

## Supporting Information

### An Efficient Synthesis of RNA Containing GS-441524: The Nucleoside Precursor of Remdesivir

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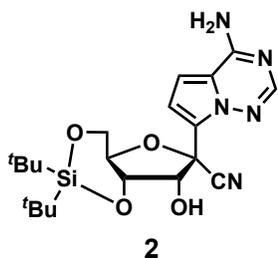
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**I. General Reaction Conditions.** All commercial chemicals were used as received. Reaction solvents acetonitrile (CH<sub>3</sub>CN), dichloromethane (CH<sub>2</sub>Cl<sub>2</sub>), and tetrahydrofuran (THF) were dried by passage over a column of activated alumina using a solvent purification system (MBraun). Dimethylformamide (DMF) was dried by passage over a column of molecular sieves using a solvent purification system (MBraun). Pyridine, *N,N*-dimethylacetamide (DMA), 2-(trimethylsilyl)ethoxymethyl chloride (SEM-Cl), *tert*-butylmagnesium chloride solution (<sup>t</sup>BuMgCl, 1M solution in THF), hydrogen fluoride pyridine complex, and 1,2-dichloroethene (1,2-DCE) were obtained from Sigma Aldrich. (2*R*,3*R*,4*S*,5*R*)-2-(4-aminopyrrolo[2,1-*f*][1,2,4]triazin-7-yl)-3,4-dihydroxy-5-(hydroxymethyl)tetrahydrofuran-2-carbonitrile (**GS-441524**) was obtained from Ontario Chemicals Inc. *N,N*-dimethylformamide dimethyl acetal was obtained from Acros Organics. *N,N*-diisopropylethylamine (DIPEA) was distilled from calcium hydride and stored with 4 Å molecular sieves under argon until use. Synthesized compounds were purified on a Teledyne-Isco Combiflash Rf-200 instrument using Redisep Rf High Performance silica gel columns from Teledyne-Isco. <sup>1</sup>H NMR (500 MHz), <sup>13</sup>C NMR (126 MHz), and <sup>31</sup>P NMR (202 MHz) spectra were recorded on a Bruker Avance NMR spectrometer (500 MHz) at room temperature. NMR chemical shifts (δ) are recorded relative to TMS (0.05% v/v; δ = 0.0) for <sup>1</sup>H NMR and the solvent signal for <sup>13</sup>C NMR (δ = 77.0 for CDCl<sub>3</sub>, and δ = 39.7 for (CD<sub>3</sub>)<sub>2</sub>SO). High-resolution mass spectra were obtained at the Analytical Biochemistry Core Facility at the University of Minnesota Masonic Cancer Center using an LTP Orbitrap Velos Mass Spectrometer (Thermo Fisher).

## II. Synthesis of the GS-441524 Phosphoramidite.

### (4aR,6R,7R,7aS)-6-(4-aminopyrrolo[2,1-f][1,2,4]triazin-7-yl)-2,2-di-tert-butyl-7-hydroxytetrahydro-4H-furo[3,2-d][1,3,2]dioxasilin-6-carbonitrile (**2**):



To a flame-dried flask under Ar was added **GS-441524** (500 mg, 1.71 mmol), di-*tert*-butyl-di-chlorosilane (0.543 mL, 2.57 mmol), AgNO<sub>3</sub> (0.437 g, 2.57 mmol), and DMF (6 mL). The reaction mixture was stirred at 0 °C for 90 minutes. The reaction was quenched with water (5 mL) and diluted with EtOAc (80 mL). The EtOAc layer was subsequently washed with brine (3 X 50 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo*. The crude material was purified by SiO<sub>2</sub>

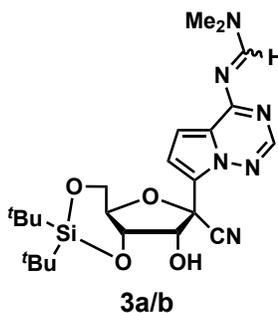
chromatography (gradient 0 – 10% CH<sub>3</sub>OH in CHCl<sub>3</sub>) to give 609 mg (82%) of **2** as a white solid. Yield range (2 runs): 82-90%; average: 86%.

**<sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):** δ 8.46 (s, 1H), 7.42 (d, *J* = 4.5 Hz, 1H), 7.23 (d, *J* = 4.6 Hz, 1H), 7.21 (d, *J* = 4.6 Hz, 1H), 5.19 (app t, *J* = 4.7 Hz, 1H), 4.99 (dd, *J* = 9.2, 5.0 Hz, 1H), 4.71 (td, *J* = 10.1, 5.0 Hz, 1H), 4.49 – 4.41 (m, 2H), 1.51 (s, 9H), 1.46 (s, 9H).

**<sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):** δ 155.5, 148.0, 123.3, 116.4, 116.3, 109.4, 101.1, 81.1, 75.9, 73.9, 73.3, 66.9, 27.1 (3C, <sup>1</sup>Butyl), 26.9 (3C, <sup>1</sup>Butyl), 22.2, 20.0.

**HRMS:** C<sub>20</sub>H<sub>30</sub>N<sub>5</sub>O<sub>4</sub>Si Calculated for [M+H]<sup>+</sup>: 432.2067; Found: 432.2056.

### N'-(7-((4aR,6R,7R,7aS)-2,2-di-tert-butyl-6-cyano-7-hydroxytetrahydro-4H-furo[3,2-d][1,3,2]dioxasilin-6-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (**3a/b**):



To a flame-dried flask under Ar was added **2** (25.1 mg, 0.0579 mmol), *N,N*-dimethylformamide dimethyl acetal (27.1 mg, 0.231 mmol), and DMF (1 mL). The reaction mixture was stirred at room temperature for 48 hours. The reaction was quenched with water (2 mL) and was diluted with EtOAc (20 mL). The EtOAc layer was subsequently washed with brine (3 X 20 mL) and dried over sodium sulfate. The drying agent was removed by vacuum filtration and the filtrate was concentrated *in vacuo*. The crude product was purified by SiO<sub>2</sub> chromatography

(gradient of 0 – 100% EtOAc in CH<sub>2</sub>Cl<sub>2</sub>) to give 19 mg (68%) of **3a/b** (mixture of separable isomers) as a yellow solid. Yield range (2 runs): 64-68%; average: 66%.

**Isomer 3a:** (R<sub>f</sub> 0.40; 4:1 EtOAc:CH<sub>2</sub>Cl<sub>2</sub>)

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.80 (s, 1H), 7.97 (s, 1H), 6.88 (d, *J* = 4.5 Hz, 1H), 6.85 (d, *J* = 4.5 Hz, 1H), 5.71 (s, 1H), 4.78 (d, *J* = 8.7 Hz, 1H), 4.46 – 4.36 (m, 2H), 4.16 – 4.06 (m, 2H), 3.26 (s, 3H), 3.25 (s, 3H), 1.11 (s, 9H), 0.98 (s, 9H).

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):** δ 160.6, 158.1, 147.5, 124.7, 123.1, 116.0, 111.4, 103.4, 79.6, 79.3, 78.7, 75.6, 67.6, 41.8, 35.5, 27.5 (3C, <sup>1</sup>Butyl), 27.1 (3C, <sup>1</sup>Butyl), 22.7, 20.2.

**HRMS:** Calculated for  $C_{23}H_{35}N_6O_4Si$   $[M+H]^+$ : 487.2489; Found: 487.2471.

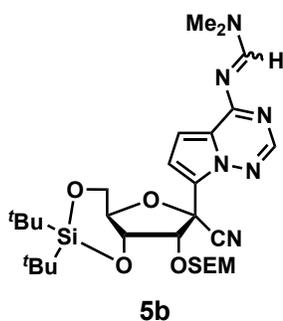
**Isomer 3b:**(Rf 0.35; 4:1 EtOAc:CH<sub>2</sub>Cl<sub>2</sub>)

**<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>): δ 8.77 (s, 1H), 8.02 (s, 1H), 6.84 – 6.81 (m, *J* = 0.9 Hz, 2H), 4.73 (d, *J* = 4.8 Hz, 1H), 4.47 (dd, *J* = 9.2, 4.8 Hz, 1H), 4.31 – 4.19 (m, 2H), 3.98 (app t, *J* = 9.5 Hz, 1H), 3.51 (s, 1H, OH), 3.18 (s, 3H), 3.16 (s, 3H), 0.99 (s, 9H), 0.98 (s, 9H).

**<sup>13</sup>C NMR** (126 MHz, CDCl<sub>3</sub>): δ 160.6, 157.7, 147.4, 123.4, 122.8, 115.6, 111.9, 102.5, 82.1, 77.3, 77.0, 76.7, 75.9, 75.1, 73.6, 67.3, 41.5, 35.3, 27.3 (3C, <sup>t</sup>Butyl), 27.2 (3C, <sup>t</sup>Butyl), 22.6, 20.4.

**HRMS:** Calculated for  $C_{23}H_{35}N_6O_4Si$   $[M+H]^+$ : 487.2489; Found: 487.2472.

**N'-(7-((4aR,6R,7R,7aR)-2,2-di-tert-butyl-6-cyano-7-((2-(trimethylsilyl)ethoxy)methoxy)tetrahydro-4H-furo[3,2-d][1,3,2]dioxasilin-6-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (5b):**



To a flame-dried flask under Ar was added **3a** (250 mg, 0.513 mmol), Bu<sub>4</sub>NI (380 mg, 1.02 mmol) and THF (5 mL). To the suspension, <sup>t</sup>BuMgCl (1M in THF solution) (0.672 mL, 5.64 mmol) was added and stirred for 10 minutes followed by the dropwise addition of SEM-Cl (0.171 g, 1.02 mmol) dissolved in THF (5 mL). The reaction stirred for 48 hours at room temperature. The reaction was quenched with water (5 mL) and diluted with EtOAc (40 mL). The EtOAc layer was subsequently washed with brine (1 X 50 mL) and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and then

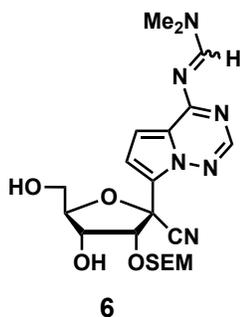
concentrated *in vacuo*. Purification by SiO<sub>2</sub> chromatography (gradient of 0 – 50% EtOAc in CH<sub>2</sub>Cl<sub>2</sub>) gave 251 mg (79%) of **5b** as a yellow solid. Yield range (2 runs): 68-79%; average: 74%.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.81 (s, 1H), 8.04 (s, 1H), 7.04 (d, *J* = 4.5 Hz, 1H), 6.87 (d, *J* = 4.5 Hz, 1H), 5.46 (d, *J* = 8.8 Hz, 1H), 4.82 (d, *J* = 6.8 Hz, 1H), 4.67 – 4.55 (m, 2H), 4.45 (app t, *J* = 9.2 Hz, 1H), 4.34 (dd, *J* = 9.2, 5.1 Hz, 1H), 4.04 (app t, *J* = 9.9 Hz, 1H), 3.55 (td, *J* = 11.6, 9.7 Hz, 1H), 3.42 (td, *J* = 11.6, 9.7, 1H), 3.23 (s, 3H), 3.20 (s, 3H), 1.09 (s, 9H), 1.04 (s, 9H), 0.72 (td, *J* = 13.6, 11.5 Hz, 1H), 0.59 (td, *J* = 13.6, 11.5 Hz, 1H), -0.08 (s, 9H).

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):** δ 160.9, 157.6, 147.5, 124.2, 123.2, 116.8, 113.9, 102.4, 94.2, 80.0, 79.5, 75.5, 67.8, 65.6, 41.5, 35.3, 27.5 (3C, <sup>t</sup>Butyl), 27.1 (3C, <sup>t</sup>Butyl), 22.7, 20.3, 20.0, 17.8, -1.3 (3C, -Si(CH<sub>3</sub>)<sub>3</sub>).

**HRMS:** Calculated for  $C_{29}H_{49}N_6O_5Si_2$   $[M+H]^+$ : 617.3303; Found: 617.3274.

**N'-(7-((2R,3R,4R,5R)-2-cyano-4-hydroxy-5-(hydroxymethyl)-3-((2-(trimethylsilyl)ethoxy)methoxy)tetrahydrofuran-2-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (6):**



To a flame-dried flask under Ar was added **5b** (250 mg, 0.405 mmol) and anhydrous pyridine (5 mL). HF-pyridine (Olah's reagent) (0.115 mL, 4.05 mmol) was added dropwise to the reaction mixture and stirred at room temperature for 1 hour. The reaction was quenched with water (5 mL) and diluted with CH<sub>2</sub>Cl<sub>2</sub> (40 mL). The CH<sub>2</sub>Cl<sub>2</sub> layer was subsequently washed with brine (1 X 20 mL) and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> followed by concentration *in vacuo*. The crude product was purified by SiO<sub>2</sub> chromatography (gradient of 0 – 40% EtOAc in CH<sub>2</sub>Cl<sub>2</sub>) to give 152 mg (79%) **6** as a

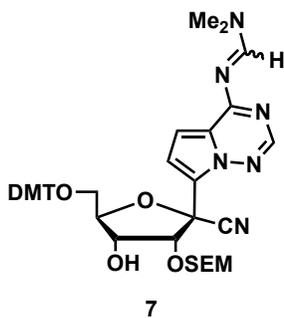
yellow solid. Yield range (2 runs): 79-85%; average: 82%.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.82 (s, 1H), 8.10 (s, 1H), 7.02 (d, *J* = 4.6 Hz, 1H), 6.89 (d, *J* = 4.6 Hz, 1H), 5.19 (d, *J* = 7.1 Hz, 1H), 4.83 (d, *J* = 7.3 Hz, 1H), 4.68 (d, *J* = 7.3 Hz, 1H), 4.53 – 4.42 (m, 2H), 3.95 (dd, *J* = 12.6, 3.1 Hz, 1H), 3.92 – 3.83 (m, 2H), 3.57 (app q, *J* = 9.2 Hz, 1H), 3.24 (s, 3H), 3.22 (s, 3H), 0.90 (t, *J* = 9.2 Hz, 2H), 0.01 (s, 9H).

