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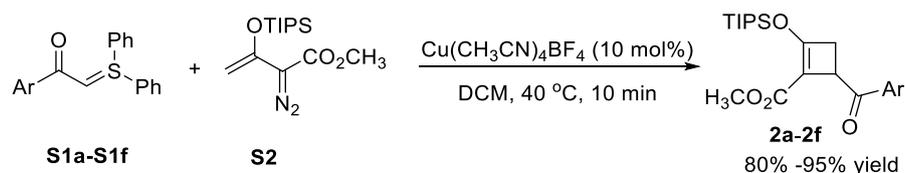
1. General information

All reactions, unless noted, were performed in oven-dried (120 °C) glassware with magnetic stirring under an inert atmosphere of dry nitrogen. Analytical thin layer chromatography (TLC) was carried out using EM Science silica gel 60 F254 plates; visualization was accomplished with UV light (254 nm). Column chromatography was performed on CombiFlash® Rf200 and Rf+ purification systems using normal phase disposable columns. ¹H, ¹³C, ¹⁹F NMR spectra were recorded on a Bruker spectrometer (300 or 500 MHz). Chemical shifts were reported in ppm downfield from tetramethylsilane (TMS) with the solvent resonance as the internal standard (CDCl₃, δ = 7.26). Spectra were reported as follows: chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, comp = composite of magnetically non-equivalent protons, dd = doublet of doublets), coupling constants (Hz), integration and assignment. ¹³C NMR spectra were collected on Bruker instruments (75 or 126 MHz) with complete proton decoupling. Chemical shifts are reported in ppm from the tetramethylsilane with the solvent resonance as internal standard (CDCl₃, δ = 77.0). High-resolution mass spectra (HRMS) were performed on a Bruker MicroTOF-ESI mass spectrometer with an ESI resource using CsI or LTQ ESI positive ion calibration solution as the standard. Enantioselectivities were determined by HPLC analysis at 25 °C using an Agilent 1260 Infinity HPLC System equipped with an G1311B quaternary pump, G1315D diode array detector, G1329B auto-sampler, G1316A thermostated column compartment and G1170A valve drive. For instrument control and data processing, Agilent OpenLAB CDS ChemStation Edition for LC & LC/MS Systems (Rev. C.01.07 [26]) software was used. The Chiralpak AD-H column was obtained from Daicel Chiral Technologies. Tetrahydrofuran, dichloromethane, chloroform, and toluene were purified using a JC-Meyer solvent purification system. Optical rotations were reported as follows: [*a*]_D²³ (c: g/100 mL, in DCM).

2. Preparation of Reactants

1*H*-Isochromene acetals (**1**)¹, donor-acceptor cyclobutenes (**2**)², donor-acceptor cycloprpenes (**4**)³, siloxyalkynes (**6**)⁴, donor-acceptor azetine **9**⁵ and other metallo-vinyl carbenes cycloaddition products (**10** and **11**)⁶ were prepared by reported methods.

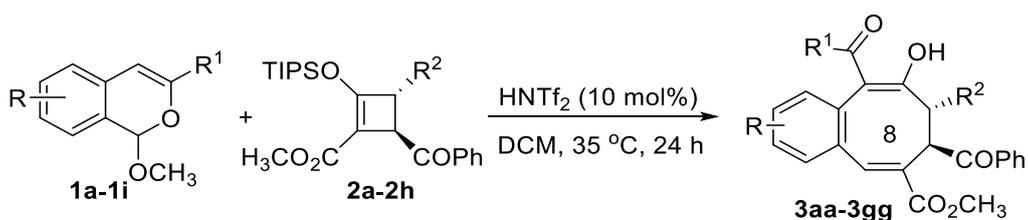
Figure 1. General procedure for the synthesis of Donor-Acceptor Cyclobutenes.



The modified procedure for the synthesis of donor-acceptor cyclobutenes (**2a-2f**) is described: To a solution of sulfur ylide (**S1a-S1f**, 3.0 mmol) and $\text{Cu(CH}_3\text{CN)}_4\text{BF}_4$ (94.3 mg, 0.3 mmol, 10 mol%) in 10 mL CH_2Cl_2 at 40 °C was slowly added a solution of enoldiazo compounds **S2** (3.6 mmol, 1.2 equiv.) in 3 mL CH_2Cl_2 over 5 minutes. The resulting solution was stirred at 40 °C for 10 min, after which the crude product was purified by flash chromatography (hexane/ethyl acetate = 19/1) to give donor-acceptor cyclobutenes (**2a-2f**) in excellent yield.

3. General procedure for the catalyzed [4+4]-, [4+3]- and [4+2]-cycloaddition reactions.

Figure 2. General procedure for the catalyzed [4+4]-cycloaddition reactions.



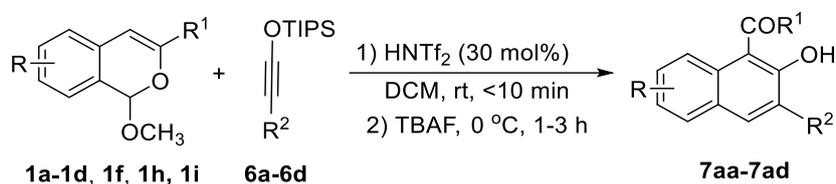
The donor-acceptor cyclobutene (**2a-2h**) (0.13 mmol, 1.3 equiv.) was dissolved in dry CH_2Cl_2 (1 mL), after which a solution of HNTf_2 (10 mol%) in dry CH_2Cl_2 (0.1 mL) was added slowly. The mixture was stirred at 35 °C for 5 min, then a solution of acetal compounds (**1a-1i**) (0.1 mmol, 1.0 equiv.) in 1 mL CH_2Cl_2 was added over 30 minutes. The reaction solution was stirred at the same temperature for 24 h, and the progress of the reaction was monitored by TLC. After complete reaction of the acetal substrate, the reaction solution was subjected to flash chromatography on silica gel with a 5:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford the desired products **3aa-3gg**.

Figure 3. General procedure for the catalyzed [4+3]-cycloaddition reactions.



The acetal compound (**1a-1j**) (0.2 mmol, 1.0 equiv.) and 4 Å molecular sieves (50 mg) were dissolved in dry CH₂Cl₂ (1 mL), and a solution of HNTf₂ (20 mol%) in dry CH₂Cl₂ (0.4 mL) was added slowly, after which a solution of donor-acceptor cyclopropene (**4a-4d**) (0.24 mmol, 1.2 equiv.) in dry CH₂Cl₂ (1.0 mL) was added over 5 min. The reaction solution was stirred at room temperature for 2 h, and the progress of the reaction was monitored by TLC. After complete reaction of the acetal substrate, the reaction solution was subjected to flash chromatography on silica gel with a 9:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford the desired product (**5aa-5ad**).

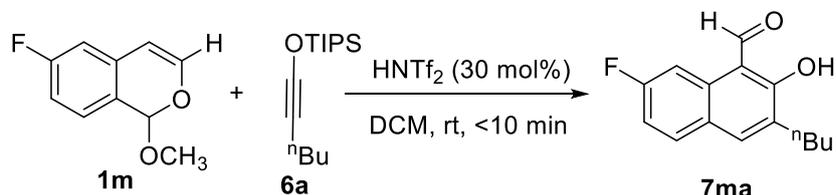
Figure 4. General procedure for the catalyzed [4+2]-cycloaddition reactions.



Acetal compound (**1a-1d**, **1f**, **1h** and **1i**) (0.2 mmol, 1.0 equiv.) and siloxyalkynes **6a-6d** (0.3 mmol, 1.5 equiv.) were dissolved in dry CH₂Cl₂ (2 mL), and a solution of HNTf₂ (20 mol%) in dry CH₂Cl₂ (0.4 mL) was added slowly, after which another solution of siloxyalkynes (**6a-6d**) (0.1 mmol, 0.5 equiv.) in dry CH₂Cl₂ (0.5 mL) was added in the reaction solution, then HNTf₂ (10 mol%) was slowly added to the reaction solution. The progress of the reaction was monitored by TLC.

After complete reaction of the acetal substrate (<10 min), the reaction solution was cooled to 0 °C, and TBAF (0.4 mmol, 2.0 equiv., 1.0 M in THF) was added. The reaction solution was then stirred at 0 °C for 1-3 h, and the progress of the reaction was monitored by TLC. Upon completion of the reaction the solvent was removed, and the residue was subjected to flash chromatography on silica gel with a 9:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford the desired products **7aa-7ad**.

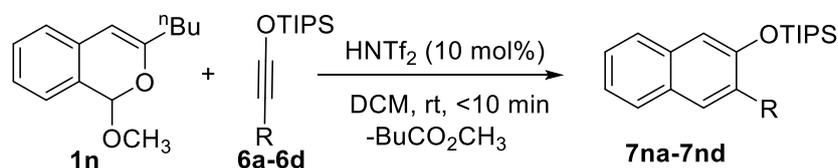
Figure 5. General procedure for the catalyzed [4+2]-cycloaddition reaction of **1m** with **6a**.



Acetal compound **1m** (36 mg, 0.2 mmol, 1.0 equiv.) and siloxyalkyne **6a** (76.2 mg, 0.3 mmol, 1.5 equiv.) were dissolved in dry CH₂Cl₂ (2 mL), and a solution of HNTf₂ (20 mol%) in dry CH₂Cl₂ (0.4 mL) was added slowly. Then another solution of siloxyalkyne **6a** (25.4 mg, 0.1 mmol, 0.5 equiv.) in dry CH₂Cl₂ (0.5 mL) was added in the reaction solution, after which HNTf₂ (10 mol%) was slowly added. The progress of the reaction was monitored by TLC. After complete reaction of the acetal substrate (<10

min), the reaction solution was subjected to flash chromatography on silica gel with a 10:1 (v/v) mixture of hexane: CH₂Cl₂ as the eluent to afford the desired products **7ma**.

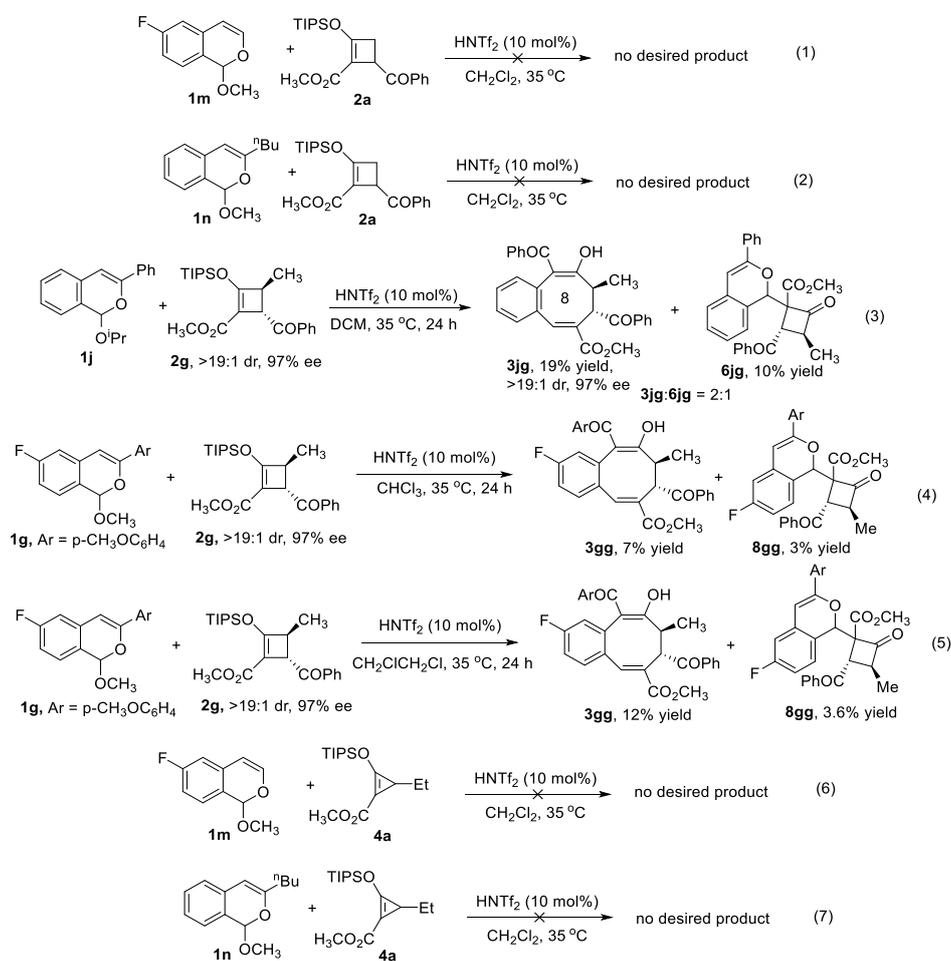
Figure 6. General procedure for the catalyzed [4+2]-cycloaddition reactions of **1n** with siloxyalkynes **6a-6d**.



Acetal compound **1n** (43.6 mg, 0.2 mmol, 1.0 equiv.) and siloxyalkynes **6a-6d** (0.3 mmol, 1.5 equiv.) were dissolved in dry CH₂Cl₂ (2 mL), and a solution of HNTf₂ (10 mol%) in dry CH₂Cl₂ (0.2 mL) was added slowly. The progress of the reaction was monitored by TLC. After complete reaction of acetal substrate (<10 min), the reaction solution was subjected to flash chromatography on silica gel with a 19:1 (v/v) mixture of hexane: ethyl acetate as the eluent to afford the desired products **7na-7nd**.

4. Constraints of these Brønsted acid catalyzed cycloadditions of benzopyrylium.

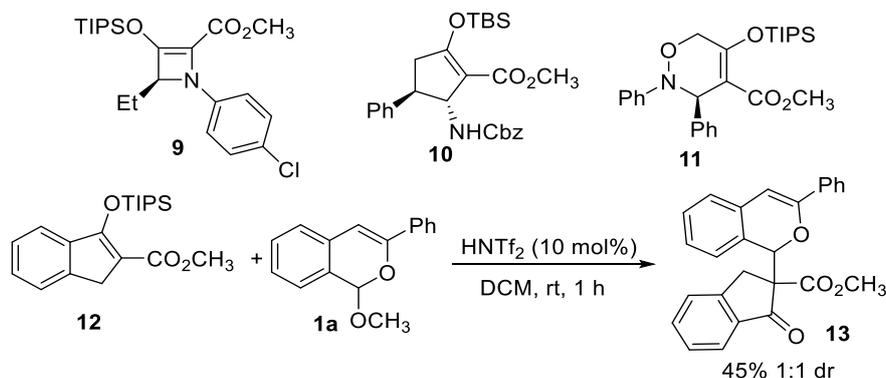
Figure 7. Constraints of these Brønsted acid catalyzed cycloadditions.



Acetal compounds **1m** and **1n** were employed in [4+4]-cycloaddition reactions with **2a** but did not produce any of the desired products. Spectral analysis showed that mostly starting material remained [Eq(1-2)]. The isopropyl acetal substrate **1j**, which forms the bulky 2-propanol nucleophile, failed to suppress the competing desilylation pathway [Eq(3)]. When CHCl_3 and $\text{CH}_2\text{ClCH}_2\text{Cl}$ were used as solvent, the [4+4] cycloaddition reaction was inefficient, and the desired product was isolated in only 7% and 12% yields [Eq(4-5)]. The reaction between acetal compounds **1m** and **1n** and donor-acceptor cyclopropene **4a** did not produce any of the desired products. Spectral analysis showed that mostly starting material remained [Eq(6-7)].

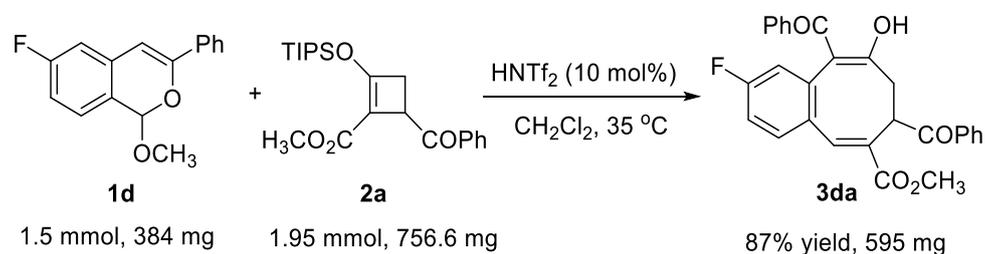
Other constraints were noted in our evaluation of this methodology (Figure 1). As expected, nitrogen-containing donor-acceptor azetine (**9**) and other metallovinylcarbenes cycloaddition products (**10** and **11**) were unreactive, and the reactants were recovered. In addition, compound **12** reacted with acetal (**1a**), giving only the direct addition product **13** in good yield, which suggests unfavorable competition for cycloaddition from desilylation.

Figure 8. Observed other reaction constraints.



5. General procedure for large scale reaction, further transformations and discussion.

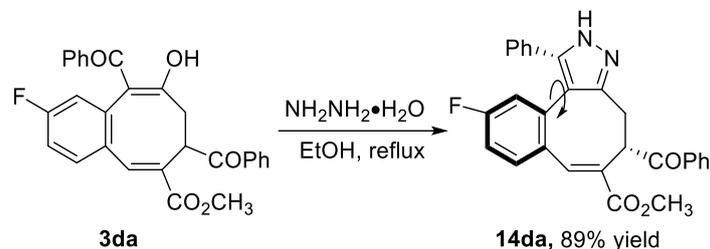
Figure 9. General procedure for large scale reaction of **1d** with **2a**.



Donor-acceptor cyclobutene **2a** (756.6 mg, 1.95 mmol, 1.3 equiv.) was dissolved in dry CH_2Cl_2 (10 mL), after which a solution of HNTf_2 (10 mol%) in dry CH_2Cl_2 (1.5 mL) was added slowly. The solution was stirred at $35\text{ }^\circ\text{C}$ for 5 min, then a solution of acetal compounds **1d** (384 mg, 1.5 mmol, 1.0 equiv.) in 5 mL CH_2Cl_2 was added over 30 minutes. The reaction solution was stirred at same temperature for 24 h, and the progress of the reaction was monitored by TLC. After complete reaction of the acetal

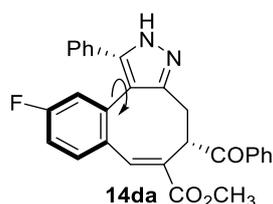
substrate, the reaction solution was subjected to flash chromatography on silica gel with a 5:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford the desired product **3da** in 87% yield.

