

Site specific role of bifunctional graphitic carbon nitride catalyst for the sustainable synthesis of 3,3-spirocyclic oxindoles in aqueous media

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1. General

All the chemicals used were of research grade (purchased from Sigma Aldrich, Acros etc.) and used without further purification. The melting points of all compounds were determined on a Toshniwal apparatus in capillary and uncorrected. ^1H and ^{13}C NMR spectra were recorded in CDCl_3 and $\text{DMSO } d_6$ using TMS as an internal standard on a JEOL NMR spectrometer at 400 and 100 MHz respectively. Chemical shifts are expressed in parts per million (ppm) using tetramethylsilane (TMS) as an internal standard. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet and m = multiplet. Mass spectrum of representative compound was recorded on Waters-Xevo G2S Q-Tof. X-ray diffraction (XRD) measurements for phase determination were recorded by Philips powder diffractometer (PW3040/60) with Cu K_α radiation (1.54060nm) operating in a continuous mode to collect 2θ values with a scan rate of $0.02^\circ/\text{min}$. SEM and EDX measurements were performed using a FEI Quanta 200F SEM. The size and morphology of the synthesized material was observed by transmission electron microscopy (TEM) using a JEOL 1011 at an accelerating voltage of 200kV. (XPS) were measured on a commercial SPECS spectrometer (Germany), equipped with an Al-K_α X-ray source (1486.5eV). The UV-Vis spectra were recorded using Ocean optics USB 2000 spectrophotometer in the solution form. The Raman spectra were recorded by micro-Raman spectrometer (Jobin Yvon Horibra LABRAM-HR visible 400-1100 nm). IR spectra were recorded on a Shimadzu FT IR- 8400S spectrophotometer using KBr pellets. The Elemental analysis of the samples has been done by Elemental Analyzer Perkin Elmer PE 2400 and ELEMENTAR Vario EL III.

2. Synthesis of g- C_3N_4

Melamine (5 g) was placed in a crucible (enclosed with a lid) and heated at 550°C (at a rate of $4^\circ\text{C}/\text{min}$) in a muffle furnace for 3 h. After that, obtained material was crushed into fine powder with the help of a pestle-mortar and referred as g- C_3N_4 .

3. Synthesis of Sg- C_3N_4

The as prepared g- C_3N_4 and 30% aqueous H_2SO_4 was placed in flask. The flask was attached to a 12 mm tip diameter probe and the reaction mixture was sonicated for the 4h at 50% power of the processor and 230W output in a 4 s pulse mode. The resulting mixture was poured in ice-cold water and centrifuged (12000 rpm for 10 min twice). The obtained solid material was dried under vacuum for 24 h and referred as Sg- C_3N_4 .

4. Synthesis of spiro-pyrano chromene derivatives

A mixture of isatins (2.0 mmol), malononitrile/ ethyl cyanoacetate (2.0 mmol), 4-hydroxycoumarin (2.0 mmol) and 20 wt% Sg-C₃N₄ in 20 ml water were mixed, and the reaction mixture was heated under reflux for appropriate time. The progress of the reaction was checked on TLC. After completion of the reaction the solid product was filtered along with the catalyst. The precipitate was dissolved in ethyl acetate and the catalyst was recovered by centrifugation (12000 rpm). The above residual solution was removed under reduced pressure. The crude product was subjected to purification by recrystallization using ethanol.

5. Synthesis of spiro indole-3,1'-naphthalene tetracyclic system

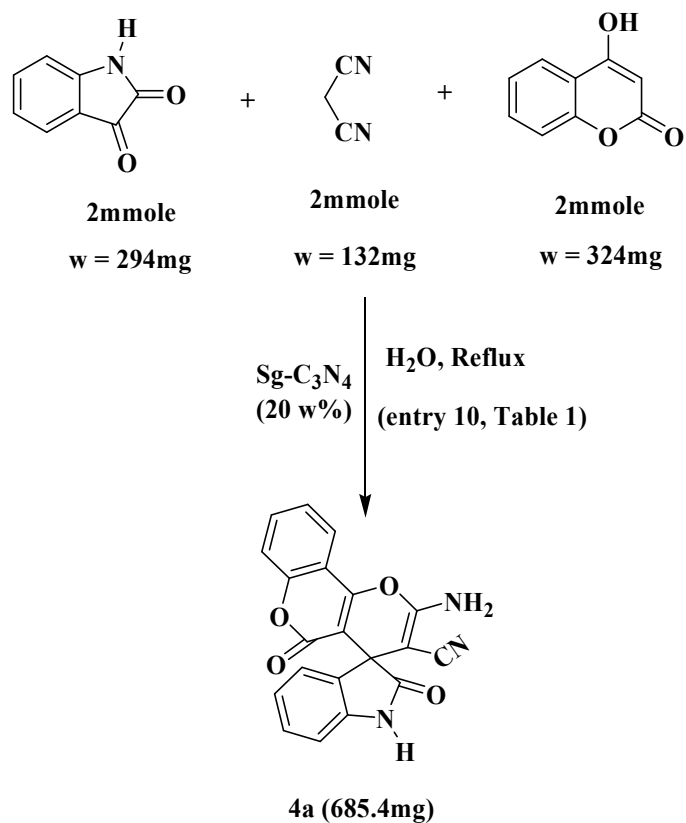
A mixture of isatins (2.0 mmol), malononitrile (4.0 mmol), cyclic ketones (2.0 mmol) and 20 wt% Sg-C₃N₄ in 20 ml water were mixed, and the reaction mixture was heated under reflux for appropriate time. The progress of the reaction was checked on TLC. After completion of the reaction the solid product was filtered along with the catalyst. The precipitate was dissolved in ethyl acetate and the catalyst was recovered by centrifugation (12000 rpm). The above residual solution was removed under reduced pressure. The crude product was subjected to purification by recrystallization using ethanol.

6. Heterogeneous nature and recyclability of Sg-C₃N₄

To confirm the heterogeneous nature of Sg-C₃N₄ in reaction, the model reaction was carried out again under similar reaction conditions with the catalyst procured from a previous cycle. After 4 min, the catalyst was separated from the reaction mixture. The reaction was continued with filtrate for another 5h and the reaction conversion was monitored for every 1h. It was observed that further increment in conversion was not observed even after 8h. These results revealed that reaction was occurring only due to the Sg-C₃N₄. This whole experiment confirms the heterogeneous nature of present catalytic system.

Recycling experiments were performed by choosing the model reaction Sg-C₃N₄ as a solid catalyst. When reaction was completed, the reaction mixture was filtered and solid precipitate was dried along with the catalyst. Then, the solid precipitate was dissolved in ethanol and catalyst was recovered by filtration. The recovered catalyst was washed with water and ethanol and reused in succeeding 7 reaction cycles with slight loss of catalytic activity. The characteristics obtained from XRD, SEM, TEM of fresh and used catalysts are similar, which suggest the retention of structure and morphology of Sg-C₃N₄ after repeated use as catalyst.

Calculation of TOF



Calculation of %ge yield

$$\%ge \text{ yield} = \frac{\text{Experimental yield}}{\text{Theoretical yield}} \times 100$$

$$\begin{aligned} \text{eg for the product 4a} &= \frac{685.4\text{mg}}{714\text{mg}} \times 100 \\ & \text{(Table 1, entry 10)} \\ &= 96\% \end{aligned}$$

Calculation of TOF Value

$$\text{TOF} = \frac{\text{No. of Moles of final product produced}}{\text{Amount of loaded Catalyst} \times \text{Time (min.)}}$$

$$\text{wt\% of loaded Catalyst} = \frac{\text{Amount loaded Catalyst}}{\text{Total Amount}} \times 100$$

For the product 4a (Table 1, entry 10)

Assuming amount of loaded Catalyst = x gm

$$\frac{20}{100} = \frac{x}{0.294 + 0.132 + 0.324 + x}$$

$$\frac{1}{5} = \frac{x}{0.750 + x}$$

$$0.750 + x = 5x$$

$$0.750 = 4x$$

$$x = \frac{0.750}{4} = 0.1875\text{gm}$$

$$\text{TOF} = \frac{0.6854}{357 \times 0.1875 \times 10} = \frac{0.6854}{669.375}$$

$$\text{TOF} = 1.024 \times 10^{-3} \text{ mol gm}^{-1} \text{ min}^{-1}$$

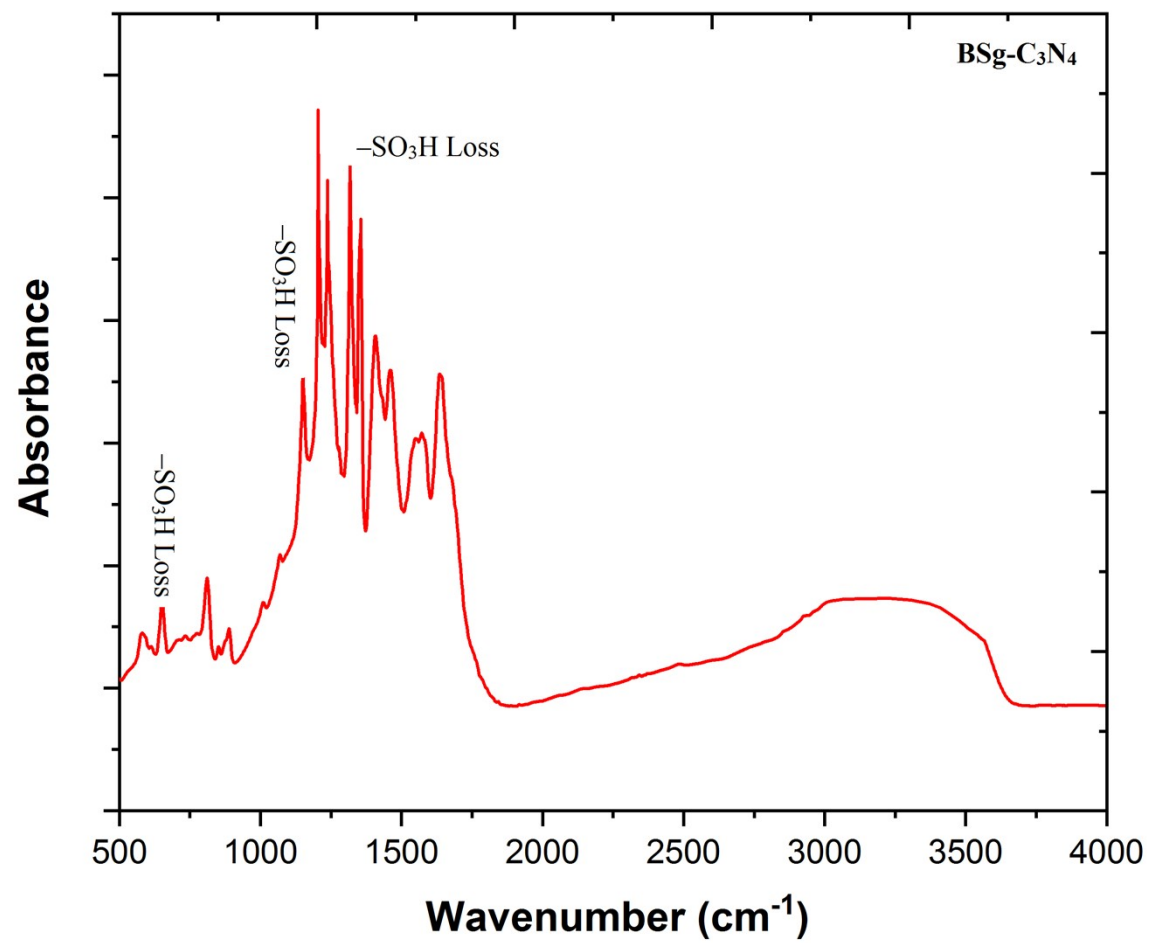


Fig S1: FT-IR spectra of BSg-C₃N₄

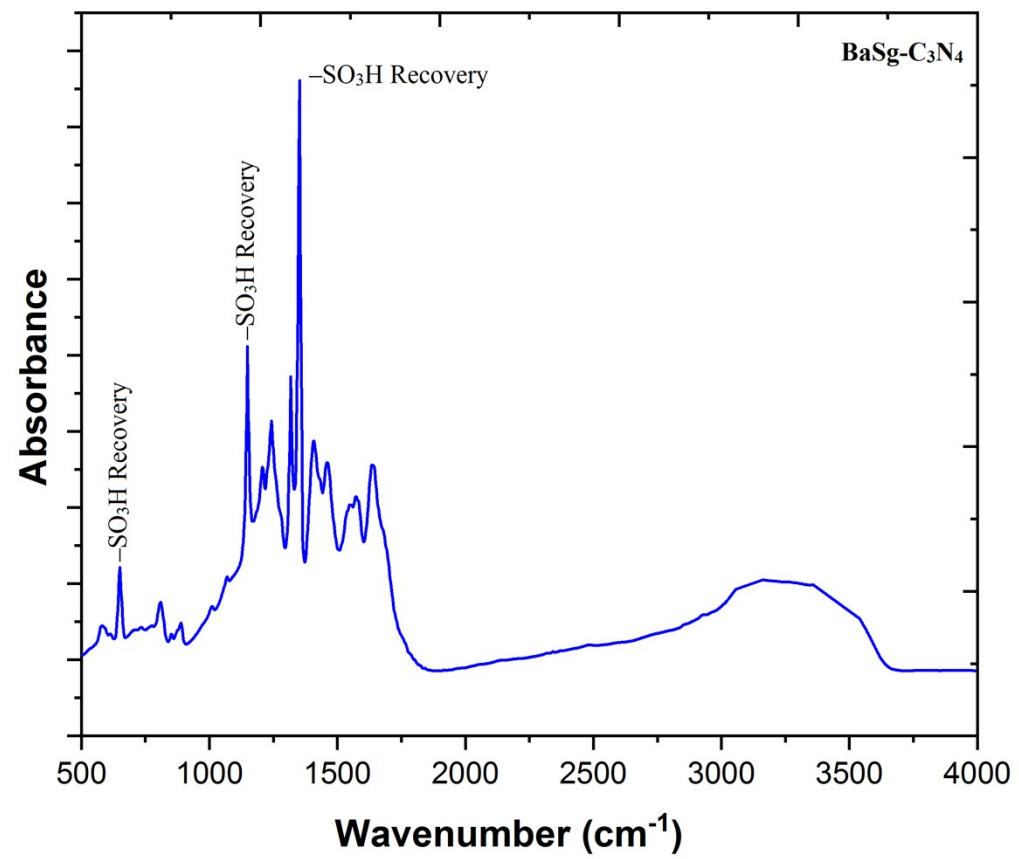


Fig S2: FT-IR spectra of BaSg-C₃N₄

Element	Atomic %
C K	33.48
O K	21.54
N K	44.98
Total	100.00

Table S1: EDAX elemental analysis of g-C₃N₄

Element	Atomic %
C K	19.00
O K	24.10
N K	49.92
S K	6.98
Total	100.00

Table S2: EDAX elemental analysis of Sg-C₃N₄

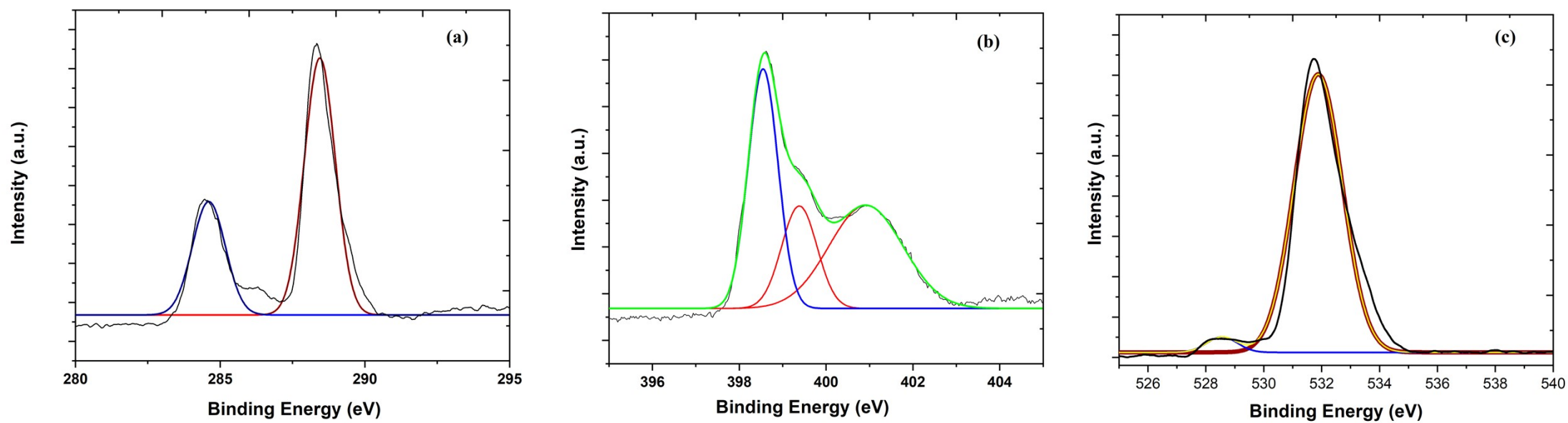


Fig.S3: XPS spectrum of (a) C1s spectrum of g-C₃N₄ (b) N1s spectrum of g-C₃N₄ (c) O1s spectrum of g-C₃N₄

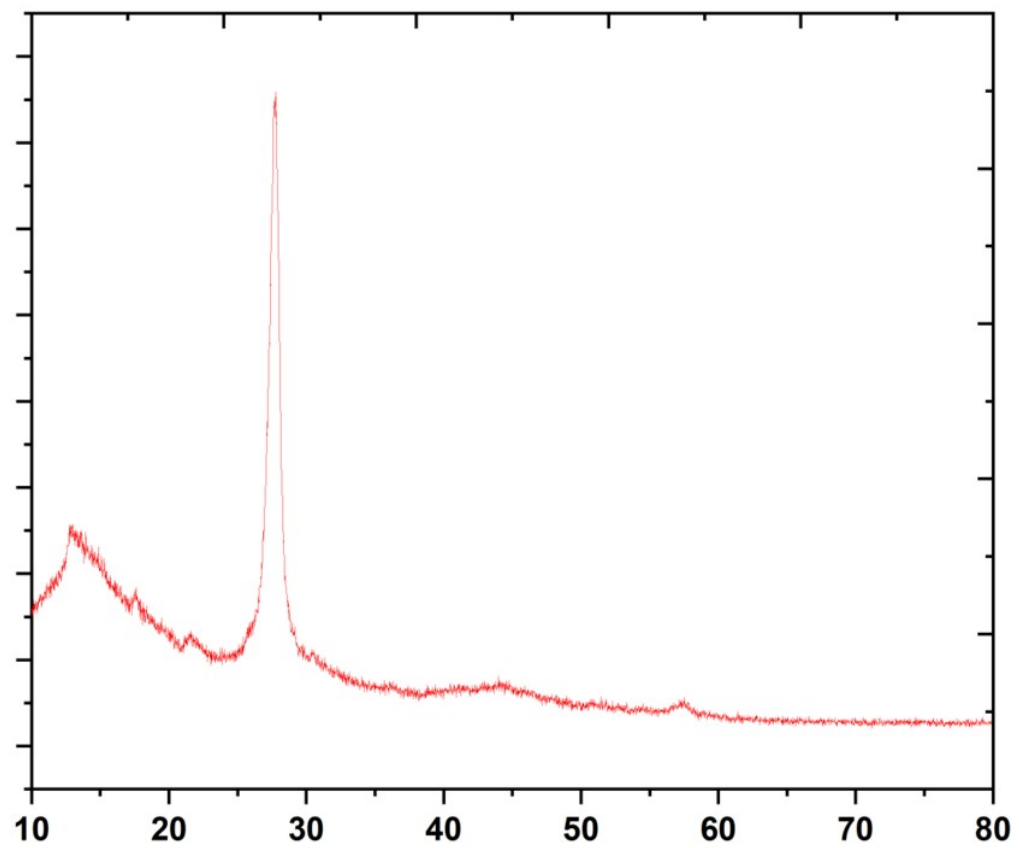


Fig.S4: XRD patterns of reused Sg-C₃N₄

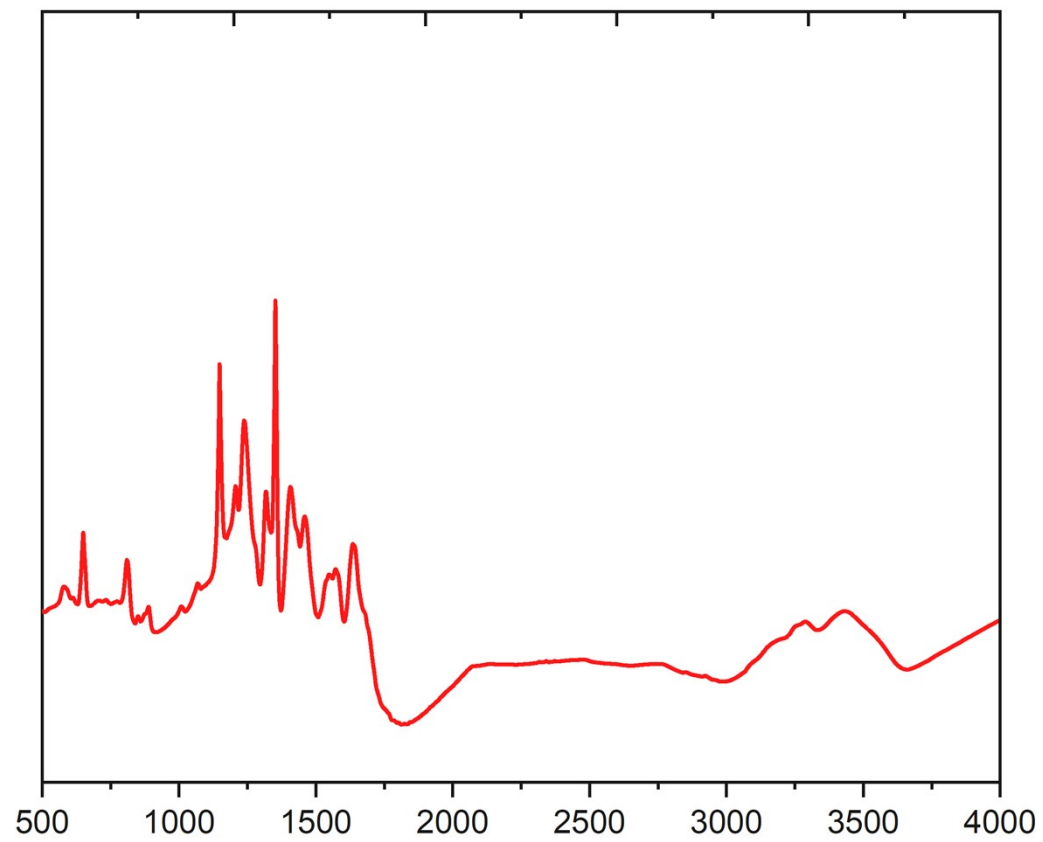


Fig.S5: FT-IR spectra of reused Sg-C₃N₄

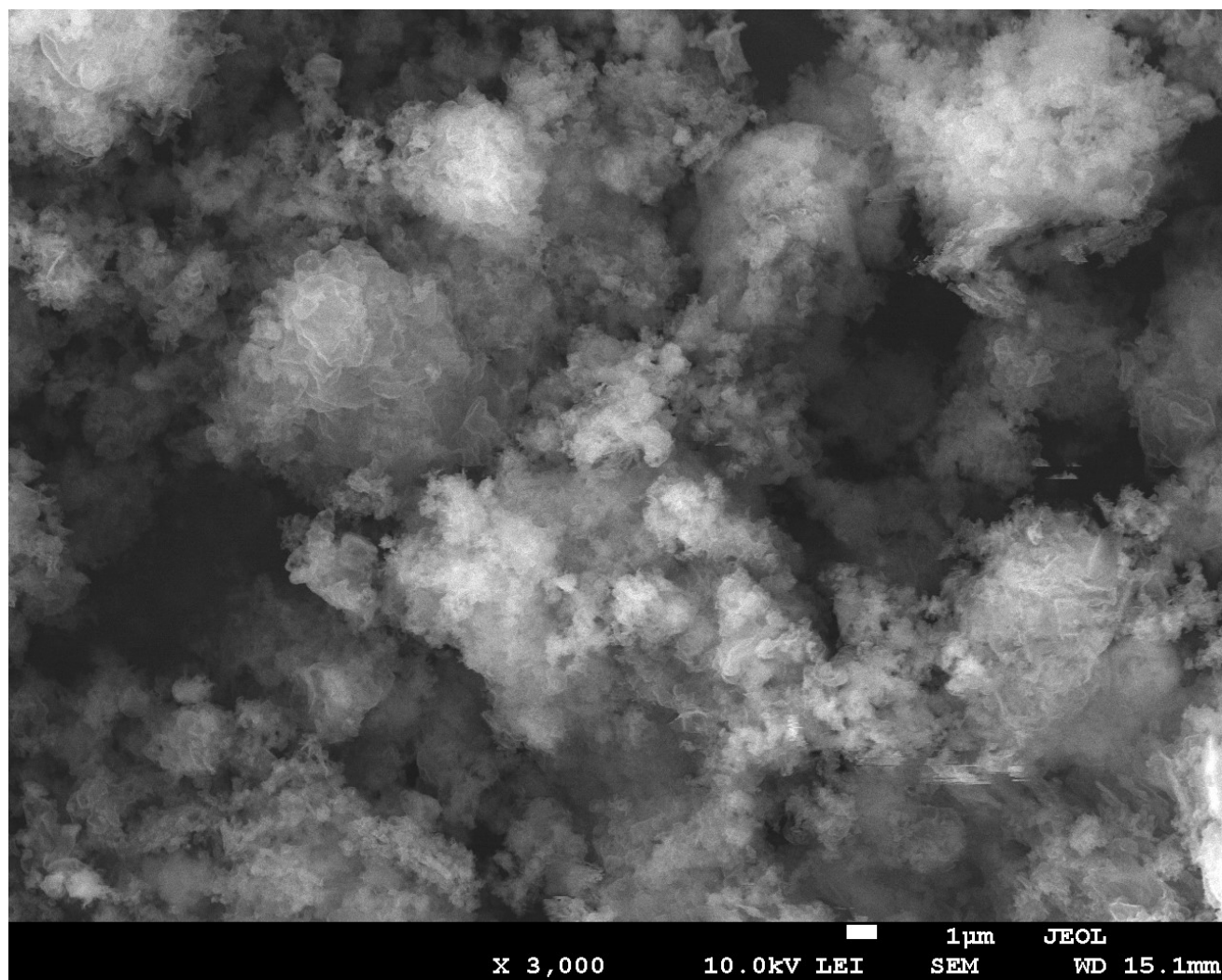


Fig.S6: SEM images of reused Sg-C₃N₄

Elemental Analysis (CNOS)

