Direct routes to functional RAFT agents from substituted *N*-alkyl maleimides

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Materials and Methods.

All solvents were purchased from Fischer Chemicals and were of analytical grade. All chemicals were purchased from Sigma-Aldrich Company, Alfa Aesar or Fisher Scientific and utilised without further purification. Azoinitiator 2,2'-azobis[2- (2-imidazolin-2-yl)propane]dihydrochloride (VA-044) was purchased from Wako.

Thin layer chromatography (TLC) was performed using pre-coated silica gel 60 ALUGRAM SIL G/UV254 and developed in the solvent system indicated. Compounds were visualised by use of UV light (254 nm) or a basic solution (10% w/w K_2CO_3 in water) of KMnO₄. Acros Organics 60Å (0.035-0.070 mm) silica gel was used for column chromatography.

Agilent 971-FP Flash Purification System was used to perform flash column chromatography on SiO₂ (SF10-8g, Si-35) or reversed-phase (KP-C18-HS) as the stationary phase.

¹H and ¹³C NMR spectra were recorded on a Bruker DPX400 UltraShield^M Spectrometer and processed with MestReNova 6.0.2[©] 2009 Mestrelab Research S.L. All chemical shifts are reported in ppm (δ) relative to tetramethylsilane or referenced to the chemical shifts of residual solvent resonances. The following abbreviations were used to explain the multiplicities: s = singlet, bs = broad singlet, d = doublet, t = triplet, q= quartet, m = multiplet, app. t = apparent triplet.

ESI-TOF Mass Spectrometry measurements were carried out using a Bruker MicroTOF spectrometer (LC-TOF instrument equipped with an ESI source).

Polymer molecular weights were determined by size exclusion chromatography (SEC) performed on a Polymer Laboratories GPC 50 system (Polymer Laboratories) equipped with refractive index (RI) detector. Separations were performed on a pair of Agilent PLgel Mixed-D columns (7.5 x 300 mm, 5 μ m bead size, Polymer Labs UK), eluting with DMF + 0.1% w/w LiBr at flow rate of 1 mL·min⁻¹ and

injection volume of 100 µL. Samples were prepared at 5 mg·mL⁻¹ concentration. The molecular weights and dispersities of the polymers were calculated according to a standard calibration method using PMMA narrow standards (500-450,000 g·mol⁻¹). Data were elaborated with Polymer Labs Cirrus 3.0 Software. For n-butyl acrylate polymers, GPC was performed on an Agilent 1200 Infinity Series equipped with a differential refractive index detector. The mobile phase was HPLC grade CHCl₃ at 25 °C and a flow rate of 1 mL·min⁻¹. Separations were performed on a pair of PLgel Mixed-D columns (7.8 × 300 mm, 5 µm bead size, Agilent) fitted with a matching guard column (50 × 7.8 mm). The molecular weights and dispersities of the polymers were calculated according to a standard calibration method using PMMA narrow molecular (200-360,000 g·mol⁻¹). Chromatographs were analysed using Astra 6.1 software.



Scheme S1. *Reagents and conditions*: (a) i. NaH; ii. CS_2 , 0°C; (b) methyl-2-bromopropionate; (c) $K_3Fe(CN)_6$; (d) I_2 ; (e) AIBN.

Sodium 2-hydroxyethyl carbonotrithioate (1).¹ NaH (60% wt% in mineral oil, 5.55 g, 138 mmol) was dispersed in 150 mL of diethyl ether and cooled to 0°C in an ice bath. Mercaptoethanol (9.00 mL, 128 mmol) was added dropwise under stirring to the organic suspension and the mixture was stirred for 10 minutes, then CS₂ (15.4 mL, 256 mmol) was added dropwise to the suspension and the reaction was stirred at ambient temperature for one hour. The resulting yellow precipitate was obtained and recovered by filtration, washed with diethyl ether, and finally desiccated under reduced pressure, to give the intermediate **(1)** as a yellow solid (20.2 g, 11.5 mmol, 94%), which was used for the next step without further purification.

¹**H NMR** (400 MHz, DMSO-d₆) δ 4.64 (t, *J* = 5.5 Hz, 1H, *H*OCH₂CH₂SC(S)S), 3.45 (q, *J* = 7.2 Hz, 2H, HOCH₂CH₂SC(S)S), 3.12 (t, *J* = 7.4 Hz, 2H, HOCH₂CH₂SC(S)S).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 239.4 (1C, SC(S)S), 60.0 (1C, HOCH₂CH₂SC(S)S), 42.7 (1C, HOCH₂CH₂SC(S)S).



Figure S1. ¹H NMR spectrum of sodium 2-hydroxyethyl carbonotrithioate (1) in DMSO-d₆.



Figure S2. ¹³C NMR spectrum of sodium 2-hydroxyethyl carbonotrithioate (1) in DMSO-d₆.

Methyl 2-((((2-hydroxyethyl)thio)carbonothioyl)thio)propanoate (2). Sodium 2-hydroxyethyl carbonotrithioate intermediate **(1)** (13.26 g, 75.25 mmol) was suspended in acetone (100 mL) and

methyl-2-bromo-propionate (9.2 mL, 82 mmol) was added dropwise under stirring. After 2 hours the solvent was removed under reduced pressure, the product was suspended in 50 mL of Et_2O and washed with water (3 x 100mL). The organic layer was dried over MgSO₄, filtered and the solvent evaporated under reduced pressure to give an orange viscous oil which was purified by flash chromatography (silica gel 60, 35-70 um) using petroleum ether/ethyl acetate 8:2 (vol/vol) as the eluent, to give the intermediate (**2**) as a yellow-orange oil (7.20 g, 30.0 mmol, 40%).

¹**H NMR** (400 MHz, CDCl₃) δ 4.9 (q, *J* = 7.4 Hz, 1H, SC*H*(CH₃)), 3.9 (t, *J* = 6.0 Hz, 2H, HOC*H*₂CH₂S), 3.8 (s, 3H, -C(O)OCH₃), 3.6 (t, *J* = 6.1 Hz, 2H, HOCH₂CH₂S), 1.9 (bs, 1H, HOCH₂), 1.6 (d, *J* = 7.4 Hz, 3H, CHCH₃).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 221.8 (1C, SC(S)S), 171.6 (1C, -*C*(O)OCH₃), 60.3 (1C, HOCH₂CH₂SC(S)S), 52.8 (1C, -SCH), 48.0 (1C, HOCH₂CH₂SC(S)S), 39.4 (1C, -OCH₃) 16.8 (1C, -CHCH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 262.98, found 262.98 (100%).



Figure S3. ¹H NMR spectrum of methyl 2-((((2-hydroxyethyl)thio)carbonothioyl)thio)propanoate **(2)** in CDCl₃.



hydroxyethyl)thio)carbonothioyl)thio)propanoate (2) in CDCl₃.



Figure S5. ¹³C spectrum of methyl 2-((((2-hydroxyethyl)thio)carbonothioyl)thio)propanoate (2) in CDCl₃.

Dithiobis-2-hydroxyethyl carbonotrithioate disulfide (3). Sodium 2-hydroxyethyl carbonotrithioate intermediate (1) (3.07 g, 17.4 mmol) was dissolved in of water (100 mL) and solid $K_3Fe(CN)_6$ (6.31 g,

19.2 mmol) was slowly added. The reaction was stirred at ambient temperature for 30 minutes. The resulting reddish-orange viscous precipitate was extracted from the aqueous mixture with ethyl acetate (100 mL). The extraction process was repeated four times and the organic layers, combined, were dried over $MgSO_4$. The mixture was filtered, and the solvent was evaporated under reduced pressure to give **(3)** (1.14 g, 37.2 mmol, 43 %) as an orange viscous oil which was used for the next step without further purification.

¹**H NMR** (400 MHz, DMSO-d₆) δ 4.64 (t, *J* = 5.5 Hz, 2H, *H*OCH₂), 3.46 (q, 4H, HOCH₂), 3.11 (t, *J* = 7.2 Hz, 4H, CH₂CH₂S).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 221.6 (2C, SC(S)S), 57.7 (2C, HOCH₂CH₂SC(S)S), 41.4 (2C, HOCH₂CH₂SC(S)S).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 328.89, found 328.89 (39%).



Figure S6. ¹H NMR spectrum of dithiobis-2-hydroxyethyl carbonotrithioate disulfide (3) in DMSO-d₆.



Figure S7. ¹³C NMR spectrum of dithiobis-2-hydroxyethyl carbonotrithioate disulfide (3) in DMSO-d₆.

