## Diphenyl Phosphate/Ethyl Diphenylphosphinite as an Efficient Organocatalytic System for Ring-opening Polymerization of $\varepsilon$ -Caprolactone and $\delta$ -Valerolactone

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## **Experimental Section**

Materials. Dichloromethane  $(CH_2Cl_2,$ ≥99.5%), toluene (>99.5%), methanol (MeOH, >99.5%) and ethyl acetate (≥99.5%) were purchased from Titan Chemical Co., Ltd (Shanghai, China). Triethylamine (>99.0%) were purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China). Ethyl diphenylphosphinite (EDPP, >93.0%) and benzoic acid (PhCO<sub>2</sub>H, >99.0%) were purchased from TCI chemicals.  $\delta$ -Valerolactone ( $\delta$ -VL,  $\geq$ 98.5%) and diphenyl phosphate (DPP, ≥99.7%) were purchased from J&K Scientific Co., Ltd. (Beijing, China). L-Lactide (L-LA,99.0%) and trimethylene carbonate (TMC, 98%) were purchased from Rhawn reagent Co., Ltd. (Shanghai, China). *e*-Caprolactone (*e*-CL, 99%), bis(trifluoromethane) sulfonamide (HNTf<sub>2</sub>, >97.0%), deuterated chloroform (CDCl<sub>3</sub>, 99.8%), and calcium hydride (CaH<sub>2.95%</sub>) were purchased from Aladdin Biochemical Technology Co., Ltd. (Shanghai, China). CH<sub>2</sub>Cl<sub>2</sub>,  $\varepsilon$ -CL,  $\delta$ -VL, and DMF were distilled from CaH<sub>2</sub> and degassed by three freeze-pump-thaw cycles under an argon atmosphere prior to use. Toluene was distilled over Na/benzophenone and degassed by three freeze-pump-thaw cycles under an argon atmosphere prior to use. L-LA was recrystallization in ethyl acetate and dried under vacuum before use. TMC was recrystallization in toluene at -20 °C before use. Other chemical were used as received.

**Measurements.** The polymerizations were conducted in a MIKROUNA stainless steel glove-box full of argon gas equipped with a gas purification system under a dry argon atmosphere (H<sub>2</sub>O, O<sub>2</sub> <0.01 ppm). The moisture and oxygen contents in the glove box were monitored by an MB- MO-SE 1 and MB-OX-SE 1, respectively. The proton, carbon, and phosphorus nuclear magnetic resonance (<sup>1</sup>H, <sup>13</sup>C, and <sup>31</sup>P NMR) spectra were recorded by a

Bruker AVANCE III 400 Hz NMR spectrometer with superconducting magnet, Bruker Biospin GmbH Rheinstetten, Germany. The <sup>31</sup>P NMR spectra were determined using P(O*n*Bu)<sub>3</sub> as an internal standard (139.1 ppm) when there was no acid in the samples. The number-average molecular weights ( $M_{n,SECS}$ ) and dispersities ( $M_w/M_nS$ ) were determined by HLC-8320 GPC at 40 °C in THF (0.35 mL min<sup>-1</sup>) equipped with a refractive index detector (+, 0.5 s) and two TSKgel Super Multipore HZ-M columns (4.6 mm I.D. × 15 cm × 2), to which 10 µL of sample is injected with the concentration of 0.2-0.3 wt% at once, calculated on the basis of a polystyrene calibration. Matrix-assisted laser desorption ionization-time of flight (MALDI-ToF) mass spectra were collected on a Bruker UltraFLEX MALDI-ToF in reflector mode with potassium trifluoroacetate as the cationic agent and 2,5-dihydroxybenzoic acid (DHB) as the matrix.

**Ring-opening polymerization of**  $\varepsilon$ -caprolactone. A typical procedure for polymerization of  $\varepsilon$ -CL is described as follows:  $\varepsilon$ -CL (2.0 mL, 2.0 mol L<sup>-1</sup> in CH<sub>2</sub>Cl<sub>2</sub>), EDPP (160 µL, 0.5 mol L<sup>-1</sup> in CH<sub>2</sub>Cl<sub>2</sub>) and DPP (320 µL, 0.5 mol L<sup>-1</sup> in CH<sub>2</sub>Cl<sub>2</sub>) were added to a test tube in glove box. Aliquots were removed from the reaction mixture to determine the conversion based on <sup>1</sup>H NMR spectrum. After stirring for 32 h, the polymerization was quenched by adding a small amount of triethylamine to the polymerization solution. The polymer product was purified by precipitation against methanol three times to give PCL as a white solid. Yield, 336.6 mg (75%); Conv. = 94.5%,  $M_{n,theo.} = 5,440$  g mol<sup>-1</sup>,  $M_{n,NMR} = 5,310$  g mol<sup>-1</sup>,  $M_{n,SEC} = 8,430$  g mol<sup>-1</sup>,  $M_w/M_n = 1.15$ .

