

Enantiopure Isothiourea@Carbon-Based Support: Stacking Interactions for Recycling a Lewis Base in Asymmetric Catalysis

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Supporting Information

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1. General Information.

Unless otherwise noted, all reactions were performed in oven-dried Schlenk tubes with Schlenk line connected to dry argon. All reagents were obtained commercially and used without further purification. Solvents were dried by distillation under argon from the followings: tetrahydrofuran and diethyl ether (sodium/benzophenone); toluene (calcium hydride); dichloromethane (calcium hydride). K_2CO_3 , Cs_2CO_3 , and $KOtBu$ were dried by heating at 110 °C for 12 h, left to cool under argon before using. Organic extracts were, in general, dried over anhydrous magnesium sulfate ($MgSO_4$). Precursors purchased from Sigma-Aldrich, Acros, TCI, Alfa Aesar and Fluorochem. Commercial reduced graphene oxide was purchased from Graphenea.

NMR spectrum: 1H and ^{13}C spectra are recorded on AM 250, AV 300 or AV 360 Bruker spectrometers, operating at 250, 300 or 360 MHz for 1H NMR, ^{19}F NMR and 75 or 90 MHz for ^{13}C NMR. Chemical shifts are reported in ppm (δ) relative with the solvent signal as reference. Chemical shifts are reported downfield from $CDCl_3$ (δ : 7.26 ppm), CD_3OD (δ : 3.31 ppm) or $(CD_3)_2SO$ (δ : 2.50 ppm) for 1H NMR. Chemical shifts of ^{13}C NMR are reported in the scale relative to the solvent of $CDCl_3$ (δ : 77.0 ppm), CD_3OD (δ : 49.0 ppm) or $(CD_3)_2SO$ (δ : 39.5 ppm) used as an internal reference. Data are represented as follows: chemical shift, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants in Hertz (Hz), integration.

Mass spectroscopy: Mass spectra were recorded on MicroTOF-Qq Bruker spectrometer by electrospray ionization (ESI) or QTOF 6540 Agilent spectrometer by atmospheric pressure photoionization (APPI).

Infrared spectra: IR spectra (in cm^{-1}) were recorded on a FTIR spectrometer (Perkin-Elmer spectrum one, NaCl pellets or Bruker Vertex 70 ATR Pike Germanium).

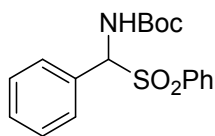
High Performance Liquid Chromatography: HPLC analysis was performed using a JASCO pump-PU 2089 associated to an UV detector (UV 100) from TSP, using Daicel Chiralpak AD-H and Chiralcel OD-H columns. Supercritical fluid chromatography (SFC) separation was performed with Investigator SFC System (Waters) equipped with a photodiode array UV detector. Data are reported as follows: column type, temperature, eluent, flow rate, pressure in column, retention time.

Spectropolarimeter: Optical rotations were measured on an Anton Paar's MCP 150 polarimeter and reported as follows: $[\alpha]_D^{T_D}$ (concentration (g/100 mL), solvent).

Chromatography: Flash column chromatography was performed using silica gel Merck (0.04–0.063 μm).

2. Preparation of *pyr*-hyperBTM.

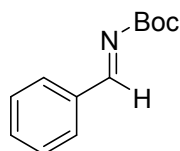
tert-Butyl (phenyl(phenylsulfonyl)methyl)carbamate



1a

Following the method described by List et al.:¹ to a stirred solution of *tert*-butyl carbamate (7.03 g, 60 mmol) in THF (26 mL) in a 250 mL three neck round bottom flask equipped with an overhead stirrer was added water (53 mL), sodium benzenesulfinate (10.0 g, 61 mmol), benzaldehyde (6.2 mL, 61 mmol) and formic acid (13 mL, 343 mmol). The reaction mixture was stirred for 24 h at rt. The resulting precipitate was filtered and washed with water (25 mL) followed by hexane/CH₂Cl₂ (91/9) (2 × 25 mL). Stirring the combined liquors from filtration and the water wash for a further 24 h provided a second, smaller quantity of solid. The combined solids were dried in an oven at 90 °C until a constant weight, giving sulfone (±)-**1a** as a white solid (14.35 g, 69%); ¹H NMR (300 MHz, *d*₆-DMSO) δ = 7.93-7.90 (m, 2H), 7.75-7.56 (m, 6H), 7.45-7.40 (m, 3H), 6.03 (d, *J* = 10.8 Hz, 1H), 1.22 (s, 9H) ppm. The spectral data match those previously reported.¹

tert-Butyl benzylidenecarbamate

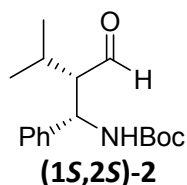


1b

A 500 mL, two-necked, round-bottomed flask equipped with a magnetic stirring bar, a rubber septum, and a reflux condenser capped with an inlet adaptor connected to an argon-vacuum manifold, was charged with anhydrous potassium carbonate (33.17 g, 240 mmol, 6.0 equiv.). The solid was placed under vacuum (0.5 mmHg) and flame-dried. The flask was purged with argon and anhydrous tetrahydrofuran (250 mL) was added via cannula under argon at 25 °C. Then, the septum was removed and compound **1a** (13.90 g, 40 mmol, 1.0 equiv.) was added into the flask. The septum was exchanged for a glass stopper, and the resulting suspension was heated to reflux at 80-85 °C (oil bath temp) with vigorous stirring under argon. After 15 h, the reaction was cooled to 25 °C and the solid is filtered off through alternating layers of Celite (1 cm thick)/Na₂SO₄(1 cm thick)/Celite (1 cm thick) using a Büchner funnel (diameter 100 mm), and washed with anhydrous tetrahydrofuran (50 mL). The filtrate is concentrated at 30 °C (water bath temperature) by rotary evaporation (20 mmHg) and dried under vacuum (0.5

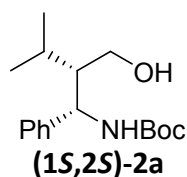
mmHg) to give the corresponding *N*-Boc-imine **1b** as colorless oil (8.21 g, 99%); ¹H NMR (300 MHz, *d*₆-acetone) δ = 8.79 (s, 1H), 7.97-7.94 (m, 2H), 7.64-7.61 (m, 1H), 7.58-7.53 (m, 2H), 1.54 (s, 9H) ppm. The spectral data match those previously reported.¹

***tert*-Butyl ((1*S*,2*S*)-2-formyl-3-methyl-1-phenylbutyl)carbamate**



Following the method described by List et al.:¹ to a stirred solution of imine **1b** (8.21 g, 40 mmol) in CH₃CN (350 mL) in a flame dried 1 L three neck round bottom flask equipped with an overhead stirrer under argon was added isovaleraldehyde (8.6 mL, 80 mmol). The resulting solution was cooled to 0 °C in an ice/brine bath. (*L*)- proline (921 mg, 8 mmol) was added and the reaction mixture was stirred for 14 h at 0 °C before warming to rt. Water (110 mL), Et₂O (70 mL) and brine (15 mL) was added and the organic layer was separated. The aqueous layer was extracted with Et₂O (2 × 30 mL). The organic extracts were combined, dried (MgSO₄), filtered and concentrated in *vacuo*. Trituration of the resulting solid with hexane (2 × 40 mL) gave aldehyde (1*S*,2*S*)-2 (>99:1 dr) as a white solid (6.27 g, 54%); ¹H NMR (300 MHz, CDCl₃) δ = 9.49 (d, *J* = 4.2 Hz, 1H), 7.36-7.22 (m, 5H), 5.17-5.01 (m, 2H), 2.52-2.46 (m, 1H), 2.16-2.03 (m, 1H), 1.40 (s, 9H), 1.14 (d, *J* = 6.6 Hz, 3H), 1.02 (d, *J* = 6.6 Hz, 3H) ppm; Chiral HPLC Chiralcel OD-H (2% IPA:hexane, flow rate 1 mL min⁻¹, 220 nm, 30 °C) *t*_R(1*R*,2*R*): 8.19 min, *t*_R(1*S*,2*S*): 9.43 min, 99% ee. The spectral data match those previously reported.¹

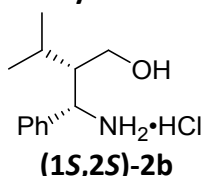
***tert*-Butyl ((1*S*,2*S*)-2-(hydroxymethyl)-3-methyl-1-phenylbutyl)carbamate**



Following a slightly modified method from that described by Smith et al.:² to a slurry of aldehyde (1*S*,2*S*)-2 (6.27 g, 21.5 mmol) in methanol (125 mL) at 2 °C in a 500 mL three neck round bottom flask equipped with an overhead stirrer, was added NaBH₄ (1.25 g, 33 mmol) portion wise over 30 min and the reaction mixture was left to stir for 1 h at rt. Saturated aqueous NaHCO₃ was added (8 mL) drop wise over 10 min forming a white precipitate. The methanol was removed in *vacuo* and water (80 mL) and CH₂Cl₂ (60 mL) were added. The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 20 mL). The organic

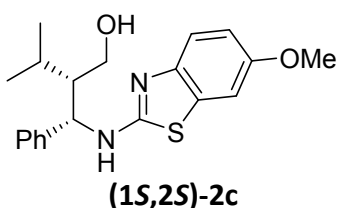
extracts were combined, dried (MgSO₄), filtered and concentrated in *vacuo* and dried for 2 h at 2 mbar to give alcohol (1*S*,2*S*)-**2a** as a white solid (6.27 g, 99%); ¹H NMR (300 MHz, CDCl₃) δ = 7.38-7.23 (m, 5H), 5.50 (d, *J* = 9.3 Hz, 1H), 5.03 (s, br, 1H), 3.70-3.66 (m, 1H), 3.51 (dd, *J* = 11.4, 8.7 Hz, 1H), 1.87-1.71 (m, 3H), 1.42 (s, 9H), 1.00 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* = 6.9 Hz, 3H) ppm. The spectral data match those previously reported.²

(*S*)-2-((*S*)-Amino(phenyl)methyl)-3-methylbutan-1-ol hydrochloride



To a slurry of alcohol (1*S*,2*S*)-**2a** (6.27 g, 21.5 mmol) in dioxane (20 mL) was added 4M HCl in dioxane (26.5 mL, 106 mmol) drop wise over 30 min and the reaction mixture was left to stir for 12 h at rt. 15 mL of dioxane was removed in *vacuo* and the solution cooled to 15 °C to precipitate a white solid. Et₂O (10 mL) was added and the precipitate was filtered, washed with Et₂O (2 × 20 mL) and the resulting white solid dried (15 mbar, 45 °C) for 2 h to give amino alcohol (1*S*,2*S*)-**2b** as a white solid (4.39 g, 89%); ¹H NMR (300 MHz, *d*₄-methanol) δ = 7.55-7.40 (m, 5H), 4.58 (d, *J* = 4.2 Hz, 1H), 3.76 (ddd, *J* = 10.8, 4.8, 0.9 Hz, 1H), 3.49 (dd, *J* = 10.5, 9.6 Hz, 1H), 2.08-2.00 (m, 1H), 1.60-1.48 (m, 1H), 1.14 (d, *J* = 6.9 Hz, 3H), 0.83 (d, *J* = 6.9 Hz, 3H) ppm. The spectral data match those previously reported.²

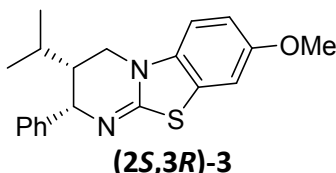
(*S*)-2-((*S*)-((6-Methoxybenzo[*d*]thiazol-2-yl)amino)(phenyl)methyl)-3-methylbutan-1-ol



Following a procedure outlined by Smith,³ 2-chloro-6-methoxybenzo[*d*]thiazole (3.3 g, 16.7 mmol, 1 equiv.) was added to a solution of (*S*)-2-((*S*)-amino(phenyl)methyl)-3-methylbutan-1-ol hydrochloride **2b** (4.0 g, 17.5 mmol, 1.05 equiv.), and *i*-Pr₂NEt (11 mL, 66.8 mmol, 4 equiv.) in *o*-dichlorobenzene (8 mL, 2.0 M). The resulting pale-yellow suspension was heated at reflux until completion. The resulting mixture was cooled to rt, H₂O (12 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 15 mL). The combined organic fractions were washed with brine, dried (MgSO₄), and concentrated in *vacuo* to afford the crude product which was purified by column chromatography (CH₂Cl₂: EtOAc = 5:1) to afford the amino

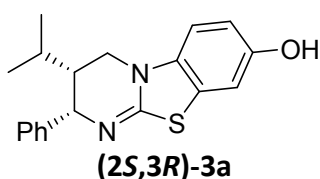
alcohol product (1*S*,2*S*)-**2c** as a brown solid (4.37 g, 73% yield); ¹H NMR (300 MHz, CDCl₃) δ = 7.46-7.26 (m, 6H), 7.05 (d, *J* = 2.4 Hz, 1H), 6.85 (dd, *J* = 9.0, 2.7 Hz, 1H), 5.01 (d, *J* = 4.5 Hz, 1H), 3.83-3.78 (m, 4H), 3.62 (t, *J* = 10.2 Hz, 1H), 2.12-2.04 (m, 1H), 1.80-1.73 (m, 1H), 1.08 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* = 6.9 Hz, 3H) ppm.

(2*S*,3*R*)-3-Isopropyl-8-methoxy-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine



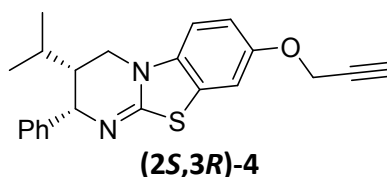
To a slurry of the crude amino alcohol (1*S*,2*S*)-**2c** (4.35 g, 12.2 mmol, 1 equiv.) in anhydrous CH₂Cl₂ (61 mL, 0.2 M) was added Et₃N (6.78 mL, 48.8 mmol, 4 equiv.) and the reaction mixture was cooled to 0 °C. Methanesulfonyl chloride (1.22 mL, 15.8 mmol, 1.3 equiv.) was added and the reaction mixture was stirred at rt for 4 h. Once complete consumption of the amino alcohol was observed (about 4 h), *i*-PrOH (4.7 mL, 5 equiv.) was added and the reaction was heated at reflux for 24 h. The reaction was quenched with 1 M aq. NaOH (16 mL) and the biphasic mixture stirred vigorously for 30 min. The aqueous layer was extracted with CH₂Cl₂ (3 × 20 mL) and the combined organic fractions washed with brine (50 mL), dried (MgSO₄) and concentrated in *vacuo* to afford the crude product which was purified by column chromatography (CH₂Cl₂: EtOAc = 5:1) to afford an off white solid that was recrystallised from 50% EtOAc in Hexane to give (2*S*,3*R*)-3-isopropyl-8-methoxy-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine **3** as a colourless solid (3.3 g, 80%); [α]_D²⁰ +308.0 (c 1.0 in CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ = 7.33-7.19 (m, 5H), 6.94 (d, *J* = 2.4 Hz, 1H), 6.78 (dd, *J* = 8.7, 2.4 Hz, 1H), 6.71 (d, *J* = 8.7 Hz, 1H), 4.90 (d, *J* = 3.9 Hz, 1H), 3.86-3.80 (m, 4H), 3.33 (dd, *J* = 11.4, 11.4 Hz, 1H), 2.00-1.90 (m, 1H), 1.37-1.25 (m, 1H), 1.13 (d, *J* = 6.6 Hz, 3H), 0.84 (d, *J* = 6.6 Hz, 3H) ppm; ¹³C{¹H} NMR (90 MHz, CDCl₃): δ = 158.4, 140.8, 134.9, 128.0, 127.1, 124.1, 111.5, 108.2, 107.8, 61.1, 56.0, 42.0, 40.8, 26.9, 22.0, 20.0 ppm; HRMS (ESI) calcd. for [M+H]⁺ C₂₀H₂₃N₂OS, 339.1526, found 339.1527 (0.3 ppm).

