

## Supplementary Information

### Effect of DNA modifications on the transition between canonical and non-canonical DNA structures in CpG islands during senescence

Saki Matsumoto<sup>a</sup>, Hisae Tateishi-Karimata<sup>a</sup>, Tatsuya Ohyama<sup>a</sup>, Naoki Sugimoto<sup>a, b, \*</sup>

<sup>a</sup> Frontier Institute for Biomolecular Engineering Research (FIBER), Konan University, 7-1-20 Minatojima-Minamimachi, Kobe 650-0047, Japan.

<sup>b</sup> Graduate School of Frontiers of Innovative Research in Science and Technology (FIRST), Konan University, 7-1-20 Minatojima-Minamimachi, Kobe 650-0047, Japan.

\*To whom correspondence should be addressed. Tel: (+81)78-303-1416; Fax: (+81)78-303-1495; Email: sugimoto@konan-u.ac.jp

Contents:

**Figure S1.** UV melting curves for i-motif forming sequences with modifications

**Figure S2.** UV melting curves for G-quadruplex forming sequences with modifications

**Figure S3.** The effect of modifications on the formation of i-motifs

**Figure S4.** UV melting curves for i-motif forming sequences at pH 5.0

**Figure S5.** UV melting curves for i-motif forming sequences at pH 7.0

**Figure S6.** The effect of PEG200 on the formation of i-motifs

**Figure S7.** The effect of cations on the formation of i-motifs

**Figure S8.** The effect of the modifications on the formation of G-quadruplexes.

**Figure S9.** CD spectra of i-motif forming DNAs in the presence of K<sup>+</sup>

**Figure S10.** CD spectra of i-motif forming DNAs in the presence of Na<sup>+</sup>

**Figure S11.** CD spectra of G-quadruplex forming DNAs in the presence of K<sup>+</sup>

**Figure S12.** Native gel electrophoresis of G-quadruplex forming sequences in the presence of K<sup>+</sup>

**Figure S13.** CD spectra of G-quadruplex forming DNAs in the presence of Na<sup>+</sup>

**Figure S14.** CD spectra of G4 with 0-40 wt% PEG200 in the presence of Na<sup>+</sup>

**Figure S15.** Native gel electrophoresis of G-quadruplex forming sequences in the presence of Na<sup>+</sup>

**Figure S16.** Native gel electrophoresis of G-quadruplex forming sequences in the presence of K<sup>+</sup> and different concentration of PEG200

**Table S1.** I-motif and G-quadruplex forming sequences in CpG island of *ELOVL2*

**Table S2.** The thermodynamic parameters of i-motif forming DNA in the presence of K<sup>+</sup>

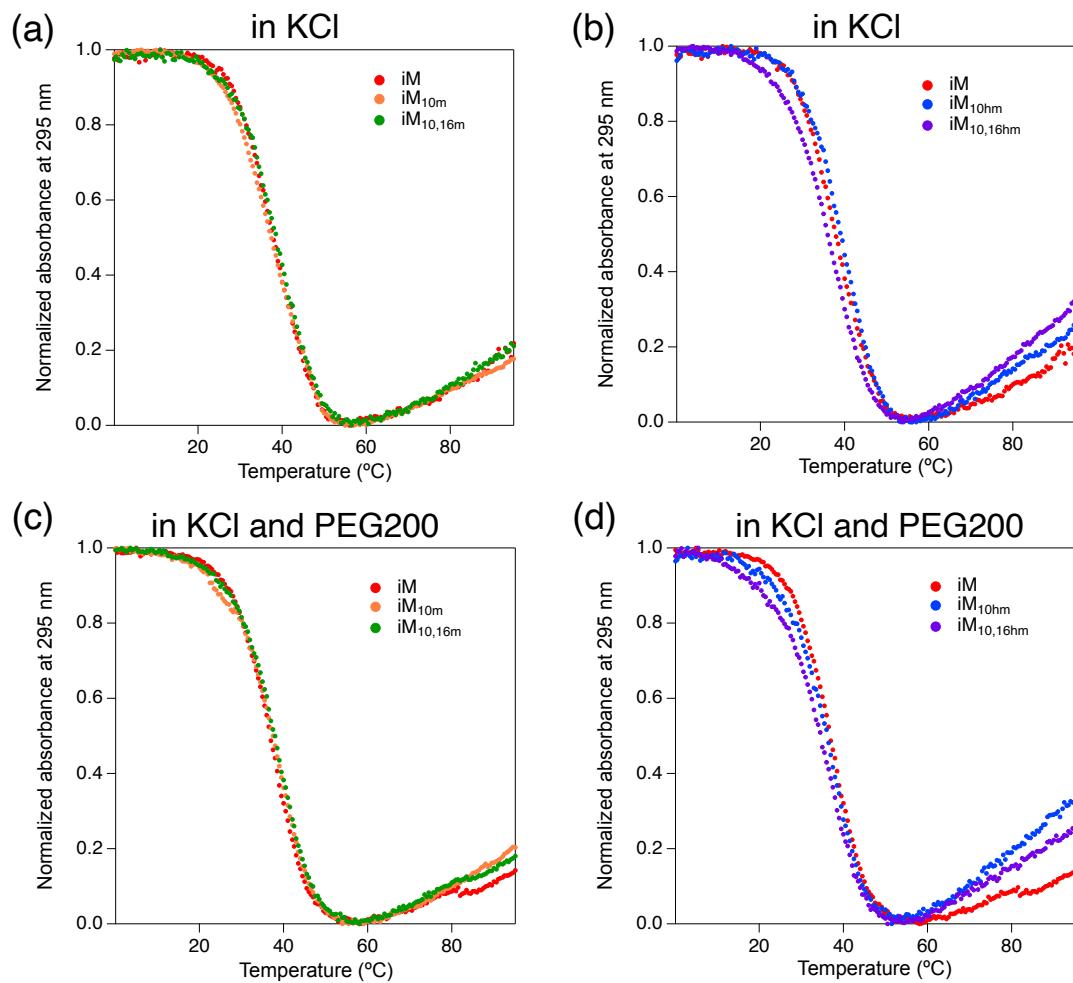
**Table S3.** The thermodynamic parameters of i-motif forming DNA in the presence of Na<sup>+</sup>

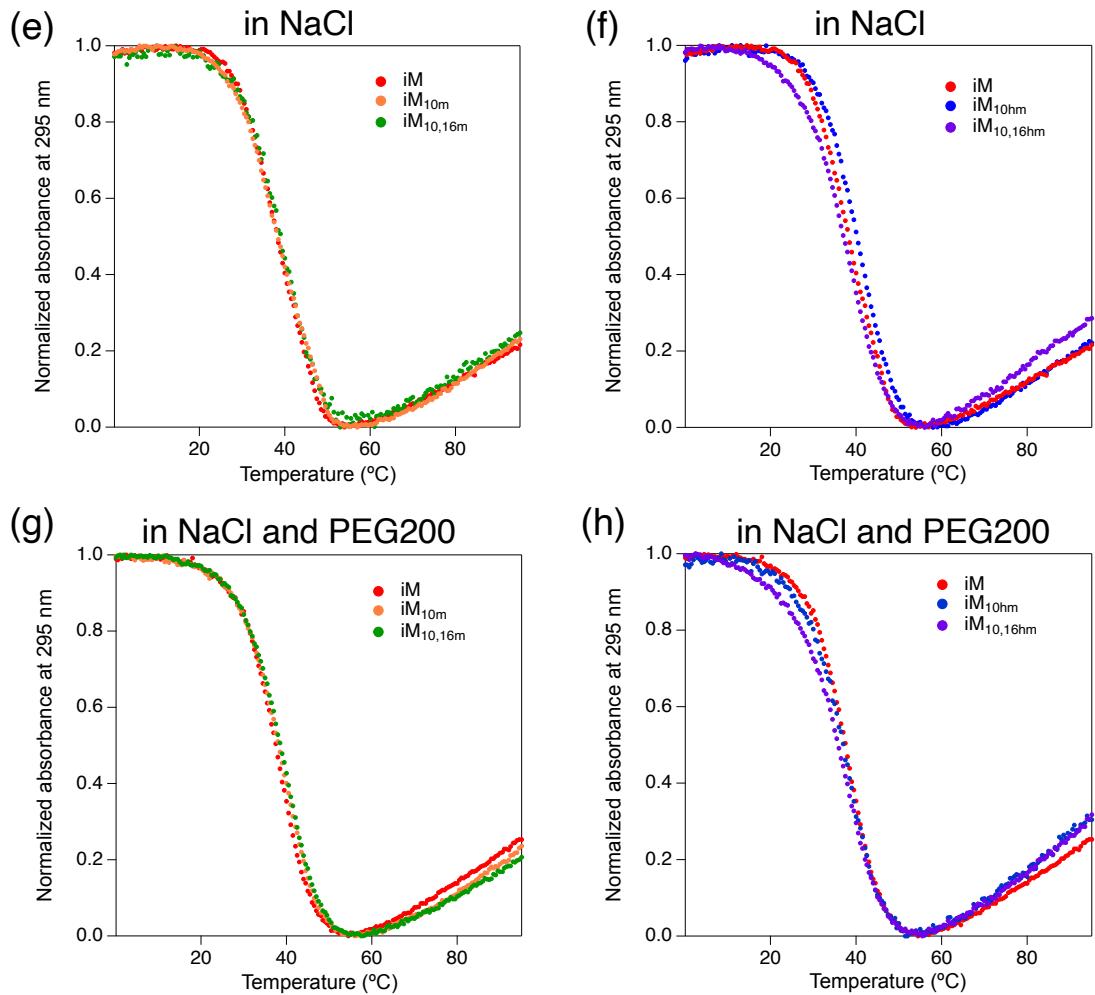
**Table S4.** The thermodynamic parameters of G-quadruplex forming DNA in the presence of K<sup>+</sup>