## **Derivation of polymerization rate equation**

$$R_{\rm p} = -\frac{d[\mathsf{M}]}{dt} = k_p[\mathsf{M}^*][\mathsf{I}] \tag{1}$$

where  $[M^*] = [activated \varepsilon-CL]$ , [I] is the concentration of active center, and  $k_p$  is the polymerization rate constant. Namely,

$$\frac{dC}{dt} = \frac{k_p [\mathsf{M}^*][\mathsf{I}]}{[\mathsf{M}]_0} \tag{2}$$

*C* is the monomer conversion and  $[M]_0$  is the initial monomer concentration. According to the proposed mechanism in Scheme S1(c), the following reactions can be known:

$$\varepsilon$$
-CL (M) + DPP  $\longrightarrow$  activated monomer (M\*) (I)

$$EDPP + DPP \stackrel{K}{\longleftarrow} EDPP - DPP \qquad (II)$$
active dormant

EDPP is the true active center and its protonated state (EDPP-DPP) is the dormant species. Thus, we can know [I] = [EDPP]. As discussed above, DPP acts as a dual-role catalyst and is divided into two parts. One role is to protonate the phosphinite species to establish an equilibrium reaction (equilibrium constant = K), and the other is the residual DPP to activate  $\varepsilon$ -CL. Thus, [M<sup>\*</sup>] = [DPP] = [DPP]\_0 - [EDPP-DPP] can be known. The following equations can be derived as:

$$[EDPP] + [EDPP - DPP] = [EDPP]_0$$
(3)

$$[DPP] + [EDPP - DPP] = [DPP]_0$$
(4)

$$[\mathsf{M}^*] = [\mathsf{DPP}] \tag{5}$$

$$K = \frac{[\text{EDPP}-\text{DPP}]}{[\text{EDPP}][\text{DPP}]} \tag{6}$$

eq. (7) can be obtained by combining (3)-(6),

$$K = \frac{[\text{EDPP}]_0 - [\text{EDPP}]}{[\text{EDPP}]_0 + [\text{EDPP}] - [\text{EDPP}]_0)}$$
(7)

According to eq. (7), [P] can be expressed by K as eq. (8),

$$[EDPP] = \sqrt{K[EDPP]_0 + (\frac{1+K([DPP]_0 - [EDPP]_0)}{2})^2} - \frac{1+K([DPP]_0 - [EDPP]_0)}{2}$$
(8)

According to *eq*. (2), (5), (6) and [I] = [EDPP],

$$\frac{dC}{dt} = \frac{k_p [M^*][I]}{[M]_0} = \frac{k_p}{[M]_0} [DPP] [EDPP] = \frac{k_p}{[M]_0} \frac{[EDPP]_0 - [EDPP]}{K}$$
(9)

Thus, the final equation can be expressed as eq. (10) after (8) is input to (9),

$$\frac{dC}{dt} = \frac{k_p}{[M]_0} \frac{K[EDPP]_0 - \{\sqrt{K[EDPP]_0 + \left(\frac{1+K([DPP]_0 - [EDPP]_0)}{2}\right)^2 - \frac{1+K([DPP]_0 - [EDPP]_0)}{2}}\}}{K^2}$$
(10)

Namely,

$$C = \frac{k_p}{[M]_0} \frac{K[EDPP]_0 - \{\sqrt{K[EDPP]_0 + \left(\frac{1 + K([DPP]_0 - [EDPP]_0)}{2}\right)^2 - \frac{1 + K([DPP]_0 - [EDPP]_0)}{2}}\}}{K^2} t$$
(11)