(2*S*,3*R*)-3-Isopropyl-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidin-8-ol



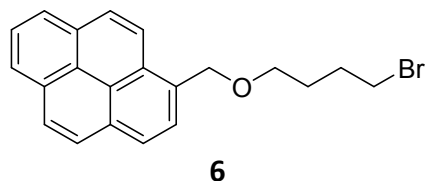
BBr₃ (42 mL, 42 mmol, 1 M in CH₂Cl₂, 6 equiv.) was added dropwise to a solution of (2*S*,3*R*)-3-isopropyl-8-methoxy-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine **3** (2.37 g, 7 mmol, 1 equiv.) in CH₂Cl₂ (47 mL) at 0 °C. The solution was stirred at 0 °C for 2 h then warmed to rt and stirred for 16 h. The reaction was carefully quenched with MeOH (50 mL). Then saturated NaHCO₃ was added until pH reached *ca.* 8. CH₂Cl₂ (50 mL) was added and the aqueous phase was extracted with CH₂Cl₂ (3 × 100 mL). The organic layers were combined, dried (MgSO₄) and concentrated in *vacuo* to afford (2*S*,3*R*)-3-isopropyl-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidin-8-ol **3a** as a colourless solid (1.69 g, 74%). ¹H NMR (360 MHz, *d*₆-DMSO) δ = 9.27 (s, 1H), 7.32 (dd, *J* = 7.9, 6.8 Hz, 2H), 7.26-7.19 (m, 3H), 6.93 (d, *J* = 2.5 Hz, 1H), 6.90 (d, *J* = 8.6 Hz, 1H), 6.65 (dd, *J* = 8.6, 2.5 Hz, 1H), 4.76 (d, *J* = 4.3 Hz, 1H), 3.90-3.85 (m, 1H), 3.32-3.26 (m, 1H), 1.91-1.83 (m, 1H), 1.19-1.10 (m, 1H), 0.99 (d, *J* = 6.5 Hz, 3H), 0.75 (d, *J* = 6.8 Hz, 3H) ppm; ¹³C{¹H} NMR (90 MHz, *d*₆-DMSO): δ = 157.1, 153.1, 142.1, 133.8, 128.6, 128.2, 127.2, 122.8, 113.2, 109.6, 109.3, 60.8, 42.1, 27.0, 22.4, 19.9 ppm; HRMS (ESI) calcd. for [M+H]⁺ C₁₉H₂₁N₂OS, 325.1369, found 325.1371 (0.5 ppm).

(2*S*,3*R*)-3-Isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine **11**



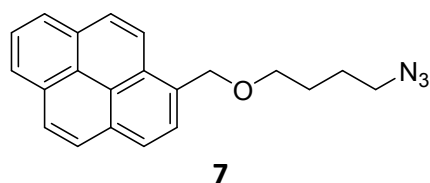
KOtBu (1.42 g, 12.74 mmol, 2.6 equiv.) was added to a solution of (2*S*,3*R*)-3-isopropyl-2-phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidin-8-ol **3a** (1.59 g, 4.9 mmol, 1 equiv.) in THF/DMSO (1:1, 36 mL) at 0 °C, and the reaction mixture was stirred for 2 h. Propargyl bromide (0.8 mL, 7.35 mmol, 80% in toluene, 1.5 equiv.) was added and the reaction mixture was allowed to warm to rt (*ca.* 2 h) and quenched with brine (30 mL). The aqueous phase was extracted with EtOAc (3 × 30 mL). The combined organic fractions were washed with brine, dried (MgSO₄) and concentrated in *vacuo*. The residue was purified by column chromatography (petroleum ether: EtOAc = 1:1) to afford (2*S*,3*R*)-3-isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine **4** as a brown solid (1.02 g, 57%); ¹H NMR (250 MHz, CDCl₃) δ = 7.32-7.17 (m, 5H), 7.01 (d, *J* = 2.5 Hz, 1H), 6.85 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.71 (d, *J* = 8.5 Hz, 1H), 4.89 (dd, *J* = 4.5, 1.5 Hz, 1H), 4.65 (d, *J* = 2.5 Hz, 2H), 3.81 (ddd, *J* = 11.5, 5.2, 1.8 Hz, 1H), 3.31 (t, *J* = 11.5 Hz, 1H), 2.53 (t, *J* = 2.5 Hz, 1H), 1.97-1.85 (m, 1H), 1.32-1.23 (m, 1H), 1.11 (d, *J* = 6.2 Hz, 3H), 0.82 (d, *J* = 6.5 Hz, 3H) ppm. The spectral data match those previously reported.³

1-((4-Bromobutoxy)methyl)pyrene



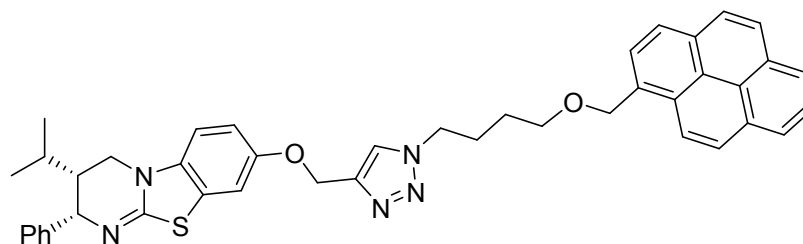
NaH (8.0 g, 200 mmol, 20 equiv.) was added to a solution of 1-pyrenemethanol (2.32 g, 10 mmol, 1 equiv.) in 100 mL dry THF under N₂ atmosphere at room temperature. The mixture was stirred for 0.5 h. Then 1,4-dibromobutane (2.16 g, 100 mmol, 10 equiv.) was added and stirred for 48 h. The solvent was removed by vacuum evaporation, and water was carefully added to the reaction mixture to quench the unreacted NaH. The solution was extracted with CH₂Cl₂ (3 × 100 mL), and the combined organic layer was washed with 5% aqueous HCl (100 mL), 10% aqueous Na₂CO₃ (100 mL) and finally with water, and then was dried over anhydrous MgSO₄. The organic phase was concentrated and purified by silica gel column chromatography (petroleum ether: EtOAc = 10:1) to afford **6** as a white solid (3.29 g, 89 %); ¹H NMR (300 MHz, CDCl₃) δ = 8.35 (d, *J* = 9.3 Hz, 1H), 8.22-8.14 (m, 4H), 8.06-8.00 (m, 4H), 5.20 (s, 2H), 3.62 (t, *J* = 6.0 Hz, 2H), 3.41 (t, *J* = 6.6 Hz, 2H), 2.03-1.94 (m, 2H), 1.85-1.76 (m, 2H) ppm. The spectral data match those previously reported.⁴

1-((4-Azidobutoxy)methyl)pyrene



The 1-((4-bromobutoxy)methyl)pyrene **6** (3.28 g, 9 mmol, 1 equiv.) was dissolved in 30 mL DMF. Sodium azide (641 mg, 10 mmol, 1.1 equiv.) was then added and the reaction mixture was stirred at rt under argon atmosphere. The reaction mixture was monitored by TLC and when complete, the DMF was removed under high vacuum. The crude compound was purified by flash chromatography on silica gel (petroleum ether: EtOAc = 10:1) to obtain the desired product **7** as a white solid (2.95 g, 99 %). ¹H NMR (360 MHz, CDCl₃): δ = 8.35 (d, *J* = 9.0 Hz, 1H), 8.20 (ddd, *J* = 7.6, 7.6, 1.1 Hz, 2H), 8.16 (s, 1H), 8.14 (d, *J* = 1.1 Hz, 1H), 8.06 (s, 2H), 8.02 (dd, *J* = 7.6, 7.6 Hz, 2H), 5.19 (s, 2H), 3.61 (t, *J* = 6.1 Hz, 2H), 3.26 (t, *J* = 6.1 Hz, 2H), 1.74-1.70 (m, 4H) ppm; ¹³C{¹H} NMR (90 MHz, CDCl₃): δ = 131.5, 131.3, 130.9, 129.3, 127.7, 127.4, 126.9, 125.3, 125.0, 124.8, 124.5, 123.4, 71.6, 69.7, 51.3, 27.1, 25.9 ppm. HRMS (ESI) calcd. for [M+Na]⁺ C₂₁H₁₉N₃ONa 352.1420, found 352.1412 (2.4 ppm).

(2*S*,3*R*)-3-isopropyl-2-phenyl-8-((1-(4-(pyren-1-ylmethoxy)butyl)-1*H*-1,2,3-triazol-4-yl)methoxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine



(2*S*,3*R*)-pyr-hyperBTM

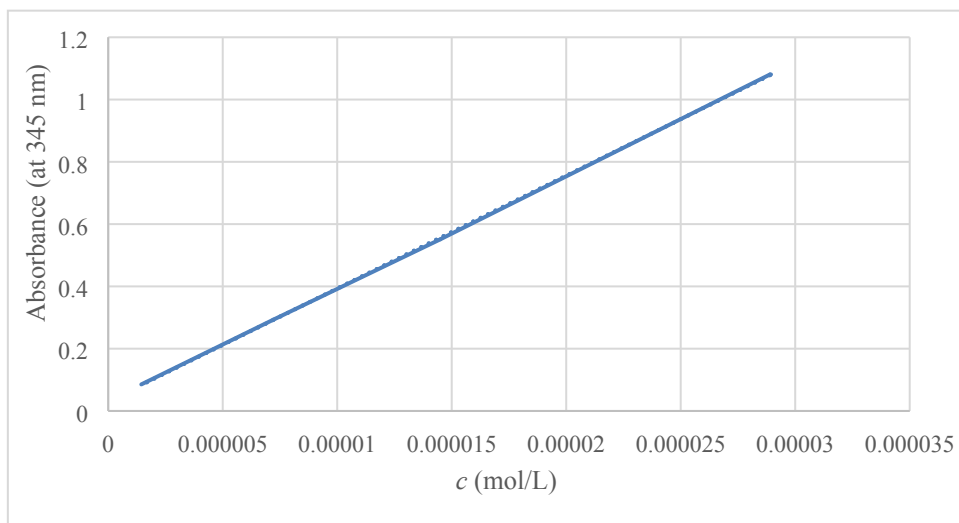
Under argon, a dry schlenk was charged with (2*S*,3*R*)-3-isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine **4** (980 mg, 2.7 mmol, 1 equiv.), 1-((4-azidobutoxy)methyl)pyrene **7** (978 mg, 2.97 mmol, 1.1 equiv.), CuI (25.7 mg, 5 mol%), degassed THF (25 ml) and a stirring bar. Anhydrous DIPEA was added (1.56 ml, 9.45 mmol, 3.5 equiv.) and the reaction mixture was stirred at 40° C for 16 h. After removing the solvent under reduced pressure, followed by a trituration in Et₂O, the resulting solid was recovered after filtration. The mixture was then diluted in DCM (10 mL) and washed with an EDTA solution (0.05 M). After drying the combined organic phases on MgSO₄, filtration and solvent evaporation, the final pure product (2*S*,3*R*)-**pyr-hyperBTM** was obtained as a yellow solid (1.0 g, 54%). $[\alpha]^{20}_{\text{D}} +105.0$ (c 1.0 in CHCl₃); ¹H NMR (360 MHz, CDCl₃) δ = 8.34 (d, *J* = 9.0 Hz, 1H), 8.20-8.12 (m, 4H), 8.06-7.98 (m, 4H), 7.37 (s, 1H), 7.34-7.21 (m, 5H), 6.95 (d, *J* = 2.5 Hz, 1H), 6.82 (dd, *J* = 8.6, 2.5 Hz, 1H), 6.66 (d, *J* = 9.0 Hz, 1H), 5.18 (s, 2H), 5.03 (s, 2H), 4.91 (d, *J* = 4.3 Hz, 1H), 4.29 (t, *J* = 7.2 Hz, 2H), 3.81-3.76 (m, 1H), 3.63 (t, *J* = 6.1 Hz, 2H), 3.31 (t, *J* = 11.5 Hz, 1H), 2.04-1.86 (m, 3H), 1.70-1.62 (m, 2H), 1.34-1.25 (m, 1H), 1.12 (d, *J* = 6.5 Hz, 3H), 0.81 (d, *J* = 6.8 Hz, 3H) ppm; ¹³C{¹H} NMR (90 MHz, CDCl₃): δ = 158.6, 153.9, 143.7, 140.6, 135.2, 131.4, 131.2, 130.8, 129.4, 128.4, 128.0, 127.8, 127.5, 127.4, 127.2, 127.1, 126.1, 125.3, 124.9, 124.7, 124.5, 123.4, 122.8, 112.6, 109.5, 107.9, 71.7, 69.4, 62.8, 61.0, 50.1, 42.0, 40.8, 27.4, 26.9, 26.6, 22.0, 20.0 ppm. HRMS (ESI) calcd. for [M+H]⁺ C₄₃H₄₂N₅O₂S 692.3054, found 692.3029 (3.6 ppm).

3. General procedure for the immobilization of *pyr*-hyperBTM onto rGO

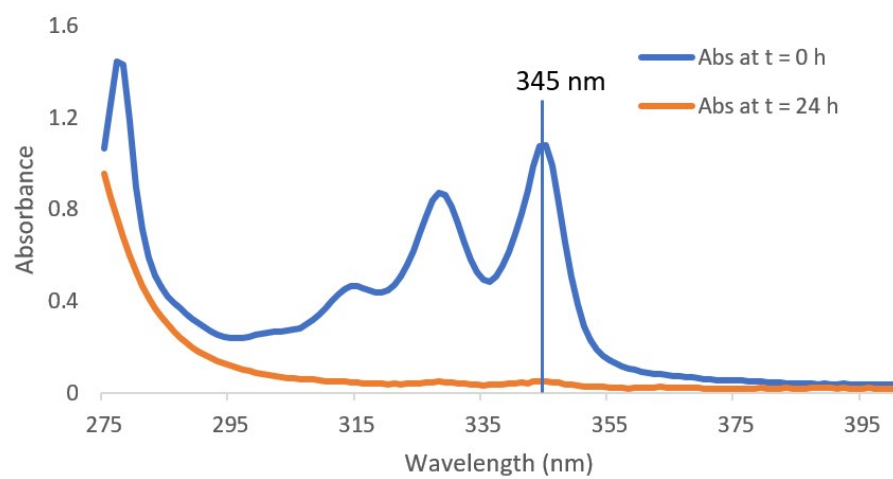
A glass vial with a screw cap septum top was charged with 80 mg of reduced graphene oxide (rGO) and 3.2 mL of anhydrous DCM. The suspension was immersed in an ultrasound bath for 1 hour, then the *pyr*-hyperBTM (20 mg) was added, and the mixture was sonicated for another 5 minutes. After stirring for 24 hours at room temperature under air, the agitation was stopped and 10 μ L were taken from the supernatant solution, diluted in 20 mL of DCM and then subjected to UV-Vis detection. The supported catalyst was recovered by filtration and centrifugation and then washed with DCM (20 mL). Residual solvent was removed under reduced pressure without heating.

The immobilization was confirmed by UV-Vis analysis, and the exact amount of immobilized catalyst was determined by UV-Vis. Furthermore, the mass of the obtained solid was equal to the sum of the mass of rGO and the mass of the introduced catalyst.

The UV-visible analysis was performed using an UV-Vis spectrum scanning method (200-500 nm). A calibration line ($A = f c$) has been drawn by measuring the absorbances (A) of diverse solutions with different concentrations (c) at the λ_{max} (345 nm).



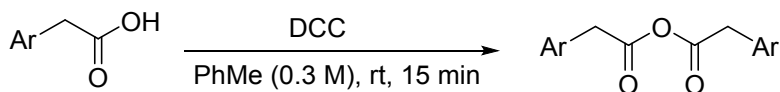
UV-Vis Calibration curve $\text{Abs} = f [\text{pyr-hyperBTM}]$



UV-Visible spectra of **pyr-hyperBTM** solution before and after immobilization

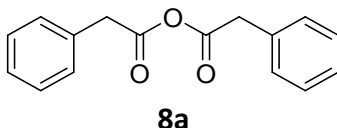
4. Preparation of substrates.

4.1 General Procedure: Synthesis of Anhydrides



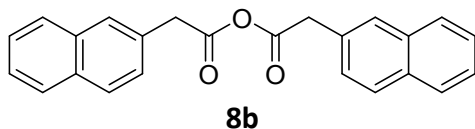
To a solution of the appropriate carboxylic acid (1 eq) in toluene (0.3 M), DCC (0.50-0.55 eq) was added and the solution stirred at room temperature for 15 min. The suspension was filtered through Celite®, which was washed with extra toluene, and the filtrate was concentrated *in vacuo* to give the crude product, which was used without further purification.