**Table S5.** The thermodynamic parameters of G-quadruplex forming DNA in the presence of Na<sup>+</sup>

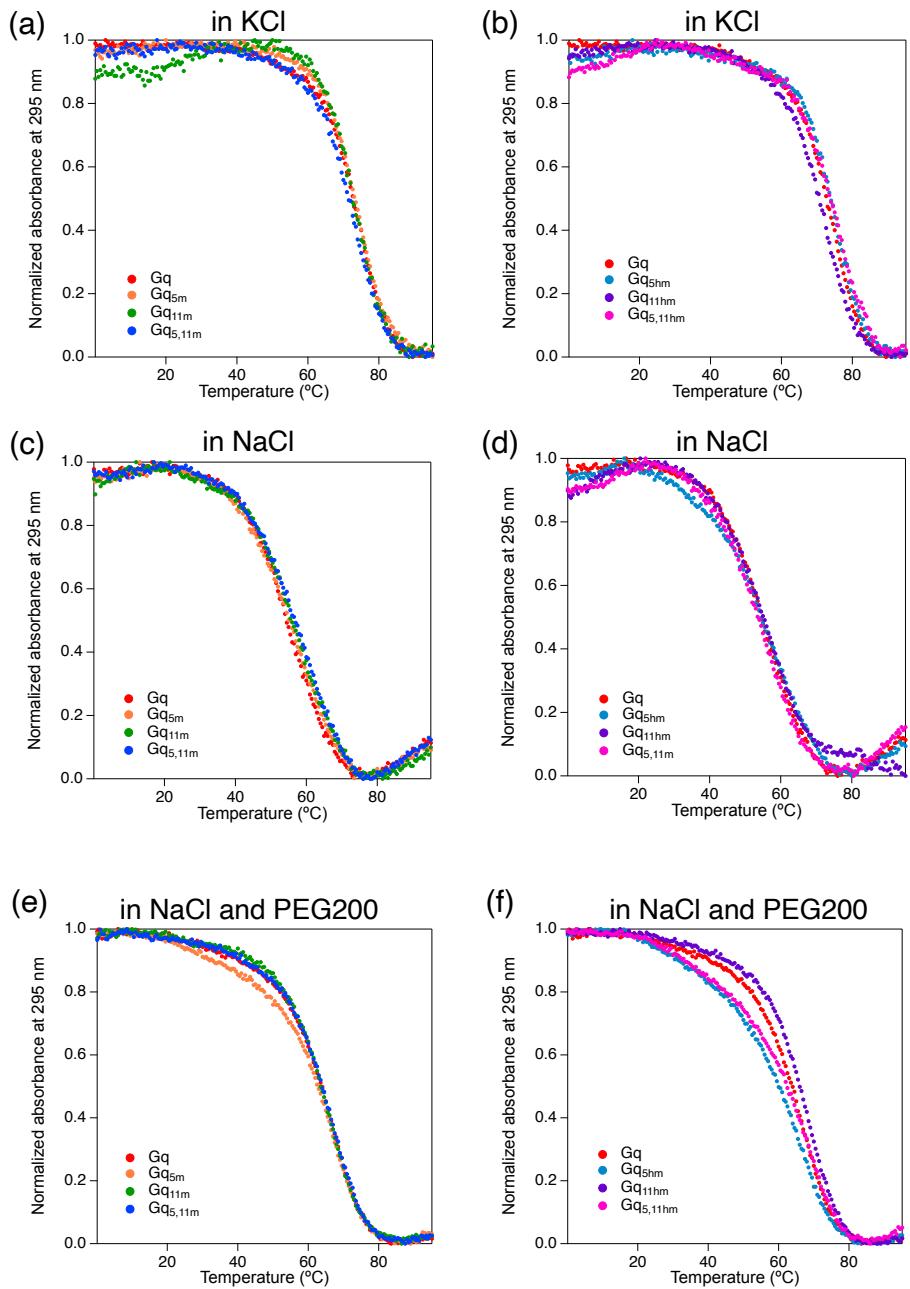
**Table S6.** Melting temperatures of i-motif forming DNA in the presence of complementary G-quadruplex forming DNA and K<sup>+</sup>

