

## Electronic Supplementary Information

### A novel hemocompatible core@shell nanosystem for selective targeting and apoptosis induction in cancer cells

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## Electronic Supplementary Information

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## Characterization techniques

Thermogravimetric analysis (TGA) and differential thermal analysis (DTA) were performed in a Perkin Elmer Pyris Diamond TG/DTA analyser (Perkin Elmer, California, USA) by placing approximately 5 mg of sample in a platinum crucible applying 5 °C/min heating ramps from room temperature to 800 °C under a flow rate of 100 mL/min of air. Chemical microanalyses were performed with a Perkin Elmer 2400 CHN and LECO CHNS-932 thermoanalyzers.

Fourier transformed infrared (FTIR) spectra were collected in a Thermo Nicolet Nexus spectrometer equipped with a Goldengate attenuated total reflectance (ATR) device.

The powder X-ray diffraction (XRD) measurements were performed in a Philips X'Pert diffractometer with Bragg-Brentano geometry operating with Cu K $\alpha$  radiation (wavelength 1.5406 Å) at 40 kV and 20 mA (Philips Electronics NV, Eindhoven, Netherlands). Low-angle XRD patterns were collected in the 2 $\theta$  range between 0.6° and 8° with a step size of 0.02° and contact time of 5 s per step. High-angle XRD patterns were collected in the 2 $\theta$  range between 15° and 100° with a step size of 0.02 s and contact time of 1 s per step.

Transmission Electron Microscopy (TEM) and energy dispersive X-ray spectroscopy (EDS) were carried out with a JEOL JEM 1400 instrument operated at 120 kV (JEOL Ltd., Tokyo, Japan). Sample preparation was performed by dispersing *ca.* 1 mg of sample in 1 mL of water followed by sonication in a low power sonicator bath (Selecta, Spain) for 5 min, and then depositing one drop of the suspension onto carbon-coated copper grids.

The textural properties of the materials were determined by N<sub>2</sub> adsorption porosimetry by using a Micromeritics ASAP 2020 (Micromeritics Co. Norcross, USA). To perform the N<sub>2</sub> measurements, 30 mg of each sample was previously degassed under vacuum for 24 h at 40 °C. The surface area ( $S_{\text{BET}}$ ) was determined using the Brunauer-Emmett-Teller (BET) method and the pore volume ( $V_{\text{p}}$ ) was estimated from the amount of N<sub>2</sub> adsorbed at a relative pressure of 0.98. The pore sized distribution between 0.5 and 40 nm was calculated from the desorption branch of the isotherm by means of the Barrett-Joyner-Halenda (BJH) method and the average mesopore size ( $D_{\text{p}}$ ) was determined from the maximum of the pore size distribution curve.

Electrophoretic mobility measurements for the materials suspended in water were used to calculate the zeta-potential ( $\zeta$ ) values of the nanosystems. Measurements were performed in a Zetasizer Nano ZS (Malvern Instruments Ltd., United Kingdom) equipped with a 633 nm “red” laser. For this purpose, 1 mg of nanoparticles were added to 10 mL of water followed by vortex and ultrasound to get a homogeneous suspension. Measurements were recorded by placing 1 mL of the suspension in a DTS1070 disposable folded capillary cells (Malvern Instruments). The hydrodynamic size of the nanoparticles was measured by dynamic light scattering (DLS) with the same Malvern instrument. Values presented are mean  $\pm$  SD from quintuplicate measurements.

**Table S1.** References of TaqMan gene expression assays used for the RT-qPCR analysis.

<b>Gene</b>	<b>Codified protein</b>	<b>RefSeq</b>	<b>Assay ID</b>
<b>TP53</b>	p53	NM_000546.5	Hs01034249_m1
<b>CDK2</b>	Cdk2	NM_001290230.1	Hs01548893_m1
<b>CCNE1</b>	Cyclin E	NM_001238.3	Hs01026536_m1
<b>CDKN1A</b>	p21	NM_000389.4	Hs00355782_m1
<b>CDK1</b>	Cdk1	NM_001170406.1	Hs00938777_m1
<b>CCNB1</b>	Cyclin B1	NM_031966.3	Hs01030099_m1
<b>CCNA2</b>	Cyclin A	NM_001237.3	Hs00996788_m1
<b>C-MYC</b>	c-Myc	NM_002467.4	Hs00153408_m1
<b>GAPDH</b>	Gapdh	NM_001256799.2	Hs03929097_g1

**Table S2.** Organic content and elemental composition from thermogravimetric and chemical analysis of Ag@MSNs and functionalized Ag@MSNs materials.

