

Electronic Supplementary Information

Experimental Section

Materials: $\text{Cu}(\text{NO}_3)_2 \cdot x\text{H}_2\text{O}$ (99.9%), S (99.9%), $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ (99.0%), HCl (99.0%), $\text{CH}_4\text{N}_2\text{S}$, $(\text{CH}_2\text{OH})_2$, thioacetamide, N-(1-naphthyl), sulfanilamide, para-(dimethylamino) benzaldehyde, salicylic acid, NaClO (5.5%), NaOH, LiNO_3 , CH_3COCH_3 , $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$, Nafion (5.0% wt), $\text{Na}_2[\text{Fe}(\text{CN})_5\text{NO}] \cdot 2\text{H}_2\text{O}$, $\text{N}_2\text{H}_4 \cdot \text{H}_2\text{O}$, NH_4Cl , H_2SO_4 , HCl (99.0%), H_2O_2 (5.0%) and $\text{C}_2\text{H}_5\text{OH}$ were obtained from Aladdin Ltd. (Shanghai, China). The ultrapure water was purified through a Millipore process. Ultra-pure Ar (99.999%) and N_2 (99.999%) were obtained from Chengdu Guoguang Chemical Co., Ltd. All reagents were analytical grade and did not further purify.

Preparation of CuS concave polyhedral superstructures (CuS-CPSs): 0.5 mmol $\text{Cu}(\text{NO}_3)_2 \cdot x\text{H}_2\text{O}$ and 1 mmol sulfur powder were added into a 40 mL Teflon-lined stainless steel autoclave. Then add 32 mL of anhydrous ethanol and stir vigorously. The autoclave was kept at 180 °C for 12 h and then cooled naturally to the room temperature. Finally, the black precipitate was centrifuged and washed three times with deionized water and ethanol, then dried in vacuum at 60 °C.¹

Preparation of CuS-CPSs/CP: Carbon paper (CP) was pretreated in 0.05 M H_2SO_4 , and then was sequentially cleaned in ethanol and deionized water several times by sonication. 10 mg CuS-CPSs and 40 μL 5 wt% Nafion solution were dispersed in 960 μL ethanol/water ($V_{\text{ethanol}}/V_{\text{water}} = 3/1$) followed by 20 min sonication to form a homogeneous ink. 10 μL ink was loaded onto a CP ($1 \times 1 \text{ cm}^2$) and dried under room temperature (CuS-CPSs loading: 0.1 mg cm^{-2}). The sample was obtained as CuS-CPSs/CP.

Preparation of CuS nanoparticles (CuS-NPs): 0.5 g of $\text{Cu}(\text{NO}_3)_2 \cdot x\text{H}_2\text{O}$, 0.7 g of $\text{CH}_4\text{N}_2\text{S}$, and 20 mL of $(\text{CH}_2\text{OH})_2$ were mixed in a beaker. The system was under initial vacuum and then nitrogen gas was introduced to minimize the exposure to air. The temperature was kept at 110 °C for ten minutes. A mixture of 5 mL of 1 M NaOH and 5 mL of $(\text{CH}_2\text{OH})_2$ was injected into the beaker. The solution was then held for an additional five minutes before cooling down to room temperature. The CuS-NPs

were separated by centrifuging for 10 min at 10000 rpm. The CuS-NPs were washed five times with water quickly to remove the excessive ions and precursors.

Preparation of CuS nanosheet assemblies (CuS-NSAs): CuS-NSAs were prepared by a simple microwave-assisted heating method. First, 4 mmol $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ and 5 mmol thioacetamide were mixed with deionized water (80 mL) and ethanol (40 mL) in a 250 mL beaker. Then, the mixture was stirred vigorously for 30 min at room temperature. Next, the synthetic reaction was carried out in a microwave reactor at 500 W for 10 min. After cooling to room temperature, large amounts of gray precipitates were obtained. Finally, the as-synthesized samples were separated by a centrifuge, washed five times with deionized water and ethanol, followed by vacuum drying for 12 h.

Characterizations: Power XRD data were acquired by a LabX XRD-6100 X-ray diffractometer with a $\text{Cu K}\alpha$ radiation (40 kV, 30 mA) of wavelength 0.154 nm (SHIMADZU, Japan). The absorbance data of spectrophotometer was measured on SHIMADZU UV-1800 UV-vis spectrophotometer. SEM images were collected on a Gemini SEM 300 scanning electron microscope (ZEISS, Germany) at an accelerating voltage of 5 kV. TEM images were acquired on a HITACHI H-8100 electron microscopy (Hitachi, Tokyo, Japan) operated at 200 kV. XPS measurements were performed on an ESCALABMK II X-ray photoelectron spectrometer using Mg as the exciting source.

Electrochemical measurements: In this paper, we use an H-type electrolytic cell separated by a Nafion membrane which was protonated by boiling in ultrapure water, H_2O_2 (5%) aqueous solution and 0.5 M H_2SO_4 at 80 °C for another 2 h, respectively. A three-electrode configuration is used for electrochemical experiments using catalyst coated carbon paper as the working electrode, Ag/AgCl (filled with saturated KCl solution) as the reference electrode, and graphite rod as the counter electrode. The electrochemical experiments were carried out with an electrochemical workstation (CHI 660E) in N_2 -saturated 0.1 M HCl solution. The potentials reported in this work were converted to reversible hydrogen electrode (RHE) scale via calibration with the following equation: $E_{\text{RHE}} = E_{\text{Ag/AgCl}} + 0.197 \text{ V} + 0.059 \times \text{pH}$ and the presented current density was normalized to the geometric surface area.

Determination of NH₃: Concentration of produced NH₃ was spectrophotometrically determined by the indophenol blue method.² 2 mL sample was removed from the cathodic chamber, then added into 1 mL of oxidizing solution containing NaClO ($p_{Cl} = 4 \sim 4.9$), then added 2 mL coloring solution containing 1 M NaOH solution, 5% C₇H₆O₃ and 5% C₆H₅Na₃O₇·2H₂O, and 200 μL catalyst solution (0.1 g Na₂[Fe(CN)₅NO]·2H₂O diluted to 10 ml with deionized water) in turn. Absorbance measurements were performed after one hour at $\lambda = 655$ nm. The concentration-absorbance curves were calibrated using standard NH₃ solution with a series of concentrations. The standard curve ($y = 0.385x + 0.048$, $R^2 = 0.999$) shows good linear relation of absorbance value with NH₃ concentration by three times independent calibrations.

Determination of N₂H₄: N₂H₄ was estimated by the method of Watt and Chrisp.³ The color reagent was a mixed solution of 5.99 g C₉H₁₁NO, 30 mL HCl and 300 mL C₂H₅OH. In detail, 5 mL electrolyte was removed from the electrochemical reaction vessel, and added into 5 mL prepared color reagent and stirred 15 min at 25 °C. The absorbance of such solution at the absorbance of 455 nm was measured to quantify the hydrazine yields with a standard curve of hydrazine ($y = 0.469x + 0.0523$, $R^2 = 0.999$).

Determination of NO₃⁻: The content of the NO₃⁻ in electrolyte can be quantitatively determined by spectrophotometer measurement.⁴ 0.10 mL of 1.0 M HCl were added into 5.0 mL of standard LiNO₃ solution. After standing for 5 min, the concentration of NO₃⁻ was measured using UV-vis spectrophotometer at wavelength range from 200 to 300 nm. The standard curve ($y = 0.084x + 0.001$, $R^2 = 0.999$) shows good linear relation of absorbance value with NO₃⁻ concentration by three times independent calibrations.