**Table S7.** Melting temperatures of duplex of i-motif and G-quadruplex forming DNAs

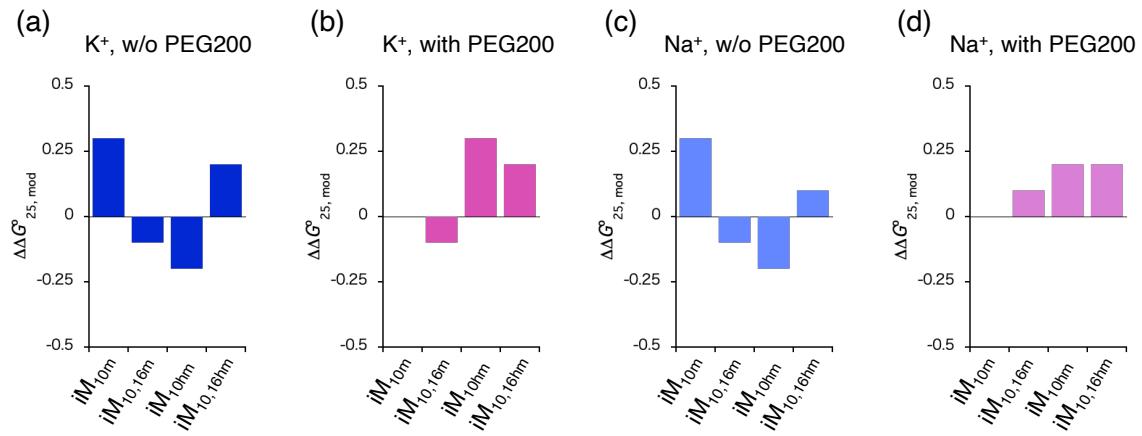




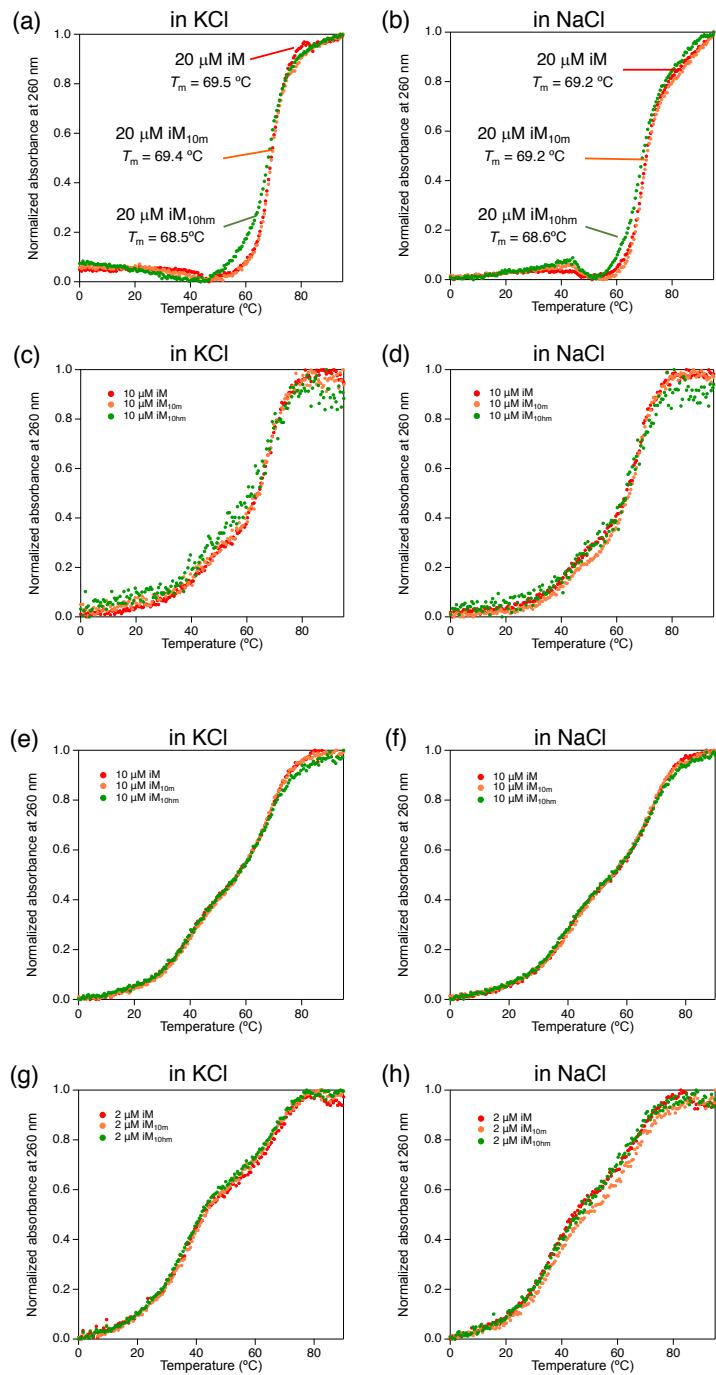
**Figure S1.** UV melting curves for 5  $\mu\text{M}$  of (a) iM, iM<sub>10m</sub>, and iM<sub>10,16m</sub> in the presence of K<sup>+</sup>, (b) iM, iM<sub>10hm</sub>, and iM<sub>10,16hm</sub> in the presence of K<sup>+</sup>, (c) iM, iM<sub>10m</sub>, and iM<sub>10,16m</sub> in the presence of K<sup>+</sup> and 40 wt% PEG200, (d) iM, iM<sub>10hm</sub>, and iM<sub>10,16hm</sub> in the presence of K<sup>+</sup> and 40 wt% PEG200, (e) iM, iM<sub>10m</sub>, and iM<sub>10,16m</sub> in the presence of Na<sup>+</sup>, (f) iM, iM<sub>10hm</sub>, and iM<sub>10,16hm</sub> in the presence of Na<sup>+</sup>, (g) iM, iM<sub>10m</sub>, and iM<sub>10,16m</sub> in the presence of Na<sup>+</sup> and 40 wt% PEG200, and (h) iM, iM<sub>10hm</sub>, and iM<sub>10,16hm</sub> in the presence of Na<sup>+</sup> and 40 wt% PEG200.



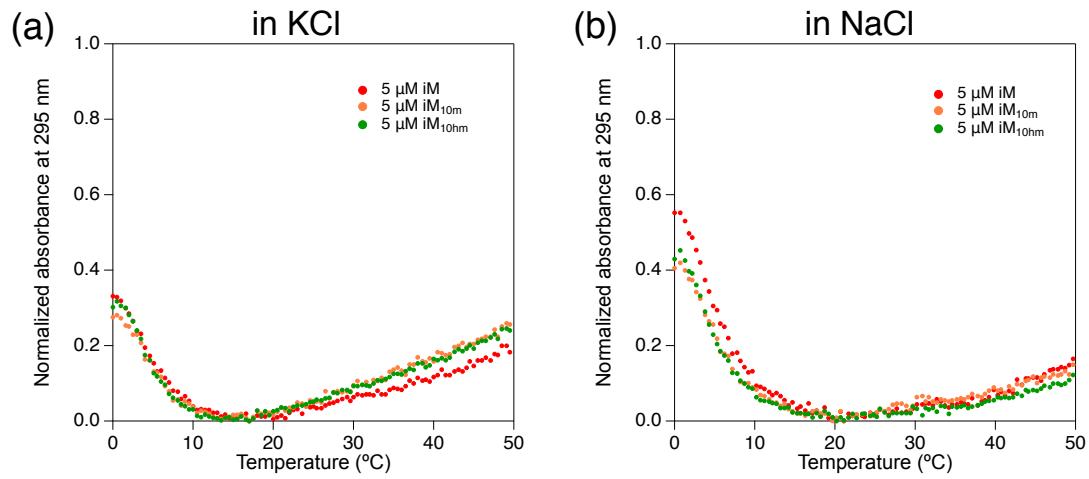
**Figure S2.** UV melting curves for 5  $\mu\text{M}$  of (a) Gq, Gq<sub>5m</sub>, Gq<sub>11m</sub>, and Gq<sub>5,11m</sub> in the presence of  $\text{K}^+$ , (b) Gq, Gq<sub>5hm</sub>, Gq<sub>11hm</sub>, and Gq<sub>5,11hm</sub> in the presence of  $\text{K}^+$ , (c) Gq, Gq<sub>5m</sub>, Gq<sub>11m</sub>, and Gq<sub>5,11m</sub> in the presence of  $\text{Na}^+$ , (d) Gq, Gq<sub>5hm</sub>, Gq<sub>11hm</sub>, and Gq<sub>5,11hm</sub> in the presence of  $\text{Na}^+$ , (e) Gq, Gq<sub>5m</sub>, Gq<sub>11m</sub>, and Gq<sub>5,11m</sub> in the presence of  $\text{Na}^+$  and 40 wt% PEG200, and (f) Gq, Gq<sub>5hm</sub>, Gq<sub>11hm</sub>, and Gq<sub>5,11hm</sub> in the presence of  $\text{Na}^+$  and 40 wt% PEG200.



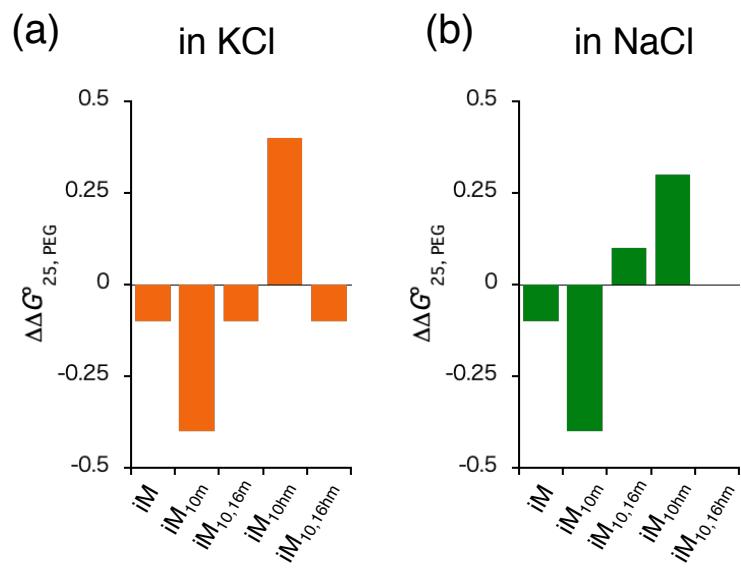
**Figure S3.** The effect of modifications on the formation of i-motifs in 100 mM KCl or NaCl in 0 wt% or 40 wt% PEG200. The  $\Delta\Delta G^\circ_{25, \text{mod}}$  values in (a) 100 mM KCl with 0 wt% PEG200, (b) 100 mM KCl with 40 wt% PEG200, (c) 100 mM NaCl with 0 wt% PEG200, and (d) 100 mM NaCl with 40 wt% PEG200.



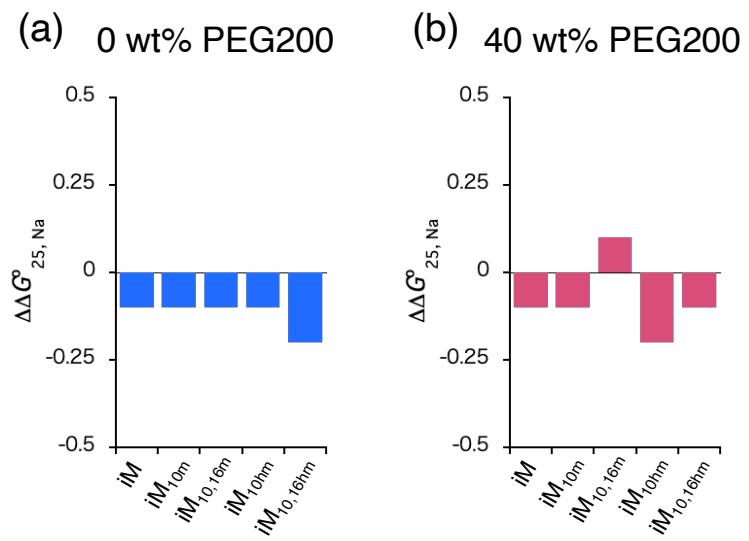
**Figure S4.** UV melting curves at pH 5.0 for (a) 20  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of K<sup>+</sup>, (b) 20  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of Na<sup>+</sup>, (c) 10  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of K<sup>+</sup>, (d) 10  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of Na<sup>+</sup>, (e) 5  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of K<sup>+</sup>, (f) 5  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of Na<sup>+</sup>, (g) 2  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of K<sup>+</sup>, and (h) 2  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of Na<sup>+</sup>.



