Unsymmetrical pentamethine cyanines visualizing physiologic acidities from whole animal to cellular scale with pH-responsive deep-red fluorescence

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Supporting Information

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1. Additional Figures and Tables

Figure. S1. ¹H-NMR spectra of **DR6** (500 MHz, DMSO-d6) with its protonated with 4% TFA (**DR6-I**) and deprotonated forms with NaHCO₃ (**DR6-II**).¹ The significant distinction between protonated form (**DR6-I**) and deprotonated form (**DR6-II**) is the proton in the indole nitrongen atom (H*). The NMR spectra of **DR6-I** and **DR6-II** were shown below (**Figure. RL4**). The NMR peak of H* was only observed in the spectra of **DR6-I**. Furthermore, addition of acid led to downfield shifts of all proton signals, which may attribute to the decrease of electron density. Besides, the chemical shift of H-8 was 4.28 and 4.19 in **DR6-I** and **DR6-II**, respectively, which correlated well with the increased electron-withdrawing ability of quaternary ammonium.



Figure. S2. pH dependence of the normalized absorbance (absorbance/absorbance_{max})

Compounds	Form	$\Delta E/ eV$	f	Theory	Experiment
				$\lambda_{Abs\ (max)}/$	$\lambda_{Abs\ (max)}/$
				nm	nm
DR1	protonated	2.47	2.13	525	637
	deprotonated	2.72	1.89	493	486
DR2	protonated	2.46	2.14	525	635
	deprotonated	2.69	1.91	497	463
DR3	protonated	2.51	2.17	512	636
	deprotonated	2.70	1.85	496	488
DR4	protonated	2.49	2.21	526	635
	deprotonated	2.79	2.09	484	475
DR5	protonated	2.38	2.02	548	647
	deprotonated	2.64	1.82	510	500
DR6	protonated	2.44	2.14	532	641
	deprotonated	2.69	1.91	499	497

Table S1. TDDFT excitation energies, oscillator strengths, calculated absorption wavelengths

 and experimental absorption wavelengths of **DR1-6**.

 $\lambda_{Abs max}$ is the absorption maxima . ΔE is The excitation HOMO–LUMO energy gap. f represents oscillator strengths.



Figure S3. Comparison of the HOMO and LUMO energy levels, excitation energies, and oscillator strengths for **DR1-6**, based on TDDFT calculations at the B3LYP/6–311 G(d,p) level.



Fig. S4. Cell viability of **DR1** on Hela cells by a standard CCK-8 assay. Data are expressed as mean values \pm standard error of the mean of three independent experiments.



Fig. S5. Fluorescence intensities of **DR1** (1.0 μ M) in the presence of endogenous ions and other potential interferents in phosphate buffer (pH 7.4) (200 μ M for NH₄⁺, Cu²⁺, Co²⁺, Ca²⁺, Mg²⁺, Fe²⁺, Fe³⁺, H₂O₂ and 5 mM for Glucose, Glutathione and Cysteine). Data were acquired in with λ ex= 640 nm. Each experiment was performed in triplicate, and error bars are determined from the mean and standard deviation (SD).



Fig. S6. Mean fluorescence intensity in the abdominal region at 30 min post administration of **DR1** (n = 3). Error bars represent mean deviation (S.D.), *P* values were calculated using two-tailed Student's t-tests.

2. NMR spectra of dyes DR1-6.



¹H NMR spectra of **DR1**





¹H NMR spectra of **DR2**





¹H NMR spectra of **DR3**





¹H NMR spectra of **DR4**





¹H NMR spectra of **DR5**







¹³C NMR spectra of **DR6**

3. High-resolution mass spectra of dyes DR1-6.



High-resolution mass spectra of DR1



High-resolution mass spectra of DR2



High-resolution mass spectra of DR3



High-resolution mass spectra of DR4



High-resolution mass spectra of DR5



High-resolution mass spectra of DR6