Determination of NO₂⁻: Spectrophotometer measurement⁴ was utilized to estimate the content of the NO₂⁻. In detail, 0.10 mL of sulfanilamide solution (0.50 g of sulfanilamide was dissolved in 50.0 mL of 2.0 M HCl solution) was added into 5.0 mL of electrolyte, then adding 1 mg/mL N-(1-naphthyl) solution in turn. Stand for 30 min at room temperature. The UV-vis spectrophotometer at wavelength ranges from 450 to 650 nm. The absorbance of such solution at 545 nm was measured to quantify the NO₃⁻ concentration by a standard curve ($y = 1.777x + 0.007$, $R^2 = 0.999$).

Determination of FE and V_{NH_3} : FE was calculated by equation:

$$\text{FE} = [3F \times c_{\text{NH}_3} \times V / (17 \times Q)] \times 100\%$$

NH_3 yield (V_{NH_3}) was calculated using the following equation:

$$V_{\text{NH}_3} = (c_{\text{NH}_3} \times V) / (t \times m)$$

The mass of NH_3 was calculated as follows:

$$m_{\text{NH}_3} = c_{\text{NH}_3} \times V$$

Where F is the Faraday constant (96500 C mol^{-1}); c_{NH_3} is the measured NH_3 concentration; Q is the total quantity of applied electricity; V is the volume of the HCl electrolyte for NH_3 collection (35 mL); t is the electrolysis time (2 h) and m is the catalyst loading mass (0.1 mg).

Computational methods

The spin-polarized first-principles DFT calculations were performed using the Vienna Ab initio Simulation Package (VASP).⁵⁻⁷ The Perdew-Burke-Ernzerhof (PBE) exchange-correlation functional⁸ within the generalized gradient approximation (GGA) was employed to describe the exchange-correlation energy. The projector-augmented-wave (PAW) method⁹ was adopted for the pseudopotentials. The energy cutoff for the plane wave basis expansion was set to 450 eV. The force on each atom was set as 0.02 eV/Å for convergence criterion. Slab model was constructed in a $1 \times 4 \times 1$ supercell with three layers of CuS (103), where the bottom layer was fixed. A vacuum layer of 15 Å in the z direction avoids the interaction between layers. The Brillouin-zone integration was sampled by single Γ point.¹⁰

The free energies of the NRR steps were calculated using the equation:¹¹

$$\Delta G = \Delta E_{\text{DFT}} + \Delta E_{\text{ZPE}} - T\Delta S$$

Where ΔE_{DFT} is the DFT calculated binding energy, the ΔE_{ZPE} and $T\Delta S$ are the zero-point energy correction and the variation of entropy, respectively, which gotten from vibration calculations.¹²

The surface energy of CuS can be evaluated by σ , which is defined as:

$$\sigma = (E_{(\text{sur})} - N \times E_{(\text{bulk})}) \times \frac{1}{2A}$$

in which $E_{(\text{sur})}$ and $E_{(\text{bulk})}$ are the energies of all research surface and bulk CuS, respectively. N is the number of bulk structure. A represents the surface area of each surface.

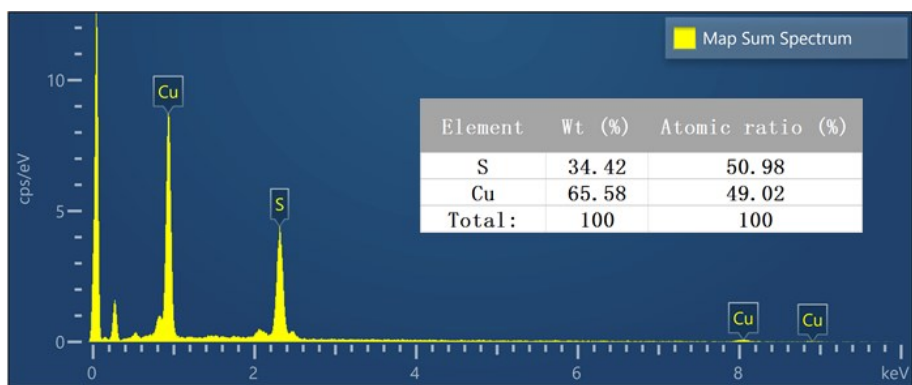


Fig. S1. EDX spectrum of CuS-CPSs.

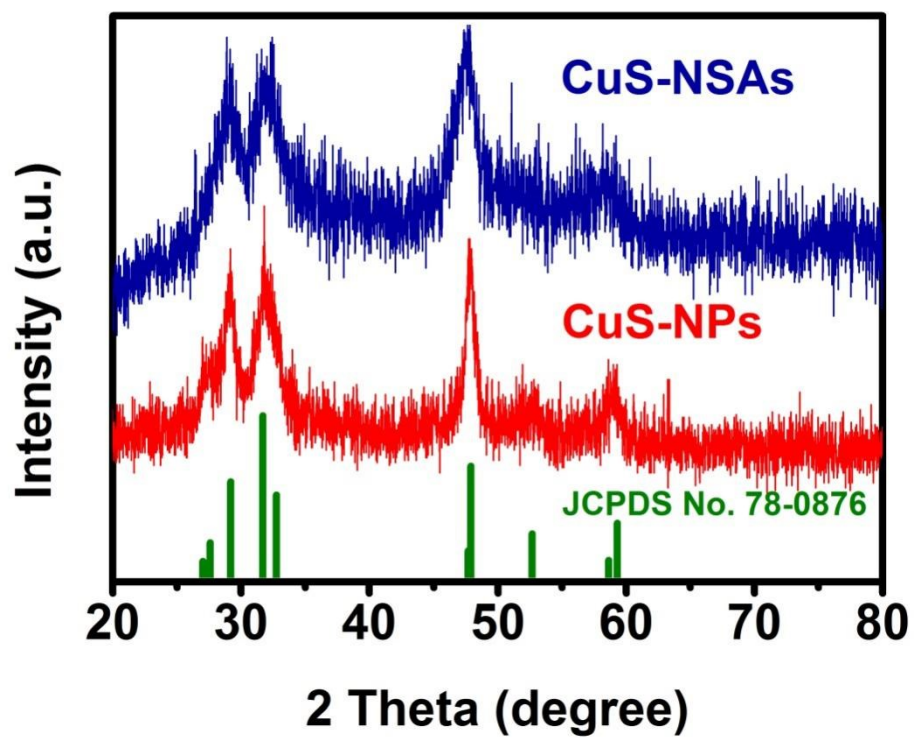


Fig. S2. XRD patterns of CuS-NPs and CuS-NSAs.