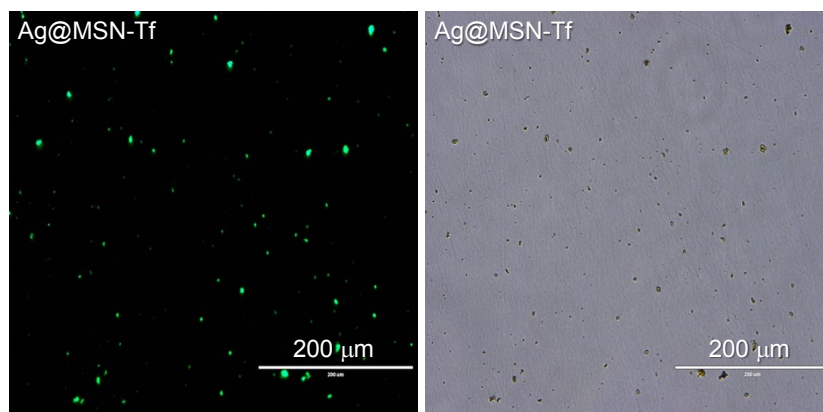
Material	Org. Content (wt %)	%C	%N	%S
Ag@MSNs	5.33 <sup>a</sup>	2.13	0.12	0.00
Ag@MSNs-COOH <sub>ext</sub>	6.95 <sup>a,b</sup>	6.53	0.33	0.01
Ag@MSNs-Tf	15.49 <sup>a,b</sup>	11.01	2.08	0.13

<sup>a</sup> Organic content (wt%) is determined from the TGA weight losses, excluding the weight loss due to the desorption of water (up to 125 °C) and <sup>b</sup> further corrected by the weight loss of the remaining alkoxy silanes after the sol-gel reaction (surfactant extracted unmodified Ag@MSNs).

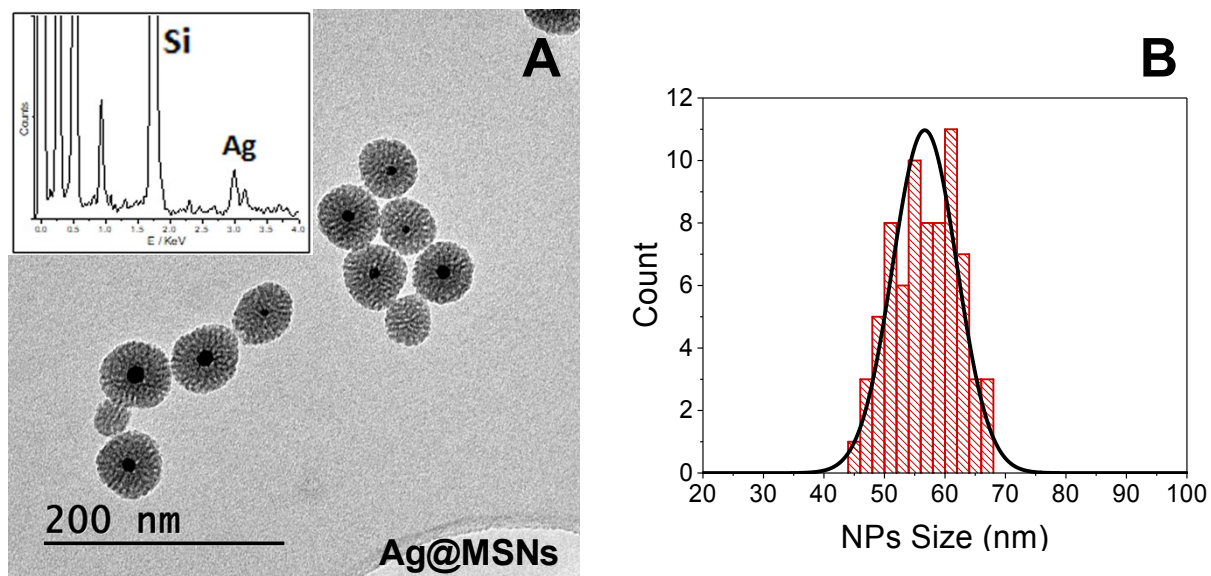
**Table S3.**  $\zeta$ -Potential values and hydrodynamic particle size in water medium of Ag@MSNs and functionalized Ag@MSNs materials.

