Microwave-assisted synthesis of 4-oxo-2-butenoic acids by aldolcondensation of glyoxylic acid

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1. Table S1. Optimisation of the transformation of 4-methoxyacetophenone into 1 via aldol-condensation.

	Conditions	ОН				
		0				
Entry	Conditions	Yield after purification				
Littiy	1.5 eq. glyoxylic acid monohydrate, 1.7 eq. NaH 60%					
1	dispersion in mineral oil,	Low conversion ¹				
	DMSO, 80 °C, 18 h					
2	1.5 eq. glyoxylic acid monohydrate,					
	1.7 eq. NaH 60% dispersion in mineral oil, DMSO, 80	Low conversion ¹				
	°C, 18h, then 1.5 eq. NaH 60% dispersion in mineral oil,					
	1.5 eq. TsCl, 80 °C, 5 min					
	1.5 eq. glyoxylic acid monohydrate					
3	1.7 eq. LiOH monohydrate	Moderate conversion ²				
	MeOH, 80°C, 18h					
	3.0 eq. glyoxylic acid monohydrate,					
4	1.0 eq. pyrrolidine, 1.0 eq. acetic acid, Moderate convers					
	MeOH, MW, 80°C, 8 h					
5	3.0 eq. glyoxylic acid monohydrate	Moderate conversion ²				
J	3.0 eq. TsCl, dioxane, 80°C, 18 h					
6	3.0 eq. glyoxylic acid monohydrate	No product				
	3.0 eq. Tf ₂ O, dioxane, 80°C, 18 h					
7	3.0 eq. glyoxylic acid monohydrate					
	1.0 eq. TsOH monohydrate	70%				
	dioxane, 80°C, 48 h					

¹Low conversion: < 10% desired product by UV and/or ELSD LC-MS analysis. ²Moderate conversion: < 50% desired product by UV and/or ELSD LC-MS analysis.





3. Table S2. Calculated HOMO and LUMO energies for different methyl ketone substrates and glyoxylic acid.

Calculations employed the RHF/6-31+G** level of theory in the Gaussian09 suite of programs. Geometries were optimised and frequencies computed to verify that they are minima.

Methyl ketone substituent	Substituent class	ENOL HOMO	ENOL LUMO	ENOLATE HOMO	ENOLATE LUMO	ENAMINE_HO MO	ENAMINE_LU MO
cyclohexyl	aliphatic	-0.33597	0.07163	-0.08481	0.16325	-0.29239	0.06944
cyclopropyl	aliphatic	-0.32685	0.06935	-0.08144	0.18209	-0.2852	0.07097
isopropyl	aliphatic	-0.33841	0.07206	-0.08243	0.18843	-0.29357	0.07363
n-propyl	aliphatic	-0.33828	0.07428	-0.08121	0.17438	-0.29256	0.07546
t-butyl	aliphatic	-0.33857	0.07477	-0.08406	0.18351	-0.30383	0.07418
p-methylphenyl	aromatic	-0.30309	0.06808	-0.09087	0.14946	-0.28913	0.07035
p-ethylphenyl	aromatic	-0.30318	0.06618	-0.09156	0.14344	-0.296	0.06673
p-fluorophenyl	aromatic	-0.31576	0.0643	-0.09751	0.1698	-0.29798	0.06697
p-chlorophenyl	aromatic	-0.3168	0.06266	-0.1016	0.1533	-0.2998	0.06417
p-methoxyphenyl	aromatic	-0.29375	0.06866	-0.09049	0.14403	-0.28246	0.06718
p-cyanophenyl	aromatic	-0.33266	0.0507	-0.11218	0.1525	-0.31016	0.05943