**Table S1.** Ring-opening polymerization (ROP) of  $\varepsilon$ -caprolactone ( $\varepsilon$ -CL) using an organocatalytic system composed of ethyl diphenylphosphinite (EDPP) and an organic acid in CH<sub>2</sub>Cl<sub>2</sub><sup>*a*</sup>

run	Organic acid (C)	[C] <sub>0</sub> /[EDPP] <sub>0</sub>	Conv. <sup>b</sup>	$M_{n,theo.}$ <sup>c</sup> $M_{n,SEC}$ <sup>d</sup>		M/M d	
			(%)	(g mol <sup>-1</sup> )	$(g mol^{-1})$	1/1 <sub>W</sub> /1/1/n "	
<b>S</b> 1	PhCO <sub>2</sub> H	1:1	0		n.d. <sup><i>h</i></sup>	n.d.	
S2	PhCO <sub>2</sub> H	2:1	0		n.d.	n.d.	
<b>S</b> 3	HNTf <sub>2</sub>	1:1	0		n.d.	n.d.	
S4	$HNTf_2$	2:1	58.8	6,760	16,100	1.40	
S5	DPP	1:1	39.9	4,600	5,840	1.17	
<b>S</b> 6	DPP	2:1	80.3	9,210	11,800	1.15	
S7 <sup>e</sup>	DPP	2:1	99.2	11,400	19,300	1.34	
<b>S</b> 8	none	0:1	0		n.d.	n.d.	
<b>S9</b> <sup><i>f</i></sup>	none	0:1	0		n.d.	n.d.	
S10 <sup>g</sup>	DPP	2:0	62.1		54,400	1.15	

<sup>*a*</sup> [ $\varepsilon$ -CL]<sub>0</sub>, 2.0 mol L<sup>-1</sup>; [ $\varepsilon$ -CL]<sub>0</sub>/[EDPP]<sub>0</sub> = 100, time = 24 h; temperature, r.t. <sup>*b*</sup> Determined by <sup>1</sup>H NMR in CDCl<sub>3</sub>. <sup>*c*</sup> Calculated from ([ $\varepsilon$ -CL]<sub>0</sub>/[EDPP]<sub>0</sub>) × Conv. × (M.W. of  $\varepsilon$ -CL) + (M.W. of EDPP residue).<sup>*d*</sup> Determined by SEC equipped with an RI detector using THF as eluent at 40 °C and the flow rate of 0.35 mL min<sup>-1</sup>. <sup>*e*</sup> Polymerization was carried out in toluene. <sup>*f*</sup> The monomer was L-lactide (LLA). <sup>*g*</sup> [ $\varepsilon$ -CL]<sub>0</sub>/[DPP]<sub>0</sub> = 100:2. <sup>*h*</sup> Not determined.

run	Monomer (M)	Time	Conv. <sup><i>b</i></sup> (%)	$M_{n,\text{theo.}}$ <sup>c</sup> (g mol <sup>-1</sup> )	$M_{n,SEC} d(g \text{ mol}^{-1})$	$M_{ m w}/M_{ m n}$ $^d$
S6	E-CL	24	80.3	9,210	11,800	1.15
S11	δ-VL	4.5	85.1	8,600	11,600	1.31
S12	ТМС	5.5	5.0	560	n.d.	n.d.
S13	LLA	3	0	n.d. <sup><i>e</i></sup>	n.d.	n.d.

Table S2. ROP of cyclic esters using EDPP/DPP as an organocatalytic system in CH<sub>2</sub>Cl<sub>2</sub> <sup>a</sup>

<sup>*a*</sup> [M]<sub>0</sub>, 2.0 mol L<sup>-1</sup>; [M]<sub>0</sub>/[DPP]<sub>0</sub>/[EDPP]<sub>0</sub> = 100/2/1; temperature, r.t. <sup>*b*</sup> Determined by <sup>1</sup>H NMR in CDCl<sub>3</sub>. <sup>*c*</sup> Calculated from ([ $\varepsilon$ -CL]<sub>0</sub>/[EDPP]<sub>0</sub>) × Conv. × (M.W. of  $\varepsilon$ -CL) + (M.W. of EDPP residue). <sup>*d*</sup> Determined by SEC equipped with an RI detector using THF as eluent at 40 °C and the flow rate of 0.35 mL min<sup>-1</sup>. <sup>*e*</sup> Not determined.