Sodium dodecyl carbonotrithioate (4). NaH (60% wt% in mineral oil, 3.26 g, 82.5 mmol) was dispersed in diethyl ether (150 mL) and cooled to 0°C in an ice bath. 1-Dodecanethiol (17.7 mL, 74.1 mmol) was added dropwise under stirring to the suspension and the resulting mixture was stirred for 12 hours at ambient temperature. Then CS₂ (13.4 mL, 222 mmol) was added dropwise to the suspension and the reaction was stirred at ambient temperature for further 12 hours. The resulting yellow precipitate was recovered by filtration, washed with diethyl ether, and finally desiccated under reduced pressure, to give the intermediate **(4)** (17.9 g, 59.4 mmol, 80%) which was used for the next step without further purification.

¹**H NMR** (400 MHz, DMSO-d₆) δ 2.98 (t, J = 7.3Hz, 2H, CH₂S), 1.50 (q, 2H, CH₂CH₂S), 1.24 (s, 18H, CH₂), 0.85 (t, *J* = 6.8 Hz, 3H, CH₂CH₃).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 239.9 (1C, SC(S)S), 39.5 (1C, CH₂S), 31.3 (1C, CH₂CH₂S), 29.1 (1C, CH₂), 29.0 (CH₂), 28.8 (1C, CH₂), 28.71 (CH₂), 28.7 (1C, CH₂), 22.1 (1C, CH₂), 14.0 (1C, CH₃).



Figure S8. ¹H NMR spectrum of sodium dodecyl trithiocarbonate (4) in DMSO-d₆.



Figure S9. ¹H-¹H COSY NMR spectrum of sodium dodecyl carbonotrithioate (4) in DMSO-d₆.



Figure S10. ¹³C NMR spectrum of sodium dodecyl carbonotrithioate (4) in DMSO-d₆.

Methyl 2-(((dodecylthio)carbonothioyl)thio)propanoate (5). Sodium dodecyl trithiocarbonate **(4)** (1.49 g, 4.96 mmol) was dissolved in acetone (30 mL) and methyl-2-bromo-propionate (0.66 mL, 5.9 mmol) was added dropwise under stirring. After 12 hours the precipitated NaBr was filtered off and the solvent was removed under reduced pressure. The resulting residue was solubilised in chloroform (50 mL) and washed with water (3 x 50mL) and, finally with brine (50 mL). The organic layer was dried over MgSO₄, filtered, and the solvent was evaporated under reduced pressure to give an orange viscous residue which was purified by flash chromatography (silica gel 60, 35-70 um) using petroleum ether as the eluent (71.4 g, 3.85 mmol, 78%).

¹**H NMR** (400 MHz, CDCl₃) δ 4.84 (d, *J* = 7.4 Hz, 1H, CHCH₃), 3.75 (s, 3H, OCH₃), 3.35 (t, *J* = 7.4 Hz, 2H, CH₂S), 1.69 (p, *J* = 7.3 Hz, 2H, CH₂CH₂S), 1.60 (d, *J* = 7.4 Hz, 3H, CHCH₃), 1.39 (m, 2H, CH₂), 1.26 (s, 16H, CH₂), 0.88 (t, *J* = 6.8 Hz, 3H, CH₂CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 222.2 (1C, SC(S)S), 171.8 (1C, CO), 53.0 (1C, OCH₃), 47.9 (1C, SCH), 37.4 (1C, CH₂S), 32.1 (1C, CH₂CH₂S), 29.7 (1C, CH₂), 29.6 (1C, CH₂), 29.5 (1C, CH₂), 29.2 (1C, CH₂), 29.1 (1C, CH₂), 28.0 (1C, CH₂), 27.5 (1C, CH₂), 23.42 (1C, CH₂), 22.8 (1C, CH₂), 17.1 (CHCH₃), 14.3 (1C, CH₂CH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 387.14, found 387.14 (100%).



Figure S11. ¹H NMR spectrum of methyl 2-(((dodecylthio)carbonothioyl)thio)propanoate (5) in CDCl₃.



Figure S12. ¹H-¹H COSY NMR spectrum of methyl 2-(((dodecylthio)carbonothioyl)thio)propanoate **(5)** in CDCl₃.



Figure S13. ¹³C NMR spectrum of methyl 2-(((dodecylthio)carbonothioyl)thio)propanoate (5) in CDCl₃.

Dithiobis-2-dodecyl carbonotrithioate disulfide (6). Sodium dodecyl trithiocarbonate intermediate **(4)** (17.9 g, 59.4 mmol) was dissolved in diethyl ether (250 mL) and I_2 (8.40 g, 33.1 mmol) was slowly added over 10 minutes. The reaction was stirred at ambient temperature for 12 hours. The resulting dark solution was washed with 5% aqueous $Na_2S_2O_3$ (250 mL), water and finally with brine. The organic layer was dried over MgSO₄, filtered and the solvent evaporated under reduced pressure to give **(5)** (14.41 g, 25.95 mmol, 87 %) as an orange viscous oil which was used for the subsequent step without further purification.

¹**H NMR** (400 MHz, CDCl₃) δ 3.32 (t, *J* = 7.4 Hz, 4H, CH₂S), 1.72 (4H, m, CH₂CH₂S), 1.42 (m, 4H, CH₂), 1.28 (s, 32H, CH₂), 0.90 (t, *J* = 6.6 Hz, 6H, CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 221.7 (2C, SC(S)S), 38.5 (2C, CH₂S), 32.1 (2C, CH₂CH₂S), 29.8 (2C, CH₂), 29.8 (2C, CH₂), 29.7 (2C, CH₂), 29.6 (2C, CH₂), 29.5 (2C, CH₂), 29.2 (2C, CH₂), 29.1 (2C, CH₂), 27.5 (2C, CH₂), 22.0 (2C, CH₂), 14.3 (1C, CH₃).



Figure S14. ¹H NMR spectrum of dithiobis-2-dodecyl carbonotrithioate disulfide (6) in CDCl₃.



Figure S15. ¹H-¹H COSY NMR spectrum of dithiobis-2-dodecyl carbonotrithioate disulfide (6) in CDCl₃.



Figure S16. ¹³C NMR spectrum of dithiobis-2-dodecyl carbonotrithioate disulfide (6) in CDCl₃.

2-Cyanopropan-2-yl dodecyl carbonotrithioate (7). Dithiobis-2-dodecyl carbonotrithioate disulfide **(6)** (7.60 g, 13.7 mmol) was dissolved in ethyl acetate (50 mL) in a three neck round bottom flask equipped with a condenser. Azobisisobutyronitrile (AIBN) (2.65 g, 16.1 mmol) was added to the reaction mixture. The reactor was sealed, cooled to 0°C in an ice bath and degassed with argon for 40 minutes. The reaction was then heated at 77°C and left to react for 48 hours. The solvent was evaporated under reduced pressure to give an orange viscous oil which was purified by flash chromatography (silica gel 60, 35-70 um) using petroleum ether/ethyl acetate (10:0 to 9.5:0.5 vol/vol) as the eluent (6.24 g, 18.0 mmol, 66%).

¹**H NMR** (400 MHz, CDCl₃) δ 3.33 (t, *J* = 7.4 Hz, 2H, CH₂CH₂S), 1.87 (s, 6H, C(CH₃)₂), 1.69 (m, 2H, CH₂CH₂S), 1.40 (m, 2H, CH₂), 1.26 (s, 16H, CH₂), 0.89 (t, *J* = 6.6 Hz, 3H, CH₂CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 218.0 (1C, SC(S)S), 120.6 (1C, CN), 43.0 (1C, SC(CH₃)₂, 37.9 (1C, SCH₂CH₂), 32.1 (1C, SCH₂CH₂), 29.8 (1C, CH₂), 29.7 (1C, CH₂), 29.6 (1C, CH₂), 29.5 (1C, CH₂), 29.2 (1C, CH₂), 29.1 (1C, CH₂), 27.9 (1C, CH₂), 27.2 (1C, CH₂), 22.8 (1C, CH₂), 14.3 (1C, CH₂CH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 368.15, found 368.15 (48.2%).



Figure S17. ¹H NMR spectrum of 2-cyanopropan-2-yl dodecyl carbonotrithioate (7) in CDCl₃.



Figure S18. ¹H-¹H COSY NMR spectrum of 2-cyanopropan-2-yl dodecyl carbonotrithioate (7) in CDCl₃.



