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SUPPORTING INFORMATION

Gold(I)-catalyzed redox transformation of *o*-nitroalkynes with

indoles for the synthesis of 2,3'-biindole derivatives

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General Information

All reactions were carried out in oven-dried glassware. Solvents were dried by the standard methods. Flash column chromatography was performed using silica gel (300-400 mesh). Analytical thin-layer chromatography was performed using glass plates pre-coated with 200-300 mesh silica gel impregnated with a fluorescent indicator (254 nm). ¹H NMR and ¹³C NMR spectra were recorded in CDCl₃ on a 400 or 500 MHz spectrometer; chemical shifts were reported in ppm with the solvent signal as reference, and coupling constants (*J*) were given in Hertz. The peak information was described as: br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, comp = composite. High-resolution mass spectra (HRMS) were recorded on a commercial apparatus (ESI or CI Source).

General Procedure for the Preparation of *o*-nitroalkynes 1.



<u>Synthesis of S-2:</u>¹ To a 50-mL oven-dried flask containing a magnetic stirring bar, S-1 (10 mmol, 1.0 equiv), trimethylsilylacetylene (15 mmol, 1.5 equiv), (PPh₃)₂PdCl₂ (70.1 mg, 0.01 equiv), CuI (19.0 mg, 0.01 equiv), and Et₃N (25 mL) were added in sequence under argon atmosphere at room temperature. The reaction mixture was stirred at 50 - 80 °C for 2.0 hours. Upon completion (monitored by TLC), the reaction mixture was filtered through a short pad of Celite and the solid was washed with EtOAc (20 mL). The combined organic phase was concentrated under reduced pressure, and the residue was purified by column chromatography on silica gel (Hexanes : EtOAc = 50:1) to afford pure products S-2 in >80% yields.



<u>Synthesis of 1:</u> To a 50-mL oven-dried flask containing a magnetic stirring bar, S-2 (8.0 mmol, 1.0 equiv), K_2CO_3 (12 mmol, 1.5 equiv), and MeOH (20 mL) were added in sequence at 0 °C. The reaction mixture was stirred at 0 °C for 10 minutes. Upon completion (monitored by TLC), H₂O (10 mL) was added to quench the reaction, and the aqueous layer was extracted with DCM (3×10 mL). The combined organic phase was washed with brine and dried over anhydrous Na₂SO₄. The solvent was evaporated in vacuo after filtration, and the residue was purified by column chromatography on silica gel (Hexanes : EtOAc = 50:1) to afford pure products **1** in >90% yields. The physical and spectral properties identical to the earlier reported.¹

General Procedure for the Gold-Catalyzed Cascade Reaction.



To a 10-mL oven-dried vial containing a magnetic stirring bar, o-nitroalkyne **1** (0.3 mmol), indole **2** (0.2 mmol), PPh₃AuNTf₂ (7.4 mg, 5.0 mol%), and anhydrous DCE (4.0 mL) were added in sequence under atmosphere of argon at 0 °C, and the reaction mixture was stirred at 0 °C for 1.0 hour (or at indicated time in Scheme 3 for the synthesis of **4**). When the reaction was completed (monitored by TLC), the crude reaction mixture was quenched with saturated aqueous sodium bicarbonate (10 mL) and extracted with EtOAc (15 mL). The organic layer was washed with brine and dried over anhydrous Na₂SO₄. The solvent was evaporated in *vacuo* after filtration. The residue was purified by flash column chromatography on silica gel (Hexanes : EtOAc = 10:1 to 4:1) to give the pure products **3** or **4** in moderate to high yields.



3-oxo-1'*H*,3*H*-[2,3'-biindole] **1-oxide** (3a). 47.7 mg, 91% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 12.11 (s, 1H), 8.94 (s, 1H), 8.58 (d, *J* = 8.0 Hz, 1H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.65 (d, *J* = 7.0 Hz, 1H), 7.60 (d, *J* = 7.6 Hz, 1H), 7.54 (comp, 2H), 7.25 (t, *J* = 7.1 Hz, 1H), 7.19 (t, *J* = 7.5 Hz, 1H). ¹³C NMR (100 MHz, DMSO) δ 187.3, 148.3, 136.7, 135.8, 133.3, 131.1, 130.6, 124.6, 124.0, 123.4, 123.3, 121.9, 121.2, 113.4, 112.7, 103.1. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₁N₂O₂ [M+H]⁺: 263.0815, found 263.0815.



4'-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3b**). 40.3 mg, 73% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 11.84 (s, 1H), 7.83 (t, *J* = 7.6 Hz, 1H), 7.75 – 7.64 (comp, 4H), 7.33 (d, *J* = 8.1 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.86 (d, *J* = 7.1 Hz, 1H), 2.38 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 187.6, 147.7, 137.0, 135.6, 134.0, 131.6, 130.6, 129.6, 126.3, 122.7, 122.1, 121.9, 114.1, 110.3, 98.9, 19.3. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0966.



