Electronic Supporting Information

Ultra-stable water-dispersive perovskite QDs encapsulated by triple siloxane coupling agent system with different hydrophilic/hydrophobic properties

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EXPERIMENTAL

Materials and chemicals NH₂-PEG was synthesized according to a pervious literature.^[1] (3-Triethoxysilyl) propyl isocyanate (ICPTES) (97%) and (3-aminopropyl) triethoxysilane (APTES) (99%) were purchased from Sinopharm. Perfluorooctyltrimethoxysilane (PFMS) was provided by Alfa Aesar chemical co. ltd. Ditin butyl dilaurate (DBTDL), PbBr₂ (99%), CsBr (99.9%), p-toluenesulfonyl chloride (p-TsCl), ethylenediamine, triethylamine, and hydrobromic acid (HBr) (48% in water) were purchased from Aladdin Reagent Co., Ltd. Poly(ethylene glycol) (mPEG₁₁₃) was obtained from Shanghai Yuanye Biological Technology Co., Ltd. Toluene, N,N-dimethylformamide (DMF), dichloromethane (DCM), methanol, ethanol, tetrahydrofuran (THF), and acetone were obtained from Beijing Chemical Works.

Synthesis of silane coupling agent modified by polyethylene glycol (Si-PEG) Into a roundbottom flask were added catalyst amount DBTDL, mPEG113 (5.0 g, 1.0 mmol), and 20 mL of anhydrous tetrahydrofuran (THF). After slowly dropwise of 0.24 mL ICPTES, the reaction was kept for the stir at room temperature for one day (molar proportion of ICPTES and mPEG113 is 1:1). The reaction mixture was precipitated in cold ethyl ether three times, and the resulted product was vacuum-dried.

Synthesis of PQDs@SiO₂-a PQDs@SiO₂-a was synthesized by using the LARP method as described below. Briefly, PbBr₂ (0.4 mmol, 0.146 g), CsBr (0.2 mmol, 0.042 g), and APTES (100 μ L) were mixed in 10 mL DMF, and then 20 μ L of HBr was injected into the above solution. The above solution was uniformly mixed quickly as the precursor. Under vigorous stirring, 500 μ L of the precursor was added to 10 mL toluene rapidly. The mixture solution immediately showed strong fluorescence emission. After stirring for 20 min to complete the reaction, the toluene solution was centrifuged at 10000 rpm for 5 min, and a precipitate with bright fluorescence was obtained.

Synthesis of PQDs@SiO₂-b PQDs@SiO₂-b was synthesized by the LARP method. Briefly, PbBr₂ (0.4 mmol, 0.146 g), CsBr (0.2 mmol, 0.042 g), APTES (100 μ L), and PMFS (250 μ L) were mixed in 10 mL DMF, and then 20 μ L of HBr was injected into the above solution and then 20 μ L of HBr was injected into the above solution. The above solution was uniformly mixed quickly as the precursor. Under vigorous stirring, 500 μ L of the precursor was added to 10 mL toluene rapidly. The mixture solution immediately showed strong fluorescence emission. After stirring for 20 min to complete the reaction, the toluene solution was centrifuged at 10000 rpm for 5 min, and a precipitate with bright fluorescence was obtained.

Synthesis of PQDs@SiO₂-c The synthesis of PQDs@SiO₂-c was similar to that of PQDs@SiO₂-b, except adding 5 mg NH₂-PEG in the toluene during the synthetic process.

Synthesis of PQDs@SiO₂-d The synthetic procedure of PQDs@SiO₂-d was similar to that of PQDs@SiO₂-b, except adding 15 mg Si-PEG in the toluene during the preparation process.