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):** δ 160.9, 157.8, 147.6, 124.1, 122.5, 117.1, 114.0, 102.5, 95.4, 85.4, 83.7, 77.6, 74.2, 66.7, 61.6, 41.6, 35.4, 18.0, -1.3 (3C, -Si(CH<sub>3</sub>)<sub>3</sub>).

**HRMS:** Calculated for C<sub>21</sub>H<sub>33</sub>N<sub>6</sub>O<sub>5</sub>Si [M+H]<sup>+</sup>: 477.2282; Found: 477.2261.

**N'-(7-((2R,3R,4R,5R)-5-((bis(4-methoxyphenyl)(phenyl)methoxy)methyl)-2-cyano-4-hydroxy-3-((2-(trimethylsilyl)ethoxy)methoxy)tetrahydrofuran-2-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (7):**



To a flame-dried flask under Ar was added 4,4'-dimethoxytrityl chloride (0.159 g, 0.472 mmol), **6** (150 mg, 0.314 mmol), 4-dimethylaminopyridine (42.1 mg, 0.345 mmol) and anhydrous pyridine (2 mL). The reaction mixture was stirred for 18 hours at room temperature. The reaction mixture was concentrated *in vacuo* and purified by chromatography on neutralized SiO<sub>2</sub> (neutralized by washing with 2 column volumes of 10% Et<sub>3</sub>N in CH<sub>2</sub>Cl<sub>2</sub>) using a gradient of 0 – 50% acetone in CH<sub>2</sub>Cl<sub>2</sub> to give 165 mg (67%) of **7** as a yellow solid. Yield range (2 runs): 66-67%;

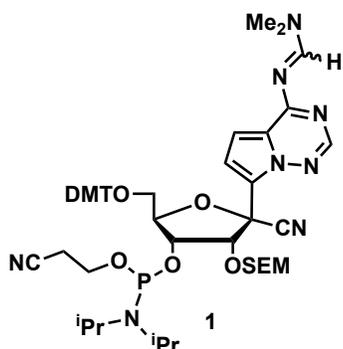
average: 66%.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.82 (s, 1H), 8.09 (s, 1H), 7.56 – 7.51 (m, 2H), 7.45 – 7.38 (m, 4H), 7.32 – 7.25 (m, 2H), 7.23 – 7.16 (m, 1H), 7.09 (d, *J* = 4.6 Hz, 1H), 6.91 (d, *J* = 4.6 Hz, 1H), 6.87 – 6.80 (m, 4H), 5.22 (d, *J* = 7.2 Hz, 1H), 4.83 (d, *J* = 7.2 Hz, 1H), 4.66 (d, *J* = 7.2 Hz, 1H), 4.62 (td, *J* = 7.6, 2.5 Hz, 1H), 4.53 – 4.46 (m, 1H), 3.98 (d, *J* = 3.2 Hz, 1H), 3.87 – 3.79 (m, 1H), 3.78 (s, 6H), 3.60 – 3.51 (m, 1H), 3.48 (dd, *J* = 10.5, 4.0 Hz, 1H), 3.40 (dd, *J* = 10.5, 4.0 Hz, 1H), 3.24 (s, 3H), 3.20 (s, 3H), 0.87 (t, *J* = 9.2 Hz, 2H), -0.02 (s, 9H).

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):** δ 160.9, 158.5, 157.6, 147.4, 144.9, 136.2 (2C), 130.2 (2C), 130.2 (2C), 129.2, 128.4 (2C), 127.9 (2C), 126.8, 124.1, 123.1, 117.1, 114.1, 113.2 (4C), 102.4, 95.2, 86.3, 84.9, 82.5, 77.6, 75.3, 66.4, 62.7, 55.3 (2C), 41.5, 35.3, 18.2, -0.7 (3C, -Si(CH<sub>3</sub>)<sub>3</sub>).

**HRMS:** Calculated for C<sub>42</sub>H<sub>51</sub>N<sub>6</sub>O<sub>7</sub>Si [M+H]<sup>+</sup>: 779.3588; Found: 779.3578.

**(2R,3R,4R,5R)-2-((bis(4-methoxyphenyl)(phenyl)methoxy)methyl)-5-cyano-5-(4-(((E)-  
(dimethylamino)methylene)amino)pyrrolo[2,1-f][1,2,4]triazin-7-yl)-4-((2-(trimethylsilyl)ethoxy)-  
methoxy)tetrahydrofuran-3-yl (2-cyanoethyl) diisopropylphosphoramidite (1):**



To a flame-dried flask under Ar was added **7** (61.1 mg, 0.0771 mmol), DIPEA (71.1 μL, 0.423 mmol) and 1,2-DCE (1 mL). The reaction was stirred at room temperature for 10 minutes. Next, 2-cyanoethyl diisopropylchlorophosphorodiamidite (25.1 μL, 0.115 mmol) was added dropwise and stirred for 2 hours at room temperature. The reaction mixture was concentrated *in vacuo* and purified by chromatography on neutralized SiO<sub>2</sub> (neutralized by washing with 2 column volumes of 10% Et<sub>3</sub>N in CH<sub>2</sub>Cl<sub>2</sub>) using a gradient from 0 – 50% EtOAc in CH<sub>2</sub>Cl<sub>2</sub> to give 50 mg (66%) of **1** as a white solid. Yield range (2 runs): 54-66%; average: 60%.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.89 – 8.71 (m, 1H), 8.14 – 8.08 (m, 1H), 7.57 – 7.49 (m, 2H), 7.46 – 7.37 (m, 4H), 7.33 – 7.25 (m, 2H), 7.24 – 7.16 (m, 1H), 7.03 (d, *J* = 4.5 Hz, 0.6H), 6.98 (d, *J* = 4.5 Hz, 0.4H), 6.90 (dd, *J* = 4.6, 1.3 Hz, 1H), 6.87 – 6.80 (m, 4H), 5.46 (d, *J* = 4.6 Hz, 0.6H), 5.43 (d, *J* = 4.6 Hz, 0.4H), 4.85 (dd, *J* = 9.8, 6.2 Hz, 0.8H), 4.78 (dd, *J* = 9.8, 6.2 Hz, 1.2H), 4.72 (dt, *J* = 10.2, 5.7 Hz, 0.6H), 4.59 (dt, *J* = 10.2, 4.4 Hz, 0.4H), 4.49 (app q, *J* = 4.8 Hz, 0.6H), 4.46 (app q, *J* = 4.8 Hz, 0.4H), 3.82 – 3.75 (m, 6H), 3.69 – 3.27 (m, 8H), 3.24 (s, 3H), 3.21 (s, 3H), 2.43 (t, *J* = 6.6 Hz, 1H), 2.36 – 2.21 (m, 1H), 1.09 (d, *J* = 6.8 Hz, 3.5H), 1.05 (d, *J* = 6.8 Hz, 2.5H), 0.99 (d, *J* = 6.7 Hz, 3.5H), 0.94 (d, *J* = 6.7 Hz, 2.5H), 0.92 – 0.55 (m, 2H), -0.02 (s, 3.5H), -0.04 (s, 5.5H). (60:40 ratio of phosphorous diastereomers in based on <sup>31</sup>P NMR).

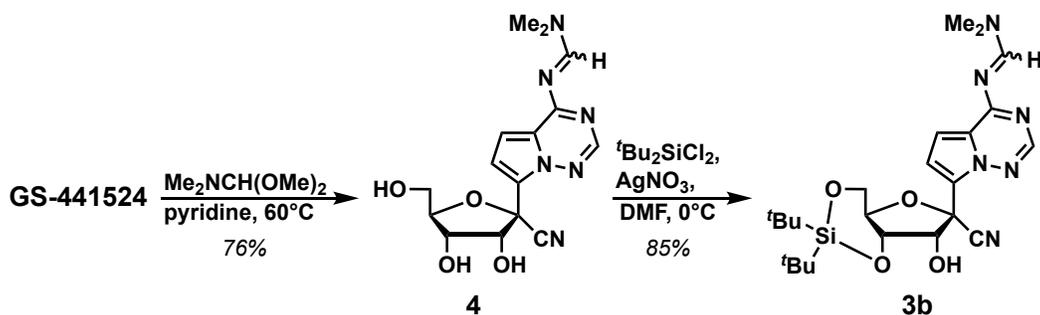
**<sup>31</sup>P NMR (202 MHz, CDCl<sub>3</sub>):** δ 150.13, 149.71.

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) (major phosphorous diastereomer):** δ 160.8, 158.5, 157.5, 147.4, 144.9, 136.2, 136.2, 130.4 (2C), 130.3 (2C), 128.6, 127.9 (2C), 126.8, 123.6, 123.3, 117.4, 116.7, 113.3, 113.2 (4C), 102.5, 94.7, 86.3, 84.9, 83.8, 82.1, 82.0, 78.8, 77.8, 66.0, 62.7, 58.5, 58.1, 55.3 (2C), 43.3, 41.5, 35.3, 24.6, 24.5, 24.4, 20.0, 19.9, 17.86, -1.2 (3C, -Si(CH<sub>3</sub>)<sub>3</sub>).

**(minor phosphorous diastereomer):** δ 160.7, 158.5, 157.6, 147.4, 144.9, 136.2, 136.1, 130.3 (2C), 130.3 (2C), 128.4, 127.9 (2C), 126.8, 123.5, 123.4, 117.5, 116.4, 113.2, 112.8 (4C), 102.6, 93.5, 86.3, 84.9, 83.2, 82.1, 81.5, 79.4, 77.9, 65.9, 62.9, 58.4, 58.3, 55.3 (2C), 43.2, 41.5, 35.3, 24.6, 24.6, 24.4, 20.1, 19.9, 18.0, -0.3 (3C, -Si(CH<sub>3</sub>)<sub>3</sub>).

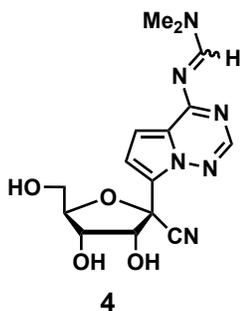
**HRMS:** Calculated for C<sub>51</sub>H<sub>68</sub>N<sub>8</sub>O<sub>8</sub>PSi [M+H]<sup>+</sup>: 979.4667; Found: 979.4622.

### III. Protecting Group Studies Towards the Synthesis of the GS-441524 Phosphoramidite.



**Scheme 1.** Synthesis of **3b**.

**N'-(7-((2R,3R,4S,5R)-2-cyano-3,4-dihydroxy-5-(hydroxymethyl)tetrahydrofuran-2-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (4).**



To a flame-dried flask under Ar was added **GS-441524** (100 mg, 0.343 mmol), *N,N*-dimethylformamide dimethyl acetal (0.171 g, 1.44 mmol), and pyridine (1 mL). The reaction mixture was stirred at room temperature for 24 hours. The reaction mixture was then concentrated *in vacuo* to remove pyridine and was diluted with EtOAc (80 mL). The EtOAc layer was subsequently washed with brine (3 X 50 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude product was purified by SiO<sub>2</sub> chromatography (gradient 0 – 20% CH<sub>3</sub>OH in CHCl<sub>3</sub>) to give 100 mg (85%) of **4**

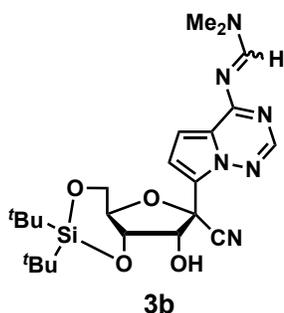
as an off-white solid. Yield range (2 runs): 68-85%; average: 76%.

**<sup>1</sup>H NMR** (500 MHz, DMSO-*d*<sub>6</sub>): δ 9.45 (s, 1H), 8.66 (s, 1H), 7.51 (d, *J* = 4.5 Hz, 1H), 7.35 (d, *J* = 4.5 Hz, 1H), 6.64 (d, *J* = 6.3 Hz, 1H), 5.74 (d, *J* = 5.1 Hz, 1H), 5.44 (app t, *J* = 5.7 Hz, 1H), 5.21 (app t, *J* = 5.8 Hz, 1H), 4.61 (app q, *J* = 4.5 Hz, 1H), 4.52 (app q, *J* = 5.2 Hz, 1H), 4.22 – 4.14 (m, 1H), 4.09 – 4.01 (m, 1H), 3.75 (s, 3H), 3.69 (s, 3H).