Figure S19. ¹³C NMR spectrum of 2-cyanopropan-2-yl dodecyl carbonotrithioate (7) in CDCl₃.

Synthesis of maleimides (B).



Scheme S2. Synthesis of maleimide intermediates (B).

3a,4,7,7a-tetrahydro-4,7-epoxyisobenzofuran-1,3-dione (8). Intermediate **(8)** was synthesised as described by Bolm *et al.*² Maleic anhydride (30.0 g, 306 mmol) was suspended in toluene (150 mL) and the suspension was heated at 80°C. Furan (33.5 mL, 459 mmol) was slowly added via syringe and the mixture was left to react under reflux in an oil bath at 80°C for 6 hours. The reaction mixture was then cooled down to ambient temperature and the resulting white crystals were collected by filtration and washed twice with petroleum ether (2 x 60 mL), to give the anhydride **(8)** as a white solid (45.2 g, 272 mmol, 89%).

¹**H NMR** (400 MHz, CDCl₃) δ 6.55 (t, J= 3.4 Hz, 2H, CH=CH), 5.43-5.54 (m, 2H, CH-O), 3.15 (dd, J=5.3, 2.6 Hz, 2H, CH).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 170.0 (2C, CO), 137.1 (2C, CH=CH), 82.2 (2C, CH-O), 48.9 (2C, CH).



Figure S20. ¹H NMR spectrum of 3a,4,7,7a-tetrahydro-4,7-epoxyisobenzofuran-1,3-dione (8) in CDCl₃.



Figure S21. ¹³C NMR spectrum of 3a,4,7,7a-tetrahydro-4,7-epoxyisobenzofuran-1,3-dione (8) in CDCl₃.

General procedure for the synthesis of intermediates (9a), (9b) and (9c).

Anhydride **(8)** was suspended in methanol and the mixture cooled at 0°C. A solution of the desired amino-alcohol and triethylamine were added dropwise to the reactor over a period of 30 minutes at a temperature of 0-2°C. After this period, the temperature of the heating block was steadily increased to 70°C and the reaction mixture was stirred at this temperature for 12-18 hours. The reactions mass was then filtered, and the solvent was removed under reduced pressure.

The reaction conditions utilised for the synthesis and purification of each intermediate are described below:

(3aR,4R,7R,7aS)-2-(2,3-dihydroxypropyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-

dione (9a). Anhydride **(8)** (10.6 g, 63.8 mmol), methanol (350 mL), 3-amino-1,2-propanediol (6.0 mL, 77 mmol), triethylamine (10.3 mL, 71.9 mmol), 70°C for 12 hours. The reaction mass was then filtered, and the solvent removed under reduced pressure, to give **(9a)** (9.53 g, 39.9 mmol, 62.5% yield). ¹H NMR (400 MHz, DMSO-d₆) δ 6.55 (t, *J* = 1.0 Hz, 2H, CHCH), 5.12 (t, *J* = 1.0 Hz, 2H, CHO), 4.78 (d, *J* = 5.3 Hz, 1H, CHOH), 4.57 (t, *J* = 5.7 Hz, 1H, CH₂OH), 3.60-3.70 (m, 1H, CHOH), 3.35 (d, *J* = 1.9 Hz, 2H, CH₂N), 3.26 (t, *J* = 5.6 Hz, 2H, CH₂OH), 2.91 (q, *J* = 2.6 Hz, 2H, CHCO).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 176.6 (2C, CO), 136.4 (2C, CHCH), 80.3 (2C, CHO), 68.1 (1C, CHOH),
63.9 (1C, CH₂OH), 47.7 (2C, CHCO), 42.3 (1C, CH₂N).



Figure S22. ¹H NMR spectrum of (3aR,4R,7R,7aS)-2-(2,3-dihydroxypropyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione **(9a)** in DMSO-d₆.



Figure S23. ¹H-¹H COSY NMR spectrum of (3aR,4R,7R,7aS)-2-(2,3-dihydroxypropyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione **(9a)** in DMSO-d₆.



Figure S24. ¹³C NMR spectrum of (3aR,4R,7R,7aS)-2-(2,3-dihydroxypropyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione**(9a)**in DMSO-d₆.

2-(2-(2-hydroxyethoxy)ethyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione (9b).

Anhydride **(8)** (23.09 g, 139.0 mmol), methanol (100 mL), 2-(2-aminoethoxy)ethanol (17.54 g, 116.8 mmol), triethylamine (19.4 g, 139 mmol), 70°C for 18 hours. The crude product was purified via flash chromatography (silica gel 60, 35–70 μ m, eluents ethyl acetate /methanol, gradient 9:1 to 1:1, vol/vol) to give **(9b)** as a white solid (20.07 g, 79.31 mmol, 57%).

¹**H NMR** (400 MHz, CDCl₃) δ 6.44 (s, 2H, CH=CH), 5.20 (s, 2H, CHOCH), 3.63 (t, *J* = 5.5 Hz, 2H, NCH₂CH₂OCH₂CH₂OH), 3.56 (t, *J* = 5.0 Hz, 4H, NCH₂CH₂OCH₂CH₂OH), 3.45 (t, *J* = 4.0 Hz, 2H, NCH₂CH₂OCH₂CH₂OH), 2.80 (s, 2H, CHCO), 2.74 (bs, 1H, OH).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 176.4 (2C, CO), 136.5 (2C, CHCH), 80.3 (2C, CHO), 71.9 (1C, OCH₂CH₂OH), 66.3 (1C, NCH₂CH₂O), 60.1 (1C, OCH₂CH₂OH), 47.1 (2C, CHCH), 37.6 (1C, NCH₂CH₂O).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 276.08, found 276.08 (100%).



Figure S25. ¹H NMR spectrum of 2-(2-(2-hydroxyethoxy)ethyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione **(9b)** in CDCl₃.



Figure S26. ¹H-¹H COSY NMR spectrum of 2-(2-(2-hydroxyethoxy)ethyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione **(9b)** in DMSO-d₆.



Figure S27. ¹³C NMR spectrum of 2-(2-(2-hydroxyethoxy)ethyl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione (**9b**) in DMSO-d₆.

2-(1,3-dihydroxypropan-2-yl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione (9c). Anhydride (8) (11.5 g, 69.3 mmol), methanol (50 mL), 2-amino-1,3-propanediol (6.31 g, 69.3 mmol), triethylamine (7.01 g, 69.3 mmol), 67°C for 18 hours. The crude product was purified by flash chromatography (silica gel 60, 35–70 μ m, eluents ethyl acetate /methanol, gradient 9:1 to 1:1, vol/vol) to give (9c) as a white solid (10.1 g, 42.2 mmol, 61%).

¹**H NMR** (400 MHz, DMSO-d₆) δ 6.54 (t, *J* = 0.9 Hz, 2H, *CHCH*), 5.10 (t, *J* = 0.9 Hz, 2H, CHO), 4.76 (t, *J* = 5.8 Hz, 2H, OH), 4.01 (p, *J* = 7.0 Hz, 1H, CH), 3.60 (t, 4H, *J* = 6.3 Hz, CH₂), 2.87 (s, 2H, *CHCO*).

¹³C {¹H} NMR (101 MHz, DMSO-d₆) δ 176.9 (2C, CO), 136.5 (2C, CHCH), 80.4 (2C, CHO), 58.1 (2C, CH₂OH), 56.8 (1C, CHN), 46.9 (2C, CHCO).



Figure S28. ¹H NMR spectrum of 2-(1,3-dihydroxypropan-2-yl)-3a,4,7,7a-tetrahydro-1H-4,7epoxyisoindole-1,3(2H)-dione **(9c)** in DMSO-d₆.



f1 (ppm)

Figure S29. ¹³C NMR spectrum of 2-(1,3-dihydroxypropan-2-yl)-3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione **(9c)** in DMSO-d₆.

