic acid at 20 g·L⁻¹ in THF with 10 μ L of polymer solution at 20 g·L⁻¹ in THF. To enhance cationization of polymers, 1 μ L of NaI at 10 g·L⁻¹ in acetone was added to solutions. Finally, 1 μ L of resulting mixture was spotted on a MALDI sample plate and air-dried.

The solid-state NMR measurements were carried out at a ¹H Larmor frequency of 400 MHz on a Bruker Avance III spectrometer with double- or triple-resonance 4 mm MAS probes. The samples were packed into 4 mm ZrO_2 rotors using Kel-F inserts and Vespel caps and spun at 10 kHz. The temperature regulation was based upon heated pressurized air using a BVT3000 temperature control unit, with an accuracy of around 1K. Fully relaxed ¹H spectra (recycle delay of 5*T*₁) were taken after a 90° pulse of 3.12 µs duration. To obtain qualitative information on mobility, *T*₂ relaxation times were measured using a rotor-synchronized Hahnecho pulse sequence.

Small angle X-ray scattering (SAXS) experiments were performed with the rotating anode from Rigaku company. The wavelength of X-ray (Cu K_a) is $\lambda = 1.54$ Å. The tube voltage and tube current were 40 kv and 60 mA. Through the optics and three pinholes, the X-ray beam become collimated and small sized. The sample holder was an aluminum disc with a hole of diameter of 1 mm in the middle. The sample was placed into the hole, and the aluminum disc was attached onto a Linkam hot stage TMS 94 by thermal conducting paste. Temperature-dependent SAXS measurements were done for all the samples at a broad temperature range, from 20 to maximum 180 °C. The measured *q* range was from 0.04 to 0.4 Å.

Synthesis of TAP-PDMS (Figure S1)



Figure S1: Synthetic route to TAP-PDMS.

In this context, a general procedure for the synthesis of TAP-PDMS1, 2 and 3 (**Figure S1**) was shown via the PDMS-N₃ with Mn = 11.4 kDa. PDMS-N₃ (Mn = 11.4 kDa, 0.15 g, 0.013 mmol) was dissolved in 2 mL toluene, compound 2^2 (0.0025 g, 0.013 mmol) was fully dissolved in 0.75 mL DMF, then the two obtained solutions were mixed together in a round-bottomed flask. Sodium L-ascorbate (0.013 g, 0.065 mmol) was added, and the mixture was degassed by bubbling N₂ for ~30 min. CuSO₄·5H₂O (0.0032 g, 0.013 mmol) was added to the mixture which was degassed again by bubbling N₂ for ~30 min, and then allowed to stir at 70 °C for 15 h. The mixture was cooled, concentrated, dissolved in CH₂Cl₂, and filtered to remove CuSO₄·5H₂O and sodium L-ascorbate, and then the filtrate was purified by column chromatography (eluent: CH₂Cl₂/MeOH, 100/3, v/v) to give TAP-PDMS3 as a yellowish viscous solid (0.13g, yield 75%).

¹H NMR (CDCl₃, **Figure S2**A) δ (ppm): 0.07 (s, ~84H), 0.52 (m, 4H), 0.88 (t, 3H), 1.31 (m, 4H), 1.48 (br, 2H), 1.59 (sext, 2H), 1.85 (m, 2H), 2.44 (br, 2H), 2.86 (br, 2H), 3.42 (t, 2H),

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3.64 (t, 2H), 4.31 (m, 2H), 5.46 (br, 2H), 5.75 (br, 2H), 6.13 (br, 4H), 7.54 (br, 1H).

¹³C NMR (CDCl₃, Figure S2B) δ (ppm): 0.88, 1.02, 10.81, 13.76, 14.07, 17.86, 23.33, 25.41,
26.32, 28.91, 29.69, 30.35, 38.88, 65.32, 68.00, 71.70, 74.07, 86.01, 128.80, 130.94, 132.54,
152.61, 167.97, 169.52.

²⁹Si NMR (CDCl₃, **Figure S2**C) δ (ppm): -22.28.



Figure S2: ¹H (A), ¹³C (B) and ²⁹Si (C) of TAP-PDMS1 recorded in CDCI₃ at 27 °C.