Glyoxylic acid	номо	LUMO
Neutral	-0.45302	0.05375
Protonated	-0.72186	-0.2152

4. General Information for the Synthesis

Chemicals were purchased from commercial suppliers and used without further purification. Thin layer chromatography (TLC) was performed on aluminium plates coated with 60 F254 silica from Merck. Flash chromatography was carried out using a Biotage SP4, Biotage Isolera or Varian automated flash system with Silicycle or GraceResolve normal phase silica gel pre-packed columns. Fractions were collected at 254 nm or if necessary on all wavelengths between 200 and 400 nm. Microwave irradiation was performed in a Biotage Initiator Sixty in sealed vials (Biotage microwave vials, Type I, Class A borosilicate, 28 mm outer diameter, 26 mm inner diameter, 83 mm long, round-bottom for 5-20 mL of total reaction volume; Biotage microwave vials, 16 mm outer diameter, 14 mm inner diameter, 83 mm long, round-bottom for 2-5 mL of total reaction volume). Reactions were irradiated at 2.45 GHz and were able to reach temperatures between 60 and 250 °C. Heating was at a rate of 2-5 °C/s and the pressure was able to reach 20 bars.

5. Analytical Equipment

Melting points were measured using a Stuart automatic melting point SMP40 apparatus or a Shanghai ShenGuang WRR apparatus. Fourier Transform InfraRed (FTIR) spectra were measured using an Agilent Cary 630 FTIR or a Bruker TENSOR II FTIR Spectrometer as a neat sample tableting with KBr. The abbreviations for peak description are as follows: b = broad; w = weak; m = medium and s = strong. Ultraviolet (UV) spectra were recorded on a Hitachi U-2900 spectrophotometer or an Agilent Cary 100 UV-Vis spectrophotometer and were performed in ethanol. High resolution mass spectrometry (HRMS) was provided by the ESPRC National Mass Spectrometry Service, University of Wales, Swansea, or the Mass Spectrometry Service, Department of Pharmacy, Naval Medical University and performed by Diya Lyu on an Agilent Technologies 6550 iFunnel Q-TOF LC-MS, or conducted using an Agilent 6550 iFunnel QTOF LC-MS with an Agilent 1260 Infinity UPLC system. The sample was eluted on Acquity UPLC BEH C18 (1.7µm, 2.1 x 50mm) with a flow rate of 0.7 mL/min and run at a gradient of 1.2 min 5-95% 0.1% aq. HCOOH in MeCN.

LC-MS analyses were conducted using a Waters Acquity UPLC system with photo diode array (PDA) and evaporating light scattering detector (ELSD) or using the ESI mass spectra which were performed by Zichao Ding on an Agilent Technologies 6120 Quadrupole LC-MS. When a 2 min gradient was used, the sample was eluted on an Acquity UPLC BEH C18, 1.7 μ m, 2.1 x 50mm, with a flow rate of 0.6 ml/min using 5-95% 0.1% HCOOH in MeCN. Analytical purity of compounds was determined using Waters XTerra RP18, 5 μ m (4.6 × 150 mm) column at 1 ml/min using either 0.1% aq. ammonia and MeCN or 0.1% aq. HCOOH and MeCN with a gradient of 5-100% over 15 min. When a 12 min gradient was used, the sample was eluted on ZORBAX Eclipse XDB-C18, 3.5 μ m, 4.6 x 100 mm, with a flow rate of 0.4 ml/min using 30-70% 0.1% HCOOH in MeCN.

¹H and ¹³C NMR spectra were obtained using a Bruker Avance III 500 spectrometer using a frequency of 500 MHz, and 123 MHz, respectively, or using a Bruker Avance III 600 spectrometer operating at a frequency of 600 MHz, and 150 MHz, respectively. ¹⁹F NMR spectra were acquired using the Bruker Avance III 300 spectrometer using a frequency of 282 MHz. The abbreviations for spin multiplicity are as follows: s = singlet; d = doublet; t = triplet; q = quartet, p = quintuplet, h = sextuplet and m = multiplet. Combinations of these abbreviations are employed to describe more complex splitting patterns (e.g. dd = doublet of doublets).

6. NMR spectra



































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