5'-methyl-3-oxo-1'*H***,3***H***-[2,3'-biindole] 1-oxide** (**3c**)**.** 45.2 mg, 82% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 11.99 (s, 1H), 8.87 (d, *J* = 3.0 Hz, 1H), 8.37 (s, 1H), 7.77 (t, *J* = 7.6 Hz, 1H), 7.64 (d, *J* = 7.1 Hz, 1H), 7.60 – 7.54 (comp, 2H), 7.40

(d, J = 8.2 Hz, 1H), 7.08 (d, J = 9.0 Hz, 1H), 2.44 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 187.3, 148.3, 135.8, 135.1, 133.3, 131.1, 130.5, 130.1, 129.8, 124.9, 123.7, 123.3, 121.9, 113.4, 112.3, 102.7, 22.1. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0988.



6'-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3d**). 44.1 mg, 80% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 11.98 (s, 1H), 8.89 (s, 1H), 8.46 (d, *J* = 8.3 Hz, 1H), 7.75 (t, *J* = 8.0 Hz, 1H), 7.63 (d, *J* = 6.9 Hz, 1H), 7.59 – 7.51 (comp, 2H), 7.30 (s, 1H), 7.01 (dd, *J* = 8.4, 1.2 Hz, 1H), 2.42 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 187.3, 148.3, 137.2, 135.7, 133.2, 132.7, 130.7, 130.5, 123.8, 123.2, 122.9, 122.4, 121.9, 113.3, 112.4, 103.1, 21.7. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0975.



7'-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3e**). 43.0 mg, 78% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 11.98 (s, 1H), 8.89 (s, 1H), 8.46 (d, *J* = 8.3 Hz, 1H), 7.75 (t, *J* = 8.0 Hz, 1H), 7.63 (d, *J* = 6.9 Hz, 1H), 7.59 – 7.51 (comp, 2H), 7.30 (s, 1H), 7.01 (dd, *J* = 8.4, 1.2 Hz, 1H), 2.42 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 187.3, 148.3, 137.2, 135.7, 133.2, 132.7, 130.7, 130.5, 123.8, 123.2, 122.9, 122.4, 121.9, 113.3, 112.4, 103.1, 21.7. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0963.



5'-methoxy-3-oxo-1*'H*,**3***H*-**[2,3'-biindole] 1-oxide** (**3f**). 40.9 mg, 70% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 11.98 (s, 1H), 8.91 (d, *J* = 3.0 Hz, 1H), 8.19 (d, *J* = 2.1 Hz, 1H), 7.76 (t, *J* = 7.6 Hz, 1H), 7.65 (d, *J* = 7.1 Hz, 1H), 7.60 – 7.52 (comp, 2H), 7.41 (d, *J* = 8.8 Hz, 1H), 6.94 – 6.85 (m, 1H), 3.83 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 187.5, 154.8, 148.3, 135.8, 133.3, 131.7, 131.4, 130.4, 125.3, 123.2, 121.9, 113.3, 113.2, 113.2, 106.1, 103.1, 55.7. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₃ [M+H]⁺: 293.0921, found 293.0927.



4'-fluoro-3-oxo-1'*H*,3*H*-[2,3'-biindole] **1-oxide** (**3g**). 47.6 mg, 85% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 12.27 (s, 1H), 8.07 (s, 1H), 7.81 (t, *J* = 7.6 Hz, 1H), 7.69 (t, *J* = 6.1 Hz, 2H), 7.64 (t, *J* = 7.4 Hz, 1H), 7.37 (d, *J* = 8.1 Hz, 1H), 7.25 – 7.16 (comp, 1H), 6.92 – 6.86 (m, 1H). ¹³C NMR (125 MHz, DMSO) δ 185.9, 157.1 (d, *J* = 248.0 Hz), 147.7, 139.4 (d, *J* = 10.1 Hz), 135.6, 132.0, 131.5, 130.7, 123.7 (d, *J* = 7.8 Hz), 123.1, 121.9, 114.6 (d, *J* = 20.0 Hz), 114.0, 109.1 (d, *J* = 3.5 Hz), 106.4 (d, *J* = 20.0 Hz), 97.1. ¹⁹F NMR (471 MHz, DMSO) δ -116.5. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀FN₂O₂ [M+H]⁺: 281.0721, found 281.0718.



5'-fluoro-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3h**). 37.5 mg, 67% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 12.23 (s, 1H), 8.95 (s, 1H), 8.31 (dd, *J* = 11.3, 2.4 Hz, 1H), 7.75 (t, *J* = 7.6 Hz, 1H), 7.63 (d, *J* = 7.0 Hz, 1H), 7.60 – 7.49 (m, 3H), 7.10 (td, *J* = 8.9, 2.4 Hz, 1H). ¹³C NMR (125 MHz, DMSO) δ 187.3, 158.0 (d, *J* = 232.5 Hz), 148.2, 135.8, 133.4, 132.5, 130.7, 130.1, 125.0 (d, *J* = 11.1 Hz), 123.25, 121.95, 113.1 (d, *J* = 10.1 Hz), 113.4, 111.5 (d, *J* = 20.0 Hz), 109.1, 108.9, 103.4 (d, *J* = 4.3 Hz). ¹⁹F NMR (376 MHz, DMSO) δ -122.3. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀FN₂O₂ [M+H]⁺: 281.0721, found 281.0718.