Table S3

SI. No	Sample	N%	C%	S%	O%
1	Sg-C ₃ N ₄	49.41	19.82	6.39	24.38
2	g-C ₃ N ₄	44.11	34.24	ND	21.65

9. Spectral data of synthesized compounds (6a-6k)

(6a)¹: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 11.34(s,1H,NH), 7.51-7.26(m,3H,NH₂, ArH), 7.06-6.86(m,3H,ArH), 5.92(s,1H, CH), 2.91(s,1H,CH₂), 2.50 (s, 1H, CH), 2.18-2.12(d,J=18MHz,1H,CH₂), 1.93(s,1H,CH₂), 1.61-1.48(m,3H,CH₂).¹³C NMR (75 MHz, DMSO-d₆): δ 173.6,143.3,142.7, 131.3, 125.8, 125.4, 124.1, 123.4, 122.9, 115.9, 111.1, 110.6, 81.9, 55.0, 42.6, 37.4, 24.9, 23.8, 20.6. +ESI MS (m/z): 342.13 [M+H]⁺

(6b)¹: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 7.55-6.92 (m, 6H, NH₂,ArH), 5.91 (s,1H, CH), 3.29(s, 3H,N-CH₃), 3.05-3.02 (d, J=9.9MHz, 1H,CH₂), 2.50(s,1H, CH₂), 2.26-1.06(m, 4H, CH₂), 0.79-0.77(d, J=6MHz,3H,CH₃).¹³C NMR (75 MHz, DMSO-d₆): δ 171.9, 144.6, 142.7, 131.4, 125.6, 125.1, 124.1, 123.9, 122.1, 115.9, 111.0, 110.5-110.2, 81.9, 56.4, 54.6, 42.7, 33.8, 32.0, 31.1, 27.5-27.1, 21.9.+ESI MS (m/z): 355.14 [M]⁺

(6c)¹: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 11.39 (s,1H, NH), 10.85 (s, 2H, NH₂), 7.66-6.84 (m, 5H, ArH, CH), 5.78 (s, 1H, CH), 3.94-3.73 (m,4H,CH₂), 3.34-3.14 (m, 2H, CH₂), 2.28-2.13 (m, 1H, CH₂), 1.52-1.44(m,1H, CH), 0.74-0.62 (m, 1H, CH₂).¹³C NMR (75 MHz, DMSO-d₆): δ 173.2, 143.3-143.2, 142.6, 131.5, 130.1, 126.9-126.3, 125.3-125.1, 124.4-124.2, 123.6, 122.4, 121.1, 111.2-111.0, 110.5-110.2, 106.6-106.2, 81.4-81.1, 64.5-64.4, 54.8-54.2, 42.5, 37.0, 35.9, 35.7, 32.6.+ESI MS (m/z): 400.14 [M+H]⁺

(6d)²: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 7.56 (s, 2H, NH₂), 7.52-7.46(t, J=7.8MHz, 1H, ArH), 7.26-7.23 (d, J=7.8MHz, 1H, ArH), 7.18-7.13 (m, 1H, ArH), 6.95-6.93 (d, J=7.5MHz, 1H, ArH), 5.93 (s, 1H, CH), 3.28 (s, 3H, N-CH₃), 2.98-2.95 (d, J=9MHz, 1H, CH), 2.50 (s, 1H, CH₂),2.18-1.45 (m, 4H, CH₂), 0.45-0.34(m,1H, CH₂). ¹³C NMR (75 MHz, DMSO-d₆): δ 172.0, 144.6, 142.6, 131.4, 125.8, 125.1, 124.2, 124.1, 122.2, 115.9, 110.5, 110.2, 82.0, 54.7, 42.6, 37.5, 27.1, 24.9, 23.8, 20.6. +ESI MS (m/z): 355.14 [M]⁺

(6e)²: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 10.64 (s,1H, NH), 8.12 (s, 2H, NH₂), 7.26 (s, 1H, ArH), 7.05-6.98 (m, 1H, ArH), 6.81-6.59 (m, 2H, ArH), 5.60 (s, 1H, CH), 3.28-2.73 (m, 2H,CH₂), 2.32 (s, 1H, CH), 2.02-1.90 (m, 2H, CH₂), 1.31-1.22(m,3H, CH₃), 0.71-0.57 (m, 1H, CH).

^{13}C NMR (75 MHz, DMSO- d_6): δ 174.6, 147.9, 142.7, 129.9-129.5, 127.8-127.0, 126.3, 124.7-124.5, 122.3, 121.5, 120.8, 117.4-117.1, 116.4, 115.5, 110.0, 109.5, 82.2, 55.3, 33.8, 32.2, 27.8, 22.0. +ESI MS (m/z): 356.14 [M+H] $^+$

(6f) 2 : ^1H -NMR (DMSO- d_6 300 MHz); δ (ppm) 7.69 (s, 2H, NH $_2$), 7.54-7.49 (m, 1H, ArH), 7.28-7.15 (m, 2H, ArH), 6.97-6.94 (d, J=7.2MHz, 1H, ArH), 5.79 (s, 1H, CH), 3.93-3.74 (m, 4H, OCH $_2$), 3.33-3.17 (m, 3H, CH $_3$), 2.50-2.16 (m, 2H, CH $_2$), 1.42-1.39 (d, J=10.5MHz, 1H, CH), 0.67-0.59 (m, 2H, CH $_2$). ^{13}C NMR (75 MHz, DMSO- d_6): δ 171.5, 144.6, 143.2, 131.7, 125.1, 124.3, 121.7, 121.2, 115.7, 110.9, 110.4, 106.2, 81.1, 64.5, 64.4, 54.2, 42.6, 37.0, 35.9, 32.6, 27.2. +ESI MS (m/z): 413.15 [M] $^+$

(6g) 2 : ^1H -NMR (DMSO- d_6 300 MHz); δ (ppm) 7.72 (s, 2H, NH $_2$), 7.69-6.89 (m, 4H, ArH), 5.79 (s, 1H, CH), 5.67 (s, 1H, CH), 5.28-5.21 (m, 2H, CH), 4.50-4.31 (m, 2H, CH $_2$), 3.90-3.64 (m, 3H, OCH $_2$), 3.35-3.18 (m, 1H, OCH $_2$), 2.29-2.11 (m, 2H, CH $_2$), 1.45-1.40 (m, 1H, CH), 0.79-0.57 (m, 2H, CH $_2$). ^{13}C NMR (75 MHz, DMSO- d_6): δ 171.4, 143.6-143.2, 131.8-131.4, 125.3-125.0, 124.3, 123.6-123.2, 122.4, 121.7-121.1, 118.3, 117.9, 115.7, 111.3-111.0, 110.9-110.5, 106.5-106.1, 81.1-81.0, 64.5-64.4, 63.0, 54.5-54.1, 42.6-42.4, 37.2, 36.9, 35.9-35.6, 32.6. +ESI MS (m/z): 439.16 [M] $^+$

(6h) 2 : ^1H -NMR (DMSO- d_6 300 MHz); δ (ppm) 11.25 (s, 1H, NH), 7.50 (s, 2H, NH $_2$), 7.20 (d, J=8.0MHz, 1H, ArH), 6.91 (d, J=8.0MHz, 1H, ArH), 6.64 (s, 1H, ArH), 5.87-5.97 (m, 1H, CH), 2.82-2.94 (m, 1H, CH), 2.30 (s, 3H, CH $_3$), 2.08-2.21 (m, 1H, CH $_2$), 1.82-2.02 (m, 1H, CH $_2$), 1.36-1.70 (m, 3H, CH $_2$), 0.47 (q, J=12.1MHz, 1H, CH $_2$). ^{13}C NMR (75 MHz, DMSO- d_6): δ 173.1, 142.2, 140.4, 131.5, 131.2, 125.6, 125.4, 123.6, 122.7, 115.5, 110.7, 110.5, 110.2, 81.5, 54.5, 42.1, 37.0, 24.4, 23.4, 21.1, 20.1. +ESI MS (m/z): 356.14 [M+H] $^+$

(6i) 1 : ^1H -NMR (DMSO- d_6 300 MHz); δ (ppm) 11.54 (s, 1H, NH), 7.65 (s, 2H, NH $_2$), 7.48 (d, J=8.4MHz, 1H, ArH), 7.05 (d, J=8.4MHz, 1H, ArH), 6.77 (s, 1H, ArH), 5.90-6.03 (m, 1H, CH), 2.86-3.02 (m, 1H, CH), 2.06-2.31 (m, 1H, CH $_2$), 1.87-2.07 (m, 1H, CH $_2$), 1.36-1.76 (m, 3H, CH $_2$), 0.49 (q, J=12.1MHz, 1H, CH $_2$). ^{13}C NMR (75 MHz, DMSO- d_6): δ 172.9, 142.0, 131.0, 126.7, 124.9, 124.7, 124.4, 124.3, 115.3, 112.4, 110.3, 110.1, 81.5, 54.8, 42.0, 37.0, 24.5, 23.4, 20.2. +ESI MS (m/z): 376.08 [M+H] $^+$

(6j)³: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 11.55 (s, 1H, NH), 7.55-7.71 (m, 3H, ArH, NH₂), 7.02 (d, J=8.4MHz, 1H, ArH), 6.91 (s, 1H, ArH), 5.91-6.02 (m, 1H, CH), 2.86-3.01 (m, 1H, CH), 2.08-2.31 (m, 1H, CH₂), 1.87-2.05 (m, 1H, CH₂), 1.37-1.75 (m, 3H, CH₂), 0.48(q, J=12.1MHz, 1H, CH₂).¹³C NMR (75 MHz, DMSO-d₆): δ 172.7, 142.3, 142.0, 133.7, 127.3, 124.6, 124.6, 124.3, 115.2, 114.1, 112.7, 110.3, 110.1, 81.5, 54.6, 42.0, 37.1, 24.4, 23.3, 20.1. +ESI MS (m/z): 419.03[M]⁺

(6k)³: ¹H-NMR (DMSO-d₆ 300 MHz); δ (ppm) 7.54 (s, 2H, NH₂), 7.47 (t, J=7.7MHz, 1H, ArH), 7.27 (d, J=7.7 MHz, 1H, ArH), 7.14 (t, J=7.7MHz, 1H, ArH), 6.95 (d, J=7.7MHz, 1H, ArH), 5.88-5.97 (m, 1H, CH), 3.78-3.92 (m, 2H, CH₂), 2.90-3.01 (m, 1H, CH), 2.07-2.21 (m, 1H, CH₂), 1.82-1.98 (m, 1H, CH₂), 1.56-1.68 (m, 1H, CH₂), 1.36-1.54 (m, 2H, CH₂), 1.17 (t, J=7.0MHz, 3H, CH₃), 0.41 (q, J=12.1MHz, 1H, CH₂).¹³C NMR (75 MHz, DMSO-d₆): δ 171.1, 143.2, 142.1, 131.0, 125.2, 124.8, 123.7, 123.6, 122.1, 115.4, 110.5, 110.0, 109.7, 81.5, 54.1, 42.1, 37.0, 34.7, 24.5, 23.2, 20.1, 12.1. +ESI MS (m/z):369.15[M]⁺

Spectral data of synthesized compounds (4a-4l)

(4a): ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm)10.63(s, 1H, NH), 7.91-7.89 (m,1H,ArH), 7.74-7.70 (m,1H,ArH), 7.62 (s, 2H, NH₂), 7.52-7.44 (m,2H,ArH), 7.19-7.15 (m,2H, ArH), 6.91-6.87 (t, J=7.6MHz, 1H,ArH), 6.82-6.80 (d, J=7.6MHz, 1H,ArH).¹³C NMR (100 MHz, DMSO-d₆): δ 177.6, 158.9, 158.8, 155.6, 152.5, 142.7, 134.2, 133.5, 129.4, 125.5, 124.6, 123.1, 122.5, 117.5, 117.2, 112.9, 110.0, 101.9, 57.5, 48.1. +ESI MS (m/z): 358.07 [M+H]⁺

(4b): ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 10.52 (s, 1H, NH), 8.15 (s, 2H, NH₂), 7.97 (s, 1H, ArH), 7.70-7.67(m, 1H, ArH), 7.48-7.39 (m, 2H, ArH), 7.15-7.13 (m, 2H, ArH),6.82-6.69(m, 1H, ArH), 3.78-3.71 (m, 2H, CH₂), 0.83-0.79(m, 3H, CH₃).¹³C NMR (100 MHz, DMSO-d₆):δ 179.2, 167.5, 159.2, 154.6, 152.4, 143.6, 137.2, 133.9, 128.1, 125.4, 125.3, 124.1, 123.4, 116.8, 113.1, 110.1, 75.5, 59.7, 56.5, 48.1, 31.2, 19.0. +ESI MS (m/z): 438.06 [M]⁺

(4c)⁴: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 7.94-7.92(m, 1H, ArH), 7.76-7.72 (m, 3H, NH₂, ArH), 7.54-7.42 (m, 4H, ArH), 7.29-7.17(m, 5H, ArH), 6.97-6.94 (m, 1H, ArH), 6.77-6.75 (d, J=8MHz,1H, ArH), 4.98-4.88 (m, 2H, CH₂). ¹³C NMR (100 MHz, DMSO-d₆): δ 176.4, 159.1, 158.9, 155.8, 152.6, 143.2, 136.4, 134.3, 132.7, 129.5, 128.9, 127.7, 127.6, 125.6, 124.6, 123.4, 123.2, 117.5, 117.2, 113.0, 109.6, 101.6, 57.2, 56.5, 19.0. +ESI MS (m/z): 448.12 [M+H]⁺

(4d)⁴: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 8.14(s, 2H, NH₂), 8.00-7.98 (d, J=7.2MHz,1H, ArH), 7.71-7.68 (m, 1H, ArH), 7.50-7.46 (m, 1H, ArH), 7.41-7.39 (d, J=8MHz,1H, ArH), 7.20-7.16 (d, J=7.6MHz, 1H, ArH), 7.05-7.03 (d, J=6.8MHz,1H, ArH), 6.90-6.82 (m, 2H, ArH), 3.69-3.64 (m, 2H, CH₂), 3.12 (m, 3H, N-CH₃), 0.73-0.69 (t, J=7.2MHz, 3H, CH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 199.7, 167.5, 159.2, 158.3, 152.4, 145.8, 134.2, 134.0, 128.6, 125.4, 123.5, 123.4, 122.3, 116.9, 112.9, 107.7, 104.1, 75.6, 59.5, 26.8, 14.0. +ESI MS (m/z): 418.11 [M]⁺

(4e)⁴: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 10.52 (s, 1H, NH), 7.91-7.89 (d, J=8MHz,1H, ArH), 7.75-7.71 (m, 1H, ArH), 7.60 (s, 2H, NH₂), 7.52-7.44 (m, 2H, ArH), 6.98-6.96 (d, J=11.2MHz, 2H, ArH), 6.70-6.68 (d, J=7.6MHz,1H,ArH), 2.15 (s, 3H, CH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 177.6, 158.9, 158.7, 155.5, 152.5, 140.2, 134.1, 133.6, 131.4, 129.6, 125.5, 125.2, 123.1, 117.5, 117.2, 113.0, 109.7, 102.0, 57.7, 21.0.+ESI MS (m/z): 372.09 [M+H]⁺

(4f)⁴: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 7.92-7.90 (t, 1H, ArH), 7.73-7.69 (m, 3H, -NH₂, ArH), 7.51-7.44 (m, 2H, ArH), 7.27-7.22 (m, 2H, ArH), 6.99-6.90 (m, 2H, ArH), 5.39-5.35 (m, 1H, =CH₂), 5.14-5.12 (d,d 1H, =CH₂), 4.39-4.25 (m, 1H, CH), 3.29 (s, 2H, CH). ¹³C NMR (100 MHz, DMSO-d₆): δ 175.9, 159.0, 158.8, 155.7, 152.6, 143.2, 134.3, 132.7, 132.0, 129.5, 125.5, 124.5, 123.3, 123.2, 117.4, 117.2, 112.9, 109.6, 101.7, 57.2, 47.8, 42.6. +ESI MS (m/z): 397.10 [M]⁺

(4g)⁴: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 7.94-7.90 (m, 1H, ArH), 7.72-7.68 (m, 3H, NH₂, ArH), 7.50-7.45 (m, 2H, ArH), 7.26-7.20 (m, 2H, ArH), 6.98-6.92 (m, 2H, ArH), 2.15 (s, 3H, CH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 175.8, 159.1, 155.6, 152.6, 143.1, 132.6, 132.1, 129.4, 125.5, 124.4, 123.2, 123.1, 117.1, 109.5, 101.7, 57.1, 47.4, 37.4. +ESI MS (m/z): 371.09 [M]⁺

(4h)⁵: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 8.17 (s, 2H, NH₂), 8.02-7.97 (d, J=7.2MHz, 1H, ArH), 7.72-7.69 (m, 1H, ArH), 7.51-7.45(m, 1H, ArH), 7.42-7.38 (d, J=8MHz, 1H, ArH), 7.21-7.17 (d, J=7.6MHz, 1H, ArH), 7.04-7.02 (d, J=6.8MHz, 1H, ArH), 6.91-6.81 (m, 2H, ArH), 5.78 (s, 1H, CH), 4.51-4.32 (m, 2H, CH₂), 3.68-3.65 (m, 2H, CH₂), 2.35-2.21 (m, 2H, CH₂), 0.72-0.68 (t, J=7.2MHz, 3H, CH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 199.7, 167.4, 159.1, 158.2, 152.6, 145.9, 134.3, 134.1, 131.5, 128.7, 125.5, 123.6, 123.3, 122.2, 116.7, 115.6, 112.8, 107.6, 104.1, 75.8, 64.6, 59.6, 14.2. +ESI MS (m/z): 418.11 [M]⁺

(4i)⁵: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 10.55 (s, 1H, NH), 8.15 (s, 2H, NH₂), 8.16-7.99 (d, J=7.2MHz, 1H, ArH), 7.72-7.69 (m, 1H, ArH), 7.50-7.44(m, 1H, ArH), 7.40-7.36 (d, J=8MHz, 1H, ArH), 7.25-7.15 (d, J=7.6MHz, 1H, ArH), 7.04-7.02 (d, J=6.8 MHz, 1H, ArH), 6.91-6.81 (m, 2H, ArH), 3.68-3.63 (m, 2H, CH₂), 0.72-0.66 (t, J=7.2 MHz, 3H, CH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 199.3, 167.33, 159.1, 158.3, 152.2, 145.9, 134.3, 134.1, 128.5, 125.2, 123.5, 123.2, 122.1, 116.4, 112.9, 107.6, 104.2, 75.4, 59.4, 14.0. +ESI MS (m/z): 405.10 [M+H]⁺

(4j)⁵: ¹H-NMR(DMSO-d₆ 400 MHz); δ (ppm): 10.61 (s, 1H, NH), 7.85 (m, 2H, ArH), 7.69 (m, 2H, ArH), 7.49 (s, 2H, NH₂), 7.37 (d, 1H, J=7.6MHz, ArH), 7.19 (d, J=7.7Hz, 1H, ArH), 6.99 (s, 1H, ArH). ¹³C NMR 100 MHz, DMSO-d₆): δ 175.9, 160.3, 158.3, 156.6, 145.9, 140.5, 133.4, 130.9, 126.7, 129.2, 125.5, 123.3, 120.8, 119.3, 117.2, 114.1, 112.4, 102.5, 59.4, 48.8. +ESI MS (m/z): 391.03 [M]⁺

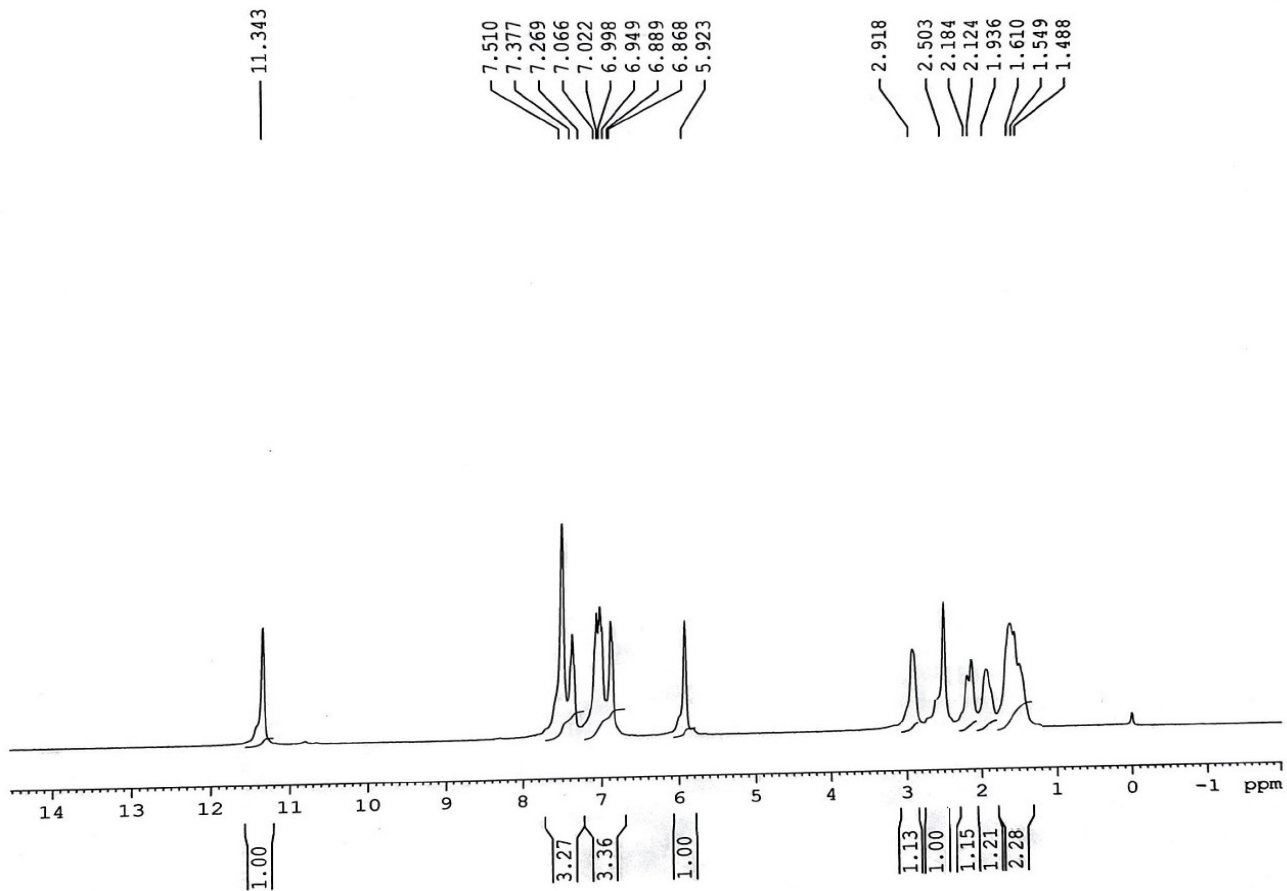
(4k)⁶: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 10.21 (s, 1H, NH), 8.01 (1H, d, J=7.9MHz, ArH), 7.99 (s, 2H, NH₂), 7.67 (d, J=7.6MHz, 1H, ArH), 7.50 (d, J=7.8MHz, 1H, ArH), 7.29 (t, J=7.2MHz, 1H, ArH), 7.10 (m, 2H, ArH), 6.90 (d, J=7.2MHz, 1H, ArH), 3.70 (s, 3H, OCH₃). ¹³C NMR (100 MHz, DMSO-d₆): δ 178.7, 160.4, 158.3, 156.2, 152.1, 145.9, 140.9, 133.3, 130.2, 125.9, 125.1, 119.6, 118.4, 117.4, 114.4, 112.9, 111.4, 101.3, 61.2, 57.9, 49.1. +ESI MS (m/z): 387.08 [M]⁺