3-((3aR,4R,7R,7aS)-1,3-dioxo-1,3,3a,4,7,7a-hexahydro-2H-4,7-epoxyisoindol-2-yl) propane-1,2-diyl dipalmitate (10a1). Compound (9a) (4.80 g, 19.9 mmol) was dissolved in dichloromethane (25 mL). A solution of palmitoyl chloride (15.1 mL, 49.8 mmol) in dichloromethane (20 mL) was added to the reactor. The resulting solution was cooled to 0°C and triethylamine (8.3 mL, 60 mmol) was then added dropwise at to the stirred solution and left overnight at room temperature. NMR analysis of an aliquot taken from the reaction mixture showed that the reaction was not complete. To push the reaction to higher conversion, (dimethyl-amino)pyridine (DMAP) (4.00 g, 39.9 mmol) organocatalyst was then added, along with more palmitoyl chloride (6.04 mL, 19.9 mmol), and the reaction stirred overnight at room temperature. Methanol (20 mL) was added dropwise at 0°C to convert the excess of acid chloride into the corresponding methyl ester and facilitate its subsequent removal. The volume of the reaction mixture was reduced under reduced pressure and the resulting semi-solid mass added of diethyl ether (500 mL). The resulting slurry was filtered and washed with more diethyl ether (60 mL), and the solvent was removed from the combined diethyl ether solutions under reduced pressure, and the solid residue was purified by silica column chromatography (gradient of diethyl ether: petroleum ether from 40:60 to 60:40 vol/vol). (10a1) was isolated as a white solid (6.6 g, 9.2 mmol, 46% yield).

¹**H NMR** (400 MHz, CDCl₃) δ 6.43 (t, *J* = 1.0 Hz, 2H, CHCH), 5.22 (m, 1H, NCH₂CH), 5.18 (m, 2H, CHOCH), 4.10 (m, 2H, NCH₂CHCH₂O), 3.72 (m, 2H, NCH₂), 2.78 (dd, *J* = 17.3, 6.5 Hz, 2H, CHCO), 2.22 (dt, *J* = 23.8, 6.5 Hz, 4H, CH₂CO), 1.51 (m, 4H, CH₂), 1.22 (m, 48H, CH₂), 0.80 (t, *J* = 7.0 Hz, 6H, CH₃),

¹³C {¹H} NMR (101 MHz, CDCl₃) 175.9 (2C, CO). 173.3 (2C, CO_{ester}), 136.6 (2C, CHCH), 81.0 (2C, CHO), 68.3 (1C, CHO), 62.7 (1C, CH₂O), 47.4 (2C, CHCO), 39.1 (1C, CH₂N), 31.9 (1C, CH₂CO), 34.1 (1C, CH₂CO), 22.71-29.7 (26C, CH₂), 14.1 (2C, CH₃).



Figure S30. ¹H NMR spectrum 3-((3aR,4R,7R,7aS)-1,3-dioxo-1,3,3a,4,7,7a-hexahydro-2H-4,7-epoxyisoindol-2-yl) propane-1,2-diyl dipalmitate (**10a1**) in CDCl₃.



Figure S31. ¹H-¹H COSY NMR spectrum 3-((3aR,4R,7R,7aS)-1,3-dioxo-1,3,3a,4,7,7a-hexahydro-2H-4,7-epoxyisoindol-2-yl) propane-1,2-diyl dipalmitate (**10a1**) in CDCl₃.



f1 (ppm)

Figure S32. ¹³C NMR spectrum 3-((3aR,4R,7R,7aS)-1,3-dioxo-1,3,3a,4,7,7a-hexahydro-2H-4,7-epoxyisoindol-2-yl) propane-1,2-diyl dipalmitate (**10a1**) in CDCl₃.

3-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)propane-1,2-diyl dipalmitate (10a). Intermediate **(10a1)** (6.6 g, 9.2 mmol) was suspended in toluene (150 mL) and the mixture heated to reflux (heating block at 130°C) for 6 hours. The solvent was then removed under reduced pressure to give **(10a)** as a white solid (5.03 g, 7.76 mmol; 84.7% yield).

¹**H NMR** (400 MHz, CDCl₃) δ 6.65 (s, 2H, CHCH), 5.24 (m, 1H, CHO), 3.99-4.20 (m, 2H, CH₂O), 3.57-3.76 (m, 2H, CH₂N), 2.23 (dt, *J* = 26.7, 7.6 Hz, 4H, CH₂CO), 1.38-1.85 (m, 4H, CH₂), 1.16-1.25 (m, 48H, CH₂), 0.81 (t, *J* = 6.6 Hz, 6H, CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 173.27 (2C, COO), 170.25 (2C, CO), 134.25 (2C, CHCH), 68.94 (1C, CHO), 62.74 (1C, CH₂O), 38.17 (1C, CH₂N), 34.10 (1C, CH₂CO), 31.94 (1C, CH₂CO), 22.71-29.72 (26C, CH₂), 14.13 (2C, CH₃).

Mass spectrometry (MALDI TOF) m/z (%): expected for [M+Na⁺] 670.50, found 670.70 (100).



Figure S33. ¹H NMR spectrum 3-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)propane-1,2-diyl dipalmitate (**10a**) in CDCl₃.



Figure S34. ¹H-¹H COSY NMR spectrum 3-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)propane-1,2-diyl dipalmitate (**10a**) in CDCl₃.



Figure S35. ¹³C NMR spectrum of 3-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)propane-1,2-diyl dipalmitate (**10a**) in CDCl₃.

1-(2-(2-hydroxyethoxy)ethyl)-1H-pyrrole-2,5-dione (10b).

Intermediate **(9b)** (6.00 g, 23.7 mmol) was suspended in toluene (150 mL) and the mixture heated to reflux (heating mantle kept at 120°C) for 16 hours. Subsequently, the solvent was removed under reduced pressure to give **(10b)** as an off-white solid (3.82 g, 20.6 mmol; 87.1% yield).

¹**H NMR** (400 MHz, CDCl₃) δ 6.71 (2H, s, CHCH), 3.73 (t, *J* = 5.2 Hz, 2H, NCH₂CH₂OCH₂CH₂OH), 3.68-3.63 (m, 4H, NCH₂CH₂OCH₂CH₂OH), 3.56 (t, *J* = 5.1 Hz, 2H, NCH₂CH₂OCH₂CH₂OH), 2.63 (bs, 1H, CH₂OH).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 170.80 (2C, CO), 134.18 (2C, CHCH), 72.11 (1C, NCH₂CH₂OCH₂),
68.07 (1C, NCH₂CH₂O) 61.55 (1C, CH₂OH), 37.33 (NCH₂).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 208.06, found 208.06 (100%).



Figure S36. ¹H NMR spectrum of 1-(2-(2-hydroxyethoxy)ethyl)-1H-pyrrole-2,5-dione (10b) in CDCl₃.



Figure S37. ¹H-¹H COSY NMR spectrum of 1-(2-(2-hydroxyethoxy)ethyl)-1H-pyrrole-2,5-dione **(10b)** in CDCl₃.



Figure S38. ¹³C NMR spectrum of 1-(2-(2-hydroxyethoxy)ethyl)-1H-pyrrole-2,5-dione (10b) in CDCl₃.



Figure S39. ¹H-¹³C HSQC NMR spectrum of 1-(2-(2-hydroxyethoxy)ethyl)-1H-pyrrole-2,5-dione **(10b)** in CDCl₃.

1-(1,3-dihydroxypropan-2-yl)-1H-pyrrole-2,5-dione (10c). Intermediate **(9c)** (1.0 g, 4.2 mmol) was suspended in toluene (150 mL) and the mixture heated to reflux (heating mantle kept at 120°C) for 16 hours. Subsequently, the solvent was removed under reduced pressure to give **(10b)** as a solid (0.58 g, 3.39 mmol; 80.7% yield).

¹**H NMR** (400 MHz, CDCl₃) δ 6.97 (2H, s, C*H*=C*H*), 4.81 (t, 2H, OH), 4.01 (m, 1H, NCH), 3.68-3.54 (m, 4H, CH₂).



Figure S40. ¹H NMR spectrum of 1-(1,3-dihydroxypropan-2-yl)-1H-pyrrole-2,5-dione **(10c)** in DMSO-d₆.



Figure S41. ¹H-¹H COSY NMR spectrum of 1-(1,3-dihydroxypropan-2-yl)-1H-pyrrole-2,5-dione (10c) in DMSO-d₆.

General procedure for the synthesis of Maleimide based RAFT agents (Mal-CTA). Maleimide and CTA starting materials were dissolved in the required solvent in a Schlenk tube containing a magnetic stirrer, and their molar ratio in the reaction feed was confirmed by ¹H NMR. The radical initiator was added as an aliquot taken from a stock solution and the tube sealed with a rubber septum. The mixture was cooled with an ice bath and was deoxygenated by argon bubbling for 15 minutes. The reaction was started by placing the Schlenk tube in a thermostated oil bath at the desired temperature.