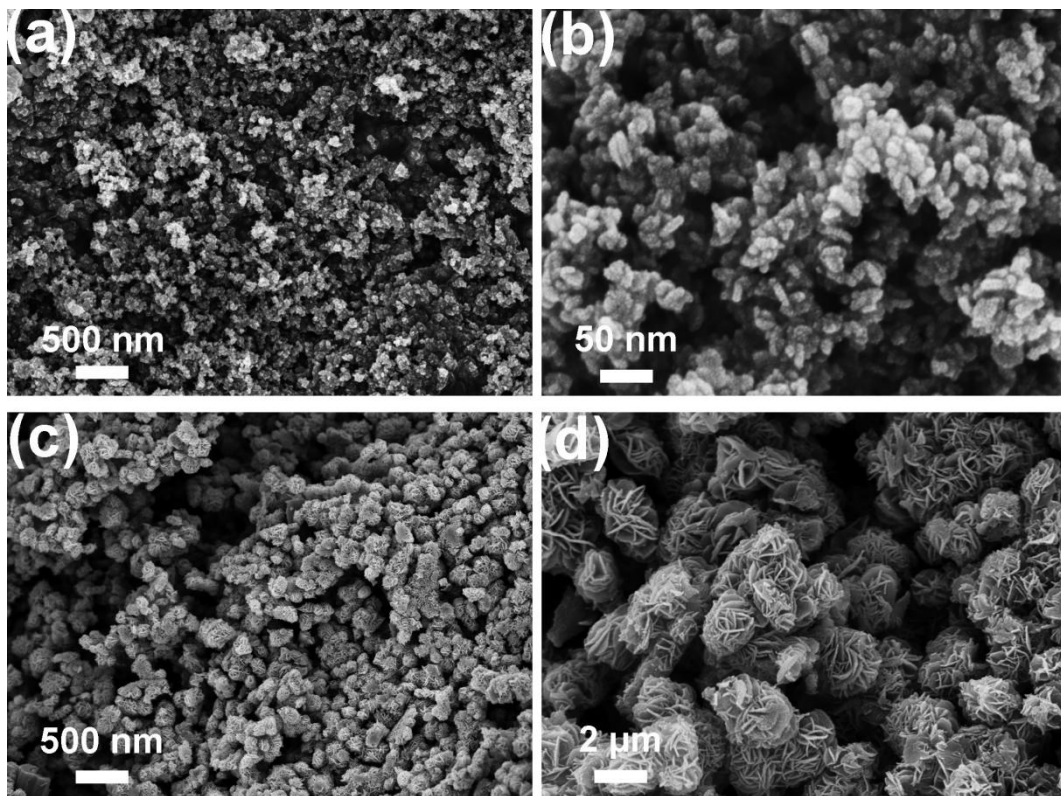


Fig. S3. SEM images of (a,b) CuS-NPs and (c,d) CuS-NSAs.

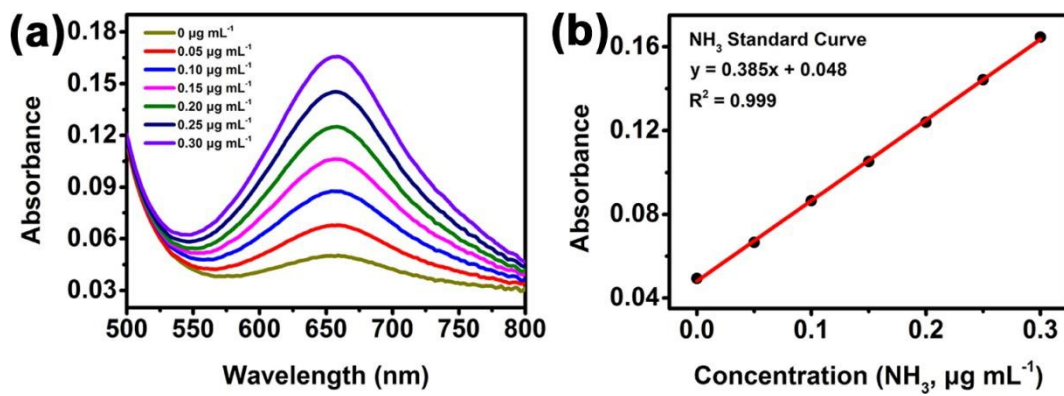


Fig. S4. (a) UV-vis absorption spectra of indophenol assays with NH_3 concentrations after incubated for 2 h at room temperature. (b) Calibration curve used for calculation of NH_3 concentrations.

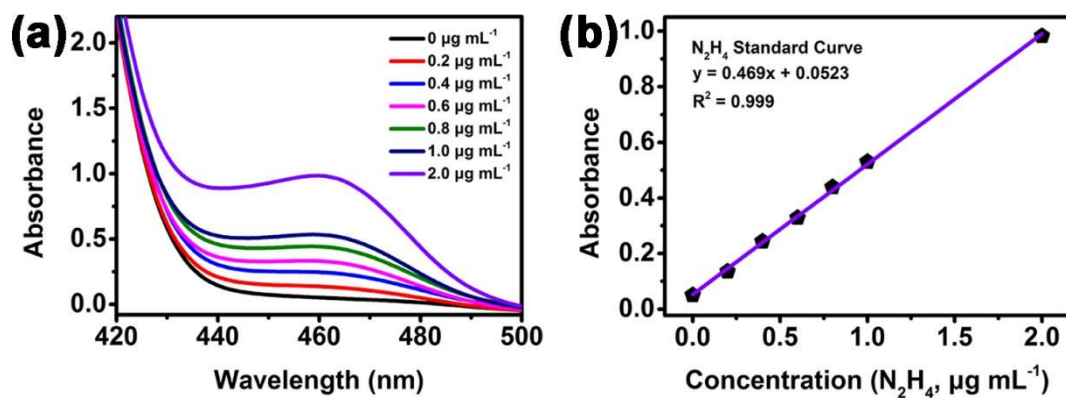


Fig. S5. (a) UV-vis absorption spectra of N_2H_4 concentrations after incubated for 20 min at room temperature. (b) Calibration curve used for calculation of N_2H_4 concentrations.

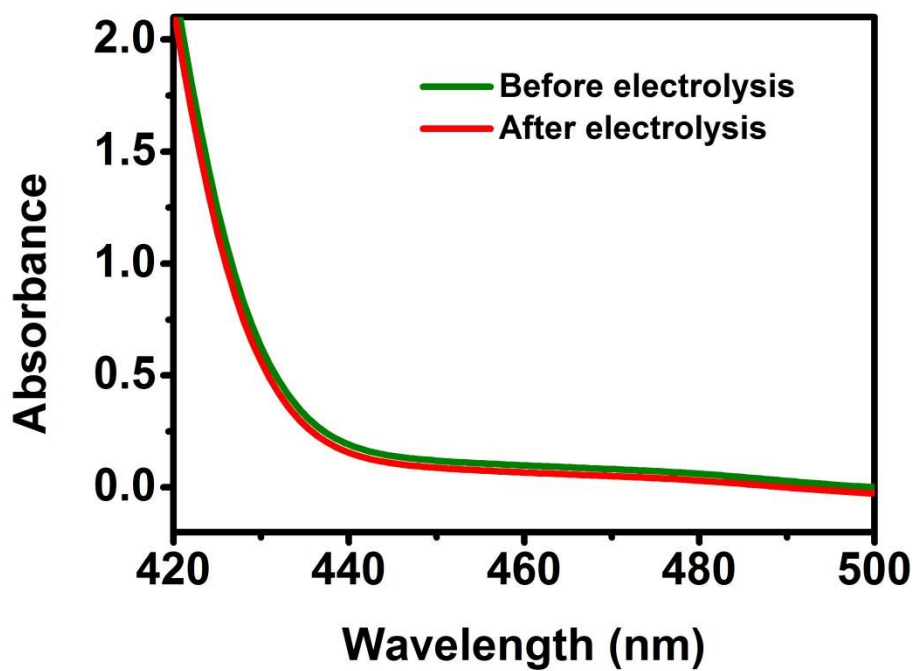


Fig. S6. UV-vis absorption spectra of electrolytes stained with para-(dimethylamino) benzaldehyde indicator before and after 2 h electrolysis at -0.15 V.