Figure 10. General procedure for further transformation of **3da** with Hydrazine.



A screw-capped 25-mL vial equipped with a magnetic stirring bar was loaded with compound **3da** (0.15 mmol) and anhydrous ethanol (2.5 mL). Hydrazine hydrate (45 wt. % N_2H_4 , 0.225 mmol, 1.5 equiv.) in anhydrous ethanol (2.5 mL) was added in one portion at room temperature, and the reaction solution was stirred at 90 °C for 1 h. The progress of the reaction was monitored by TLC. After complete conversion of starting material **3da**, the solvent was evaporated, and the residue was purified by column chromatography on silica gel with a 2:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford pyrazole **14da**.

Methyl (*E*)-5-Benzoyl-10-fluoro-1-phenyl-4,5-dihydro-2*H*-benzo[3,4]-cycloocta[1,2-*c*]pyrazole-6-carboxylate.



White solid (60 mg, 89% yield), 0.2 mmol scale reaction. m.p. 138 – 139 °C.

Major rotamer:

$^1\text{H NMR}$ (500 MHz, chloroform-*d*) δ 11.57 (s, 1H), 8.05 (s, 1H), 7.60 (d, $J = 7.5$ Hz, 2H), 7.51 – 7.48 (comp., 2H), 7.38 (t, $J = 7.5$ Hz, 2H), 7.28 – 7.23 (comp., 3H), 7.18 – 7.13 (comp., 3H), 6.81 (dd, $J = 10.0, 2.5$ Hz, 1H), 4.51 (dd, $J = 10.0, 6.0$ Hz, 1H), 3.61 – 3.53 (comp., 5H).

$^{13}\text{C NMR}$ (126 MHz, chloroform-*d*) δ 197.98, 165.46, 162.55 (d, $J = 254$ Hz, 1C), 142.14, 136.63, 135.95 (d, $J = 9$ Hz, 1C), 132.66, 131.17, 129.05, 128.98, 128.49, 128.39, 128.00, 127.89, 119.93 (d, $J = 21$ Hz, 1C), 114.62 (d, $J = 21$ Hz, 1C), 114.63, 52.04, 44.54, 25.91.

$^{19}\text{F NMR}$ (470.5 MHz, chloroform-*d*) δ -112.2 (m, 1F).

Minor rotamer:

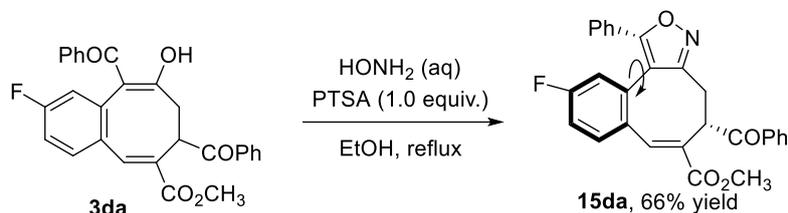
$^1\text{H NMR}$ (500 MHz, chloroform-*d*) δ 7.90 (s, 1H), 7.65 – 7.58 (m, 1H), 7.52 – 7.48 (comp, 2H), 7.39 – 7.36 (m, 1H), 7.33 – 7.23 (comp, 3H) 7.19 – 7.13 (comp, 3H), 6.73 (t, $J = 8.5$ Hz, 1H), 6.61 (d, $J = 8.5$ Hz, 2H), 4.92 (t, $J = 5.5$ Hz, 1H), 3.88 (s, 3H), 3.45 (dd, $J = 15.5, 5.5$ Hz, 1H), 3.03 (dd, $J = 15.5, 5.5$ Hz, 1H).

$^{13}\text{C NMR}$ (126 MHz, chloroform-*d*) δ 197.76, 167.37, 162.38 (d, $J = 250$ Hz, 1C), 142.28, 135.53, 133.37, 131.83, 130.13, 128.30, 128.18, 128.09, 118.16 (d, $J = 21$ Hz, 1C), 113.89 (d, $J = 21$ Hz, 1C), 52.78, 45.76, 25.47.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -112.6 (m, 1F).

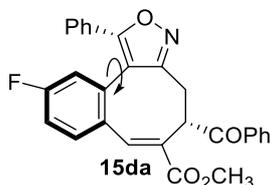
HRMS (ESI) scaled for [M+H]⁺: C₂₈H₂₂FN₂O₃, m/z: 453.1609, observed: 453.1598.

Figure 11. General procedure for further transformation of **3da** with Hydroxylamine.



A screw-capped 25 mL vial equipped with a magnetic stirring bar was loaded with compound **3da** (0.1 mmol), *p*-toluenesulfonic acid monohydrate (PTSA·H₂O, 0.1 mmol) and anhydrous ethanol (2.5 mL). Hydroxylamine (50% wt. in water, 0.3 mmol) in anhydrous ethanol (2.5 mL) was added in one portion at room temperature, and the reaction solution was stirred at 90 °C for 3 days. After that, hydroxylamine (50% wt. in water, 0.3 mmol) in anhydrous ethanol (2.5 mL) was added again, and the reaction solution was stirred at 90 °C for an additional 3 days. Solvents were evaporated, and the residue was purified by column chromatography on silica gel with a 3:1 (v/v) mixture of hexane:ethyl acetate as the eluent to afford pyrazole **15da**.

Methyl (E)-5-Benzoyl-10-fluoro-1-phenyl-4,5-dihydrobenzo[3,4]-cycloocta[1,2-*c*]isoxazole-6-carboxylate.



White solid (30 mg, 66% yield), 0.1 mmol scale reaction. m.p. 218 – 219 °C.

Major rotamer:

¹H NMR (500 MHz, chloroform-*d*) δ 7.97 (s, 1H), 7.64 (d, *J* = 7.2 Hz, 1H), 7.59 – 7.53 (comp., 2H), 7.51 (t, *J* = 7.0 Hz, 2H), 7.56 – 7.54 (comp., 3H), 7.36 – 7.29 (comp, 3H), 7.27 – 7.22 (comp., 1H), 7.03 (dd, *J* = 9.5, 2.5 Hz, 1H), 4.42 (dd, *J* = 12.5, 5.5 Hz, 1H), 3.75 (dd, *J* = 17.0, 12.5 Hz, 1H), 3.59 (s, 3H), 3.44 (dd, *J* = 17.0, 5.5 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 197.07, 167.20, 164.92, 162.80 (d, *J* = 254 Hz, 1C), 160.53, 141.02, 136.67, 135.70, 133.08, 132.66, 131.38 (d, *J* = 9 Hz, 1C), 130.13, 128.73, 128.42, 127.82, 127.51, 119.26 (d, *J* = 21 Hz, 1C), 116.25 (d, *J* = 21 Hz, 1C), 113.36, 52.10, 44.16, 25.10.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -110.9 (m, 1F).

Minor rotamer:

¹H NMR (500 MHz, chloroform-*d*) δ 7.86 (s, 1H), 6.93 (td, *J* = 8.5, 2.5 Hz, 1H), 6.88 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.77 (dd, *J* = 8.5, 5.5 Hz, 1H), 5.11 (dd, *J* = 7.0, 5.0 Hz, 1H), 3.86 (s, 3H), 3.55 (dd, *J* = 16.0, 5.0 Hz, 1H), 3.35 (dd, *J* = 16.0, 5.0 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 197.36, 166.91, 165.97, 162.60 (d, *J* = 254 Hz, 1C), 161.09, 141.23, 135.21, 133.21, 130.88, 128.39, 128.34, 127.16, 127.04, 117.78, 117.69 (d, *J* = 22 Hz, 1C), 115.49 (d, *J* = 22 Hz, 1C), 114.00, 52.88, 45.45, 23.25.

^{19}F NMR (470.5 MHz, chloroform- d) δ -111.5 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{28}\text{H}_{21}\text{FNO}_4$, m/z : 454.1449, observed: 454.1431.

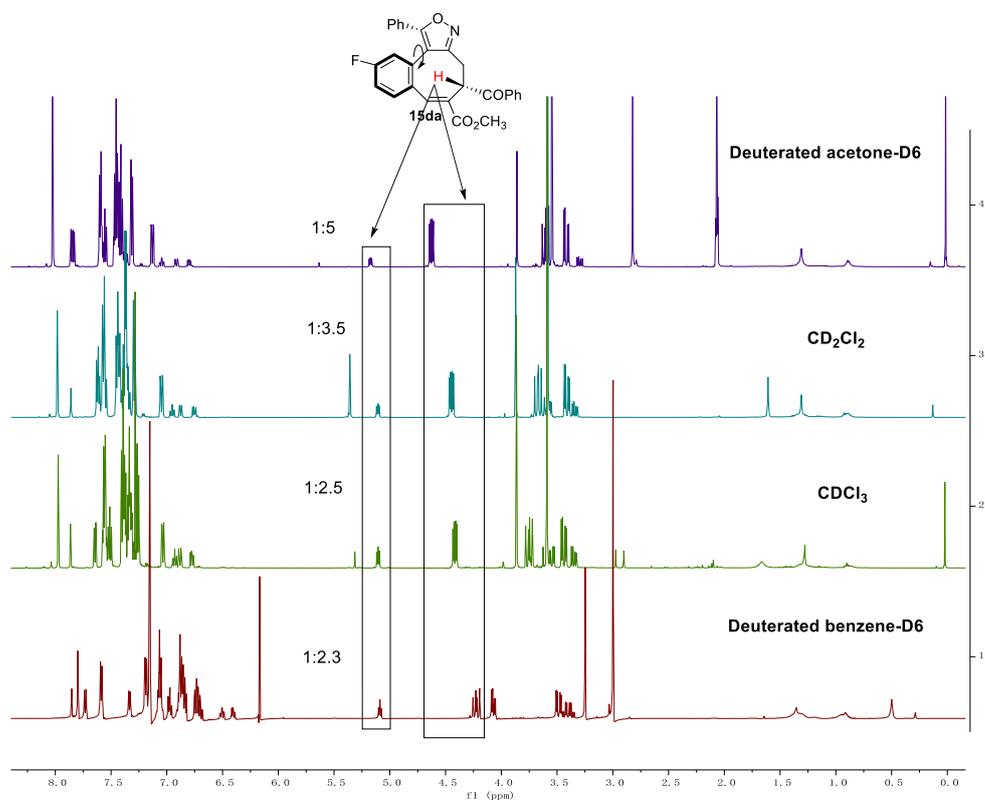


Figure 12. NMR spectra of compound **15da** in different deuterated solvents.

Discussion 1. The ratio of the proton NMR signals are different in different deuterated solvents, indicating that rotamers exist in solution of product **15da**.

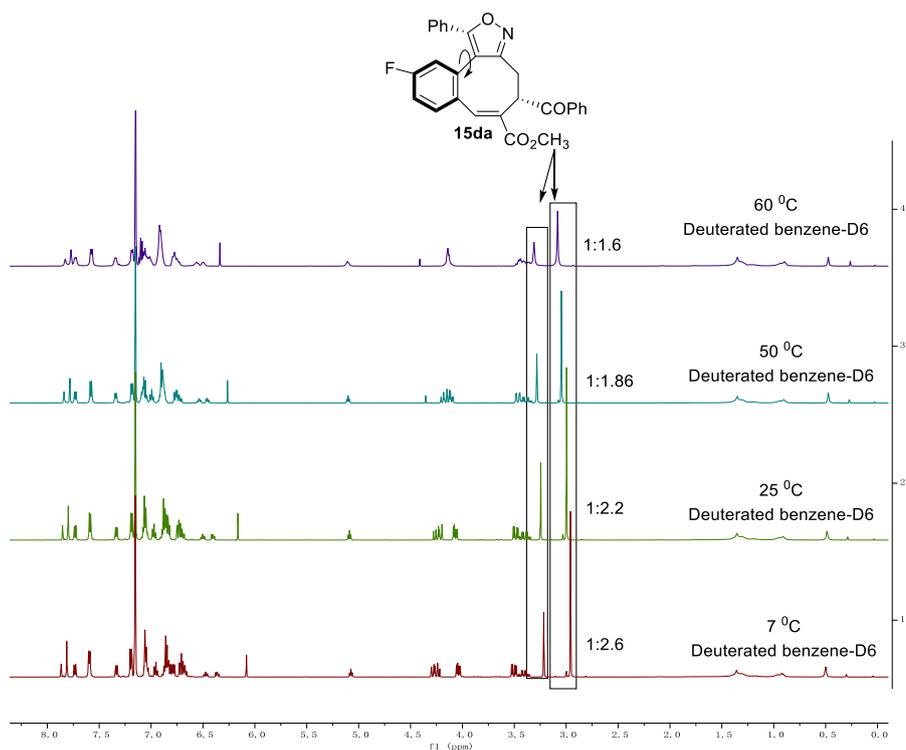


Figure 13. The NMR spectra of compound **15da** in different temperatures.

Discussion 2. The ratio of the proton NMR spectra reversibly change at different temperatures, indicating the existence of rotamers.

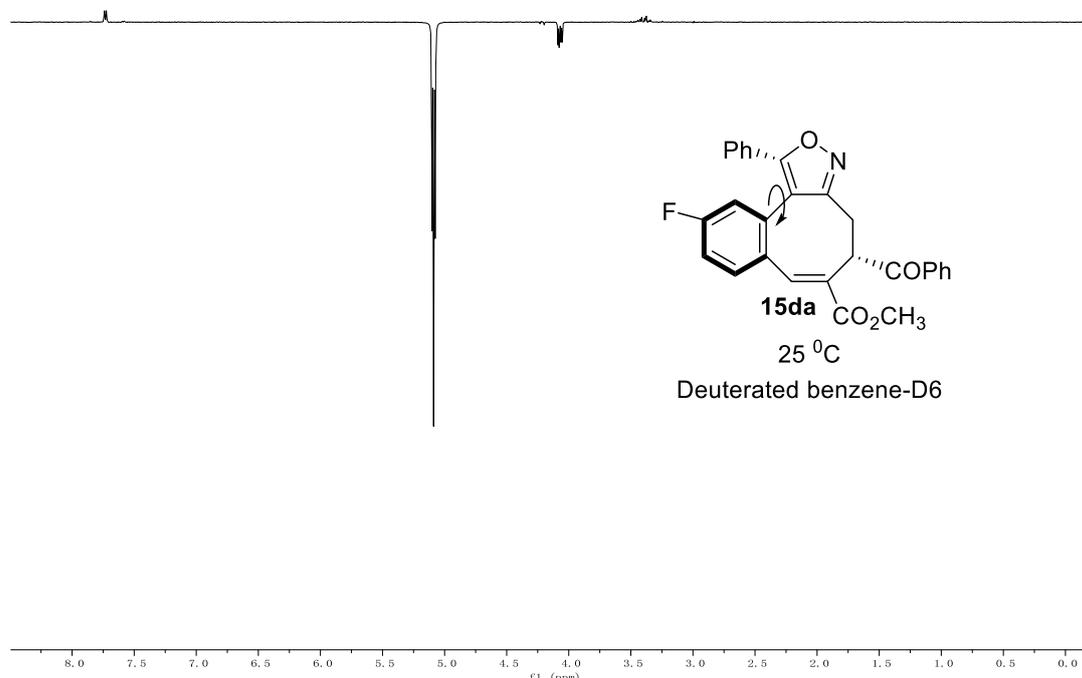
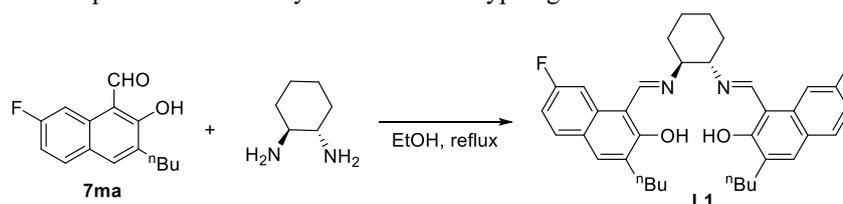


Figure 14. The 1D- gradient NOE spectra experiment of **15da**.

6. General procedure for the synthesis of salen type ligands L1 and L2

Figure 15. General procedure for the synthesis of salen type ligand L1.



1,1'-((1E,1'E)-((1S,2S)-Cyclohexane-1,2-diyl)bis(azanylylidene))bis(methanylylidene))bis(3-butyl-7-fluoronaphthalen-2-ol)

To a solution of (*S,S*)-1,2-cyclohexanediamine (57 mg, 0.5 mmol, 1.0 equiv.) in EtOH (5 mL) was added a solution of **7ma** (270.6, 1.1 mmol, 2.1 equiv.) in EtOH (5 mL). The suspension was refluxed for 12 h at which time a yellow solution was evident. The mixture was cooled to room temperature, and the solvent was removed. The reaction mixture was purified by flash chromatography on silica gel (15% EtOAc/hexanes) to give **L1** (151 mg, 53%) as an amorphous yellow solid: mp 60 – 61 °C.

¹H NMR (500 MHz, chloroform-*d*) δ 14.63 (s, 2H), 8.46 (s, 2H), 7.32 (dd, *J* = 8.5, 6.0 Hz, 2H), 7.26 (s, 2H), 7.22 (dd, *J* = 11.5, 2.5 Hz, 2H), 6.83 (td, *J* = 8.5, 2.5 Hz, 2H), 3.45 – 3.29 (m, 2H), 2.71 – 2.65 (m, 2H), 2.57 – 2.51 (m, 2H), 2.24 (d, *J* = 14.0 Hz, 2H), 1.96 (d, *J* = 8.5 Hz, 2H), 1.85 – 1.70 (m, 2H), 1.66 – 1.55 (m, 4H), 1.55 – 1.39 (comp., 6H), 0.97 (t, *J* = 7.5 Hz, 6H).