(4l)⁶: ¹H-NMR (DMSO-d₆ 400 MHz); δ (ppm): 10.21 (s, 1H, NH), 7.90 (s, 2H, NH₂), 7.54 (d, J=8.0MHz, 1H, ArH), 7.49 (m, 2H, ArH), 7.28 (t, J=7.5MHz, 1H, ArH), 7.09 (m, 2H, ArH). ¹³C NMR (100MHz, DMSO-d₆): δ 176.5, 158.9, 158.7, 157.0, 146.7, 141.4, 140.8, 132.7, 131.2, 126.7, 125.9, 122.8, 122.3, 120.1, 116.0, 114.1, 109.2, 102.6, 61.8, 49.3. +ESI MS (m/z): 403.06 [M+H]⁺

References

- (1). Hegade, P. G., Chinchkar, S. D., & Pore, D. M. (2016). DABCO catalyzed pseudo multi-component synthesis of functionalized spirooxindoles. *Monatshefte für Chemie-Chemical Monthly*, 147(7), 1243-1249.
- (2). H. Yongke, C. Lei, L. Bindong, *RSC Adv.*, 2013, 00, 1-3.
- (3). K. L. Joydev, K. S. S. Tummalapalli, P. J. Krupal, *Org. Biomol. Chem.*, 2016, 14, 2473-2479.
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C 1



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PROCNO    1

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PULPROG   zg30
TD         65536
SOLVENT   DMSO
NS         16
DS         2
SWH        6172.839 Hz
FIDRES     0.094190 Hz
AQ         5.3084660 sec
RG         203.2
DN         91.000 usec
DE         6.50 usec
TE         300.0 K
D1         1.00000000 sec
TD0        1

----- CHANNEL f1 -----
NUC1       1H
P1         10.70 usec
PL1        -2.00 dB
SFO1       300.1318534 MHz

F2 - Processing parameters
SI         32768
SF         300.1299976 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
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Figure 1: ¹H-NMR spectrum of (6a)

C1

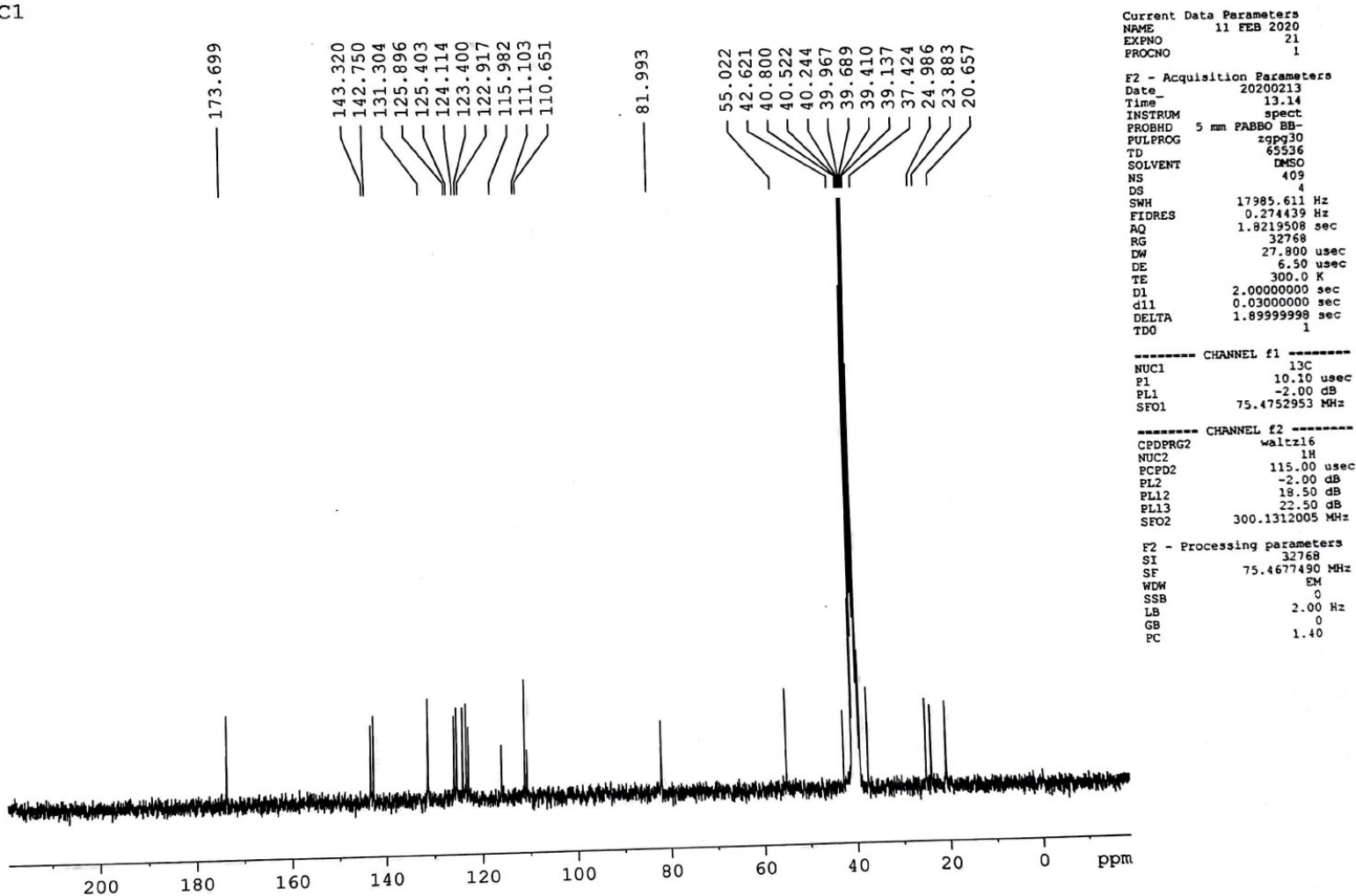
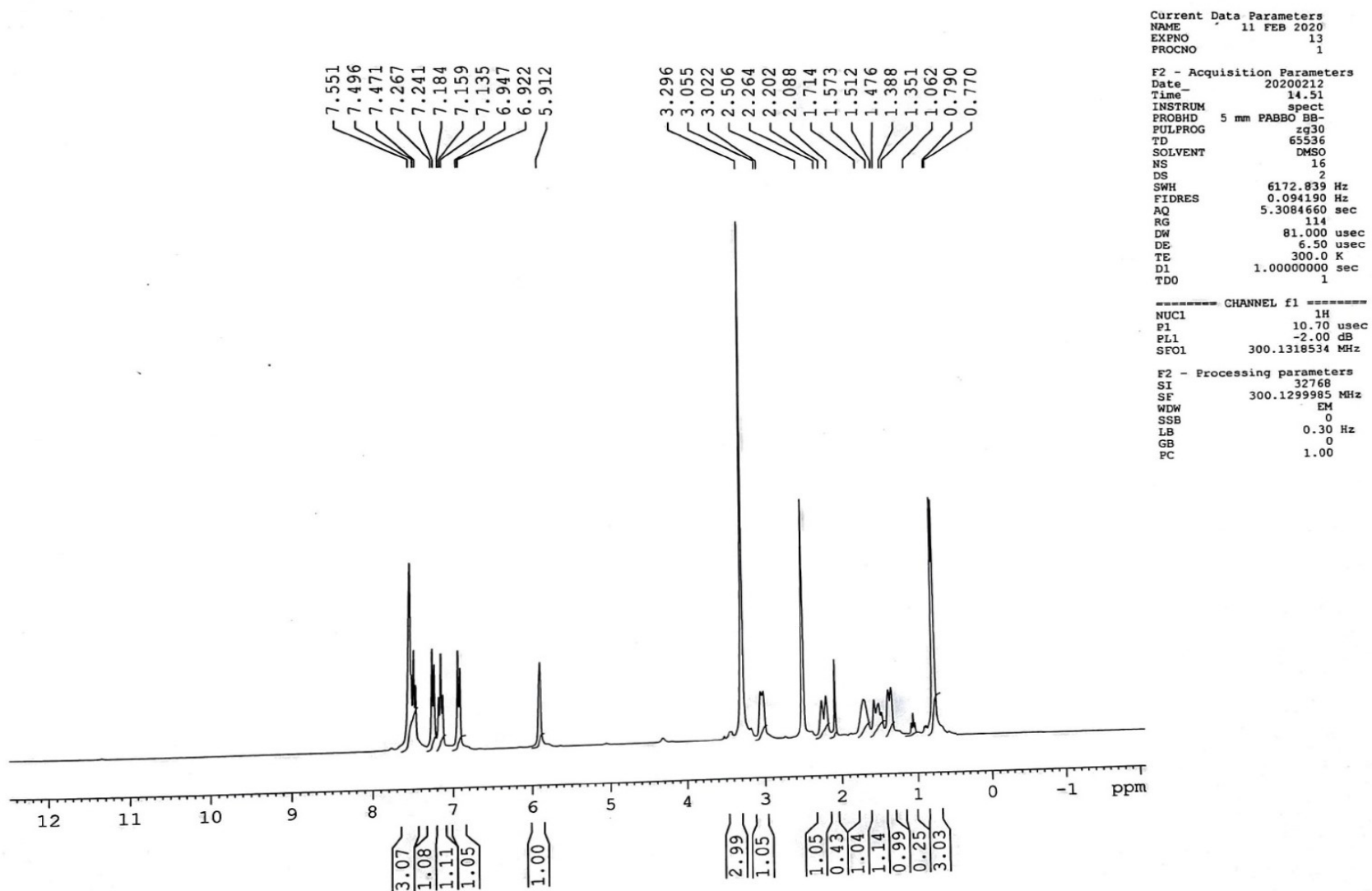


Figure 2: ^{13}C NMR spectrum of (6a)

Figure 3: ¹H-NMR spectrum of (6b)

C2

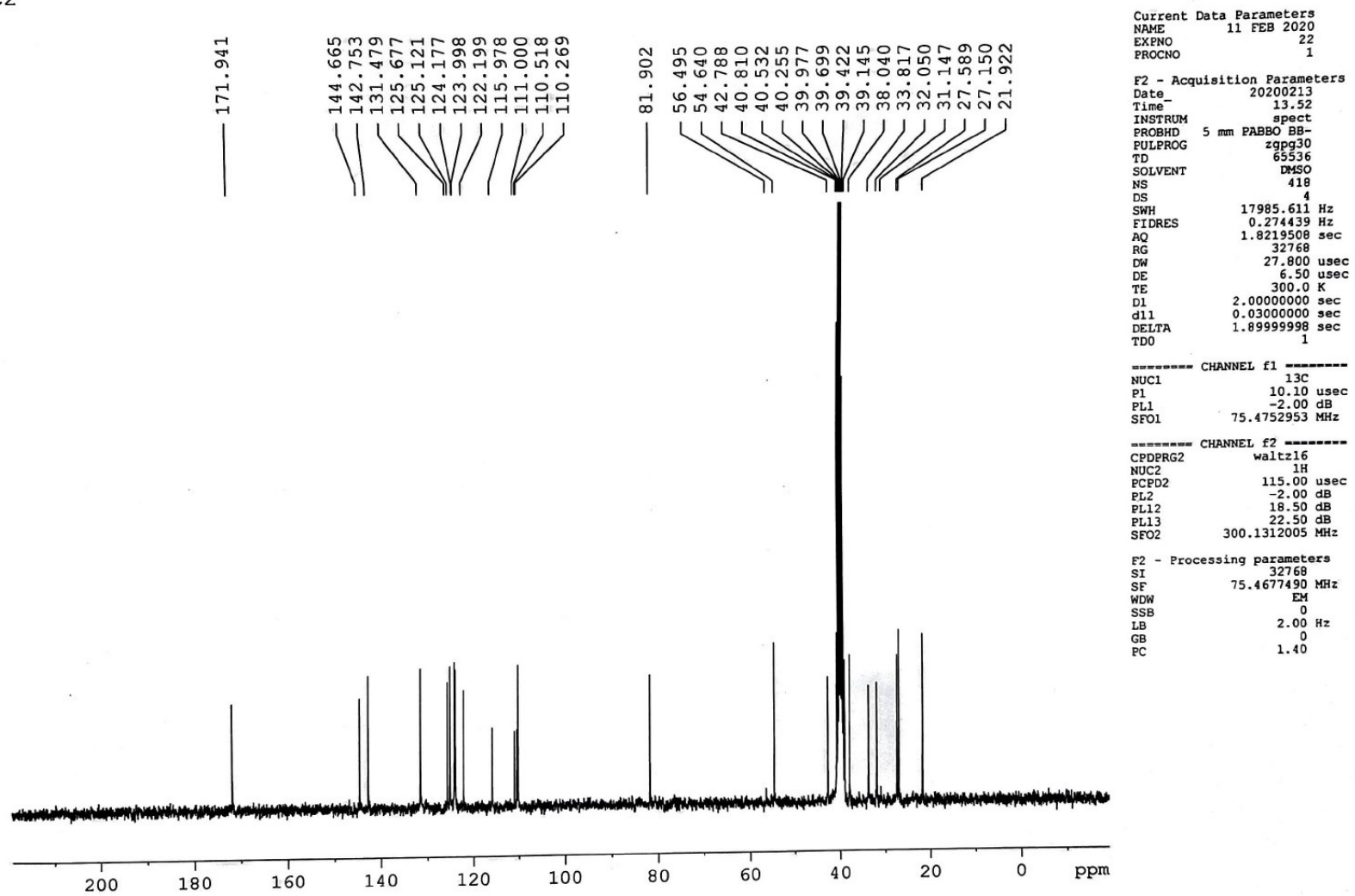
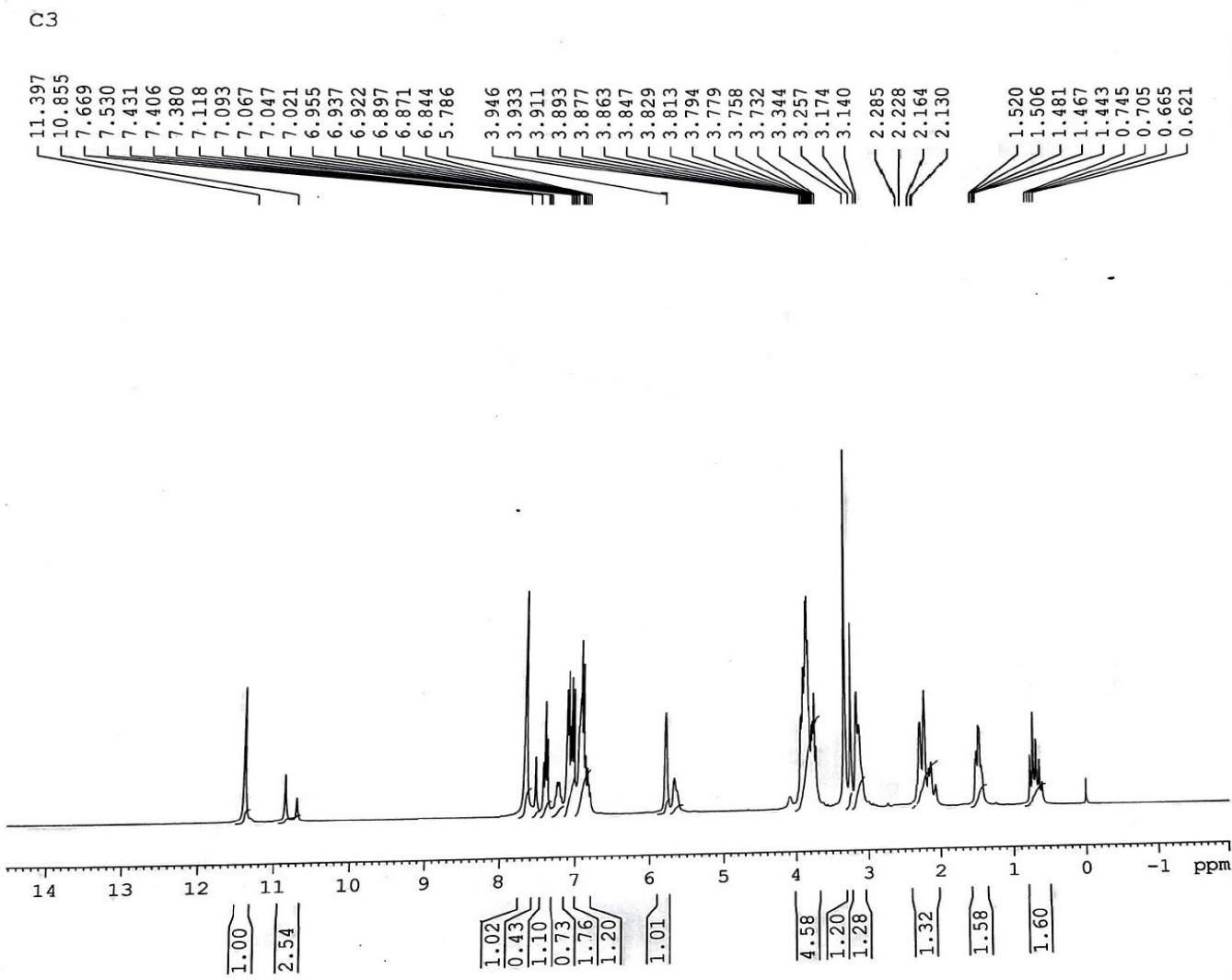


Figure 4: ¹³C NMR spectrum of (6b)



```

Current Data Parameters
NAME      11 FEB 2020
EXPNO    15
PROCNO   1

F2 - Acquisition Parameters
Date_    20200212
Time     14.58
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       11
DS       2
SWH      6172.839 Hz
FIDRES   0.094190 Hz
AQ       5.3084660 sec
RG       203.2
DW       81.000 usec
DE       6.50 usec
TE       300.0 K
D1       1.00000000 sec
TD0      1

----- CHANNEL f1 -----
NUC1     1H
P1       10.70 usec
PL1      -2.00 dB
SFO1     300.1318534 MHz

F2 - Processing parameters
SI       32768
SF       300.1299977 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

```

Figure 5: ¹H-NMR spectrum of (6c)

C3

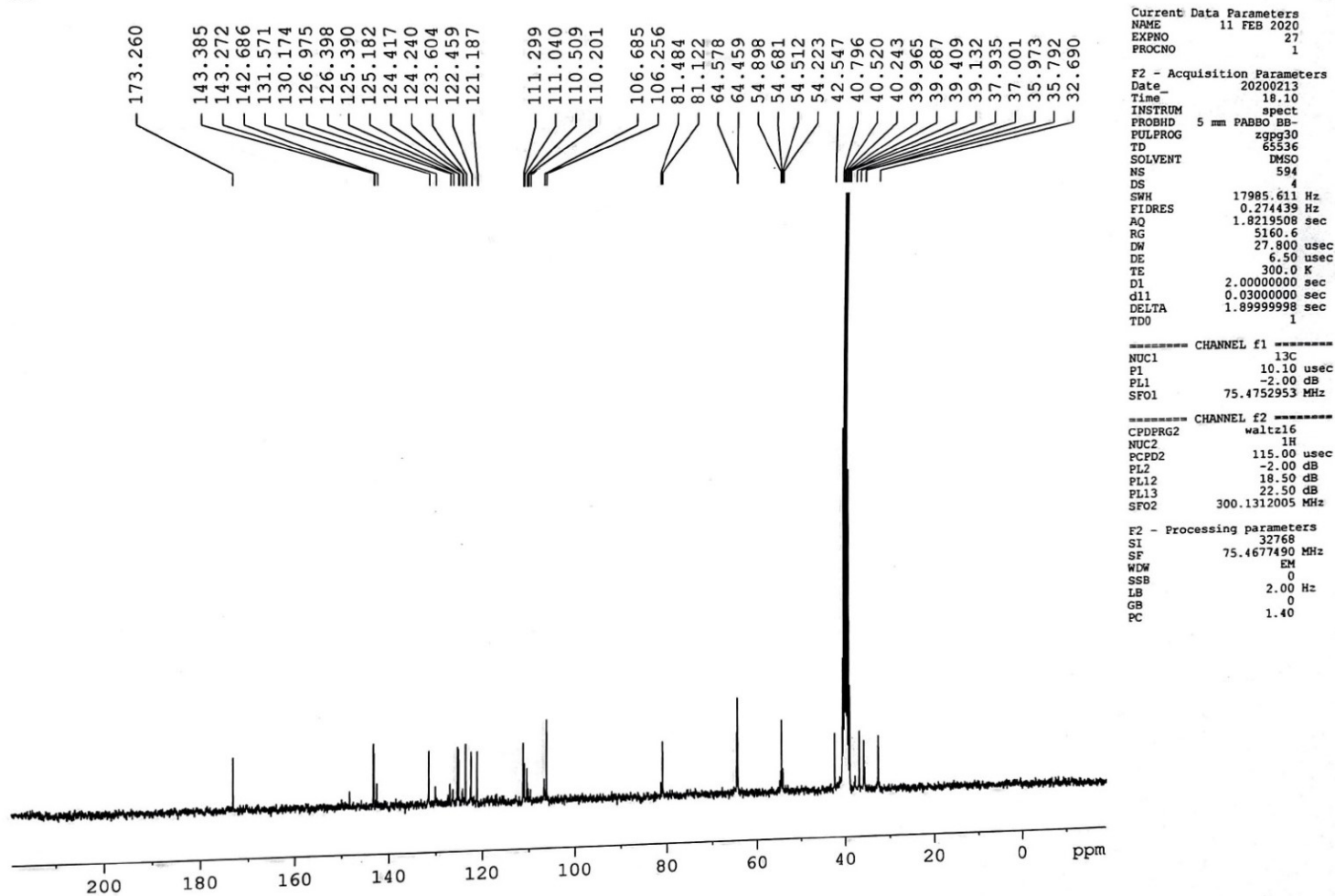


Figure 6: ¹³C NMR spectrum of (6c)

C4

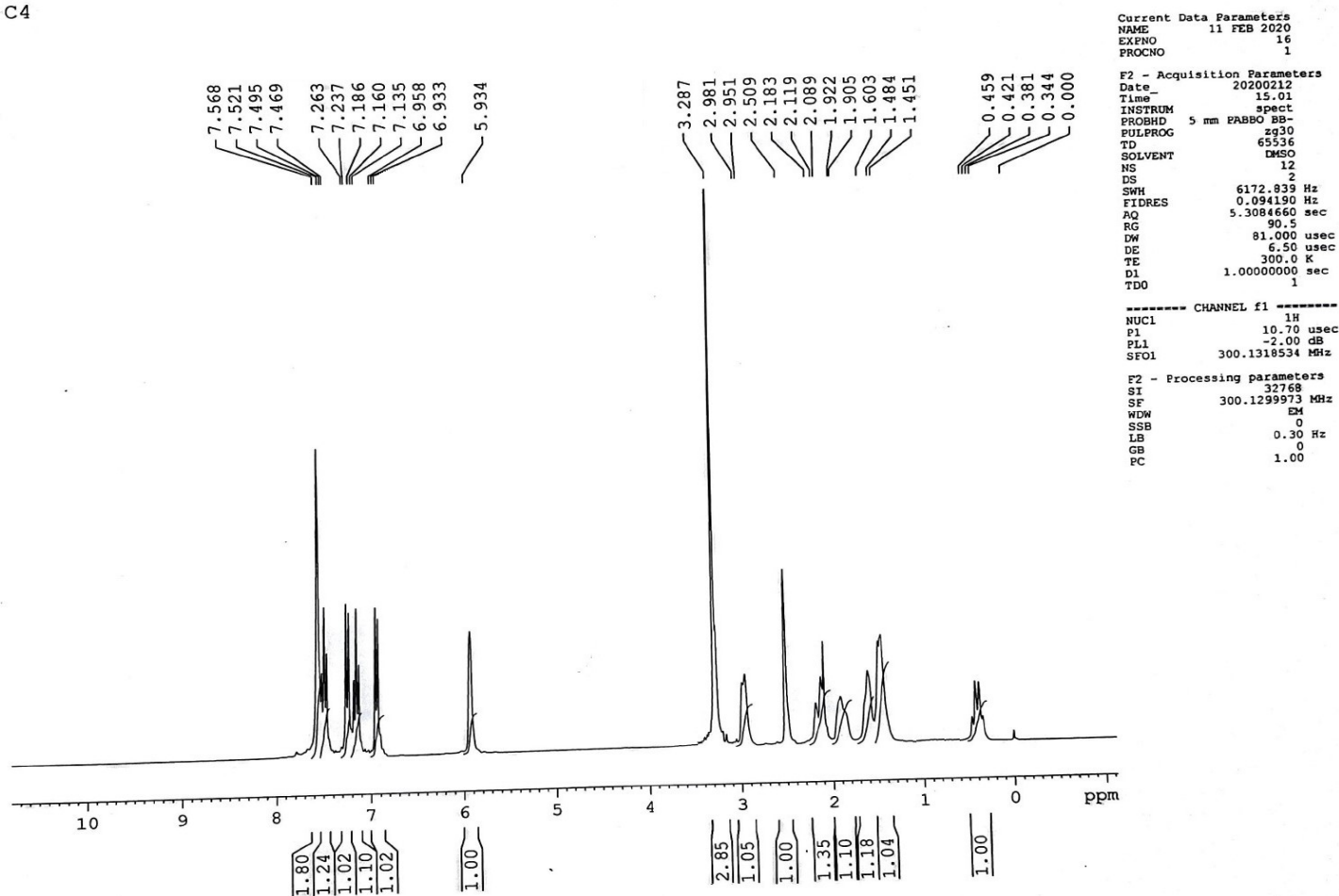


Figure 7: ¹H-NMR spectrum of (6d)