Synthesis of Maleimide based RAFT agents

	Route 1a		Route 1b	
	(13b)	(14b)	(13a)	(13b)
Maleimide	10b	10b	10a	10b
m _{maleimide added} (mg)	71.3	69.9	118	60.5
mmol	0.385	0.378	0.182	0.327
СТА			(7)	(7)
bis(thioacyl) disulfide	(6)	(3)	-	-
m _{CTA added} (mg)	230.3	123.7	69	124
mmol	0.4155	0.4042	0.20	0.360
Initiator	AIBN	AIBN	AIBN	AIBN
m _{initiator added} (mg)	122.9	122.8	1.65	2.95
mmol	0.7484	0.7478	0.0100	0.0180
Solvent 1	Toluene	EtOAc	Toluene	Toluene
$V_{solvent 1 added}$ (mL)	1	1	0.183	0.327
[monomer] (mol·L ⁻¹)	0.4	0.4	1	1
[CTA or bis(thioacyl) disulfide] / [initiator]	0.5	0.5	20	20
Temperature (°C)	70	70	75	75
Time (h)	16	16	48	48
m _{product} (mg)	108.9	96.8	132.3	72.0
mmol	0.21	0.24	0.13	0.14
Yield (%)	55	63	73	42

 Table S1. Synthesis of maleimide-RAFT agents (Mal-CTAs) via route 1: reaction conditions.

	11a	11b	11c	12a	12b
Maleimide	10a	10b	10c	10a	10b
m _{maleimide added} (mg)	131	824.9	200	124	57.8
mmol	0.183	4.454	1.17	0.174	0.284
СТА	(2)	(2)	(2)	(5)	(5)
m _{CTA added} (mg)	48.5	1176	309.6	70.0	114
mmol	0.202	4.900	1.288	0.192	0.312
Initiator	AIBN	VA-044	VA-044	AIBN	VA-044
m _{initiator added} (mg)	0.301	15.8	3.79	1.43	1.01
mmol	1.83 [.] 10 ⁻³	0.0489	0.117	8.71 [.] 10 ⁻³	3.12 [.] 10 ⁻³
Solvent 1	Toluene	DMSO	DMSO	Toluene	DMSO
V _{solvent 1 added} (mL)	0.183	1.485	0.390	0.174	0.95
Solvent 2	-	H ₂ O	H ₂ O	-	H ₂ O
V _{solvent 2 added} (mL)	-	0.742	0.195	-	0.47
[monomer] (mol·L ⁻¹)	1	2	2	1	2
[CTA]/[initiator]	100	100	100	20	100
Temperature (°C)	75	100	65	75	90
Time (h)r	24	4	24	48	12
m _{product} (mg)	48.0	0.60	223	43.6	37.4
mmol	0.05	1.41	0.544	0.04	0.07
Yield (%)	30	32	46	26	22

 Table S2.
 Synthesis of maleimide-RAFT agents (Mal-CTAs) via route 2: reaction conditions.

At the end of each reaction, the reaction mixture was purified *via* flash column chromatography (Agilent 971-FP Flash Purification System) according to the following condition. Compounds **11b** and **11c** were purified on reversed-phase (KP-C18-HS) as the stationary phase, and acetonitrile (from 5% to 20% and from 5% to 15%, respectively) in water as mobile phase.

All the other compounds were purified on SiO₂ as the stationary phase, mobile phases were as follow: petroleum ether/diethyl ether 1:1 (vol/vol) for **11a**; diethyl ether/petroleum ether 3:7 (vol/vol) for **12a**; diethyl ether/petroleum ether 2:8 (vol/vol) for **13a**; diethyl ether for **12b** and **13b**; ethyl acetate for **14b**.

Characterisation:

Mal-CTA **(11a)**.

¹**H NMR** (400 MHz, CDCl₃) δ 5.31-5.22 (m, 1H, NCH₂CHCH₂O), 5.15-5.09-4.99-4.93 (d, 1H, SC(S)SCH) , 4.31-4.10 (m, 2H, NCH₂CHCH₂O), 3.91 (t, 2H, HOCH₂CH₂), 3.86-3.74 (m, 2H, NCH₂CHCH₂O), 3.71 -3.70 - 3.69 (s, 3H, OCH₃), 3.61 (t, 2H, HOCH₂CH₂), 3.39-3.34 and 3.16-3.10 (1H, CHCHCH₃) 3.28-3.18 (1H, CHCHCH₃), 2.34-2.26 (m, 4H, C(O)CH₂(CH₂)₁₃CH₃), 1.59 (m, 4H, C(O)CH₂CH₂(CH₂)₁₂CH₃), 1.42-1.25 (m, 58H, (CH₂)₁₂CH₃, and CHCHCH₃), 0.88 (t, 6H, (CH₂)_nCH₃).

¹³**C NMR** (101 MHz, CDCl₃) δ 221.06-220.63 (1C, SC(S)S), 175.70-175.25 (1C, *C*(O)OCH₃), 173.92-172.30 (2C, CO_{Mal} and 2C, CO_{ester}), 68.77 (1C, NCH₂CHCH₂O), 62.91 (1C, NCH₂CHCH₂O), 60.30 (1C, HOCH₂CH₂), 52.56 (1C, OCH₃), 49.20 (1C, SC(S)SCH), 48.76 (1C, CHCHCH₃), 40.30 (1C, HOCH₂CH₂), 39.32-38.35 (1C, NCH₂CHCH₂O), 34.21 (2C, CHOC(O)CH₂(CH₂)₁₃CH₃), 32.07, 29.85-29.29 (24C, (*CH₂*)₁₂CH₃), 25.08-24.81 (2C, CH₂CH₂(CH₂)₁₂CH₃), 22.84 (1C, CHCHCH₃), 14.26 (2C, (CH₂)_nCH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 910.50, found 910.50 (100%).



igure S42. ¹H NMR spectrum of Mal-CTA (11a) in CDCl₃.



Figure S43. ¹H-¹H COSY NMR spectrum of Mal-CTA (11a) in CDCl₃.



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 f1(ppm)

Figure S44. ¹³C NMR spectrum of Mal-CTA (11a) in CDCl₃.



Figure S45. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (11a) in CDCl₃.

Mal-CTA (11b). Anti:Syn molar ratio: 95:5

¹**H NMR** (400 MHz, CDCl₃, anti adduct) δ 5.02 (d, 1H, *J* = 6.6 Hz, SC(S)SC*H*), 4.90 (d, 1H, , *J* = 6.6 Hz, SC(S)SC*H*), 3.89 (q, 2H, *J* = 5.9 Hz, HOC*H*₂CH₂S), 3.83-3.75 (m, 2H, NCH₂CH₂OC*H*₂), 3.71 (s, 3H, OCH₃), 3.70 (s, 3H, OCH₃), 3.70-3.65 (m, 4H, NCH₂C*H*₂OC*H*₂C*H*₂OH), 3.61-3.56 (m, 4H, HOCH₂C*H*₂S and NC*H*₂), 3.45-3.38 and 3.16-3.11 (m, 1H, CHC*H*CH₃, respectively *J* = 7.6, 3.9 Hz and *J* = 7.4, 3.7 Hz), 3.28-3.22 (td, 1H, *J* = 6.6, 3.7 Hz, CHCHCH₃), 2.85 (t, 1H, *J* = 6.2 Hz, OCH₂CH₂OH), 2.74 (t, 1H, *J* = 6.2 Hz, OCH₂CH₂OH), 2.31 (t, 1H, *J* = 5.7 Hz, HOCH₂CH₂CH₂S), 1.46 (d, 3H, *J* = 7.4 Hz, CHCHCH₃), 1.36 (d, 3H, *J* = 7.6 Hz, CHCHCH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃, anti adduct) δ 221.0 (1C, SC(S)S), 220.5 (1C, SC(S)S), 176.1 (1C,CO_{Mal}), 175.6 (1C,CO_{Mal}), 174.3 (1C,CO_{ester}), 174.2 (1C,CO_{ester}), 173.0 (1C,CO_{Mal}), 172.6 (1C,CO_{Mal}), 72.5 (1C, NCH₂CH₂OCH₂CH₂OH), 67.2 (1C, NCH₂CH₂OCH₂CH₂OH), 61.8 (1C, NCH₂CH₂O), 60.2 (1C, HOCH₂CH₂S), 52.7 (1C, OCH₃), 52.7 (1C, OCH₃), 50.2 (1C, SC(S)SCH), 49.3 (1C, SC(S)SCH), 48.8 (1C, CHCHCH₃), 48.7 (1C, CHCHCH₃), 40.3 (1C, HOCH₂CH₂S), 40.2 (1C, HOCH₂CH₂S), 39.5 (1C, NCH₂CH₂OCH₂CH₂OH), 39.3 (1C, CHCHCH₃), 38.5 (1C, CHCHCH₃), 15.2 (1C, CHCHCH₃), 14.9 (1C, CHCHCH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 448.05, found 448.05 (100%).