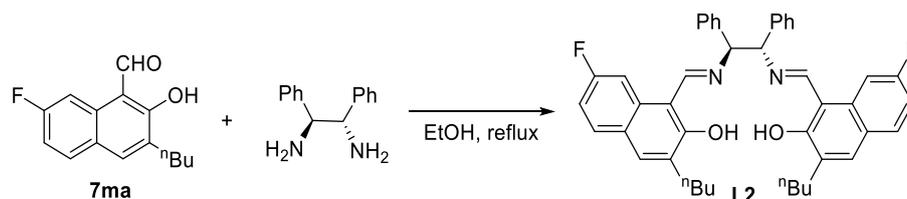
^{13}C NMR (126 MHz, chloroform-*d*) δ 174.27, 161.92 (d, $J = 245$ Hz, 1C), 158.76, 134.84, 134.00, 133.92, 130.19 (d, $J = 9$ Hz, 1C), 123.04, 111.35 (d, $J = 24$ Hz, 1C), 106.04 (d, $J = 3.9$ Hz, 1C), 102.90 (d, $J = 24$ Hz, 1C), 68.55, 32.44, 31.04, 29.53, 24.38, 22.81, 14.02.

^{19}F NMR (470.5 MHz, chloroform-*d*) δ -113.1 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{36}\text{H}_{41}\text{F}_2\text{N}_2\text{O}_2$, m/z : 571.3131, observed: 571.3114.

$[\alpha]_{\text{D}}^{23} = +460.98$ (c 0.31, CH_2Cl_2).

Figure 16. General procedure for the synthesis of salen type ligand **L2**.



1,1'-(1*E*,1'*E*)-[(1*S*,2*S*)-1,2-Diphenylethane-1,2-diyl]bis(azanylylidene))bis(methanylylidene)]-bis(3-butyl-7-fluoronaphthalen-2-ol)

To a solution of (*S,S*)-1,2-diphenylethylenediamine (40.3 mg, 0.19 mmol, 1.0 equiv.) in EtOH (5 mL) was added a solution of **7ma** (93.5 mg, 0.38 mmol, 2 equiv.) in EtOH (5 mL). The suspension was refluxed for 12 h at which time a yellow solution was evident. The mixture was cooled to room temperature, and the solvent was removed. The crude mixture was purified by flash chromatography on silica gel (35% EtOAc/hexanes) to give **L2** (94 mg, 74%) as an amorphous yellow solid: mp 65 – 66 °C. ^1H NMR (500 MHz, chloroform-*d*) δ 15.41 (s, 2H), 8.82 (s, 2H), 7.45 – 7.38 (comp., 4H), 7.34 (s, 2H), 7.32 – 7.25 (comp., 10H), 6.93 (td, $J = 8.5, 2.5$ Hz, 2H), 4.85 (s, 2H), 2.73 – 2.57 (m, 4H), 1.69 – 1.52 (m, 4H), 1.47 – 1.40 (m, 4H), 0.99 (t, $J = 7.5$ Hz, 6H).

^{13}C NMR (126 MHz, chloroform-*d*) δ 169.12, 161.76 (d, $J = 245$ Hz, 1C), 160.84, 137.97, 133.43, 133.34, 133.16, 130.33 (d, $J = 9.7$ Hz, 1C), 128.72, 128.09, 127.76, 123.63, 111.96 (d, $J = 24$ Hz, 1C), 107.07 (d, $J = 4$ Hz, 1C), 103.31 (d, $J = 24$ Hz, 1C), 77.95, 30.96, 29.41, 22.74, 14.05.

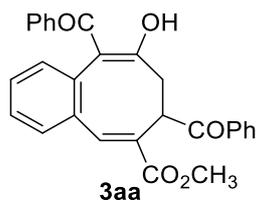
^{19}F NMR (470.5 MHz, chloroform-*d*) δ -113.1 (m, 1F).

HRMS (ESI) scaled for; $[\text{M}+\text{H}]^+$: $\text{C}_{44}\text{H}_{43}\text{F}_2\text{N}_2\text{O}_2$, m/z : 669.3293, observed: 669.3284.

$[\alpha]_{\text{D}}^{23} = +134.44$ (c 0.30, CH_2Cl_2).

7. Analytical and spectral characterization data for reaction products

Methyl (*5E*, *9Z*)-7,10-Dibenzoyl-9-hydroxy-7,8-dihydrobenzo[8]-annulene-6-carboxylate.



Colorless oil (33 mg, 76% yield), 0.1 mmol scale reaction.

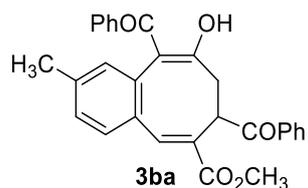
^1H NMR (500 MHz, chloroform-*d*) δ 8.32 (s, 1H), 8.11 (d, $J = 7.5$ Hz, 2H), 7.62 (t, $J = 7.5$ Hz, 1H), 7.53 – 7.52 (comp, 3H), 7.34 – 7.31 (comp, 2H), 7.25 – 7.20 (comp, 4H), 7.04 (t, $J = 7.5$ Hz, 1H), 6.77

(d, $J = 7.5$ Hz, 1H), 5.27 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.74 (s, 3H), 2.95 (t, $J = 12.5$ Hz, 1H), 2.50 (dd, $J = 12.5, 4.5$ Hz, 1H).

^{13}C NMR (126 MHz, chloroform- d) δ 198.2, 192.2, 186.7, 167.6, 142.3, 135.9, 135.8, 135.1, 133.9, 133.5, 132.2, 131.1, 130.7, 129.9, 129.3, 128.9, 128.8, 128.0, 127.8, 127.7, 52.5, 46.9, 37.0.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{28}\text{H}_{23}\text{O}_5$, m/z : 439.1540, observed: 439.1541.

Methyl (5*E*, 9*Z*)-7,10-Dibenzoyl-9-hydroxy-2-methyl-7,8-dihydro-benzo[8]annulene-6-carboxylate.



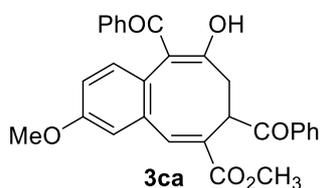
Colorless oil (29.4 mg, 65% yield), 0.1 mmol scale reaction.

^1H NMR (500 MHz, chloroform- d) δ 8.31 (s, 1H), 8.12 (d, $J = 8.0$ Hz, 2H), 7.62 (t, $J = 7.5$ Hz, 1H), 7.53 (t, $J = 7.5$ Hz, 2H), 7.43 (d, $J = 8.0$ Hz, 1H), 7.35 (t, $J = 7.0$ Hz, 1H), 7.28 – 7.19 (comp, 4H), 7.15 (d, $J = 8.0$ Hz, 1H), 6.59 (s, 1H), 5.26 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.73 (s, 3H), 2.98 (t, $J = 12.5$ Hz, 1H), 2.50 (dd, $J = 12.5, 4.5$ Hz, 1H), 2.10 (s, 3H).

^{13}C NMR (126 MHz, chloroform- d) δ 198.3, 192.2, 186.6, 167.8, 142.5, 138.0, 135.9, 135.1, 133.7, 133.4, 132.9, 132.7, 131.0, 130.2, 129.9, 129.3, 128.9, 128.8, 128.6, 127.8, 112.8, 52.5, 47.0, 37.0, 20.9.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{29}\text{H}_{25}\text{O}_5$, m/z : 453.1697, observed: 453.1698.

Methyl (5*E*, 9*Z*)-7,10-Dibenzoyl-9-hydroxy-3-methoxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



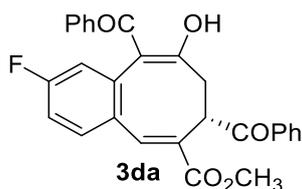
Colorless oil (26 mg, 55% yield), 0.1 mmol scale reaction.

^1H NMR (500 MHz, chloroform- d) δ 8.29 (s, 1H), 8.12 (d, $J = 7.5$ Hz, 2H), 7.63 (t, $J = 7.5$ Hz, 1H), 7.54 (t, $J = 7.5$ Hz, 2H), 7.35 (t, $J = 7.0$ Hz, 1H), 7.29 – 7.22 (comp, 5H), 7.06 (s, 1H), 6.67 (d, $J = 8.5$ Hz, 1H), 6.61 (d, $J = 8.5$ Hz, 1H), 5.28 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.85 (s, 3H), 3.75 (s, 3H), 3.00 (t, $J = 12.5$ Hz, 1H), 2.51 (dd, $J = 12.5, 4.5$ Hz, 1H).

^{13}C NMR (126 MHz, chloroform- d) δ 198.2, 192.5, 186.4, 167.7, 158.9, 142.1, 136.8, 136.0, 135.1, 133.5, 133.3, 131.0, 130.7, 129.4, 129.0, 128.8, 127.9, 126.2, 114.8, 113.9, 112.3, 55.4, 52.5, 47.1, 37.0.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{29}\text{H}_{25}\text{O}_6$, m/z : 469.1646, observed: 469.1651.

Methyl (5*S*, 5*E*, 9*Z*)-7,10-Dibenzoyl-2-fluoro-9-hydroxy-7,8-dihydro-benzo[8]annulene-6-carboxylate.



White solid (64 mg, 81% yield, 80% ee), 0.2 mmol scale reaction. m.p. 171 – 172 °C.

Optical purity determined by HPLC analysis [Daicel chiralpak ADH, *n*-hexane/*i*-PrOH = 75/25, 1.0 mL/min, λ = 254 nm, t_1 = 9.10 min, t_2 = 19.63 min].

¹H NMR (500 MHz, chloroform-*d*) δ 8.27 (s, 1H), 8.11 (d, J = 8.5 Hz, 2H), 7.63 (t, J = 7.5 Hz, 1H), 7.56 – 7.49 (comp, 3H), 7.42 – 7.36 (m, 1H), 7.28 – 7.19 (comp, 4H), 7.06 (td, J = 8.5, 2.5 Hz, 1H), 6.49 (dd, J = 9.5, 2.5 Hz, 1H), 5.28 (dd, J = 13.5, 4.5 Hz, 1H), 3.74 (s, 3H), 2.92 (t, J = 12.0 Hz, 1H), 2.53 (dd, J = 12.0, 4.5 Hz, 1H).

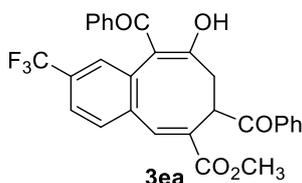
¹³C NMR (126 MHz, chloroform-*d*) δ 198.02, 192.00, 187.00, 167.49, 161.70 (d, J = 254 Hz, 1C), 141.27, 136.31 (d, J = 8.4 Hz, 1C), 135.48, 134.99, 133.58, 131.81, 131.74, 131.42, 131.03, 129.20, 128.98, 128.78, 128.03, 118.74 (d, J = 21 Hz, 1C), 115.21 (d, J = 21 Hz, 1C), 112.06, 52.57, 46.75, 37.04.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₈H₂₂FO₅, m/z : 457.1446, observed: 457.1453.

$[\alpha]_D^{23}$ = +55.4 (*c* 0.43, CH₂Cl₂).

Methyl (5E, 9Z)-7,10-Dibenzoyl-9-hydroxy-2-(trifluoromethyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (25.3 mg, 50% yield), 0.1 mmol scale reaction.

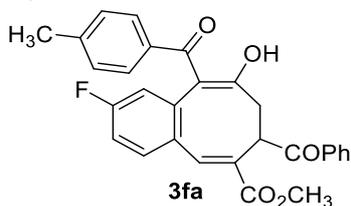
¹H NMR (500 MHz, chloroform-*d*) δ 8.30 (s, 1H), 8.11 (d, J = 8.0 Hz, 2H), 7.82 (s, 1H), 7.64 (t, J = 7.5 Hz, 2H), 7.55 (t, J = 7.5 Hz, 2H), 7.26 - 7.20 (m, 5H), 6.90 (d, J = 8.0 Hz, 1H), 5.30 (dd, J = 12.5, 4.5 Hz, 1H), 3.76 (s, 3H), 2.87 (t, J = 12.5 Hz, 1H), 2.55 (dd, J = 12.5, 4.5 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 197.71, 191.69, 187.58, 167.22, 140.47, 137.85, 136.39, 135.47, 134.86, 133.65, 132.85, 132.28, 131.48, 130.00 (q, J = 32 Hz, 1C), 129.23, 129.00, 128.80, 128.07, 126.91 (q, J = 3.6 Hz, 1C), 123.62 (q, J = 271 Hz, 1C), 124.50 (q, J = 3.6 Hz, 1C), 111.89, 52.69, 46.75, 36.81.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -63.2 (s, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₉H₂₂F₃O₅, m/z : 507.1414, observed: 507.1422.

Methyl (5E, 9Z)-7-Benzoyl-2-fluoro-9-hydroxy-10-(4-methyl-benzoyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (40 mg, 85% yield), 0.1 mmol scale reaction.

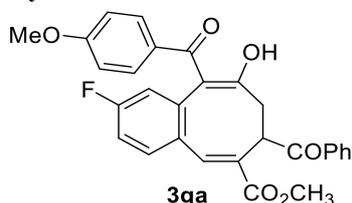
¹H NMR (500 MHz, chloroform-*d*) δ 8.26 (s, 1H), 8.11 (d, $J = 7.5$ Hz, 2H), 7.63 (t, $J = 7.5$ Hz, 1H), 7.58 – 7.47 (comp, 3H), 7.14 (d, $J = 8.0$ Hz, 2H), 7.05 (d, $J = 8.0$ Hz, 3H), 6.52 (dd, $J = 9.5$ Hz, 4.5, 1H), 5.26 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.74 (s, 3H), 2.91 (t, $J = 12.5$ Hz, 1H), 2.51 (dd, $J = 12.5, 4.5$ Hz, 1H), 2.34 (s, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 198.11, 191.63, 186.97, 167.52, 161.69 (d, $J = 254$ Hz, 1C), 142.17, 141.30, 136.56 (d, $J = 8.4$ Hz, 1C), 135.00, 133.55, 132.62, 131.94, 131.75, 131.68, 130.99, 129.38, 128.96, 128.78, 128.75, 118.75 (d, $J = 21$ Hz, 1C), 115.14 (d, $J = 21$ Hz, 1C), 111.79, 52.54, 46.77, 37.01, 21.55.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -113.1 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₉H₂₄FO₅, m/z : 471.1602, observed: 471.1602.

Methyl (5*E*, 9*Z*)-7-Benzoyl-2-fluoro-9-hydroxy-10-(4-methoxy-benzoyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (41 mg, 84% yield), 0.1 mmol scale reaction.

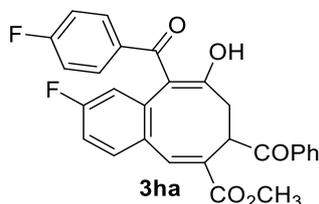
¹H NMR (500 MHz, chloroform-*d*) δ 8.25 (s, 1H), 8.11 (d, $J = 7.5$ Hz, 2H), 7.63 (t, $J = 7.5$ Hz, 1H), 7.58 – 7.50 (comp, 3H), 7.23 (d, $J = 8.5$ Hz, 2H), 7.08 (td, $J = 8.5, 2.5$ Hz, 1H), 6.75 (d, $J = 9.0$ Hz, 2H), 6.56 (dd, $J = 9.0, 2.5$ Hz, 1H), 5.26 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.82 (s, 3H), 3.73 (s, 3H), 2.90 (t, $J = 12.5$ Hz, 1H), 2.50 (dd, $J = 12.5, 4.5$ Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 198.14, 190.80, 186.52, 167.52, 162.21, 161.75 (d, $J = 254$ Hz, 1C), 141.21, 136.80 (d, $J = 8.4$ Hz, 1C), 135.01, 133.53, 131.92, 131.81, 131.74, 131.60, 131.06, 128.95, 128.77, 127.74, 118.74 (d, $J = 21$ Hz, 1C), 115.17 (d, $J = 21$ Hz, 1C), 113.40, 111.34, 55.34, 52.52, 46.80, 36.88.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₉H₂₄FO₆, m/z : 487.1551, observed: 487.1553.

Methyl (5*E*, 9*Z*)-7-Benzoyl-2-fluoro-10-(4-fluorobenzoyl)-9-hydroxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (37 mg, 78% yield), 0.1 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 8.25 (s, 1H), 8.10 (d, $J = 7.5$ Hz, 2H), 7.64 (t, $J = 7.5$ Hz, 1H), 7.55 - 7.51 (comp, 3H), 7.26 - 7.24 (comp, 2H), 7.09 (td, $J = 8.0, 2.5$ Hz, 1H), 6.95 (t, $J = 9.0$ Hz, 2H), 6.51 (dd, $J = 9.0, 2.5$ Hz, 1H), 5.27 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.74 (s, 3H), 2.91 (t, $J = 12.5$ Hz, 1H), 2.53 (dd, $J = 12.5, 4.5$ Hz, 1H).

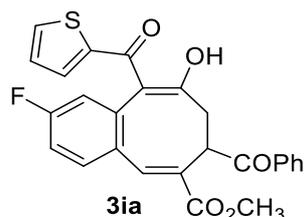
¹³C NMR (126 MHz, chloroform-*d*) δ 197.94, 191.64, 185.92, 167.44, 165.39 (d, $J = 254$ Hz, 1C),

161.75(d, $J = 254$ Hz, 1C), 141.06, 136.15 (d, $J = 8.4$ Hz, 1C), 134.93, 133.61, 131.97, 131.91, 131.78, 131.70, 131.14, 128.98, 128.77, 118.67 (d, $J = 21$ Hz, 1C), 115.41(d, $J = 21$ Hz, 1C), 115.30 (d, $J = 21$ Hz, 1C), 52.59, 46.71, 36.93.

^{19}F NMR (470.5 MHz, chloroform- d) δ -106.8 (m, 1F), -112.7 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{28}\text{H}_{21}\text{F}_2\text{O}_5$, m/z : 475.1352, observed: 475.1350.

Methyl (5E, 9Z)-7-Benzoyl-2-fluoro-9-hydroxy-10-(thiophene-2-carbonyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (40 mg, 87% yield), 0.1 mmol scale reaction.