2-Phenylacetic anhydride **8a**



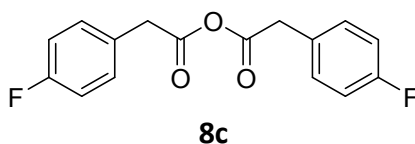
Following General Procedure 4.1, *N,N'*-Dicyclohexylcarbodiimide (DCC) (1.03 g, 5 mmol, 0.5 equiv.) was added to a solution of phenylacetic acid (1.36g, 10 mmol, 1 equiv.) in toluene (0.3 M, 34 mL) at rt to give product **8a** as a white solid (1.26 g, >99%). ¹H NMR (300 MHz, CDCl₃) δ = 7.36-7.29 (m, 6H), 7.24-7.19 (m, 4H), 3.72 (s, 4H) ppm. The spectral data match those previously reported.⁵

2-(Naphthalen-2-yl)acetic anhydride **8b**



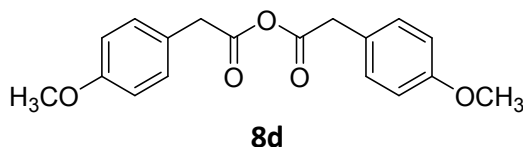
Following General Procedure 4.1, *N,N'*-Dicyclohexylcarbodiimide (DCC) (2.06 g, 10 mmol, 0.5 equiv.) was added to a solution of 2-naphthylacetic acid (3.72 g, 20 mmol, 1 equiv.) in toluene (0.3 M, 67 mL) at rt to give product **8b** as a white solid (1.4 g, 40%). ¹H NMR (300 MHz, CDCl₃) δ = 7.82-7.79 (m, 2H), 7.72-7.70 (m, 4H), 7.63 (s, 2H), 7.50-7.46 (m, 4H), 7.27 (dd, *J* = 8.4, 1.8 Hz, 2H), 3.88 (s, 4H) ppm. The spectral data match those previously reported.⁵

2-(4-Fluorophenyl)acetic anhydride **8c**



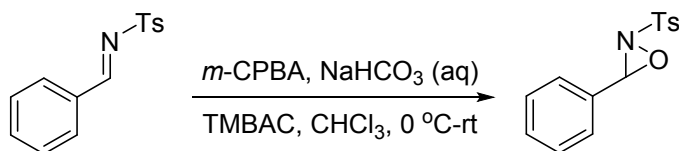
Following General Procedure 4.1, *N,N'*-Dicyclohexylcarbodiimide (DCC) (1.03g, 5 mmol, 0.5 equiv.) was added to a solution of 4-fluorophenylacetic acid (1.54 g, 10 mmol, 1 equiv.) in toluene (0.3 M, 34 mL) at rt to give product **8c** as a white solid (1.50 g, >99%). ¹H NMR (300 MHz, CDCl₃) δ = 7.21-7.14 (m, 4H), 7.05-6.97 (m, 4H), 3.70 (s, 4H) ppm; ¹⁹F{¹H} NMR (235 MHz, CDCl₃): δ = -114.56 ppm. The spectral data match those previously reported.⁵

2-(4-Methoxyphenyl)acetic anhydride **8d**



Following General Procedure 4.1, *N,N'*-Dicyclohexylcarbodiimide (DCC) (1.03 g, 5 mmol, 0.5 equiv.) was added to a solution of 4-methoxyphenylacetic acid (1.66 g, 10 mmol, 1 equiv.) in toluene (0.3 M, 34 mL) at rt to give product **8d** as a white solid (1.57 g, >99%). ¹H NMR (300 MHz, CDCl₃) δ = 7.12 (d, *J* = 8.7 Hz, 4H), 6.85 (d, *J* = 8.7 Hz, 4H), 3.80 (s, 6H), 3.66 (s, 4H) ppm. The spectral data match those previously reported.⁵

4.2 General Procedure 2: Synthesis of *N*-Sulfonyl Oxaziridine

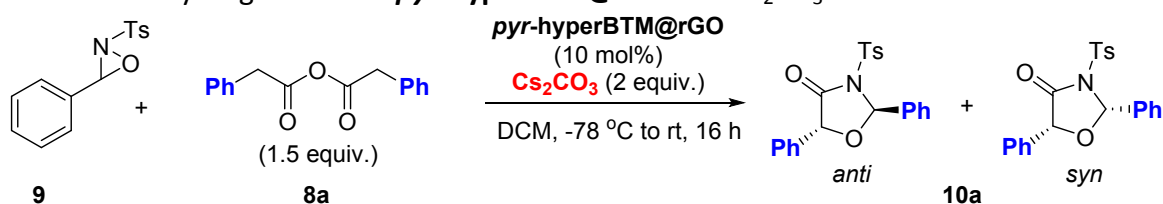


To a solution of saturated aqueous sodium bicarbonate (10 mL, 1 M) and benzyltrimethylammonium chloride (102 mg, 0.55 mmol, 0.11 eq), the appropriate *N*-sulfonyl imine (1.3 g, 5 mmol, 1 eq) was added as a solution in CHCl₃ (5 mL, 1 M). The mixture was cooled to 0 °C and a solution of *m*-CPBA (1.28 g, 5.5 mmol, 1.1 eq) in CHCl₃ (11 mL, 0.5 M) was added dropwise at 0 °C and stirred for 1 h. The organic layer was separated, washed with water, 10% sodium sulphite solution, water and brine before drying (MgSO₄) and concentrating in vacuo, keeping the bath temperature below 40 °C. The crude oxaziridine was recrystallised from a small amount of ethyl acetate and petrol ether without heating (60% yield). ¹H NMR (250 MHz, CDCl₃) δ = 7.93 (d, *J* = 8.5 Hz, 2H), 7.46-7.40 (m, 7H), 5.45 (s, 1H), 2.50 (s, 3H) ppm. The spectral data match those previously reported.⁵

5. Recycling tests for screening reaction conditions

The appropriate oxaziridine **9** (1 equiv) and **pyr-hyperBTM@rGO** (10 mol%) were added to a solution of the 2-phenylacetic anhydride **8a** (1.5 equiv) and base (2 equiv) in CH₂Cl₂ (0.2 M) at -78 °C. The reaction mixture was stirred at -78 °C then warmed slowly to room temperature over 24h. The **pyr-hyperBTM@rGO** was filtered off, washed with the appropriate solvent (3 x 3 mL). The recovered catalyst was dried under vacuum for 2-3 h and the dried catalyst was directly used in the next recycling run. The combined organic phases were dried over MgSO₄, filtered and concentrated under reduced pressure. The crude residue was purified by column chromatography on silica gel (petroleum ether:EtOAc=10:1) to afford the desired product **10a**.

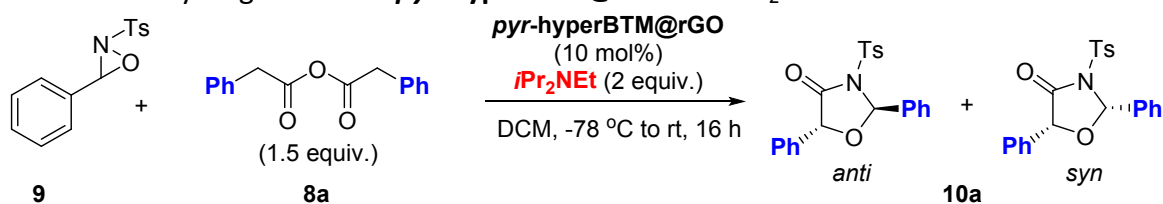
Table S1. Recycling tests with **pyr-hyperBTM@rGO** and Cs₂CO₃ as a base.



	Work-up washing	Yield ^a	d.r. ^a _{anti/syn} ^b	ee ^a _{anti/syn} ^b
1st attempt	DCM			
	1 st run	64	69 / 31	87 / 81
Without additional base	2 nd run	20	88 / 12	87 / -6
With 2 eq Cs ₂ CO ₃	3 rd run	< 5	-	-
2nd attempt	H₂O + acetone			
	1 st run	81	67 / 33	85 / 90
	2 nd run	40	74 / 26	67 / 92
	3 rd run ^c	23	67 / 33	54 / 75

^aisolated yield of **anti-10a** + **syn-10a**. ^bdetermined by HPLC analysis. ^cbefore the 3rd run, **pyr-hyperBTM@rGO** was stirred at 60 °C for 30 min in 2 mL toluene, toluene was removed and the catalyst dried under vacuum))

Table S2. Recycling tests with **pyr-hyperBTM@rGO** and *i*Pr₂NEt as a base.



	Work-up washing	Yield ^a	d.r. ^a _{anti/syn} ^b	ee ^a _{anti/syn} ^b
1st attempt	acetone			
	1 st run	76	64 / 36	92 / 93
	2 nd run	47	67 / 33	73 / 89
	3 rd run	55	60 / 40	90 / 92
2nd attempt	DCM			
	1 st run	75	59 / 41	88 / 92
	2 nd run	65	56 / 44	89 / 88
	3 rd run	60	50 / 50	93 / 85

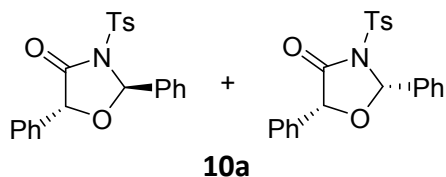
^aisolated yield of **anti-10a** + **syn-10a**. ^bdetermined by HPLC analysis

6. General procedure for the asymmetric reaction recycling

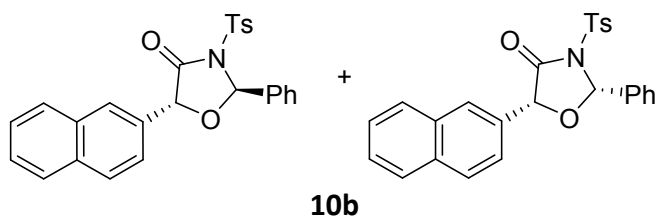
General procedure for the asymmetric organocatalytic formation of oxazolidin-4-ones:

The appropriate oxaziridine **9** (1 equiv) and **pyr-hyperBTM@rGO** (10 mol%) were added to a solution of the appropriate homoanhydride (1.5 equiv) and cesium carbonate (2 equiv) in CH₂Cl₂ (0.2 M) at -78 °C. The reaction mixture was stirred at -78 °C then warmed slowly to room temperature over 24h. The **pyr-hyperBTM@rGO** was filtered off, washed with CH₂Cl₂ (3 x 3 mL). The recovered catalyst was dried under vacuum for 2-3 h and the dried catalyst was directly used in the next recycling run. The combined organic phases were dried over MgSO₄, filtered and concentrated under reduced pressure. The crude residue was purified by column chromatography on silica gel (petroleum ether: EtOAc = 10:1) to afford the desired products **10a-10d**.

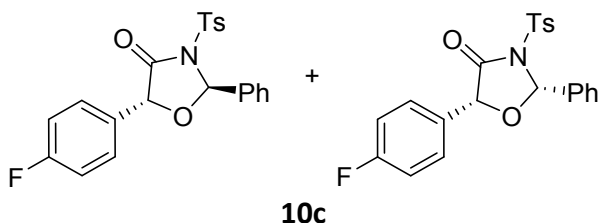
7. Characterization data of products 10a-10d.



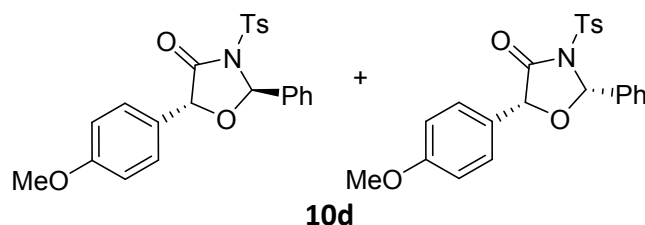
(2R,5R)-2,5-Diphenyl-3-tosyloxazolidin-4-one (*anti*-10a) and (2S,5R)-2,5-diphenyl-3-tosyloxazolidin-4-one (*syn*-10a): White solid. ^1H NMR (300 MHz, CDCl_3): δ = 7.59-7.34 (m, 12H), 7.20-7.15 (m, 2H), 6.73 (s, 1H, *anti*-10a), 6.59 (s, 1H, *syn*-10a), 5.44 (s, 1H, *anti*-10a), 5.40 (s, 1H, *syn*-10a), 2.40 (s, 3H, *anti*-10a), 2.38 (s, 3H, *syn*-10a), ppm; $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3): δ = 168.8 (*anti*-10a), 168.6 (*syn*-10a), 145.7 (*anti*-10a), 145.6 (*syn*-10a), 136.6 (*anti*-10a), 136.4 (*syn*-10a), 135.1 (*syn*-10a), 134.8 (*anti*-10a), 134.4 (*anti*-10a), 134.0 (*syn*-10a), 130.4 (*syn*-10a), 130.2 (*anti*-10a), 129.5, 129.2, 128.9, 128.8, 128.63, 128.56, 128.4, 128.3, 128.2, 127.5, 126.5 (*anti*-10a), 126.4 (*syn*-10a), 91.3 (*anti*-10a), 90.9 (*syn*-10a), 79.0 (*syn*-10a), 78.7 (*anti*-10a), 21.8 ppm. The spectral data match those previously reported.⁵



(2R,5R)-5-(Naphthalen-2-yl)-2-phenyl-3-tosyloxazolidin-4-one (*anti*-10b) and (2S,5R)-5-(naphthalen-2-yl)-2-phenyl-3-tosyloxazolidin-4-one (*syn*-10b): White solid. ^1H NMR (300 MHz, CDCl_3): δ = 7.87-7.75 (m, 4H), 7.60-7.39 (m, 10H), 7.19-7.14 (m, 2H), 6.79 (s, 1H, *anti*-10b), 6.66 (s, 1H, *syn*-10b), 5.61 (s, 1H, *anti*-10b), 5.57 (s, 1H, *syn*-10b), 2.38 (s, 3H, *anti*-10b), 2.37 (s, 3H, *syn*-10b) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3): δ = 168.8 (*anti*-10b), 168.6 (*syn*-10b), 145.8, 145.6, 136.7 (*anti*-10b), 136.5 (*syn*-10b), 135.1 (*syn*-10b), 134.8 (*anti*-10b), 133.6, 133.5, 133.1 (*anti*-10b), 133.0 (*syn*-10b), 131.7 (*syn*-10b), 131.5 (*anti*-10b), 130.5, 130.3, 129.6, 128.9, 128.8, 128.6, 128.5, 128.4, 128.3, 128.2, 127.8, 127.5, 126.8, 126.6, 126.5, 126.1, 125.8, 123.7 (*anti*-10b), 123.6 (*syn*-10b), 91.4 (*syn*-10b), 91.0 (*anti*-10b), 79.1, 78.8 (*anti*-10b), 21.8 ppm. The spectral data match those previously reported.⁵



(2*R*,5*R*)-5-(4-Fluorophenyl)-2-phenyl-3-tosyloxazolidin-4-one (*anti*-10c) and (2*S*,5*R*)-5-(4-fluorophenyl)-2-phenyl-3-tosyloxazolidin-4-one (*syn*-10c): White solid. ^1H NMR (300 MHz, CDCl_3): δ = 7.55-7.33 (m, 9H), 7.20-7.15 (m, 2H), 7.09-7.01 (m, 2H), 6.71 (s, 1H, *anti*-10c), 6.58 (s, 1H, *syn*-10c), 5.41 (s, 1H, *anti*-10c), 5.37 (s, 1H, *syn*-10c), 2.41 (s, 3H, *anti*-10c), 2.38 (s, 3H, *syn*-10c) ppm; $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3): δ = 168.6 (*anti*-10c), 168.5 (*syn*-10c), 164.9 (*anti*-10c), 164.7 (*syn*-10c), 161.6, 161.5, 145.9 (*anti*-10c), 145.7 (*syn*-10c), 136.5 (*anti*-10c), 136.3 (*syn*-10c), 135.0 (*syn*-10c), 134.7 (*anti*-10c), 130.5 (*syn*-10c), 130.3 (*anti*-10c), 129.9, 129.6, 128.8, 128.6, 128.5, 128.3, 128.24, 128.19, 127.4, 116.1, 115.8, 115.5, 91.2 (*anti*-10c), 90.9 (*syn*-10c), 78.4 (*syn*-10c), 78.1 (*anti*-10c), 21.8 ppm; $^{19}\text{F}\{^1\text{H}\}$ NMR (235 MHz, CDCl_3): δ = -112.2 (*anti*-10c), -112.8 (*syn*-10c). The spectral data match those previously reported.⁵