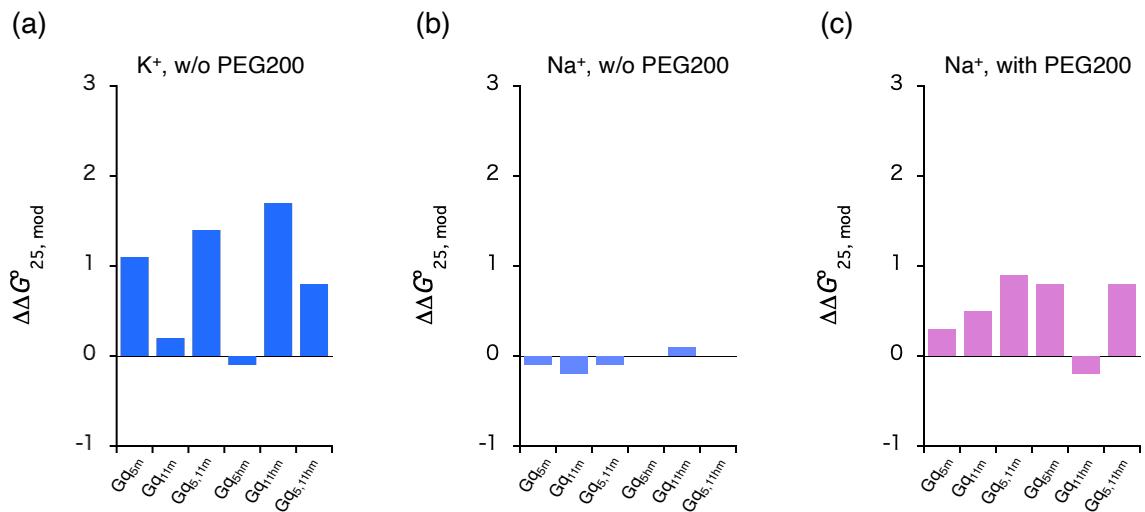
**Figure S5.** UV melting curves at pH 7.0 for (a) 5  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of K<sup>+</sup> and (b) 5  $\mu\text{M}$  of iM, iM<sub>10m</sub>, and iM<sub>10hm</sub> in the presence of Na<sup>+</sup>.



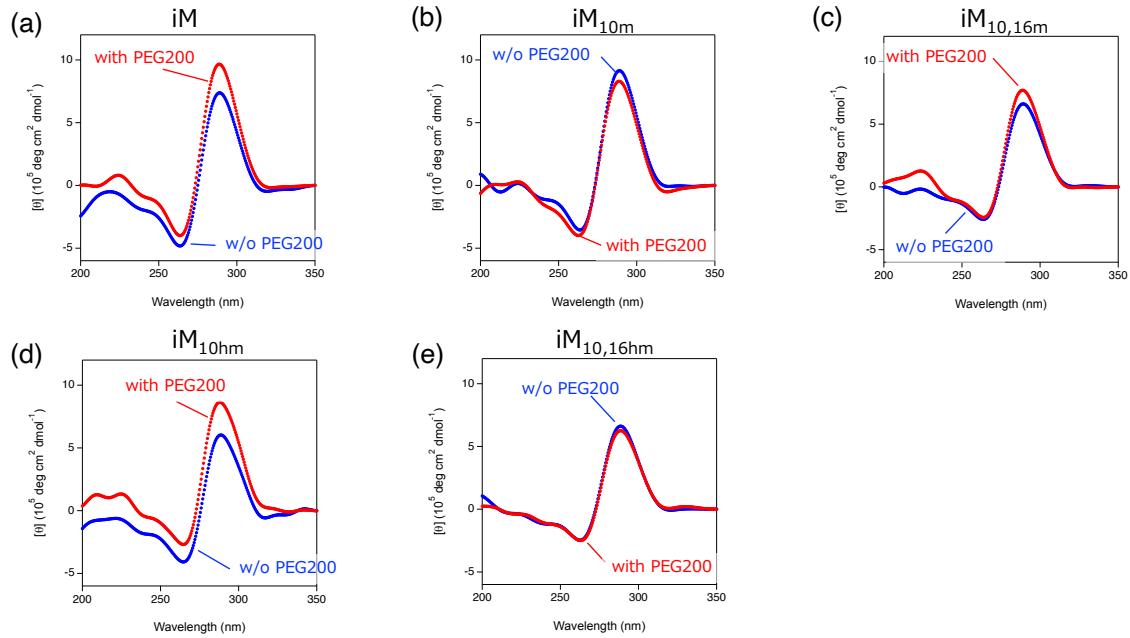
**Figure S6.** The effect of PEG200 on the formation of i-motifs. The  $\Delta\Delta G^{\circ}_{25,\text{PEG}}$  values in (a) 100 mM KCl and (b) 100 mM NaCl



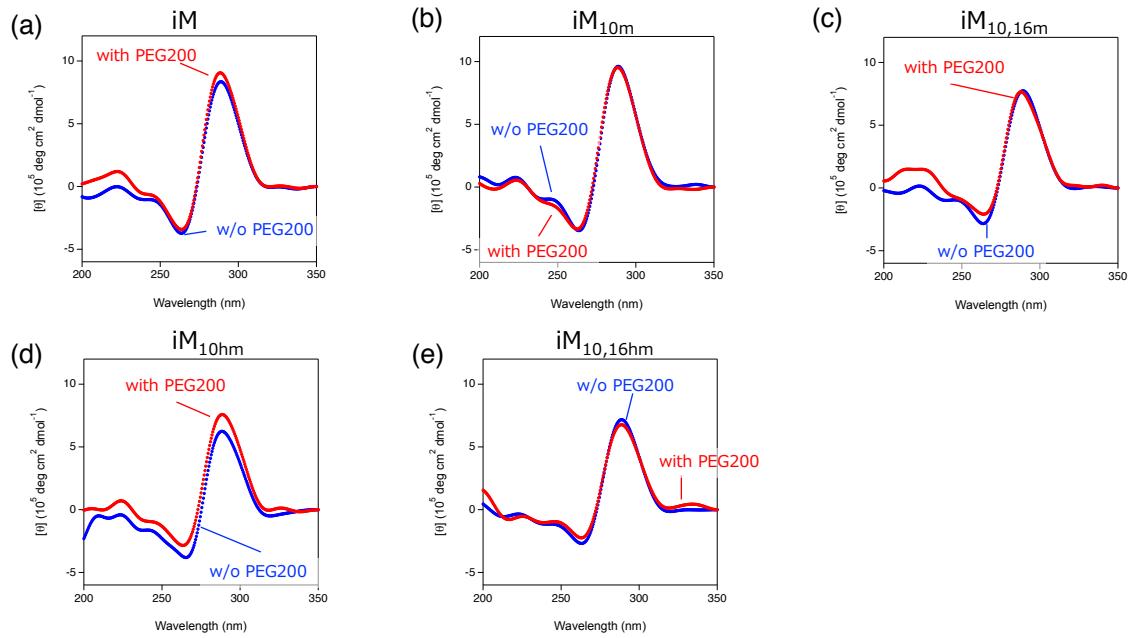
**Figure S7.** The effect of cations on the formation of *i*-motifs. The  $\Delta\Delta G^{\circ}_{25,Na}$  values in (a) 0 wt% PEG200 and (b) 40 wt% PEG200