5'-chloro-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3i**). 40.2 mg, 68% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 12.26 (s, 1H), 8.93 (s, 1H), 8.62 (d, *J* = 1.8 Hz, 1H), 7.77 (t, *J* = 7.5 Hz, 1H), 7.65 (d, *J* = 7.0 Hz, 1H), 7.61 – 7.53 (comp, 3H), 7.26 (dd, *J* = 8.6, 1.9 Hz, 1H). ¹³C NMR (125 MHz, DMSO) δ 187.2, 148.2, 135.8, 135.3, 132.8, 132.2, 130.7, 125.7, 125.6, 123.4, 123.31, 123.27, 122.0, 114.2, 113.5, 102.9. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀ClN₂O₂ [M+H]⁺: 297.0425, found 297.0429.



2'-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3**j). 39.2 mg, 71% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 11.76 (s, 1H), 7.79 (t, *J* = 7.2 Hz, 1H), 7.68 – 7.65 (comp, 2H), 7.61 (t, *J* = 7.3 Hz, 1H), 7.45 (d, *J* = 7.9 Hz, 1H), 7.36 (d, *J* = 8.0 Hz, 1H), 7.11 (t, *J* = 7.1 Hz, 1H), 7.02 (t, *J* = 7.5 Hz, 1H), 2.49 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 186.7, 148.2, 140.6, 136.1, 135.4, 134.2, 131.2, 127.0, 123.9, 121.9, 121.8, 121.3, 120.2, 113.9, 111.5, 97.9, 14.7. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0963.



1'-methyl-3-oxo-1'*H*,3*H*-[**2**,3**'-biindole**] **1-oxide** (**3k**). 40.8 mg, 74% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 8.95 (s, 1H), 8.59 (d, *J* = 8.1 Hz, 1H), 7.77 - 7.73 (comp, 1H), 7.65 (d, *J* = 7.1 Hz, 1H), 7.60 – 7.53 (comp, 3H), 7.34 – 7.31 (comp, 1H), 7.25 – 7.22 (comp, 1H), 3.93 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 187.2, 148.2, 137.3, 135.8, 134.6, 132.9, 130.6, 125.1, 124.3, 123.5, 123.2, 121.9, 121.4, 113.4, 111.0, 102.1, 33.8. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0973.



1'-allyl-3-oxo-1'*H***,3***H***-[2,3'-biindole] 1-oxide** (**3).** 47.1 mg, 78% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 8.99 (s, 1H), 8.60 (d, *J* = 8.0 Hz, 1H), 7.76 (d, *J* = 7.6

Hz, 1H), 7.65 (d, J = 7.1 Hz, 1H), 7.62 - 7.50 (comp, 3H), 7.30 (t, J = 7.5 Hz, 1H), 7.23 (t, J = 7.5 Hz, 1H), 6.12 – 5.99 (comp, 1H), 5.23 (d, J = 10.3 Hz, 1H), 5.14 (d, J = 17.0 Hz, 1H), 5.00 (d, J = 5.3 Hz, 2H). ¹³C NMR (100 MHz, DMSO) δ 187.1, 148.2, 136.5, 135.8, 134.0, 133.7, 132.9, 130.6, 125.2, 1244, 123.5, 123.2, 121.9, 121.5, 118.1, 113.4, 111.3, 102.6, 49.1. HRMS (TOF MS ESI⁺) calculated for C₁₉H₁₅N₂O₂ [M+H]⁺: 303.1128, found 303.1126.



2-(1-methyl-1*H***-pyrrol-2-yl)-3-oxo-3***H***-indole 1-oxide (3m). 39.8 mg, 88% yield. Purple solid. ¹H NMR (500 MHz, DMSO) \delta 7.81 - 7.74 (comp, 1H), 7.69 - 7.57 (comp, 3H), 7.15 – 7.10 (comp, 1H), 6.85 - 6.80 (comp, 1H), 6.28 – 6.24 (comp, 1H), 3.78 (s, 3H). ¹³C NMR (125 MHz, DMSO) \delta 185.8, 147.9, 135.6, 131.4, 128.9, 123.4, 129.9, 122.0, 118.8, 116.6, 114.1, 109.7, 37.4. HRMS (TOF MS ESI⁺) calculated for C₁₃H₁₁N₂O₂ [M+H]⁺: 227.0815, found 227.0817.**



3-oxo-2-(1*H***-pyrrol-2-yl)-3***H***-indole 1-oxide (3n). 35.2 mg, 83% yield. Purple solid. ¹H NMR (400 MHz, DMSO) \delta 11.73 (s, 1H), 7.78 - 7.71 (comp, 1H), 7.63 – 7.50 (comp, 3H), 7.32 - 7.28 (comp, 1H), 7.21 - 7.15 (comp, 1H), 6.40 - 6.32 (comp, 1H). ¹³C NMR (100 MHz, DMSO) \delta 186.2, 148.3, 135.9, 130.7, 124.8, 123.4, 122.2, 118.9, 114.6, 113.6, 111.2. HRMS (TOF MS ESI⁺) calculated for C₁₂H₉N₂O₂ [M+H]⁺: 213.0659, found 213.0656.**



7-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3o**). 41.4 mg, 75% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 12.05 (s, 1H), 8.92 (d, *J* = 3.0 Hz, 1H), 8.55 (d, *J* = 8.1 Hz, 1H), 7.54 – 7.46 (comp, 3H), 7.41 (t, *J* = 7.4 Hz, 1H), 7.26 – 7.21 (comp, 1H), 7.19 – 7.15 (comp, 1H), 2.72 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 187.3, 143.8, 139.7, 136.7, 133.7, 130.5, 130.2, 127.0, 124.6, 124.1, 124.0, 123.3, 121.0, 120.1, 112.6, 103.0, 17.0. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0977.