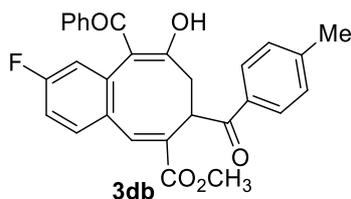
^1H NMR (500 MHz, chloroform- d) δ 8.17 (s, 1H), 8.09 (d, $J = 7.5$ Hz, 2H), 7.63 (t, $J = 7.5$ Hz, 1H), 7.60 – 7.49 (comp, 4H), 7.22 (td, $J = 8.5, 2.5$ Hz, 1H), 6.96 (d, $J = 8.5$ Hz, 3H), 5.24 (dd, $J = 12.5, 4.5$ Hz, 1H), 3.70 (s, 3H), 2.85 (t, $J = 12.5$ Hz, 1H), 2.51 (dd, $J = 12.5, 4.5$ Hz, 1H).

^{13}C NMR (126 MHz, chloroform- d) δ 198.05, 188.64, 160.93, 167.48, 161.92 (d, $J = 254$ Hz, 1C), 141.09, 139.86, 135.27 (d, $J = 8.4$ Hz, 1C), 133.97, 133.81, 133.57, 132.54, 131.97, 131.90, 130.84, 128.96, 128.77, 127.80, 119.18 (d, $J = 21$ Hz, 1C), 116.16 (d, $J = 21$ Hz, 1C), 110.47, 56.70, 46.80, 36.59.

^{19}F NMR (470.5 MHz, chloroform- d) δ -112.6 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{26}\text{H}_{20}\text{FO}_5\text{S}$, m/z : 463.1010, observed: 463.1011.

Methyl (5E, 9Z)-10-Benzoyl-2-fluoro-9-hydroxy-7-(4-methyl-benzoyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (38 mg, 81% yield), 0.1 mmol scale reaction.

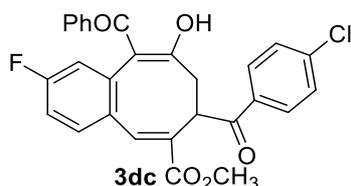
^1H NMR (500 MHz, chloroform- d) δ 8.25 (s, 1H), 8.01 (d, $J = 8.5$ Hz, 2H), 7.51 (dd, $J = 8.5, 6.0$ Hz, 1H), 7.39 – 7.36 (m, 1H), 7.33 (d, $J = 8.0$ Hz, 2H), 7.28 – 7.22 (comp, 4H), 7.05 (td, $J = 8.5, 2.5$ Hz, 1H), 6.49 (dd, $J = 9.0, 3.0$ Hz, 1H), 5.25 (dd, $J = 13.5, 4.5$ Hz, 1H), 3.73 (s, 3H), 2.92 (t, $J = 12.0$ Hz, 1H), 2.52 (dd, $J = 12.0, 4.5$ Hz, 1H), 2.45 (s, 3H).

^{13}C NMR (126 MHz, chloroform- d) δ 197.59, 192.12, 186.97, 167.50, 161.67 (d, $J = 254$ Hz, 1C), 144.53, 141.19, 136.30 (d, $J = 8.4$ Hz, 1C), 135.50, 132.38, 132.03, 132.01, 131.79, 131.72, 131.39, 131.13, 129.66, 129.20, 128.93, 128.02, 118.71 (d, $J = 21$ Hz, 1C), 115.19 (d, $J = 21$ Hz, 1C), 112.07, 52.54, 46.59, 37.20, 21.72.

^{19}F NMR (470.5 MHz, chloroform- d) δ -113.2 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{29}\text{H}_{24}\text{FO}_5$, m/z : 471.1602, observed: 471.1609.

Methyl (5*E*, 9*Z*)-10-Benzoyl-7-(4-chlorobenzoyl)-2-fluoro-9-hydroxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (33 mg, 67% yield), 0.1 mmol scale reaction.

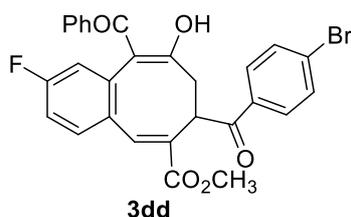
¹H NMR (500 MHz, chloroform-*d*) δ 8.26 (s, 1H), 8.05 (d, *J* = 8.5 Hz, 2H), 7.52 (d, *J* = 8.5 Hz, 3H), 7.38 (d, *J* = 8.5 Hz, 1H), 7.29 – 7.21 (comp, 5H), 7.06 (td, *J* = 8.5, 2.5 Hz, 1H), 6.50 (dd, *J* = 9.5, 2.5 Hz, 1H), 5.20 (dd, *J* = 12.5, 4.5 Hz, 1H), 3.75 (s, 3H), 2.93 (t, *J* = 12.5 Hz, 1H), 2.47 (dd, *J* = 12.5, 4.5 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 196.92, 191.76, 187.01, 167.43, 161.73 (d, *J* = 254 Hz, 1C), 141.37, 140.12, 136.29 (d, *J* = 8.4 Hz, 1C), 135.40, 133.33, 131.80, 131.73, 131.46, 130.76, 130.20, 129.30, 129.20, 128.04, 118.79 (d, *J* = 21 Hz, 1C), 115.26 (d, *J* = 21 Hz, 1C), 52.60, 46.72, 36.94.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -112.8 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺:C₂₈H₂₁ClFO₅, m/z: 491.1056, observed: 491.1063.

Methyl (5*E*, 9*Z*)-10-Benzoyl-7-(4-bromobenzoyl)-2-fluoro-9-hydroxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (38 mg, 72% yield), 0.1 mmol scale reaction.

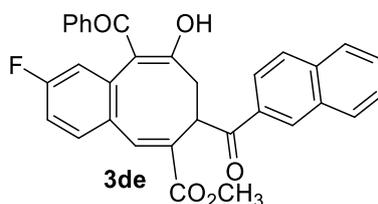
¹H NMR (500 MHz, chloroform-*d*) δ 8.26 (s, 1H), 7.97 (d, *J* = 8.5 Hz, 2H), 7.68 (d, *J* = 8.5 Hz, 2H), 7.50 (dd, *J* = 8.5, 6.0 Hz, 1H), 7.39 (t, *J* = 7.0 Hz, 1H), 7.27 – 7.22 (comp, 5H), 7.06 (td, *J* = 8.5, 3.0 Hz, 1H), 6.50 (dd, *J* = 9.0, 3.0 Hz, 1H), 5.19 (dd, *J* = 12.5, 4.5 Hz, 1H), 3.75 (s, 3H), 2.93 (t, *J* = 12.5 Hz, 1H), 2.47 (dd, *J* = 12.5, 4.5 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 197.14, 191.74, 187.02, 167.43, 161.73 (d, *J* = 254 Hz, 1C), 141.39, 136.29 (d, *J* = 8.4 Hz, 1C), 135.39, 133.74, 132.31, 131.80, 131.74, 131.48, 130.73, 130.29, 129.20, 128.90, 128.05, 118.80 (d, *J* = 21 Hz, 1C), 115.26 (d, *J* = 21 Hz, 1C), 52.62, 46.70, 36.91.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -112.8 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺:C₂₈H₂₁BrFO₅, m/z: 535.0551, observed: 535.0561.

Methyl (5*E*, 9*Z*)-7-(2-Naphthoyl)-10-benzoyl-2-fluoro-9-hydroxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (43 mg, 85% yield), 0.1 mmol scale reaction.

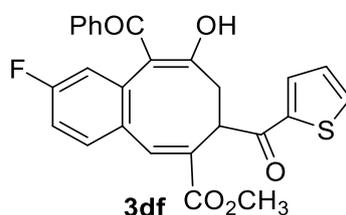
¹H NMR (500 MHz, chloroform-*d*) δ 8.29 (s, 1H), 8.27 - 8.23 (comp, 2H), 8.04 (d, J = 8.0 Hz, 1H), 7.95 - 7.86 (m, 1H), 7.61 (t, J = 8.0 Hz, 1H), 7.58 - 7.50 (comp, 3H), 7.39 (t, J = 7.0 Hz, 1H), 7.31 - 7.21 (comp, 5H), 7.11 (td, J = 8.0, 2.5 Hz, 1H), 6.51 (dd, J = 9.5, 2.5 Hz, 1H), 5.25 (dd, J = 12.5, 4.5 Hz, 1H), 3.78 (s, 3H), 2.99 (t, J = 12.5 Hz, 1H), 2.49 (dd, J = 12.5, 4.5 Hz, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.92, 192.34, 186.66, 167.60, 161.76 (d, J = 254 Hz, 1C), 141.33, 136.42 (d, J = 8.4 Hz, 1C), 135.43, 134.57, 133.90, 132.82, 132.03, 132.01, 131.76, 131.69, 131.41, 130.92, 130.63, 129.19, 128.48, 128.03, 127.91, 127.57, 126.52, 125.26, 124.67, 118.78 (d, J = 21 Hz, 1C), 115.26 (d, J = 21 Hz, 1C), 112.12, 52.61, 50.81, 36.67.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -112.9 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₃₂H₂₄FO₅, m/z : 507.1602, observed: 507.1608.

Methyl (5*E*, 9*Z*)-10-Benzoyl-2-fluoro-9-hydroxy-7-(thiophene-2-carbonyl)-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (47 mg, 51% yield), 0.2 mmol scale reaction.

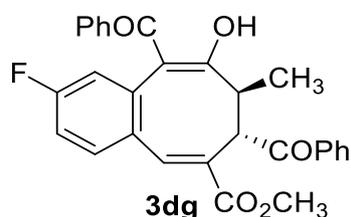
¹H NMR (500 MHz, chloroform-*d*) δ 8.17 (s, 1H), 8.08 (d, J = 8.5 Hz, 2H), 7.64 - 7.48 (comp, 5H), 7.21 (td, J = 8.5, 2.5 Hz, 1H), 7.00 - 6.86 (comp, 3H), 5.25 (dd, J = 13.5, 4.5 Hz, 1H), 3.69 (s, 3H), 2.85 (t, J = 12.5 Hz, 1H), 2.51 (dd, J = 12.0, 4.5 Hz, 1H).

¹³C NMR (126 MHz, Chloroform-*d*) δ 198.02, 188.66, 179.43, 167.47, 161.91 (d, J = 254 Hz, 1C), 141.07, 139.85, 135.27 (d, J = 8.4 Hz, 1C), 134.98, 133.97, 133.84, 133.57, 132.52, 131.98, 131.91, 130.86, 128.96, 128.77, 127.81, 119.18 (d, J = 21 Hz, 1C), 116.15 (d, J = 21 Hz, 1C), 110.47, 52.52, 46.81, 36.58.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₆H₂₀FO₅S, m/z : 463.1010, observed: 463.1021.

Methyl (5*E*, 7*S*, 8*S*, 9*Z*)-7,10-Dibenzoyl-2-fluoro-9-hydroxy-8-methyl-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (21 mg, 45% yield, >19:1 dr, 97% ee), 0.1 mmol scale reaction.

Optical purity determined by HPLC analysis [Daicel chiralpak ADH, *n*-hexane/*i*-PrOH = 80/20, 1.0 mL/min, λ = 254 nm, t_1 = 4.67 min, t_2 = 5.44 min].

¹H NMR (500 MHz, chloroform-*d*) δ 8.11 (d, J = 7.5 Hz, 2H), 8.08 (s, 1H), 7.59 (t, J = 7.5 Hz, 1H), 7.50 (t, J = 7.5 Hz, 2H), 7.42 (dd, J = 8.5, 6.0 Hz, 1H), 7.37 - 7.32 (m, 1H), 7.25 - 7.17 (comp, 4H),

7.07 (td, $J = 8.5, 2.5$ Hz, 1H), 6.50 (dd, $J = 9.5, 2.5$ Hz, 1H), 5.02 (d, $J = 11.5$ Hz, 1H), 3.67 (s, 3H), 3.46 (dq, $J = 11.5, 6.5$ Hz, 1H), 0.95 (d, $J = 6.5$ Hz, 3H).

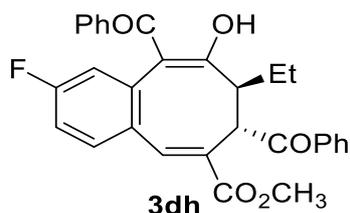
^{13}C NMR (126 MHz, chloroform- d) δ 199.32, 197.86, 184.12, 167.35, 161.93(d, $J = 254$ Hz, 1C), 141.75, 137.41, 136.08 (d, $J = 8.4$ Hz, 1C), 135.32, 133.53, 133.03, 131.99, 131.13, 130.75, 130.68, 129.13, 129.10, 128.70, 127.98, 118.44 (d, $J = 21$ Hz, 1C), 115.18 (d, $J = 21$ Hz, 1C), 52.45, 51.26, 41.30, 15.52.

^{19}F NMR (470.5 MHz, chloroform- d) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{29}\text{H}_{24}\text{FO}_5$, m/z : 471.1602, observed: 471.1616.

$[\alpha]_{\text{D}}^{23} = +146.3$ (c 0.61, CH_2Cl_2).

Methyl (5*E*, 7*S*, 8*S*, 9*Z*)-7,10-Dibenzoyl-8-ethyl-2-fluoro-9-hydroxy-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (17 mg, 35% yield, >19:1 dr, 90% ee), 0.1 mmol scale reaction.

Optical purity determined by HPLC analysis [Daicel chiralpak ADH, n -hexane/ i -PrOH = 80/20, 1.0 mL/min, $\lambda = 254$ nm, $t_1 = 4.93$ min, $t_2 = 6.90$ min].

^1H NMR (500 MHz, chloroform- d) δ 8.13 (d, $J = 7.5$ Hz, 2H), 8.10 (s, 1H), 7.62 (t, $J = 7.5$ Hz, 1H), 7.52 (t, $J = 7.5$ Hz, 2H), 7.45 (dd, $J = 8.5, 6.0$ Hz, 1H), 7.38 (t, $J = 7.3$ Hz, 1H), 7.28 – 7.19 (comp, 4H), 7.10 (td, $J = 8.5, 2.5$ Hz, 1H), 6.53 (dd, $J = 9.5, 2.5$ Hz, 1H), 5.07 (d, $J = 11.5$ Hz, 1H), 3.68 (s, 3H), 3.35 (td, $J = 11.5, 3.0$ Hz, 1H), 1.85 – 1.77 (m, 1H), 1.08 – 1.03 (m, 1H), 0.67 (t, $J = 7.5$ Hz, 3H).

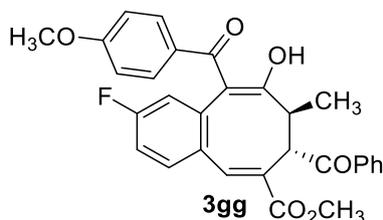
^{13}C NMR (126 MHz, chloroform- d) δ 199.36, 196.69, 184.44, 167.35, 162.02 (d, $J = 254$ Hz, 1C), 141.73, 137.30, 136.25 (d, $J = 8.4$ Hz, 1C), 135.38, 133.53, 133.07, 132.03, 131.14, 130.77, 130.70, 129.08, 128.70, 127.96, 118.15 (d, $J = 21$ Hz, 1C), 115.18 (d, $J = 21$ Hz, 1C), 52.43, 50.16, 48.74, 23.66, 12.09.

^{19}F NMR (470.5 MHz, chloroform- d) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{30}\text{H}_{26}\text{FO}_5$, m/z : 485.1759, observed: 485.1775.

$[\alpha]_{\text{D}}^{23} = +327.7$ (c 0.14, CH_2Cl_2).

Methyl (5*E*, 7*S*, 8*S*, 9*Z*)-7-Benzoyl-2-fluoro-9-hydroxy-10-(4-methoxybenzoyl)-8-methyl-7,8-dihydrobenzo[8]annulene-6-carboxylate.



Colorless oil (25 mg, 50% yield, >19:1 dr, 97% ee), 0.1 mmol scale reaction.

Optical purity determined by HPLC analysis [Daicel chiralpak ADH, n -hexane/ i -PrOH = 95/5, 1.0 mL/min, $\lambda = 254$ nm, $t_1 = 13.59$ min, $t_2 = 17.50$ min].

^1H NMR (500 MHz, chloroform- d) δ 8.13 (d, $J = 8.5$ Hz, 2H), 8.09 (s, 1H), 7.61 (t, $J = 7.5$ Hz, 1H),

7.52 (t, $J = 7.5$ Hz, 2H), 7.46 (dd, $J = 8.5, 6.0$ Hz, 1H), 7.20 – 7.18 (comp, 2H), 7.11 (td, $J = 8.5, 2.5$ Hz, 1H), 6.76 – 6.73 (comp, 2H), 6.60 (dd, $J = 9.5, 2.5$ Hz, 1H), 5.04 (d, $J = 11.5$ Hz, 1H), 3.82 (s, 3H), 3.68 (s, 3H), 3.50 – 3.39 (m, 1H), 0.96 (d, $J = 6.5$ Hz, 3H).

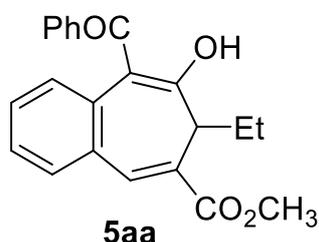
^{13}C NMR (126 MHz, chloroform- d) δ 199.50, 196.55, 183.96, 167.39, 161.99 (d, $J = 254$ Hz, 1C), 161.96, 141.69, 137.46, 136.53 (d, $J = 8.4$ Hz, 1C), 133.49, 133.00, 132.06, 131.41, 130.79, 130.72, 129.12, 128.67, 127.62, 118.42 (d, $J = 21$ Hz, 1C), 115.14 (d, $J = 21$ Hz, 1C), 113.36, 110.99, 55.32, 52.40, 51.25, 41.07, 15.53.

^{19}F NMR (470.5 MHz, chloroform- d) δ -112.9 (m, 1F).

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{30}\text{H}_{26}\text{FO}_6$, m/z : 501.1708, observed: 501.1686.

$[\alpha]_{\text{D}}^{23} = +384.0$ (c 0.30, CH_2Cl_2).

Methyl 9-Benzoyl-7-ethyl-8-hydroxy-7H-benzo-[7]annulene-6-carboxylate.