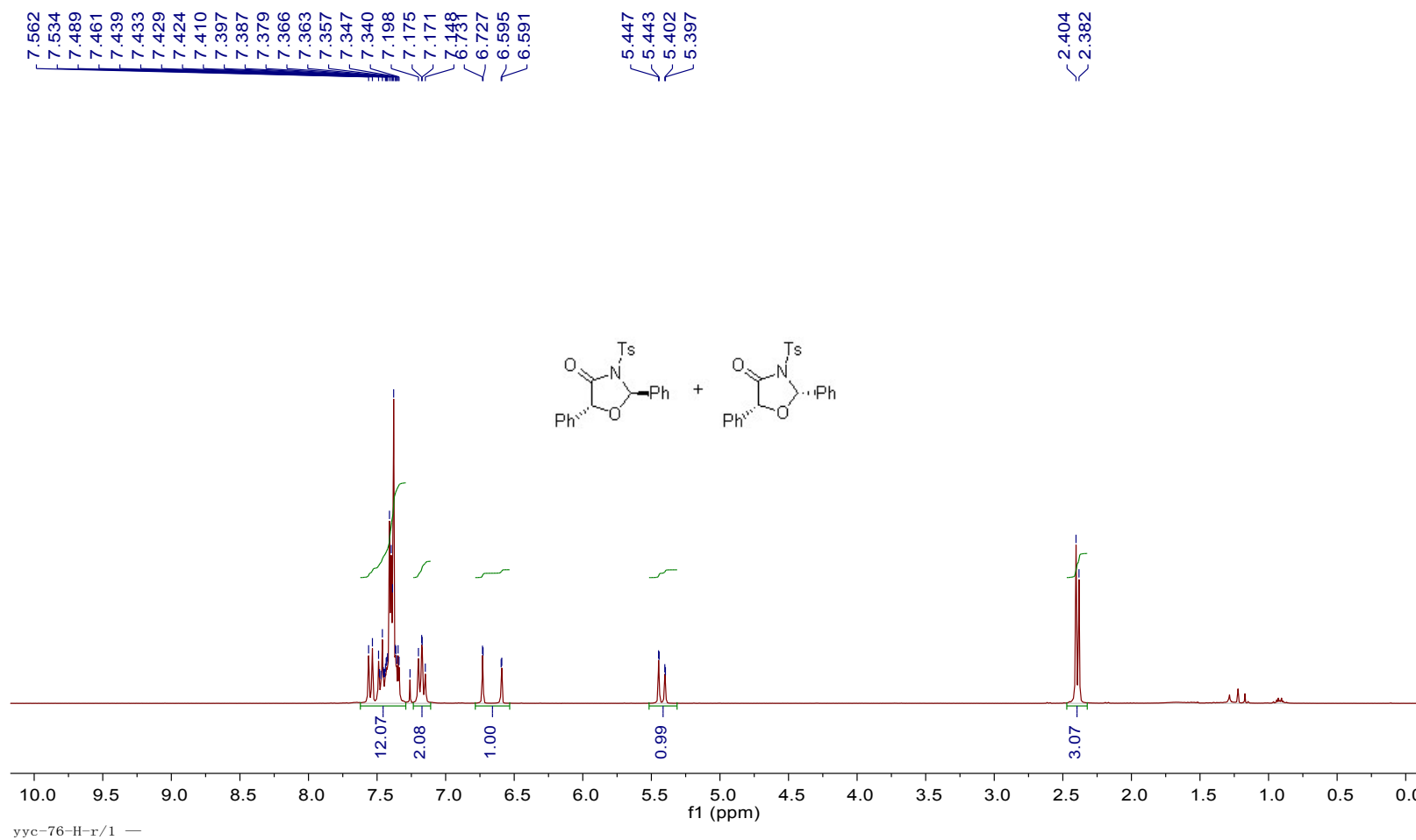


(2*R*,5*R*)-5-(4-Methoxyphenyl)-2-phenyl-3-tosyloxazolidin-4-one and (*anti*-10d) (2*S*,5*R*)-5-(4-methoxyphenyl)-2-phenyl-3-tosyloxazolidin-4-one (*syn*-10d): Colourless oil. ^1H NMR (300 MHz, CDCl_3): δ = 7.47-7.26 (m, 7H), 7.22-7.14 (m, 2H), 7.11-7.05 (m, 2H), 6.81-6.76 (m, 2H), 6.58 (d, J = 1.2 Hz, 1H, *anti*-10d), 6.45 (d, J = 1.2 Hz, 1H, *syn*-10d), 5.27 (s, 1H, *anti*-10d), 5.24 (s, 1H, *syn*-10d), 3.70 (s, 3H, *anti*-10d), 3.68 (s, 3H, *syn*-10d), 2.31 (s, 3H, *anti*-10d), 2.29 (s, 3H, *syn*-10d), ppm; $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3): δ = 169.1 (*anti*-10d), 169.0 (*syn*-10d), 160.4, 160.2, 145.7 (*anti*-10d), 145.6 (*syn*-10d), 136.7 (*anti*-10d), 136.4 (*syn*-10d), 135.2 (*syn*-10d), 134.9 (*anti*-10d), 130.4 (*syn*-10d), 130.2 (*anti*-10d), 129.6, 128.8, 128.6, 128.43, 128.39, 128.27, 128.1, 127.5, 126.6 (*anti*-10d), 126.2 (*syn*-10d), 114.4 (*anti*-10d), 114.2 (*syn*-10d), 91.1 (*anti*-10d), 90.8 (*syn*-10d), 79.0 (*syn*-10d), 78.6 (*anti*-10d), 55.4, 21.8 ppm. The spectral data match those previously reported.⁵

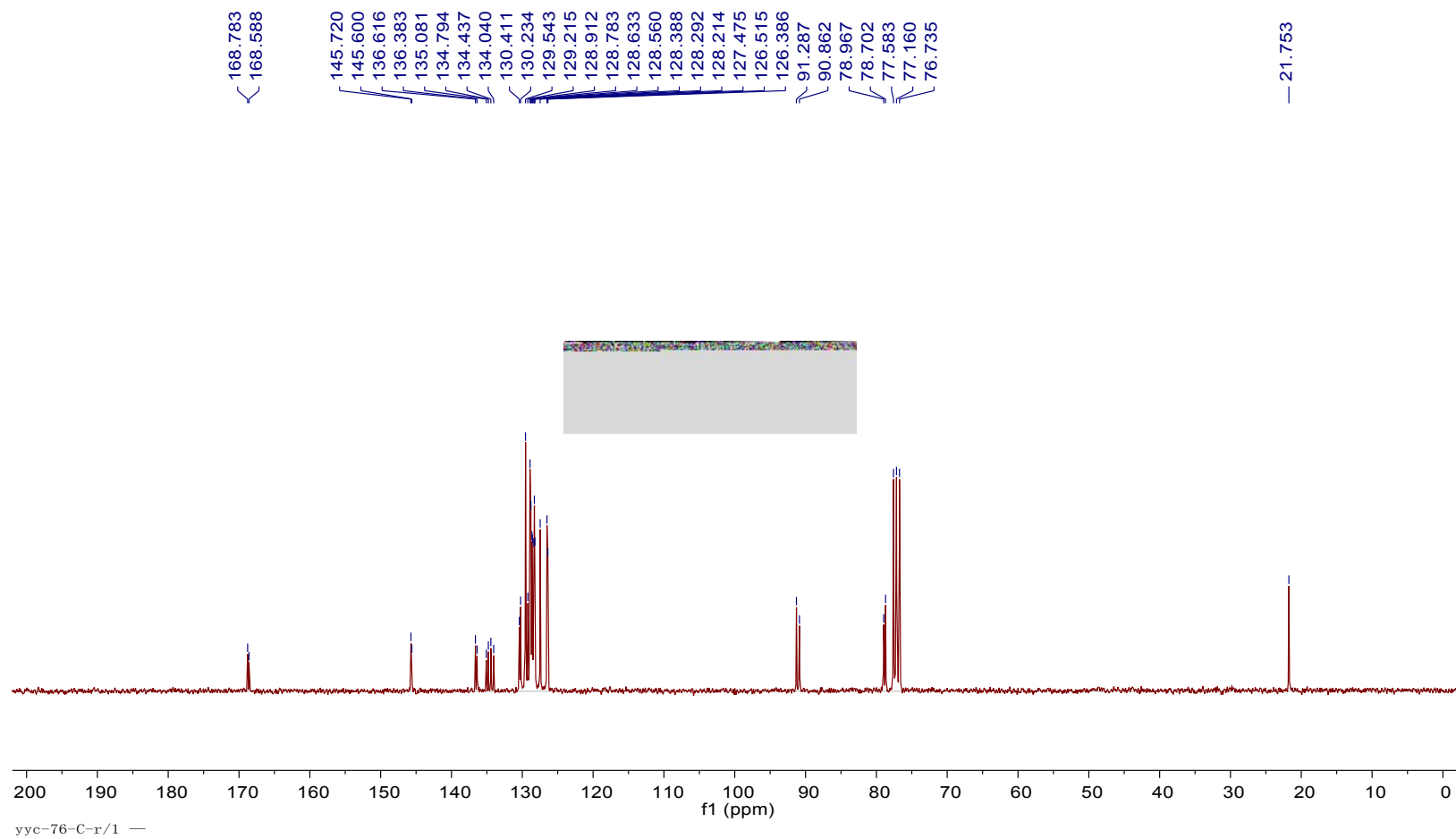
8. References.

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- [2] C. Joannesse, C. P. Johnston, C. Concellon, C. Simal, D. Philp, A. D. Smith, *Angew. Chem. Int. Ed.*, **2009**, *48*, 8914.
- [3] D. S. B. Daniels, S. R. Smith, T. Lebl, P. Shapland, A. D. Smith, *Synthesis* **2015**, *47*, 34.
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- [5] S. R. Smith, C. Fallan, J. E. Taylor, R. McLennan, D. S. B. Daniels, L. C. Morrill, A. M. Z. Slawin and A. D. Smith, *Chem. Eur. J.*, **2015**, *21*, 10530.

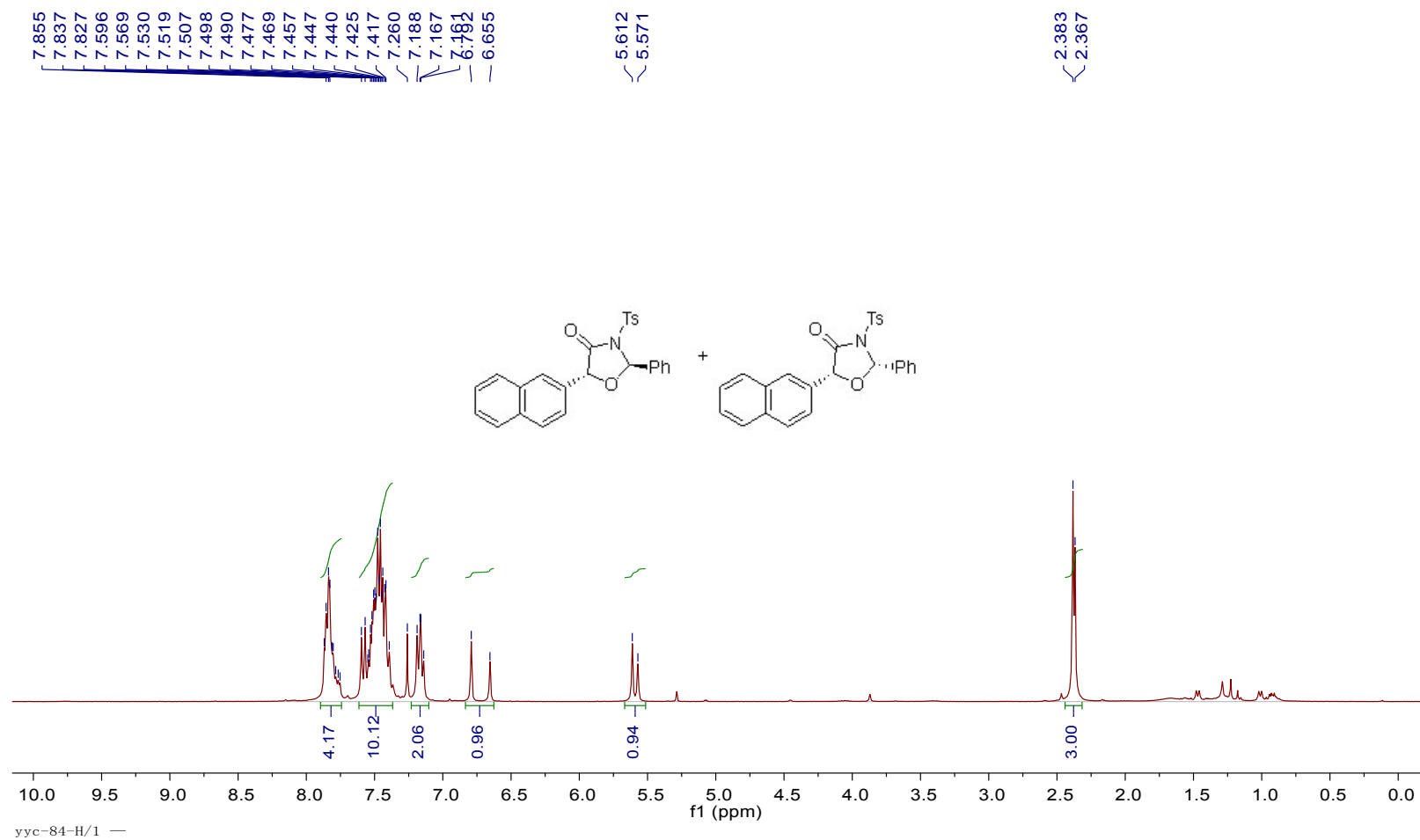
9. NMR spectra of products 10a-10d and *pyr*-hyperBTM