5-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3p**). 38.6 mg, 70% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 12.06 (s, 1H), 8.90 (s, 1H), 8.55 (d, *J* = 8.1 Hz, 1H), 7.55 (d, *J* = 7.6 Hz, 1H), 7.51 (d, *J* = 8.0 Hz, 1H), 7.49 – 7.43 (m, 2H), 7.24 (t, *J* = 7.5 Hz, 1H), 7.17 (t, *J* = 7.5 Hz, 1H), 2.43 (s, 3H). ¹³C NMR (125 MHz, DMSO) δ 187.4, 146.2, 140.8, 136.7, 135.5, 133.0, 130.8, 124.6, 124.0, 123.4, 123.3, 122.6, 121.1, 113.2, 112.6, 103.1, 21.3. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0971.



4-methyl-3-oxo-1'*H***,3***H***-[2,3'-biindole] 1-oxide** (**3q**). 44.2 mg, 80% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 12.05 (s, 1H), 8.92 (d, *J* = 2.9 Hz, 1H), 8.55 (d,

J = 8.0 Hz, 1H), 7.56 - 7.44 (comp, 3H), 7.40 (t, J = 7.4 Hz, 1H), 7.27 - 7.23 (comp, 1H), 7.20 - 7.14 (comp, 1H), 2.71 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 187.3, 143.7, 139.7, 136.7, 133.7 130.6, 130.5, 130.2, 127.0, 124.6, 124.0, 123.2, 121.0, 120.0, 112.5, 103.0, 17.0. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0976.



4-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3r**). 42.6 mg, 76% yield. Purple solid. ¹H NMR (400 MHz, DMSO) δ 12.12 (s, 1H), 8.94 (d, J = 2.8 Hz, 1H), 8.54 (d, J = 8.0 Hz, 1H), 7.84 - 7.76 (comp, 1H), 7.52 (d, J = 8.0 Hz, 1H), 7.45 (d, J = 7.6 Hz, 1H), 7.38 (t, J = 8.7 Hz, 1H), 7.25 (t, J = 7.4 Hz, 1H), 7.18 (t, J = 7.5 Hz, 1H). ¹³C NMR (125 MHz, DMSO) δ 183.61, 155.2 (d, J = 261.7 Hz), 149.2 (d, J = 4.6 Hz), 138.2 (d, J = 8.7 Hz), 136.8, 133.4, 131.4, 124.5, 124.1, 123.5, 121.3, 119.2, 119.1, 112.7, 110.1, 103.0, 79.7. ¹⁹F NMR (376 MHz, DMSO) δ -115.4. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀FN₂O₂ [M+H]⁺: 281.0721, found 281.0726.



6-methyl-3-oxo-1'*H*,3*H*-[**2**,3'-biindole] **1-oxide** (**3s**). 30.4 mg, 55% yield. Purple solid. ¹H NMR (500 MHz, DMSO) δ 12.10 (s, 1H), 8.94 (s, 1H), 8.57 (d, *J* = 7.6 Hz, 1H), 7.52 (d, *J* = 7.1 Hz, 2H), 7.43 (s, 1H), 7.33 (d, *J* = 6.4 Hz, 1H), 7.25 (t, *J* = 6.9 Hz, 1H), 7.18 (t, *J* = 6.9 Hz, 1H), 2.47 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 186.9, 148.7, 147.1, 136.7, 131.0, 130.5, 124.6, 124.1, 123.4, 121.9, 121.1, 120.8, 114.2, 112.6, 103.2, 22.3. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O₂ [M+H]⁺: 277.0972, found 277.0967.



1'H,3H-[2,3'-biindol]-3-one (**4a**). Deep red solid. ¹H NMR (500 MHz, DMSO) δ ¹H NMR (500 MHz, DMSO) δ 12.16 (s, 1H), 8.52 (s, 1H), 8.41 (d, J = 7.1 Hz, 1H), 7.62 - 7.56 (comp, 3H), 7.36 (d, J = 6.8 Hz, 1H), 7.31 - 7.24 (comp, 2H), 7.19 (t, J = 6.3 Hz, 1H). ¹¹³C NMR (125 MHz, DMSO) δ 195.7, 163.4, 158.5, 137.8, 137.3, 133.8, 126.8, 126.4, 124.9, 124.0, 123.0, 122.9, 122.3, 121.1, 113.0, 106.9. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₁N₂O [M+H]⁺: 247.0871, found 247.0863.