Series	Species	m/z measured	m/z simulated
		g·mol ^{−1}	g·mol⁻¹
1	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₇ C ₆ H ₁₅ Si + H] ⁺	1039.482	1039.469
2	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₇ C ₆ H ₁₅ Si + Li ₂ - H] ⁺	1051.534	1051.486
3	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₇ C ₆ H ₁₅ Si + Na]⁺	1061.509	1061.451
4	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₈ C ₆ H ₁₅ Si + H] ⁺	1113.533	1113.488
5	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₈ C ₆ H ₁₅ Si + Li]⁺	1119.429	1119.496
6	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₉ C ₆ H ₁₅ Si + H] ⁺	1188.106	1188.507
7	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₉ C ₆ H ₁₅ Si + Li]⁺	1193.526	1193.515
8	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₀ C ₆ H ₁₅ Si + H] ⁺	1262.557	1262.526

 Table S2:
 MALDI-TOF MS results of TAP-PDMS2.

Series	Species	m/z measured	m/z simulated
		g·mol ^{−1}	g·mol⁻¹
1	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₆₇ C ₆ H ₁₅ Si + H] ⁺	5489.503	5489.596
2	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₆₇ C ₆ H ₁₅ Si + Na]⁺	5511.497	5511.578
3	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₆₈ C ₆ H ₁₅ Si + H]⁺	5563.536	5563.615
4	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₆₈ C ₆ H ₁₅ Si + Na]⁺	5585.508	5585.597
5	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₆₉ C ₆ H ₁₅ Si + Na]⁺	5659.531	5659.615
6	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₇₀ C ₆ H ₁₅ Si + H] ⁺	5711.574	5711.652
7	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₇₀ C ₆ H ₁₅ Si + Na]⁺	5733.566	5733.634

Table S3: MALDI-TOF MS results of TAP-PDMS3.

Series	Species	m/z measured	m/z simulated
		g·mol⁻¹	g·mol⁻¹
1	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₁₈ C ₆ H ₁₅ Si + H] ⁺	9271.486	9271.553
2	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₁₈ C ₆ H ₁₅ Si + Na]⁺	9293.462	9293.535
3	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₁₉ C ₆ H ₁₅ Si + H] ⁺	9345.503	9345.571
4	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₁₉ C ₆ H ₁₅ Si + Na]⁺	9367.479	9367.553
5	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₂₀ C ₆ H ₁₅ Si + Na]⁺	9441.508	9441.572
6	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₂₁ C ₆ H ₁₅ Si + H] ⁺	9493.517	9493.609
7	[C ₁₈ H ₂₉ N ₈ O ₃ (C ₂ H ₆ OSi) ₁₂₁ C ₆ H ₁₅ Si + Na]⁺	9515.536	9515.591

Synthesis of Ba-PDMS (Figure S3)



Figure S3: Synthetic route to Ba-PDMS.

In this context, a general procedure for the synthesis of Ba-PDMS1, 2 and 3 (Figure S3) was

shown via the PDMS- N₃ with Mn = 11.4 kDa. PDMS-N₃ (0.33 g, 0.26 mmol) and compound **3** (0.91 g, 0.4 mmol) were added to a round-bottomed flask and dissolved in ethanol (5 mL). Sodium L-ascorbate (0.021 g, 0.1 mmol) was added, and the mixture was degassed by bubbling N₂ for ~30 min. CuSO₄·5H₂O (0.0064 g, 0.026 mmol) was added, the flask was degassed again by bubbling N₂ for ~30 min, and then allowed to stir at 60 °C for 15 h. The mixture was cooled, concentrated, dissolved in CH₂Cl₂, and filtered to remove CuSO₄·5H₂O and sodium L-ascorbate, and then the filtrate was purified by column chromatography (eluent: CH₂Cl₂/MeOH, 100/3, v/v) to give Ba-PDMS as a yellowish solid (yield 70%).

¹H NMR (CDCl₃, **Figure S4**A) δ (ppm): 0.07 (s, ~84H), 0.54 (m, 4H), 0.88 (t, 3H), 0.92 (t, 3H), 1.25-1.31 (m, 6H), 1.56-1.68 (m, 4H), 1.82 (d, 3H), 2.03 (m, 4H), 2.74 (br, 3H), 3.45 (t, 2H), 3.67 (t, 2H), 4.34 (m, 2H), 5.46 (q, 1H), 7.43 (s, 1H), 8.56 (br.s, 2H).