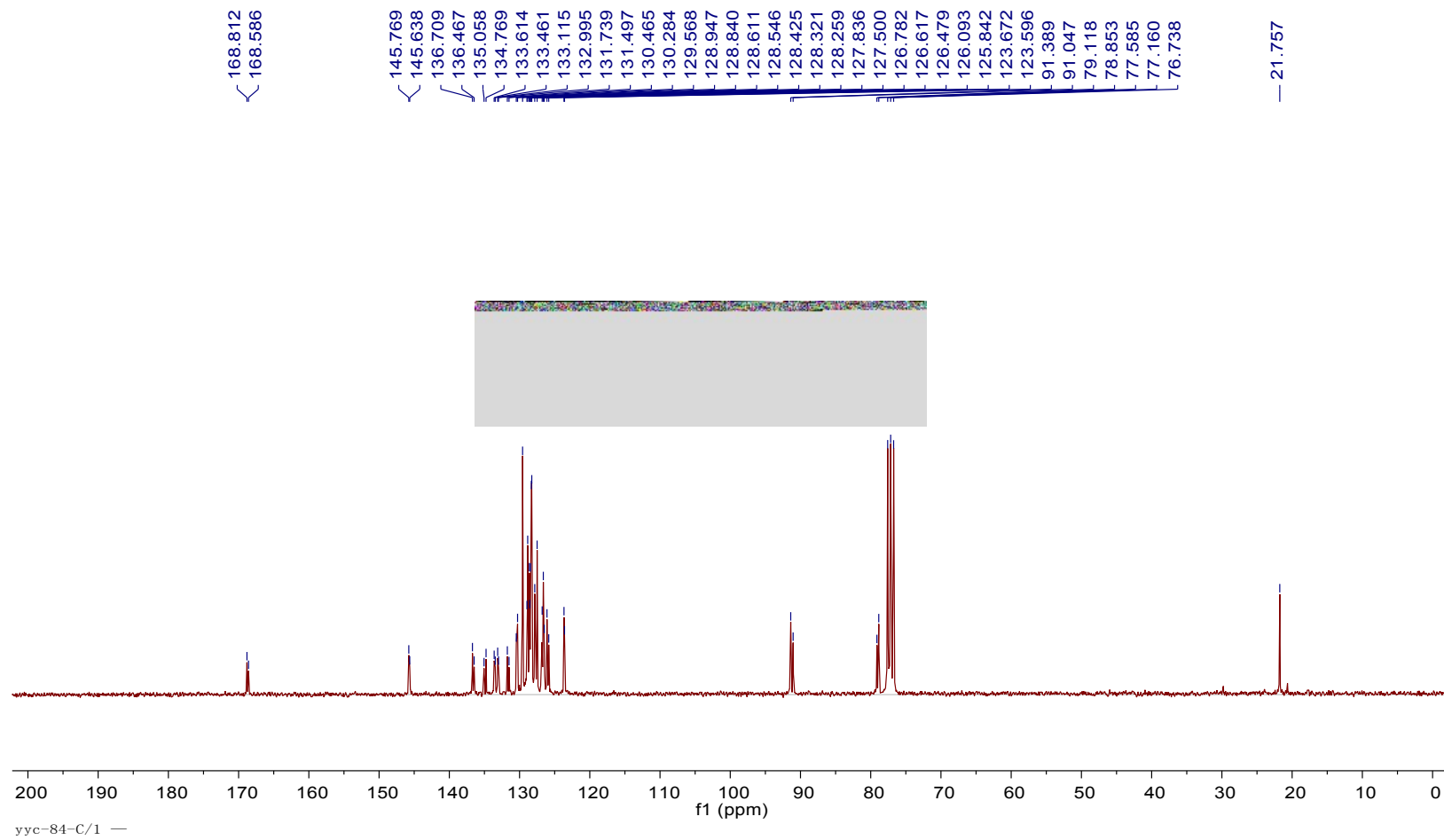
^1H NMR spectrum of **10a**



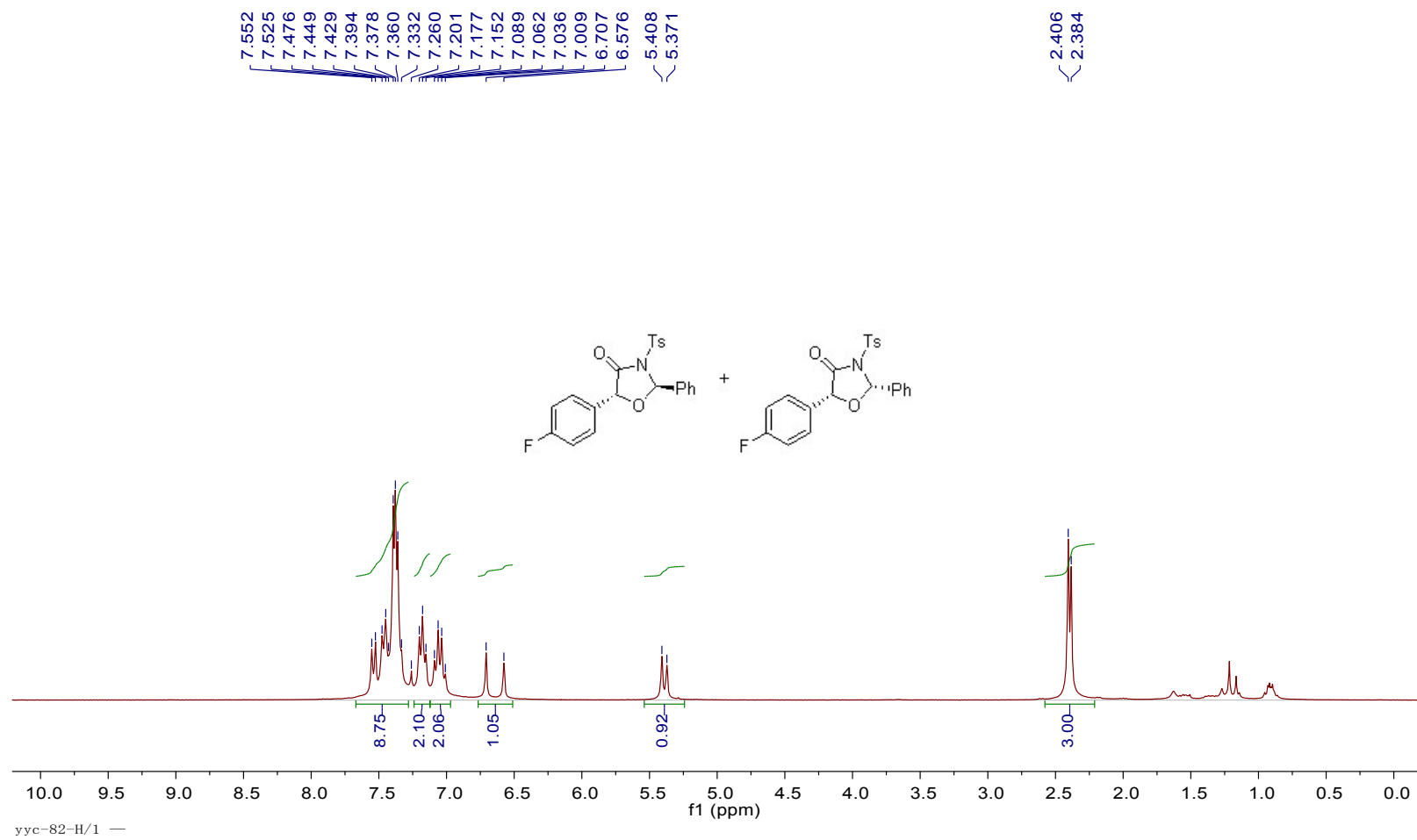
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **10a**