6-methyl-1'*H*,3*H*-[**2**,3'-biindol]-3-one (4s). 31.7 mg, 61% yield. Deep red solid. ¹H NMR (400 MHz, DMSO) δ 12.13 (s, 1H), 8.51 (s, 1H), 8.42 – 8.37 (comp, 1H), 7.57 – 7.52 (comp, 1H), 7.42 (d, *J* = 7.3 Hz, 1H), 7.31 – 7.24 (comp, 2H), 7.20 (s, 1H), 7.00 (d, *J* = 7.3 Hz, 1H), 2.39 (s, 3H). ¹³C NMR (100 MHz, DMSO) δ 195.1, 164.0, 159.2, 149.2, 137.2, 133.7, 127.0, 126.4, 124.9, 123.9, 122.9, 122.2, 122.1, 120.6, 113.0, 107.0, 22.4. HRMS (TOF MS ESI⁺) calculated for C₁₇H₁₃N₂O [M+H]⁺: 261.1022, found 261.1028.



7-fluoro-1'*H***,3***H***-[2,3'-biindol]-3-one (4t).** 37.0 mg, 70% yield. Deep red solid. ¹H NMR (400 MHz, DMSO) δ 12.23 (s, 1H), 8.55 (d, *J* = 3.0 Hz, 1H), 8.47 - 8.22 (comp, 1H), 7.61 – 7.53 (comp, 1H), 7.52 – 7.45 (comp, 1H), 7.40 (d, *J* = 7.0 Hz, 1H), 7.36 – 7.26 (comp, 2H), 7.25 – 7.18 (comp, 1H). ¹³C NMR (100 MHz, DMSO) δ 194.5,

158.6, 153.2 (d, J = 254.4 Hz), 148.7 (d, J = 12.3 Hz), 137.33, 134.36, 128.4 (d, J = 5.8 Hz), 126.31, 126.0 (d, J = 3.7 Hz), 125.5 (d, J = 19.7 Hz), 124.15, 122.97, 122.52, 121.1(d, J = 2.5 Hz), 113.1, 107.1. ¹⁹F NMR (376 MHz, DMSO) δ -130.0. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀FN₂O [M+H]⁺: 265.0772, found 265.0770.



5-fluoro-1'*H*,**3***H*-**[2,3'-biindol]-3-one** (**4u**). 35.9 mg, 68% yield. Deep red solid. ¹H NMR (400 MHz, DMSO) δ 12.14 (s, 1H), 8.48 (d, J = 2.4 Hz, 1H), 8.43 – 8.34 (comp, 1H), 7.57 - 7.50 (comp, 1H), 7.42 – 7.34 (comp, 3H), 7.33 – 7.21 (comp, 2H). ¹³C NMR (125 MHz, DMSO) δ 194.7, 161.2 (d, J = 244.2 Hz), 159.3, 158.9 (d, J = 3.8 Hz), 137.2, 133.7, 126.2, 124.3 (d, J = 7.9 Hz), 124.0, 123.1 (d, J = 23.6 Hz), 122.9, 122.3, 122.1 (d, J = 7.5 Hz), 113.1, 112.6 (d, J = 25.4 Hz), 106.9. ¹⁹F NMR (376 MHz, DMSO) δ -116.5. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀FN₂O [M+H]⁺: 265.0772, found 265.0768.



1'-benzyl-1'*H*,3*H*-[**2**,3'-biindol]-3-one (4v). 47.7 mg, 71% yield. Deep red solid. ¹H NMR (500 MHz, CDCl₃) δ 8.61 (d, *J* = 7.8 Hz, 1H), 8.49 (d, *J* = 3.5 Hz, 1H), 7.52 - 7.47 (comp, 2H), 7.37 – 7.27 (comp, 7H), 7.18 (d, *J* = 6.6 Hz, 2H), 7.12 – 7.09 (comp, 1H), 5.40 (s, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 195.9, 163.6, 158.3, 137.1, 137.0, 135.9, 135.6, 129.0, 128.2, 127.3, 126.9, 126.2, 124.5, 123.9, 123.6, 122.9, 122.6, 121.2, 110.5, 107.3, 51.01. HRMS (TOF MS ESI⁺) calculated for C₂₃H₁₇N₂O [M+H]⁺: 337.1335, found 337.1340.



1'-(4-chlorobenzyl)-1'*H*,3*H*-[2,3'-biindol]-3-one (4w). 56.3 mg, 76% yield. Deep red solid. ¹H NMR (400 MHz, DMSO) δ 8.72 (s, 1H), 8.58 - 8.42 (comp, 1H), 7.62 - 7.56 (comp, 2H), 7.53 (d, *J* = 6.9 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 2H), 7.37 (d, *J* = 7.5 Hz, 1H), 7.35 - 7.25 (comp, 4H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.62 (s, 2H). ¹³C NMR (100 MHz, DMSO) δ 195.4, 163.2, 158.3, 137.8, 137.1, 136.6, 136.5, 132.8, 130.1, 129.7, 129.2, 127.1, 127.0, 125.0, 124.3, 124.2, 123.3, 122.9, 122.7, 121.3, 111.9, 106.6, 49.4. HRMS (TOF MS ESI⁺) calculated for C₂₃H₁₆ClN₂O [M+H]⁺: 371.0946, found 371.0948.