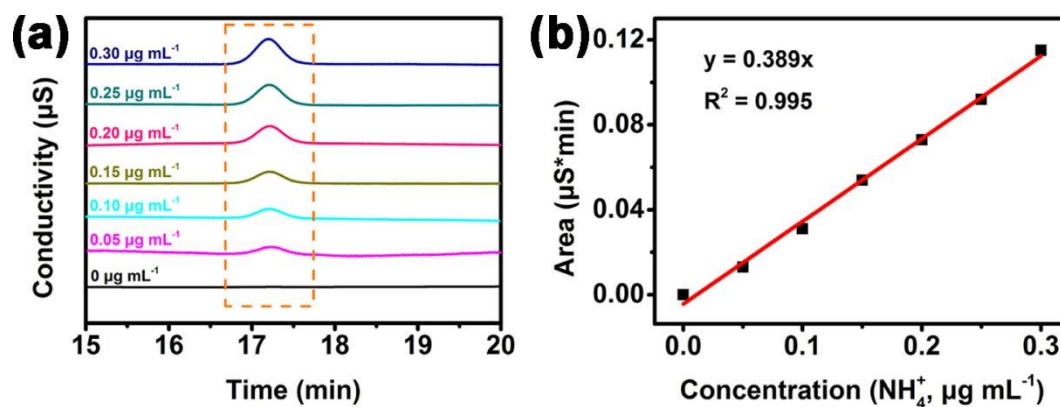


Fig. S7. (a) Ion chromatograms of NH_4^+ with different concentrations in 0.1 M HCl and (b) corresponding standard curve.

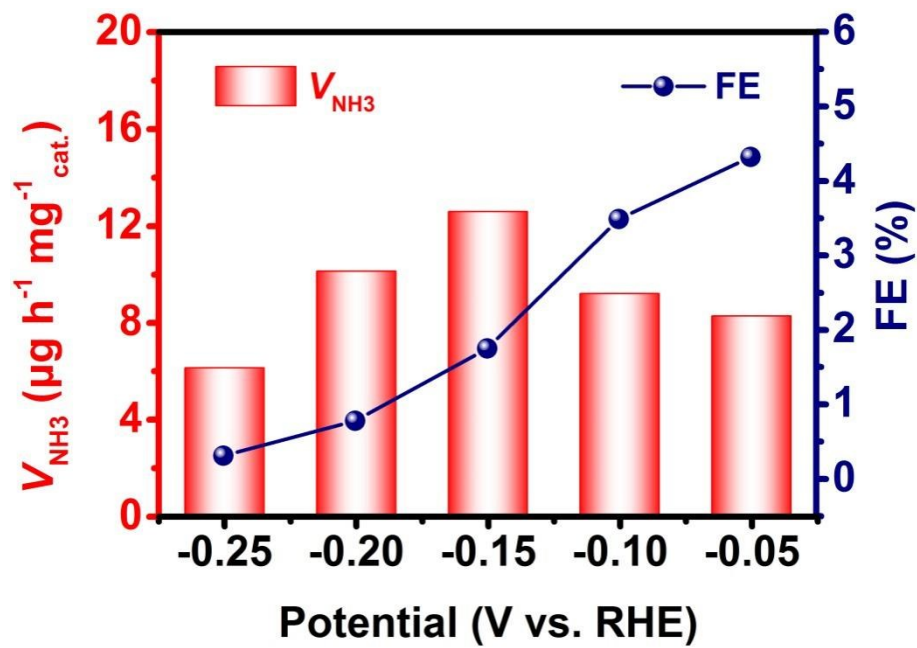


Fig. S8. V_{NH_3} and FE for CuS-CPSs/CP tested in 0.1 M Na_2SO_4 .

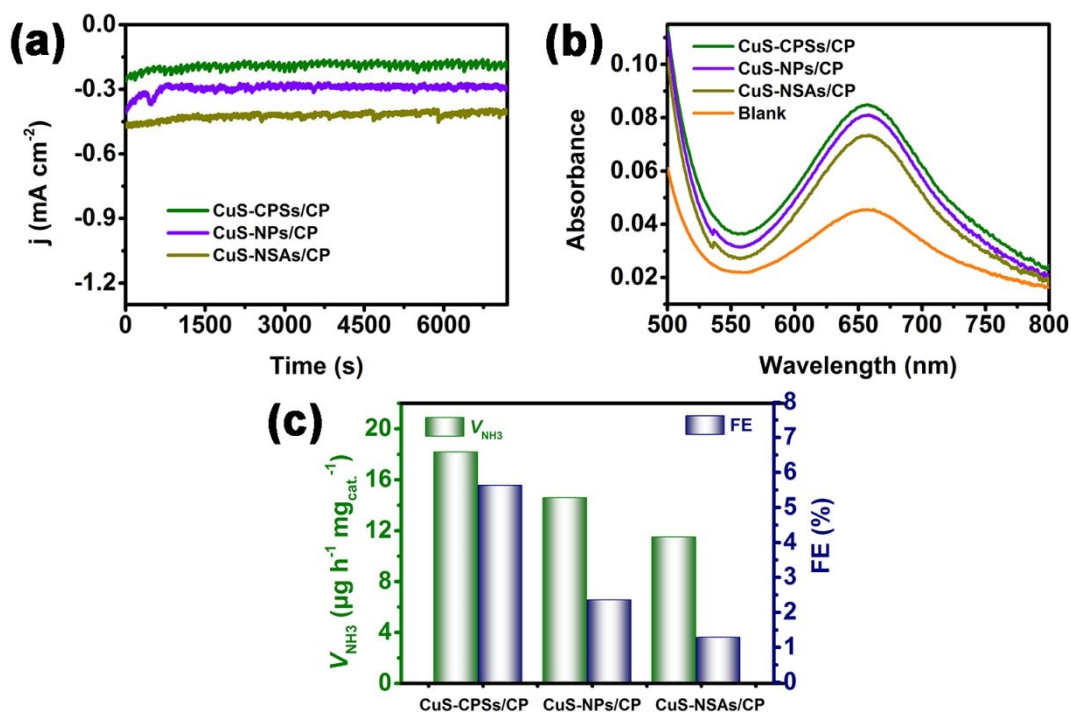


Fig. S9. (a) Time-dependent current density curves for CuS-CPSs/CP, CuS-NPs/CP and CuS-NSAs/CP at -0.15 V. (b) UV-vis absorption spectra of the electrolytes stained with indophenol indicator after 2 h electrolysis using CuS-CPSs/CP, CuS-NPs/CP and CuS-NSAs/CP. (c) Comparison of V_{NH_3} and FE for CuS-CPSs/CP, CuS-NPs/CP and CuS-NSAs/CP.

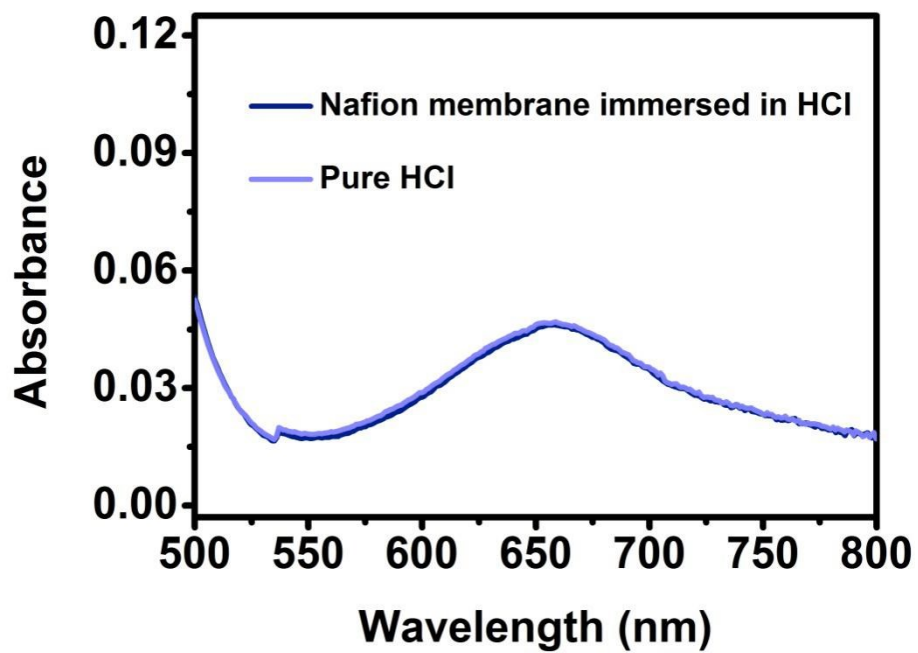


Fig. S10. Comparison of UV-vis absorption spectra of Nafion membrane immersed in 0.1 M HCl for 2 h and pure 0.1 M HCl.