Cytotoxicity evaluation and cellular imaging HeLa cells were selected to the cytotoxicity evaluation and cellular imaging and were cultured in Dulbecco's modified Eagle's medium (DMEM) with fetal bovineserum (10%) (37 °C and 5% CO₂). The cytotoxicity evaluation of PQDs@SiO₂-d was proved by the methylthiazolyldiphenyltetrazolium bromide (MTT) assays. After 24h of incubation of HeLa cells in a 96-well plate, a fresh cell culture media with different concentrations of PQDs@SiO₂-d (20–120 μ g mL⁻¹) was used to replace the old one and further cultured for 24 h. After 20h incubation, adding 15 μ L DDB to each well was incubated for 4 h. Finally, using a Tecan Infinite M1000 plate reader (Durham, NC, USA) determined the OD value of each well under the wavelength at 490 nm.

For cell imaging, HeLa cells were incubated on 8-well chamber (37 °C and 5% CO₂) for 24 h firstly, and then fresh medium containing PQDs@SiO₂-d (20 μ g mL⁻¹) was used to replace the old one. After 4 h incubation, the obtained HeLa cells were washed with PBS, and 4`,6-

diamidino-2-phenylindole (DAPI) was added to 10 min prior to the imaging for nuclear counterstaining. Prior to imaging, cells were washed with PBS for three times. After washing, the cell imaging as performed on a confocal laser scanning microscope (Nikon C1Si).

Characterization TEM images were measured on JEOL-2100F electron microscope. All PL spectra, PLQYs, and PL lifetimes were recorded with a fluorescence spectrophotometer (Edinburgh FLSP920). UV–vis absorption spectra were performed on a UV–vis spectrophotometer (SHIMADZU UV-2550). FTIR spectra were performed with a FTIR spectrometer (Magna 560). XRD measurements were recorded using a Rigaku D/max-TTR-III diffractometer using Cu Ka radiation (l=0.15405 nm). XPS spectra were performed with a VGESCALAB 220-IXL spectrometer.

Exponential fitting of PL decays and calculation Decay time of the samples was obtained from the decay curves, which is simulated by using a biexponential decay function (1):

$$I(t) = A_0 + A_1 \exp(-t / \tau_1) + A_2 \exp(-t / \tau_2)$$
(1)

where I(t) is the PL intensity at time t, A_i are the weight constants, τ_1 and τ_2 represent the PL decay time. The mean lifetime τ_{ave} is calculated by the formula (2):

$$\tau_{\text{ave=}} \frac{A\tau_1^2 + B\tau_2^2}{A\tau_1 + B\tau_2} \qquad (2)$$

where τ_1 and τ_2 are the PL decay time, and A and B represent the ratio of component contribution to the decay. Actually, τ_1 is due to an internal reorganization, while τ_2 is from the surface state reorganization. The radiative lifetime (τ_r) and non-radiative lifetime (τ_{nr}) can be calculated by the following formula:

$$\tau_{ave} = \frac{1}{k_f + \Sigma k_i} \quad (3)$$
$$PLQY = \frac{k_f}{k_f + \Sigma k_i} \quad (4)$$

$$\tau_{\rm r} = \frac{1}{k_f} \qquad (5)$$
$$\tau_{\rm nr} = \frac{1}{\Sigma k_i} \qquad (6)$$

 Σk_i where k_f and Σki represent the radiative recombination and non-radiative recombination rate constant, respectively.



Scheme S1. Synthetic route of NH₂-PEG.



Fig. S1. ¹H-NMR spectra for a) mPEG₁₁₃-OTs and b) NH₂-PEG.



Fig. S2. FTIR spectra for $mPEG_{113}$, $mPEG_{113}$ -OTs, and NH_2 -PEG.

As shown in Scheme S1, the NH₂-PEG was synthesized by the reaction of ethylenediamine with active intermediate mPEG₁₁₃-OTs obtained from m-PEG₁₁₃ and p-toluenesulfonyl chloride (p-TsCl). The chemical structure of NH₂-PEG was confirmed by ¹H NMR and FTIR (Fig. S1 and S2). The signal 1 at 3.39 ppm can be attributed to the terminal methoxy of mPEG₁₁₃ and the broad signal at 3.5-4.0 ppm is assigned to the methylene hydrogen of the PEG main chain (Fig. S1a). The peak at 4.17 ppm is ascribed to the methylene hydrogen which connected to p-toluenesulfonate. The signals at 7.0-8.3 ppm are characteristic peaks of the benzene ring in p-toluenesulfonate. In Fig. S1b, the broad signals at 3.0-3.5 ppm are assigned to methylene which connected to the amino group and the signals corresponding to the p-toluenesulfonate group disappeared. From the FTIR spectra (Fig. S2) of m-PEG₁₁₃-OTs and NH₂-PEG, the characteristic peak of N-H at 1680 cm⁻¹ also can be observed, which indicates that -OTs group has been replaced by ethylenediamine to obtain the target product NH₂-PEG.