**<sup>13</sup>C NMR** (126 MHz, DMSO-*d*<sub>6</sub>): δ 160.2, 158.1, 147.3, 123.8, 122.6, 117.3, 111.9, 101.8, 85.5, 78.4, 74.2, 70.1, 60.9, 41.1, 34.9.

**HRMS:** Calculated for C<sub>15</sub>H<sub>19</sub>N<sub>6</sub>O<sub>4</sub> [M+H]<sup>+</sup>: 347.1468; Found: 347.1455.

**N'-(7-((4aR,6R,7R,7aS)-2,2-di-*tert*-butyl-6-cyano-7-hydroxytetrahydro-4H-furo[3,2-d][1,3,2]dioxasilin-6-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (3b):**

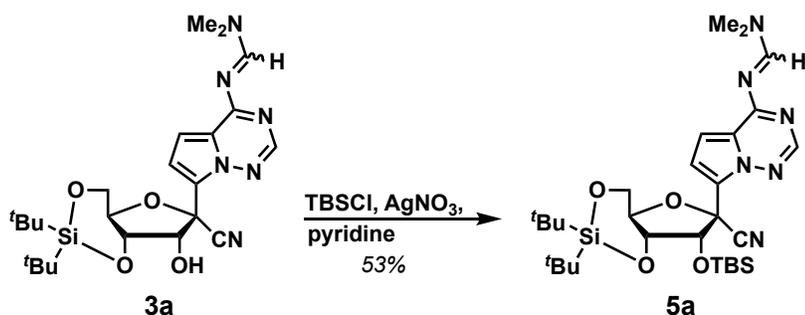


To a flame-dried flask under Ar was added **4** (100 mg, 0.288 mmol), di-*tert*-butyl dichlorosilane (91.1 mL, 0.433 mmol), AgNO<sub>3</sub> (73.5 mg, 0.433 mmol), and DMF (2 mL). The reaction mixture was stirred at 0 °C for 90 minutes. The reaction was quenched with water (5 mL) and diluted with EtOAc (40 mL). The EtOAc layer was subsequently washed with brine (3 X 40 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude product was purified by SiO<sub>2</sub> chromatography

(gradient 0 – 100% EtOAc in CH<sub>2</sub>Cl<sub>2</sub>) to give 120 mg (85%) of **3b** as an off-white solid. Yield range (2 runs): 85-85%; average: 85%.

<sup>1</sup>H and <sup>13</sup>C NMR are shown in **Section II**.

**N'-(7-((4aR,6R,7R,7aR)-2,2-di-tert-butyl-7-((tert-butyldimethylsilyl)oxy)-6-cyanotetrahydro-4H-furo[3,2-d][1,3,2]dioxasilin-6-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl)-N,N-dimethylformimidamide (5a):**



To a flame-dried flask under Ar was added **3a** (40.5 mg, 0.0822 mmol), *t*-butyldimethylsilylchloride (17.5 mg, 0.115 mmol), AgNO<sub>3</sub> (19.5 mg, 0.115 mmol), and pyridine (1 mL). The reaction mixture was stirred at room temperature for 48 hours under dark conditions; the

reaction flask was wrapped in aluminum foil. The reaction mixture was concentrated *in vacuo* to remove pyridine, followed by the dilution in CH<sub>2</sub>Cl<sub>2</sub> (40 mL). The CH<sub>2</sub>Cl<sub>2</sub> layer was subsequently washed with saturated NaHCO<sub>3</sub> (3 X 30 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo*. The crude product was purified by SiO<sub>2</sub> chromatography (gradient of 0 – 50% EtOAc in CH<sub>2</sub>Cl<sub>2</sub> to give 26 mg (53%) of **5a** as a yellow solid. Yield range (2 runs): 50-53%; average: 52%.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):** δ 8.84 (s, 1H), 8.03 (s, 1H), 7.04 (d, *J* = 4.6 Hz, 1H), 6.89 (d, *J* = 4.6 Hz, 1H), 5.38 (d, *J* = 8.7 Hz, 1H), 4.66 (td, *J* = 10.4, 5.1 Hz, 1H), 4.39 – 4.32 (m, 2H), 4.05 (dd, *J* = 10.7, 9.1 Hz, 1H), 3.25 (s, 3H), 3.22 (s, 3H), 1.09 (s, 9H), 1.06 (s, 9H), 0.81 (s, 9H), -0.16 (s, 3H), -0.23 (s, 3H).

**<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>):** δ 161.0, 157.8, 147.4, 124.0, 123.2, 117.0, 114.6, 102.4, 80.7, 78.8, 77.0, 75.6, 68.0, 41.6, 35.4, 27.4 (3C, <sup>t</sup>Butyl), 27.2 (3C, <sup>t</sup>Butyl), 25.5 (3C, <sup>t</sup>Butyl), 22.7, 20.3, 18.0, -5.0, -5.2.

**HRMS:** Calculated for C<sub>29</sub>H<sub>49</sub>N<sub>6</sub>O<sub>4</sub>Si<sub>2</sub> [M+H]<sup>+</sup>: 601.3354; Found: 601.3318

#### IV. Preparation of GS-441524-Containing RNA.

RNA was synthesized using standard solid-phase synthesis on a BioAutomation MerMade4 DNA/RNA synthesizer. Remdesivir-containing phosphoramidite **1** was synthesized as described in **Section II**. All other phosphoramidites and reagents were purchased from Glen Research Corporation. The CPG universal solid support (1.0 μmol scale; MM1-3500) was purchased from BioAutomation Technologies. Phosphoramidites and reagents were freshly prepared prior to synthesis. Trap packs (NC1758846) were added to the anhydrous acetonitrile (40-4050-50, Glen Research) and activator (30-3140-57, Glen Research) solutions. Automated RNA synthesis was paused immediately prior to the incorporation of **1**. Freshly prepared **1** (30.0 mg) was dissolved in anhydrous acetonitrile (200.0 μL, ~50 mM) and was loaded onto the instrument prior to incorporation for on-system coupling. Reaction conditions for the synthesis of

**RNA-1** were optimized based on vendor recommendations and literature precedence (**Supplemental Table 1**).<sup>1-5</sup>

**Supplemental Table 1.** Reagent table of optimized coupling conditions used for RNA synthesis.

Composition of Reagent	Reaction Time on MerMade4	Number of times reagent added during one base coupling	Product number from Glen Research
<b>Deblocking Mix:</b> 3 % dichloroacetic acid in dichloromethane	1-minute rxn	2X	40-4040-57
<b>Cap A:</b> Tetrahydrofuran:acetic anhydride:pyridine (80:10:10)	1-minute rxn	2X	40-4110-57
<b>Cap B:</b> 16 % 1-methylimidazole in tetrahydrofuran	1-minute rxn	2X	40-4220-57
<b>Oxidation Solution:</b> 0.02 M I <sub>2</sub> in tetrahydrofuran:pyridine:water (88:10:2)	1-minute rxn	2X	40-4032-57
<b>Activator Solution:</b> 0.25 M 5-ethylthio-1H-tetrazole in anhydrous acetonitrile	12-minute rxn	2X <sup>a</sup> or 3X <sup>b</sup>	30-3140-57

<sup>a</sup> Standard RNA phosphoramidites were coupled onto the resin 2X. <sup>b</sup> RNA phosphoramidite **1** was coupled onto the resin 3X.

Following the synthesis of **RNA-1** (5'-CCGGGCGGCR-3', where R = **1**), the resin was dried under N<sub>2</sub> for 1 hour before being transferred to a fritted reaction vessel. Concentrated aqueous NH<sub>4</sub>OH (2.5 mL) was added, and the aluminum foil-covered vessel was placed in a shaker for 30-hours at room temperature. After the removal of the resin, the solution was filtered into a centrifuge tube and diluted with DNase/RNase-free water (2 mL, cat# 10977015 Thermo Fisher). The solution was transferred into microcentrifuge tubes and concentrated *in vacuo* in a SpeedVac. The crude sample was completely dry before being deprotected with 1.0 M tetrabutylammonium fluoride (TBAF) in tetrahydrofuran (2 mL) for 12-hours in a 4-mL glass vial equipped with a stir bar. The solution was concentrated *in vacuo* and the crude material was purified by HPLC. **RNA-1** was HPLC purified on an Agilent 1200 series instrument equipped with a diode array detector and a PLRP-S column (8 μmol, 100 Å, 4.6 X 150 mm, Agilent Technologies). The analysis method (2.750 mL/min flow rate) involved isocratic 100 mM triethylammonium acetate (TEAA, pH 7.0, Sigma Aldrich; 0-5 minutes, Solvent A) followed by a linear gradient to 10 % 100 mM TEAA:MeCN (1:1, 5-10 minutes, Solvent B) and finally a linear gradient of 10-70% 100 mM TEAA/MeCN (1:1, 10-45 minutes, Solvent B). Wavelengths monitored = 260 nm. After purification **RNA-1** was desalted with DNase/RNase-free water using an Illustra NAP-5 column (Sephadex G-25 DNA grade, GE Healthcare) according to the manufacturer's instructions. The desalted oligoribonucleotide was quantified by UV-vis Spectroscopy (using the molar extinction coefficient of 765 M<sup>-1</sup> cm<sup>-1</sup> for Remdesivir calculated at 260 nm in H<sub>2</sub>O, **Supplemental Information V**) and confirmed by LC-MS. The purity was assessed by HPLC

reinjection of the purified oligonucleotides. LC-MS was performed on a Thermo Scientific LTQ XL instrument equipped with a diode array detector and using a Zorbax C-18 column (5  $\mu\text{m}$ , 80  $\text{\AA}$ , 0.5 X 150 mm, Agilent Technologies). The analysis method (15  $\mu\text{L}/\text{min}$  flow rate) involved 15 mM aqueous  $\text{NH}_4\text{OAc}$  containing 2%  $\text{CH}_3\text{CN}$  followed by a linear gradient of 2-15%  $\text{CH}_3\text{CN}$  (0-15 min) and 25-60%  $\text{CH}_3\text{CN}$  (15-25 min). Wavelengths monitored = 260 nm.

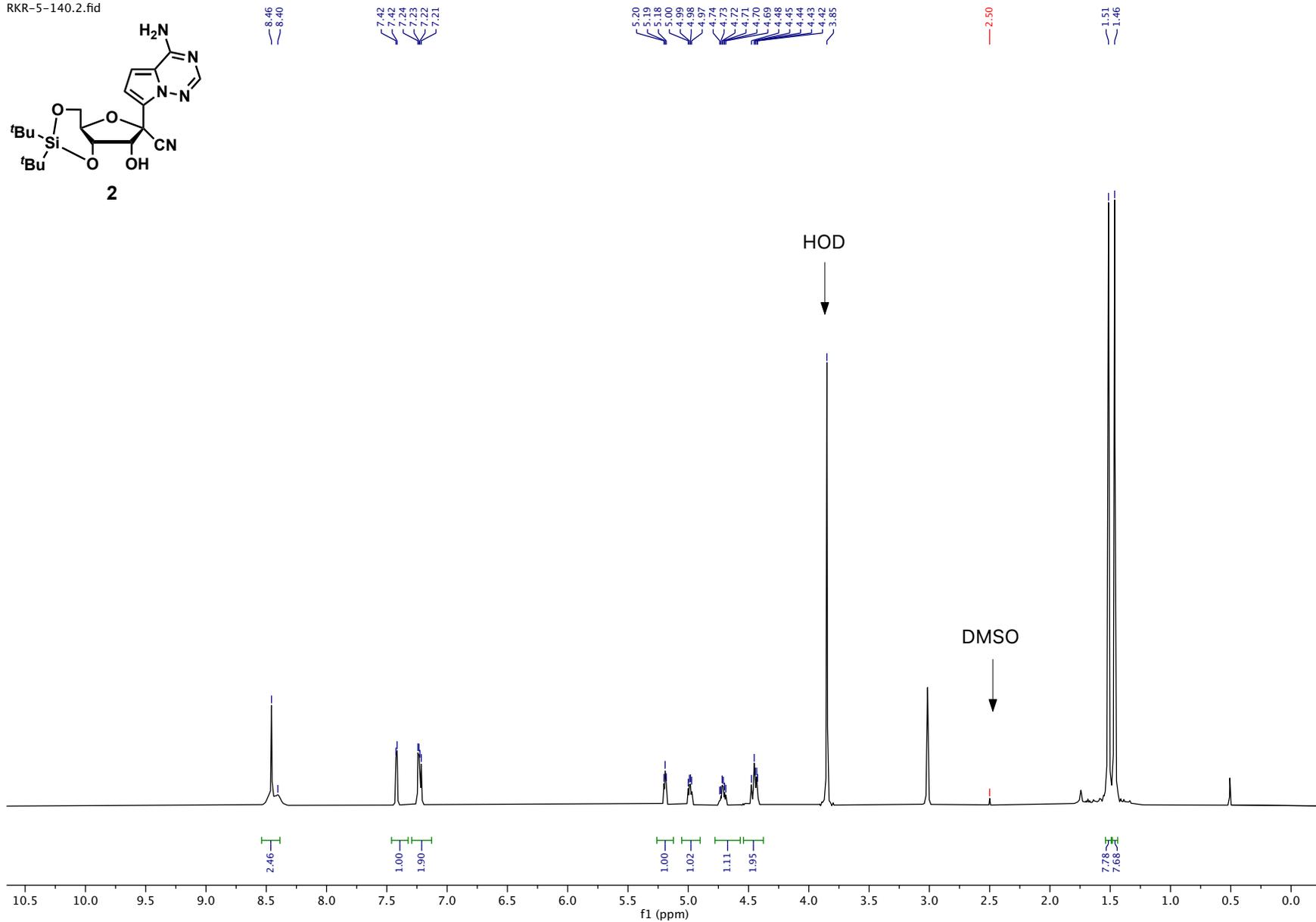
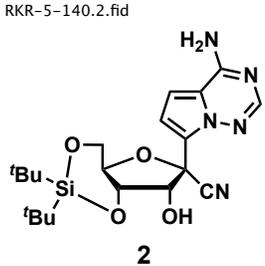
## V. Molar Extinction Coefficient Calculation of Remdesivir.