Figure S46. ¹H NMR spectrum of Mal-CTA **(11b)** in CDCl₃. f' and f" refer to the-CHSC(S)SZ proton of the trans (anti) and cis (syn) adducts, respectively.



Figure S47. ¹H-¹H COSY NMR spectrum of Mal-CTA (11b) in CDCl₃.



Figure S48. ¹³C NMR spectrum of Mal-CTA (11b) in CDCl₃.



Figure S49. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (11b) in CDCl₃.

Mal-CTA (11c). Anti:Syn molar ratio: 94:6

¹**H NMR** (400 MHz, CDCl₃, anti adducts) δ_{H} (400 MHz, CDCl₃) 4.94 (d, *J* = 6.4 Hz, 1H, SC(S)SC*H*), 4.86 (d, *J* = 6.5 Hz, 1H, SC(S)SC*H*), 4.42 (m, 1H, NCH), 4.06 (m, 2H, NCHC*H*₂OH) 3.91 (m, 2H,NCHC*H*₂OH overlapping t, *J* = 6.0 Hz, 4H, HOC*H*₂CH₂S), 3.75, 3.73, 3.72, 3.69 (4s, 3H, OCH₃), 3.61 (t, *J* = 6.0 Hz, 2H, HOCH₂C*H*₂S), 3.48-3.43 and 3.20- 3.13 (m, 1H, CHC*H*CH₃), 3.28-3.24 (m, 1H, CHCHCH₃), 3.07 (bs, 1H, NCHCH₂O*H*), 2.91 (bs, 1H, NCHCH₂O*H*), 2.08 (bs, 1H, HOCH₂CH₂S), 1.51 (d, *J* = 7.4 Hz, 3H, CHCHCH₃), 1.38 (d, *J* = 7.6 Hz, 3H, CHCHCH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃, anti adducts) δ 221.2 (1C, SC(S)S), 220.6 (1C, SC(S)S), 177.8 (1C,CO_{Mal}), 177.4 (1C,CO_{Mal}), 177.0 (1C,CO_{ester}), 174.5 (1C,CO_{ester}), 174.2 (1C,CO_{Mal}), 173.8 (1C,CO_{Mal}), 60.7 (1C, NCHCH₂O), 60.5 (1C, NCHCH₂O), 60.2 (1C, HOCH₂CH₂S), 57.3 (1C, NCH), 53.1 (1C, OCH₃), 52.9 (1C, OCH₃), 50.5 (1C, SC(S)SCH), 49.0 (1C, SC(S)SCH), 48.8 (1C, CHCHCH₃), 48.5 (1C, CHCHCH₃), 40.5 (1C, HOCH₂CH₂S), 40.2 (1C, HOCH₂CH₂S), 39.3 (1C, CHCHCH₃), 38.2 (1C, CHCHCH₃), 15.7 (1C, CHCHCH₃), 14.7 (1C, CHCHCH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 434.04, found 434.04 (100%).



Figure S50. Top: ¹H NMR spectrum of Mal-CTA **(11c)** in CDCl₃. Bottom: NOESY spectrum showing spatial proximity between protons f'_1 and f'_2 and the methyl group of the 2-methylpropionate

substituent (h'_1 and h'_2), confirming the *anti* (trans) structure of the major adduct. Carbon in position g is also chiral, resulting in the observed split of all signals, shown here as f'_1 and f'_2 for group f'.



Figure S51. ¹H-¹H COSY NMR spectrum of Mal-CTA (11c) in CDCl₃.



Figure S52. ¹³C NMR spectrum of Mal-CTA (11c) in CDCl₃.



Figure S53. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (11c) in CDCl₃.

Mal-CTA (12a).

¹**H NMR** (400 MHz, CDCl₃) δ 5.31-5.21 (m, 1H, NCH₂CHCH₂O), 5.16 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 5.09 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 5.00 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 4.93 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 4.31 - 4.10 (m, 2H, NCH₂CHCH₂O), 3.89 - 3.73 (m, 2H, NCH₂CHCH₂O), 3.70 (s, 3H, OCH₃), 3.69 (s, 3H, OCH₃), 3.69 (s, 3H, OCH₃), 3.69 (s, 3H, OCH₃), 3.68 (s, 3H, OCH₃), 3.35 (t, 2H, *J* = 7.4 Hz, CH₂SC(S)S), 3.27-3.18 (m, 1H, CHCHCH₃), 3.38 and 3.17 (m, 1H, CHCHCH₃), 2.34-2.24 (m, 4H, C(O)CH₂CH₂(CH₂)₁₂CH₃), 1.69 (m, 2H, CH₂CH₂SC(S)S), 1.59 (m, 4H, C(O)CH₂CH₂(CH₂)₁₂CH₃), 1.41-1.25 (bs, 66H, (CH₂)₁₂CH₃, (CH₂)₉CH₃) overlapping 1.35 (3H, d, *J* = 2.7 Hz, CHCHCH₃), 0.88 (t, 9H, *J* = 7.0 Hz, (CH₂)_nCH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 221.1 (1C, SC(S)S), 220.6 (1C, SC(S)S), 175.8 (1C, *C*(O)OCH₃), 175.3 (1C, *C*(O)OCH₃), 173.9 (1C, CO_{Mal}), 173.4 (2C, CO_{ester}), 172.5 (1C, CO_{Mal}), 68.8 (1C, NCH₂CH), 62.9 (1C, NCH₂CHCH₂O), 52.5 (1C, OCH₃), 52.5 (1C, OCH₃), 49.7 (1C, SC(S)SCH), 49,5 (1C, SC(S)SCH), 49,4 (1C, SC(S)SCH), 49.3 (1C, SC(S)SCH), 48.9 (1C, CHCHCH₃), 48.5 (1C, CHCHCH₃), 40.1 (1C, NCH₂CHCH₂O), 39.9 (1C, NCH₂CHCH₂O), 39.4 (1C, CHCH₃), 39.3 (1C, CHCH₃), 38.4 (1C, CH₂SC(S)S), 38.1 (1C, CH₂SC(S)S), 34.3 (1C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 34.2 (1C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 32.1 - 22.8 (33C, (CH₂)₁₂CH₃, (CH₂)₉CH₃), 27.9 (1C, CH₂CH₂SC(S)S), 25.0 (1C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 24.8 (1C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 15.1 (1C, CHCH₃), 14.8 (1C, CHCH₃), 14.7 (1C, CHCH₃), 14.3 (3C, (CH₂)_nCH₃).



Figure S55. ¹H-¹H COSY NMR spectrum (CDCl₃) of Mal-CTA (12a) in CDCl₃.



Figure S56. ¹³C NMR spectrum (CDCl₃) of Mal-CTA (12a) in CDCl₃.



Figure S57. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (12a) in CDCl₃.

Mal-CTA (12b). Anti:Syn molar ratio: 92:8

¹**H NMR** (400 MHz, CDCl₃, anti isomer) δ 5.05 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 4.94 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 3.84-3.65 (m, 2H, NCH₂CH₂OCH₂CH₂OH and m, 4H, NCH₂CH₂OCH₂CH₂OH) overlapping with 3.71 (s, 3H, OCH₃) and 3.70 (s, 3H, OCH₃), 3.57 (m, 2H, 2H, NCH₂CH₂O), 3.43-3.41 and 3.16-3.14 (m, 1H, *J* = 7.4, 3.7 Hz, CHCHCH₃), 3.35 (t, 2H, *J* = 7.4 Hz, CH₂SC(S)S), 3.25-3.21 (td, 1H, *J* = 6.6, 3.8 Hz, CHCHCH₃), 1.70 (m, 2H, *J* = 7.2 Hz, CH₂CH₂SC(S)S), 1.47 (d, 3H, *J* = 7.4 Hz CHCHCH₃), 1.38 (d, 3H, *J* = 7.4 Hz, CHCHCH₃), 1.42 – 1.21 (m and bs, 2H, 18H, CH₂CH₃), 0.88 (t, 9H, *J* = 6.9 Hz, CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃, anti isomer) δ 221.0 (1C, SC(S)S), 220.1 (1C, SC(S)S), 176.1 (1C,CO_{Mal}), 175.7 (1C,CO_{Mal}), 174.2 (1C,CO_{ester}), 174.2 (1C,CO_{ester}), 173.1 (1C,CO_{Mal}), 172.8 (1C,CO_{Mal}), 72.5 (1C, OCH₂CH₂OH), 67.2 (1C, OCH₂CH₂OH), 61.8 (1C, NCH₂CH₂), 52.7 (1C, OCH₃), 52.6 (1C, OCH₃), 49.9 (1C, SC(S)SCH), 49.5 (1C, SC(S)SCH), 48.9 (1C, CHCHCH₃), 48.6 (1C, CHCHCH₃), 39.5 (1C, NCH₂), 39.5 (1C, NCH₂CH₂OCH₂CH₂OH), 39.3 (1C, CHCHCH₃), 38.5 (1C, CHCHCH₃), 38.1 (1C, CH₂SC(S)S), 38.0 (1C, CH₂SC(S)S),32.0 - 22.8 (9C, (CH₂)₉CH₃), 27.8 (1C, CH₂CH₂SC(S)S), 15.3 (1C, CHCHCH₃), 14.9 (1C, CHCHCH₃), 14.3 (3C, (CH₂)₉CH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 572.22, found 572.21 (92.4%).