C 4

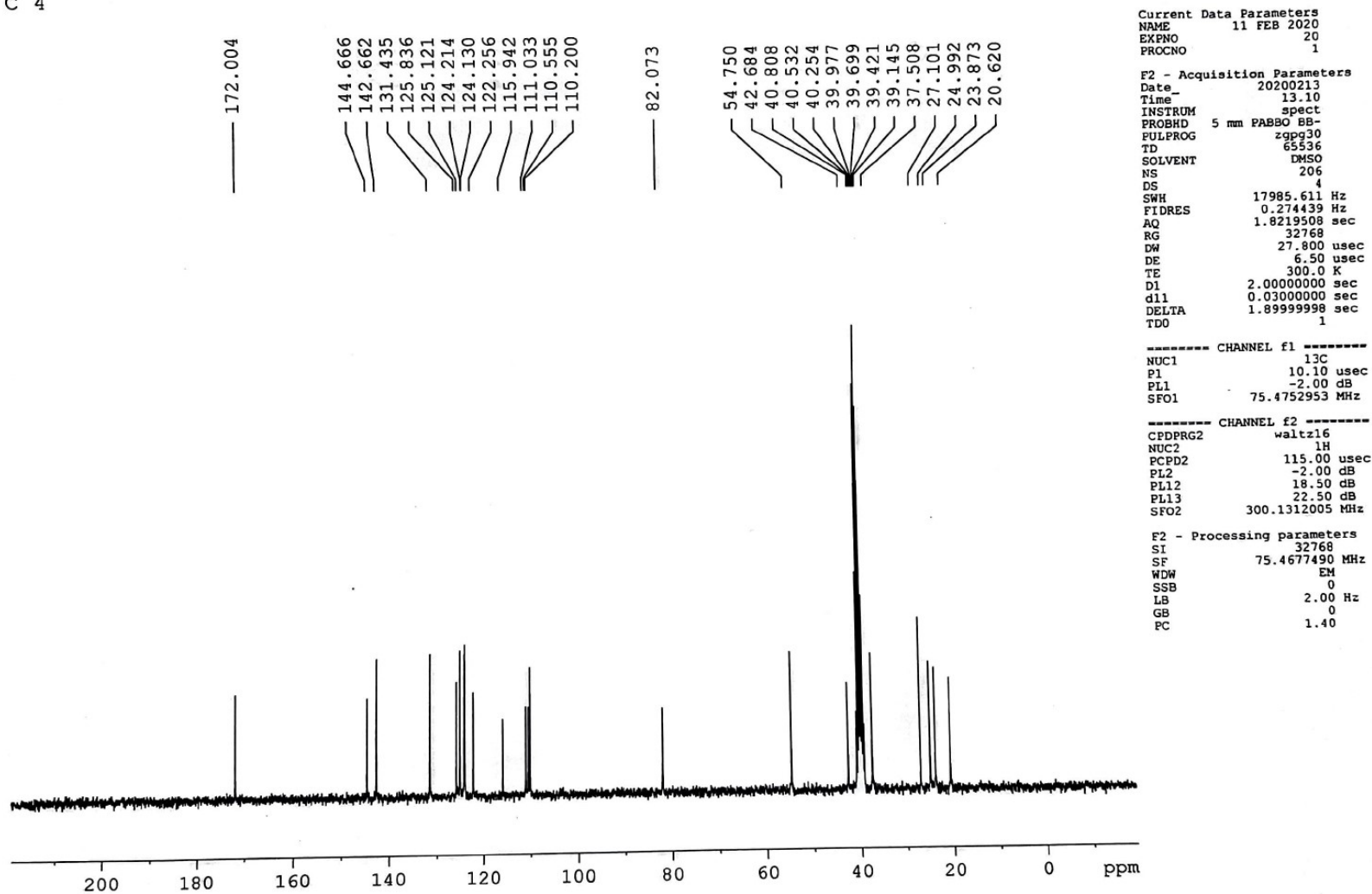
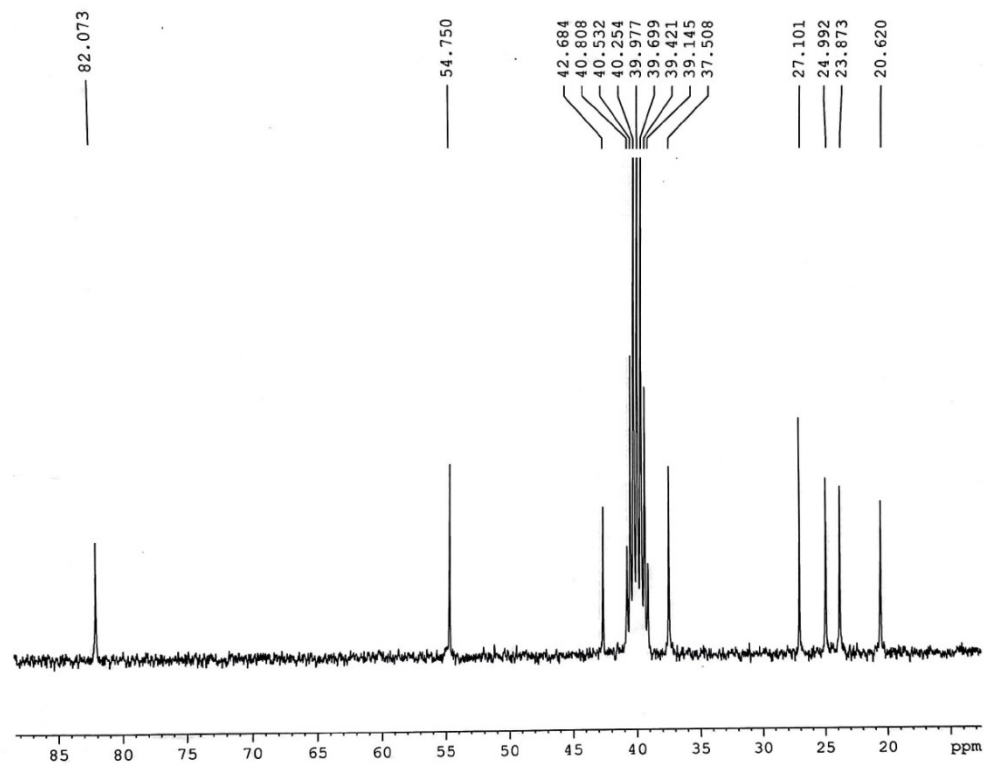


Figure 8: ^{13}C NMR spectrum of (6d)

C 4



```
Current Data Parameters
NAME      11 FEB 2020
EXPNO    20
PROCNO    1

F2 - Acquisition Parameters
Date_     20200213
Time      13.10
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD         65536
SOLVENT   DMSO
NS         206
DS         4
SWH        17985.611 Hz
FIDRES     0.274439 Hz
AQ         1.8219508 sec
RG         32768
DW         27.800 usec
DE         6.50 usec
TE         300.0 K
D1         2.0000000 sec
d11        0.0300000 sec
DELTA     1.8999999 sec
TDO        1

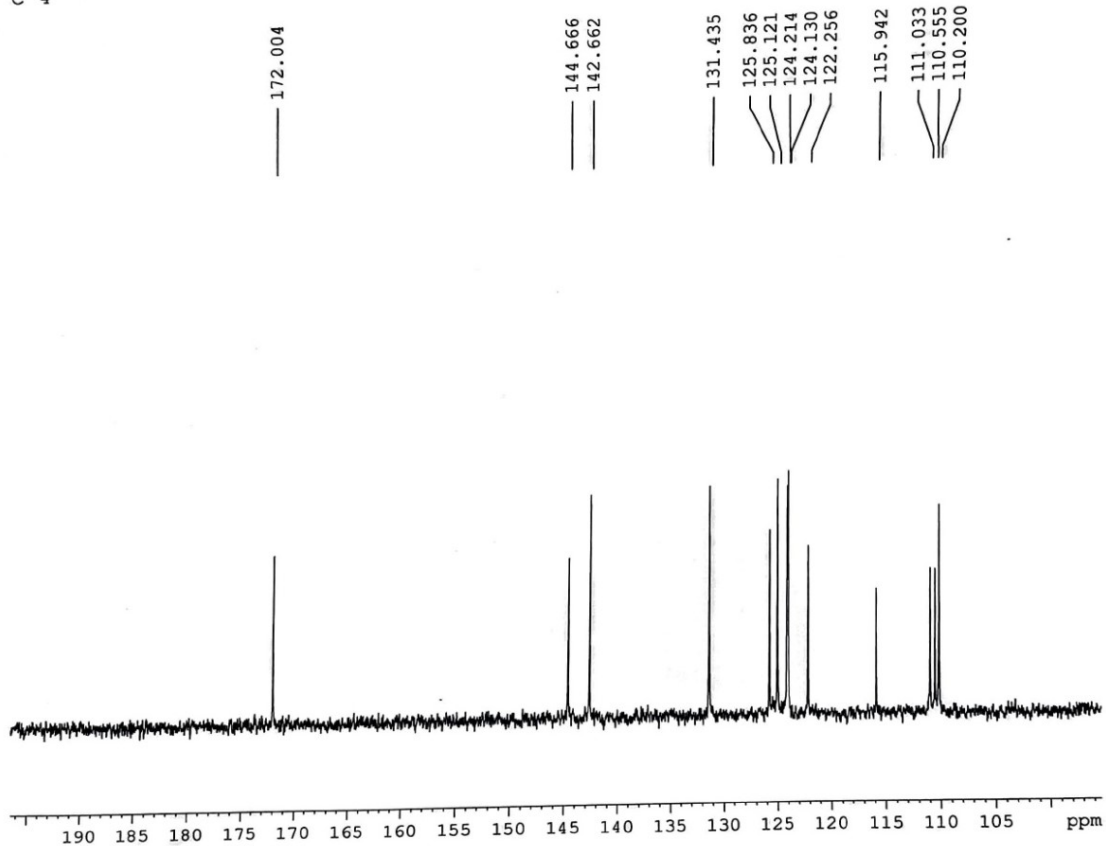
----- CHANNEL f1 -----
NUC1       13C
P1         10.10 usec
PL1        -2.00 dB
SFO1       75.4752953 MHz

----- CHANNEL f2 -----
CPOBPG2    waltz16
NUC2        1H
PCPD2      115.00 usec
PL2         -2.00 dB
PL12        18.50 dB
PL13        22.50 dB
SFO2       300.1312005 MHz

F2 - Processing parameters
SI         32768
SF         75.4677490 MHz
WDW        EM
SSB         0
LB         2.00 Hz
GB          0
PC         1.40
```

Figure 8a: ^{13}C NMR spectrum of (6d) Zoom

C 4



```
Current Data Parameters
NAME      11 FEB 2020
EXPNO    20
PROCNO   1

F2 - Acquisition Parameters
Date     20200213
Time     13.10
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  DMSO
NS       206
DS       4
SWH      17985.611 Hz
FIDRES   0.274439 Hz
AQ       1.8219508 sec
RG       32768
DW       27.800 usec
DE       6.50 usec
TE       300.0 K
D1       2.0000000 sec
d11      0.0300000 sec
DELTA    1.89999998 sec
TD0      1

----- CHANNEL f1 -----
NUC1     13C
P1       10.10 usec
PL1      -2.00 dB
SFO1     75.4752953 MHz

----- CHANNEL f2 -----
CPDPRG2  waltz16
NUC2     1H
PCPD2    115.00 usec
PL2      -2.00 dB
PL12     18.50 dB
PL13     22.50 dB
SFO2     300.1312005 MHz

F2 - Processing parameters
SI       32768
SF       75.4677490 MHz
WDW      EM
SSB      0
LB       2.00 Hz
GB       0
PC       1.40
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Figure 8b: ^{13}C NMR spectrum of (6d) Zoom

C5

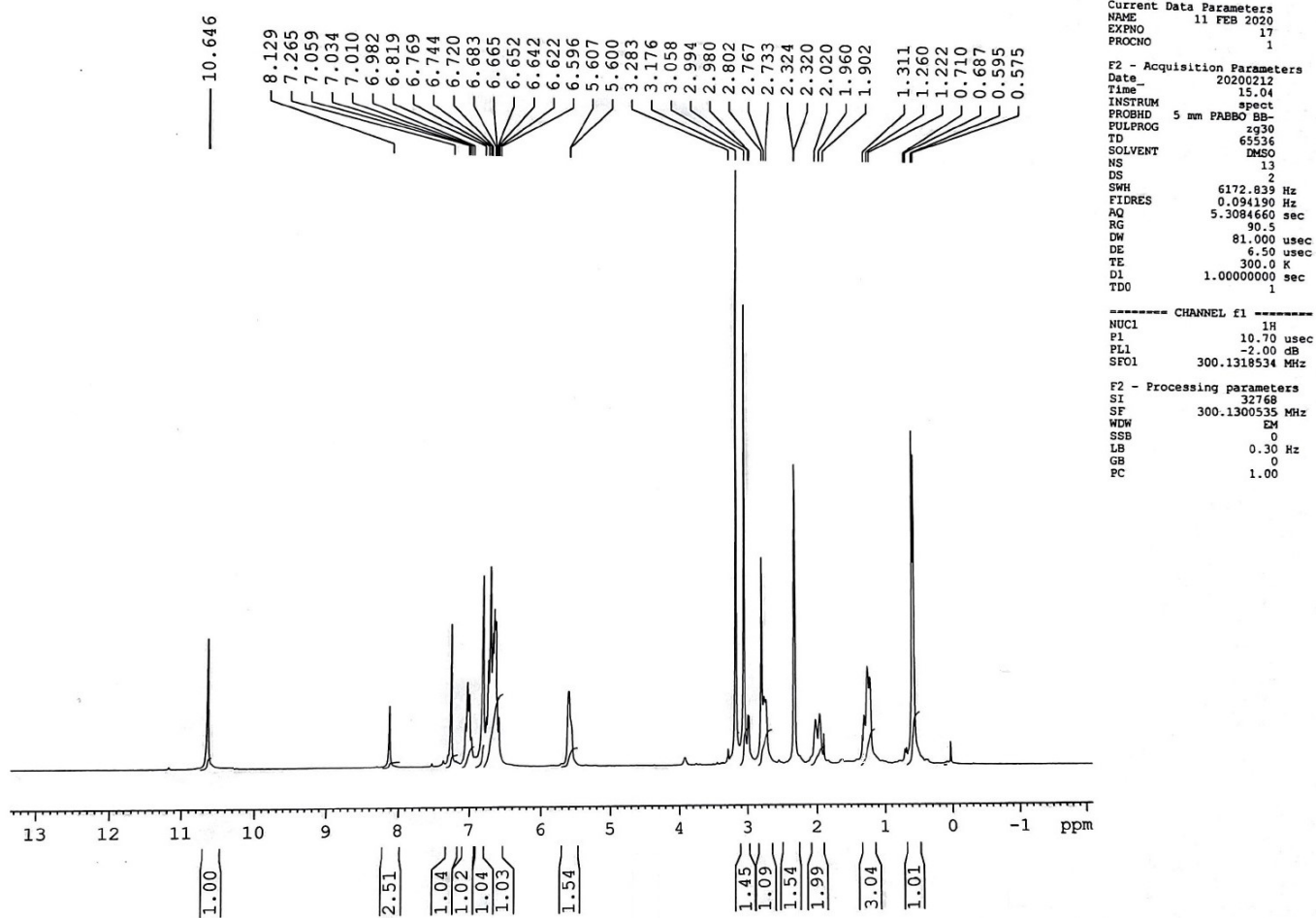


Figure 9: ¹H-NMR spectrum of (6e)

C 5

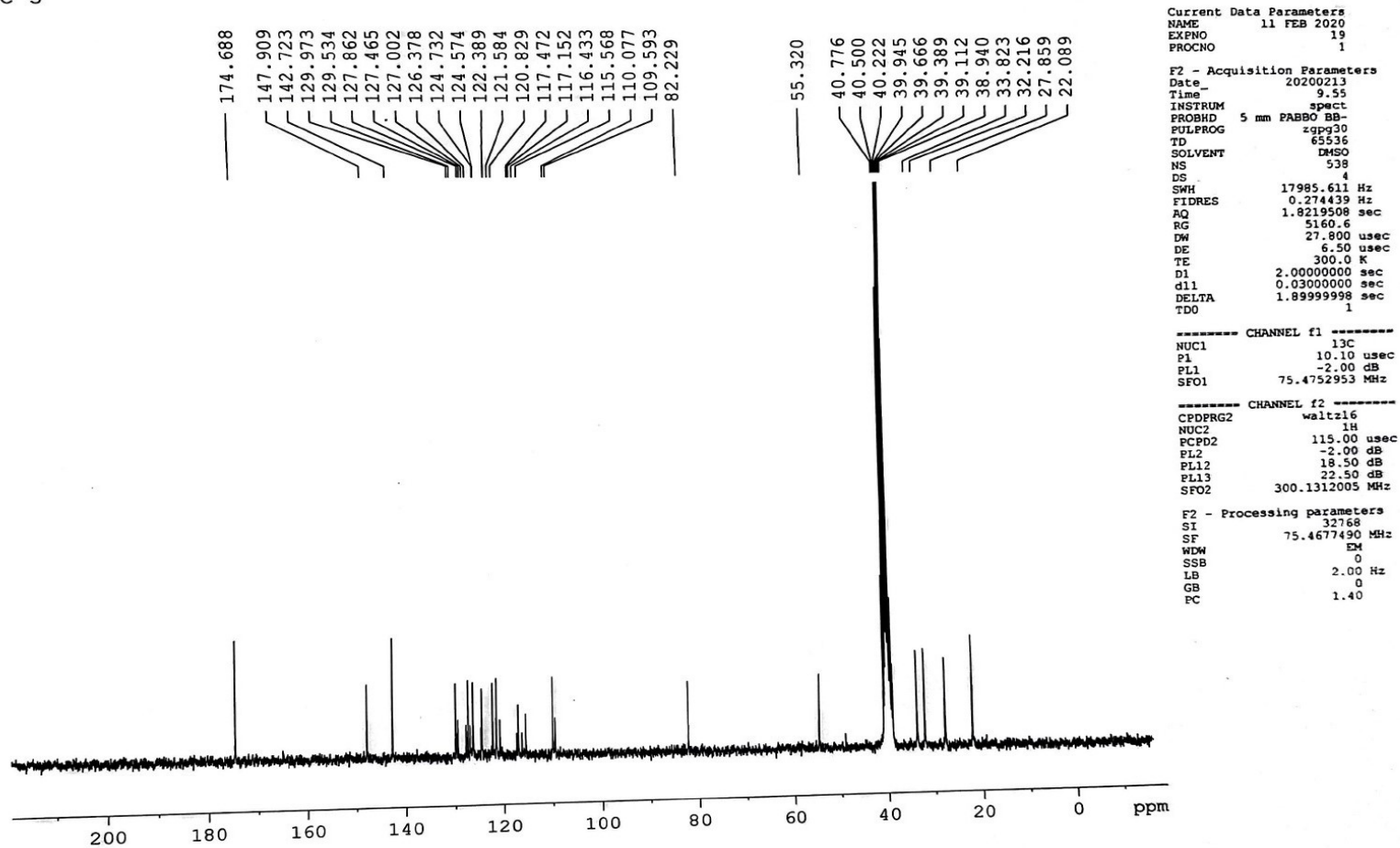
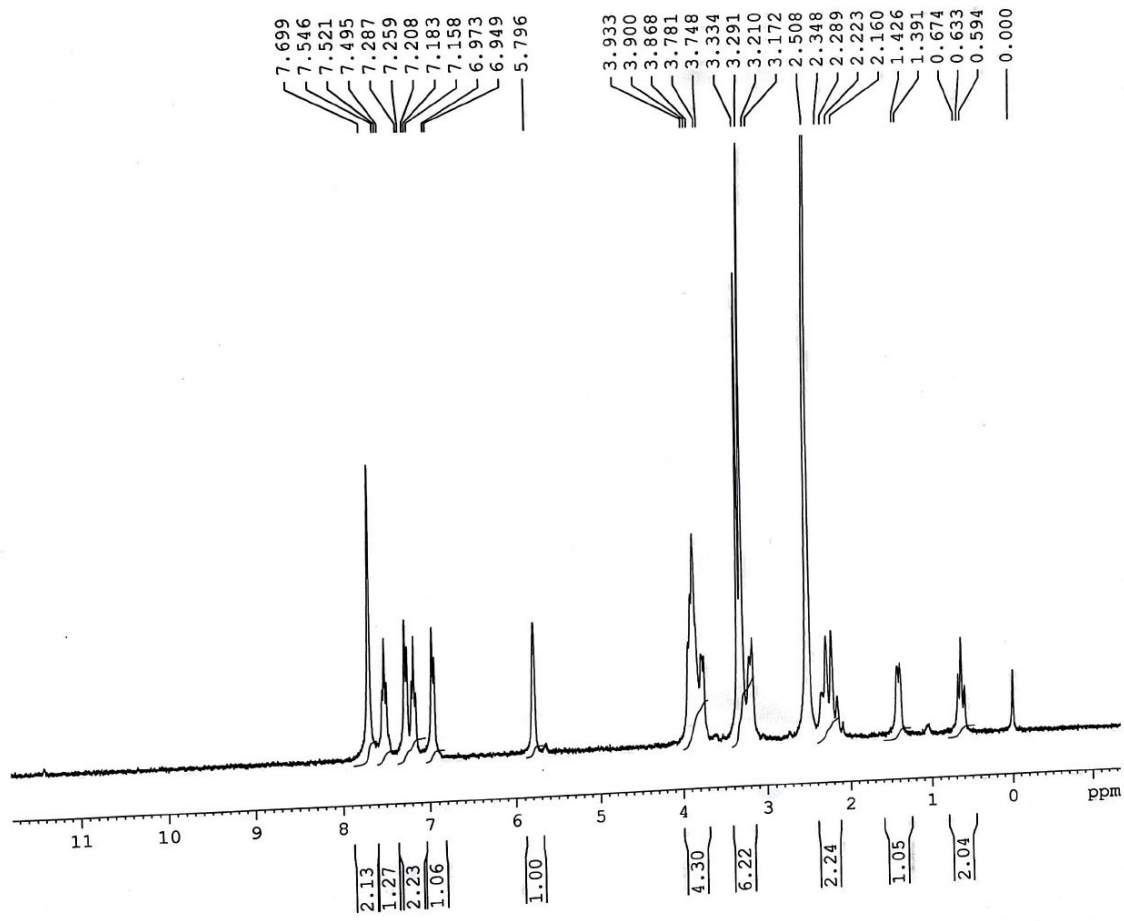


Figure 10: ^{13}C NMR spectrum of (6e)

C6



```

Current Data Parameters
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EXPNO    23
PROCNO   1

F2 - Acquisition Parameters
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Time     14.17
INSTRUM  spect
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PULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      6172.839 Hz
FIDRES   0.094190 Hz
AQ       5.3084660 sec
RG       4
DW       81.000 usec
DE       6.50 usec
TE       300.0 K
D1       1.0000000 sec
TDO     1

----- CHANNEL f1 -----
NUC1     1H
F1       10.70 usec
PL1     -2.00 dB
SFO1    300.1318534 MHz

F2 - Processing parameters
SI       32768
SF       300.1299975 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00
  
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Figure 11: ¹H-NMR spectrum of (6f)

C6

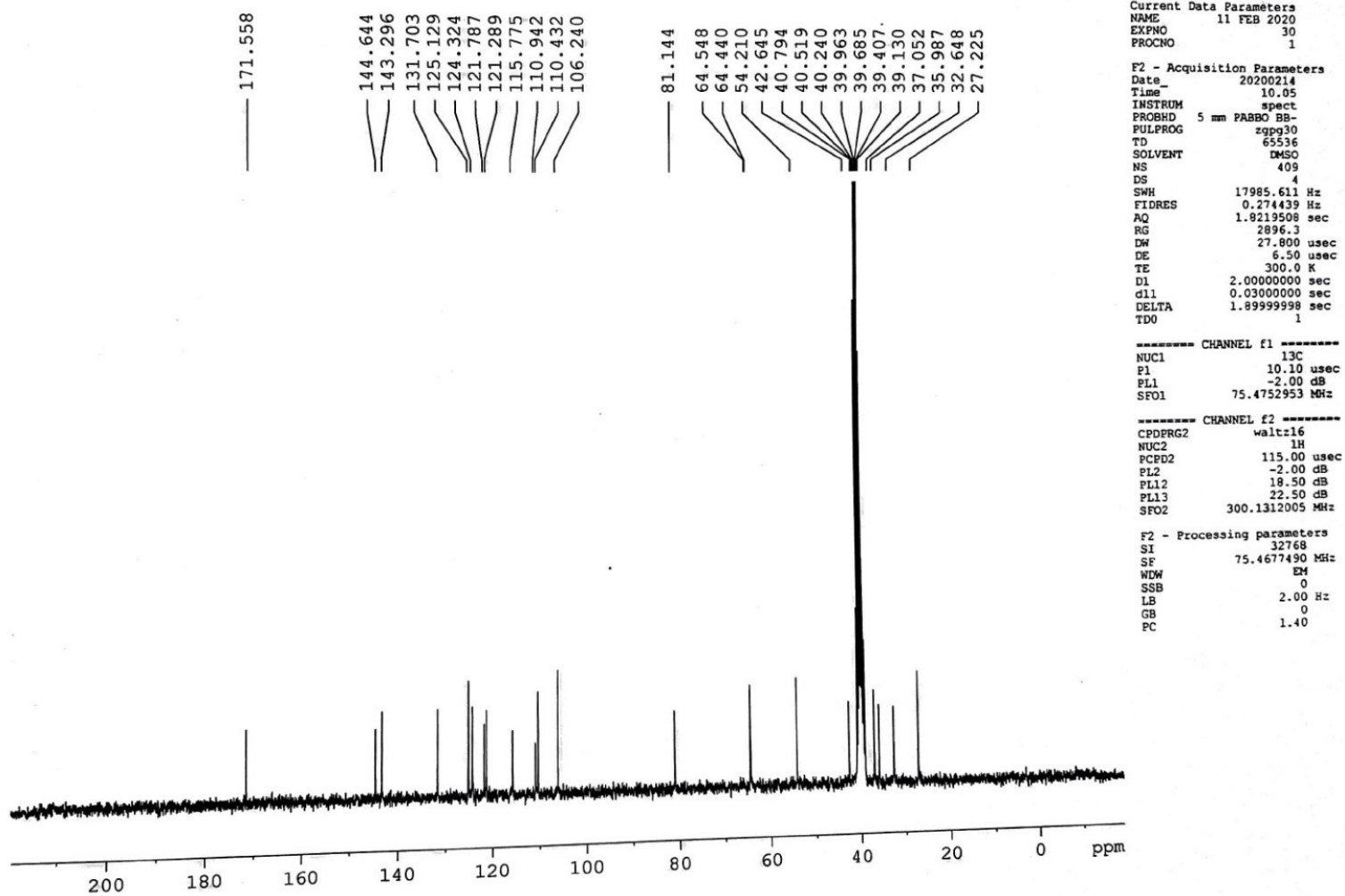


Figure 12: ^{13}C NMR spectrum of (6f)