1'-phenyl-1'*H***,3***H***-[2,3'-biindol]-3-one (4x).** 42.5 mg, 66% yield. Deep red solid. ¹H NMR (400 MHz, DMSO) δ 8.60 (s, 1H), 8.58 – 8.49 (m, 1H), 7.73 – 7.65 (m, 4H), 7.60 – 7.52 (m, 4H), 7.45 – 7.36 (m, 3H), 7.23 (dd, *J* = 14.3, 7.0 Hz, 1H). ¹³C NMR (125 MHz, DMSO) δ 195.0, 162.8, 158.3, 138.2, 137.9, 136.8, 135.0, 130.6, 130.1, 128.6, 127.4, 127.3, 125.1, 125.0, 123.5, 123.4, 122.9, 121.6, 111.9, 108.3. HRMS (TOF MS ESI⁺) calculated for C₂₂H₁₅N₂O [M+H]⁺: 323.1179, found 323.1185.



2-(4-(dibenzylamino)phenyl)-3*H***-indol-3-one (4y).** 26.5 mg, 33% yield. Purple solid. ¹H NMR (400 MHz, CDCl₃) δ 8.32 (d, *J* = 9.2 Hz, 2H), 7.50 (d, *J* = 7.3 Hz, 2H), 7.40 - 7.36 (comp, 4H), 7.35 - 7.30 (d, *J* = 4.7 Hz, 4H), 7.25 (d, *J* = 6.9 Hz, 4H), 6.85 (d, *J* = 9.2 Hz, 2H), 4.77 (s, 4H). ¹³C NMR (100 MHz, DMSO) δ 195.2, 161.2, 159.7, 152.0, 138.4, 137.5, 131.2, 129.1, 127.5, 127.1, 127.0, 124.8, 123.5, 121.2, 117.7, 112.9, 54.3. HRMS (TOF MS ESI⁺) calculated for $C_{28}H_{23}N_2O$ [M+H]⁺: 403.1824, found 403.1805.

General Procedure for Scale up.



To a 100-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **1a** (6.0 mmol), indole **2a** (4.0 mmol), PPh₃AuNTf₂ (88.6 mg, 3.0 mol%), and anhydrous DCE (50 mL) were added in sequence under atmosphere of argon at 0 °C, and the reaction mixture was stirred at 0 °C for 1.0 hours. When the reaction was completed (monitored by TLC), the crude reaction mixture was quenched with saturated aqueous sodium bicarbonate (30 mL) and extracted with EtOAc (30 mL). The organic layer was washed with brine and dried over anhydrous Na₂SO₄. The solvent was evaporated in *vacuo* after filtration. The residue was purified by flash column chromatography on silica gel (Hexanes : EtOAc = 6:1 to 4:1) to give 0.89 g pure **3a** in 85% yield.

Derivatizations.



<u>Synthesis of 5:</u>² To a 10-mL oven-dried round-bottom flask containing a magnetic stirring bar, **3a** (26.2 mg, 0.1 mmol) in EtOH (2.0 mL), was added NHOH·HCl (10.4 mg, 0.3 mmol, 3.0 equiv) and pydrine (12.0 mg, 0.3 mmol, 3.0 equiv) under stirring at

room temperature. And the reaction mixture was stirred for 24 h under these conditions. When the reaction was completed (monitored by TLC), the solvent was evaporated under vacuum after filtering through a pad of Celite. The residue was purified by column chromatography on silica gel (Hexanes : EtOAc = 2:1) to give 16.6 mg of the product **5** as orange solid, 60% yield. ¹H NMR (500 MHz, DMSO) δ 13.53 (s, 1H), 11.95 (s, 1H), 8.77 (s, 1H), 8.40 (d, *J* = 8.1 Hz, 1H), 8.24 (d, *J* = 7.3 Hz, 1H), 7.66 (d, *J* = 3.9 Hz, 2H), 7.56 – 7.53 (comp, 1H), 7.51 (d, *J* = 8.0 Hz, 1H), 7.22 (t, *J* = 7.5 Hz, 1H), 7.14 – 7.10 (comp, 1H). ¹³C NMR (125 MHz, DMSO) δ 148.5, 145.7, 136.6, 135.0, 131.9, 131.1, 129.7, 126.9, 125.2, 124.2, 122.8, 120.5, 118.7, 113.3, 112.4, 103.3. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₁N₃O₂ [M+H]⁺: 278.0924, found 278.0922.



<u>Synthesis of 6:</u>³ To a 10-mL oven-dried round-bottom flask containing a magnetic stirring bar, and **3a** (26.2 mg, 0.1 mmol) in Et₂O (2.0 mL), was added LiAlH₄ (3.8 mg, 0.1 mmol, 1.0 equiv) under stirring at 0 °C, and the reaction mixture was stirred at room temperature for additional 1.0 hour. When the reaction was completed (monitored by TLC), the solvent was evaporated under vacuum after filtering through a pad of Celite. The residue was purified by column chromatography on silica gel (Hexanes : EtOAc = 6:1) to give 20.9 mg of the reduction product as deep-red solid, 85% yield.