<b>Material</b>	<b><math>\zeta</math>-Potential <sup>a</sup> (mV)</b>	<b>Hydrodynamic size <sup>a,b</sup> (nm)</b>
<b>Ag@MSNs</b>	$-14 \pm 3$	$68 \pm 6$
<b>Ag@MSNs-COOH<sub>ext</sub></b>	$-26 \pm 6$	$59 \pm 5$
<b>Ag@MSNs-Tf</b>	$-20 \pm 6$	$79 \pm 8$

<sup>a</sup> Samples were measured in quintuplicate (n = 5). <sup>b</sup> Maximum of the size distribution measured by dynamic light scattering.



**Figure S1.** Fluorescence microscopy image of Ag@MSNs-Tf nanosystem (left). The bright field image of the same region is shown of the right image. To take the images a suspension of the nanosystem in ethanol is deposited in a well and left to dry before examination under the microscope. Fluorescein green fluorescence from the nanoparticles aggregates, which are formed during solvent evaporation, can be observed.

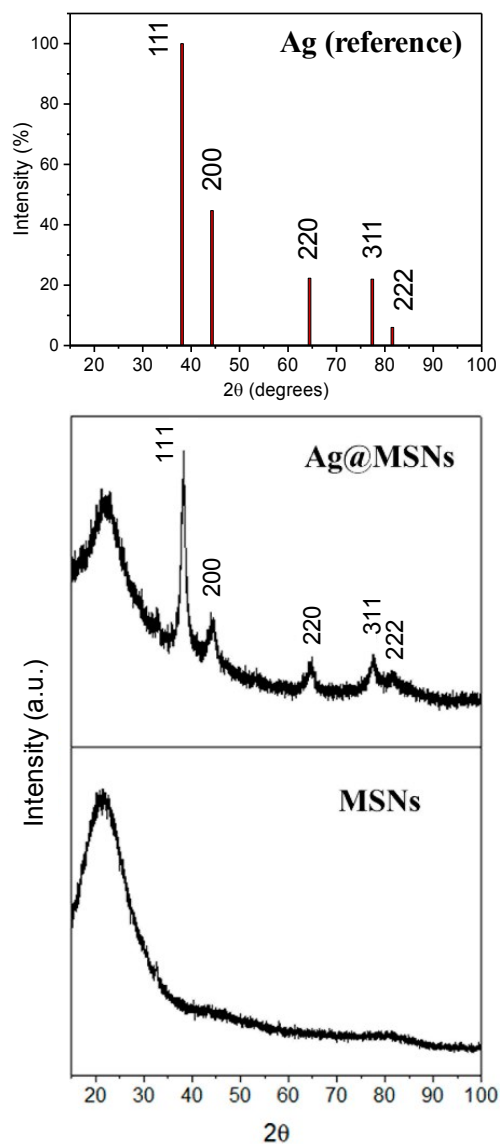


**Figure S2.** A) TEM micrograph of Ag@MSNs, EDS analysis of Ag@MSNs is shown as an inset. B) Histogram of measured nanoparticle diameters ( $n = 73$ ) in the TEM images with the best-fit lognormal distribution superimposed. The normality test using the Kolmogorov-Smirnov approximation gives a p-value of 0.91563, which indicates that at the 0.05 level the data are significantly drawn from a normally distributed population.

The polydispersity index calculated as the square of the relation between the width of the distribution and the mean value of the distribution has a value of 0.138. Therefore, it can be considered that the size distribution of Ag@MSNs nanosystem, with a value less than 0.2 and close to 0.1, tends to be highly monodisperse because a sample is considered monodisperse when the PDI value is less than 0.1.