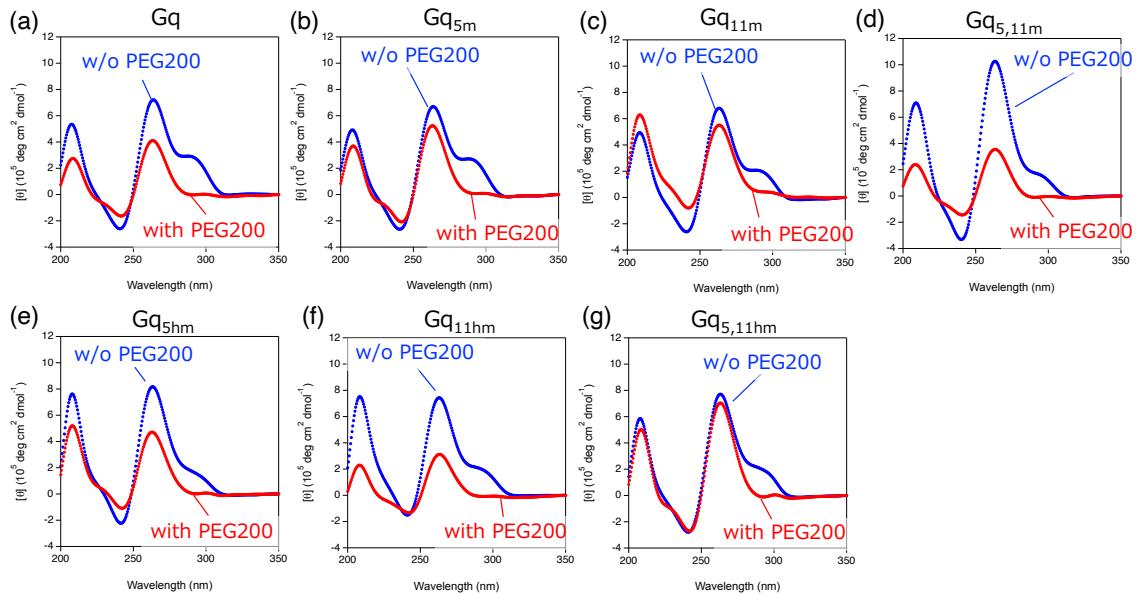
**Figure S8.** The effect of the modifications on the formation of G-quadruplexes. The  $\Delta\Delta G^\circ_{25, \text{mod}}$  values in (a) 100 mM KCl with 0 wt% PEG200, (b) 100 mM NaCl with 0 wt% PEG200, (c) 100 mM NaCl with 40 wt% PEG200.



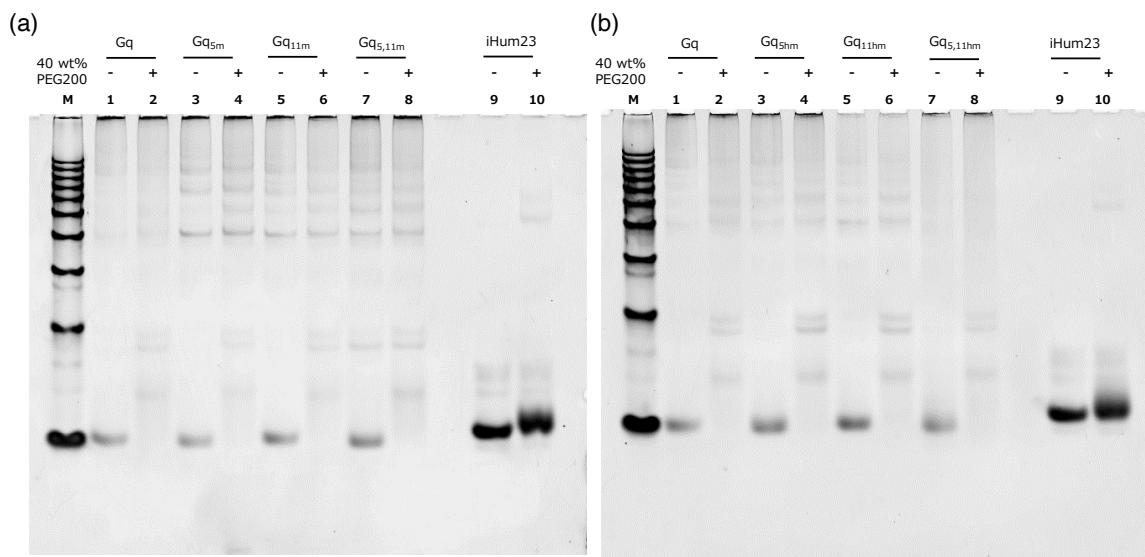
**Figure S9.** CD spectra of (a) iM, (b) iM<sub>10m</sub> (c) iM<sub>10,16m</sub>, (d) iM<sub>10hm</sub>, and (e) iM<sub>10,16hm</sub> in the presence of K<sup>+</sup>. All experiments were carried out at 4 °C in 50 mM MES-LiOH (pH 6.0) containing 100 mM KCl containing 0 wt% (blue) or 40 wt% PEG200 (red).



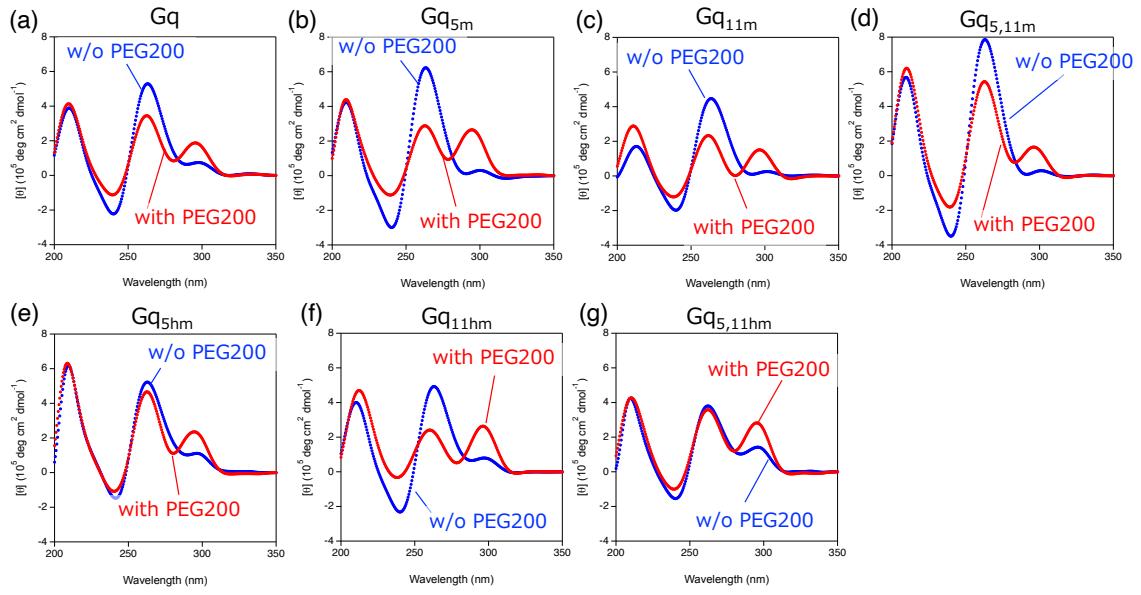
**Figure S10.** CD spectra of (a) iM, (b) iM<sub>10m</sub> (c) iM<sub>10,16m</sub>, (d) iM<sub>10hm</sub>, and (e) iM<sub>10,16hm</sub> in the presence of Na<sup>+</sup>. All experiments were carried out at 4 °C in 50 mM MES-LiOH (pH 6.0) containing 100 mM NaCl containing 0 wt% (blue) or 40 wt% PEG200 (red).



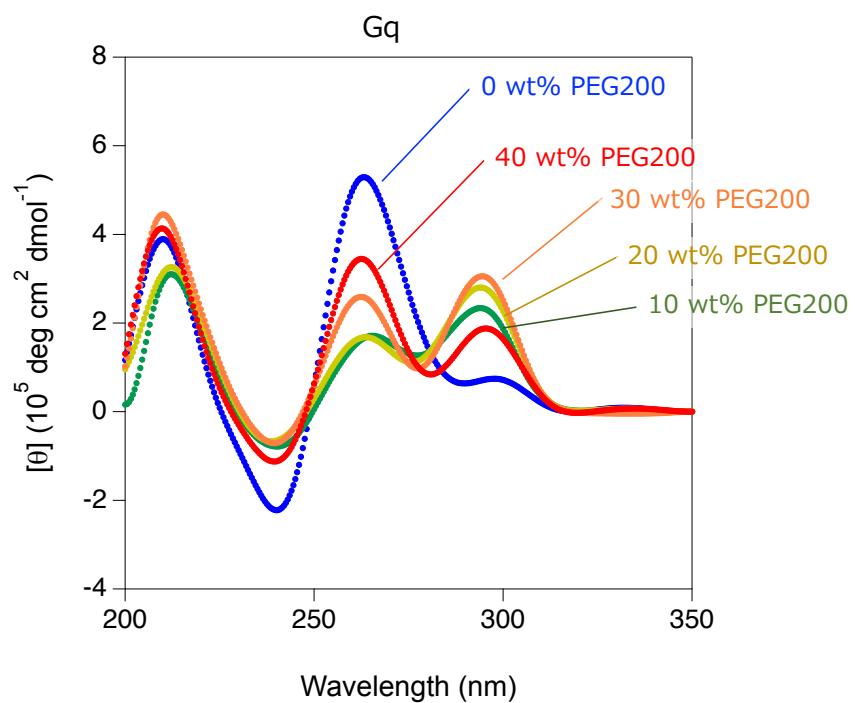
**Figure S11.** CD spectra of (a) Gq, (b) Gq<sub>5m</sub> (c) Gq<sub>11m</sub>, (d) Gq<sub>5,11m</sub>, (e) Gq<sub>5hm</sub>, (f) Gq<sub>11hm</sub>, and (g) Gq<sub>5,11hm</sub> in the presence of K<sup>+</sup>. All experiments were carried out at 4 °C in 50 mM MES-LiOH (pH 6.0) with 100 mM KCl containing 0 wt% (blue) or 40 wt% PEG200 (red).