Then to another 10-mL oven-dried round-bottom flask containing a magnetic stirring bar, the above obtained product (20.9 mg, 0.085 mmol), and Li_2CO_3 (1.9 mg, 30 mol %), in DCM (1.0 mL), was added a solution of *m*-CPBA (29.2 mg, 0.17 mmol 2.0 equiv) in DCM (1.0 mL) under stirring at 0 °C, and the reaction mixture was stirred at room temperature for 5 minutes. Then, the reaction mixture was quenched

with aqueous Na₂SO₃, and extracted with ether, washed with cold aqueous NaHCO₃ and aqueous NaCl in sequence. Then the organic phase was dried over anhydrous MgSO₄. After evaporating the solvent after filtration, the crude product was purified by flash chromatography on silica gel (Hexanes : EtOAc = 15:1) to give 17.1 mg of pure product **6** as white solid, 66% yield over 2 steps. ¹H NMR (400 MHz, DMSO) δ 12.13 (s, 1H), 8.44 (comp, 1H), 8.30 (s, 1H), 8.12 (d, *J* = 7.8 Hz, 1H), 7.91 (t, *J* = 7.5 Hz, 1H), 7.68 (d, *J* = 8.0 Hz, 1H), 7.56 - 7.46 (comp, 2H), 7.33 - 7.22 (comp, 2H). ¹³C NMR (125 MHz, DMSO) δ 159.2, 155.6, 147.5, 136.9, 136.7, 131.5, 127.9, 126.8, 126.1, 124.8, 122.8, 121.4, 121.2, 116.2, 112.4, 106.3. HRMS (TOF MS ESI⁺) calculated for C₁₆H₁₀N₂O₂ [M+H]⁺: 263.0815, found 263.0815.

Control Experiment.



To a 10-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **1a** (14.7 mg, 0.1 mmol), PPh₃AuNTf₂ (3.7 mg, 5.0 mol%), and anhydrous DCE (2.0 mL) were added in sequence under atmosphere of argon at 0 °C. Then the reaction mixture was subjected to proton NMR analysis after 24 hours under these conditions. All the material **1a** was consumed and decomposed in to a complex mixture after 24 hours (see Figure S1, the third spectrum). Then, indole **2a** (7.8 mg, 0.07 mmol) was added to the above reaction mixture, leading to no further reaction at all (see Figure S1, the fourth spectrum).



Figure S1. Proton NMR spectra of crude reaction mixture of 1a.



To a 10-mL oven-dried vial containing a magnetic stirring bar, o-nitroalkyne **1a** (14.7 mg, 0.1 mmol), AuCl₃ (4.6 mg, 7.0 mol%), and PhMe (1.0 mL) were added in sequence under atmosphere of argon at room temperature. Then the reaction was monitored by TLC and LCMS. All the material **1a** was consumed and decomposed in to a complex mixture after 12 hours (see Figure S2, the MS signal 293.09 might be the dimer product of **1a**). Then, indole **2a** (7.8 mg, 0.07 mmol) was added shortly to the above reaction mixture, leading to no further reaction at all.



Figure S2. LCMS monitoring of the crude reaction of 1a with 2a.



To a 10-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **8** (22.3 mg, 0.1 mmol), PPh₃AuNTf₂ (3.7 mg, 5.0 mol%), and anhydrous DCE (2.0 mL) were added in sequence under atmosphere of argon at 0 °C. Then the reaction mixture was subjected to proton NMR analysis after 1.0 hour under these conditions. Most of **8** remained intact, and minor amount of **9** was detected by proton NMR (10%, see Figure S3).



Figure S3. Proton NMR spectra of crude reaction mixture of 8.



To a 10-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **8** (33.5 mg, 0.15 mmol), indole **2a** (11.7 mg, 0.1 mmol), PPh₃AuNTf₂ (3.7 mg, 5.0 mol%), and anhydrous DCE (2.0 mL) were added in sequence under atmosphere of argon at 0 °C. Then the reaction mixture was subjected to proton NMR analysis after 1.0 hour under these conditions. Most of **8** and indole remained intact, and minor amount of **9** was detected by proton NMR (13%, see Figure S4).



Figure S4. Proton NMR spectra of crude reaction mixture of 8 with indole 2a.



To a 10-mL oven-dried vial containing a magnetic stirring bar, **9** (22.3 mg, 0.1 mmol), indole **2a** (11.7 mg, 0.1 mmol), PPh₃AuNTf₂ (3.7 mg, 5.0 mol%), and anhydrous DCE

(2.0 mL) were added in sequence under atmosphere of argon at 0 °C. Then the reaction mixture was subjected to proton NMR analysis after 1.0 hour under these conditions. Most of **9** and indole **2a** remained intact, and the desired product **10** reported by Jia was not detected (see Figure S5).⁴



Figure S5. Proton NMR spectra of crude reaction mixture of 9 with indole 2a.



To a 10-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **1a** (44.1 mg, 0.3 mmol), indole **2a** (23.4 mg, 0.2 mmol), PPh₃AuNTf₂ (7.4 mg, 5.0 mol%), and anhydrous DCE (4.0 mL) were added in sequence under atmosphere of argon at 0 °C, and the reaction mixture was stirred at 0 °C for 1.0 hours. The the reaction was subjected to the LCMS analysis. MS signal for intermediate **IV** was observed (see Figure S6).