¹³C NMR (CDCl₃, Figure S4B) δ (ppm): 0.94, 9.14, 13.75, 13.77, 17.80, 22.99, 25.32, 26.00,
30.99, 32.27, 33.61, 57.02, 57.99, 64.93, 67.90, 73.87, 121.01, 142.18, 147.78, 168.89,
172.16.

²⁹Si NMR (CDCl₃, **Figure S4**C) δ (ppm): -22.38.

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Figure S4: ¹H (A), ¹³C (B) and ²⁹Si (C) of Ba-PDMS1 recorded in CDCl₃ at 27 °C.

MALDI-TOF MS measurement of Ba-PDMS1 and Ba-PDMS3

The structure of Ba-PDMS was further studied via MALDI-TOF MS spectrometry (Figure S5). The mass spectrum of Ba-PDMS1 was observed in the linear mode, Figure S5A was obtained by ionization of chains assisted with 2,5-dihydroxybenzoic acid as matrix and Nal as cationization agent. MALDI-TOF spectrum (Figure S5B) underlined the presence of main series (1) and (13), along with other minor series (2)-(12), the mass difference between the m/z values of molecular ions between series (1), (6), (10) and (13) was equal to ~74.0 g/mol, reflecting the repeating dimethylsiloxane units (calculated 74.02 g/mol). The observation of the main series (1) visible at 1552.533 g/mol (Figure S5C) could be assigned to a Ba functionalized species [C₆H₁₅Si-P(DMS)₁₃-Ba + Na]⁺, i.e., [C₆H₁₅Si-P(C₂H₆OSi)₁₃-C₂₁H₃₄N₅O₆ + Na]⁺, which matched well with simulated pattern ($m/z_{average}$ = 1552.579 g/mol). The observed isotopic patterns of other minor series (2-13), illustrated in Figure S5B, also agreed well with the simulated structures, illustrating the click reaction proceeded effectively, and ascertaining the successful formation of barbiturate functionalized PDMS (Ba-PDMS1). Additionally, the structure of Ba-PDMS3, which possess higher molar mass 11600 g/mol, was also proven by MALDI-TOF MS spectroscopy (Figure S7).



Figure S5: MALDI-TOF MS of Ba-PDMS1 (A) full spectrum, (B) expansion and (C) simulation of the isotope pattern.



Figure S6: MALDI-TOF MS of Ba-PDMS2 (A) full spectrum, (B) expansion and (C) simulation of the isotope pattern.



Figure S7: MALDI-TOF MS of Ba-PDMS3 (A) full spectrum, (B) expansion and (C) simulation of the isotope pattern.

MALDI-TOF MS measurement of HW-PDMS1 and HW-PDMS3

The mass spectrum of HW-PDMS1 (**Figure S8**), acquired in the linear mode, disclosed the series \blacklozenge \bigstar and \Re separated by ~74.0 g/mol mass units, i.e., a dimethylsiloxane repeat unit (**Figure S8**B). The main series \blacklozenge located at 1398.594 g/mol (**Figure S8**C) was corresponding to a HW functionalized species [C₆H₁₅Si-P(DMS)₇-HW + Na]⁺, i.e., [C₆H₁₅Si-

 $P(C_2H_6OSi)_7-C_{37}H_{44}N_9O_8 + Na]^+$, in good agreement with simulated pattern (*m/z* average = 1398.546 g/mol). As expected, the simulated and measured isotopic patterns of the other main and minor species were also well-matched (**Figure S8**B). In a similar manner, the structure of HW-PDMS3 (Mn = 11900 g/mol), was also evidenced by MALDI-TOF MS spectroscopy (**Figure S9**).



Figure S8: MALDI-TOF MS of HW-PDMS1 (A) full spectrum, (B) expansion and (C) simulation of the isotope pattern.



Figure S9: MALDI-TOF MS of HW-PDMS3 (A) full spectrum, (B) expansion and (C) simulation of the isotope pattern.



Figure S10: Evolution of the SEC traces of H-bonding PDMS.

¹H NMR spin-lattice relaxation time T1 calculation.