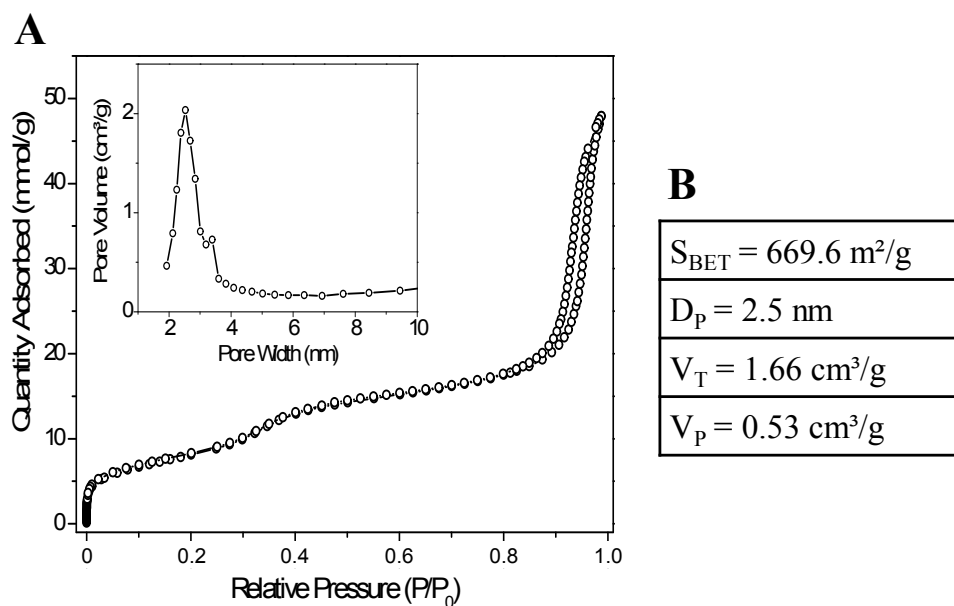
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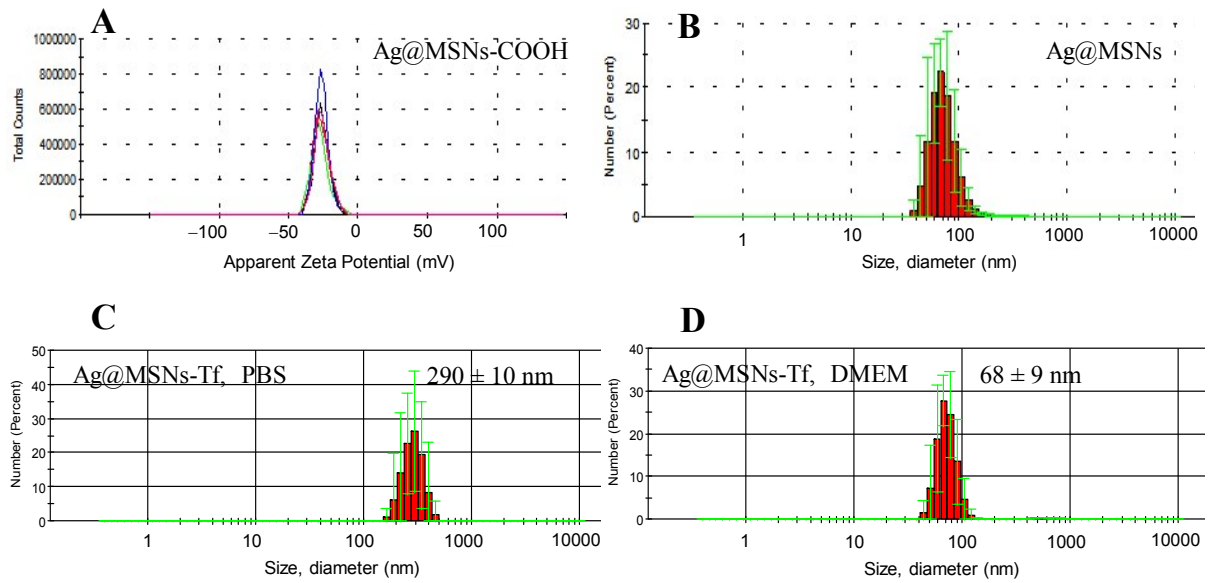


**Figure S3.** A) High angle powder X-ray diffraction pattern of the starting material Ag@MSNs and silver diffraction pattern reference for direct comparison. High angle powder X-ray diffraction pattern of mesoporous silica nanoparticles (MSNs) is also displayed (bottom) to assess that the broad signal appearing between 20 and 30 ° in the diffractogram of Ag@MSNs material is ascribed to the amorphous silica network.





**Figure S4.** A)  $\text{N}_2$  adsorption-desorption isotherms of the Ag@MSN material. Inset: Pore-size distribution for the mesoporous sample. B) Textural parameters of the Ag@MSN material obtained by  $\text{N}_2$  sorption measurements, where  $S_{\text{BET}}$  is the specific surface area obtained by using the BET equation,  $D_{\text{p}}$  is the pore diameter calculated by using the BJH method,  $V_{\text{T}}$  is the total pore volume obtained at  $P/P_0 = 0.99$  and  $V_{\text{p}}$  is the total pore volume obtained at  $P/P_0 = 0.60$ .



**Figure S5.** A) Zeta potential analysis (five independent measurements) and B) hydrodynamic size distribution obtained by dynamic light scattering of Ag@MSNs-COOH and Ag@MSNs materials suspended in water media as representative examples, respectively. Hydrodynamic size distribution of Ag@MSNs-Tf obtained by dynamic light scattering suspended in PBS (C) and cell culture media DMEM (D).