C7

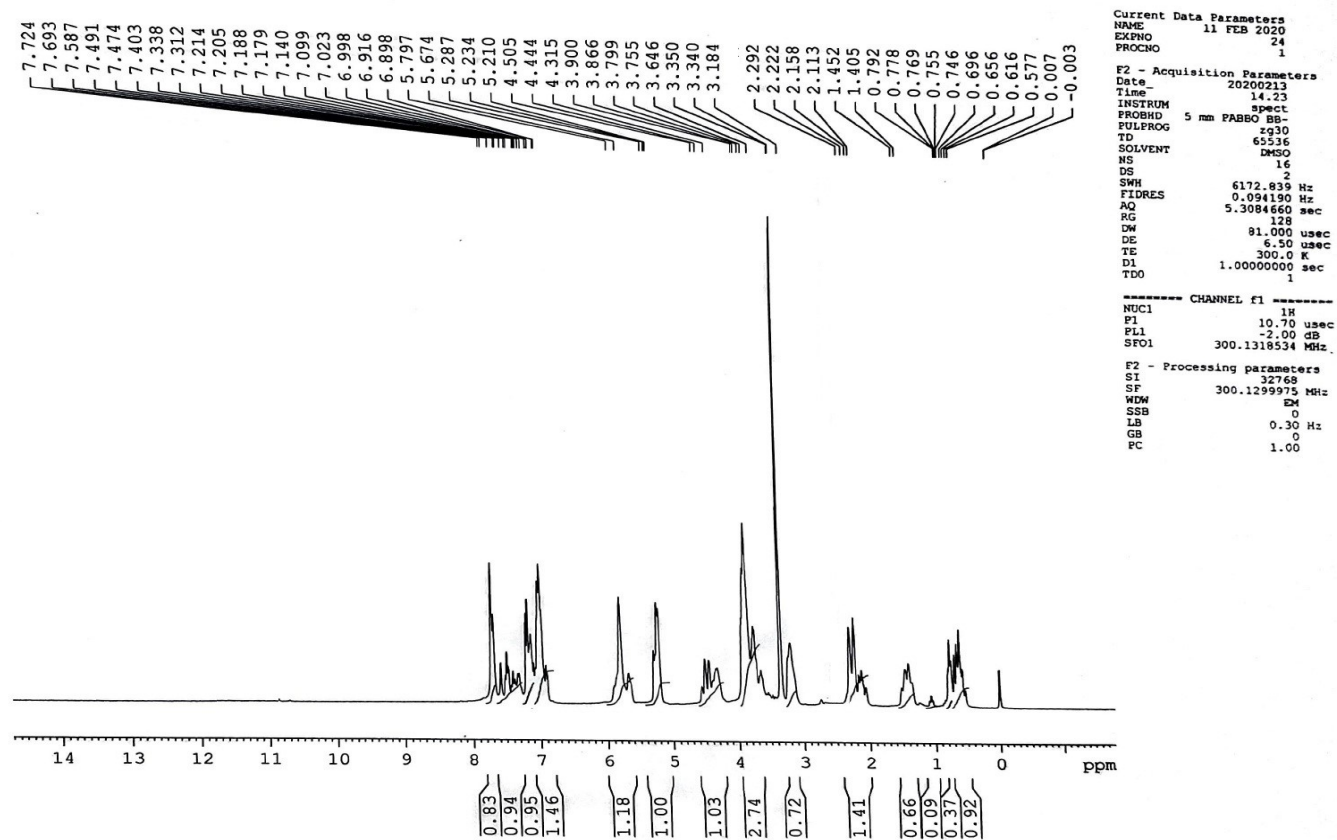


Figure 13: ¹H-NMR spectrum of (6g)

C7

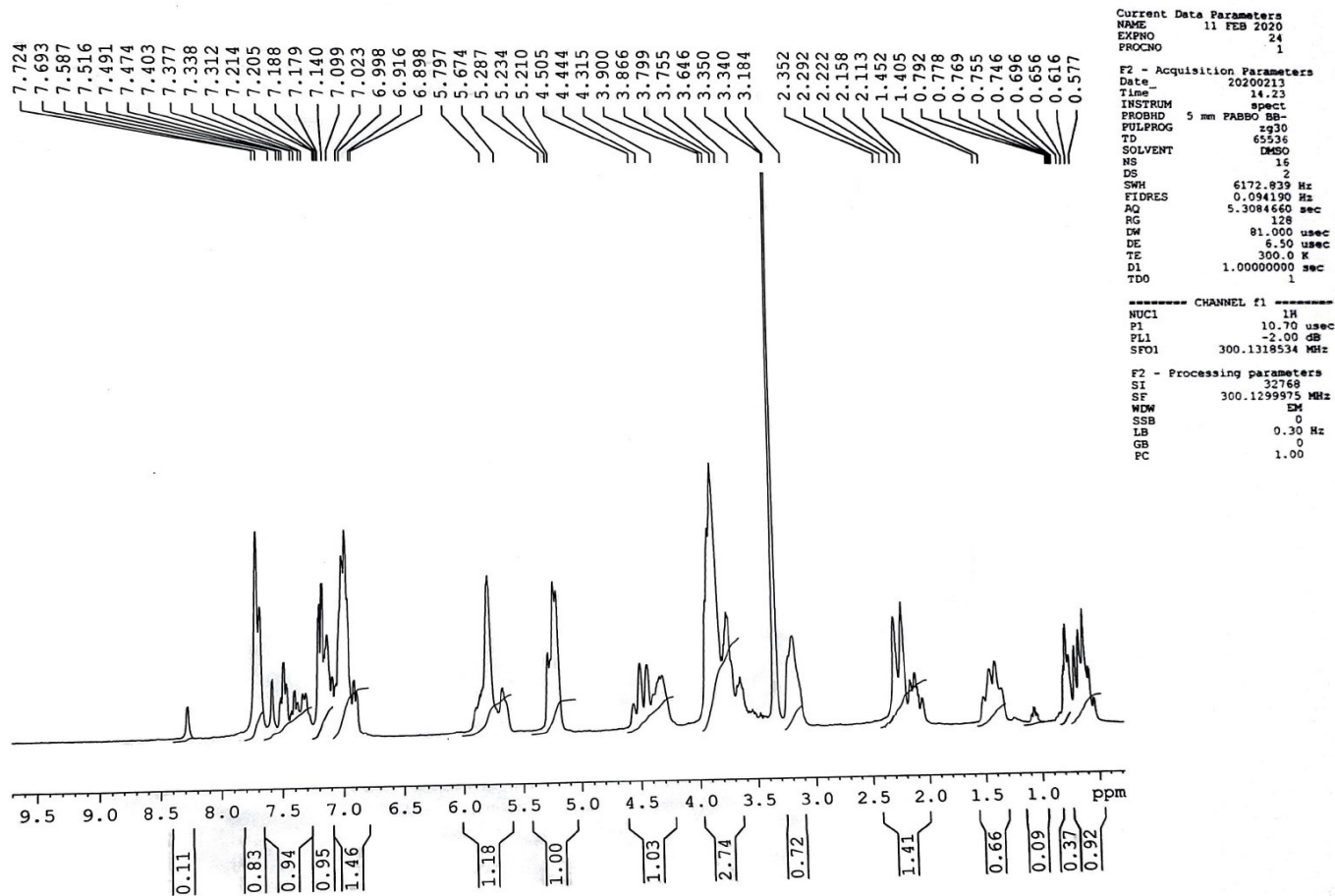


Figure 13A: ¹H-NMR spectrum of (6g) Zoom

C7

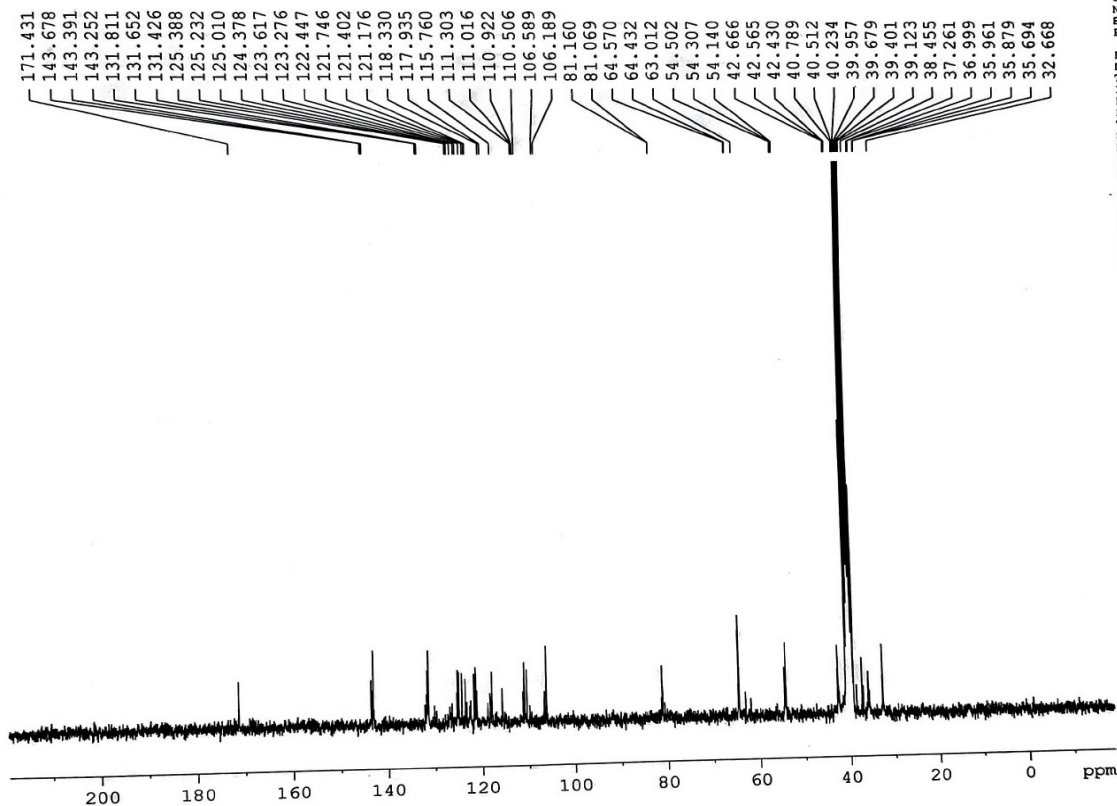


Figure 14: ¹³C NMR spectrum of (6g)

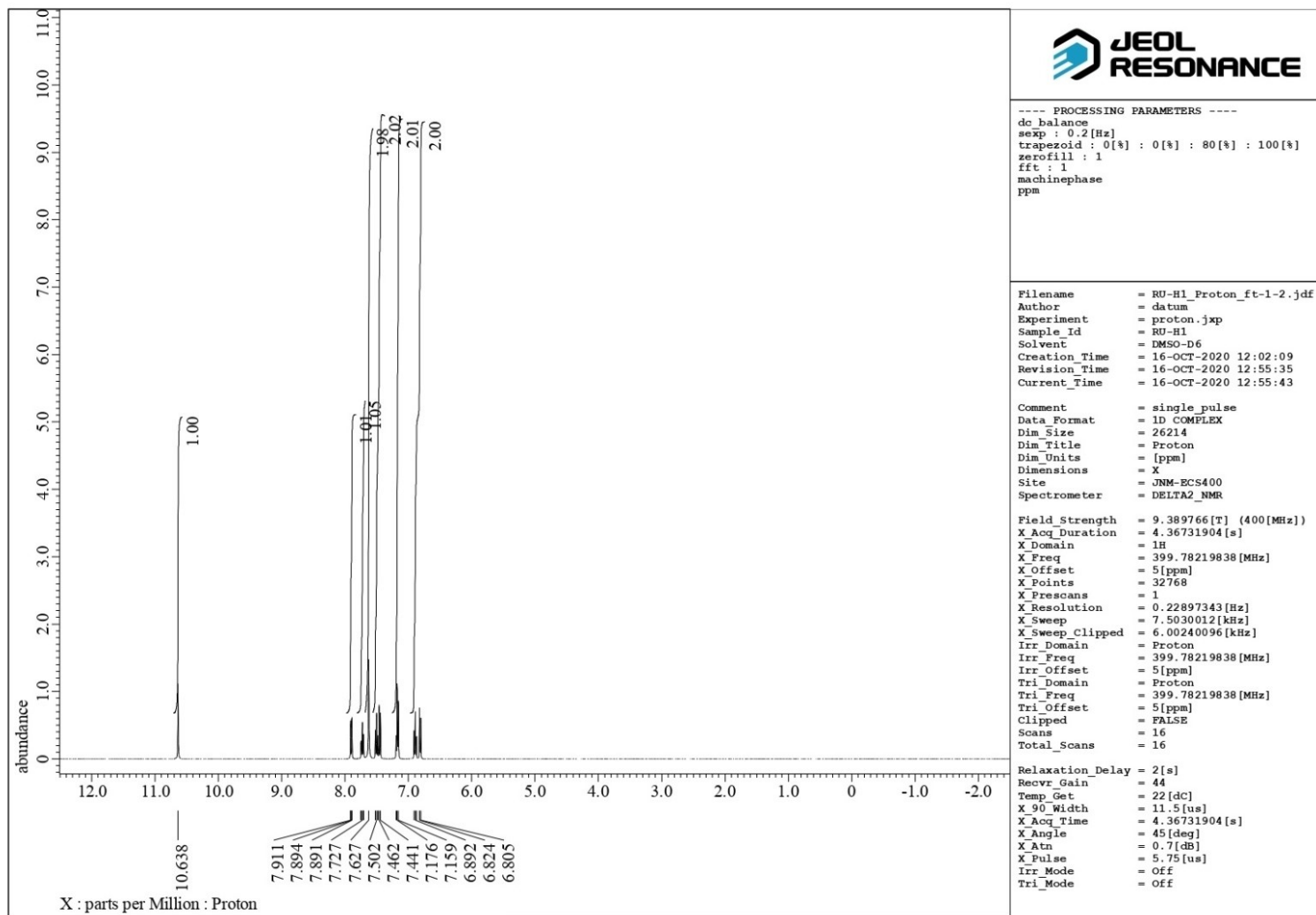


Figure 15: ^1H -NMR spectrum of (4a)

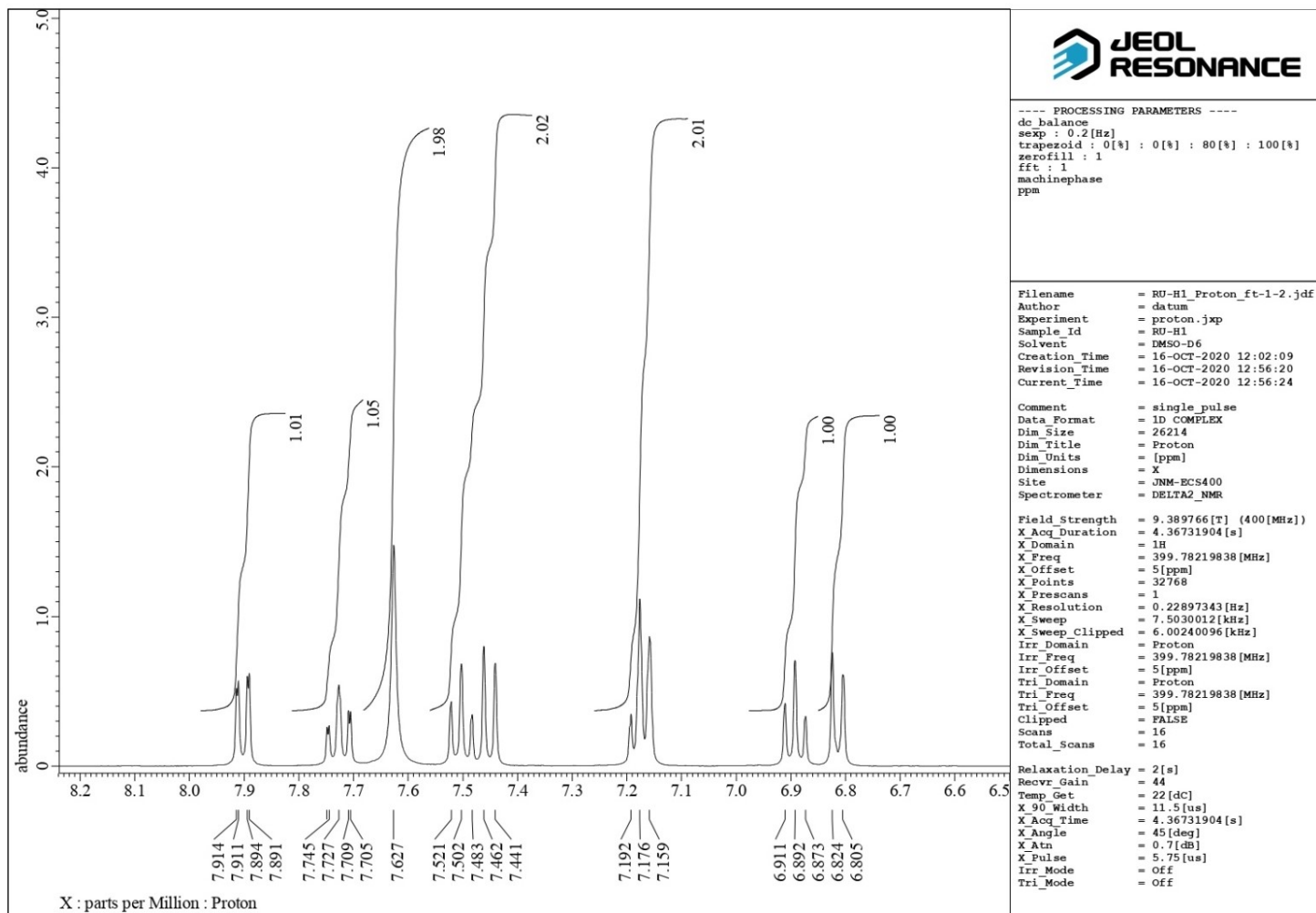


Figure 15A: ¹H-NMR spectrum of (4a) zoom

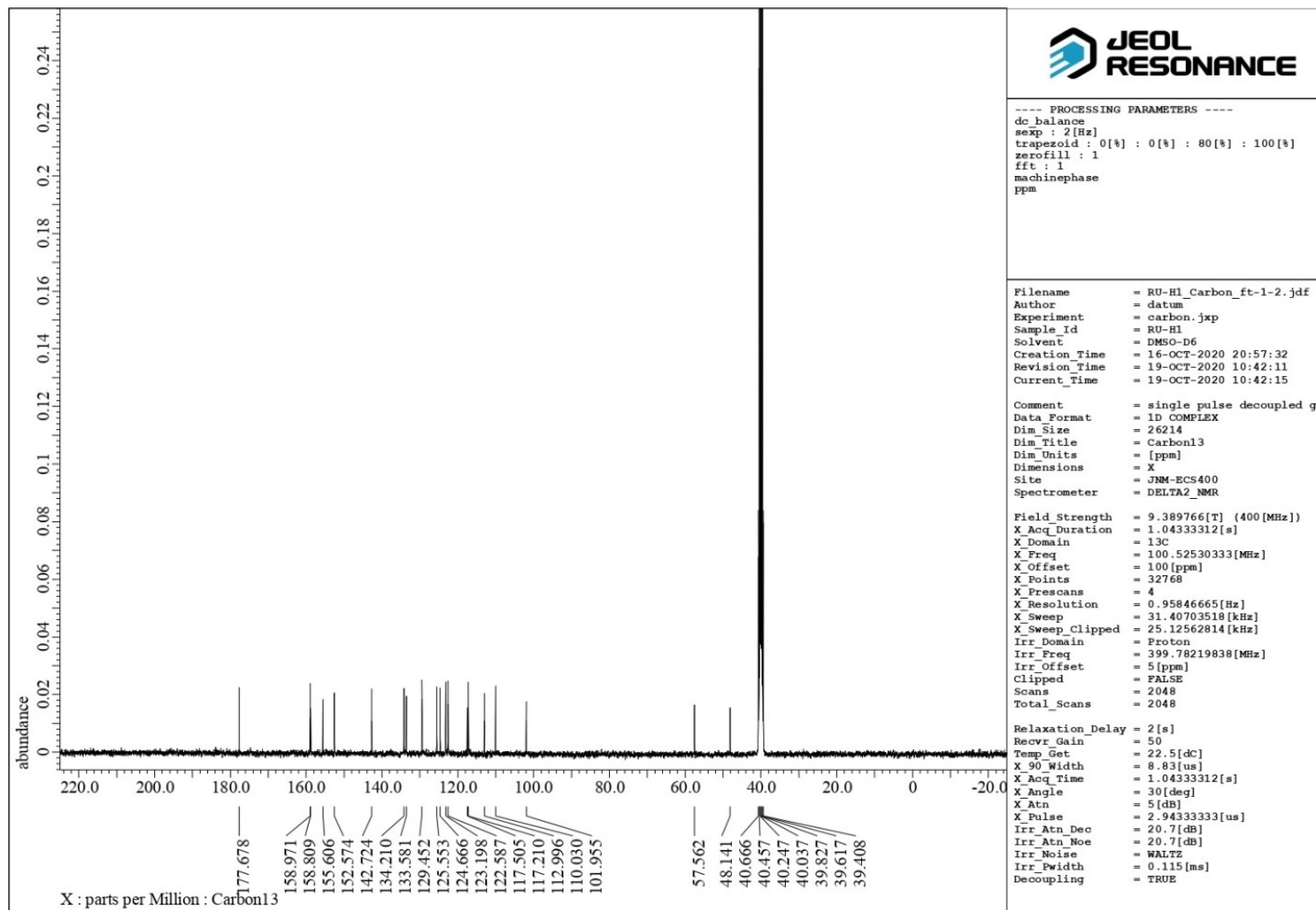


Figure 16: ^{13}C NMR spectrum of (4a)