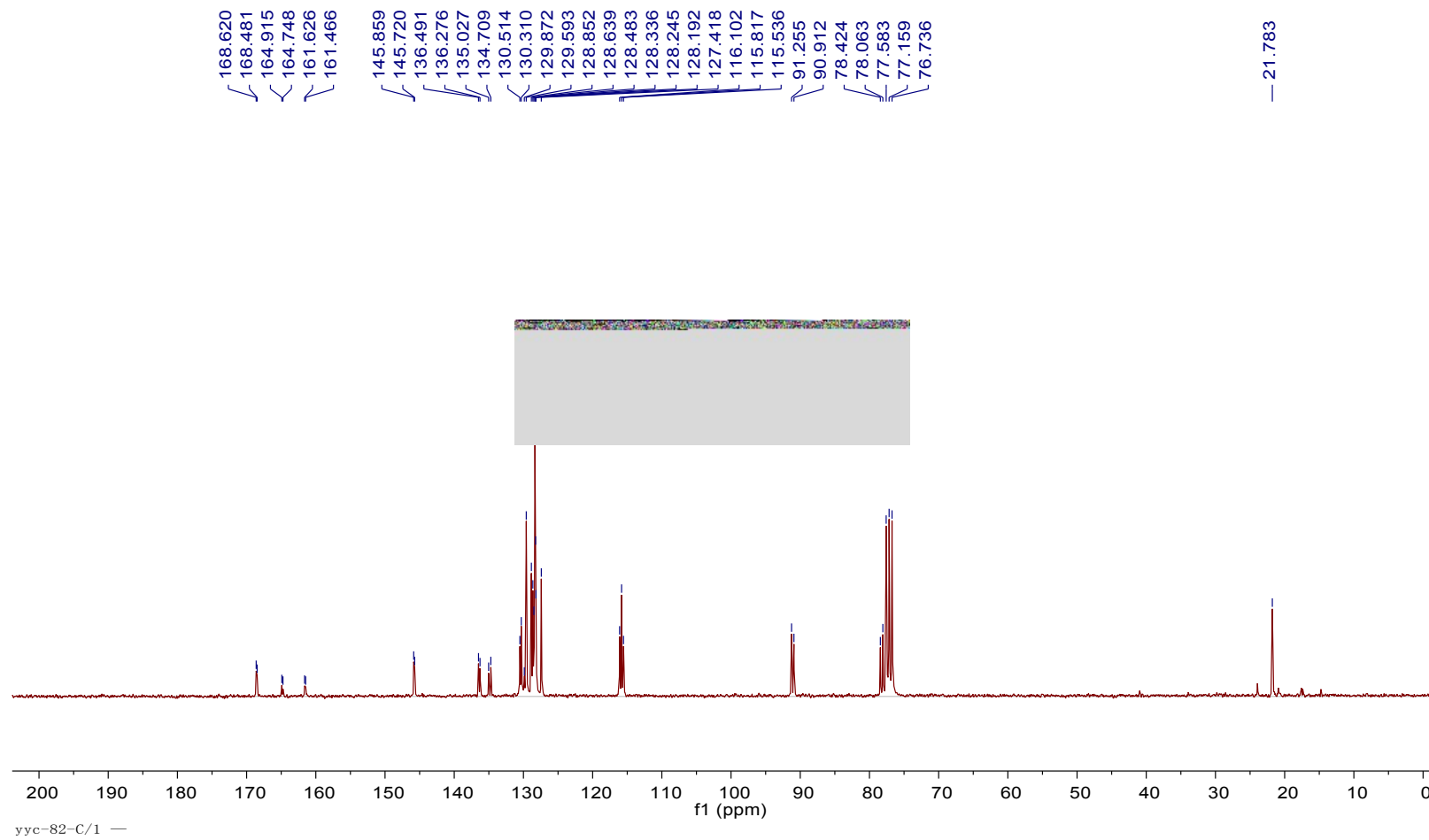
¹H NMR spectrum of **10b**

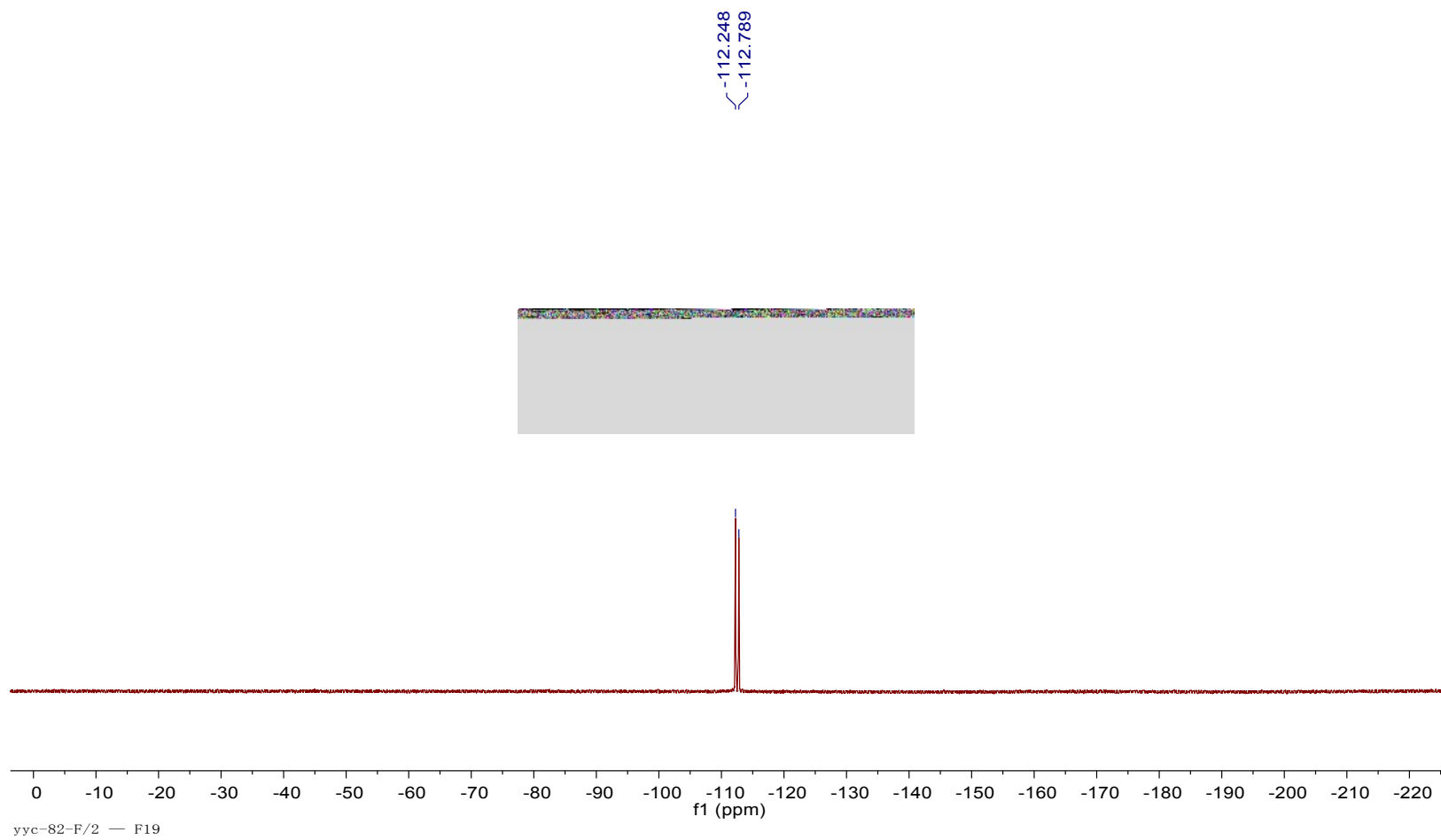


$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **10b**

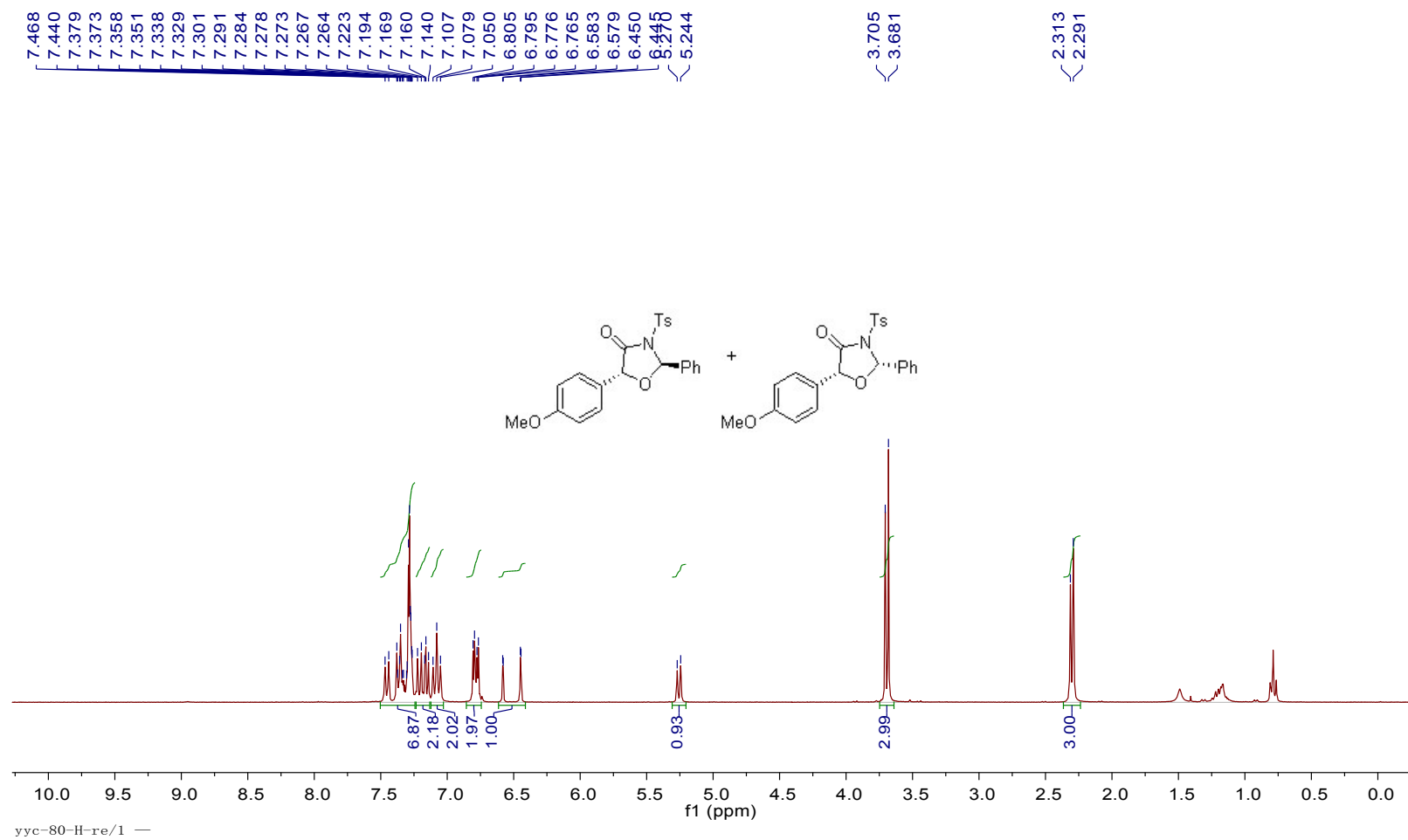


¹H NMR spectrum of **10c**

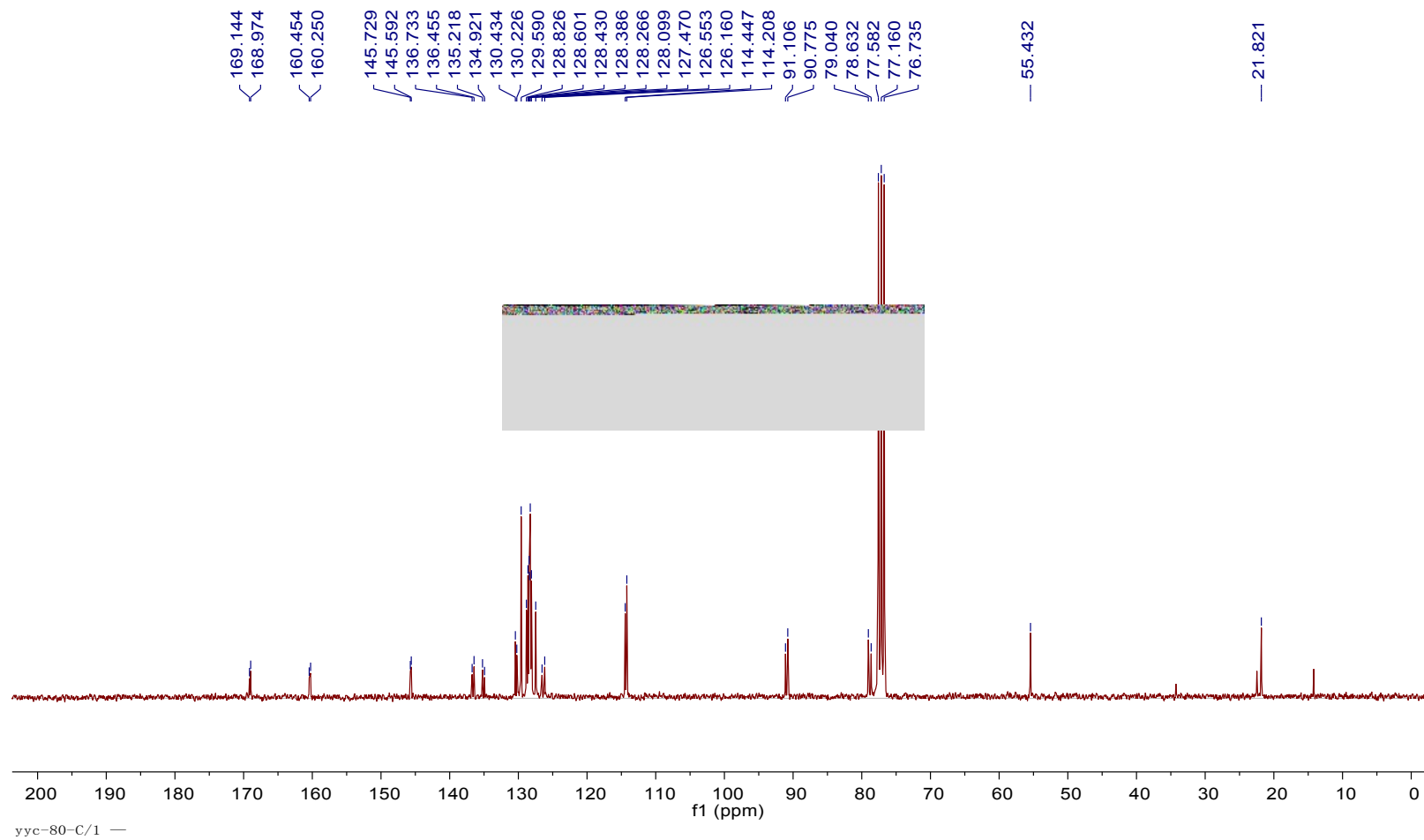




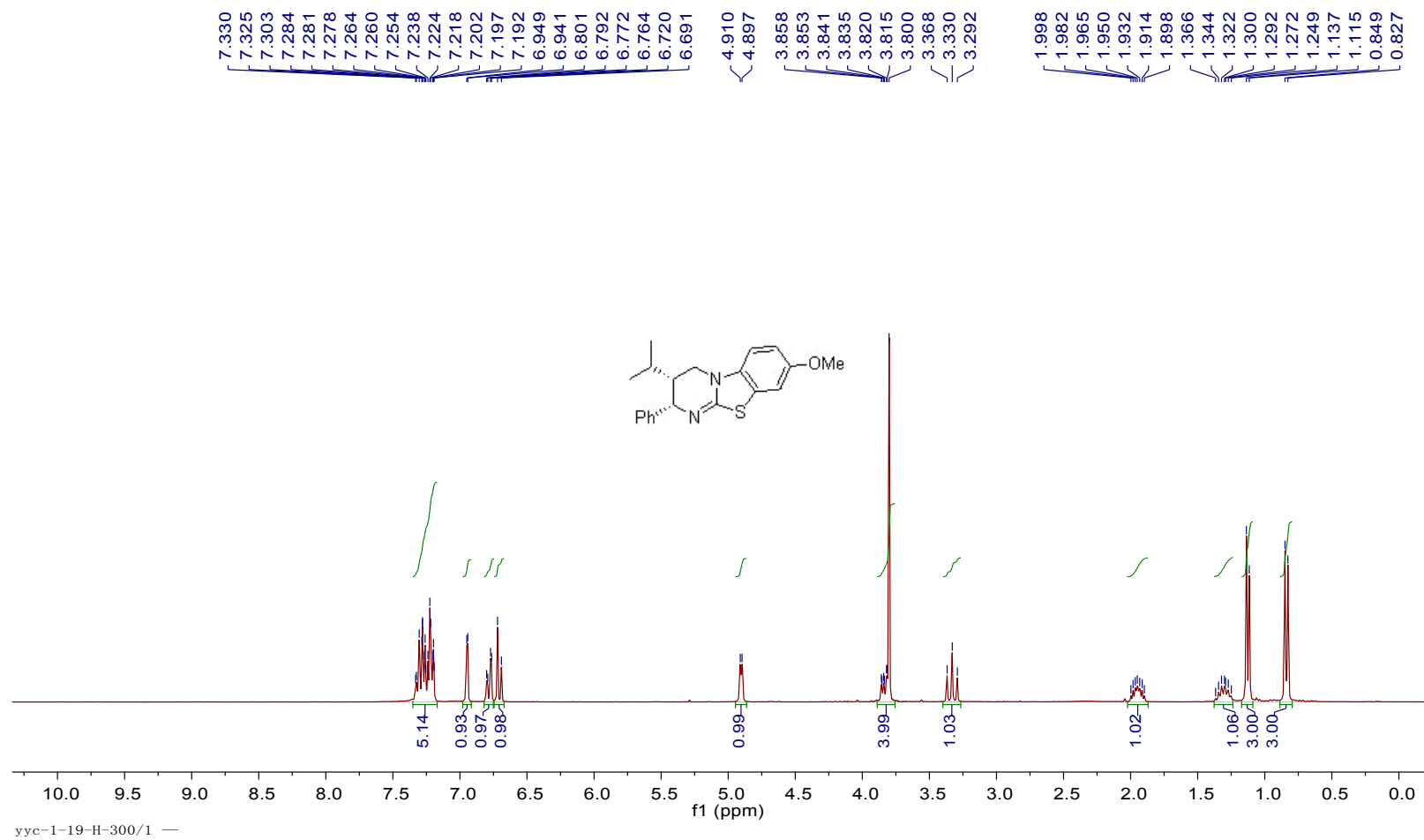
$^{19}\text{F}\{^1\text{H}\}$ NMR spectrum of **10c**



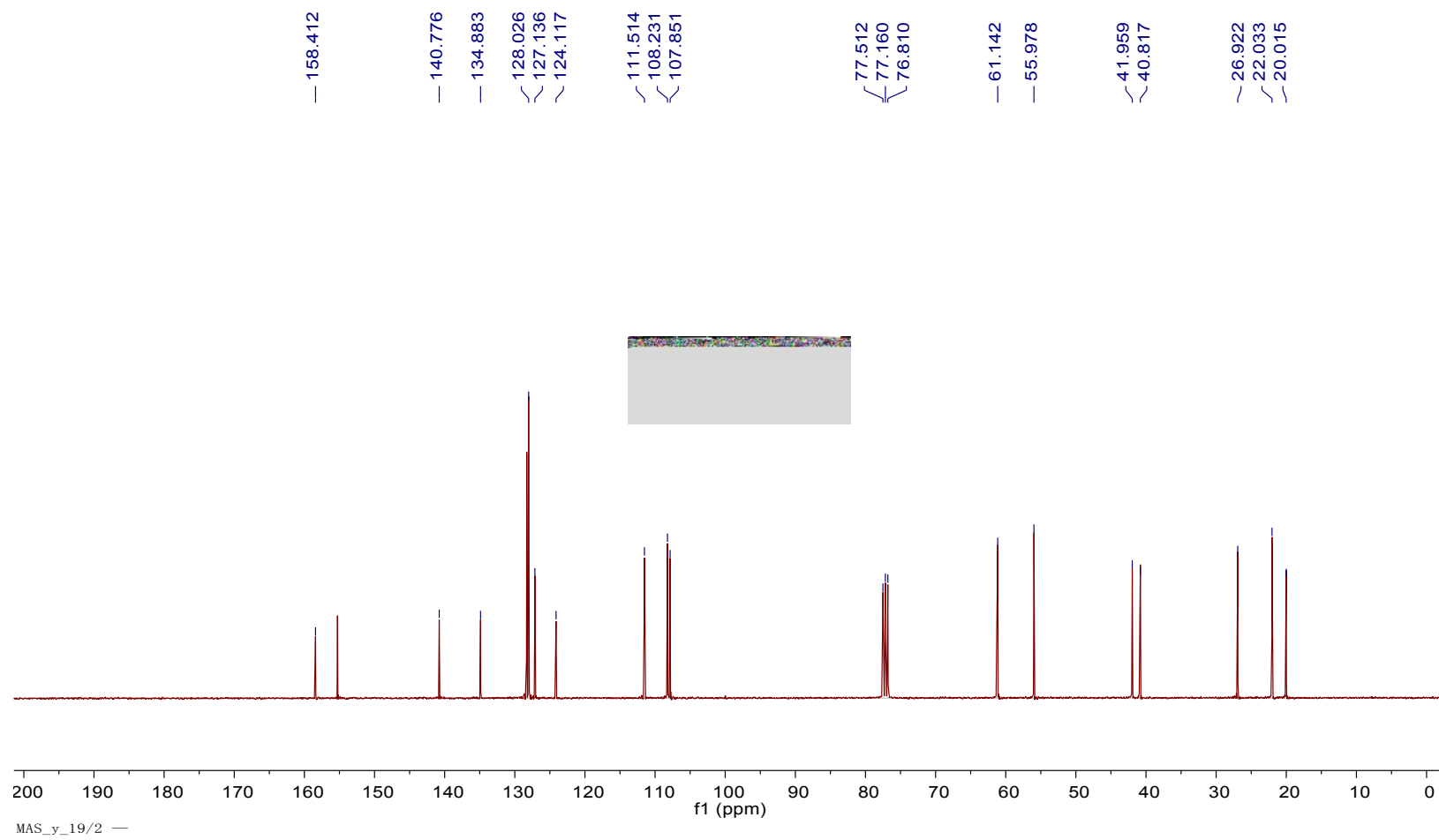
¹H NMR spectrum of **10d**



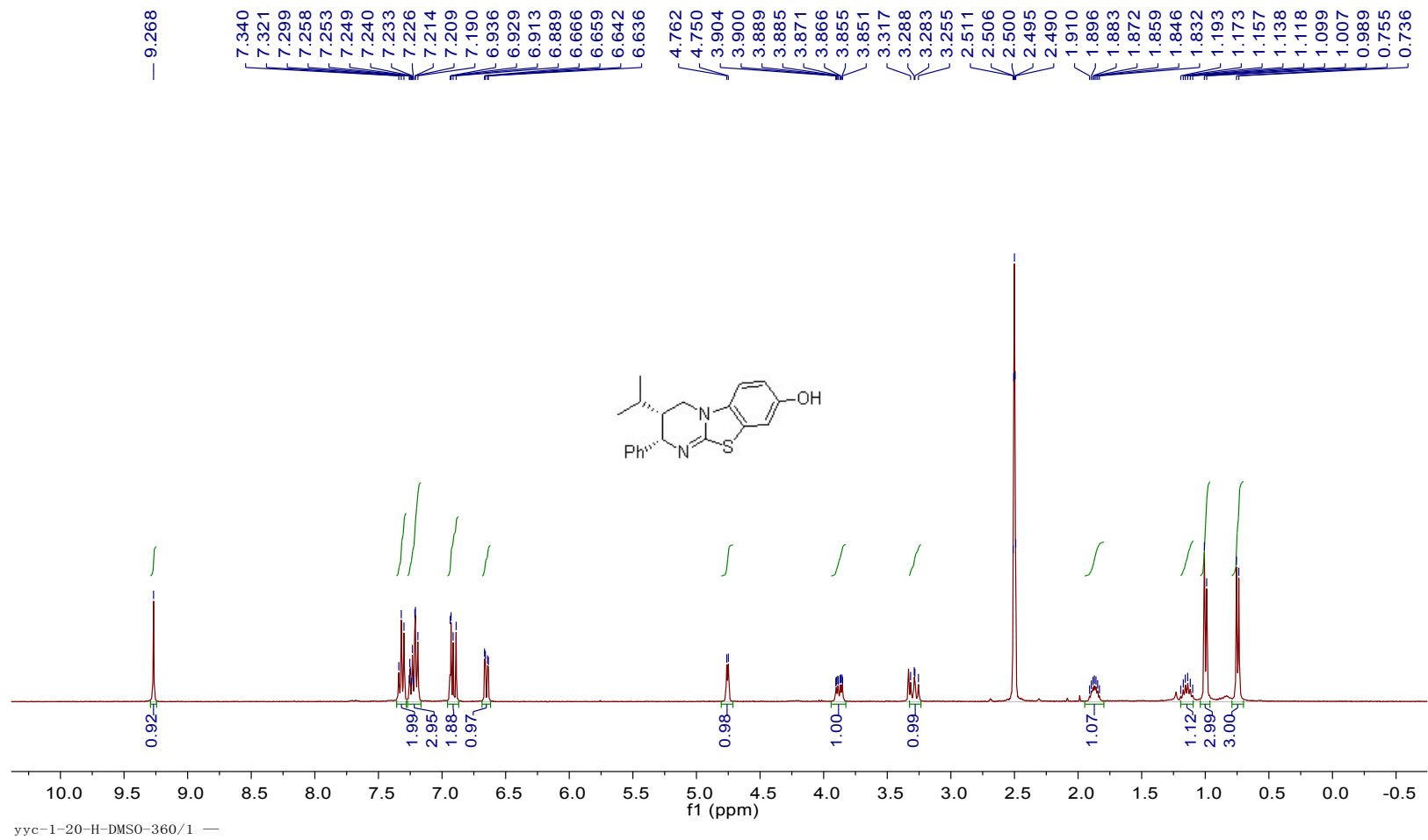
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **10d**