Figure S58. Top: ¹H NMR spectrum of Mal-CTA **(12b)** in CDCl₃. Bottom: NOESY spectrum showing spatial proximity between protons f'_1 and f'_2 and the methyl group of the 2-methylpropionate substituent (i'_1 and i'_2), confirming the *anti* (trans) structure of the major adduct. Carbon in position g is also chiral, resulting in the observed split of all signals, shown here as f'_1 and f'_2 for group f'.



Figure S59. ¹H-¹H COSY NMR spectrum of Mal-CTA (12b) in CDCl₃.





Figure S61. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (12b) in CDCl₃.

Mal-CTA (13a).

¹**H NMR** (400 MHz, CDCl₃) δ 5.31-5.21 (m, 1H, NCH₂CH), 4.92 (d, 1H, *J* = 6.6 Hz, SC(S)SCH) , 4.77 (d, 1H, *J* = 6.6 Hz, SC(S)SCH), 4.28-4.09 (m, 2H, CH₂O), 3.91-3.71 (m, 2H, NCH₂), 3.38-3.33 (m, 2H, CH₂SC(S)S), 3.20 (app. t, 1H, *J* = 6.5 Hz, CHC(CH₃)₂CN), 2.32 (t, 2H, *J* = 7.5 Hz, C(O)CH₂CH₂(CH₂)₁₂CH₃), 2.26 (t, 2H, *J* = 7.5 Hz, C(O)CH₂CH₂(CH₂)₁₂CH₃), 2.26 (t, 2H, *J* = 7.5 Hz, C(O)CH₂CH₂(CH₂)₁₂CH₃), 1.70 (m, 2H, CH₂CH₂SC(S)S), 1.63-1.51 (m overlapping with four s, 10H, C(O)CH₂CH₂(CH₂)₁₂CH₃, C(CH₃)₂CN), 1.39- 1.25 (bs, 66H, (CH₂)₁₂CH₃, (CH₂)₉CH₃), 0.87 (t, 9H, *J* = 6.8 Hz, CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 220.2 (1C, SC(S)S), 220.1 (1C, SC(S)S), 173.6 (1C,CO_{Mal}), 172.8 (1C,CO_{Mal}), 171.4 (1C, CO_{ester}), 171.3 (1C, CO_{ester}), 122.0 (1C, CN), 121.9 (1C, CN), 68.8 (1C, NCH₂CHCH₂O), 68.6 (1C, NCH₂CHCH₂O), 62.9 (1C, NCH₂CHCH₂O), 62.8 (1C, NCH₂CHCH₂O), 52.8 (1C, CHC(CH₃)₂CN), 52.7 (1C,CHC(CH₃)₂CN), 48.4 (1C, SC(S)SCH), 48.4 (1C, SC(S)SCH), 40.4 (1C, NCH₂CHCH₂O), 40.4 (1C, NCH₂CHCH₂O), 38.3 (1C, CH₂SC(S)S), 38.3 (1C, CH₂SC(S)S), 34.4-34.2 (2C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 34.0(1C, C(CH₃)₂CN), 32.1-22.8 (33C, (CH₂)₁₂CH₃, (CH₂)₉CH₃), 27.8 (1C, CH₂CH₂SC(S)S), 25.5 – 24.5 (2C, C(CH₃)₂CN), 25.0 (2C, C(O)CH₂CH₂(CH₂)₁₂CH₃), 14.3 (3C, (CH₂)_nCH₃).

ESI-TOF Mass spectrometry: expected m/z [M+Na]⁺ 1015.66, found 1015.73 (33%).



Figure S62. ¹H NMR spectrum of Mal-CTA (13a) in CDCl₃.



Figure S63. ¹H-¹H COSY NMR spectrum of Mal-CTA (13a) in CDCl₃.



240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 f1 (ppm)

Figure S64. ¹³C {¹H} NMR spectrum of Mal-CTA (13a) in CDCl₃.



Figure S65. ¹H-¹³C HSQC NMR spectrum of Mal-CTA (13a) in CDCl₃.

¹**H NMR** (400 MHz, CDCl₃) δ 4.72 (d, 1H, *J* = 6.2 Hz, SC(S)SCH), 3.87-3.74 (m, 2H, NCH₂CH₂O), 3.71-3.64 (m, 4H, CH₂OCH₂CH₂OH), 3.55 (t, 2H, *J* = 4.5 Hz NCH₂CH₂O), 3.35 (t, 2H, *J* = 7.5 Hz, CH₂SC(S)S), 3.22 (d, 1H, *J* = 6.2 Hz, CHC(CH₃)₂CN), 2.30 (t, 1H, *J* = 6.4 Hz, OCH₂CH₂OH), 1.70 (m, 2H, CH₂CH₂SC(S)S), 1.65 (s, 3H, C(CH₃)₂CN), 1.54 (s, 3H, C(CH₃)₂CN), 1.39 (m, 2H, CH₂CH₂CH₂SC(S)S), 1.25(bs, 16H, (CH₂)₈CH₃), 0.87 (t, 9H, *J* = 6.9 Hz, CH₃).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 220.0 (1C, SC(S)S), 173.1 (1C,CO), 171.5 (1C,CO), 121.9 (1C, CN), 72.5 (1C, OCH₂CH₂OH), 67.1 (1C, NCH₂CH₂O), 61.9 (1C, NCH₂CH₂O), 53.0 (1C, CHC(CH₃)₂CN), 48.6 (1C, SC(S)SCH), 39.5 (1C, NCH₂CH₂O), 38.3 (1C, CH₂SC(S)S), 34.8 (1C, C(CH₃)₂CN), 32.0-22.8 (9C, (CH₂)₉CH₃), 27.8 (1C, CH₂CH₂SC(S)S), 25.6 (1C, C(CH₃)₂CN), 24.9 (1C, (CH₃)₂CN), 14.3 (1C, (CH₂)₉CH₃).



Figure S66. ¹H NMR spectrum of Mal-CTA (13b) in CDCl₃.



Figure S67. ¹H-¹H COSY NMR spectrum (CDCl₃) of Mal-CTA (13b) in CDCl₃.



Figure S68. ¹³C NMR spectrum (CDCl₃) of Mal-CTA (13b) in CDCl₃.



Figure S69. ¹H-¹³C HSQC NMR spectrum (CDCl₃) of Mal-CTA (13b) in CDCl₃.

¹**H NMR** (400 MHz, CDCl₃) δ 4.71 (d, 1H, *J* = 6.2 Hz, SC(S)SCH), 3.93-3.88 (m, 2H, HOCH₂CH₂S), 3.83-3.75 (m, 2H, NCH₂CH₂O), 3.72-3.66 (m, 4H, CH₂OCH₂CH₂OH), 3.60 (t, 2H, *J* = 5.9 Hz, HOCH₂CH₂S), 3.56 (m, 2H, OCH₂CH₂OH), 3.22 (d, 1H, *J* = 6.2 Hz, CHC(CH₃)₂CN), 2.8 (bs, 1H, *J* = 6.4 Hz, OCH₂CH₂OH), 1.91 (bs, 1H, *J* = 5.7 Hz, HOCH₂CH₂S), 1.65 (s, 3H, C(CH₃)₂CN), 1.54 (s, 3H, C(CH₃)₂CN).