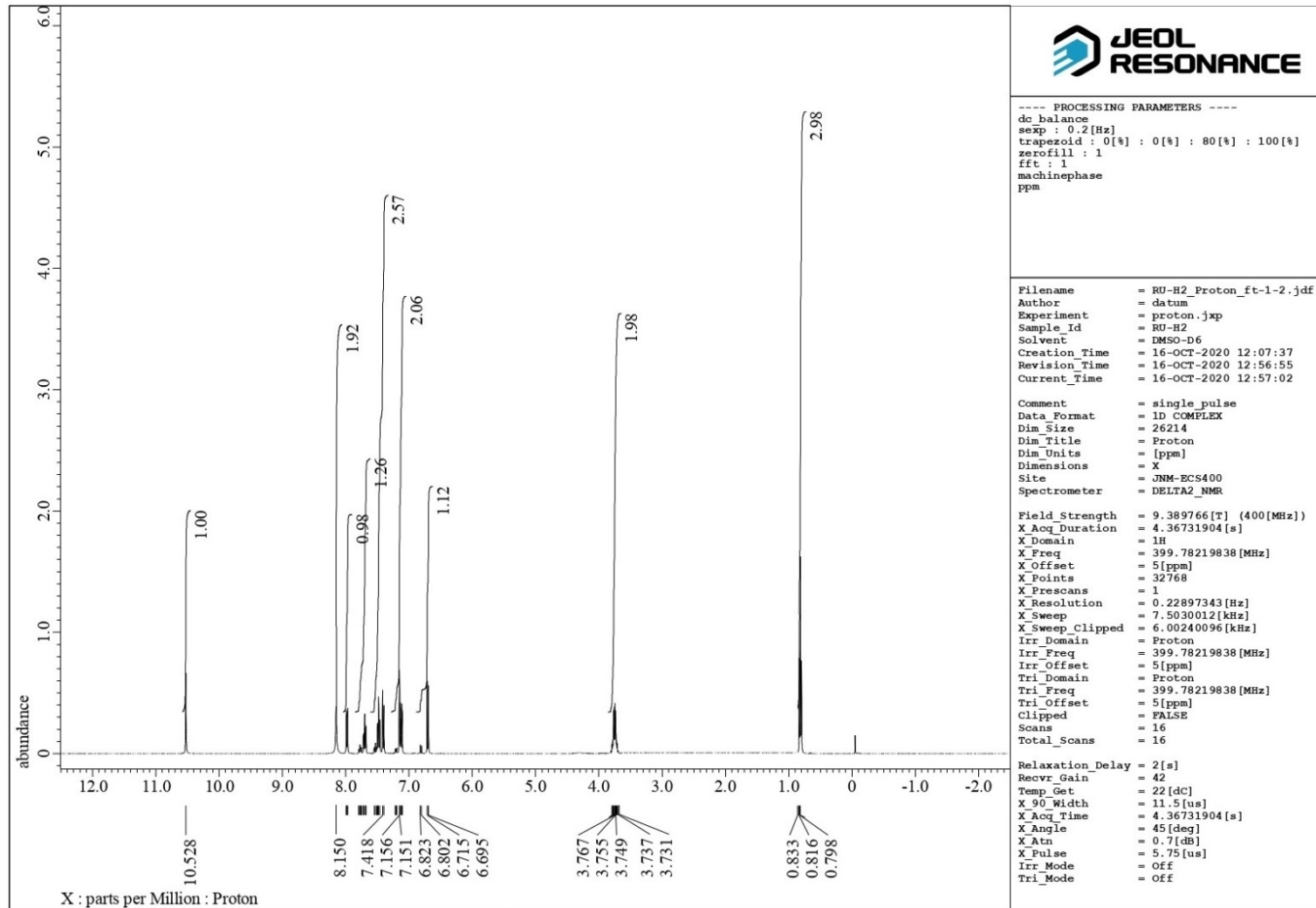


Figure 17: ^1H -NMR spectrum of (4b)

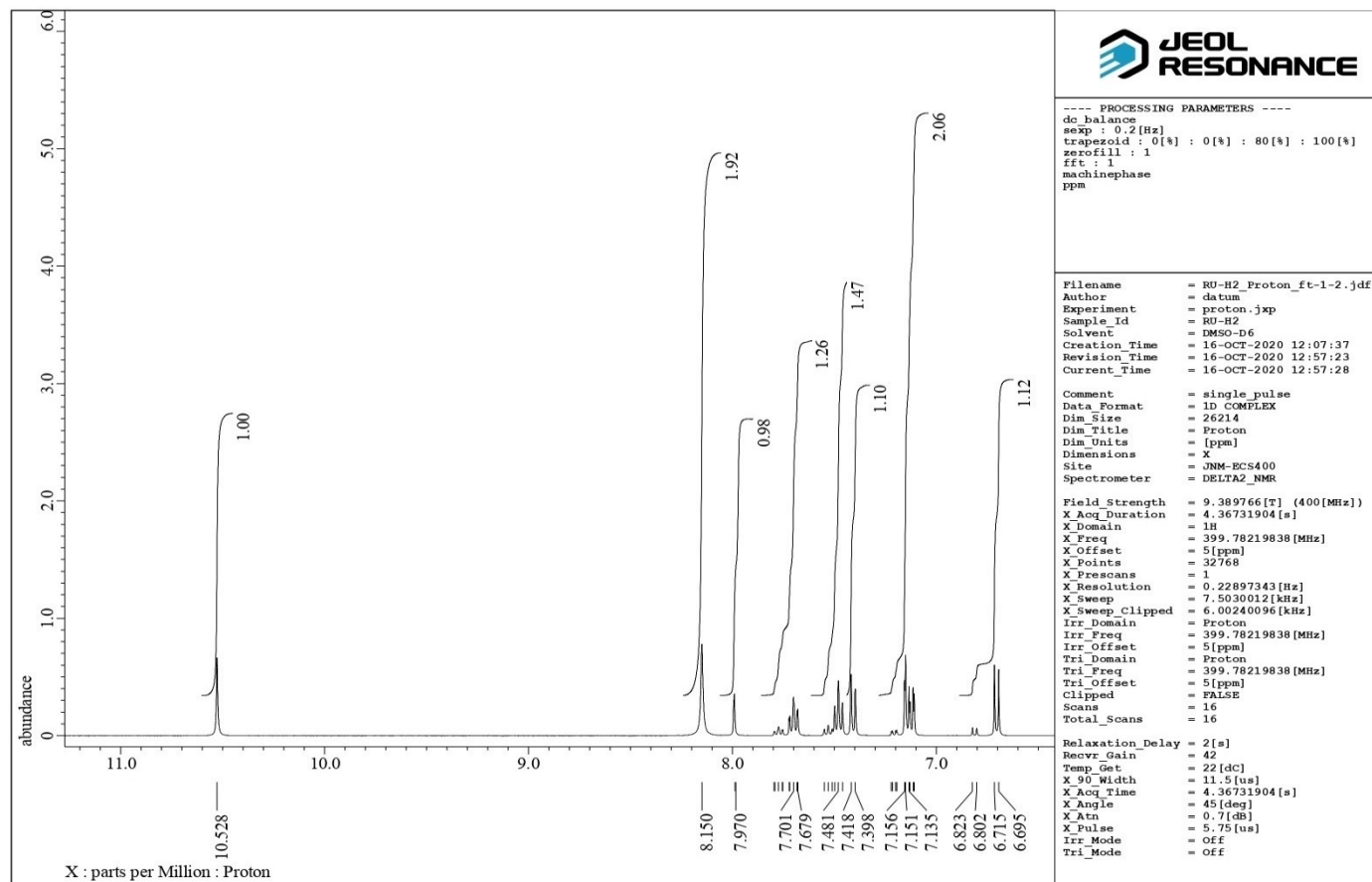


Figure 17A: ¹H-NMR spectrum of (4b) zoom

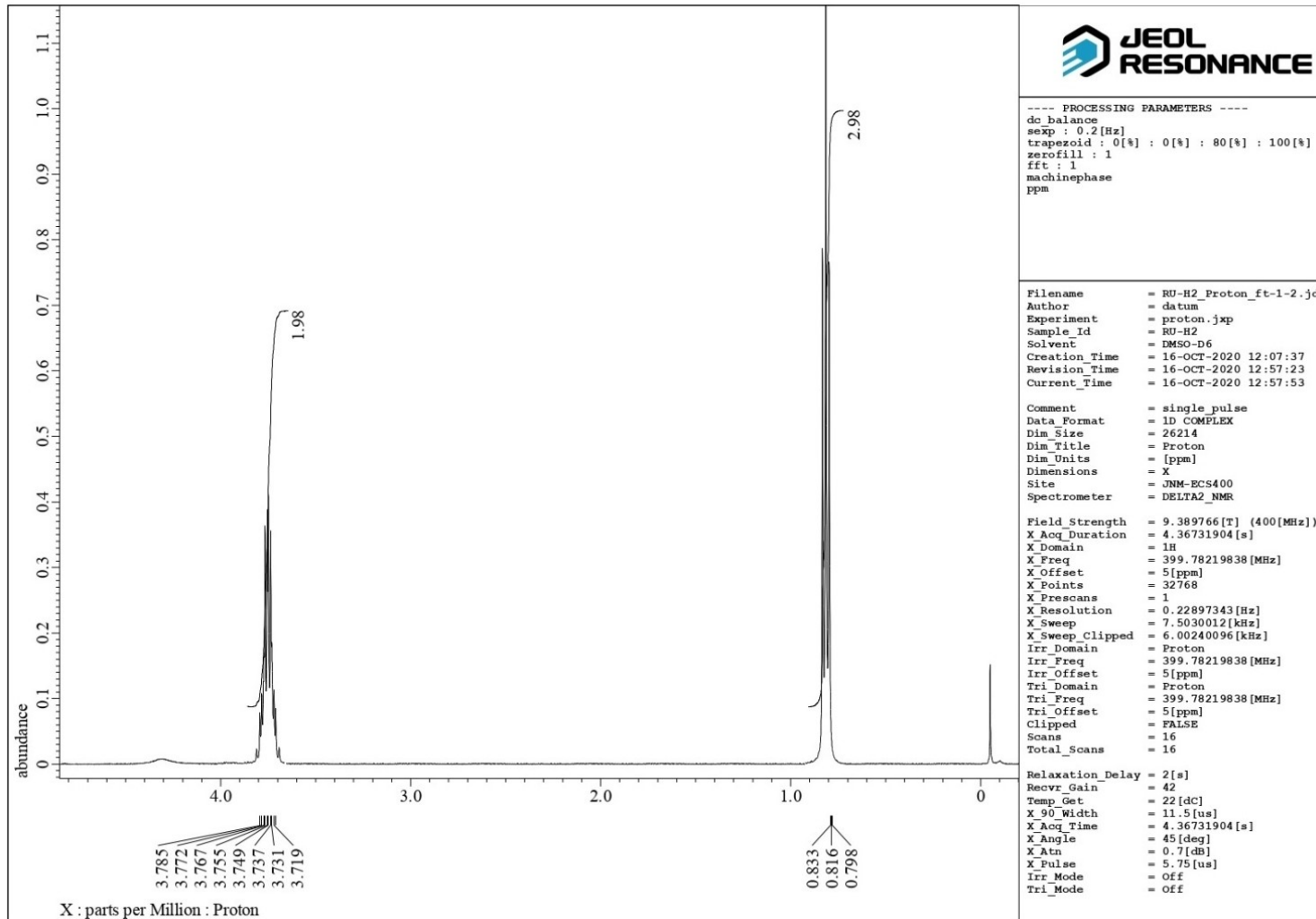


Figure 17B: ^1H -NMR spectrum of (4b) zoom

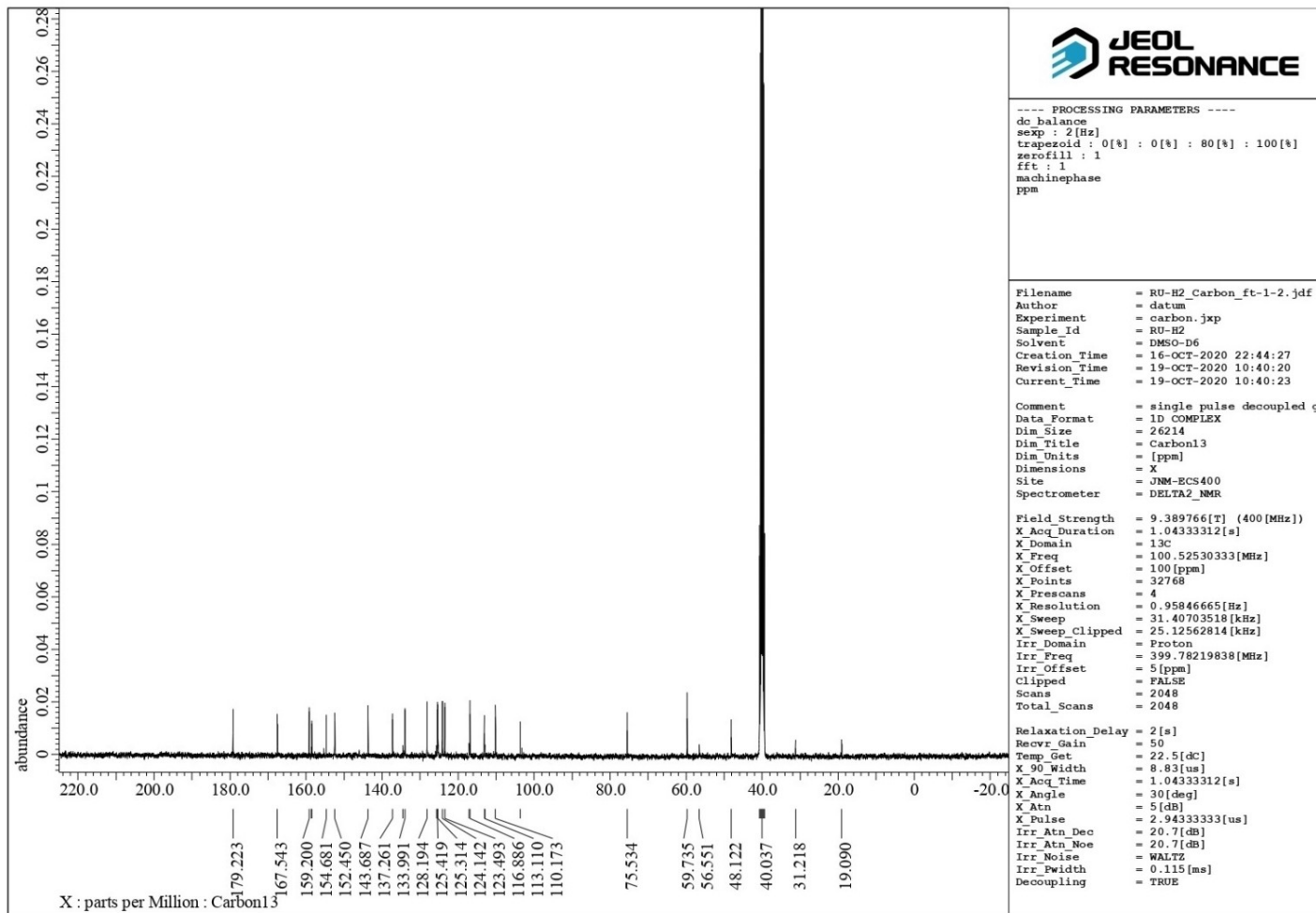


Figure 18: ¹³C NMR spectrum of (4b)

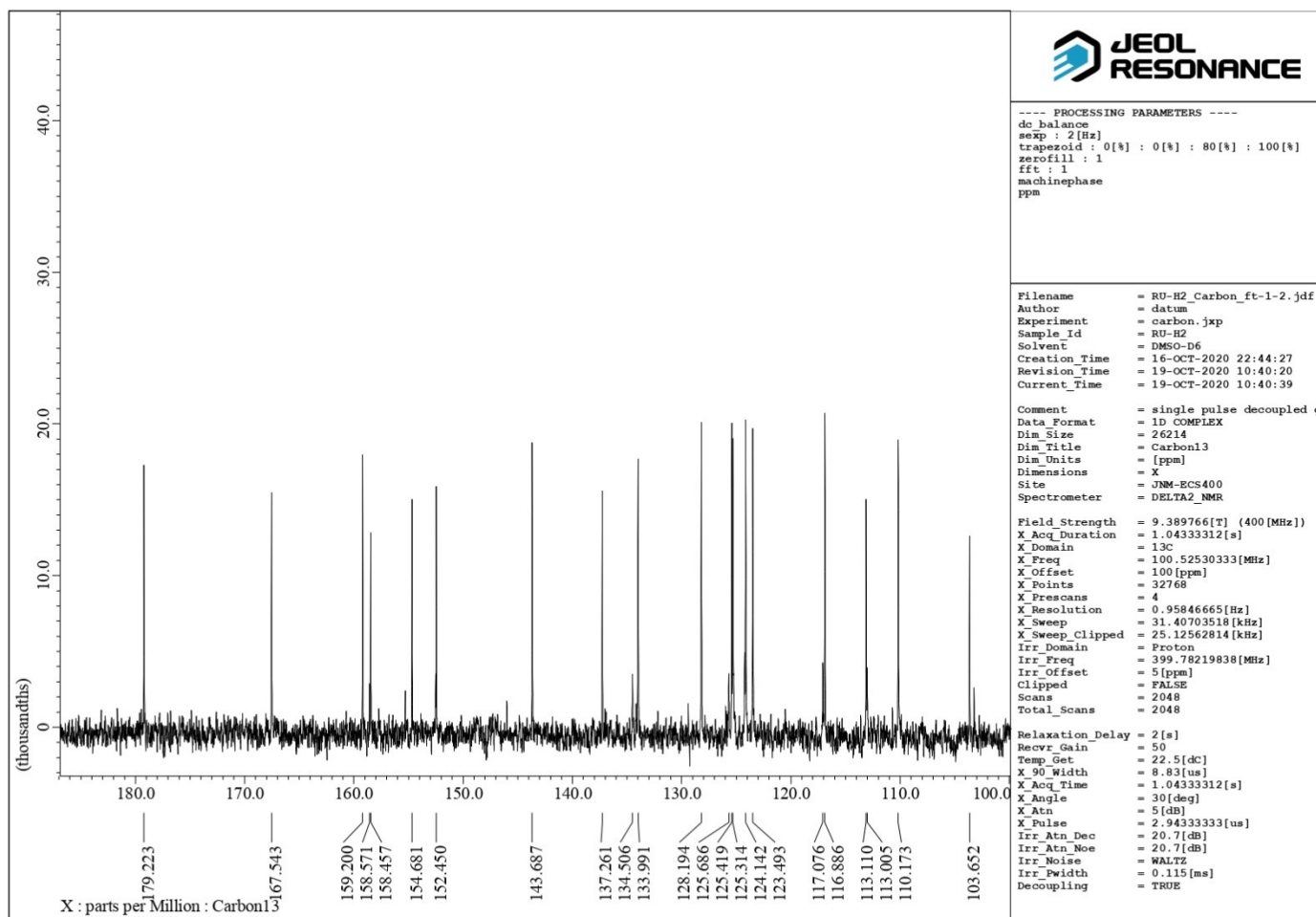


Figure 18A: ^{13}C NMR spectrum of (4b) Zoom

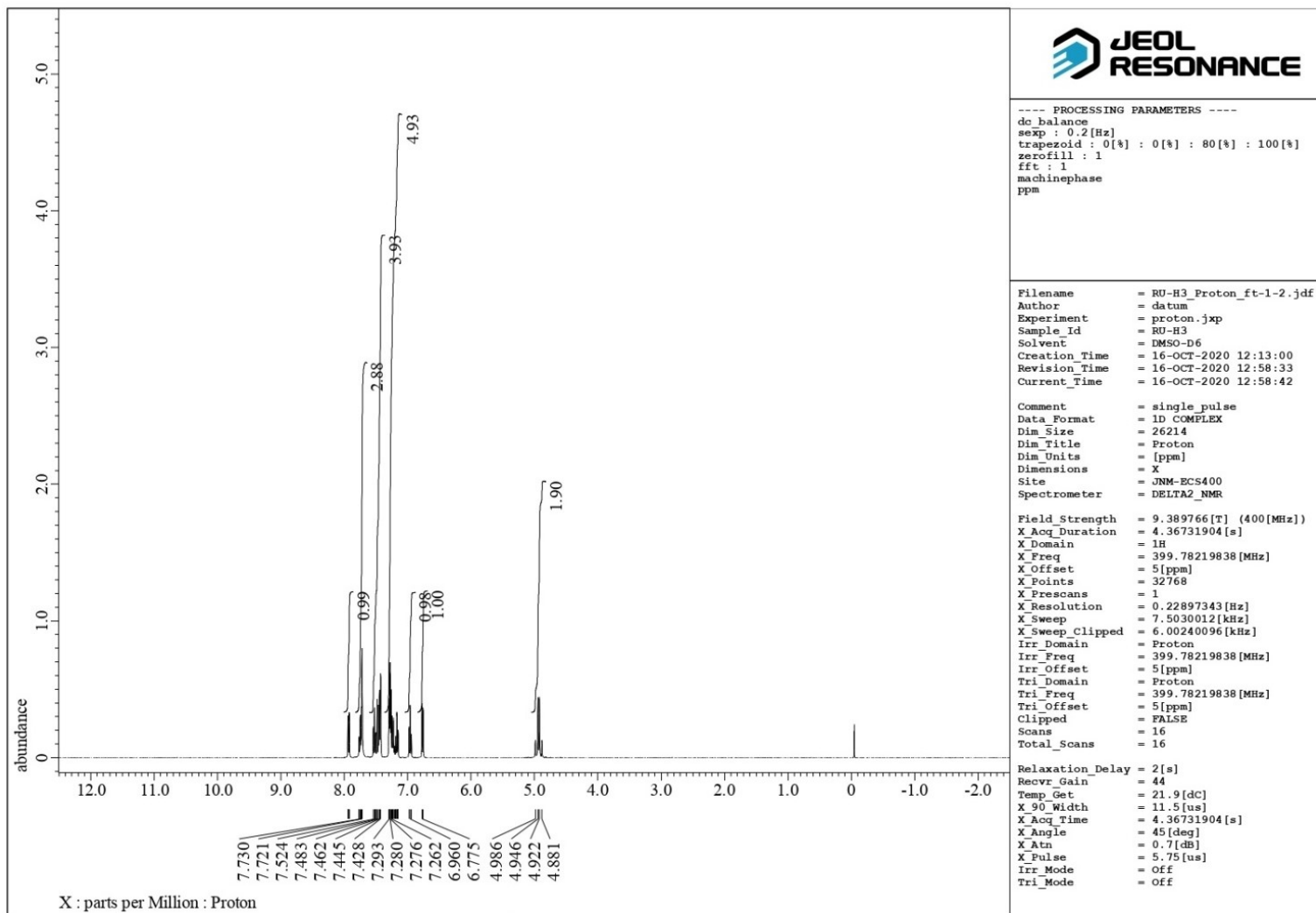


Figure 19: ^1H -NMR spectrum of (4c)

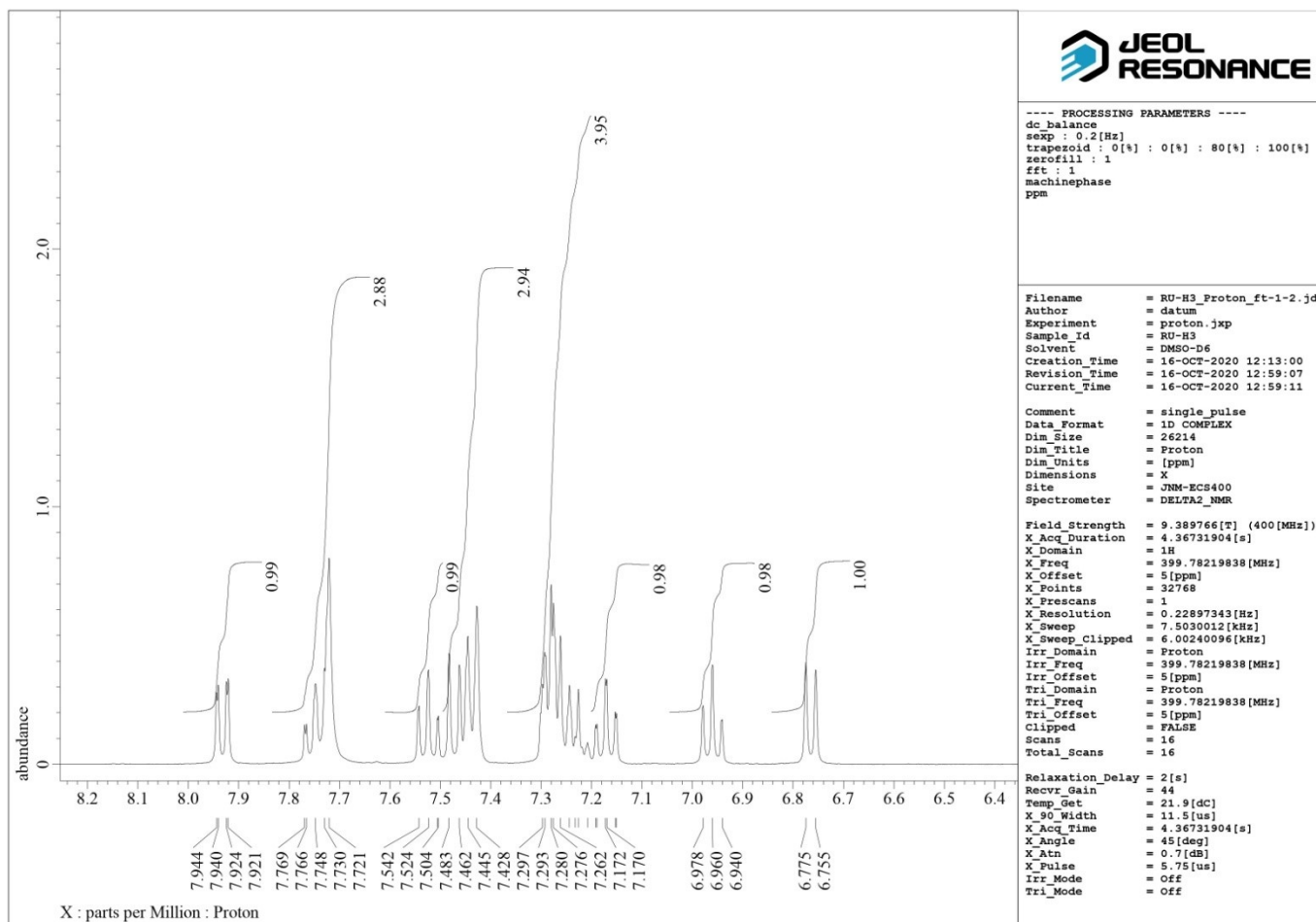


Figure 19A: ^1H -NMR spectrum of (4c) Zoom

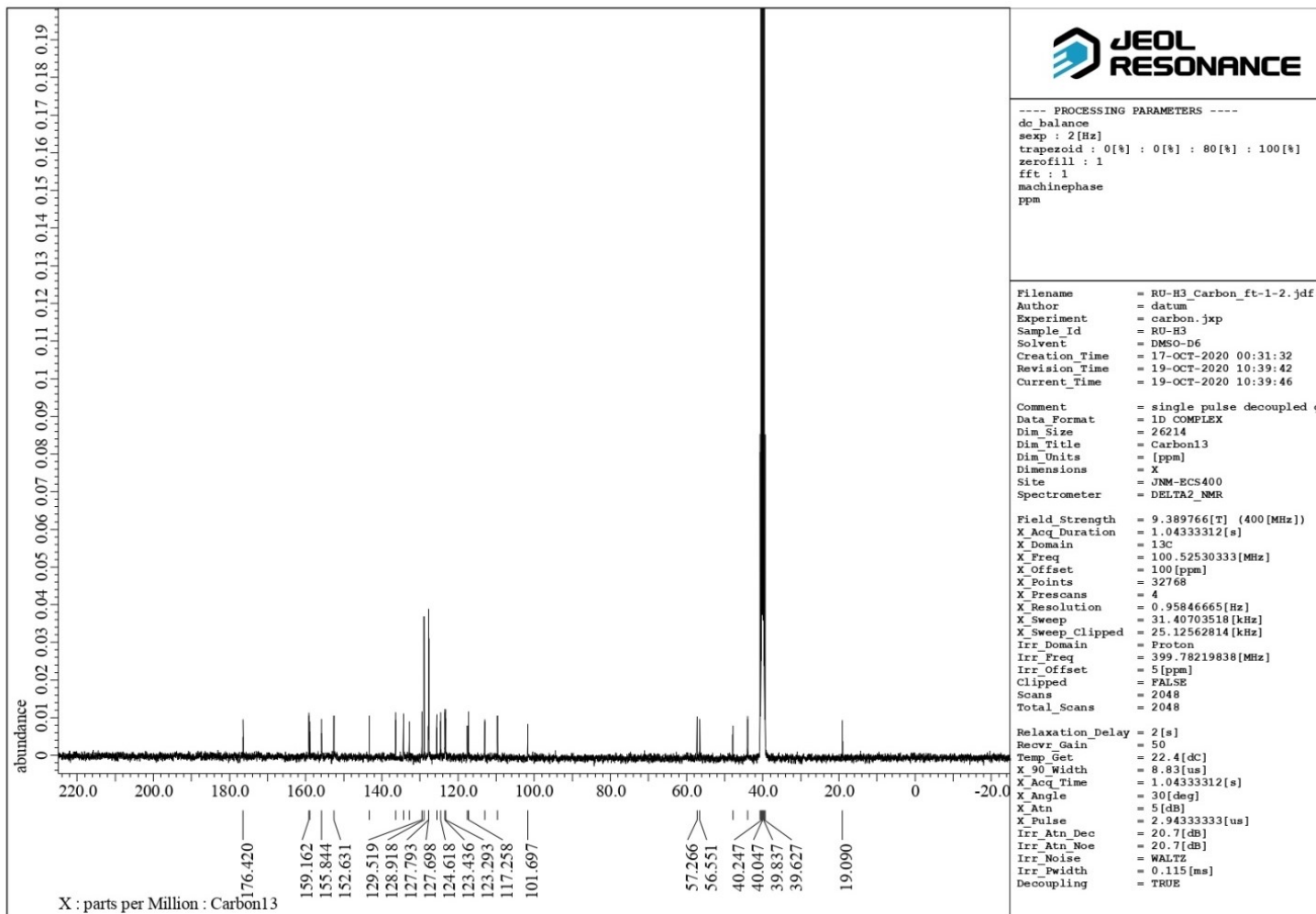


Figure 20: ¹³C NMR spectrum of (4c)