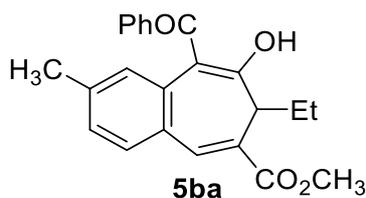
Pale yellow oil (49.5 mg, 71% yield), 0.2 mmol scale reaction.

^1H NMR (300 MHz, chloroform- d) δ 7.99 (s, 1H), 7.42 (d, $J = 7.5$ Hz, 1H), 7.35 – 7.29 (m, 1H), 7.22 – 7.11 (comp, 5H), 6.92 (t, $J = 7.0$ Hz, 1H), 6.78 (d, $J = 7.8$ Hz, 1H), 4.13 – 4.03 (m, 1H), 3.89 (s, 3H), 1.32 – 1.13 (m, 2H), 0.82 (t, $J = 7.5$ Hz, 3H).

^{13}C NMR (75 MHz, Chloroform- d) δ 196.3, 189.0, 166.9, 138.8, 133.2, 132.0, 130.7, 129.7, 129.6, 127.8, 127.7, 126.1, 52.5, 49.9, 17.7, 12.2.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{22}\text{H}_{21}\text{O}_4$, m/z : 349.1434, observed: 349.1437.

Methyl 9-Benzoyl-7-ethyl-8-hydroxy-2-methyl-7H-benzo[7]-annulene-6-carboxylate.



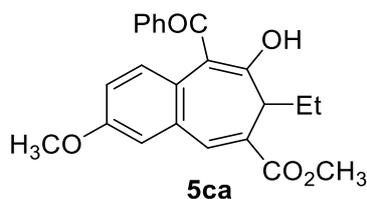
Pale yellow oil (58 mg, 80% yield), 0.2 mmol scale reaction.

^1H NMR (500 MHz, chloroform- d) δ 7.95 (s, 1H), 7.33 – 7.28 (comp, 2H), 7.18 (t, $J = 7.5$ Hz, 2H), 7.12 (d, $J = 7.5$ Hz, 2H), 6.99 (d, $J = 7.5$ Hz, 1H), 6.56 (s, 1H), 4.13 – 4.03 (m, 1H), 3.88 (s, 3H), 1.98 (s, 3H), 1.33 – 1.28 (m, 1H), 1.20 – 1.17 (m, 1H), 0.82 (t, $J = 7.5$ Hz, 3H).

^{13}C NMR (126 MHz, chloroform- d) δ 195.9, 189.5, 167.0, 138.9, 137.8, 134.0, 130.8, 130.5, 129.8, 129.4, 127.7, 127.0, 52.4, 49.9, 21.0, 17.7, 12.2.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{23}\text{H}_{23}\text{O}_4$, m/z : 363.1591, observed: 363.1594.

Methyl 9-Benzoyl-7-ethyl-8-hydroxy-3-methoxy-7H-benzo[7]annulene-6-carboxylate.



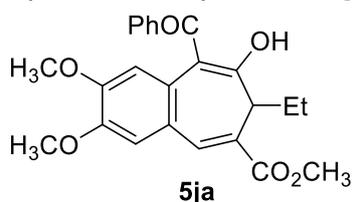
Pale yellow oil (60 mg, 79% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.93 (s, 1H), 7.33 – 7.29 (m, 1H), 7.20 (t, $J = 7.5$ Hz, 2H), 7.15 (d, $J = 7.5$ Hz, 2H), 6.92 (s, 1H), 6.68 (d, $J = 8.6$ Hz, 1H), 6.51 (dd, $J = 8.5, 3.0$ Hz, 1H), 4.09 – 4.06 (m, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 1.32 – 1.27 (m, 1H), 1.25 – 1.17 (m, 1H), 0.82 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 196.6, 188.0, 166.9, 157.5, 138.7, 134.4, 132.2, 130.5, 129.6, 127.8, 114.6, 113.5, 55.3, 52.5, 50.2, 20.9, 12.3.

HRMS (ESI) scaled for $[M+H]^+$: C₂₃H₂₃O₅, m/z : 379.1540, observed: 379.1540.

Methyl 9-Benzoyl-7-ethyl-8-hydroxy-2,3-dimethoxy-7H-benzo[7]annulene-6-carboxylate.



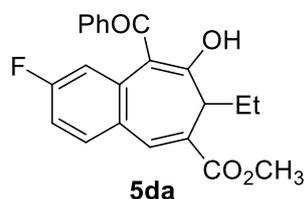
Pale yellow oil (66 mg, 81% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.90 (s, 1H), 7.32 (t, $J = 7.5$ Hz, 1H), 7.22 (t, $J = 7.5$ Hz, 2H), 7.16 (d, $J = 7.5$ Hz, 2H), 6.86 (s, 1H), 6.17 (s, 1H), 4.11 – 4.06 (m, 1H), 3.91 (s, 3H), 3.88 (s, 3H), 3.26 (s, 3H), 1.36 – 1.29 (m, 1H), 1.23 (s, 1H), 0.83 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 195.5, 189.5, 167.0, 148.1, 138.6, 130.5, 129.7, 129.2, 128.0, 116.37, 111.6, 55.8, 55.4, 49.8, 20.5, 17.9, 12.2.

HRMS (ESI) scaled for $[M+H]^+$: C₂₄H₂₅O₆, m/z : 409.1646, observed: 409.1645.

Methyl 9-Benzoyl-7-ethyl-2-fluoro-8-hydroxy-7H-benzo[7]annulene-6-carboxylate.



Pale yellow oil (57 mg, 78% yield), 0.2 mmol scale reaction.

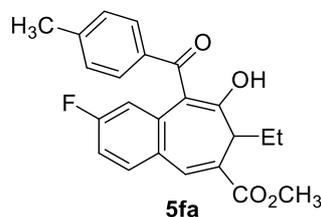
¹H NMR (500 MHz, chloroform-*d*) δ 7.94 (s, 1H), 7.39 (t, $J = 7.5$ Hz, 1H), 7.35 (d, $J = 7.5$ Hz, 1H), 7.23 (t, $J = 7.5$ Hz, 2H), 7.15 (d, $J = 7.5$ Hz, 2H), 6.89 (t, $J = 7.5$ Hz, 1H), 6.47 (d, $J = 10.0$ Hz, 1H), 4.15 – 4.03 (m, 1H), 3.89 (s, 3H), 1.30 – 1.27 (m, 1H), 1.23 – 1.16 (m, 1H), 0.83 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 195.89, 191.57, 166.76, 161.32 (d, $J = 254$ Hz, 1C), 137.73, 131.74, 131.65, 131.09, 129.40, 128.03, 119.57 (d, $J = 22$ Hz, 1C), 113.61 (d, $J = 22$ Hz, 1C), 52.54, 49.73, 20.89, 12.18.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -111.7 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺:C₂₂H₂₀FO₄, m/z: 367.1340, observed: 367.1339.

Methyl 7-Ethyl-2-fluoro-8-hydroxy-9-(4-methylbenzoyl)-7H-benzo[7]annulene-6-carboxylate.



Pale yellow oil (57 mg, 75% yield), 0.2 mmol scale reaction.

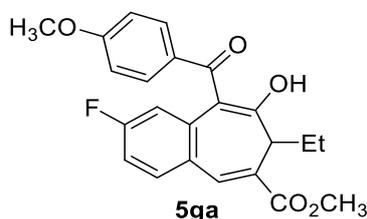
¹H NMR (500 MHz, chloroform-*d*) δ 7.94 (s, 1H), 7.43 – 7.36 (m, 1H), 7.03 (comp, 4H), 6.90 (t, *J* = 8.0 Hz, 1H), 6.51 (d, *J* = 11.5 Hz, 1H), 4.13 – 4.02 (m, 1H), 3.88 (s, 3H), 2.33 (s, 3H), 1.30 – 1.24 (m, 1H), 1.23 – 1.16 (m, 1H), 0.83 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 195.43, 189.85, 166.78, 161.31 (d, *J* = 250 Hz, 1C), 141.76, 137.72, 131.71, 131.68, 131.61, 129.56, 128.73, 119.57 (d, *J* = 22 Hz, 1C), 113.51 (d, *J* = 22 Hz, 1C), 52.51, 49.65, 21.50, 20.90, 12.18.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -111.8 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺:C₂₃H₂₂FO₄, m/z: 381.1497, observed: 381.1499.

Methyl 7-Ethyl-2-fluoro-8-hydroxy-9-(4-methoxybenzoyl)-7H-benzo[7]annulene-6-carboxylate.



Pale yellow oil (61 mg, 78% yield), 0.2 mmol scale reaction.

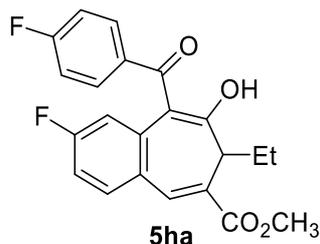
¹H NMR (500 MHz, chloroform-*d*) δ 7.93 (s, 1H), 7.44 – 7.38 (m, 1H), 7.12 (d, *J* = 8.5 Hz, 2H), 6.91 (t, *J* = 8.5 Hz, 1H), 6.72 (d, *J* = 9.0 Hz, 2H), 6.55 (d, *J* = 10.0 Hz, 1H), 4.11 – 4.01 (m, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 1.27 – 1.22 (m, 1H), 1.21 – 1.15 (m, 1H), 0.82 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 194.66, 189.52, 166.78, 161.37 (d, *J* = 250 Hz, 1C), 137.63, 131.88, 131.69, 119.51 (d, *J* = 22 Hz, 1C), 113.50 (d, *J* = 22 Hz, 1C), 113.36, 55.31, 52.50, 49.40, 20.90, 12.17.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -111.7 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺:C₂₃H₂₂FO₅, m/z: 397.1446, observed: 397.1449.

Methyl 7-Ethyl-2-fluoro-9-(4-fluorobenzoyl)-8-hydroxy-7H-benzo[7]annulene-6-carboxylate.



Pale yellow oil (46 mg, 60% yield), 0.2 mmol scale reaction.

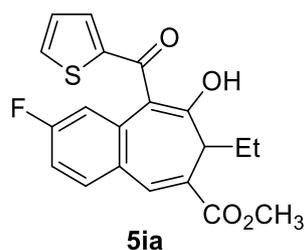
¹H NMR (500 MHz, chloroform-*d*) δ 7.93 (s, 1H), 7.44 – 7.37 (m, 1H), 7.20 – 7.12 (comp, 2H), 6.95 – 6.88 (comp, 3H), 6.48 (d, $J = 9.5$ Hz, 1H), 4.15 – 4.02 (m, 1H), 3.88 (s, 3H), 1.29 – 1.23 (m, 1H), 1.22 – 1.15 (m, 1H), 0.82 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 195.52, 188.64, 166.68, 164.17 (d, $J = 254$ Hz, 1C), 161.36 (d, $J = 254$ Hz, 1C), 137.58, 131.92, 131.84, 119.48 (d, $J = 22$ Hz, 1C), 115.26 (d, $J = 22$ Hz, 1C), 113.78 (d, $J = 22$ Hz, 1C), 52.55, 49.54, 20.92, 12.15.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -107.2 (m, 1F), -111.5 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₂H₁₉F₂O₄, m/z : 385.1246, observed: 385.1249.

Methyl 7-Ethyl-2-fluoro-8-hydroxy-9-(thiophene-2-carbonyl)-7H-benzo[7]annulene-6-carboxylate.



Pale yellow oil (33.5 mg, 45% yield), 0.2 mmol scale reaction.

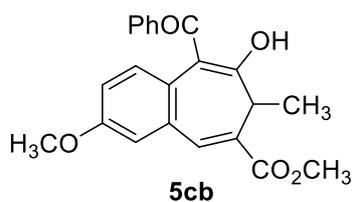
¹H NMR (500 MHz, chloroform-*d*) δ 7.92 (s, 1H), 7.49 (d, $J = 5.0$ Hz, 1H), 7.48 – 7.44 (m, 1H), 7.03 (t, $J = 8.5$ Hz, 1H), 6.95 (d, $J = 8.5$ Hz, 1H), 6.90 – 6.85 (m, 1H), 6.76 (d, $J = 5.0$ Hz, 1H), 4.06 – 3.99 (m, 1H), 3.87 (s, 3H), 1.28 – 1.22 (m, 1H), 1.17 – 1.12 (m, 1H), 0.82 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 192.19, 182.51, 166.69, 161.36 (d, $J = 250$ Hz, 1C), 137.43, 133.51, 132.81, 132.43, 131.68 (d, $J = 9$ Hz, 1C), 127.42, 119.70 (d, $J = 21$ Hz, 1C), 114.32 (d, $J = 21$ Hz, 1C), 109.61, 52.52, 48.69, 20.87, 12.16.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -111.5 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₀H₁₈FO₄S, m/z : 373.0904, observed: 373.0907.

Methyl 9-Benzoyl-8-hydroxy-3-methoxy-7-methyl-7H-benzo[7]annulene-6-carboxylate.



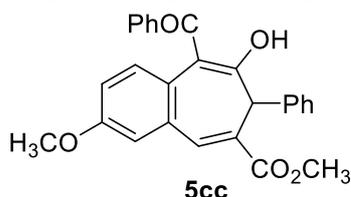
Pale yellow oil (59 mg, 81% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.88 (s, 1H), 7.31 (t, *J* = 7.5 Hz, 1H), 7.20 (t, *J* = 7.5 Hz, 2H), 7.16 (d, *J* = 7.5 Hz, 2H), 6.93 (d, *J* = 2.5 Hz, 1H), 6.70 (d, *J* = 9.0 Hz, 1H), 6.53 (dd, *J* = 9.0, 2.5 Hz, 1H), 4.35 – 4.15 (m, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 1.03 – 0.68 (m, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 191.10, 188.02, 166.68, 157.57, 134.47, 130.54, 129.61, 127.84, 114.70, 55.28, 52.42.

HRMS (ESI) scaled for [M+H]⁺:C₂₂H₂₁O₅, m/z: 365.1384, observed: 365.1388.

Methyl 9-Benzoyl-8-hydroxy-3-methoxy-7-phenyl-7H-benzo[7]-annulene-6-carboxylate.



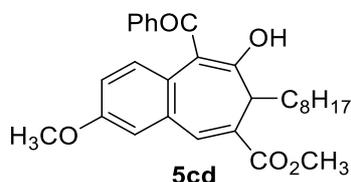
Pale yellow oil (58.8 mg, 69% yield), 0.2 mmol scale reaction.

¹H NMR (300 MHz, chloroform-*d*) δ 8.19 (s, 1H), 7.32 – 7.28 (m, 1H), 7.23 – 7.10 (comp, 4H), 7.04 – 6.89 (comp, 5H), 6.81 (d, *J* = 2.5 Hz, 1H), 6.38 (d, *J* = 8.5 Hz, 1H), 6.25 (dd, *J* = 8.5, 2.5 Hz, 1H), 3.94 (s, 3H), 3.71 (s, 3H).

¹³C NMR (75 MHz, chloroform-*d*) δ 196.5, 186.7, 166.9, 157.4, 140.9, 136.4, 135.2, 134.4, 134.2, 130.7, 130.6, 129.6, 127.9, 127.8, 126.7, 126.5, 126.2, 114.5, 112.9, 55.2, 52.8.

HRMS (ESI) scaled for [M+H]⁺:C₂₇H₂₃O₅, m/z: 427.1540, observed: 427.1545.

Methyl 9-Benzoyl-8-hydroxy-3-methoxy-7-octyl-7H-benzo[7]-annulene-6-carboxylate.



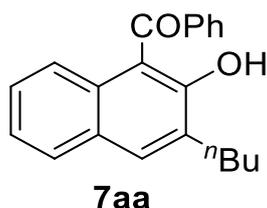
Pale yellow oil (58 mg, 63% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.93 (s, 1H), 7.31 (t, *J* = 7.5 Hz, 1H), 7.23 – 7.12 (comp, 4H), 6.92 (s, 1H), 6.67 (d, *J* = 8.5 Hz, 1H), 6.51 (d, *J* = 11.0 Hz, 1H), 4.17 – 4.11 (m, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 1.26 – 1.11 (m, 14H), 0.86 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 196.73, 187.81, 166.90, 157.52, 138.72, 137.04, 134.51, 134.41, 132.42, 130.52, 129.58, 127.81, 126.78, 114.56, 113.47, 110.75, 55.25, 52.51, 48.69, 31.79, 29.14, 29.09, 29.03, 27.68, 27.47, 22.60, 14.08.

HRMS (ESI) scaled for [M+H]⁺:C₂₉H₃₅O₅, m/z: 463.2479, observed: 463.2488.

(3-Butyl-2-hydroxynaphthalen-1-yl)(phenyl)methanone.



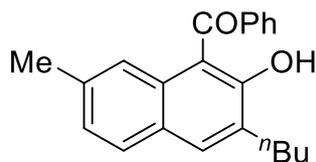
Yellow oil (52 mg, 86% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.84 (s, 1H), 7.81 (s, 1H), 7.72 (d, $J = 7.5$ Hz, 1H), 7.69 – 7.63 (m, 2H), 7.61 – 7.54 (m, 1H), 7.43 (t, $J = 8.0$ Hz, 2H), 7.27 (t, $J = 7.5$ Hz, 2H), 7.15 – 7.09 (m, 1H), 2.94 – 2.83 (m, 2H), 1.85 – 1.73 (m, 2H), 1.55 – 1.47 (m, 2H), 1.04 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.9, 161.2, 140.7, 135.0, 132.5, 129.4, 128.5, 128.2, 128.0, 126.1, 125.7, 123.6, 113.7, 31.5, 30.1, 22.7, 14.1.

HRMS (ESI) scaled for $[M+H]^+$: C₂₁H₂₁O₂, m/z : 305.1536, observed: 305.1538.

(3-Butyl-2-hydroxy-7-methylnaphthalen-1-yl)(phenyl)methanone.