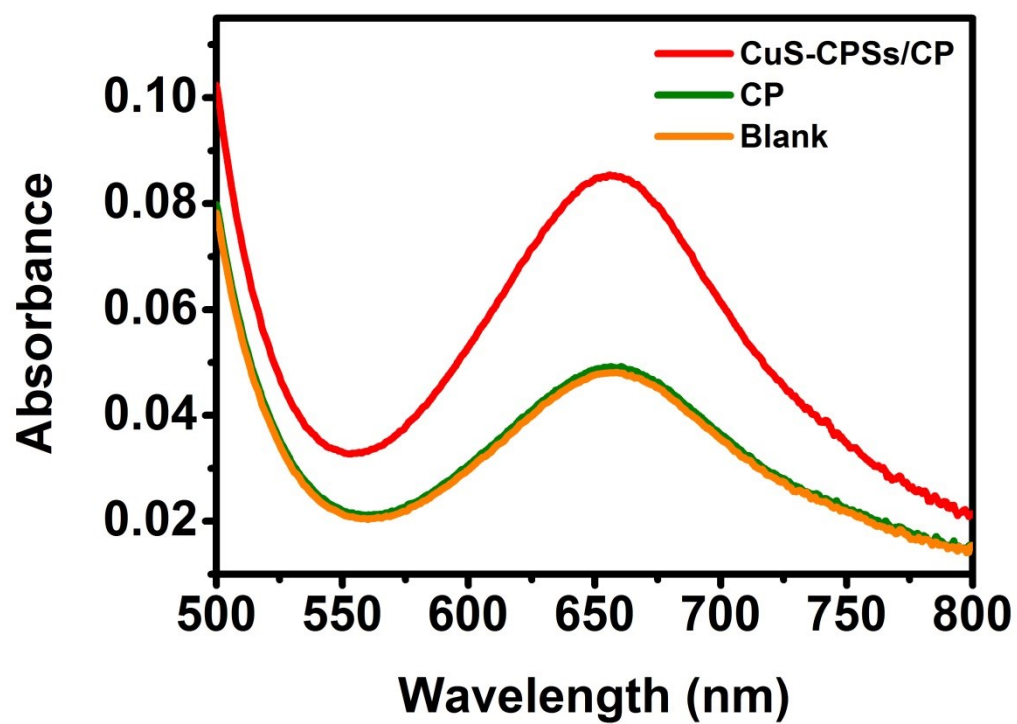


Fig. S11. UV-vis absorption spectra of the electrolytes colored with the indophenol indicator after NRR electrolysis at different electrodes for 2 h.

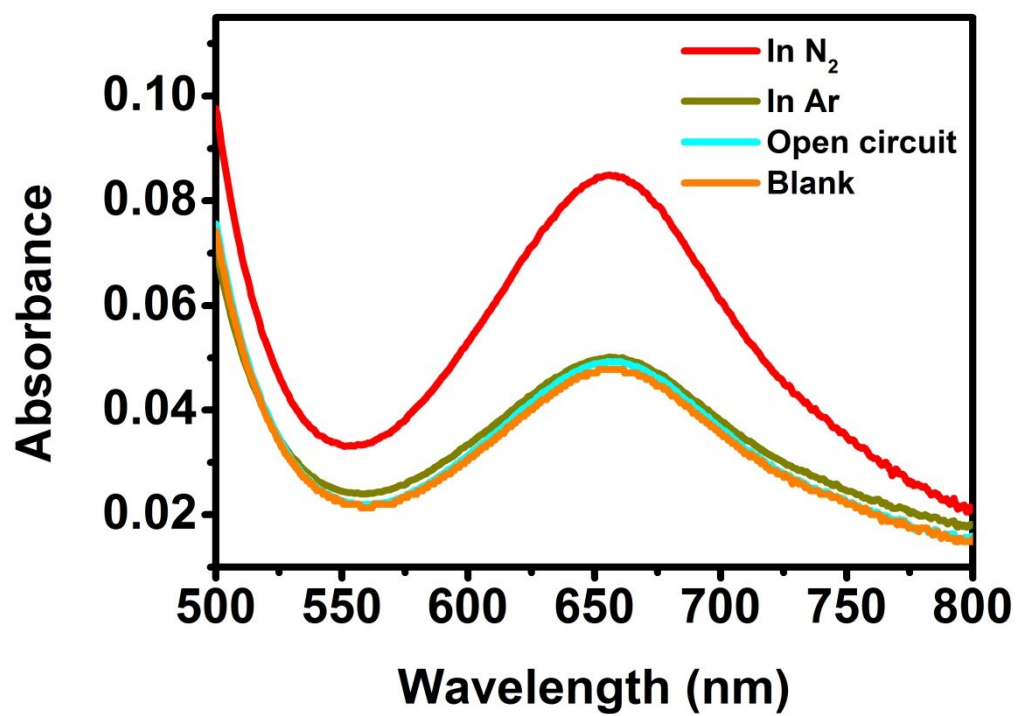


Fig. S12. UV-vis absorption spectra of the electrolytes stained with indophenol indicator after 2 h electrolysis under different electrochemical conditions using CuS-CPSs/CP.

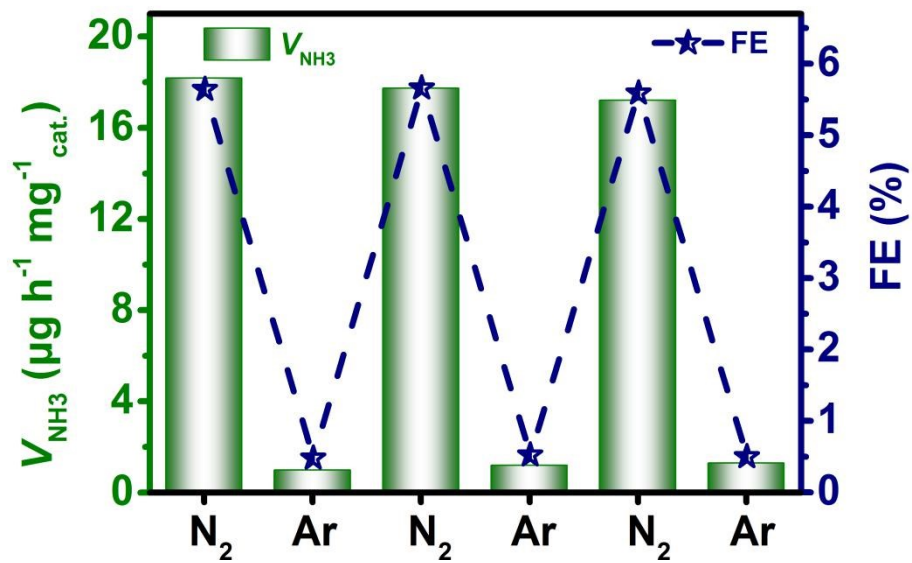


Fig. S13. V_{NH_3} and FE of alternative N₂-saturated and Ar-saturated tests over CuS-CPSs/CP at -0.15 V for 2 h.