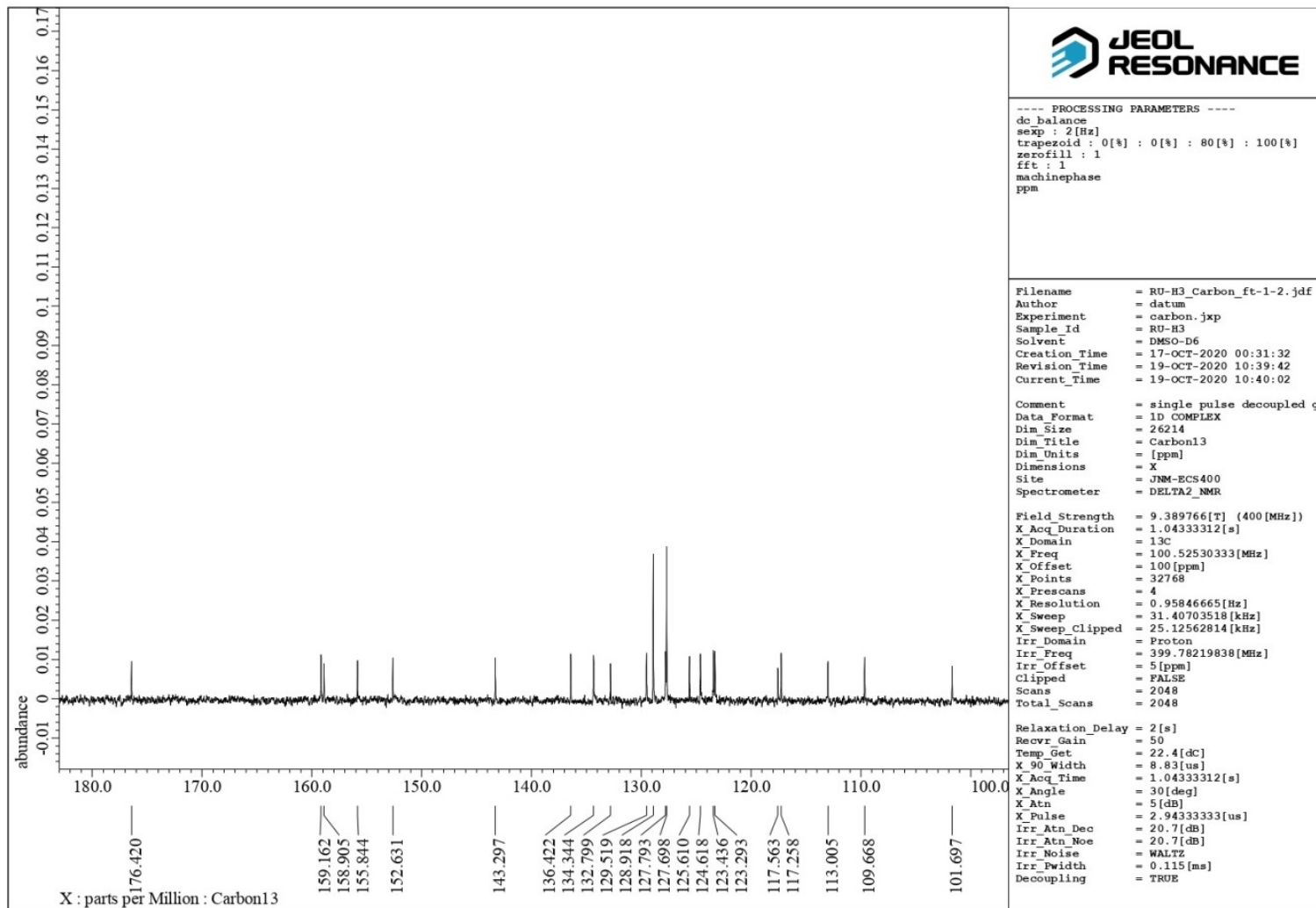


Figure 20A: ^{13}C NMR spectrum of (4c) zoom

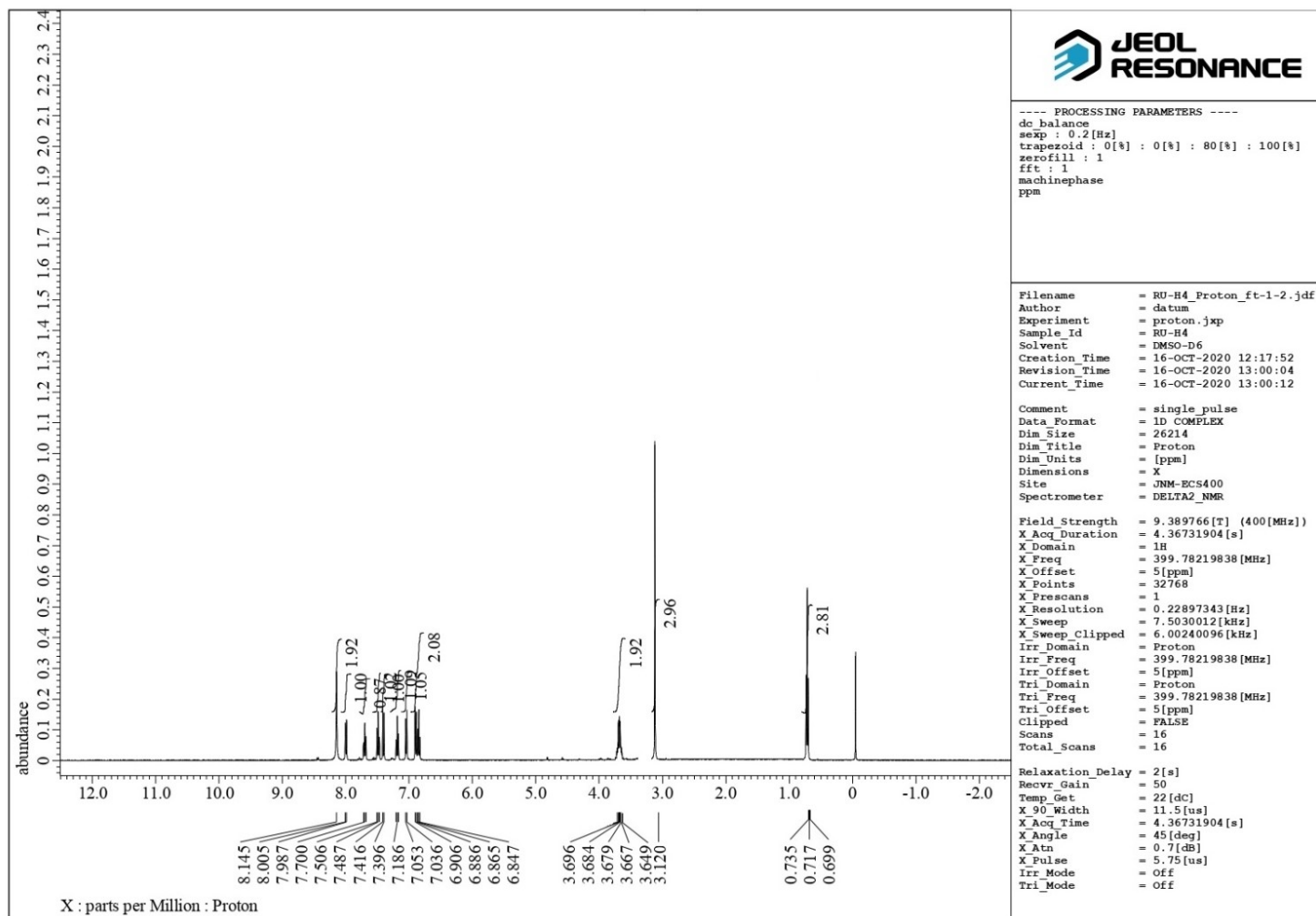
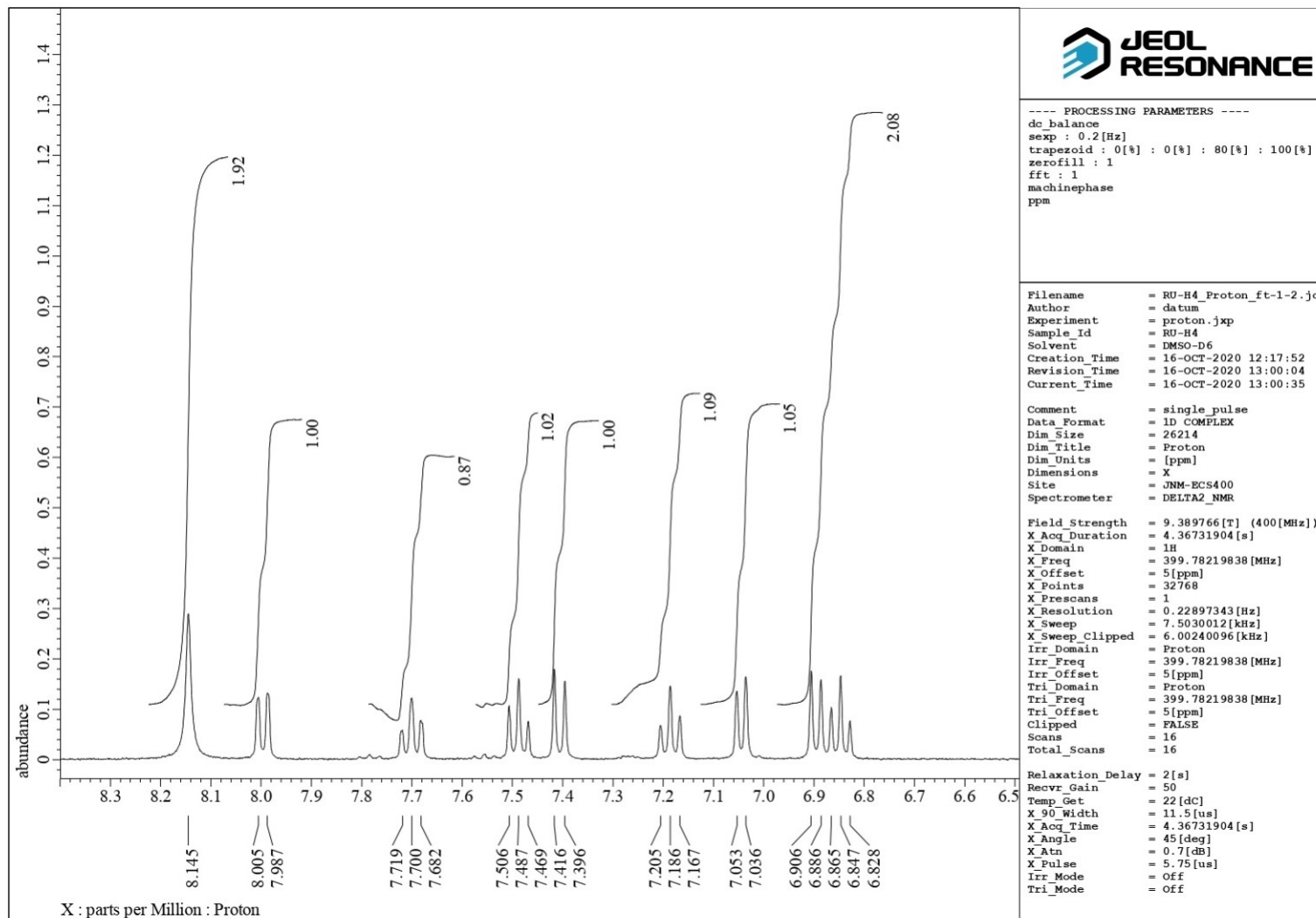


Figure 21: ¹H-NMR spectrum of (4d)



```

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fft : 1
machinephase
ppm

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Revision_Time = 16-OCT-2020 13:00:04
Current_Time = 16-OCT-2020 13:00:35

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Dim_Title    = Proton
Dim_Units    = [ppm]
Dimensions   = X
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Spectrometer = DELTA2_NMR

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X_Sweep_Clipped = 6.00240096[kHz]
Irr_Domain     = Proton
Irr_Freq       = 399.78219838[MHz]
Irr_Offset     = 5[ppm]
Tri_Domain     = Proton
Tri_Freq       = 399.78219838[MHz]
Tri_Offset     = 5[ppm]
Clipped        = FALSE
Scans          = 16
Total_Scans    = 16

Relaxation_Delay = 2[s]
Recvr_Gain       = 50
Temp_Get         = 22[dc]
X_90_Width      = 11.5[us]
X_Acq_Time      = 4.36731904[s]
X_Angle         = 45[deg]
X_Atn           = 0.7[db]
X_Pulse         = 5.75[us]
Irr_Mode        = Off
Tri_Mode        = Off

```

Figure 21A: ¹H-NMR spectrum of (4d) zoom

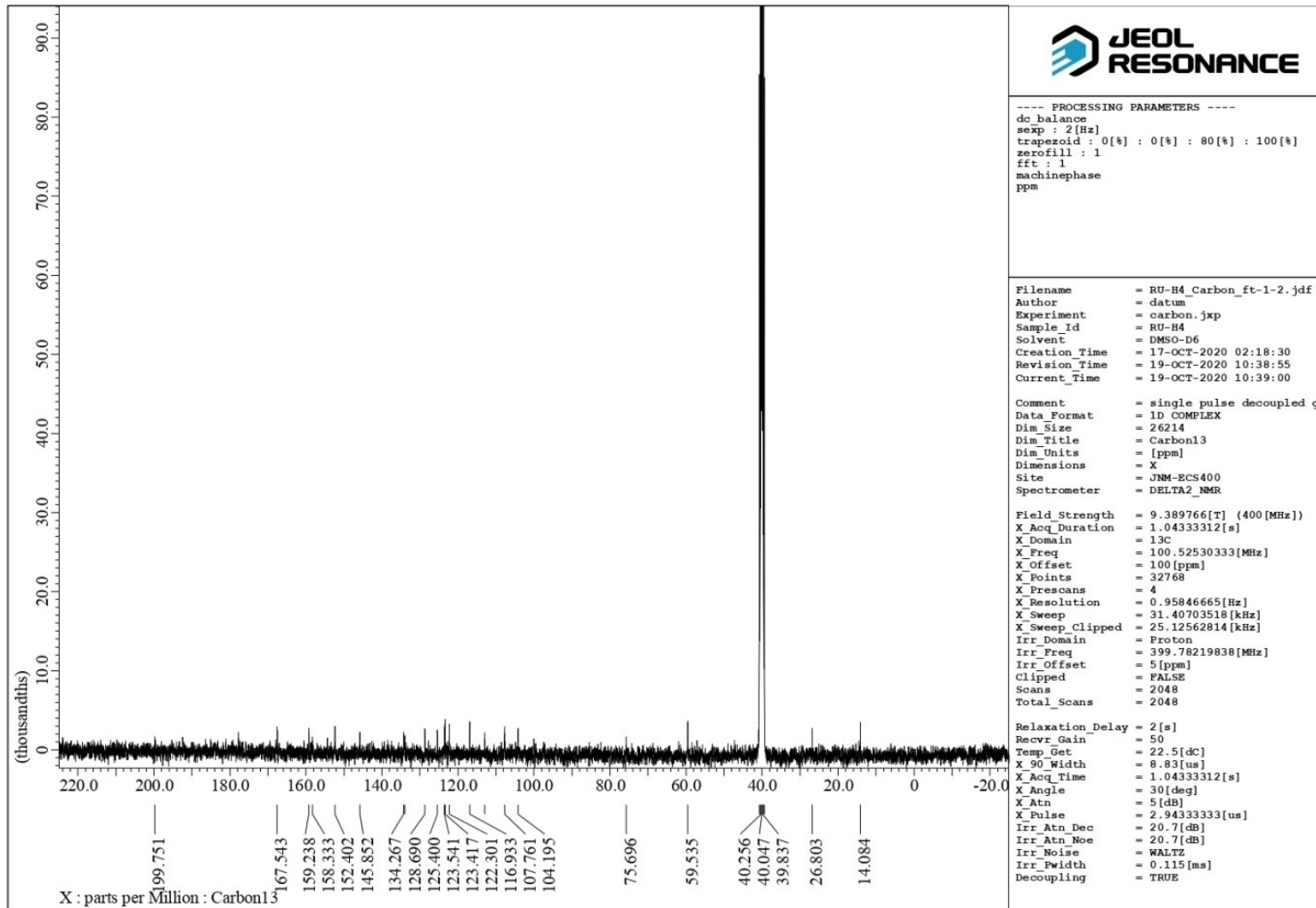


Figure 22: ^{13}C NMR spectrum of (4d)

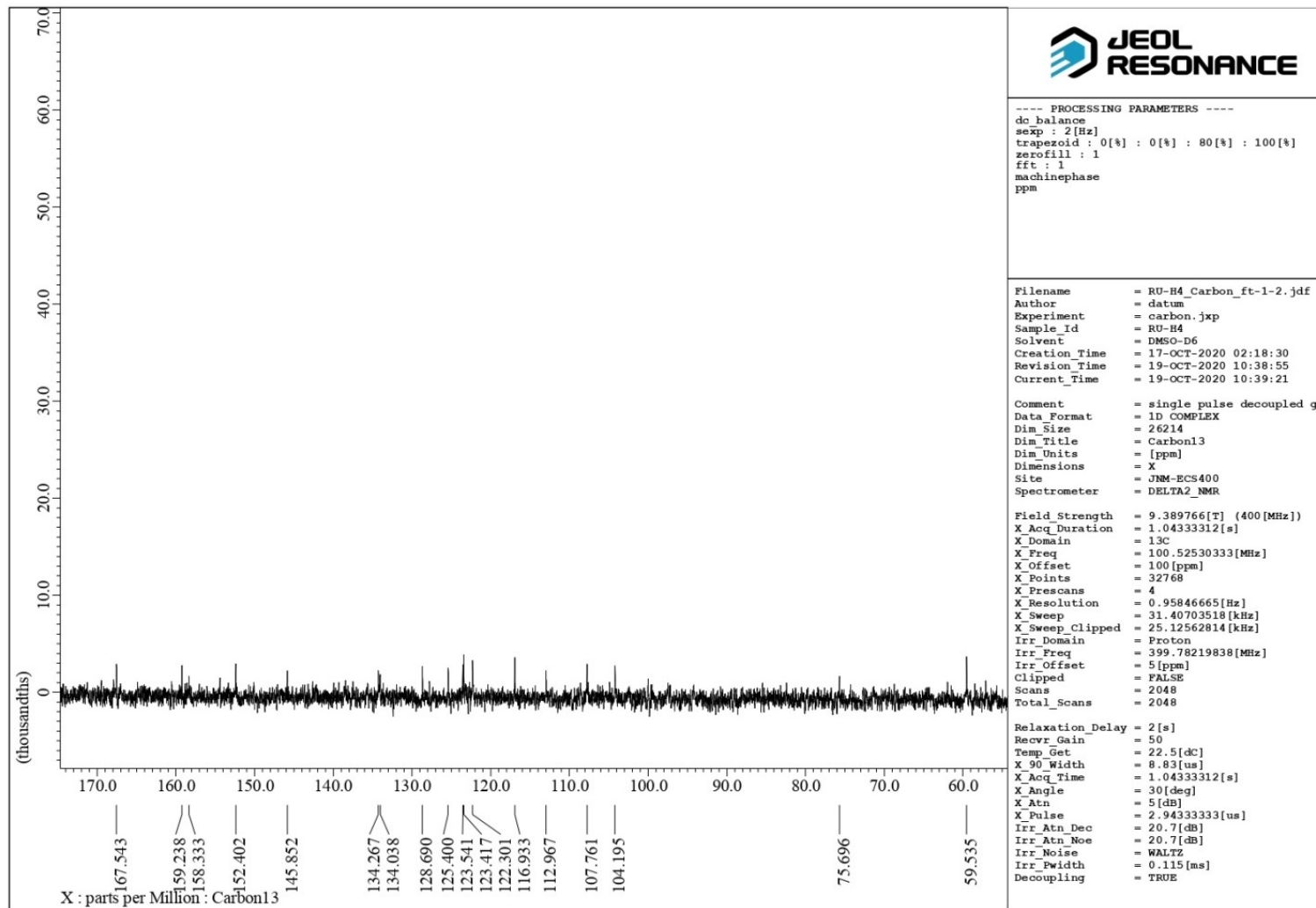


Figure 22A: ^{13}C NMR spectrum of (4d) zoom

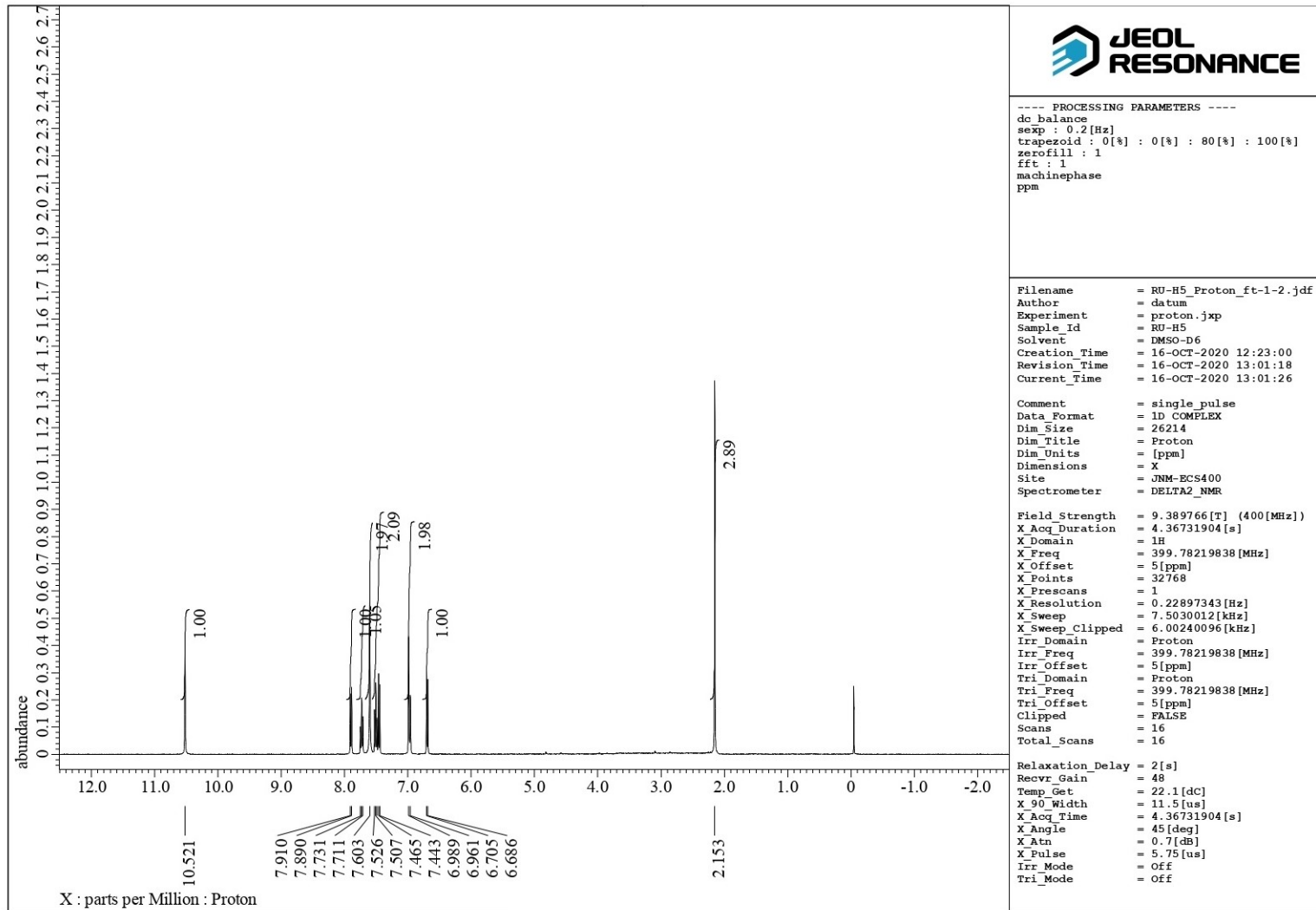


Figure 23: ^1H -NMR spectrum of (4e)