The extinction coefficient of Remdesivir was calculated as follows: a glass vial was dried *in vacuo* for 48 hours before addition of a known mass of commercial Remdesivir. The charged vial with dried *in vacuo* for an additional 24 hours before dissolving the Remdesivir in  $\text{H}_2\text{O}$  such that the exact concentration of the solution was known. Beer's Law plots were then constructed by taking the solution and diluting serially such that 6 samples of a known concentration could be measured at 260 nm using a UV spectrometer (all absorbances < 1.0) using quartz cuvettes (Starna Cells, Inc., 10 mm path length). The slope of the line absorbance vs concentration (in molar, M) is the extinction coefficient  $\epsilon_{260}$  in  $\text{M}^{-1}\text{cm}^{-1}$ . This process was repeated four times with four different stock solutions, the slopes averaged, and standard deviation calculated using Microsoft Excel. The  $\epsilon_{260}$  calculated for Remdesivir was  $765 \pm 77 \text{ M}^{-1} \text{ cm}^{-1}$  using the nearest-neighbors model equation (**Equation 1**).

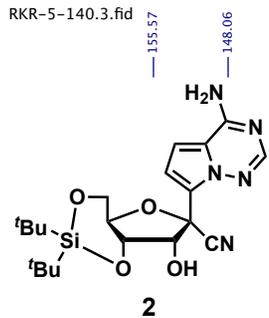
$$\epsilon_{260} = \sum_1^{N-1} \epsilon \text{ Nearest Neighbor} - \sum_1^{N-1} \epsilon \text{ Individual Bases}$$