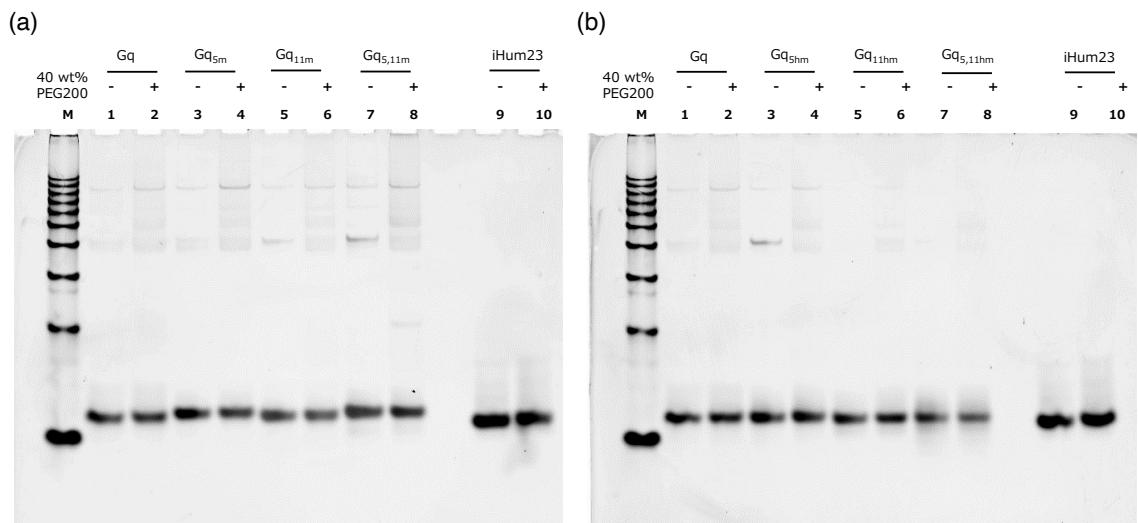
**Figure S12.** Nondenaturing gel electrophoresis of (a) Gq, Gq<sub>5m</sub>, Gq<sub>11m</sub>, and Gq<sub>5,11m</sub> and (b) Gq, Gq<sub>5hm</sub>, Gq<sub>11hm</sub>, and Gq<sub>5,11hm</sub> at 25 °C in 50 mM MES-LiOH (pH 6.0) containing 100 mM KCl without or with 40 wt% of PEG200. Lane M: 10 bp DNA standard; Lanes 1, 3, 5, 7 and 9; without PEG200: lanes 2, 4, 6, 8, and 10; with 40 wt% PEG200. iHum23 indicates telomeric G-quadruplex, TA(G<sub>3</sub>TTA)<sub>3</sub>G<sub>3</sub> as the marker of monomer G-quadruplex with antiparallel.



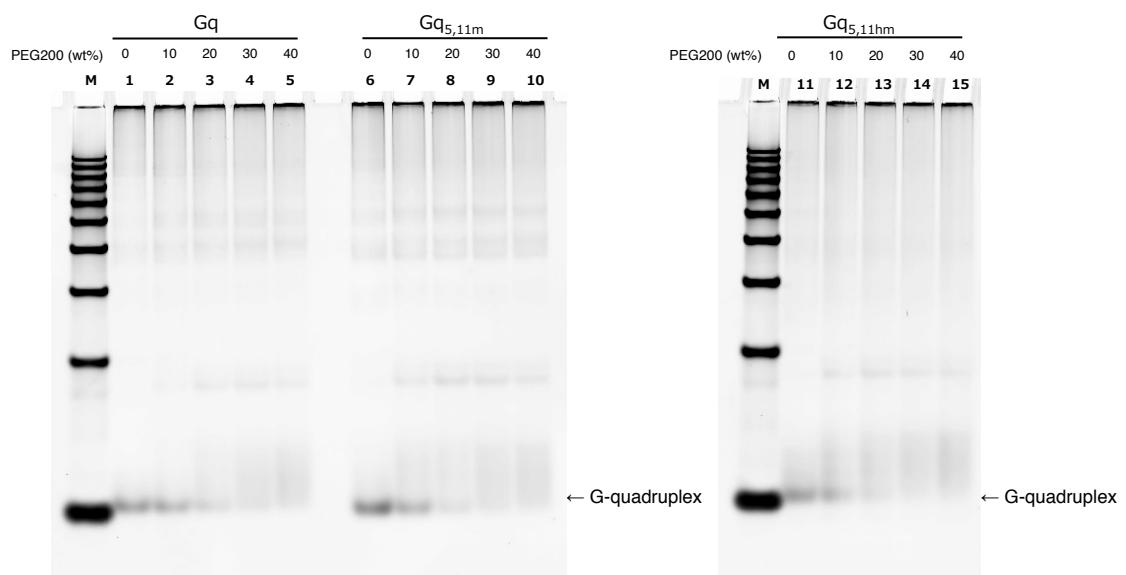
**Figure S13.** CD spectra of (a) Gq, (b) Gq<sub>5m</sub> (c) Gq<sub>11m</sub>, (d) Gq<sub>5,11m</sub>, (e) Gq<sub>5hm</sub>, (f) Gq<sub>11hm</sub>, and (g) Gq<sub>5,11hm</sub> in the presence of Na<sup>+</sup>. All experiments were carried out at 4 °C in 50 mM MES-LiOH (pH 6.0) with 100 mM NaCl containing 0 wt% (blue) or 40 wt% PEG200 (red).



**Figure S14.** CD spectra of Gq in the presence of  $\text{Na}^+$ . All experiments were carried out at 4 °C in 50 mM MES-LiOH (pH 6.0) with 100 mM NaCl containing 0 wt% (blue), 10 wt% (green), 20 wt% (yellow), 30 wt% (orange), or 40 wt% PEG200 (red).



**Figure S15.** Nondenaturing gel electrophoresis of (a) Gq, Gq<sub>5m</sub>, Gq<sub>11m</sub>, and Gq<sub>5,11m</sub> and (b) Gq, Gq<sub>5m</sub>, Gq<sub>11hm</sub>, and Gq<sub>5,11hm</sub> at 25 °C in 50 mM MES-LiOH (pH 6.0) containing 100 mM NaCl without or with 40 wt% of PEG200. Lane M: 10 bp DNA standard; Lanes 1, 3, 5, 7 and 9; without PEG200: lanes 2, 4, 6, 8, and 10; with 40 wt% PEG200. iHum23 indicates telomeric G-quadruplex, TA(G<sub>3</sub>TTA)<sub>3</sub>G<sub>3</sub> as the marker of monomer G-quadruplex with antiparallel.



**Figure S16.** Nondenaturing gel electrophoresis of Gq, Gq<sub>5,11m</sub>, and Gq<sub>5,11hm</sub>, at 25 °C in 50 mM MES-LiOH (pH 6.0) containing 100 mM KCl with 0, 10, 20, 30, and 40 wt% of PEG200. Lane M: 10 bp DNA standard; lanes 1, 6, and 11; 0 wt % PEG200: lanes 2, 7, and 12; with 10 wt% PEG200: lanes 3, 8, and 13; with 20 wt% PEG200: lanes 4, 9, and 14; with 30 wt% PEG200: lanes 5, 10, and 15; with 40 wt% PEG200.