Scheme S2. Synthetic route of Si-PEG.



Fig. S3. ¹H-NMR spectra for Si-PEG.



Fig. S4. FTIR spectra for ICPTES, mPEG₁₁₃, and Si-PEG.

The Si-PEG was prepared by the polyaddition reaction between the isocyanate group (3triethoxysilyl) propyl isocyanate (ICPTES) and the hydroxyl group on the end of m-PEG₁₁₃ (Scheme S2). The peak at 5.3-5.4 ppm can be attributed to the newly formed N-H group by polyaddition reaction (Fig. S3). As shown in Fig. S3, the characteristic signal at 3.39 ppm can be attributed to the methylene of m-PEG₁₁₃. The peaks at 0.78-0.83, 1.38 ppm are assigned to the methylene in ICPTES. The broad peak at 1.8-2.0 ppm is attributed to the methyl in ICPTES. From the FTIR spectra (Fig. S4), the characteristic peak of -N=C=O at 2273 cm⁻¹ disappeared and a new peak corresponding to the urethane group was observed at 1760-1530 cm⁻¹. The above results indicated the successful synthesis of Si-PEG.



Fig. S5. (a) PL spectra of PQD's colloidal solutions obtained by different feeding amount of APTES with 20 μ L of HBr. (b) PL spectra of PQD's colloidal solutions obtained by different concentrations of PFMS with 100 μ L of APTES and 20 μ L of HBr. (c) PL spectra of PQD's colloidal solutions obtained by different concentrations of NH₂-PEG with 100 μ L of APTES, 20 μ L of HBr, and 250 μ L of PFMS. (d) PL spectra of PQD's colloidal solutions obtained by different by different concentrations of PFMS. 20 μ L of HBr, and 250 μ L of PFMS. (d) PL spectra of PQD's colloidal solutions obtained by PFMS.



Fig. S6. High-resolution Br 3d spectra of (a) PQDs@SiO₂-b, (b) PQDs@SiO₂-c, and (c) PQDs@SiO₂-d.



Fig. S7. High-resolution Cs 3d spectra of (a) PQDs@SiO₂-b, (b) PQDs@SiO₂-c, and (c) PQDs@SiO₂-d.



Fig. S8. High-resolution Pb 4f spectra of (a) PQDs@SiO₂-b, (b) PQDs@SiO₂-c, and (c) PQDs@SiO₂-d.



Fig. S9. High-resolution N 1s spectra of (a) PQDs@SiO₂-b, (b) PQDs@SiO₂-c, and (c) PQDs@SiO₂-d.



Fig. S10. High-resolution O 1s spectra of d) PQDs@SiO₂-b, e) PQDs@SiO₂-c, and f) PQDs@SiO₂-d.



Fig. S11. High-resolution F 1s spectra of g) PQDs@SiO₂-b, h) PQDs@SiO₂-c, and i) PQDs@SiO₂-d.



Fig. S12. High-resolution Si 2p spectra of j) PQDs@SiO₂-b, k) PQDs@SiO₂-c, and l) PQDs@SiO₂-d.



Fig. S13. Time-resolved FL decay curves of PQDs@SiO₂-a, PQDs@SiO₂-b, PQDs@SiO₂-c, and PQDs@SiO₂-d.



Fig. S14. PL spectra of PQDs@SiO₂-c (1.0 mg mL⁻¹) in toluene, water, THF, acetone, ethanol, PBS buffer solution, HCl solution (pH=1) and NaOH solution (pH=14).