## VI. NMR Spectroscopic Data.

RKR-5-140.2.fid



RKR-5-140.3.fid



155.57

148.06

123.34

116.45  
116.34

109.44

101.12

81.17

75.90

73.94

73.36

66.93

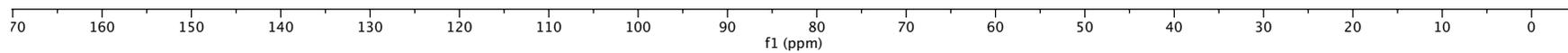
DMSO

27.10

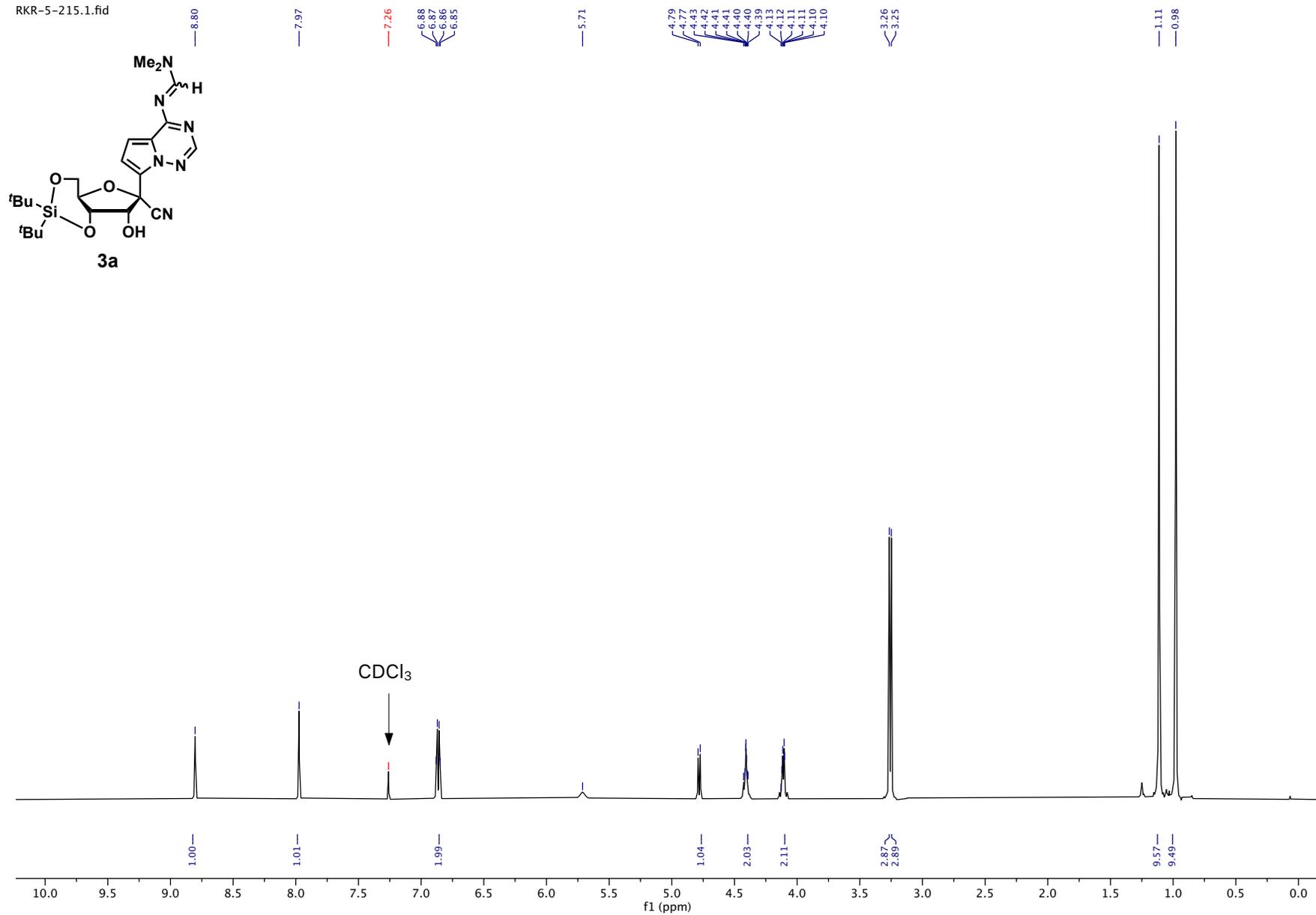
26.93

22.21

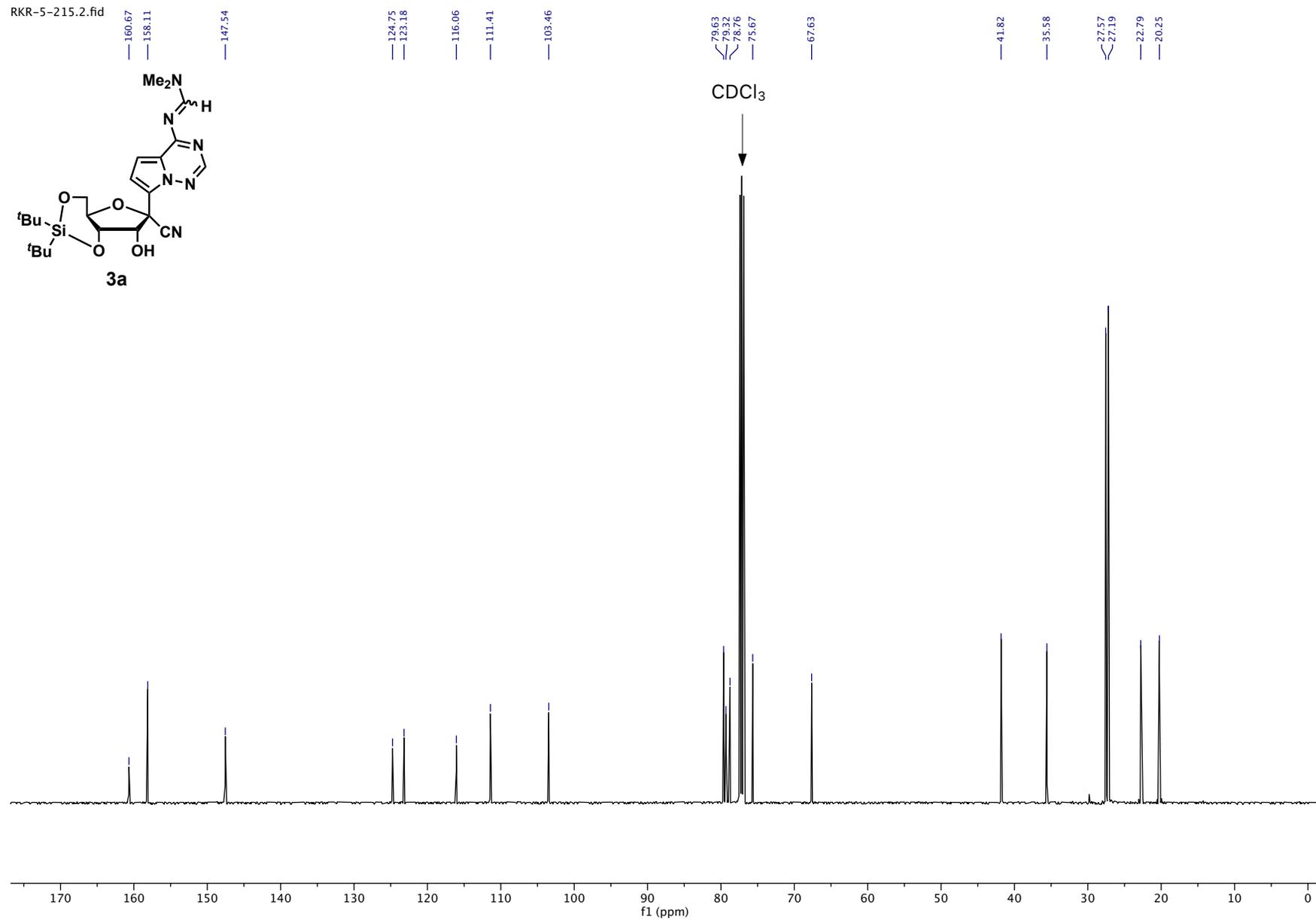
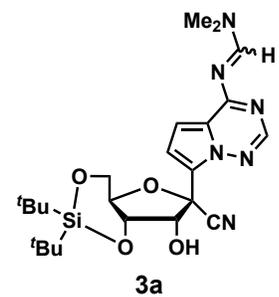
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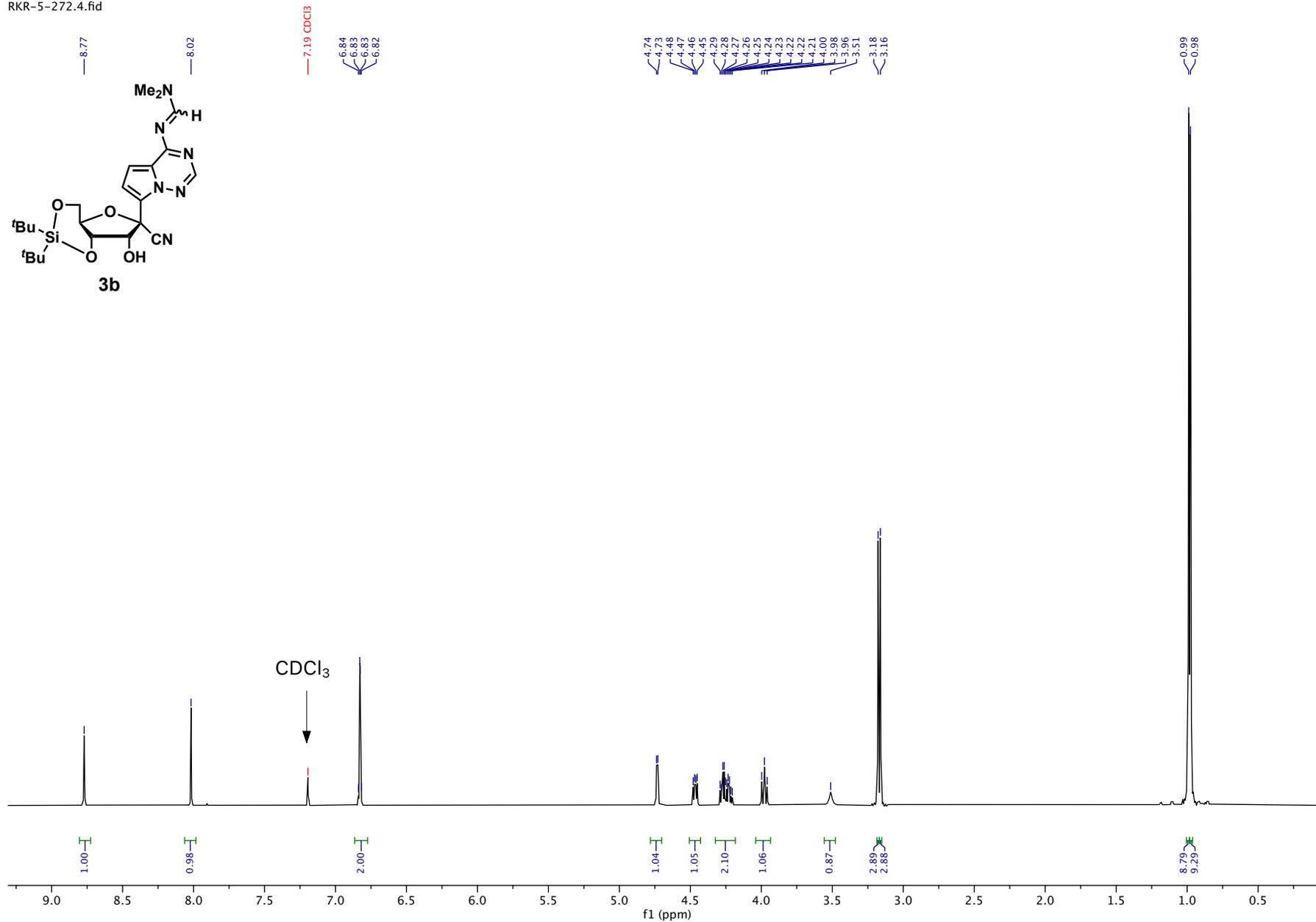
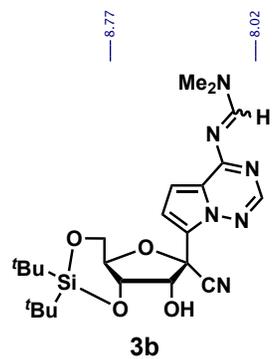
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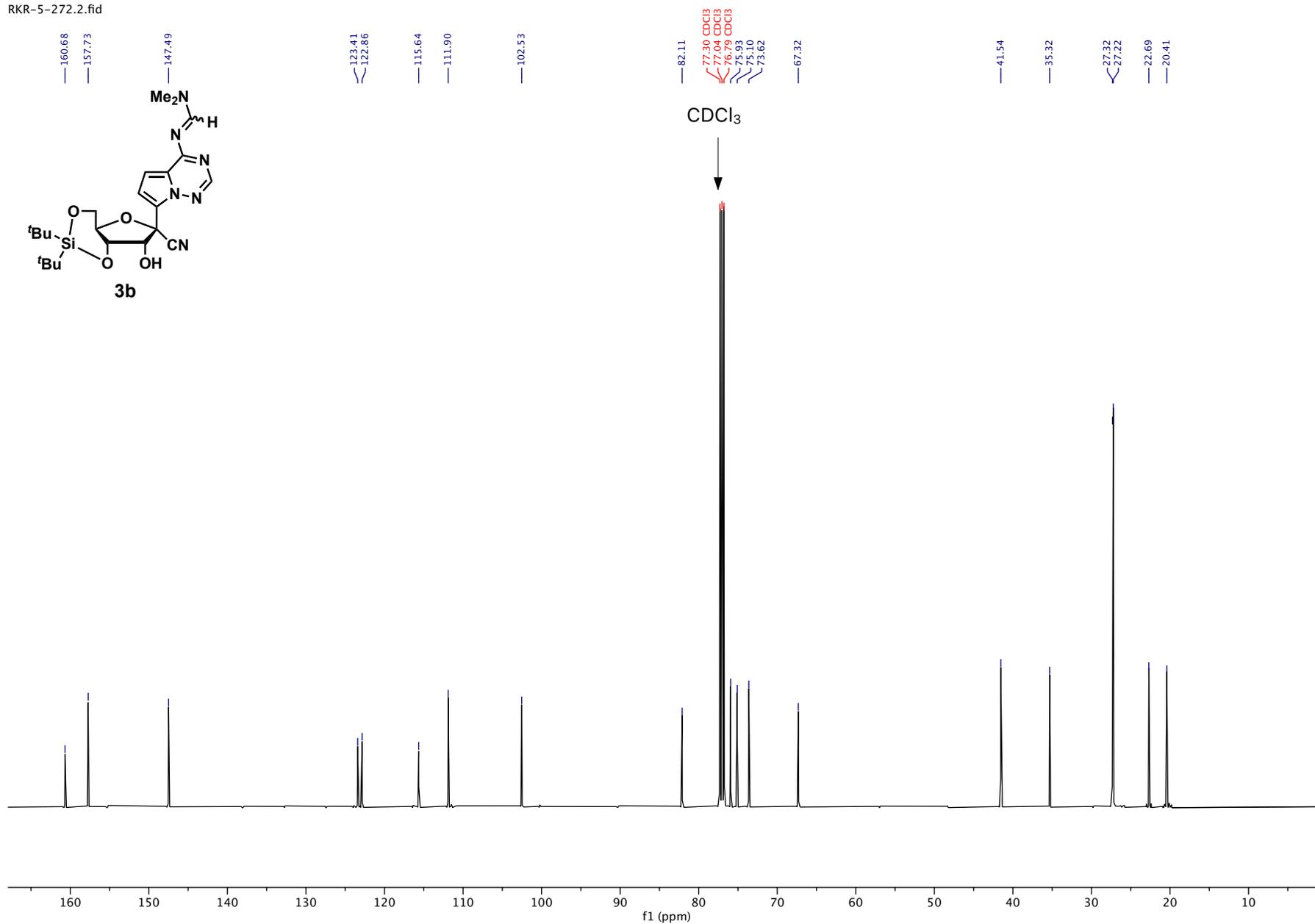
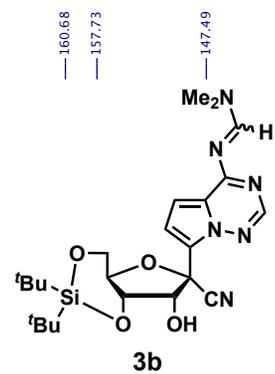
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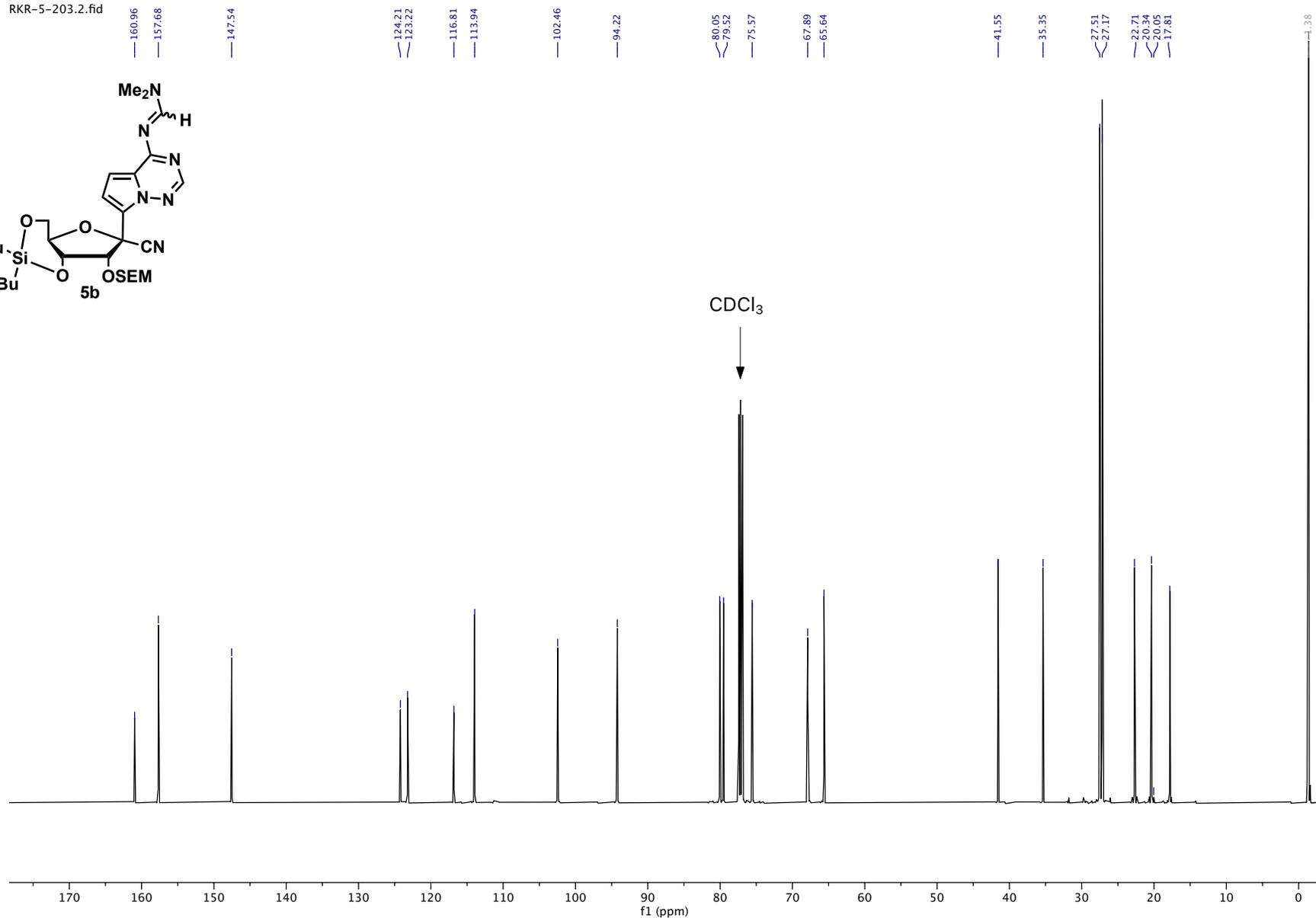
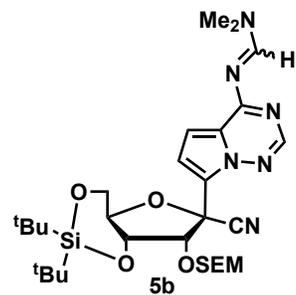


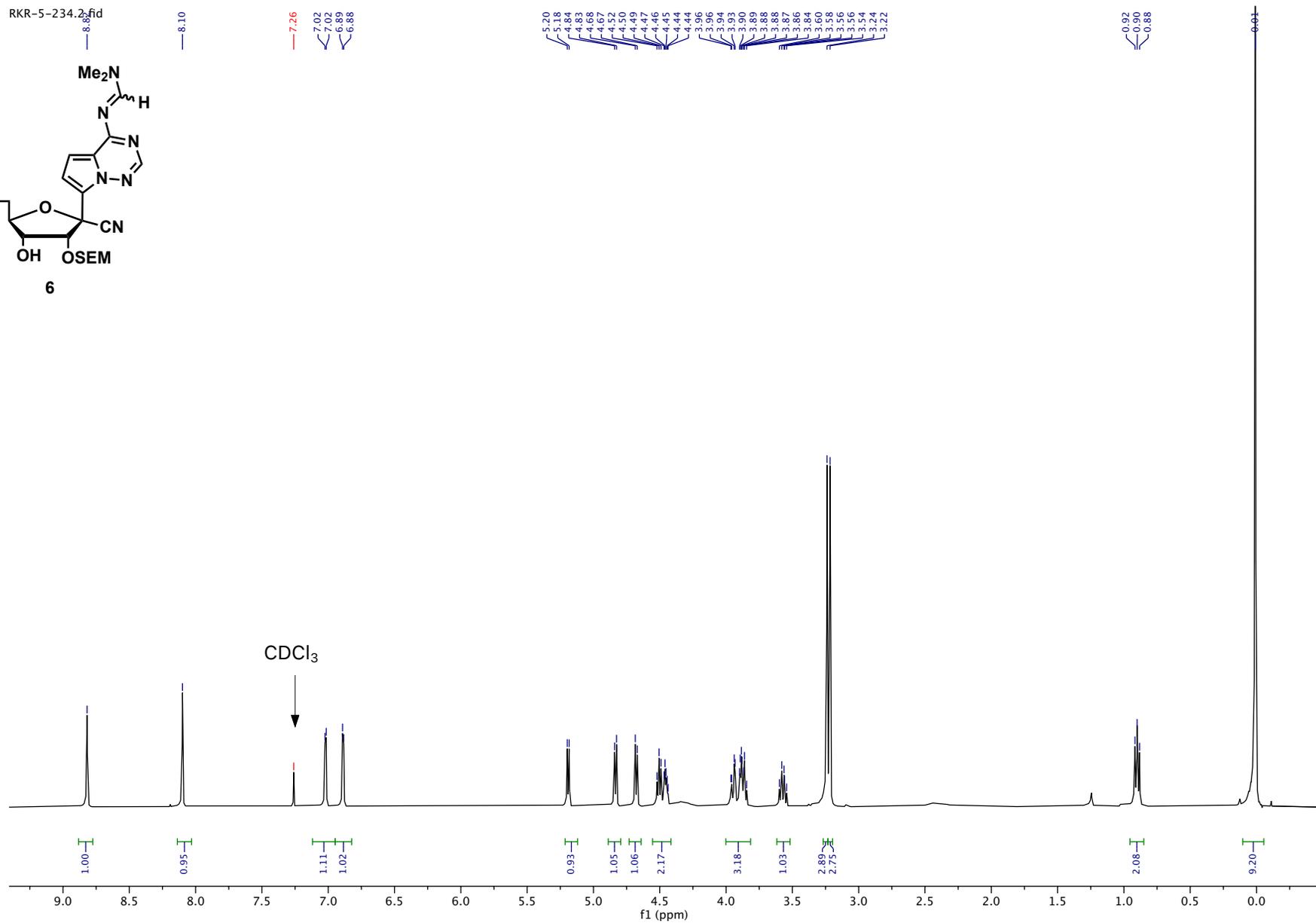
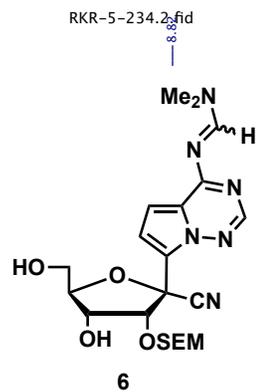
RKR-5-272.2.fid