**Table S1.** i-motif and G-quadruplex forming sequences in CpG island of *ELOVL2*

No.	sense sequences	$T_m$ (°C) <sup>a</sup>	antisense sequences	$T_m$ (°C) <sup>a</sup>	$N_{CpG}^b$
1	GGGCAG <b>C</b> GGGTGGGTATTCTGGGG	N.D. <sup>c</sup>	CCCCAGGAATACCCACCC <b>G</b> CTGCC	34.8	1
2	GGGGGC <b>GGGG</b> GAGG <b>CGCGGGCGGG</b>	60.6	<b>CCCGCCCGCGC</b> CCTCCCC <b>GCCCC</b>	36.8	4
3	GGG <b>CGT</b> GGGTGTGGGTGGGG	58.7	CCCCCACCCACACCC <b>A</b> CGCCC	34.3	1
4	GGG <b>CGGGAAAGGGCCGAGCGGG</b>	N.D. <sup>c</sup>	<b>CCCGCTCGGCC</b> CTTCCCGCCC	27.8	3
5	GGGG <b>CCGGGTCCGCGCGGG</b> CTGGGGAG <b>CGGG</b>	72	<b>CCCGCT</b> CCCCAGGCC <b>CGCGCGG</b> ACCCGGCCCC	34.4	5
6	CCCACCC <b>TCCGACCC</b> TCC <b>GGACCCCC</b>	34.5	GGGG <b>GTCCGGAGGGTCGGAGGGT</b> GGG	45.2	2
7	CCC <b>CTCCCCGGTCCC</b> GCCCC	45.4	GGGG <b>CGGGACC</b> GGGGAGAGGGG	56.0	2
8	CC <b>CTGCGCCC</b> CTCCCC <b>CGCGCGCC</b>	24.5	GGG <b>CGCGCGGGGGAGGGGCGCAGGG</b>	60.5	4
9	CCCCCAGAGAAACCCACCCACCC	26.3	GGGG <b>GGTTGGGTGG</b> TTCTCTGGGG	N.D. <sup>c</sup>	0
10	CCC <b>ACCGCACCCCCAGAGAAACCCACCC</b>	36.0	GGGTGGTTCTCTGGGG <b>TGCGGT</b> GGG	43.2	1

<sup>a</sup>The value of melting temperature ( $T_m$ ) was determined by differentiation of UV melting curve of each DNA (5 μM) in 50 mM MES-LiOH (pH 6.0) and 100 mM NaCl. <sup>b</sup> $N_{CpG}$  represents the number of CpG sites within structure forming sequences and CpG sites are marked in bold.

<sup>c</sup>“N.D.” indicates that values could not be obtained from the melting curves.

**Table S2.** The thermodynamic parameters of i-motif forming DNA in the presence of K<sup>+</sup><sup>a</sup>

	without PEG200				40 wt% PEG200			
	$\Delta H^\circ$ ( $\Delta\Delta H^\circ_{\text{mod}}\text{b}$ ) [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [ $\Delta(T\Delta S^\circ_{\text{mod}})$ ] <sup>b</sup> [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ ( $\Delta\Delta G^\circ_{25, \text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T_m$ ( $\Delta T_m$ , mod) <sup>b</sup> [°C]	$\Delta H^\circ$ ( $\Delta\Delta H^\circ_{\text{mod}}\text{b}$ ) [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [ $\Delta(T\Delta S^\circ_{\text{mod}})$ ] <sup>b</sup> [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ ( $\Delta\Delta G^\circ_{25, \text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T_m$ ( $\Delta T_m$ , mod) <sup>b</sup> [°C]
	iM	$-37.1 \pm 0.7$	$-35.5 \pm 0.8$	$-1.7 \pm 0.0$	$39.1 \pm 0.5$	$-41.2 \pm 0.7$	$-39.4 \pm 0.7$	$-1.8 \pm 0.0$
iM <sub>10m</sub>	$-32.6 \pm 0.8$ (+4.5)	$-31.1 \pm 0.9$ (+4.4)	$-1.4 \pm 0.1$ (+0.3)	$38.8 \pm 1.0$ (- 0.3)	$-38.7 \pm 4.4$ (+2.5)	$-37.0 \pm 4.2$ (+2.4)	$-1.8 \pm 0.3$ (0.0)	$39.5 \pm 0.5$ (+ 0.9)
iM <sub>10,16m</sub>	$-35.9 \pm 1.0$ (+1.2)	$-34.2 \pm 1.0$ (+1.3)	$-1.8 \pm 0.1$ (-0.1)	$40.4 \pm 0.7$ (+ 1.3)	$-38.1 \pm 1.8$ (+3.1)	$-36.1 \pm 1.8$ (+3.3)	$-1.9 \pm 0.1$ (-0.1)	$40.9 \pm 0.7$ (+ 2.3)
iM <sub>10hm</sub>	$-36.9 \pm 0.5$ (+0.2)	$-35.0 \pm 0.5$ (+0.5)	$-1.9 \pm 0.1$ (-0.2)	$41.2 \pm 0.7$ (+ 2.1)	$-35.2 \pm 2.4$ (+6.0)	$-33.7 \pm 2.3$ (+5.7)	$-1.5 \pm 0.1$ (+0.3)	$38.4 \pm 0.1$ (- 0.2)
iM <sub>10,16hm</sub>	$-34.9 \pm 0.5$ (+2.2)	$-33.4 \pm 0.5$ (+2.1)	$-1.5 \pm 0.0$ (+0.2)	$38.8 \pm 0.1$ (- 0.3)	$-36.6 \pm 1.1$ (+4.6)	$-35.1 \pm 1.1$ (+4.3)	$-1.6 \pm 0.0$ (+0.2)	$38.2 \pm 0.5$ (- 0.4)

<sup>a</sup>The values were determined in 50 mM MES-LiOH (pH 6.0) and 100 mM KCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5 μM of DNA. <sup>b</sup>The amount of change resulting from the modifications was calculated by using  $\Delta X = [X(\text{without modification}) - X(\text{modification})]$ , in which  $X$  is  $\Delta H^\circ$ ,  $T\Delta S^\circ$ ,  $\Delta G^\circ_{25}$ , or  $T_m$  and included in parentheses.

**Table S3.** The thermodynamic parameters of i-motif forming DNA in the presence of Na<sup>+</sup><sup>a</sup>

	without PEG200				40 wt% PEG200			
	$\Delta H^\circ$ ( $\Delta\Delta H^\circ_{\text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [ $\Delta(T\Delta S^\circ_{\text{mod}})$ ] <sup>b</sup> [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ ( $\Delta\Delta G^\circ_{25,\text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T_m$ ( $\Delta T_{m,\text{mod}}$ ) <sup>b</sup> [°C]	$\Delta H^\circ$ ( $\Delta\Delta H^\circ_{\text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [ $\Delta(T\Delta S^\circ_{\text{mod}})$ ] <sup>b</sup> [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ ( $\Delta\Delta G^\circ_{25,\text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T_m$ ( $\Delta T_{m,\text{mod}}$ ) <sup>b</sup> [°C]
	iM	-36.8 ± 1.1	-35.0 ± 1.0	-1.8 ± 0.0	40.1 ± 0.4	-41.6 ± 2.8	-39.7 ± 2.7	-1.9 ± 0.1
iM <sub>10m</sub>	-31.4 ± 1.0 (+5.4)	-29.9 ± 1.0 (+5.1)	-1.5 ± 0.1 (+0.3)	40.2 ± 0.8 (+ 0.1)	-40.0 ± 2.8 (+1.6)	-38.1 ± 2.6 (+1.6)	-1.9 ± 0.2 (0.0)	39.7 ± 0.2 (+ 0.4)
iM <sub>10,16m</sub>	-37.5 ± 0.6 (-0.7)	-35.7 ± 0.6 (-0.7)	-1.9 ± 0.0 (-0.1)	40.7 ± 0.1 (+ 0.6)	-40.2 ± 2.0 (+1.4)	-38.5 ± 2.0 (+1.2)	-1.8 ± 0.1 (+0.1)	38.6 ± 1.0 (-0.7)
iM <sub>10hm</sub>	-38.6 ± 1.0 (-1.8)	-36.5 ± 1.0 (-1.5)	-2.0 ± 0.1 (-0.2)	41.7 ± 0.4 (+ 1.6)	-37.8 ± 2.5 (+3.8)	-36.1 ± 2.3 (+3.6)	-1.7 ± 0.2 (+0.2)	38.8 ± 0.8 (- 0.5)
iM <sub>10,16hm</sub>	-35.5 ± 2.8 (+1.3)	-33.8 ± 2.7 (+1.2)	-1.7 ± 0.1 (+0.1)	40.0 ± 0.7 (- 0.1)	-37.0 ± 2.2 (+4.6)	-35.3 ± 2.1 (+4.4)	-1.7 ± 0.0 (+0.2)	39.1 ± 0.6 (- 0.2)

<sup>a</sup>The values were determined in 50 mM MES-LiOH (pH 6.0) and 100 mM NaCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5 μM of DNA. <sup>b</sup>The amount of change resulting from the modifications was calculated by using  $\Delta X = [X(\text{without modification}) - X(\text{modification})]$ , in which  $X$  is  $\Delta H^\circ$ ,  $T\Delta S^\circ$ ,  $\Delta G^\circ_{25}$ , or  $T_m$  and included in parentheses.