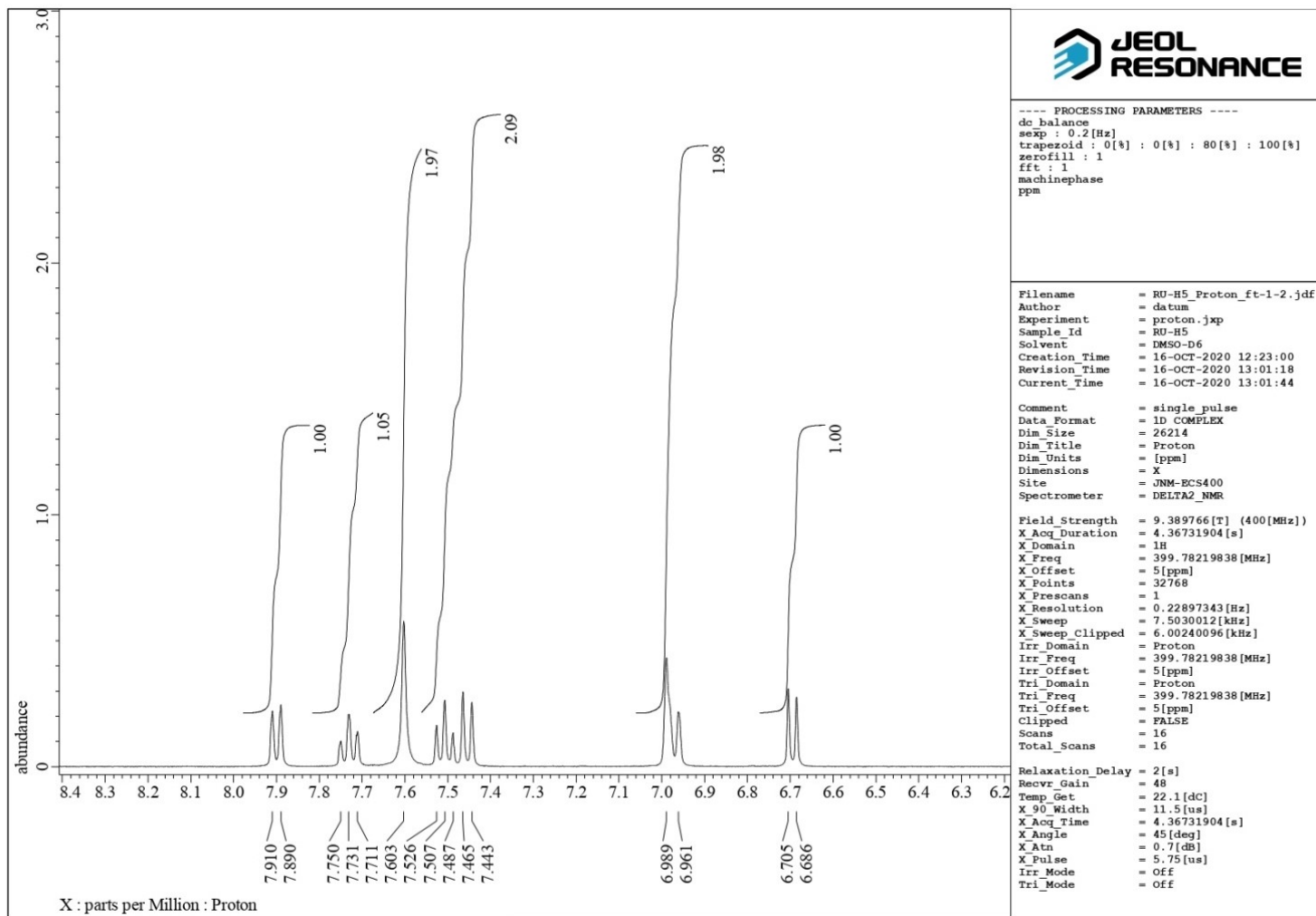


Figure 23A: ¹H-NMR spectrum of (4e) zoom

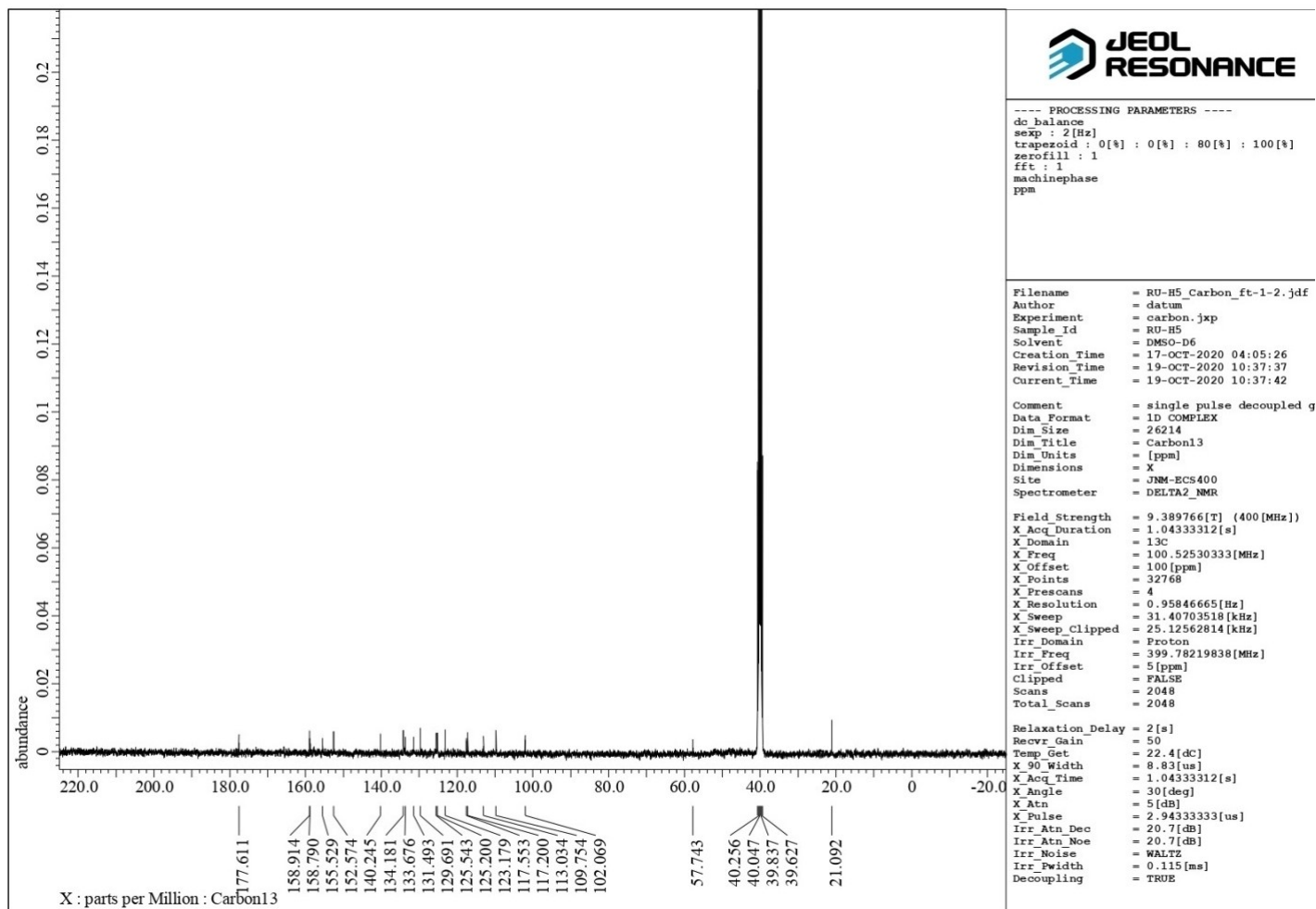


Figure 24: ¹³C NMR spectrum of (4e)

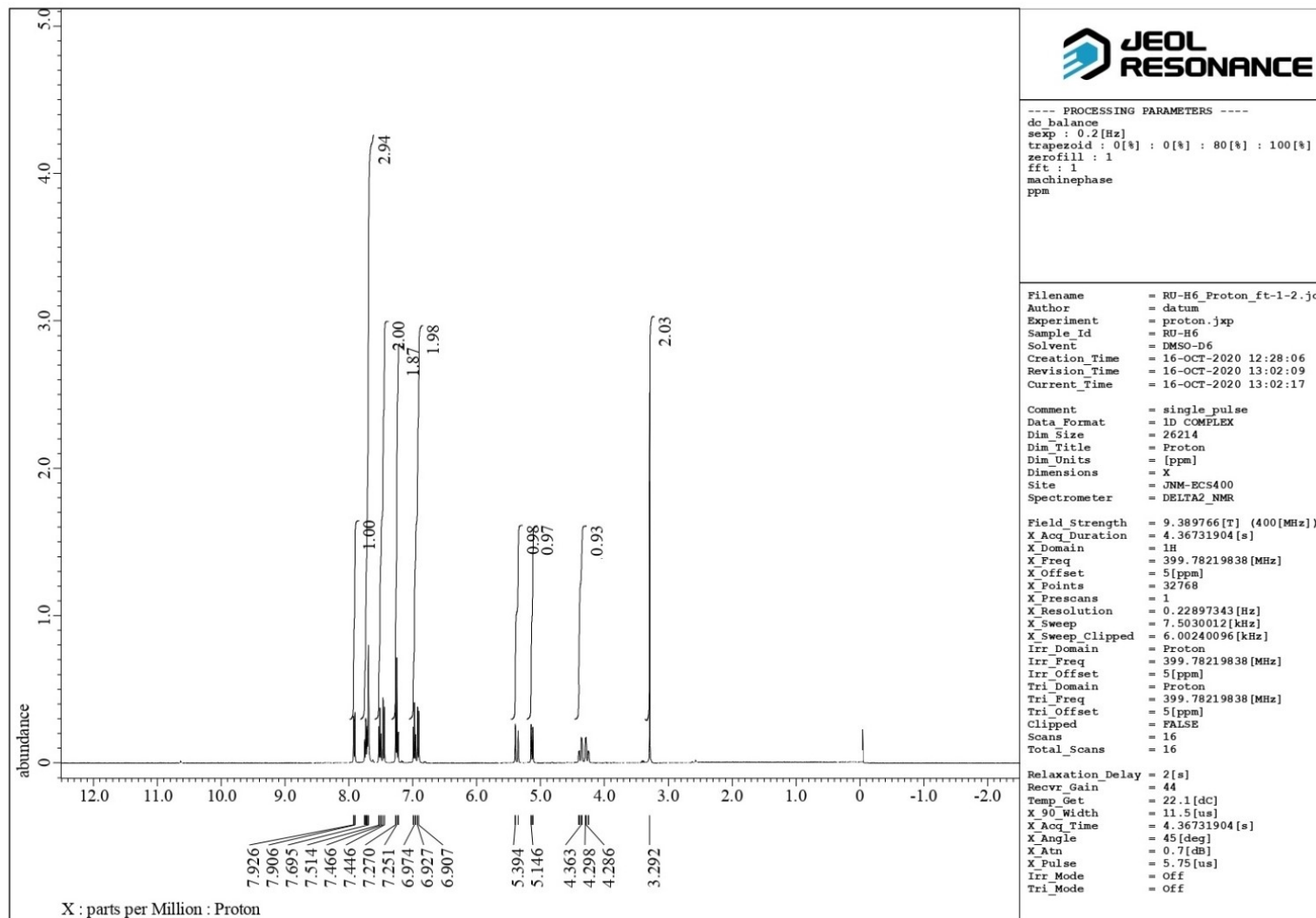


Figure 25: $^1\text{H-NMR}$ spectrum of (4f)

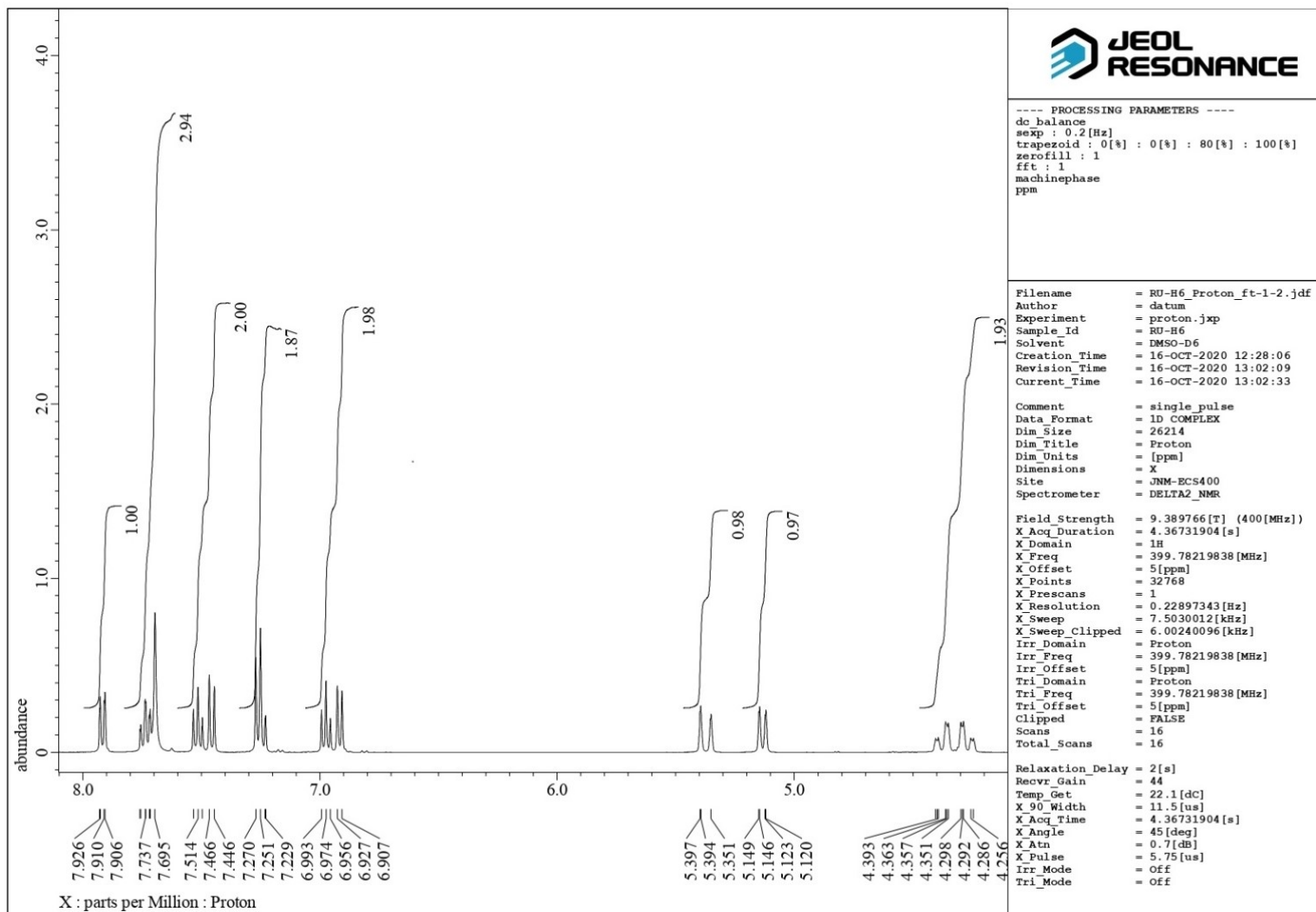


Figure 25A: ¹H-NMR spectrum of (4f) zoom

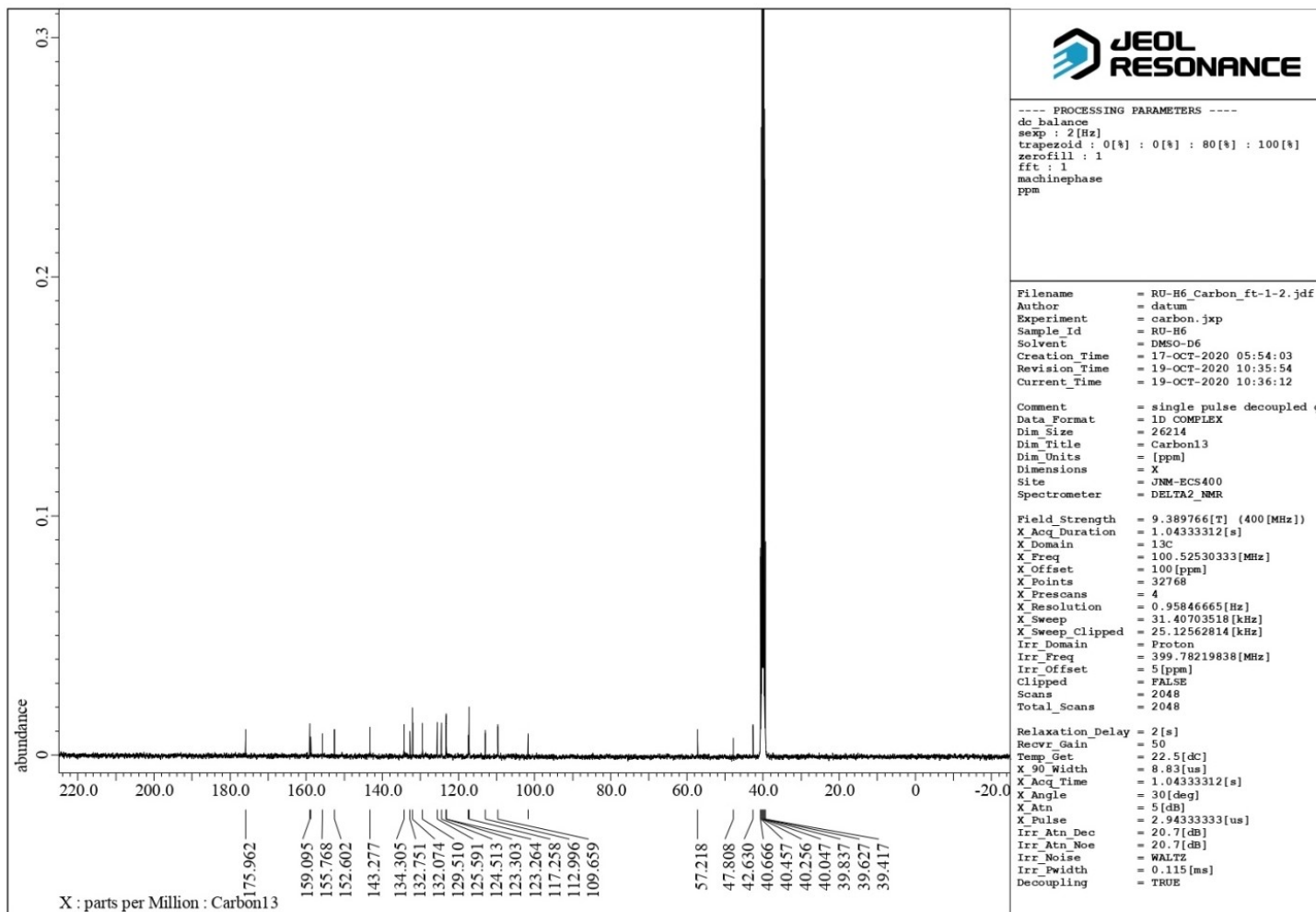


Figure 26: ¹³C NMR spectrum of (4f)

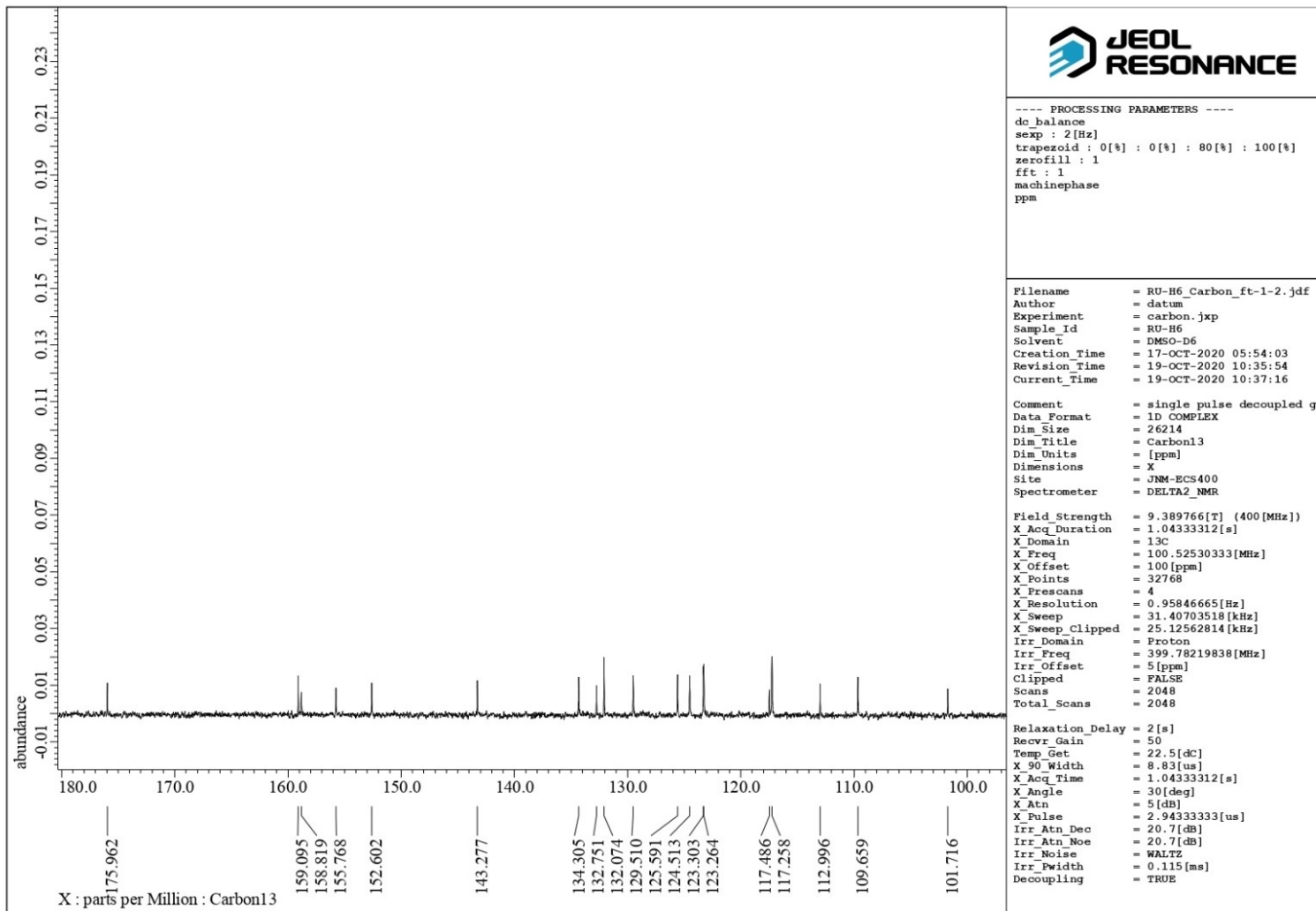


Figure 26A: ^{13}C NMR spectrum of (4f) zoom