¹³C {¹H} NMR (101 MHz, CDCl₃) δ 220.0 (1C, SC(S)S), 172.9 (1C,CO), 171.4 (1C,CO), 121.8 (1C, CN), 72.5 (1C, OCH₂CH₂OH), 67.1 (1C, OCH₂CH₂OH), 61.9 (1C, NCH₂CH₂O), 59.9 (1C, HOCH₂CH₂S), 52.9 (1C, CHC(CH₃)₂CN), 48.9 (1C, SC(S)SCH), 39.9 (1C, CH₂SC(S)S), 39.5 (1C, NCH₂CH₂O), 35.1 (1C, C(CH₃)₂CN), 25.6 (1C, C(CH₃)₂CN), 24.9 (1C, (CH₃)₂CN).



Figure S70. ¹H NMR spectrum of Mal-CTA (14b) in CDCl₃.



Figure S71. ¹H-¹H COSY NMR spectrum (CDCl₃) of Mal-CTA (14b) in CDCl₃.



Figure S72. 13 C NMR spectrum (CDCl₃) of Mal-CTA (14b) in CDCl₃.



Figure S73. ¹H-¹³C HSQC NMR spectrum (CDCl₃) of Mal-CTA (14b) in CDCl₃.

RAFT polymerisation mediated by Mal-CTAs. General polymerisation procedure: maleimide-based RAFT agent (**Mal-CTA**), monomer, initiator (aliquot from stock solution), and solvent were introduced in a Schlenk tube along with a magnetic stirrer and the tube was sealed with a rubber septum. The mixture was cooled with an ice bath and was degassed by argon bubbling for 15 minutes. The reaction was started by placing the Schlenk tube in a thermostated oil bath at the desired temperature.

	/	ast	Ultrafast					
Polymer	(11a) - BA ₄₈	(12a) - BA ₃₄	(11b) - HEA ₄₃	(11b) - NAM ₄₉	(11b) - DMA ₃₉	(11b) - DMA ₉₇	(11b) - DMA ₁₈₄	(11b) - DMA ₄₇₅
	15	16	17	18	19	20	21	22
Monomer	BA	BA	HEA	NAM	DMA	DMA	DMA	DMA
$DP_{targeted}$	50	50	50	50	50	100	200	500
mg	152.9	134.1	148.8	182.5	128.1	256.3	256.3	640.6
mmol	1.193	1.046	1.293	1.293	1.293	2.585	2.585	6.462
Mal-CTA	(11a)	(12a)	(11b)	(11b)	(11b)	(11b)	(11b)	(11b)
mg	16.3	16.3	10.0	10.0	10.0	10.0	5.0	5.0
mmol	0.018	0.016	0.024	0.024	0.024	0.024	0.012	0.012
Initiator	AIBN	AIBN	VA-044	VA-044	VA-044	VA-044	VA-044	VA-044
mg	0.30	0.26	0.076	0.076	0.076	0.076	0.038	0.038
mmol	1.8·10 ⁻³	1.6·10 ⁻³	2.4·10 ⁻⁴	2.4·10 ⁻⁴	2.4.10-4	2.4.10-4	1.2·10 ⁻⁴	1.2·10 ⁻⁴
V _{total} (mL)	0.596	0.523	0.646	0.646	0.646	1.293	1.293	3.231
Solvent 1	DMF	DMF	H_2O	H_2O	H ₂ O	H_2O	H_2O	H_2O
V _{solvent 1 added} (mL)	0.425	0.373	0.307	0.290	0.308	0.616	0.616	1.54
Solvent 2	-	-	dioxane	dioxane	dioxane	dioxane	dioxane	dioxane
V _{solvent 2 added} (mL)	-	-	0.205	0.193	0.205	0.410	0.410	1.026
[CTA]/[initiator]	10	10	100	100	100	100	100	100
Temp. (°C)	80	80	100	100	75	75	75	75
Time (min)	90	90	15	15	15	15	15	15

 Table S3. Synthesis of (Mal-CTA)-polymers: reaction conditions.

BA = n-butyl acrylate, HEA = N-hydroxyethylacrylamide, NAM = 4-acryloylmorpholine, DMA = N, N-dimethylacrylamide.

At the end of the polymerization, polymers (**11a**)-BA₄₈ and (**12a**)-BA₃₄ (**14** and **15**) were precipitated in H₂O/MeOH; polymers (**11b**)-HEA₄₃, (**11b**)-NAM₄₉, (**11b**)-DMA₃₉, (**11b**)-DMA₉₇, (**11b**)-DMA₁₈₄, (**11b**)-DMA₄₇₅ (**16**, **17**, **18**, **19**, **20**, and **21**) were purified by dialysis (MWCO: 1 kDa) against water for 24 hours, with three water changes, followed by lyophilisation.



Figure S74. ¹H NMR spectrum of polymer 15 (11a)-BA₄₈ in CDCl₃.



Figure S75. SEC analysis of polymer 15 - (11a)- BA_{48} , using $CHCI_3$ as the mobile phase.



Figure S76. ¹H NMR spectrum of polymer 16 - (12a)-BA₃₄ in CDCl₃.



Figure S77. SEC analysis of polymer 16 - (12a)-BA₃₄, using CHCl₃ as the mobile phase.



Figure S78. ¹H NMR spectrum of polymer **17** - (**11b**)-HEA₄₃ in DMSO-d₆.



Figure S79. ¹H NMR spectrum of polymer 18 - (11b)-NAM₄₉ in DMSO-d₆.



Figure S80. ¹H NMR spectrum of polymer **19** - (**11b**)-DMA₃₉ in DMSO-d₆.



Figure S81. ¹H NMR spectrum of polymer 20 - (11b)-DMA₉₇ in DMSO-d₆.



Figure S82. ¹H NMR spectrum of polymer 21 - (11b)-DMA₁₈₄ in DMSO-d₆.



Figure S83. ¹H NMR spectrum of polymer 22 - (11b)-DMA₄₇₅ in DMSO-d₆.

RAFT polymerisation kinetics. The kinetic sampling experiment was performed as with the other polymerisations described in the general procedure. Briefly, a 4 mL test tube was charged with Mal-CTA **(13b)** (13.25 mg, 25 μ mol), DMA (0.5 g, 5 mmol), 1,4-dioxane (1.84 mL) and 140 μ L of an 4,4'-Azobis(4-cyanovaleric acid) stock solution (5 mg mL⁻¹, 700 μ g, 2.5 μ mol). A trace amount of 1,3,5-trioxane was added as an internal reference to monitor monomer conversions. The tube was fitted with an appropriate magnetic stir bar, sealed with a rubber septum and purged with N₂ for 15 minutes. The tube was then immersed in a preheated oil bath at 70°C. 100 μ L samples were withdrawn using a degassed gas tight syringe at 15, 30, 60, 120 and 360 min.



Figure S84 Polymerisation kinetics of DMA monomer mediated by CTA **(13b)**. (A) SEC chromatograms using DMF with 0.1% (wt%) LiBr additive. (B) Evolution of conversion with time. (C) Evolution of $M_{n,SEC}$ with conversion, black filled circles refer to $M_{n,SEC}$ and red filled circles $M_{n,th}$. (D) Pseudo-first order kinetic plot.

Conditions for methacrylate polymerisations. A 4 mL test tube was charged with either Mal-CTA **(13b)** (8.46 mg, 16 μ mol) or CTA **(7)** (5.5 mg, (16 μ mol), methyl methacrylate (MMA, 0.2 g, 2 mmol), 1,4-dioxane (0.69 mL) and 90 μ L of an 4,4'-azobis(4-cyanovaleric acid) stock solution (5 mg mL⁻¹, 440 μ g, 1.6 μ mol). The tube was fitted with an appropriate magnetic stir bar, sealed with a rubber

septum, and purged with N_2 for 15 minutes. The tube was then immersed in a preheated oil bath at 70 $^\circ C$ for 16 h.



Figure S85 SEC analysis of MMA polymerisations mediated by CTA (7) and CTA (13b) using DMF with 0.1% (wt%) LiBr additive.

Table S4 $M_{n,SEC}$ and D of MMA polymers prepared using CTAs (7) and (13b)

	$M_{n,SEC}$ (kg mol ⁻¹) ^a	Monomer conversion (%) ^b	\mathcal{D}^{a}
CTA (7)	13.2	84%	1.19
CTA (13b)	51.0	86%	2.05

^aDetermined by SEC, using DMF with 0.1% (wt%) LiBr as the mobile phase. ^bDetermined by ¹H NMR spectroscopy.

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