Figure S6. LCMS monitoring of the crude reaction of 1a with 2a.



To a 10-mL oven-dried vial containing a magnetic stirring bar, *o*-nitroalkyne **1a** (44.1 mg, 0.3 mmol), indole **2a** (23.4 mg, 0.2 mmol), PPh₃AuNTf₂ (7.4 mg, 5.0 mol%), and anhydrous DCE (4.0 mL) were added in sequence under atmosphere of argon at 0 °C, and the reaction mixture was stirred at 0 °C for several hours. The reaction mixture was monitored by crude proton NMR after 2 and 6 hours, the ratio of **3a**:**4a** was decreased from 10:1.4 to 10:2.9, and this ratio could reach to 10:5.3 by adding additional 5 mol% of gold catalyst to the reaction mixture and stirring for 2 hours. (see Figure S7).



Figure S7. Proton NMR monitoring of the model reaction at different reaction times.





40.59 40.17 40.17 39.76 39.76 39.35











210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

















ZS-T64-F







10 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -1 f1 (ppm)











































10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)













6.0 5.5 5.0 f1 (ppm) 7.5 4.5 4.0 3.5 8.5 7.0 0.5 0.0 10.0 9.5 9.0 6.5 2.5 2.0 1.5 1.0 8.0 3.0











90 80 fl (ppm)

Crystallographic Data for Compound 3a



R(reflections) =	0.0945(4707)	wR2(reflections)=	0.3277(9707)
		,			- · - · /

S = 1.034 Npar= 722

Crystallographic Data for Compound 4w



CCDC: 2022225



Bond precision	: $C-C = 0.0082$ A		Waveleng	gth=1.54184
Cell:	a=8.3666(3)	b=15.635	9(10)	c=16.4108(8)
Temperature:	alpha=113.858(5) 100 K	beta=92.	451(4)	gamma=103.993(5)
	Calculated		Reporte	ed
Volume	1881.09(19)		1881.08	3(18)
Space group	P -1		P -1	
Hall group	-P 1		-P 1	
Moiety formula	2(C23 H15 C1 N2 C12	O), C H2	2(C23 H C12	H15 Cl N2 O), C H2
Sum formula	C47 H32 C14 N4 O	2	C47 H32	2 Cl4 N4 O2
Mr	826.57		826.56	
Dx,g cm-3	1.459		1.459	
Z	2		2	
Mu (mm-1)	3.243		3.243	
F000	852.0		852.0	
F000′	856.96			
h,k,lmax	10,19,20		10,19,2	20
Nref	7876		7466	
Tmin, Tmax	0.759,0.723		0.747,3	1.000
Tmin'	0.689			
Correction met AbsCorr = MULT	hod= # Reported T I-SCAN	Limits: T	min=0.74	7 Tmax=1.000
Data completen	ess= 0.948	Theta(m	nax)= 76	.178
R(reflections)	= 0.1241(5798)	wR2(ref	lection	s = 0.3400(7466)

S = 1.318 Npar= 514

General Procedure for the in vitro Anti-tumor Activity Study

Cell viability was measured by CCK-8 assay

Human small cell lung cancer cell lines H446 and H128 were obtained from American Type Culture Collection (ATCC). Cells were cultured in RPMI1640 medium containing 10% fetal bovine serum and 1% penicillin/ streptomycin (Gibco) in a humidified incubator containing 5% CO₂ at 37 °C. For cell viability, cells were seeded in 96-well plates at 1000 cells per well. After 24 hours, serially diluted compounds were added and cells were cultured for another 96 hours. Cell viability was measured using a Cell Counting Kit-8 (CCK-8) assay according to the manufacturer's instructions (Beyotime Biotechnology, China). The results were presented as percentages and vehicle-treated cells set at 100 (Figure S7).



Figure S8. Compounds 3c-6 on the viability of H446 and H128 cells.

These representative products **3c-3d**, **3h**, **3i**, **3k**, **3r**, **3s**, **4s**, and **6** on cell viability was evaluated *via* CCK8 assay in SCLC cells (H446 and H128), and the *in vitro* anti-tumor activity results are listed in Table S1. Most of these 2,3'-biindole derivatives showed significant anti-cancer activity in comparison to the reported 2-aryl analogous, compounds **3d** and **4s** were more potent than other compounds.

Moreover, the substitutions of the left benzene ring on isatogens might affect the activity and showed lower cytotoxicity (the following order of potency: 3c, 3d, 3e, 3h, 3i > 3k, 3r, 3s). The structural differences between 3s and 4s showed that the reduction product 4s without the *N*-oxide part was more effective. Whereas, the B-V oxidation product 6 was less potent than the other tested compounds in SCLC cells.

Compound	H446	H128
3c	0.68	2.84
3d	0.83	2.25
3e	0.48	3.01
3h	0.50	3.74
3i	0.56	3.46
3k	0.93	4.19
3r	1.24	4.94
3 s	1.66	>10
4 s	0.47	1.67
6	>10	>10

Table S1 Anti-tumor activities of compounds 3c-6 (IC₅₀, μ M)^a

^{*a*}IC₅₀ is the half maximal inhibitory concentration.

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