7ba

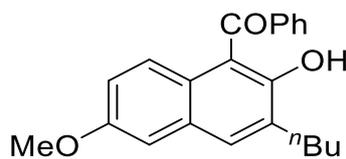
Yellow oil (60 mg, 94% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.92 (s, 1H), 7.76 (s, 1H), 7.65 (d, $J = 7.5$ Hz, 2H), 7.61 (d, $J = 8.0$ Hz, 1H), 7.58 (t, $J = 7.5$ Hz, 1H), 7.43 (t, $J = 7.5$ Hz, 2H), 7.10 (d, $J = 8.0$ Hz, 1H), 7.01 (s, 1H), 2.91 – 2.82 (m, 2H), 2.16 (s, 3H), 1.83 – 1.73 (m, 2H), 1.54 – 1.46 (h, $J = 7.5$ Hz, 2H), 1.03 (t, $J = 7.4$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 201.0, 161.4, 140.7, 135.4, 134.9, 132.3, 131.5, 131.3, 129.4, 128.4, 127.8, 126.3, 125.8, 125.5, 113.3, 31.5, 30.0, 22.7, 21.9, 14.1.

HRMS (ESI) scaled for $[M+H]^+$: C₂₂H₂₃O₂, m/z : 319.1693, observed: 319.1706.

(3-Butyl-2-hydroxy-6-methoxynaphthalen-1-yl)(phenyl)methanone.



7ca

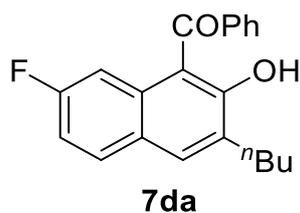
Yellow oil (64 mg, 96% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.53 (s, 1H), 7.71 (s, 1H), 7.64 (d, $J = 9.0$ Hz, 2H), 7.57 (t, $J = 8.0$ Hz, 1H), 7.42 (t, $J = 8.0$ Hz, 2H), 7.16 (d, $J = 9.0$ Hz, 1H), 7.08 (d, $J = 3.0$ Hz, 1H), 6.79 (dd, $J = 9.0, 2.5$ Hz, 1H), 3.89 (s, 3H), 2.91 – 2.82 (m, 2H), 1.81 – 1.70 (m, 2H), 1.51 – 1.47 (m, 2H), 1.02 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.8, 159.3, 155.7, 140.7, 133.9, 132.9, 132.5, 129.4, 129.2, 128.6, 127.5, 126.1, 117.4, 114.0, 106.8, 55.2, 31.5, 30.1, 22.7, 14.1.

HRMS (ESI) scaled for $[M+H]^+$: C₂₂H₂₃O₃, m/z : 335.1642, observed: 335.1629.

(3-Butyl-7-fluoro-2-hydroxynaphthalen-1-yl)(phenyl)methanone.



Yellow oil (48.3 mg, 75% yield), 0.2 mmol scale reaction.

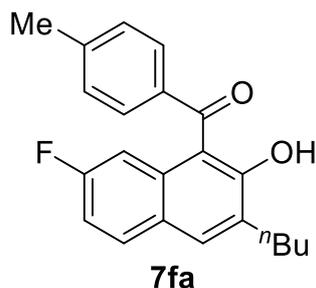
¹H NMR (500 MHz, chloroform-*d*) δ 12.00 (s, 1H), 7.77 (s, 1H), 7.69 (dd, *J* = 9.0, 6.0 Hz, 1H), 7.66 – 7.62 (m, 2H), 7.60 (t, *J* = 7.5 Hz, 1H), 7.45 (t, *J* = 7.5 Hz, 2H), 7.03 (td, *J* = 7.5, 2.5 Hz, 1H), 6.85 (dd, *J* = 12.0, 2.5 Hz, 1H), 2.92 – 2.80 (m, 2H), 1.78 – 1.72 (m, 2H), 1.51 – 1.48 (m, 2H), 1.02 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.67, 162.26, 160.39 (d, *J* = 245 Hz, 1C), 140.12, 134.80, 132.76, 132.51 (d, *J* = 9 Hz, 1C), 131.79, 130.23, 130.16, 129.30, 128.70, 125.05, 113.31 (d, *J* = 21 Hz, 1C), 110.51 (d, *J* = 21 Hz, 1C), 31.43, 29.90, 22.68, 14.03.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -112.6 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺: C₂₁H₂₀FO₂, m/z: 323.1442, observed: 323.1441.

(3-Butyl-7-fluoro-2-hydroxynaphthalen-1-yl)(p-tolyl)methanone.



Yellow oil (53 mg, 79% yield), 0.2 mmol scale reaction.

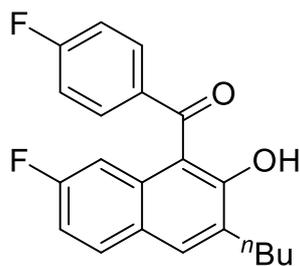
¹H NMR (500 MHz, chloroform-*d*) δ 11.75 (s, 1H), 7.76 (s, 1H), 7.69 (dd, *J* = 9.0, 6.0 Hz, 1H), 7.55 (d, *J* = 8.0 Hz, 2H), 7.24 (d, *J* = 8.0 Hz, 2H), 7.03 (td, *J* = 8.5, 2.5 Hz, 1H), 6.92 (dd, *J* = 12.0, 2.5 Hz, 1H), 2.90 – 2.80 (m, 2H), 2.46 (s, 3H), 1.80 – 1.70 (m, 2H), 1.52 – 1.44 (m, 2H), 1.01 (t, *J* = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.28, 161.63, 160.46 (d, *J* = 245 Hz, 1C), 143.77, 137.30, 134.39, 132.53 (d, *J* = 9 Hz, 1C), 131.72, 130.14, 130.06, 129.61, 129.37, 125.04, 113.26 (d, *J* = 21 Hz, 1C), 110.49 (d, *J* = 21 Hz, 1C), 31.44, 29.92, 22.67, 21.75, 14.02.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -111.8 (m, 1F).

HRMS (ESI) scaled for [M+H]⁺: C₂₂H₂₂FO₂, m/z: 337.1598, observed: 337.1613.

(3-Butyl-7-fluoro-2-hydroxynaphthalen-1-yl)(4-fluorophenyl)-methanone.



7ha

Yellow oil (56 mg, 83% yield), 0.2 mmol scale reaction.

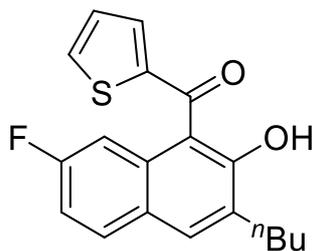
¹H NMR (500 MHz, chloroform-*d*) δ 11.68 (s, 1H), 7.77 (s, 1H), 7.72 – 7.65 (m, 3H), 7.13 (t, J = 8.5 Hz, 2H), 7.05 (ddd, J = 12.0, 8.5, 2.5 Hz, 1H), 6.87 (dd, J = 12.0, 2.5 Hz, 1H), 2.89 – 2.78 (comp, 2H), 1.77 – 1.71 (m, 2H), 1.52 – 1.44 (m, 2H), 1.01 (t, J = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 198.87, 165.52 (d, J = 255 Hz, 1C), 161.87, 160.56 (d, J = 245 Hz, 1C), 136.11, 134.76, 132.35, 132.28, 132.16, 132.09, 131.80, 130.32 (d, J = 9 Hz, 1C), 125.07, 115.92 (d, J = 21 Hz, 1C), 113.44 (d, J = 21 Hz, 1C), 110.34 (d, J = 21 Hz, 1C), 31.42, 29.89, 22.66, 14.01.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -104.9 (m, 1F), -112.3 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₂₁H₁₉F₂O₂, m/z : 341.1348, observed: 341.1363.

(3-Butyl-7-fluoro-2-hydroxynaphthalen-1-yl)(thiophen-2-yl)methanone.



7ia

Yellow oil (40.6 mg, 62% yield), 0.2 mmol scale reaction.

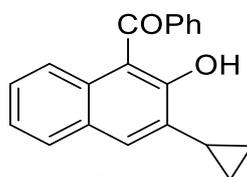
¹H NMR (500 MHz, chloroform-*d*) δ 10.41 (s, 1H), 7.76 (dd, J = 5.0, 1.0 Hz, 1H), 7.74 (s, 1H), 7.71 (dd, J = 9.0, 6.0 Hz, 1H), 7.41 (dd, J = 4.0, 1.0 Hz, 1H), 7.37 (dd, J = 12.0, 2.5 Hz, 1H), 7.10 – 7.06 (comp, 2H), 2.88 – 2.79 (m, 2H), 1.80 – 1.67 (m, 2H), 1.50 – 1.43 (m, 2H), 1.00 (t, J = 7.5 Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 190.81, 160.40 (d, J = 245 Hz, 1C), 159.16, 144.51, 135.59, 134.91, 133.73, 132.0 (d, J = 9 Hz, 1C), 131.59, 130.00 (d, J = 9 Hz, 1C), 128.16, 125.28, 113.64 (d, J = 21 Hz, 1C), 110.15 (d, J = 21 Hz, 1C), 31.46, 29.99, 22.65, 14.01.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -110.5 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₁₉H₁₈FO₂S, m/z : 329.1006, observed: 329.1021.

(3-Cyclopropyl-2-hydroxynaphthalen-1-yl)(phenyl)methanone.



7ab

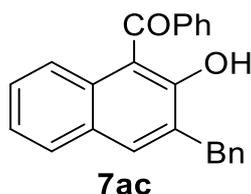
Yellow oil (46 mg, 80% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.48 (s, 1H), 7.70 – 7.66 (comp, 3H), 7.61 – 7.55 (comp, 2H), 7.43 (t, *J* = 8.0 Hz, 2H), 7.31 – 7.23 (comp, 2H), 7.12 (ddd, *J* = 8.4, 6.9, 1.4 Hz, 1H), 2.38 – 2.33 (m, 1H), 1.15 – 1.08 (m, 2H), 0.89 – 0.84 (m, 2H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.7, 160.8, 140.4, 133.1, 132.6, 131.0, 130.7, 129.5, 128.5, 128.2, 128.0, 125.9, 125.8, 123.7, 114.0, 10.0, 7.5.

HRMS (ESI) scaled for [M+H]⁺: C₂₀H₁₇O₂, *m/z*: 289.1223, observed: 289.1237.

(3-benzyl-2-hydroxynaphthalen-1-yl)(phenyl)methanone.



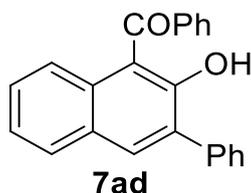
Yellow solid (57 mg, 85% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.79 (s, 1H), 7.70 (s, 1H), 7.68 (d, *J* = 8.0 Hz, 1H), 7.66 – 7.62 (comp., 2H), 7.58 (t, *J* = 7.5 Hz, 1H), 7.43 (t, *J* = 7.5 Hz, 2H), 7.39 – 7.34 (comp., 4H), 7.26 (dd, *J* = 15.0, 8.0 Hz, 3H), 7.12 (t, *J* = 7.5 Hz, 1H), 4.23 (s, 2H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.8, 160.7, 140.5, 139.7, 135.7, 132.6, 131.5, 131.1, 129.4, 129.2, 128.5, 128.52, 128.2, 128.1, 126.3, 126.1, 126.0, 123.7, 36.1.

HRMS (ESI) scaled for [M+H]⁺: C₂₄H₁₉O₂, *m/z*: 339.1380, observed: 339.1387.

(2-Hydroxy-3-phenylnaphthalen-1-yl)(phenyl)methanone.



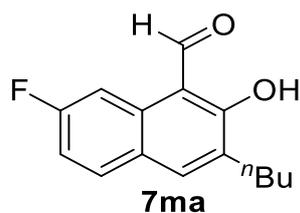
Yellow oil (22 mg, 35% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 11.11 (s, 1H), 8.00 (s, 1H), 7.82 (d, *J* = 8.0 Hz, 1H), 7.76 – 7.70 (comp, 4H), 7.61 (t, *J* = 7.5 Hz, 1H), 7.55 – 7.51 (comp, 2H), 7.46 (td, *J* = 7.5, 1.5 Hz, 3H), 7.38 – 7.31 (m, 2H), 7.23 – 7.20 (m, 1H).

¹³C NMR (126 MHz, chloroform-*d*) δ 200.5, 158.3, 140.2, 136.8, 135.8, 132.8, 132.0, 131.6, 129.6, 128.6, 128.5, 128.3, 128.0, 126.7, 125.9, 124.0, 115.2.

HRMS (ESI) scaled for [M+H]⁺: C₂₃H₁₇O₂, *m/z*: 325.1223, observed: 325.1234.

3-butyl-7-fluoro-2-hydroxy-1-naphthaldehyde.



White solid (31 mg, 61% yield), 0.2 mmol scale reaction.

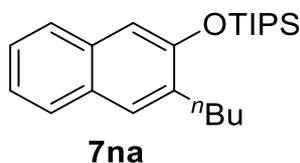
¹H NMR (500 MHz, chloroform-*d*) δ 13.61 (s, 1H), 10.64 (s, 1H), 7.89 (dd, $J = 11.0, 2.5$ Hz, 1H), 7.78 (s, 1H), 7.73 (dd, $J = 9.0, 6.0$ Hz, 1H), 7.17 (td, $J = 8.0, 2.5$ Hz, 1H), 2.83 – 2.69 (m, 2H), 1.76 – 1.65 (m, 2H), 1.48 – 1.41 (m, 2H), 0.99 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 192.79, 165.20, 162.40 (d, $J = 245$ Hz, 1C), 137.16, 133.2 (d, $J = 9$ Hz, 1C), 131.74, 131.19, 131.11, 124.52, 113.90 (d, $J = 21$ Hz, 1C), 110.60, 103.12 (d, $J = 21$ Hz, 1C), 31.25, 29.17, 22.58, 13.96.

¹⁹F NMR (470.5 MHz, chloroform-*d*) δ -113.0 (m, 1F).

HRMS (ESI) scaled for $[M+H]^+$: C₁₅H₁₆FO₂, m/z : 247.1129, observed: 247.1133.

((3-butyl-naphthalen-2-yl)oxy)triisopropylsilane.

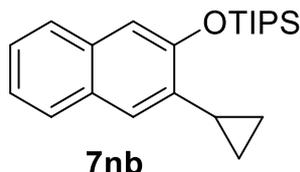


Colorless oil (65 mg, 91% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.76 (dd, $J = 8.0, 1.0$ Hz, 1H), 7.69 (dd, $J = 8.0, 1.0$ Hz, 1H), 7.63 (s, 1H), 7.40 (ddd, $J = 8.0, 7.0, 1.0$ Hz, 1H), 7.34 (ddd, $J = 8.0, 7.0, 1.0$ Hz, 1H), 7.17 (s, 1H), 2.89 – 2.81 (m, 2H), 1.78 – 1.69 (m, 2H), 1.51 – 1.42 (m, 5H), 1.22 (d, $J = 7.5$ Hz, 18H), 1.02 (t, $J = 7.5$ Hz, 3H).

¹³C NMR (126 MHz, chloroform-*d*) δ 153.0, 135.0, 133.3, 129.1, 128.2, 127.0, 126.1, 123.4, 112.4, 32.3, 31.2, 22.8, 18.2, 13.2. The spectra is consist with previous report.⁴

((3-cyclopropyl-naphthalen-2-yl)oxy)triisopropylsilane.

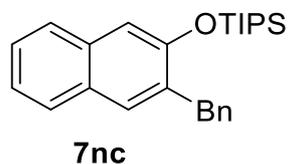


Colorless oil (62.5 mg, 92% yield), 0.2 mmol scale reaction.

¹H NMR (500 MHz, chloroform-*d*) δ 7.72 (d, $J = 8.0$ Hz, 1H), 7.68 (d, $J = 8.0$ Hz, 1H), 7.39 (ddd, $J = 8.0, 7.0, 1.0$ Hz, 1H), 7.35 (s, 1H), 7.34 – 7.31 (m, 1H), 7.18 (s, 1H), 2.42 – 2.32 (m, 1H), 1.53 – 1.43 (m, 3H), 1.23 (d, $J = 7.5$ Hz, 18H), 1.07 – 1.01 (m, 2H), 0.85 – 0.77 (m, 2H).

^{13}C NMR (126 MHz, chloroform-*d*) δ 153.8, 135.9, 132.9, 129.1, 127.1, 126.0, 125.1, 123.7, 123.5, 112.4, 18.2, 13.1, 10.9, 8.0. The spectra is consist with previous report.⁴

((3-Benzyl)naphthalen-2-yl)oxy)triisopropylsilane.



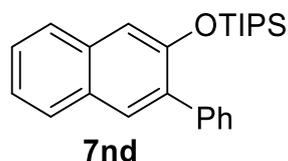
Colorless oil (74mg, 96% yield), 0.2 mmol scale reaction.

^1H NMR (500 MHz, chloroform-*d*) δ 7.76 – 7.71 (m, 2H), 7.52 (s, 1H), 7.44 (t, $J = 7.5$ Hz, 1H), 7.39 – 7.33 (comp, 3H), 7.32 – 7.28 (comp, 3H), 7.24 (s, 1H), 4.26 (s, 2H), 1.51 – 1.40 (m, 3H), 1.22 – 1.19 (m, 18H).

^{13}C NMR (126 MHz, chloroform-*d*) δ 152.9, 140.7, 133.2, 129.4, 129.1, 129.0, 128.4, 127.3, 126.1, 126.0, 125.5, 123.6, 112.5, 36.9, 18.2, 13.2.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{26}\text{H}_{35}\text{OSi}$, m/z : 391.2452, observed: 391.2448.

Triisopropyl((3-phenyl)naphthalen-2-yl)oxy)silane.

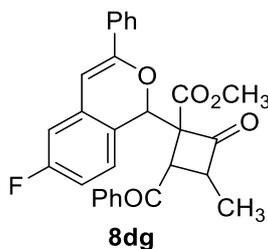


Colorless oil (18.8 mg, 25% yield), 0.2 mmol scale reaction.

^1H NMR (300 MHz, chloroform-*d*) δ 7.85 – 7.78 (comp, 2H), 7.73 (s, 1H), 7.64 – 7.59 (comp, 2H), 7.50 – 7.42 (comp, 3H), 7.41 – 7.33 (comp, 2H), 7.29 (s, 1H), 1.33 – 1.20 (m, 3H), 1.04 (d, $J = 7.5$ Hz, 18H).