RKR-5-203.2.fid





RKR-5-234.3.fid

160.96  
157.81

147.60

124.16  
122.53

117.11  
114.03

102.56

95.42

85.47  
83.78

77.61  
74.21

66.71

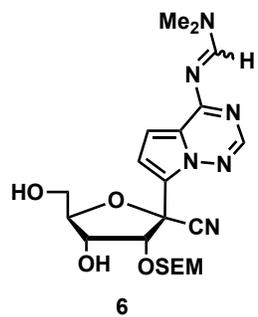
61.64

41.61

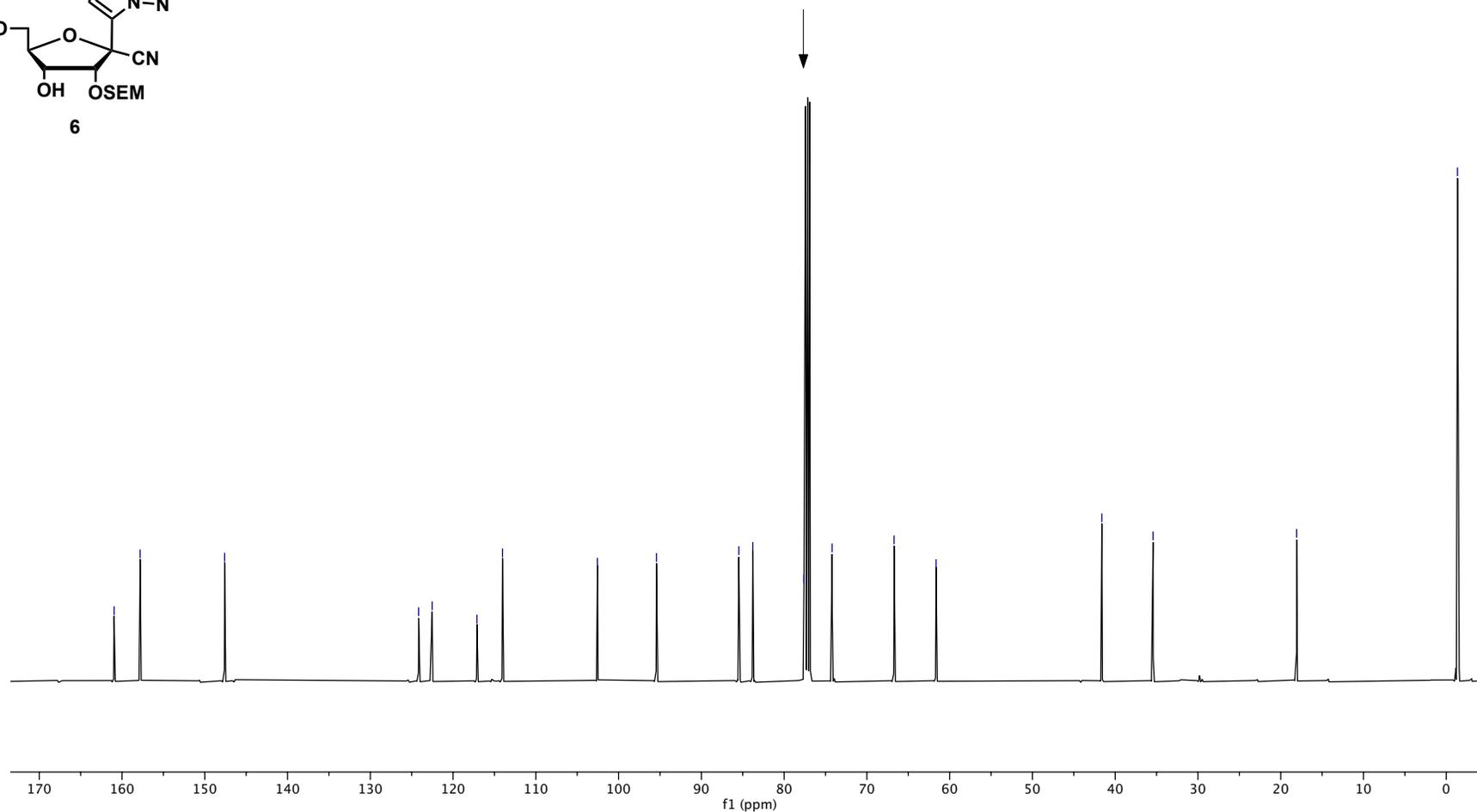
35.42

18.08

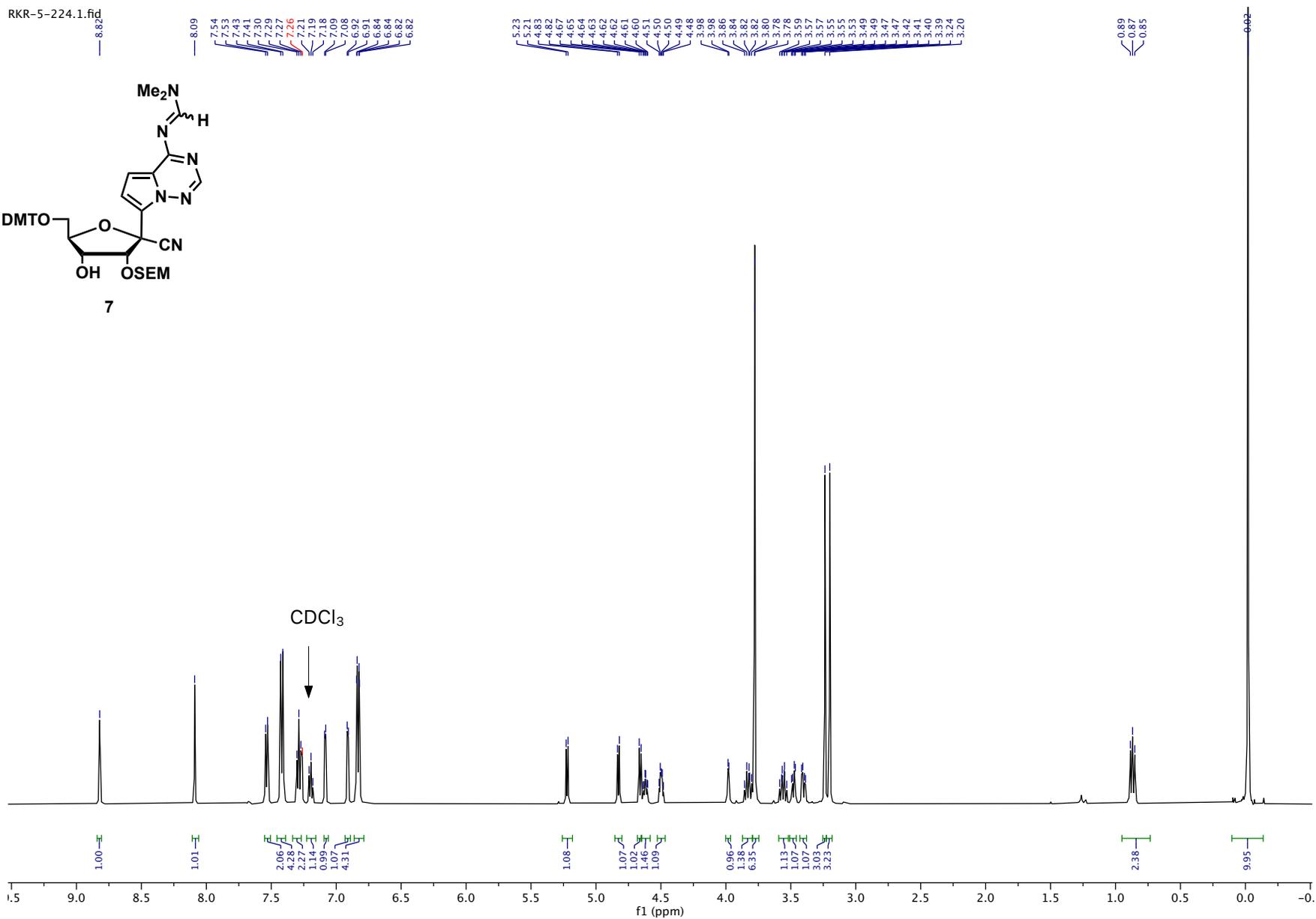
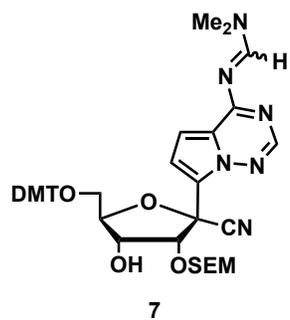
1.36



CDCl<sub>3</sub>



RKR-5-224.1.fid



RKR-5-224.2.fid

160.93  
158.54  
157.69

147.49  
144.91

136.22

130.79  
130.27  
129.25  
128.44  
127.94  
126.81  
124.10  
123.11

117.19  
114.11  
113.26

102.48

95.22

86.30  
84.90  
82.58

77.65  
75.32

66.44  
62.75

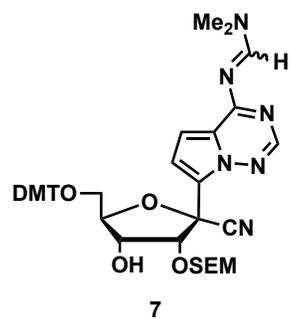
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41.56