**Table S4.** The thermodynamic parameters of G-quadruplex forming DNA in the presence of K<sup>+</sup><sup>a</sup>

	without PEG200				40 wt% PEG200			
	$\Delta H^\circ$ ( $\Delta \Delta H^\circ_{\text{mod}}$ ) <sup>b</sup> [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [ $\Delta(T\Delta S^\circ)_{\text{mod}}$ ] [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ ( $\Delta \Delta G^\circ_{25,\text{mod}}$ ) [kcal mol <sup>-1</sup> ] <sup>b</sup>	$T_m(\Delta T_m)$ [°C] <sup>b</sup>	$\Delta H^\circ$ [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ [kcal mol <sup>-1</sup> ]	$T_m$ [°C]
Gq	-53.1 ± 1.1	-45.7 ± 0.9	-7.4 ± 0.2	73.6 ± 0.3	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>5m</sub>	-45.4 ± 2.0 (+7.7)	-39.1 ± 1.8 (+6.6)	-6.3 ± 0.2 (+1.1)	73.0 ± 0.5 (-0.6)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>11m</sub>	-52.5 ± 0.5 (+0.6)	-45.2 ± 0.4 (+0.5)	-7.2 ± 0.1 (+0.2)	72.5 ± 0.1 (-1.1)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>5,11m</sub>	-44.1 ± 3.4 (+9.0)	-38.2 ± 3.0 (+7.5)	-6.0 ± 0.4 (+1.4)	71.8 ± 0.5 (-1.8)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>5hm</sub>	-52.6 ± 2.0 (+0.5)	-45.2 ± 1.7 (+0.5)	-7.5 ± 0.3 (-0.1)	74.4 ± 0.1 (+0.8)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>11hm</sub>	-42.1 ± 3.9 (+11.0)	-36.4 ± 3.3 (+9.3)	-5.7 ± 0.6 (+1.7)	71.3 ± 1.1 (-2.3)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>
Gq <sub>5,11hm</sub>	-47.1 ± 1.6 (+6.0)	-40.5 ± 1.3 (+5.2)	-6.6 ± 0.4 (+0.8)	73.6 ± 1.2 (0.0)	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>	N.D. <sup>c</sup>

<sup>a</sup>The values were determined in 50 mM MES-LiOH (pH 6.0) and 100 mM KCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5 μM of DNA. <sup>b</sup> The amount of change resulting from the modifications was calculated by using  $\Delta X = [X(\text{without modification}) - X(\text{modification})]$ , in which  $X$  is  $\Delta H^\circ$ ,  $T\Delta S^\circ$ ,  $\Delta G^\circ_{25}$ , or  $T_m$  and included in parentheses. <sup>c</sup> “N.D.” indicates that values could not be obtained from the melting curves.

**Table S5.** The thermodynamic parameters of G-quadruplex forming DNA in the presence of  $\text{Na}^+$ <sup>a</sup>

	without PEG200				40 wt% PEG200			
	$\Delta H^\circ$ $(\Delta\Delta H^\circ_{\text{mod}})^b$ [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ $[\Delta(T\Delta S^\circ)_{\text{mod}}]^b$ [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ $(\Delta\Delta G^\circ_{25,\text{mod}})^b$ [kcal mol <sup>-1</sup> ]	$T_m(\Delta T_m)$ [°C] <sup>b</sup>	$\Delta H^\circ$ $(\Delta\Delta H^\circ_{\text{mod}})^b$ [kcal mol <sup>-1</sup> ]	$T\Delta S^\circ$ $[\Delta(T\Delta S^\circ)_{\text{mod}}]^b$ [kcal mol <sup>-1</sup> ]	$\Delta G^\circ_{25}$ $(\Delta\Delta G^\circ_{25,\text{mod}})^b$ [kcal mol <sup>-1</sup> ]	$T_m(\Delta T_m)$ [°C] <sup>b</sup>
Gq	-31.9 ± 0.9	-29.1 ± 0.8	-2.8 ± 0.1	53.7 ± 0.6	-42.2 ± 0.5	-37.3 ± 0.4	-4.9 ± 0.0	64.0 ± 0.3
Gq <sub>5m</sub>	-31.6 ± 0.3 (+0.3)	-28.7 ± 0.2 (+0.4)	-2.9 ± 0.0 (-0.1)	54.9 ± 0.2 (+1.2)	-39.8 ± 0.0 (+2.4)	-35.2 ± 0.0 (+2.1)	-4.6 ± 0.0 (+0.3)	63.7 ± 0.3 (-0.3)
Gq <sub>11m</sub>	-31.8 ± 0.6 (+0.1)	-28.8 ± 0.5 (+0.3)	-3.0 ± 0.1 (-0.2)	55.7 ± 0.3 (+2.0)	-38.6 ± 0.2 (+3.6)	-34.2 ± 0.2 (+3.1)	-4.4 ± 0.1 (+0.5)	63.0 ± 0.5 (-1.0)
Gq <sub>5,11m</sub>	-30.5 ± 0.5 (+1.4)	-27.6 ± 0.5 (+1.5)	-2.9 ± 0.1 (-0.1)	55.8 ± 0.4 (+2.1)	-35.4 ± 0.4 (+6.8)	-31.4 ± 0.4 (+5.9)	-4.0 ± 0.0 (+0.9)	63.0 ± 0.2 (-1.0)
Gq <sub>5hm</sub>	-30.4 ± 0.2 (+1.5)	-27.6 ± 0.2 (+1.5)	-2.8 ± 0.0 (0.0)	55.3 ± 0.2 (+1.6)	-37.3 ± 0.8 (+4.9)	-33.1 ± 0.7 (+4.2)	-4.1 ± 0.1 (+0.8)	62.2 ± 0.1 (-1.8)
Gq <sub>11hm</sub>	-30.2 ± 0.5 (+1.7)	-27.5 ± 0.5 (+1.6)	-2.7 ± 0.0 (+0.1)	54.3 ± 0.1 (+0.6)	-42.0 ± 1.9 (+0.2)	-37.0 ± 1.6 (+0.3)	-5.1 ± 0.2 (-0.2)	66.0 ± 0.2 (+2.0)
Gq <sub>5,11hm</sub>	-31.9 ± 0.9 (0.0)	-29.2 ± 0.8 (-0.1)	-2.8 ± 0.1 (0.0)	53.2 ± 0.8 (-0.5)	-35.0 ± 1.3 (+7.2)	-30.9 ± 1.2 (+6.4)	-4.1 ± 0.2 (+0.8)	64.2 ± 0.4 (+0.2)

<sup>a</sup>The values were determined in 50 mM MES-LiOH (pH 6.0) and 100 mM NaCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5  $\mu\text{M}$  of DNA. <sup>b</sup> The amount of change resulting from the modifications was calculated by using  $\Delta X = [X(\text{without modification}) - X(\text{modification})]$ , in which  $X$  is  $\Delta H^\circ$ ,  $T\Delta S^\circ$ ,  $\Delta G^\circ_{25}$ , or  $T_m$  and included in parentheses.

**Table S6.** Melting temperatures of i-motif forming DNA in the presence of complementary G-quadruplex forming DNA and K<sup>+</sup><sup>a</sup>

	$T_m$ (°C)	100 mM KCl 40 wt% PEG200
iM and Gq	39.3 ± 1.1	39.5 ± 0.4
iM <sub>10,16m</sub> and Gq <sub>5,11m</sub>	N.D. <sup>b</sup>	41.1 ± 0.3
iM <sub>10,16hm</sub> and Gq <sub>5,11hm</sub>	N.D. <sup>b</sup>	37.5 ± 0.3

<sup>a</sup>The values were determined by UV melting curves at 295 nm in 50 mM MES-LiOH (pH 6.0) and 100 mM KCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5 μM of DNA. <sup>b</sup>“N.D.” indicates that values could not be obtained from the melting curves.

**Table S7.** Melting temperatures of mixture of i-motif forming DNA and G-quadruplex forming DNA in the presence of  $\text{Na}^+$ <sup>a</sup>

	$T_m$ (°C)	
	100 mM NaCl	100 mM NaCl 40 wt% PEG200
iM and Gq	78.5 ± 1.1	61.4 ± 1.1
iM <sub>10,16m</sub> and Gq <sub>5,11m</sub>	79.8 ± 0.7	62.4 ± 1.7
iM <sub>10,16hm</sub> and Gq <sub>5,11hm</sub>	77.6 ± 0.8	60.7 ± 1.7

<sup>a</sup>The values were determined by UV melting curves at 260 nm in 50 mM MES-LiOH (pH 6.0) and 100 mM NaCl with or without 40 wt% PEG200. Each value is the average of three determinations and the standard deviation is shown with the average. Melting temperatures were measured at 5  $\mu\text{M}$  of DNA.