^{13}C NMR (75 MHz, chloroform-*d*) δ 151.8, 139.1, 135.0, 134.0, 130.0, 129.1, 127.8, 127.0, 126.2, 126.0, 123.9, 113.9, 17.9, 13.0. The spectra is consist with previous report.⁴

Methyl 2-Benzoyl-1-(6-fluoro-3-phenyl-1H-isochromen-1-yl)-3-methyl-4-oxocyclobutane-1-carboxylate.



Colorless oil (10.8 mg, 23% yield, 5:1 dr).

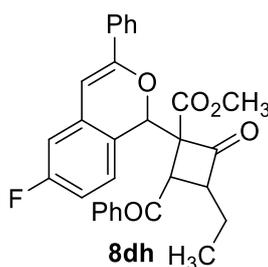
Major isomer: ^1H NMR (500 MHz, chloroform-*d*) δ 8.05 (d, $J = 7.5$ Hz, 2H), 7.78 – 7.71 (comp., 2H),

7.52 (t, $J = 7.5$ Hz, 2H), 7.44 - 7.39 (comp., 2H), 7.24 (s, 1H), 7.09 - 7.01 (comp., 2H), 6.96 - 6.85 (comp., 2H), 6.47 (s, 1H), 6.08 (s, 1H), 4.45 (d, $J = 8.8$ Hz, 1H), 4.20 - 4.05 (m, 1H), 3.63 (s, 3H), 0.90 (d, $J = 7.5$ Hz, 3H).

^{13}C NMR (126 MHz, Chloroform- d) δ 201.67, 197.10, 167.26, 163.20 (d, $J = 245$ Hz), 151.18, 137.09, 133.64, 133.22, 129.45, 128.87, 128.71, 128.46, 128.3, 128.30, 127.84, 127.14, 127.06, 124.90, 124.43, 113.78 (d, $J = 21$ Hz), 111.22 (d, $J = 21$ Hz), 102.39, 76.10, 55.11, 52.93, 45.50, 12.63.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{29}\text{H}_{24}\text{FO}_5$, m/z : 471.1608, observed: 471.19.1607.

Methyl 2-Benzoyl-1-(6-fluoro-3-phenyl-1H-isochromen-1-yl)-3-methyl-4-oxocyclobutane-1-carboxylate.



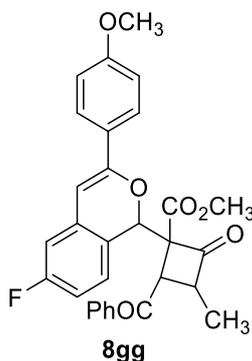
Colorless oil (8.4 mg, 18% yield, 5:1 dr).

Major isomer: ^1H NMR (500 MHz, Chloroform- d) δ 8.06 (d, $J = 7.5$ Hz, 2H), 7.74 - 7.72 (comp, 2H), 7.66 - 7.61 (m, 1H), 7.53 - 7.50 (comp, 2H), 7.44 - 7.39 (comp, 3H), 7.25 - 7.22 (m, 1H), 7.09 - 7.07 (m, 1H), 6.93 - 6.85 (m, 2H), 6.45 (s, 1H), 6.07 (s, 1H), 4.49 (d, $J = 9.0$ Hz, 1H), 4.12 - 4.04 (m, 1H), 3.62 (s, 3H), 1.51 - 1.43 (m, 1H), 1.17 - 1.07 (m, 1H), 0.71 (t, $J = 7.5$ Hz, 3H).

^{13}C NMR (126 MHz, Chloroform- d) δ 201.16, 197.37, 162.26, 160.42 (d, $J = 245$ Hz), 129.42, 128.85, 128.66, 128.46, 128.35, 127.84, 124.90, 124.42, 113.74(d, $J = 21$ Hz),, 111.18 (d, $J = 21$ Hz), 102.26, 76.20, 61.85, 52.93, 46.94, 43.57, 21.45, 11.19.

HRMS (ESI) scaled for $[\text{M}+\text{H}]^+$: $\text{C}_{30}\text{H}_{26}\text{FO}_5$, m/z : 485.1764, observed: 485.1760.

Methyl 2-benzoyl-1-(6-fluoro-3-(4-methoxyphenyl)-1H-isochromen-1-yl)-3-methyl-4-oxocyclobutane-1-carboxylate.



Colorless oil (6 mg, 12% yield).

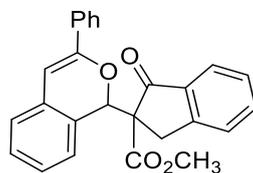
Major isomer: ^1H NMR (500 MHz, chloroform- d) δ 8.04 (d, $J = 7.5$ Hz, 2H), 7.68 (d, $J = 9.0$ Hz, 2H), 7.62 (t, $J = 7.5$ Hz, 1H), 7.52 (t, $J = 7.5$ Hz, 2H), 7.45 - 7.38 (m, 1H), 7.01 (dd, $J = 8.4, 5.5$ Hz, 1H), 6.96 - 6.92 (comp, 2H), 6.91 - 6.82 (comp, 2H), 6.34 (s, 1H), 6.07 (s, 1H), 4.43 (d, $J = 8.5$ Hz, 1H), 4.19 - 4.07 (m, 1H), 3.86 (s, 3H), 3.63 (s, 3H), 0.90 (d, $J = 7.5$ Hz, 3H).

^{13}C NMR (126 MHz, chloroform- d) δ 201.71, 200.06, 164.20, 162.55 (d, $J = 245$ Hz), 151.24, 137.12,

133.57, 128.82, 128.34, 126.04, 114.09, 113.29 (d, $J = 21$ Hz), 110.85 (d, $J = 21$ Hz), 100.67, 76.13, 55.35; 55.16; 52.89; 45.50; 12.68.

HRMS (ESI) scaled for $[M+H]^+$: $C_{30}H_{26}FO_6$, m/z : 501.1708, observed: 501.1686.

Methyl 1-oxo-2-(3-phenyl-1H-isochromen-1-yl)-2,3-dihydro-1H-indene-2-carboxylate.



13

Colorless oil (17 mg, 45% yield, 1:1 dr), 0.1 mmol scale reaction.

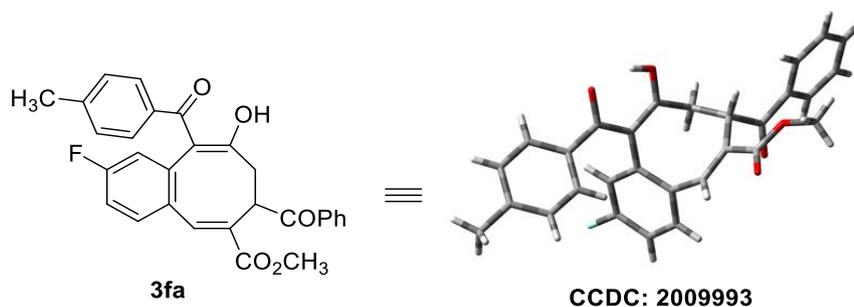
13 was reported as 1:1 mixture spectrum:

1H NMR (500 MHz, chloroform-*d*) δ 7.91 (d, $J = 7.5$ Hz, 1H), 7.73 (d, $J = 7.5$ Hz, 1H), 7.66 – 7.62 (comp., 2H), 7.57 (td, $J = 7.5, 1.0$ Hz, 1H), 7.52 – 7.47 (m, 1H), 7.45 (d, $J = 7.5$ Hz, 1H), 7.43 – 7.37 (comp., 4H), 7.35 – 7.29 (comp., 5H), 7.25 – 7.19 (comp., 4H), 7.16 (tt, $J = 7.5, 2.5$ Hz, 2H), 7.07 (d, $J = 7.5$ Hz, 1H), 7.01 (d, $J = 7.5$ Hz, 1H), 6.99 – 6.95 (comp, 2H), 6.70 (s, 1H), 6.40 (s, 1H), 6.37 (s, 1H), 6.35 (s, 1H), 4.04 (d, $J = 17.5$ Hz, 1H), 3.92 (d, $J = 17.5$ Hz, 1H), 3.92 (s, 3H), 3.84 (s, 3H), 3.69 (d, $J = 17.3$ Hz, 1H), 3.42 (d, $J = 17.3$ Hz, 1H).

^{13}C NMR (126 MHz, chloroform-*d*) δ 199.22, 198.48, 168.88, 168.12, 154.48, 154.37, 153.45, 151.99, 135.5, 135.4, 134.85, 134.75, 133.83, 133.04, 131.49, 129.02, 128.85, 128.76, 128.53, 128.37, 128.15, 127.63, 127.44, 127.00, 126.78, 126.33, 126.18, 125.79, 125.08, 124.84, 124.67, 124.39, 124.33, 123.50, 101.01, 99.89, 80.14, 79.77, 70.24, 65.15, 53.43, 53.32, 33.24, 31.68.

HRMS (ESI) scaled for $[M+H]^+$: $C_{26}H_{21}O_4$, m/z : 419.1254, observed: 419.1261.

8. X-ray crystal structure of the product 3fa, 7ma and 15da



Single crystals of $C_{29}H_{22}FO_5$ were prepared by slow evaporation of a EA/hexane solution. A suitable colorless plate-like crystal with dimensions of 0.021 mm \times 0.107 mm \times 0.141 mm, was mounted in paratone oil onto a nylon loop. All data were collected at 100.0(1) K, using a XtaLAB Synergy/ Dualflex, HyPix fitted with $CuK\alpha$ radiation ($\lambda = 1.54184$ Å). Data collection and unit cell refinement were performed using *CrysAlisPro* software.^[1] The total number of data were measured in the $6.76^\circ < 2\theta < 144.8^\circ$, using ω scans. Data processing and absorption correction, giving minimum and maximum

transmission factors (0.769, 1.000) were accomplished with *CrysAlisPro*^[1] and *SCALE3 ABSPACK*^[2], respectively. The structure, using *Olex2*^[3], was solved with the *ShelXT*^[4] structure solution program using direct methods and refined (on F^2) with the *ShelXL*^[5] refinement package using full-matrix, least-squares techniques. All non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atom positions were determined by geometry and refined by a riding model.

^[1] *CrysAlisPro* 1.171.40.63a (Rigaku Oxford Diffraction, 2019)

^[2] *SCALE3 ABSPACK* -An Oxford Diffraction program(1.0.4,gui:1.0.3) (C) 2005 Oxford Diffraction Ltd.

^[3] O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard, H. Puschmann, *J. Appl. Cryst.* 2009, **42**, 339.

^[4] G. M. Sheldrick, *Acta Cryst.* 2015, **A71**, 3.

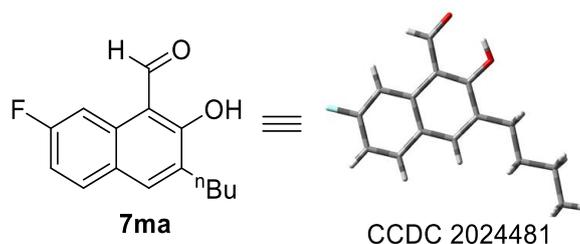
^[5] G. M. Sheldrick, *Acta Cryst.* 2008, **A64**, 112.

Table S1: Crystallographic data and structure refinement for **3fa**

Identification code	3fa
Empirical formula	C ₂₉ H ₂₂ FO ₅
Formula weight	469.46
Crystal system	Monoclinic
Space group	<i>P</i> 2 ₁
<i>a</i> (Å)	9.9631(3)
<i>b</i> (Å)	9.1494(2)
<i>c</i> (Å)	13.3605(4)
<i>α</i> (°)	90
<i>β</i> (°)	102.001(3)
<i>γ</i> (°)	90
Volume (Å ³)	1191.28(6)
<i>Z</i>	2
ρ (calc.)	1.309
λ	1.54184
Temp. (K)	100.0(1)
F(000)	490
μ (mm ⁻¹)	0.779
T _{min} , T _{max}	0.769, 1.000
2 θ _{range} (°)	6.76 to 144.8
Reflections collected	14112
Independent reflections	4208 [R(int) = 0.0406]
Completeness	99.9%
Data / restraints / parameters	4208 / 1 / 319
Observed data [<i>I</i> > 2 σ (<i>I</i>)]	3887
$wR(F^2$ all data)	0.1190
$R(F$ obsd data)	0.0482
Goodness-of-fit on F^2	1.04
largest diff. peak and hole (e Å ⁻³)	0.45 / -0.24

$$wR_2 = \{ \Sigma [w(F_o^2 - F_c^2)^2] / \Sigma [w(F_o^2)^2] \}^{1/2}$$

$$R_1 = \Sigma ||F_o| - |F_c|| / \Sigma |F_o|$$



Single crystals of $C_{15}H_{15}FO_2$ were prepared by slow evaporation of a dichloromethane and hexane solution. A suitable colorless block-like crystal, with dimensions of 0.175 mm \times 0.104 mm \times 0.072 mm, was mounted in paratone oil onto a nylon loop. All data were collected at 100.0(1) K, using a XtaLAB Synergy/ Dualflex, HyPix fitted with $CuK\alpha$ radiation ($\lambda = 1.54184 \text{ \AA}$). Data collection and unit cell refinement were performed using *CrysAlisPro* software.^[1] The total number of data were measured in the $9.27^\circ < 2\theta < 152.5^\circ$, using ω scans. Data processing and absorption correction, giving minimum and maximum transmission factors (0.600, 1.000) were accomplished with *CrysAlisPro*^[6] and *SCALE3 ABSPACK*^[7], respectively. The structure, using Olex2^[8], was solved with the ShelXT^[9] structure solution program using direct methods and refined (on F^2) with the ShelXL^[10] refinement package using full-matrix, least-squares techniques. All non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atom positions were determined by geometry and refined by a riding model.

[1] CrysAlisPro 1.171.40.63a (Rigaku Oxford Diffraction, 2019)

[7] SCALE3 ABSPACK -An Oxford Diffraction program(1.0.4,gui:1.0.3) (C) 2005 Oxford Diffraction Ltd.

[8] O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard, H. Puschmann, *J. Appl. Cryst.* 2009, **42**, 339.

[9] G. M. Sheldrick, *Acta Cryst.* 2015, **A71**, 3.

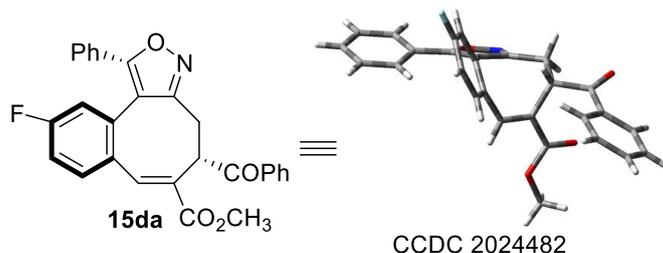
[10] G. M. Sheldrick, *Acta Cryst.* 2008, **A64**, 112.

Table S2: Crystallographic data and structure refinement for **7ma**

Identification code	7ma
Empirical formula	C ₁₅ H ₁₅ FO ₂
Formula weight	246.27
Crystal system	Monoclinic
Space group	<i>P2₁/n</i>
<i>a</i> (Å)	7.3867(2)
<i>b</i> (Å)	19.0785(5)
<i>c</i> (Å)	9.3357(2)
<i>α</i> (°)	90
<i>β</i> (°)	111.972(3)
<i>γ</i> (°)	90
Volume (Å ³)	1220.09(6)
<i>Z</i>	4
ρ (calc.)	1.341
λ	1.54184
Temp. (K)	100.0(1)
F(000)	520
μ (mm ⁻¹)	0.808
T _{min} , T _{max}	0.600, 1.000
2θ _{range} (°)	9.27 to 152.5
Reflections collected	13853
Independent reflections	2429 [R(int) = 0.0414]
Completeness	99.8%
Data / restraints / parameters	2429 / 0 / 165
Observed data [I > 2σ(I)]	2122
wR(<i>F</i> ² all data)	0.1096
<i>R</i> (<i>F</i> obsd data)	0.0415
Goodness-of-fit on <i>F</i> ²	1.09
largest diff. peak and hole (e Å ⁻³)	0.21 / -0.32

$$wR_2 = \{ \sum [w(F_o^2 - F_c^2)^2] / \sum [w(F_o^2)^2] \}^{1/2}$$

$$R_1 = \sum ||F_o| - |F_c|| / \sum |F_o|$$



Single crystals of $C_{28}H_{21}FNO_4$ were prepared by slow evaporation of a dichloromethane and hexane solution. A suitable colorless plank-like crystal, with dimensions of 0.369 mm \times 0.180 mm \times 0.058 mm, was mounted in paratone oil onto a nylon loop. All data were collected at 100.0(1) K, using a XtaLAB Synergy/ Dualflex, HyPix fitted with CuK α radiation ($\lambda = 1.54184$ Å). Data collection and unit cell refinement were performed using *CrysAlisPro* software.^[1] The total number of data were measured in the $7.29^\circ < 2\theta < 153.4^\circ$, using ω scans. Data processing and absorption correction, giving minimum and maximum transmission factors (0.437, 1.000) were accomplished with *CrysAlisPro*^[11] and *SCALE3 ABSPACK*^[12], respectively. The structure, using Olex2^[13], was solved with the ShelXT^[14] structure solution program using direct methods and refined (on F^2) with the ShelXL^[15] refinement package using full-matrix, least-squares techniques. All non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atom positions were determined by geometry and refined by a riding model.

^[1] CrysAlisPro 1.171.40.63a (Rigaku Oxford Diffraction, 2019)

^[12] SCALE3 ABSPACK -An Oxford Diffraction program(1.0.4,gui:1.0.3) (C) 2005 Oxford Diffraction Ltd.

^[13] O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard, H. Puschmann, *J. Appl. Cryst.* 2009, **42**, 339.

^[14] G. M. Sheldrick, *Acta Cryst.* 2015, **A71**, 3.

^[15] G. M. Sheldrick, *Acta Cryst.* 2008, **A64**, 112.