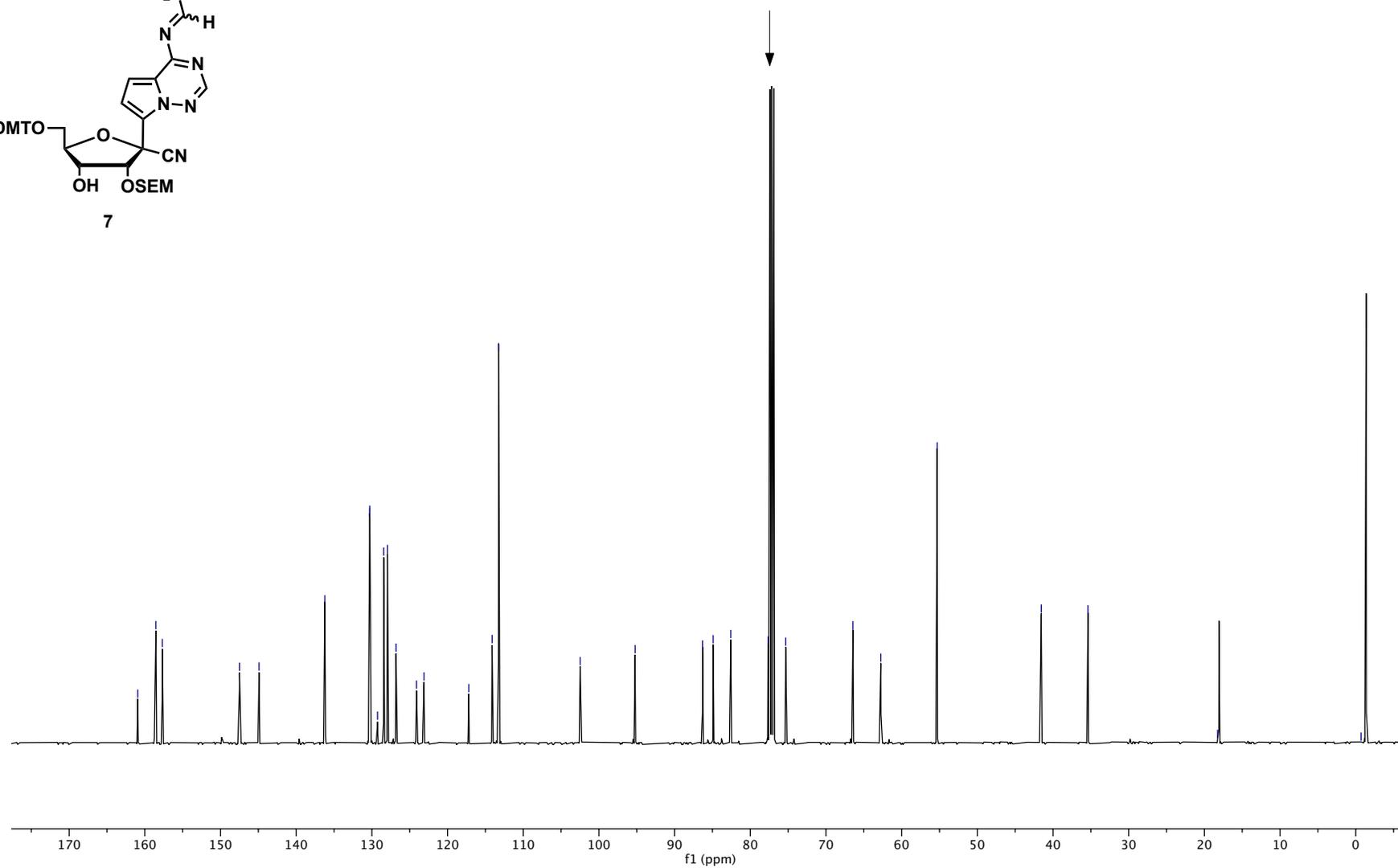
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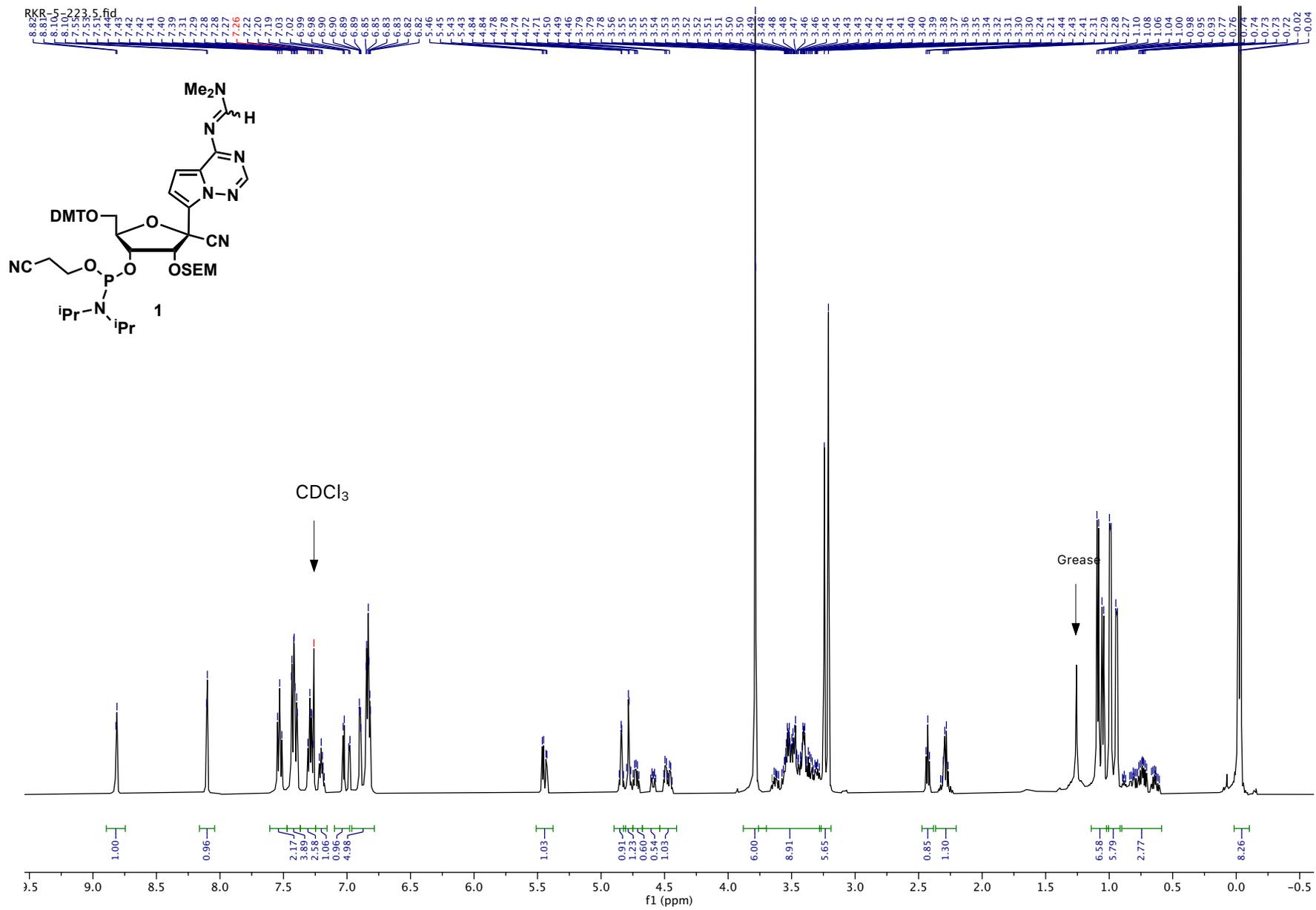
18.25

-0.70



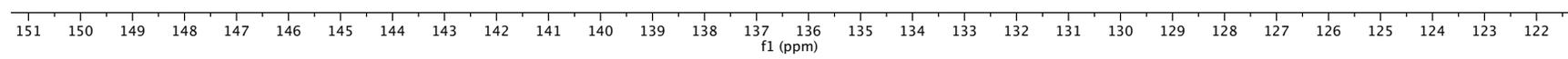
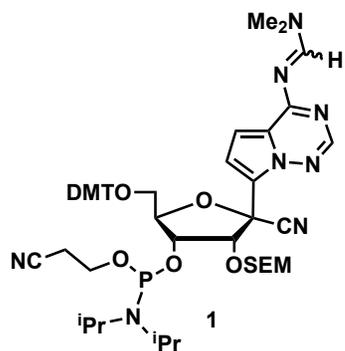
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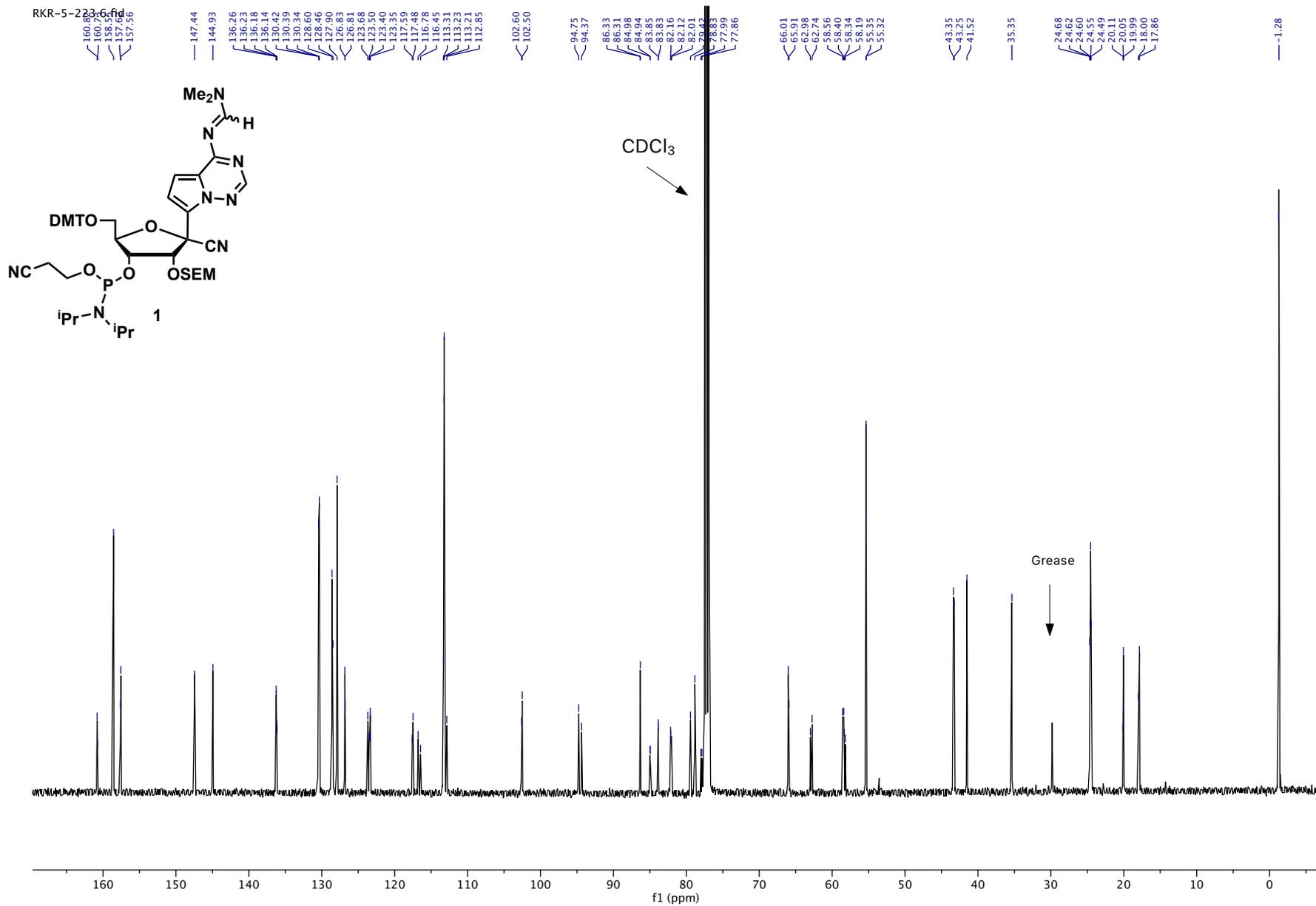