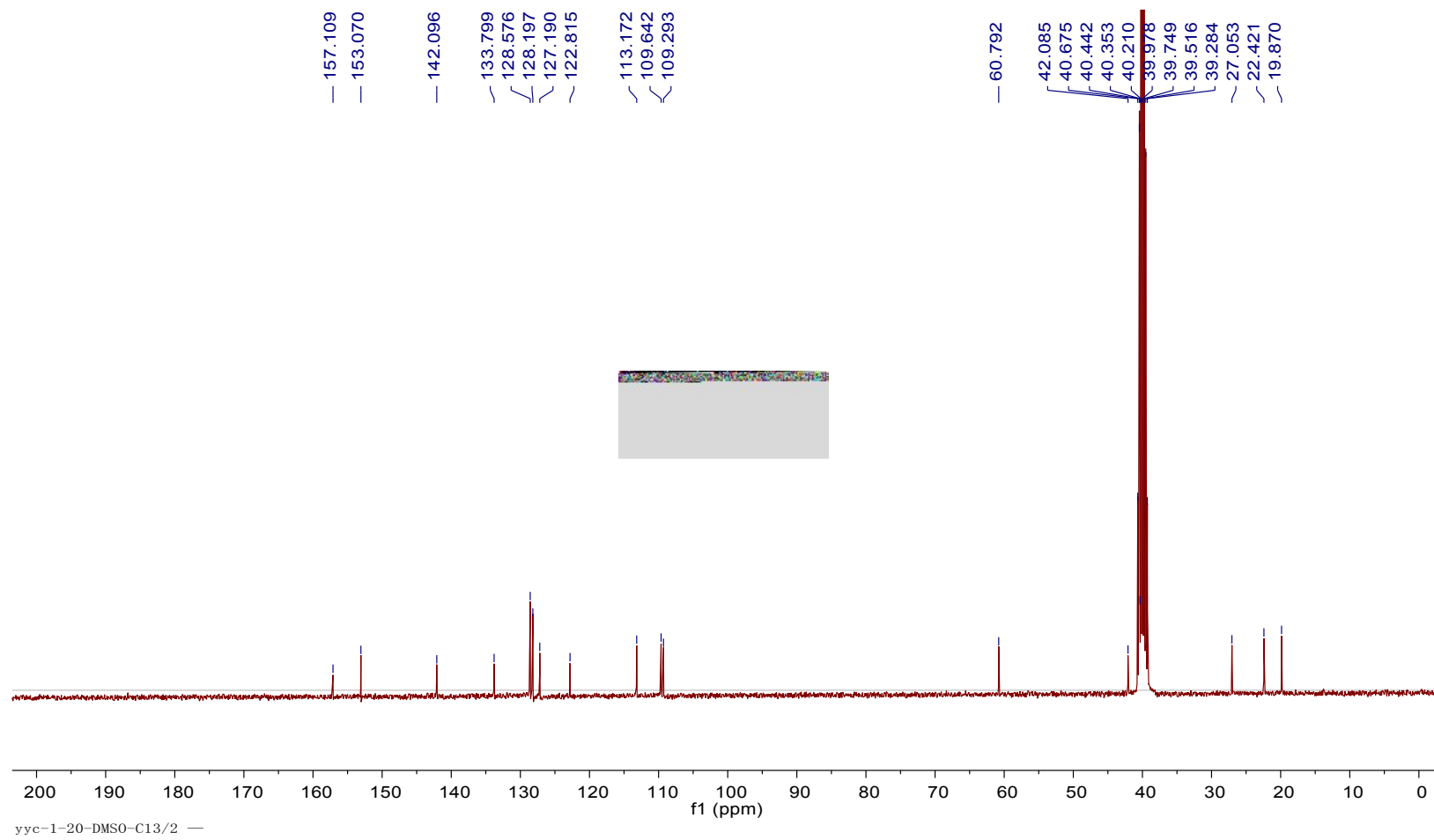


¹H NMR spectrum of (2*S*,3*R*)-3

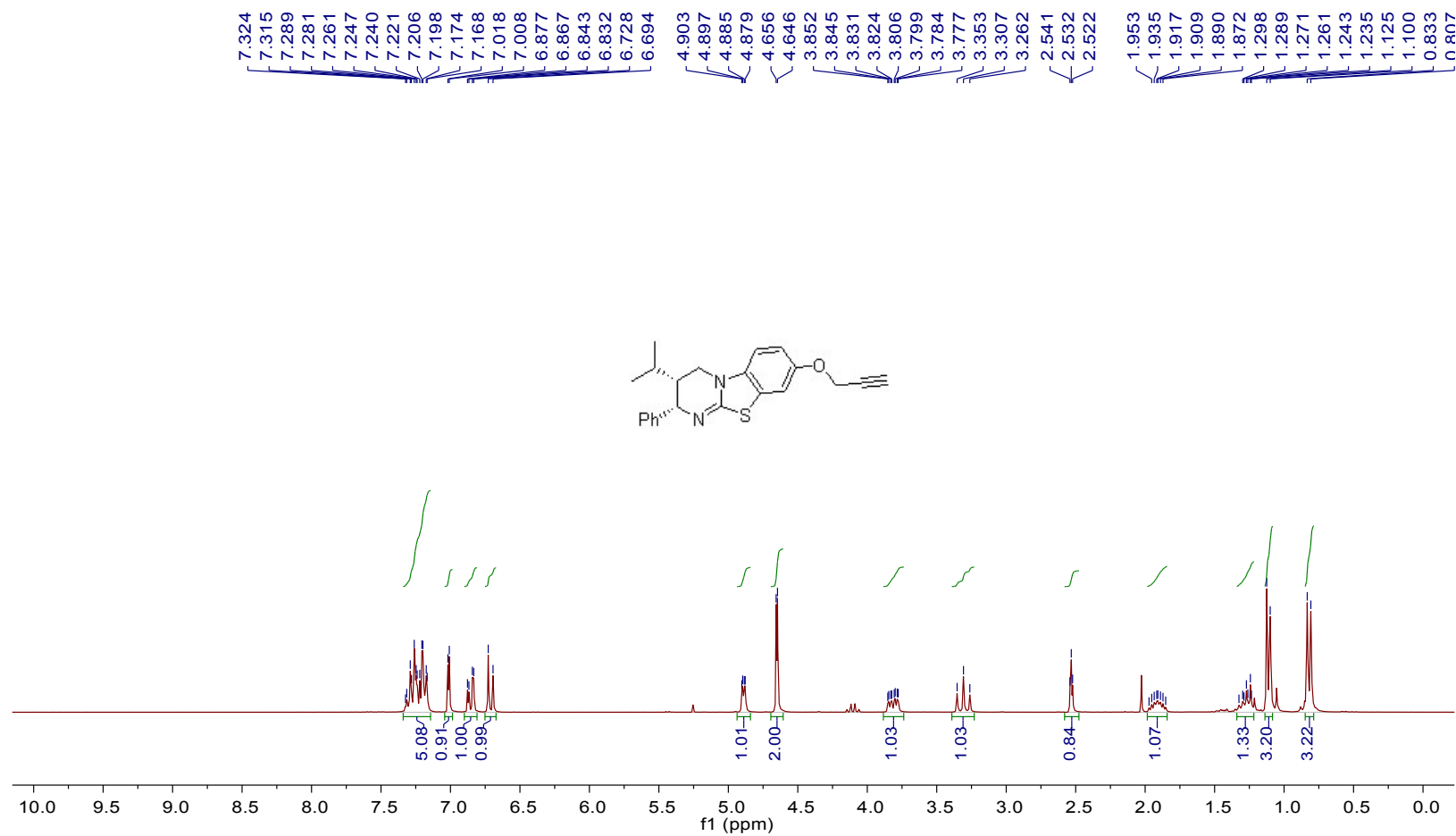


$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of (2S,3R)-3

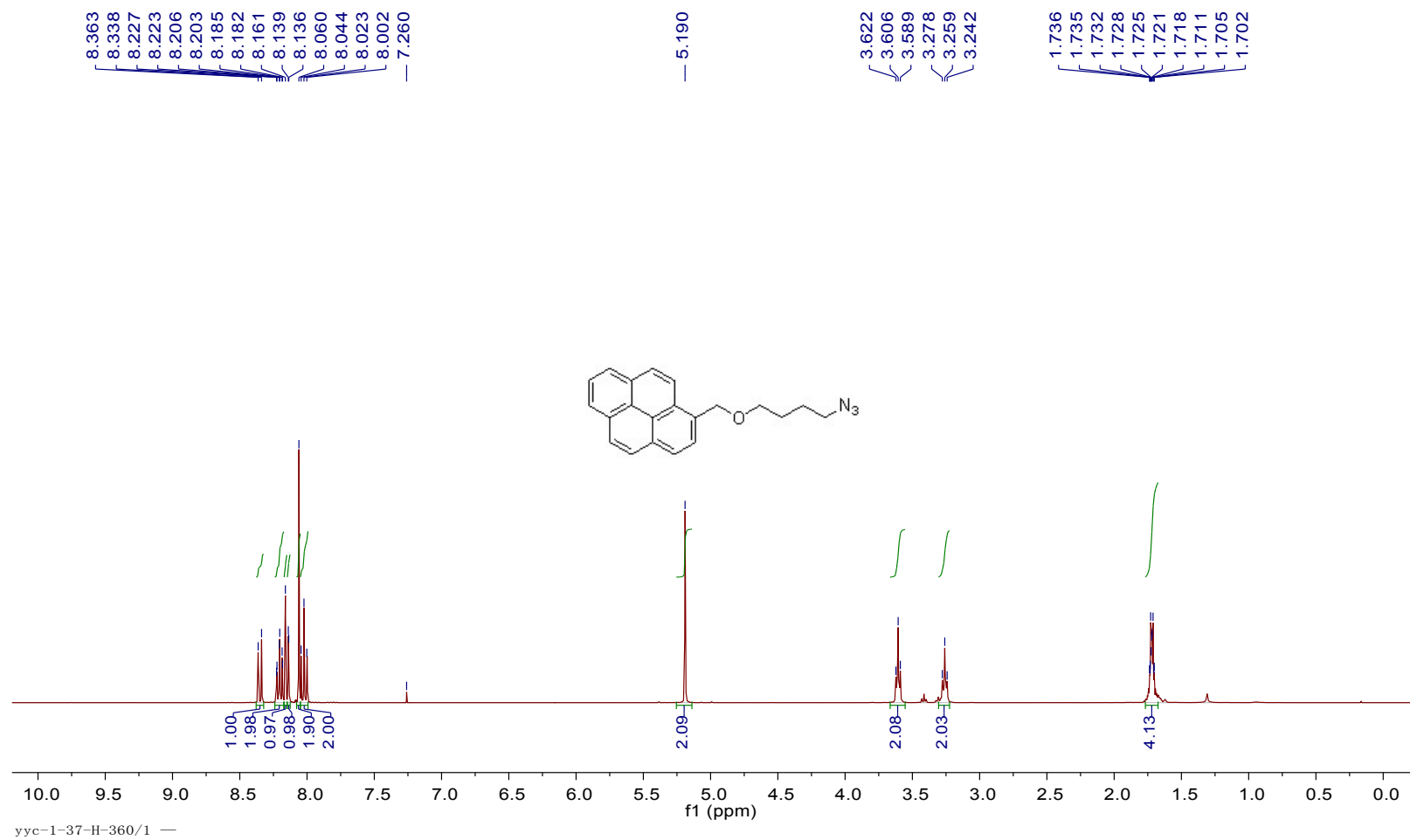




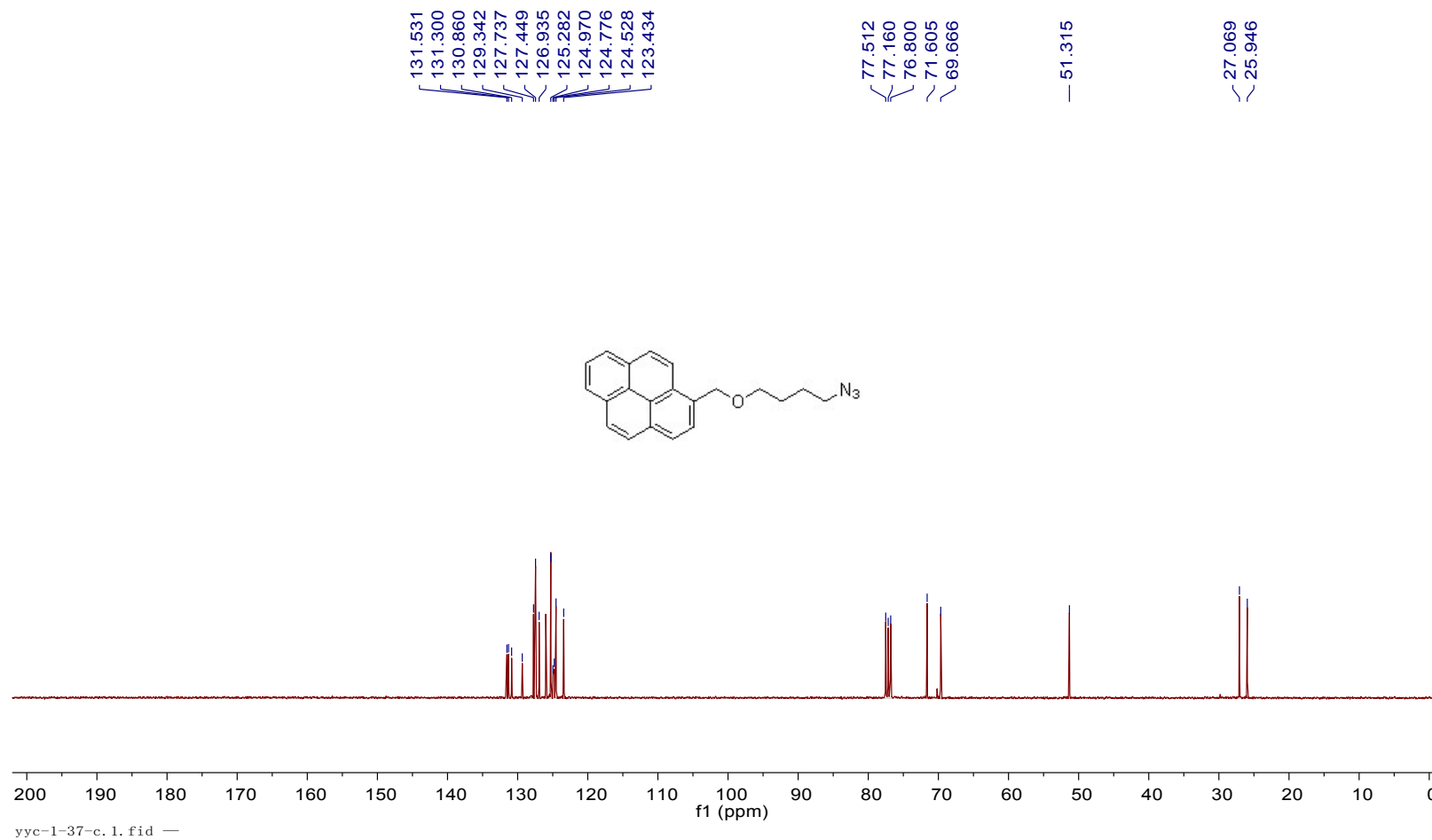
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of (2*S*,3*R*)-3a



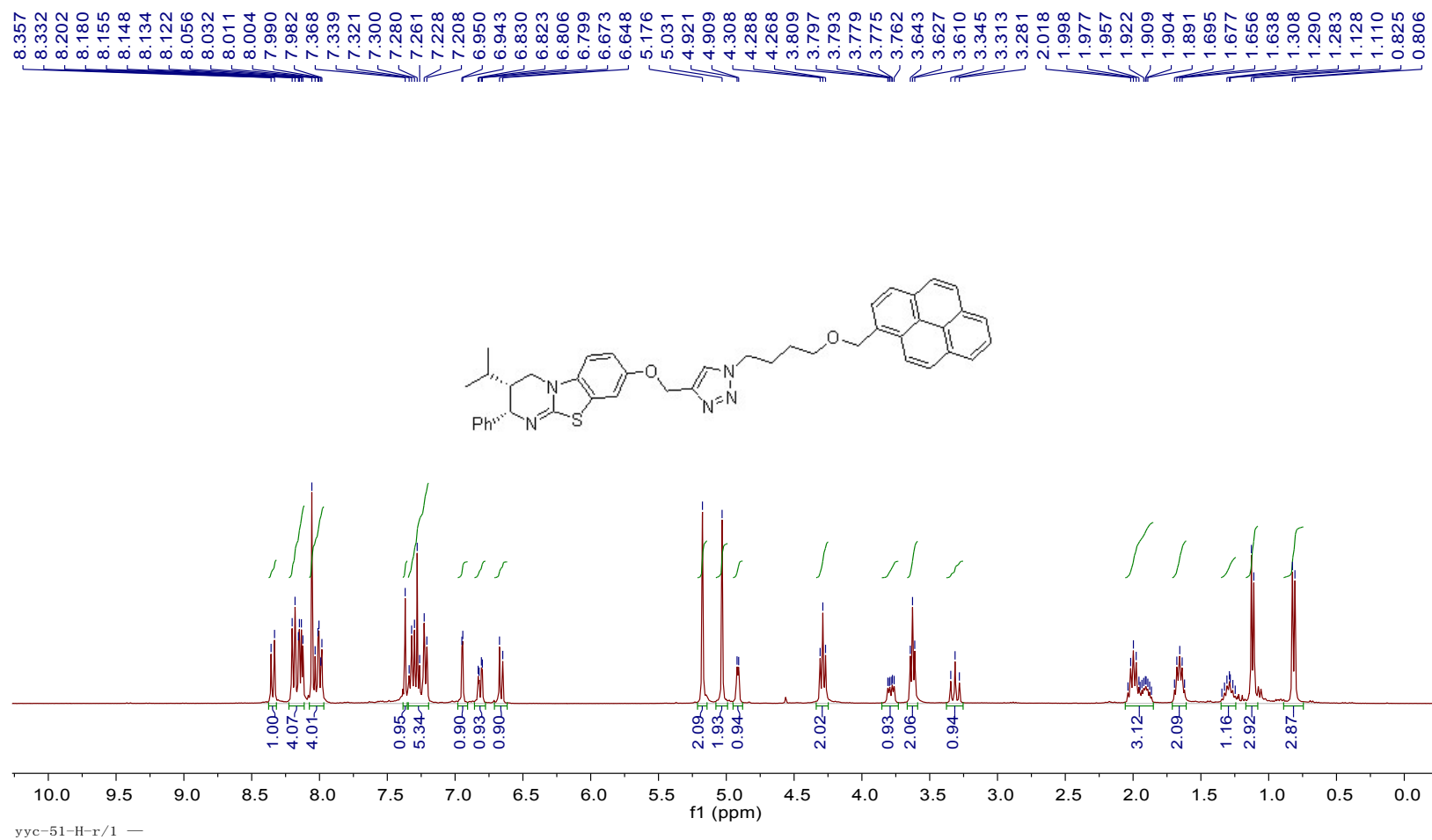
¹H NMR spectrum of (2S,3R)-4



¹H NMR spectrum of **7**

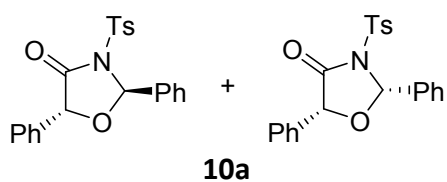


$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum of **7**



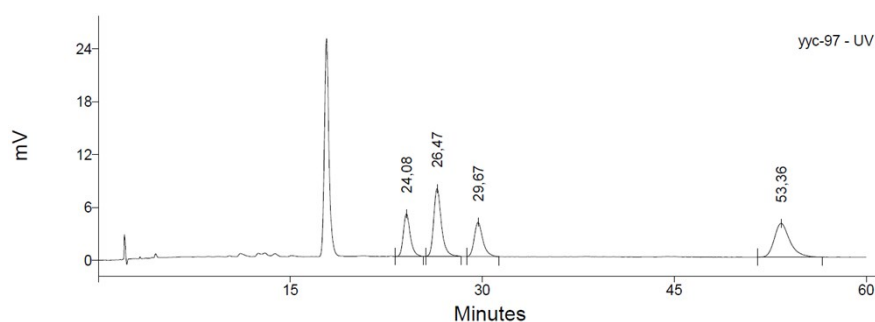
¹H NMR spectrum of (2*S*,3*R*)-pyr-hyperBTM