**Table S3.** Block copolymerization of  $\delta$ -VL and  $\varepsilon$ -CL using EDPP/DPP as an organocatalytic system in CH<sub>2</sub>Cl<sub>2</sub> by sequential monomer addition method <sup>*a*</sup>

Polymer	$[\delta - VL + \varepsilon - CL]_0$	time	Conv. $(\delta$ -VL) <sup>b</sup>	Conv. $(\mathcal{E}$ -CL) <sup>b</sup>	$M_{n,,theo.}$ <sup>c</sup>	$M_{n,NMR}$ <sup>b</sup>	$M_{n,SEC}^{d}$	$M_{\rm w}/M_{\rm n}{}^d$	
	/[EDPP]0	(h)	(%)	(%)	(gmol <sup>-1</sup> )	(gmol <sup>-1</sup> )	(gmol <sup>-1</sup> )		
PVL-b-poly	(50+0)/1	3	78.0	0	3,950	4,720	5,960	1.25	
(VL-co-CL)	(0+50)/1	32	97.3	82.2	9,610	8,940	17,100	1.21	
PCL-b-poly	(0+50)/1	32	0	84.7	4,880	5,380	7,200	1.14	
(CL-co-VL)	(50+0)/1	5	71.5	88.0	8,650	8,130	12,400	1.25	

<sup>*a*</sup> [ $\varepsilon$ -CL]<sub>0</sub>, 2.0 mol L<sup>-1</sup>; [EDPP]<sub>0</sub>, 0.5 mol L<sup>-1</sup>; [DPP]<sub>0</sub>, 0.5 mol L<sup>-1</sup>; temperature, r.t. <sup>*b*</sup> Determined by <sup>1</sup>H NMR in CDCl<sub>3</sub>. <sup>*c*</sup> Calculated from ([ $\varepsilon$ -CL]<sub>0</sub>/[EDPP]<sub>0</sub>) × Conv. × (M.W. of  $\varepsilon$ -CL) + (M.W. of EDPP residue). <sup>*d*</sup> Determined by SEC equipped with an RI detector using THF as eluent at 40 °C and the flow rate of 0.35 mL min<sup>-1</sup>. <sup>*e*</sup> not determined.



**Figure S1.** SEC traces of (a) run 4, (b) run 5, (c) run 6, (d) run 7, and run 9 determined by a SEC equipped with an RI detector using THF as eluent at 40 °C and the flow rate of 0.35 mL min<sup>-1</sup>.



**Figure S2.** <sup>1</sup>H NMR spectra of (a) &-CL, (b) EDPP, (c) HNTf<sub>2</sub> (d) &-CL + HNTf<sub>2</sub> (molar ratio, 1:1, the same for the following mixtures), (e) EDPP + HNTf<sub>2</sub>, and (f) &-CL + EDPP + HNTf<sub>2</sub> determined in CDCl<sub>3</sub>.



**Figure S3.** <sup>1</sup>H NMR spectra of (A): (a)  $\varepsilon$ -CL, (b) EDPP, (c) PhCO<sub>2</sub>H, (d)  $\varepsilon$ -CL + PhCO<sub>2</sub>H (molar ratio, 1:1, the same for the following mixtures), (e) PhCO<sub>2</sub>H + EDPP, and (f)  $\varepsilon$ -CL + PhCO<sub>2</sub>H + EDPP determined in CDCl<sub>3</sub>.



**Figure S4.** <sup>1</sup>H NMR spectrum of a PCL (theoretically 25-mer) purified after reprecipitation from hexane for three times and then preparative SEC in CH<sub>2</sub>Cl<sub>2</sub>, determined in CDCl<sub>3</sub>.



**Figure S5.** (a) Conv. vs time and  $\ln[1/(1-\text{Conv.})]$  vs time plots, and (b) the dependence of  $M_{n,\text{SEC}}$  and  $M_w/M_n$  on Conv. of the ROP of  $\delta$ -VL carried out at r.t. in CH<sub>2</sub>Cl<sub>2</sub> under various  $[\delta$ -VL]<sub>0</sub>/[EDPP]<sub>0</sub>/[DPP]<sub>0</sub> ratios of 100/2/1 ( $\circ$ ) and 100/1/1 ( $\circ$ ) ([ $\delta$ -VL]<sub>0</sub> = 2.0 mol L<sup>-1</sup>).



**Figure S6.** SEC traces of copolymerization of  $\delta$ -VL and  $\varepsilon$ -CL by sequential monomer addition method: (a) PVL in first polymerization (red line) and PVL-*b*-poly(VL-*co*-CL) (blue line) in the following copolymerization and (b) PCL in first polymerization (red line) and PCL-*b*-poly(CL-*co*-VL) (blue line) in the following copolymerization determined by a SEC equipped with an RI detector using THF as eluent at 40 °C and the flow rate of 0.35 mL min<sup>-1</sup>.