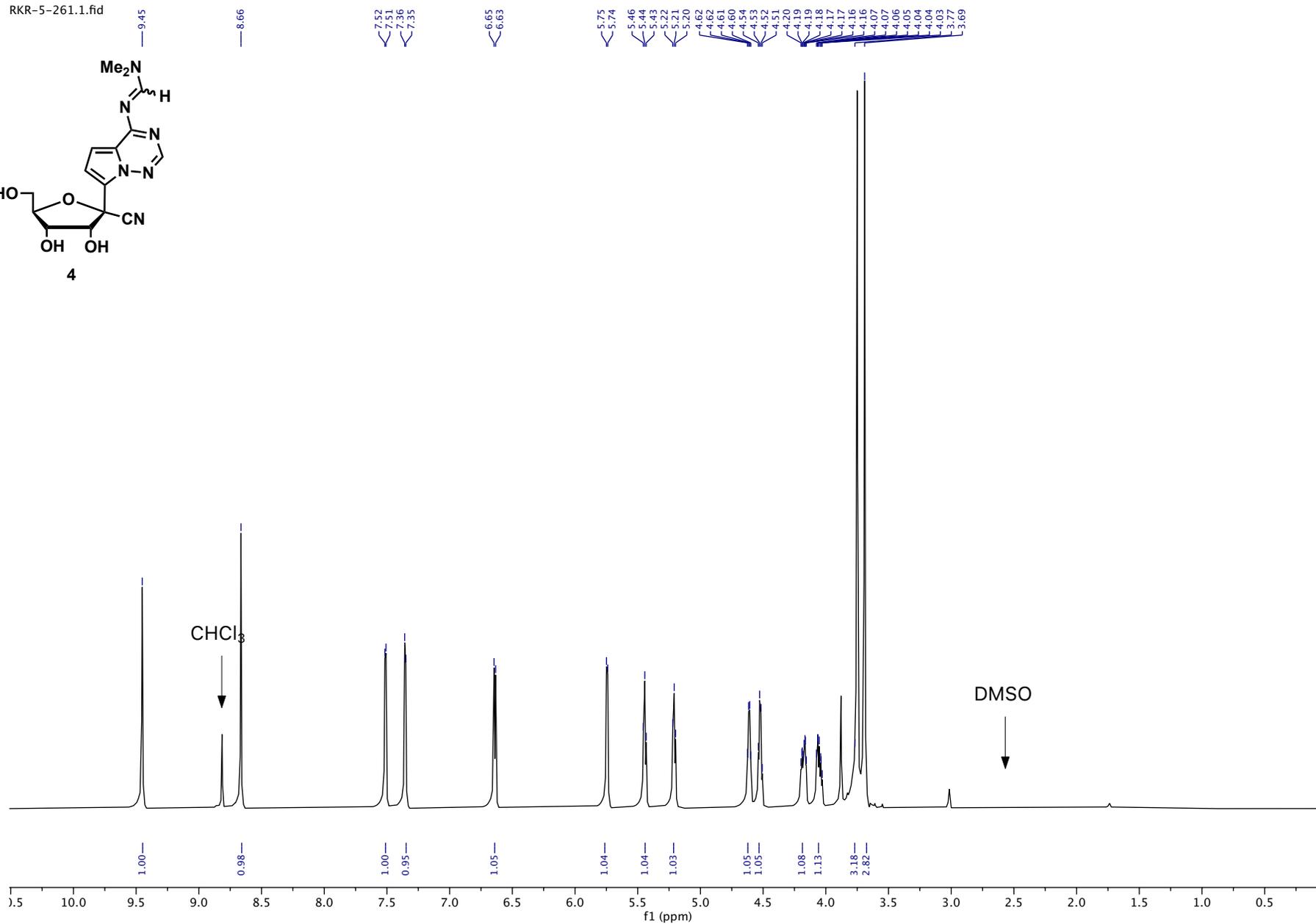
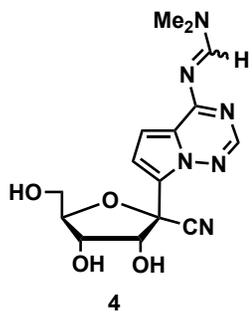
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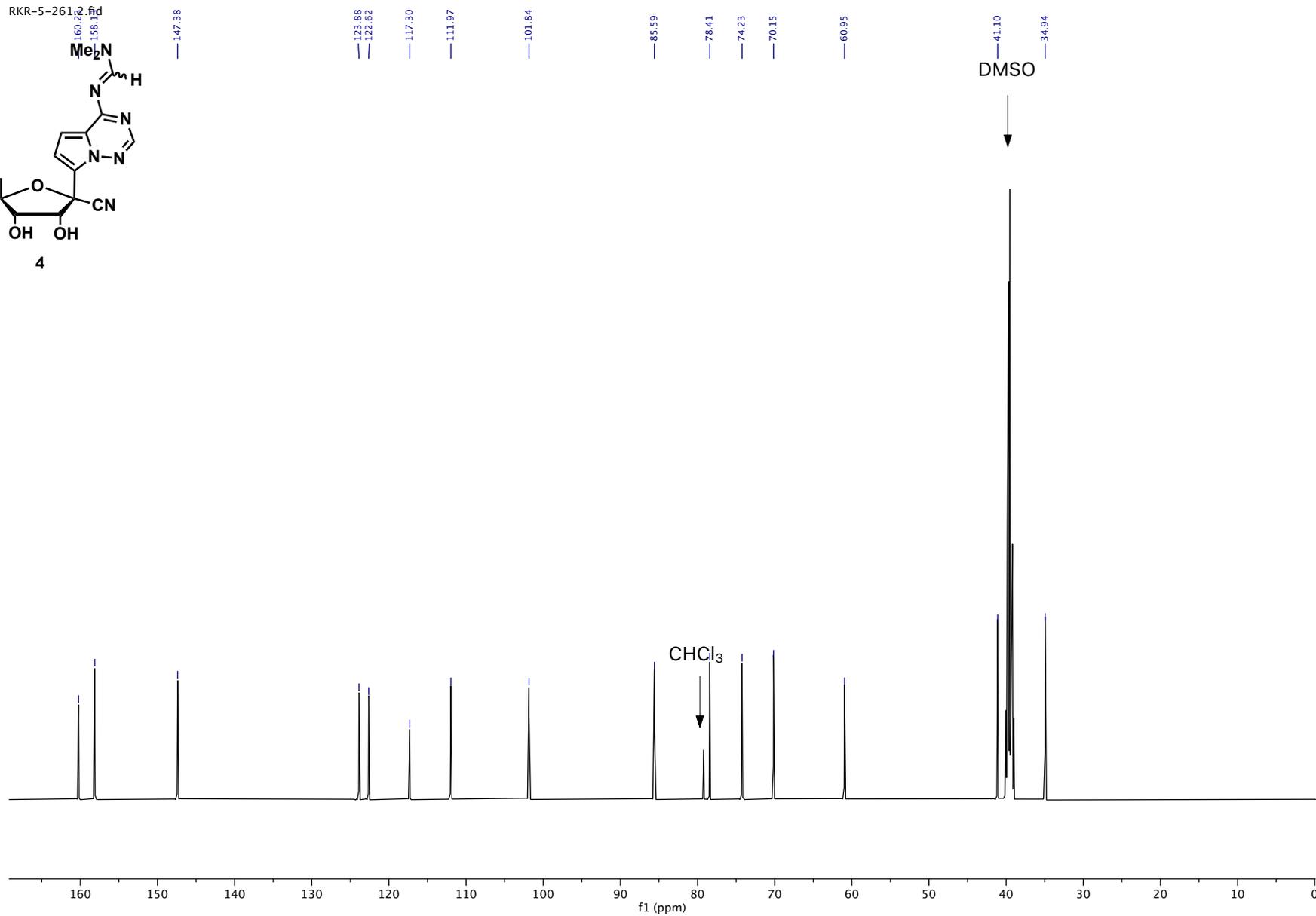
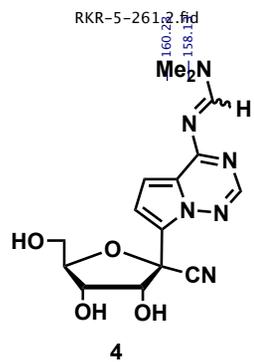
150.18  
149.74



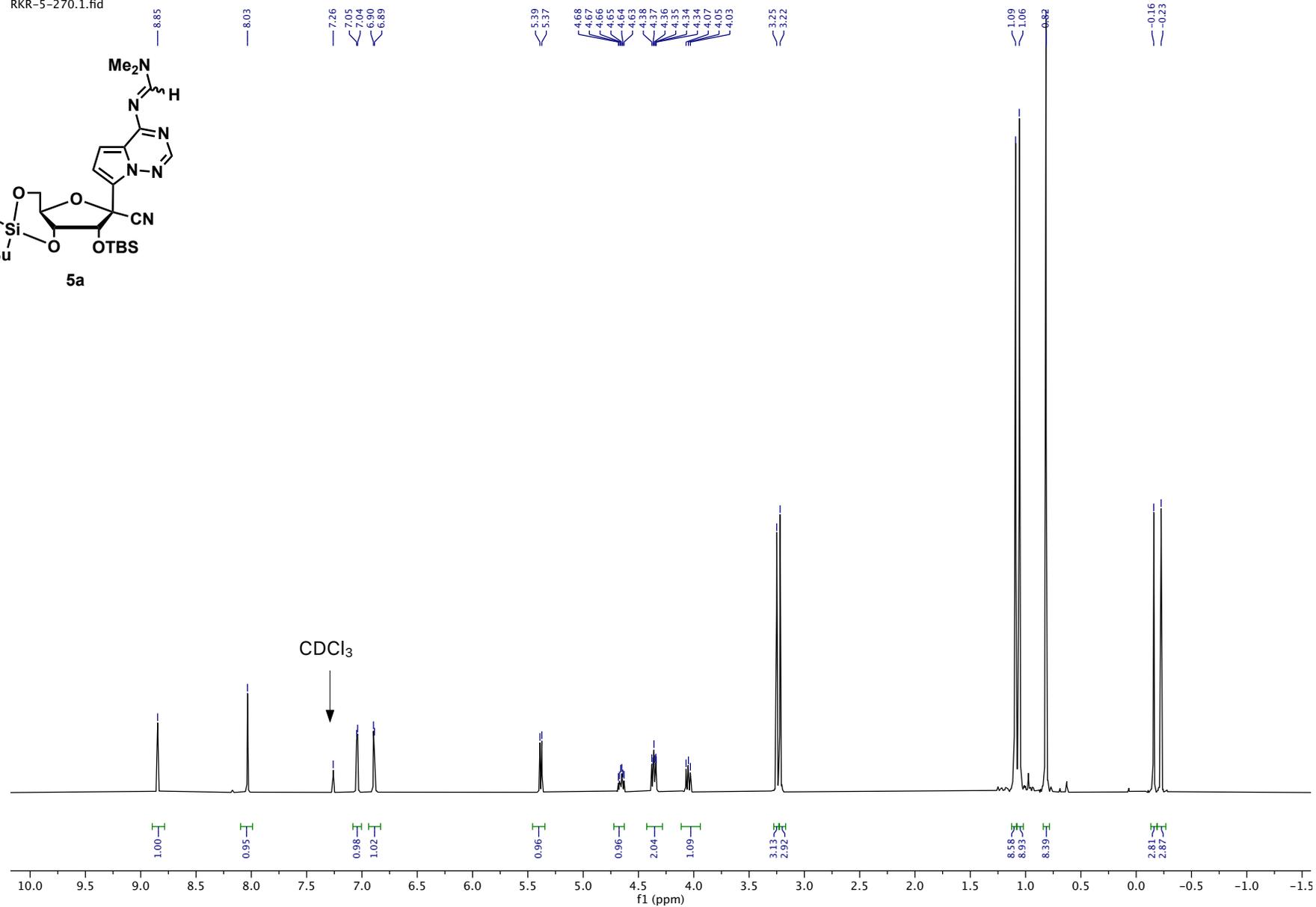
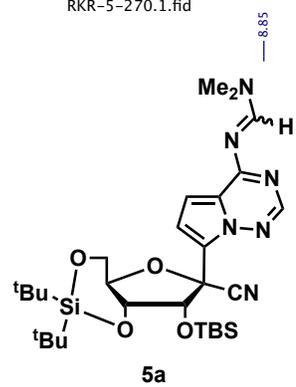


RKR-5-261.1.fid





RKR-5-270.1.fid



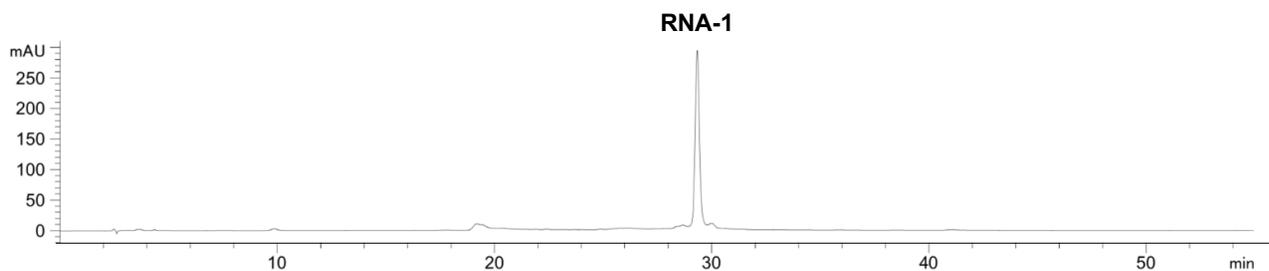


## VII. LC-MS Trace and Mass Spectra of RNA-1.

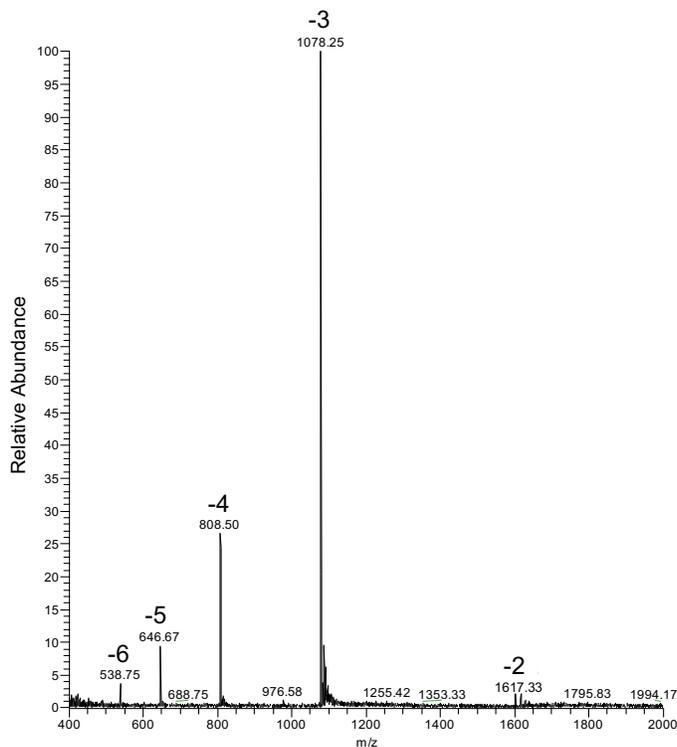
RNA-1: 5'—CCGGGCGGCR—3'

Purity – 91% (monitored at 260 nm); MS calculated ( $m/z + Na^+$ ): 3234.9; found ( $m/z + Na^+$ ): 3238.0

### HPLC Trace of RNA-1



### Mass Spectrometry Analysis of RNA-1



## VIII. References.

1. Reddy, M. P., Hanna, N. B., Farooqui, F. *Tetrahedron Lett.*, **1994**, 35, 4311-4314.
2. Wincott, F., DiRenzo, A., Shaffer, C., Grimm, S., Tracz, D., Workman, C., Sweedler, D., Gonzalez, C., Scaringe, S., Usman, N. *Nucleic Acids Res.*, **1995**, 23, 2677-2684.
3. Sproat, B., Colonna, F., Mullah, B., Tsou, D., Andrus, A., Hampel, A., Vinayak, R. *Nucleosides Nucleotides*, **1995**, 14, 255-273.
4. Pitsch, S., Weiss, P. A., Jenny, L., Stutz, A., Wu, X. *Helv. Chim. Acta*, **2001**, 84, 3773-3795.
5. Kocic, G., Hillen, H. S., Tegunov, D., Dienemann, C., Seitz, F., Schmitzova, J., Farnung, L., Siewert, A., Hobartner, C., Cramer, P. *Nat. Commun.*, **2021**, 12, 1-7.