10. HPLC Traces



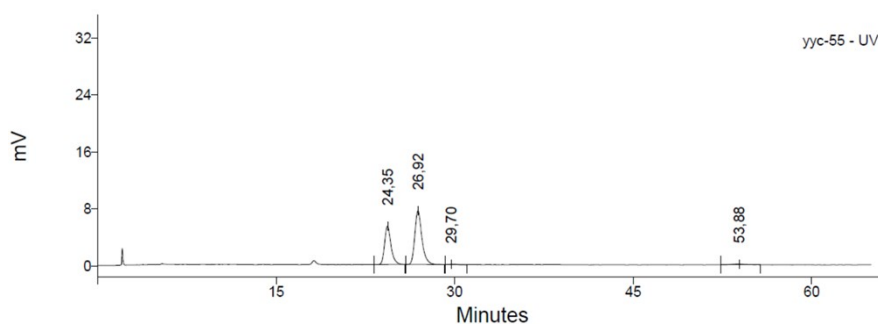
Data for the anti diastereoisomer: Chiral HPLC analysis, Chiralcel AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 39 °C) t_R (2*R*,5*R*): 26.9 min, t_R (2*S*,5*S*): 53.9 min, 96% ee.

Data for the syn diastereoisomer: Chiral HPLC analysis, Chiralcel AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 39 °C) t_R (2*S*,5*R*): 24.3 min, t_R (2*R*,5*S*): 29.7 min, 98% ee.



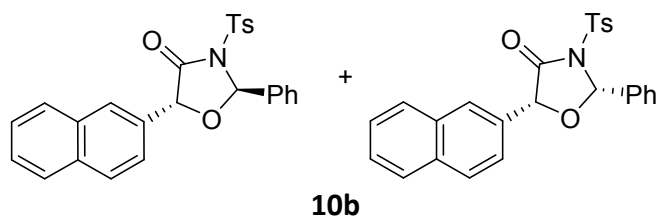
Informations :
AD-H- hex/IPA: 95/5 - 1.5 ml/min - 211nm 39°C

Résultats d'intégration						
#	Nom du pic	Tr.	Aire	% Aire	Asymétrie (USP, EP)	Plateaux (EP)
1		24,08	181,54	18,17	1,29	10241,33
2		26,47	320,43	32,06	1,32	10354,41
3		29,67	179,24	17,94	1,32	10369,27
4		53,36	318,12	31,83	1,34	10340,54
SOMME			999,34	100,00		



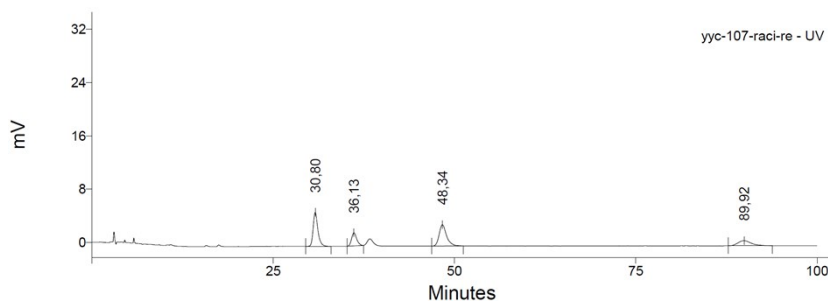
Informations :
AD-H- hex/IPA: 95/5 - 1.5 ml/min - 211nm 39°C

Résultats d'intégration						
#	Nom du pic	Tr.	Aire	% Aire	Asymétrie (USP, EP)	Plateaux (EP)
1		24,35	201,78	38,56	1,30	10571,21
2		26,92	312,21	59,67	1,30	10738,02
3		29,70	2,40	0,46	#	9278,77
4		53,88	6,83	1,31	1,16	12541,96
SOMME			523,21	100,00		



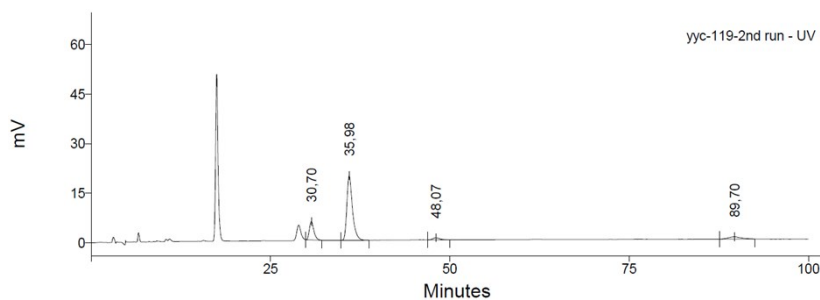
Data for the anti diastereoisomer: Chiral HPLC analysis, Chiralcel AD-H (90:10 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 40 °C) t_R (2*R*,5*R*): 36.0 min, t_R (2*S*,5*S*): 89.7 min, 86% ee.

Data for the syn diastereoisomer: Chiral HPLC analysis, Chiralcel AD-H (90:10 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 40 °C) t_R (2*S*,5*R*): 30.7 min, t_R (2*R*,5*S*): 48.1 min, 69% ee.



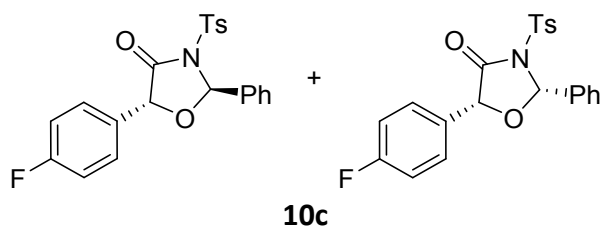
Informations :
AD-H - hex/IPA: 90/10 - 1.0 mL/min - 211 nm 40°C

Résultats d'intégration						
#	Nom du pic	Tr.	Aire	% Aire	Asymétrie (USP, EP)	Plateaux (EP)
1		30,80	228,85	35,27	1,31	11992,30
2		36,13	95,55	14,73	1,34	12844,34
3		48,34	225,88	34,81	1,33	12063,38
4		89,92	98,59	15,19	1,33	12260,32
SOMME			648,87	100,00		



Informations :
AD-H - hex/IPA: 90/10 - 1.0 ml/min - 211 nm 40°C

Résultats d'intégration						
#	Nom du pic	Tr.	Aire	% Aire	Asymétrie (USP, EP)	Plateaux (EP)
1		30,70	244,88	17,13	1,27	12220,91
2		35,98	1057,95	74,01	1,42	11344,14
3		48,07	44,64	3,12	1,40	11825,84
4		89,70	82,02	5,74	1,28	12168,98
SOMME			1429,49	100,00		



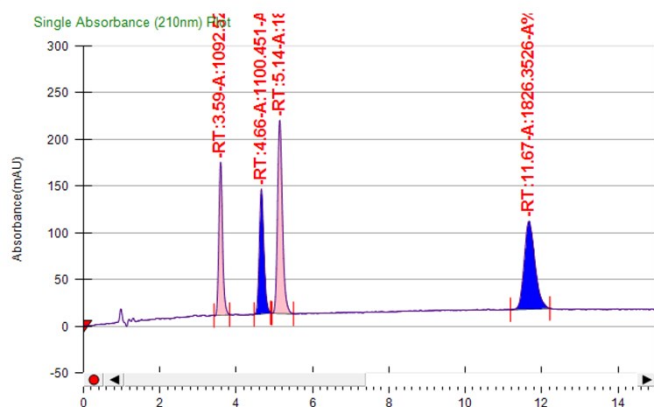
SFC Analysis: CHIRALPAK® AD-H, 35 °C, 5% MeOH, 4.0 mL min⁻¹, 150 bar, retention times:

Data for the anti diastereoisomer: t_R (2*S*,5*S*): 4.9 min, t_R (2*R*,5*R*): 11.6 min, 85% ee.

Data for the syn diastereoisomer: t_R (2*S*,5*R*): 4.6 min, t_R (2*R*,5*S*): 3.5 min, 60% ee.

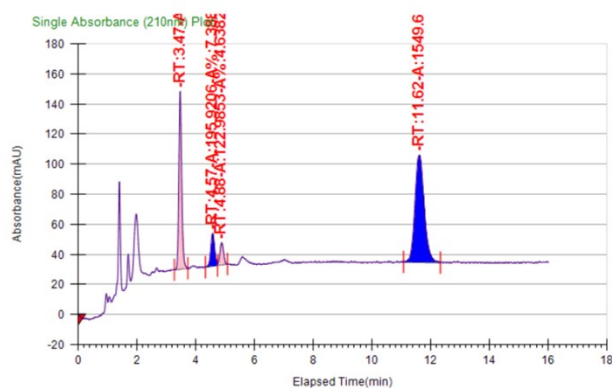
The *anti/syn* attribution is made according to the NMR which indicates the main diastereoisomer. The attribution of the configurations is made by analogy with the data of ref 5.

Sample	Column	Co-Solvent	Back Pressure	Col. Temp.
YYC-106 racemic	AD-H 250*4.6	EtOH	150	34.1
Instrument Method	Inj. Vol.	Total Flow	Pressure Drop	Wavelength
Iso150b-4ml-15cos-35deg	5	4	53	0

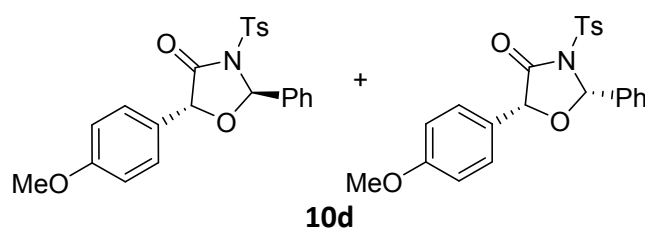


Peak #	Peak Name	Area %	Area	Ret. Time	Height
1	Peak1	18.6805	1092.5212	3.59 min	163.5718
2	Peak2	18.816	1100.451	4.66 min	133.4745
3	Peak3	31.2756	1829.1474	5.14 min	206.6156
4	Peak4	31.2279	1826.3526	11.67 min	94.1105

Sample	Column	Co-Solvent	Back Pressure	Col. Temp.
YYC-82	AD-H 250*4.6	EtOH	150	34.4
Instrument Method	Inj. Vol.	Total Flow	Pressure Drop	Wavelength
Iso150b-4ml-15cos-35deg	5	4	56	0



Peak #	Peak Name	Area %	Area	Ret. Time	Height
1	Peak1	29.5278	782.9463	3.47 min	118.1412
2	Peak2	7.3889	195.9206	4.57 min	21.7665
3	Peak3	4.6382	122.9853	4.88 min	14.6723
4	Peak4	58.445	1549.6999	11.62 min	71.3037



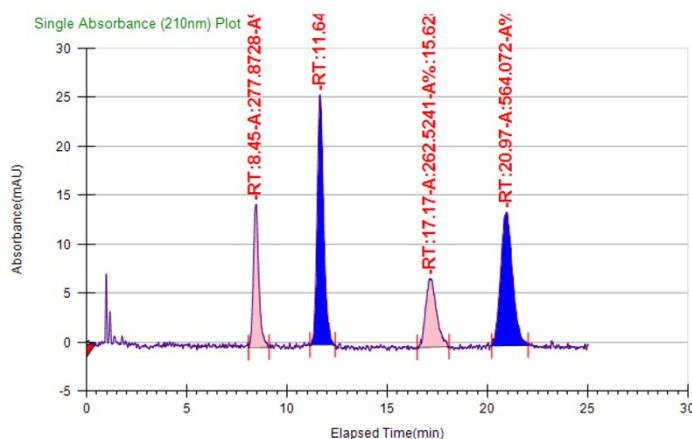
SFC Analysis: [CHIRALPAK® AD-H, 35 °C, 5% MeOH, 4.0 mL min⁻¹, 150 bar, retention times:

Data for the anti diastereoisomer: t_R (2*R*,5*R*): 22.8 min, t_R (2*S*,5*S*): 11.9 min, 87% ee.

Data for the syn diastereoisomer: t_R (2*S*,5*R*): 8.4 min, t_R (2*R*,5*S*): 17.8 min, 64% ee.

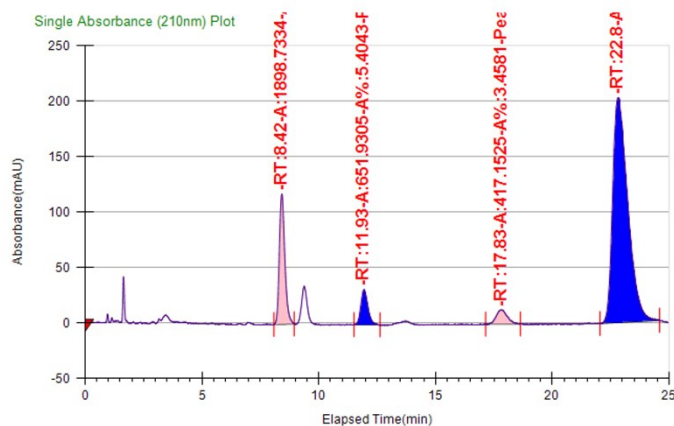
The *anti/syn* attribution is made according to the NMR which indicates the main diastereoisomer. The attribution of the configurations is made by analogy with the data of ref 5.

Sample	Column	Co-Solvent	Back Pressure	Col. Temp.
YYC-98 racemic	AD-H 250*4.6	Methanol	150	35
Instrument Method	Inj. Vol.	Total Flow	Pressure Drop	Wavelength
Iso150b-4ml-10cos-35deg	5	4	51	0



Peak #	Peak Name	Area %	Area	Ret. Time	Height
1	Peak1	16.5418	277.8728	8.45 min	14.6069
2	Peak2	34.2508	575.3513	11.64 min	25.525
3	Peak3	15.6281	262.5241	17.17 min	6.9836
4	Peak4	33.5793	564.072	20.97 min	13.6293

Sample	Column	Co-Solvent	Back Pressure	Col. Temp.
YYC-121	AD-H 250*4.6	Methanol	150	36.1
Instrument Method	Inj. Vol.	Total Flow	Pressure Drop	Wavelength
Iso150b-4ml-10cos-35deg	5	4	50	0



Peak #	Peak Name	Area %	Area	Ret. Time	Height
1	Peak1	15.74	1898.7334	8.42 min	117.5466
2	Peak2	5.4043	651.9305	11.93 min	31.4974
3	Peak3	3.4581	417.1525	17.83 min	12.8126
4	Peak4	75.3976	9095.3156	22.8 min	202.6987