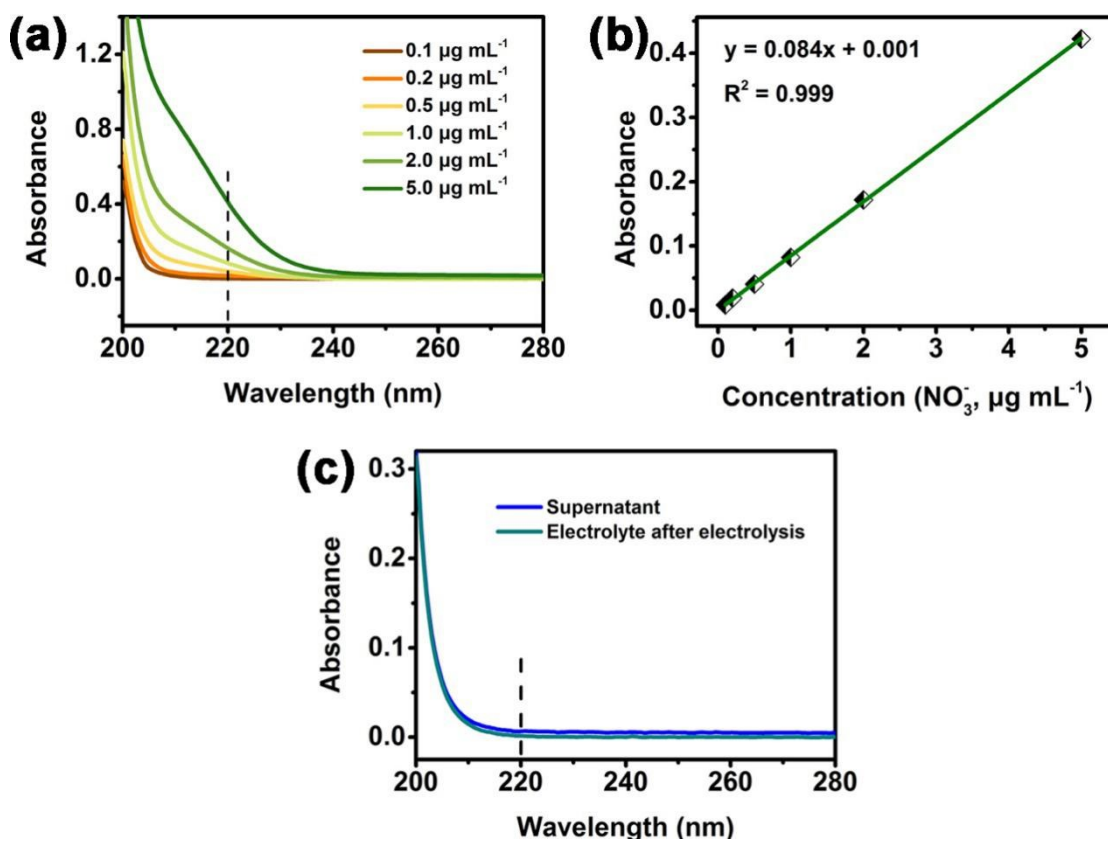


Fig. S14. (a) UV-vis absorption spectra of different concentrations of LiNO_3 . (b) Calibration curve used for calculating the concentration of NO_3^- . (c) UV-vis absorption spectra for the determination of NO_3^- in the supernatant of the washed CuS-CPSs and in 0.1 M HCl after electrolysis.

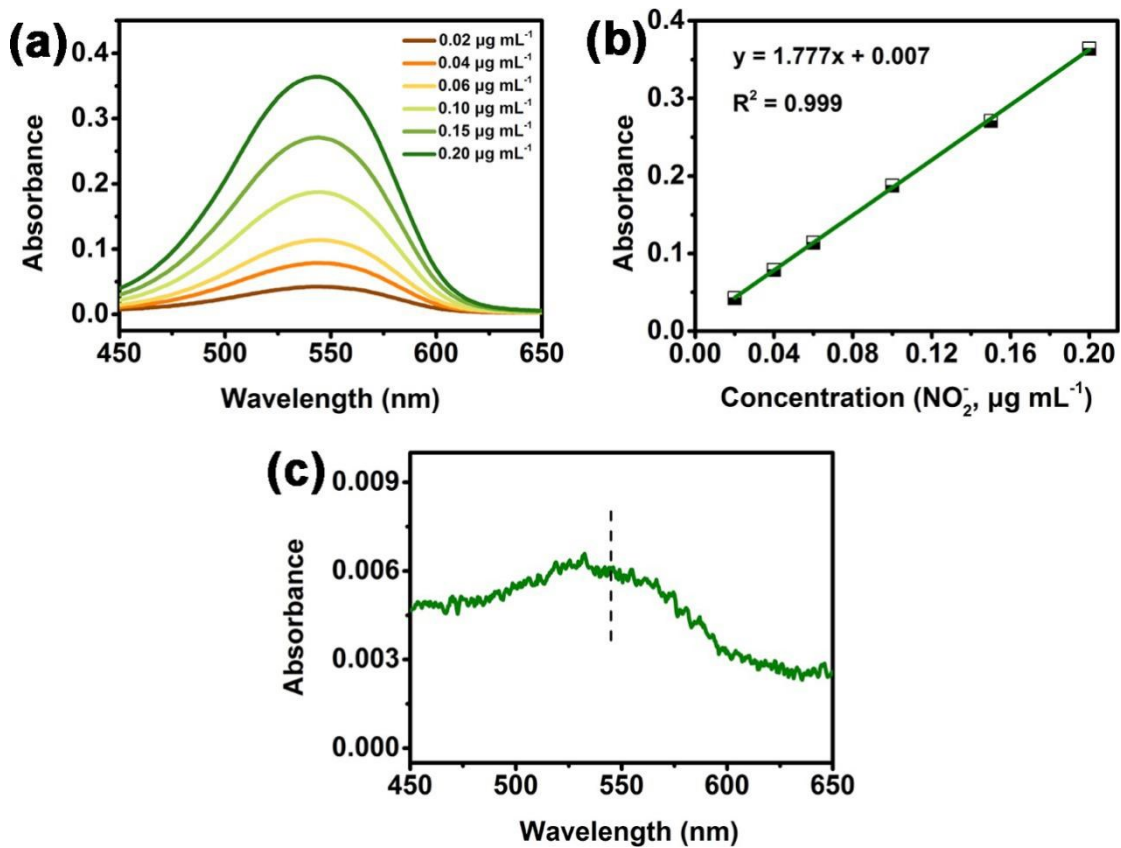


Fig. S15. (a) UV-vis spectra of various concentrations of NaNO_2 . (b) Calibration curve used for calculating the concentration of NO_2^- . (c) UV-vis spectra for the determination of NO_2^- in 0.1 M HCl after electrolysis.