Fig. S15. PL spectra of PQDs@SiO₂-d (1.0 mg mL⁻¹) in toluene, water, THF, acetone, ethanol, PBS buffer solution, HCl solution (pH=1) and NaOH solution (pH=14).



Fig. S16. Temperature-dependent PL intensity of (a) PQDs@SiO₂-c and (b) PQDs@SiO₂-d in water (1.0 mg mL⁻¹).

	Ligand in precusors					Ligand in	
Samples					nucleation solvent		
	PbBr ₂	CsBr	APTES	HBr	PFMS	NH ₂ -PEG	Si-PEG
	(g)	(g)	(µL)	(µL)	(µL)	(mg)	(mg)
PQDs@SiO ₂ -a	0.146	0.042	100	20	none	none	none
PQDs@SiO ₂ -b	0.146	0.042	100	20	250	none	none
PQDs@SiO ₂ -c	0.146	0.042	100	20	250	5	none
PQDs@SiO ₂ -d	0.146	0.042	100	20	250	none	15

Table S1. Experimental conditions of preparing samples via LARP method. a

^a Precursor solvent: DMF. Nucleation solvent: toluene. Temperature: room temperature.

Table S2. Cs/Pb, Pb/Br, and Cs/Pb/Br ratios of PQDs@SiO2-b, PQDs@SiO2-c, andPQDs@SiO2-d measured by XPS.

Samples	Cs/Pb	Pb/Br	Cs/Pb/Br
PQDs@SiO ₂ -b	1:1.3	1:4.34	1:1.3:5.64
PQDs@SiO ₂ -c	1:2.56	1:4.49	1:2.56:11.51
PQDs@SiO ₂ -d	1:0.79	1:4.72	1:0.79:3.73

Table S3. Summary of time-resolved FL decay measurements, where the lifetimes of samples are tested in toluene solution. τ_1 and τ_2 are the detailed recombination lifetimes, A and B represent the ratio of component contribution to the decay. k_f and Σki represent the radiative recombination and the non-radiative recombination rate constant.

Samples	τ_1 (ns)	A%	$ au_2$ (ns)	В%	<i>k</i> _f	Σki
PQDs@SiO ₂ -a	45.92	43.61	276.64	56.39	2.56×10 ⁻³	1.44×10 ⁻³
PQDs@SiO ₂ -b	18.42	53.23	142.47	46.27	7.74×10 ⁻³	0.16×10 ⁻³
PQDs@SiO ₂ -c	19.27	34.25	142.31	65.75	4.54×10 ⁻³	3.15×10 ⁻³
PQDs@SiO ₂ -d	24.53	42.46	112.67	57.54	9.26×10 ⁻³	0.70×10 ⁻³

Table S4. Overview and comparison of the PL spectroscopic results from Fig. S14 of $PQDs@SiO_2$ -c in toluene, water, THF, acetone, ethanol, PBS buffer solution, HCl solution(pH=1) and NaOH solution(pH=14) (1.0 mg mL⁻¹).

Solution	Peak (nm)	Intensity	Ratio
Toluene	505	43399	1
Water	512	37091	0.85
THF	509	31178	0.72
Acetone	505	24318	0.56
Ethanol	505	22027	0.51
PBS	506	27555	0.63
pH=1	492	25182	0.58
pH=14	506	11378	0.26

Table S5. Overview and comparison of the PL spectroscopic results from Fig. S15 of $PQDs@SiO_2$ -d in toluene, water, THF, acetone, ethanol, PBS buffer solution, HCl solution (pH=1) and NaOH solution (pH=14) (1.0 mg mL⁻¹).

Solution	Peak (nm)	Intensity	Ratio
Toluene	505	107240	1
Water	510	101000	0.94
THF	501	95108	0.89
Acetone	489	88370	0.82
Ethanol	506	79810	0.74
PBS	512	95067	0.87
pH=1	506	101536	0.95
pH=14	510	97663	0.91

REFERENCES

[1] Y. Chen, X. Song, F. Peng, J. Li, Preparation of Monomethoxy Poly(ethylene glycol) Amine, *Fine Chemicals* **2015**, 32, 434.