Figure S11: ¹H NMR spin-lattice relaxation time T1 of Ba-PDMS1, TAP-PDMS1, and HW-PDMS1 at 27 °C.

SAXS profile of HO-PDMS1.



Figure S12: SAXS profile of HO-PDMS1 at 20 °C.

Mathematical model chosen to determine K_a between Ba-PDMS1 and TAP-PDMS1.

Data related to the concentration-dependence of the chemical shift of Ba N*H* proton for a mixture of a Ba-PDMS1 ([Ba-P]= 3×10^{-3} mol·L⁻¹) and TAP-PDMS1 were fitted according to the following equation to provide K_a:

$$\delta_{\text{mixt}} = \delta_{\text{Ba} \rightarrow \text{P}} + \frac{(\delta_{\infty} - \delta_{\text{Ba} \rightarrow \text{P}})(([\text{Ba} - \text{P}] + [\text{TAP} - \text{P}] + \frac{1}{K_a}) - (([\text{Ba} - \text{P}] + [\text{TAP} - \text{P}] + \frac{1}{K_a})^2 - 4[\text{Ba} - \text{P}][\text{TAP} - \text{P}])^{1/2})}{2[\text{Ba} - \text{P}]}$$

where the experimental parameters are: [Ba-P] and [TAP-P], the respective molar concentrations of Ba-PDMS1 and TAP-PDMS1; δ_{mixt} , the measured NH chemical shift for Ba-P and TAP-P mixture; δ_{Ba-P} , the measured NH chemical shift for TAP-P solution. The fitted parameters are: K_a, the association constant and δ_{∞} , the NH chemical shift of the fully associated system.



Figure S13: Concentration-dependence of the chemical shift of Ba NH proton for a mixture of Ba-PDMS1 ([Ba] = 3 x 10⁻³ mol·L⁻¹) and TAP-PDMS1 in CDCl₃ at 27 °C (filled symbols); fit corresponding to K_a = 130 M⁻¹ and δ_{∞} = 13 ppm (curve).



Figure S14: ¹H NMR spectra of Ba-PDMS1 (A), HW-PDMS1 (B), and a stoichiometric Ba-PDMS1/HW-PDMS1 mixture (C) at 27 °C in CDCl₃ ([Ba] = [HW] = 3 mM). Green cross: Ba NH protons, red spot: HW NH protons.

Mathematical model chosen to determine K_a between Ba-PDMS1 and HW-PDMS1.

Data related to the concentration-dependence of the chemical shift of Ba N*H* proton for a mixture of a Ba-PDMS1 ([Ba-P]= 3×10^{-3} mol·L⁻¹) and HW-PDMS1 were fitted according to the following equation to provide K_a:

$$\delta_{\text{mixt}} = \delta_{\text{Ba} \rightarrow P} + \frac{(\delta_{\infty} - \delta_{\text{Ba} \rightarrow P})(([\text{Ba} - P] + [\text{HW} - P] + \frac{1}{K_a}) - (([\text{Ba} - P] + [\text{HW} - P] + \frac{1}{K_a})^2 - 4[\text{Ba} - P][\text{HW} - P])^{1/2})}{2[\text{Ba} - P]}$$

where the experimental parameters are: [Ba-P] and [HW-P], the respective molar

concentrations of Ba-PDMS1 and HW-PDMS1; δ_{mixt} , the measured NH chemical shift for Ba-P and HW-P mixture; δ_{Ba-P} , the measured NH chemical shift for HW-P solution. The fitted parameters are: K_a , the association constant and δ_{∞} , the NH chemical shift of the fully associated system.



Figure S15: Concentration-dependence of the chemical shift of Ba NH proton for a mixture of Ba-PDMS1 ([Ba] = 3 x 10⁻³ mol·L⁻¹) and HW-PDMS1 in CDCl₃ at 27 °C (filled symbols); fit corresponding to $K_a = 4.3 \times 10^4 \text{ M}^{-1}$ and $\delta_{\infty} = 13 \text{ ppm}$ (curve).

Viscosities of H-bonding PDMS.



Figure S16: logarithmic plot of the real part of viscosity vs. temperature

SAXS profiles of H-bonding PDMS.



Figure S17: SAXS profiles of H-bonding PDMS at 20 °C.



Figure S18: SAXS patterns of H-bonding PDMS at 20 °C.

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