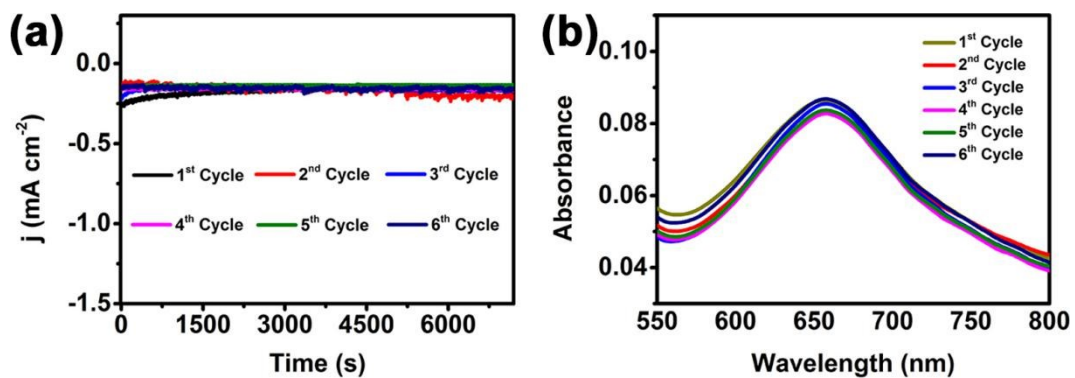


Fig. S16. (a) Time-dependent current density curves for CuS-CPSs/CP at -0.15 V over 6 cycles. (b) UV-vis absorption spectra of the electrolytes stained with indophenol indicator after electrolysis for CuS-CPSs/CP at -0.15 V over 6 cycles.

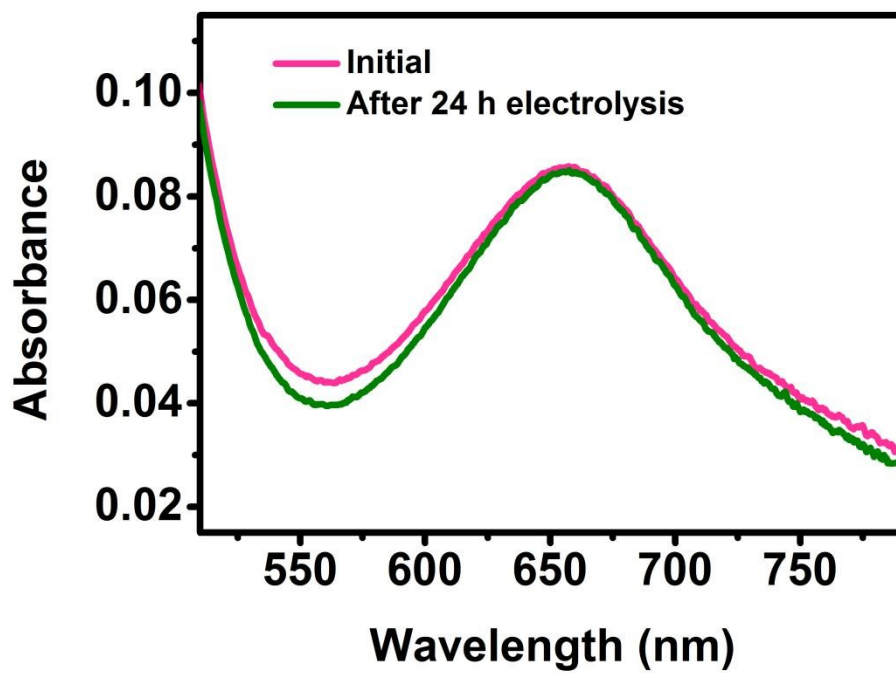


Fig. S17. UV-vis absorption spectra of the electrolytes stained with indophenol indicator at -0.15 V after electrolysis for 2 h with the initial CuS-CPSs/CP and the CuS-CPSs/CP after 24 h electrolysis.

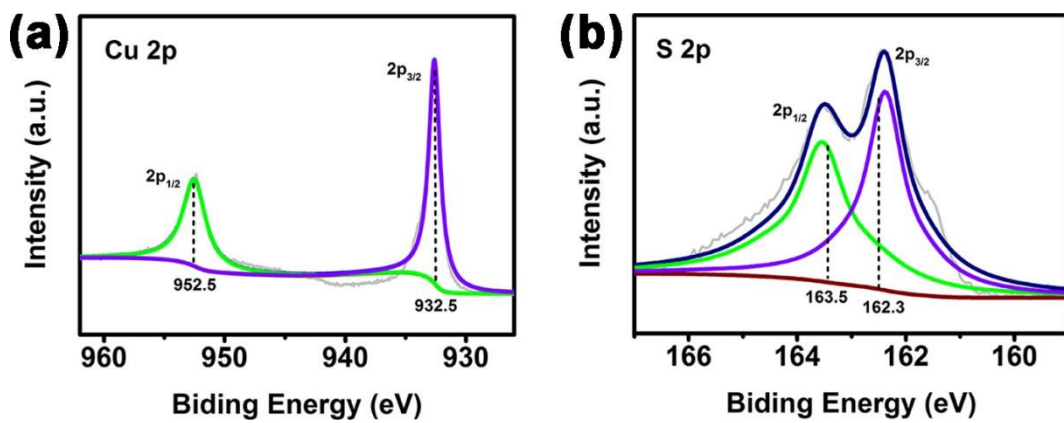


Fig. S18. XPS spectra of post-NRR CuS-CPSs in the (a) Cu 2p and (b) S 2p regions.

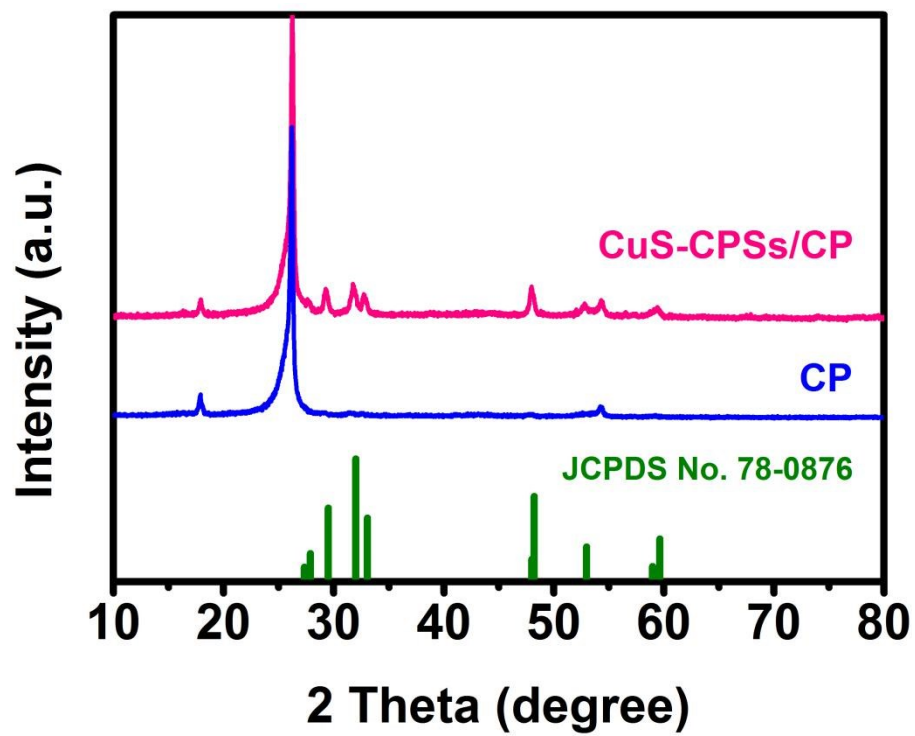


Fig. S19. XRD patterns of CP and post-NRR CuS-CPSs/CP.

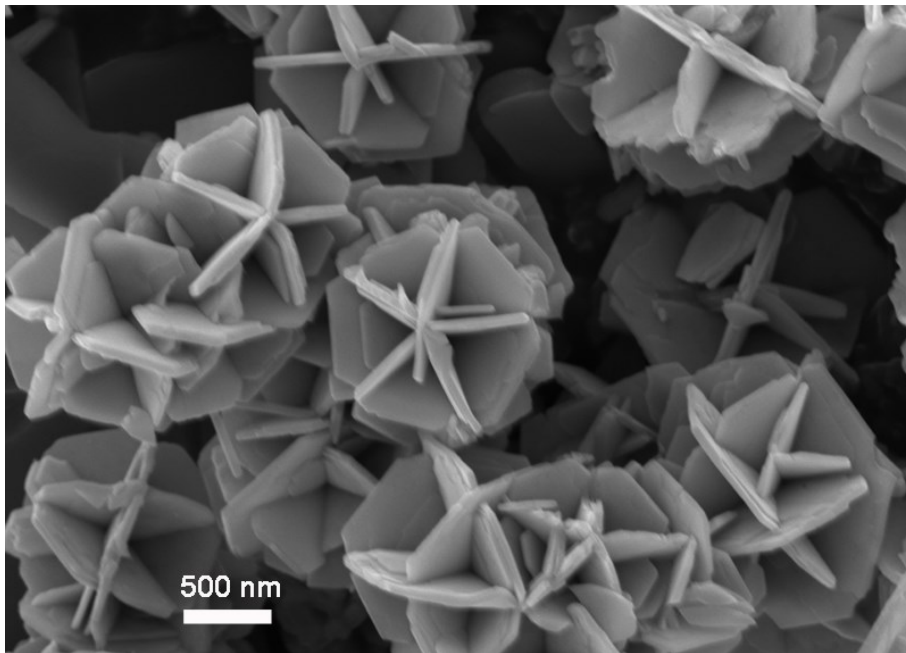


Fig. S20. SEM image of post-NRR CuS-CPSs/CP.

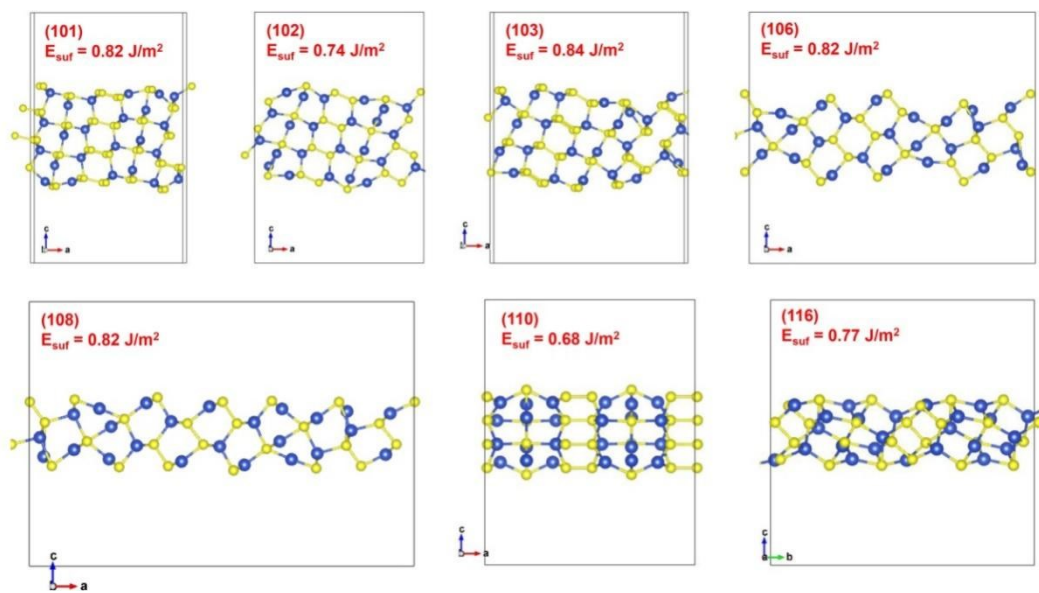


Fig. S21. Surface energies of different facets for CuS.

Table S1. Comparison of electrocatalytic N₂ reduction performance of CuS-CPSs with other NRR electrocatalysts in acidic electrolytes at ambient conditions.

Catalyst	Electrolyte	NH ₃ yield	FE (%)	Ref.
CuS-CPSs	0.1 M HCl	18.18 μg h ⁻¹ mg ⁻¹ _{cat.}	5.63	This work
CoS ₂ @NC	0.1 M HCl	17.45 μg h ⁻¹ mg ⁻¹ _{cat.}	4.6	13
FeMoS	0.1 M HCl	8.45 μg h ⁻¹ mg ⁻¹ _{cat.}	2.96	14
Co-dopd MoS _{2-x}	0.01 M H ₂ SO ₄	0.6 μg h ⁻¹ mg ⁻¹ _{cat.}	10	15
MoS ₂ /C ₃ N ₄	0.1 M HCl	18.5 μg h ⁻¹ mg ⁻¹ _{cat.}	17.8	16
ZrS ₂ NF-Vs	0.1 M HCl	30.72 μg h ⁻¹ mg ⁻¹ _{cat.}	10.33	17
W ₂ N ₃	0.1 M HCl	11.66 μg h ⁻¹ mg ⁻¹ _{cat.}	11.67	18
MoO ₂ -OVs	0.1 M HCl	12.2 μg h ⁻¹ mg ⁻¹ _{cat.}	8.2	19
NPC	0.1 M HCl	0.97 μg h ⁻¹ mg ⁻¹ _{cat.}	4.2	20
ZrO ₂	0.1 M HCl	24.74 μg h ⁻¹ mg ⁻¹ _{cat.}	5.0	21
Ru single-atoms/Zn	0.1 M HCl	3.665 μg h ⁻¹ mg ⁻¹ _{cat.}	21	22
La ₂ Ti ₂ O ₇ nanosheet	0.1 M HCl	25.15 μg h ⁻¹ mg ⁻¹ _{cat.}	4.55	23
Mo nanofilm	0.01 M H ₂ SO ₄	1.89 μg h ⁻¹ mg ⁻¹ _{cat.}	0.72	24
B-doped graphene	0.05 M H ₂ SO ₄	9.8 μg h ⁻¹ mg ⁻¹ _{cat.}	10.8	25
N-doped porous carbon	0.05 M H ₂ SO ₄	15.7 μg h ⁻¹ mg ⁻¹ _{cat.}	1.45	26
LiMn ₂ O ₄ nanofiber	0.1 M HCl	15.83 μg h ⁻¹ mg ⁻¹ _{cat.}	7.44	27
TiC/C nanofiber	0.1 M HCl	14.1 μg h ⁻¹ mg ⁻¹ _{cat.}	5.8	28
Ag nanosheets	0.1 M HCl	2.83 μg h ⁻¹ mg ⁻¹ _{cat.}	4.8	29
α-Au/CeO _x -RGO	0.1 M HCl	8.3 μg h ⁻¹ mg ⁻¹ _{cat.}	10.10	30
Bi nanosheet array	0.1 M HCl	5.26 μg h ⁻¹ mg ⁻¹ _{cat.}	10.26	31
MoO ₃ nanosheets	0.1 M HCl	29.43 μg h ⁻¹ mg ⁻¹ _{cat.}	1.9	32
Black Phosphorus	0.1 M HCl	31.37 μg h ⁻¹ mg ⁻¹ _{cat.}	5.07	33

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