Table S3: Crystallographic data and structure refinement for **15da**

Identification code	15da
Empirical formula	C ₂₈ H ₂₁ FNO ₄
Formula weight	454.46
Crystal system	Monoclinic
Space group	<i>P2₁/c</i>
<i>a</i> (Å)	12.28538(14)
<i>b</i> (Å)	20.3181(2)
<i>c</i> (Å)	8.91344(10)
<i>α</i> (°)	90
<i>β</i> (°)	99.0372(11)
<i>γ</i> (°)	90
Volume (Å ³)	2197.31(4)
<i>Z</i>	4
ρ (calc.)	1.374
λ	1.54184
Temp. (K)	100.0(1)
F(000)	948
μ (mm ⁻¹)	0.804
T _{min} , T _{max}	0.437, 1.000
2 θ _{range} (°)	7.29 to 153.4
Reflections collected	21382
Independent reflections	4485 [R(int) = 0.0402]
Completeness	99.9%
Data / restraints / parameters	4485 / 0 / 308
Observed data [<i>I</i> > 2 σ (<i>I</i>)]	4040
$wR(F^2$ all data)	0.1051
$R(F$ obsd data)	0.0383
Goodness-of-fit on F^2	1.05
largest diff. peak and hole (e Å ⁻³)	0.29 / -0.78

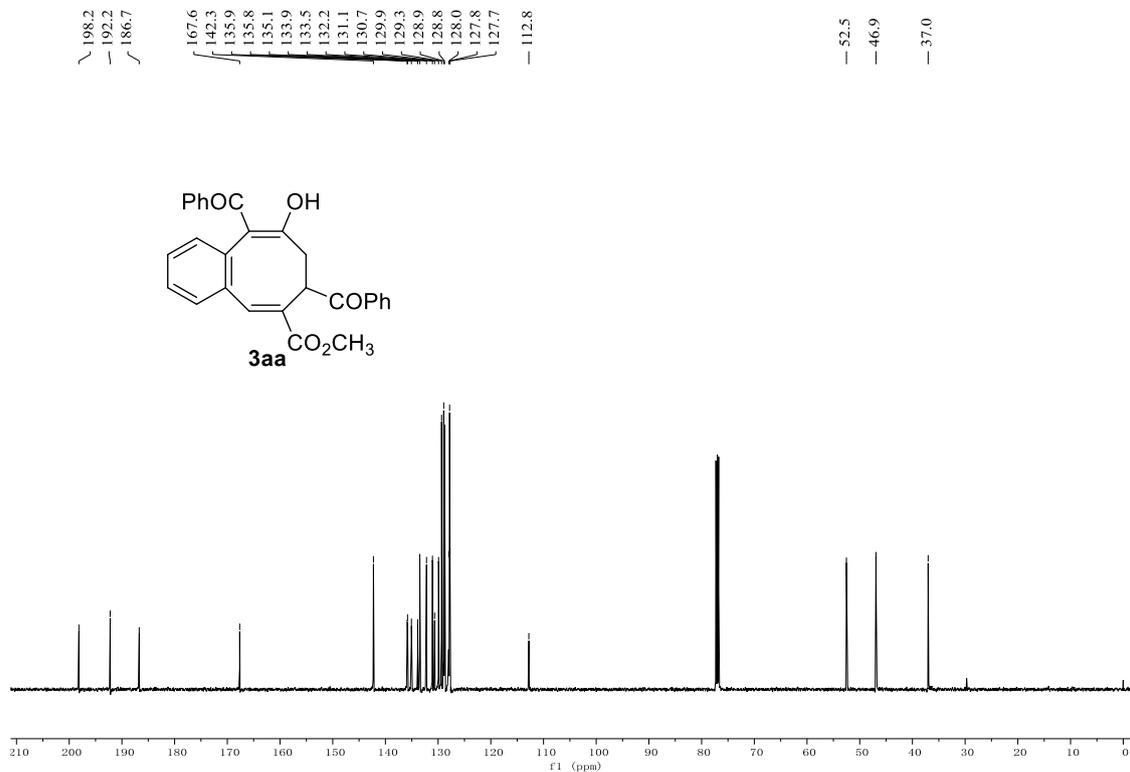
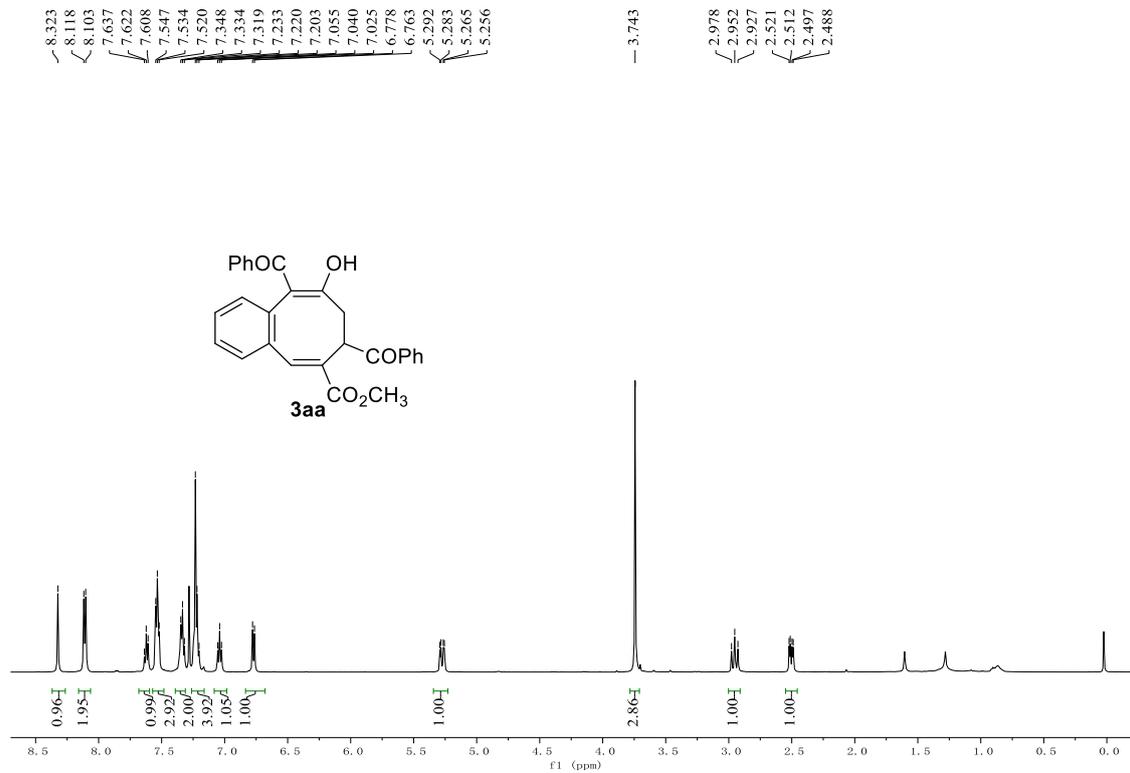
$$wR_2 = \{ \sum [w(F_o^2 - F_c^2)^2] / \sum [w(F_o^2)^2] \}^{1/2}$$

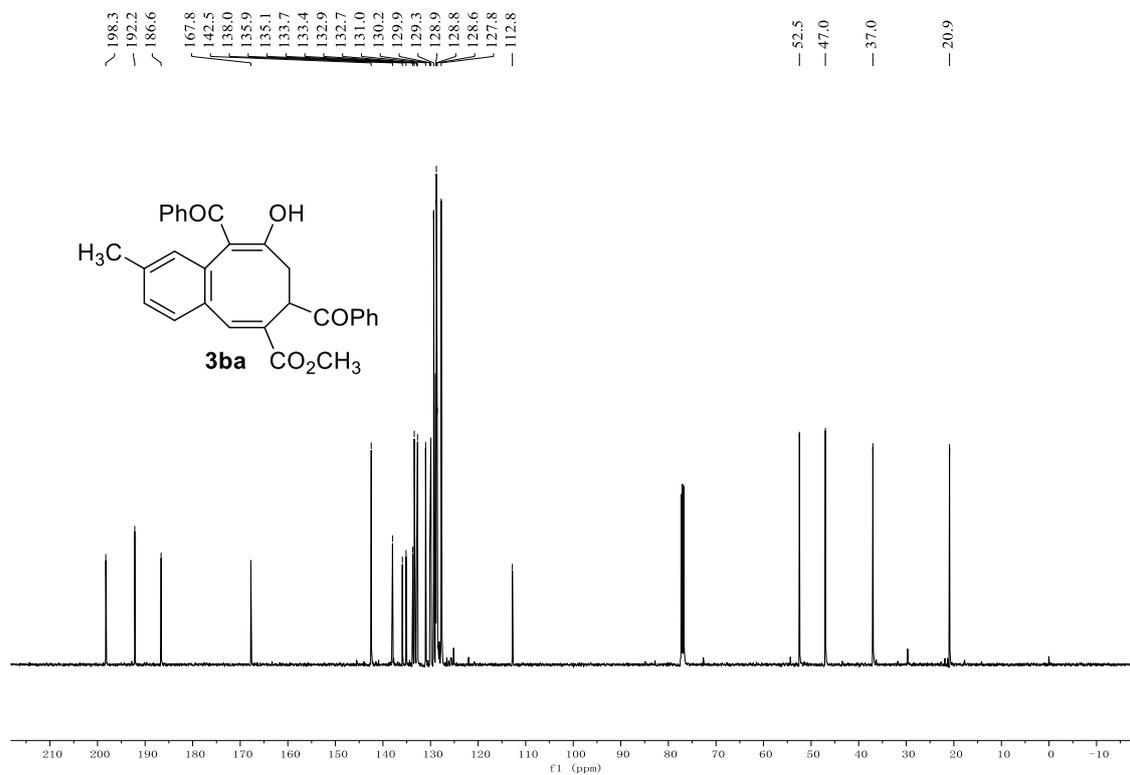
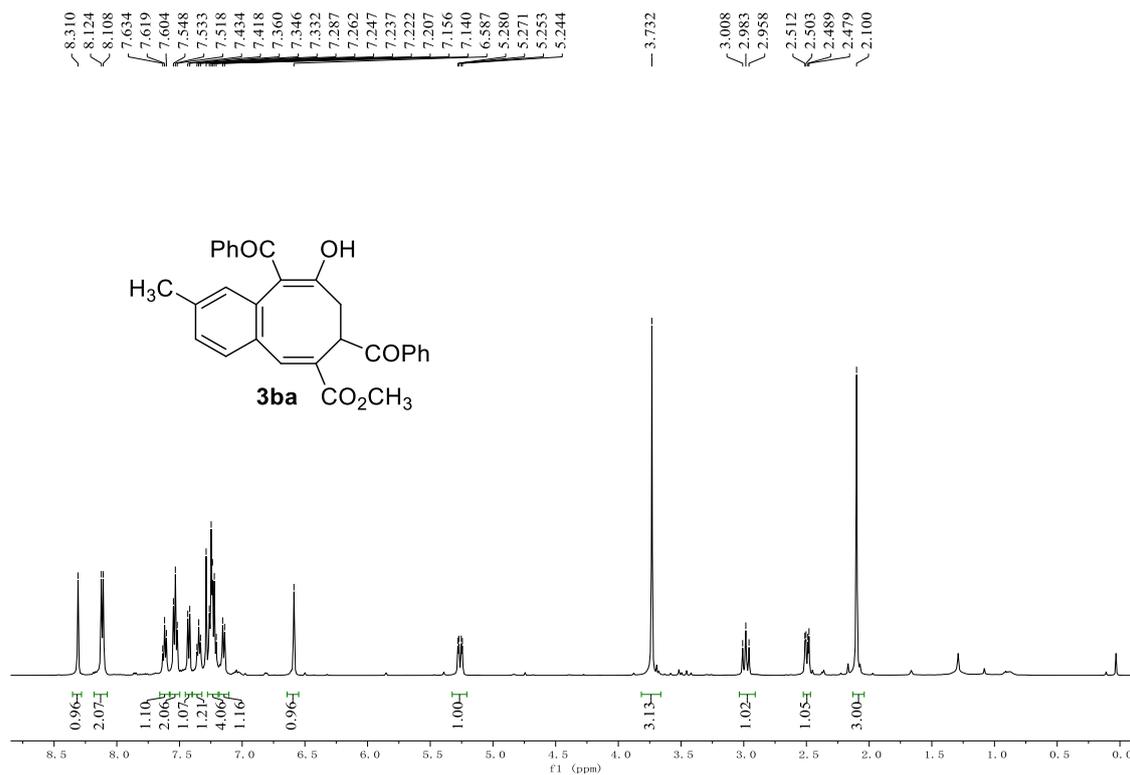
$$R_1 = \sum ||F_o| - |F_c|| / \sum |F_o|$$

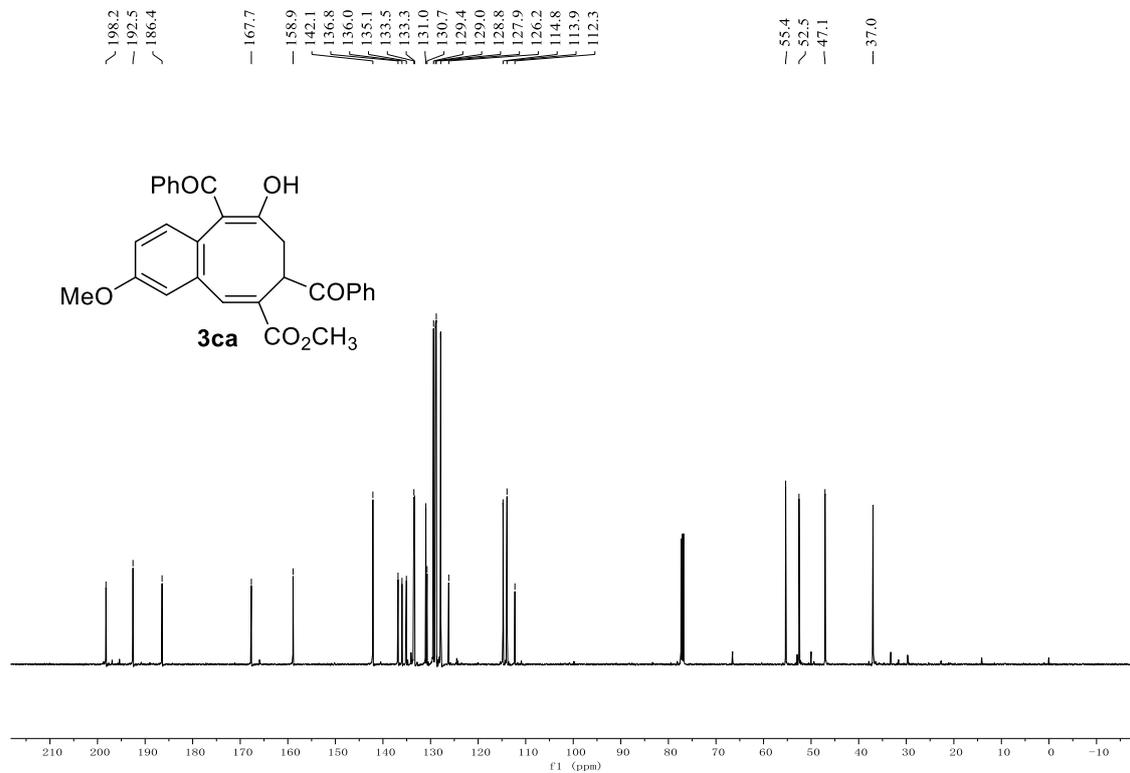
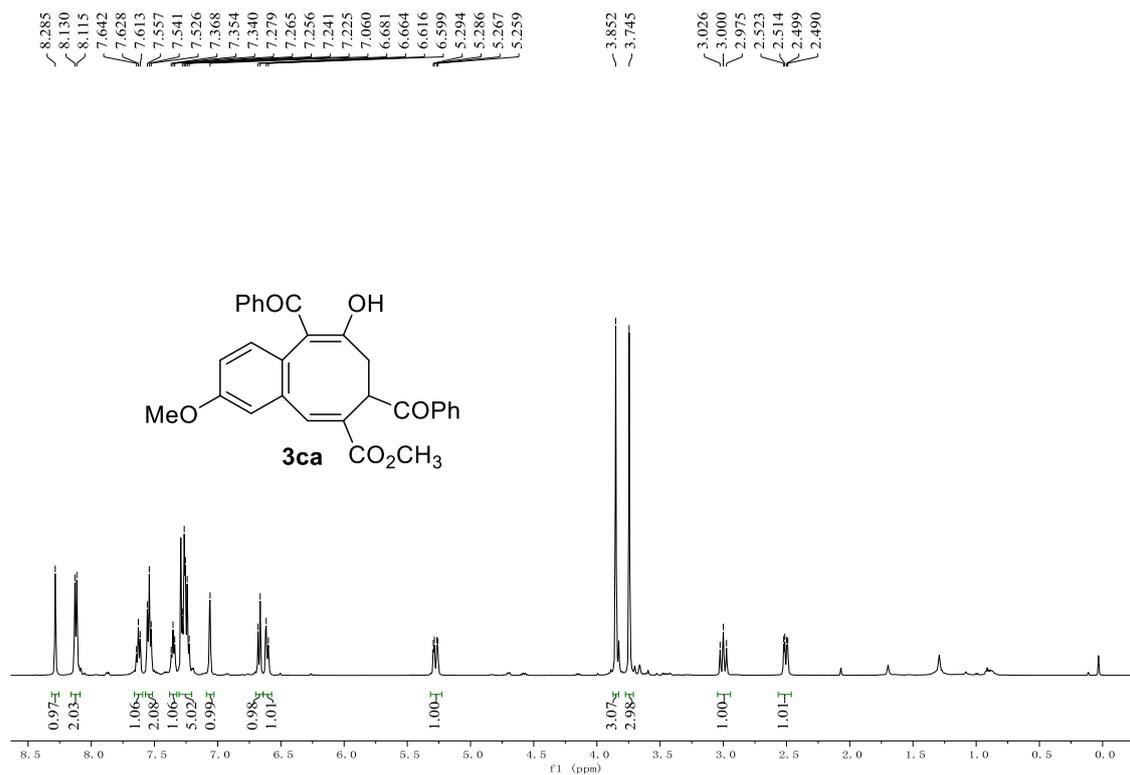
9. References

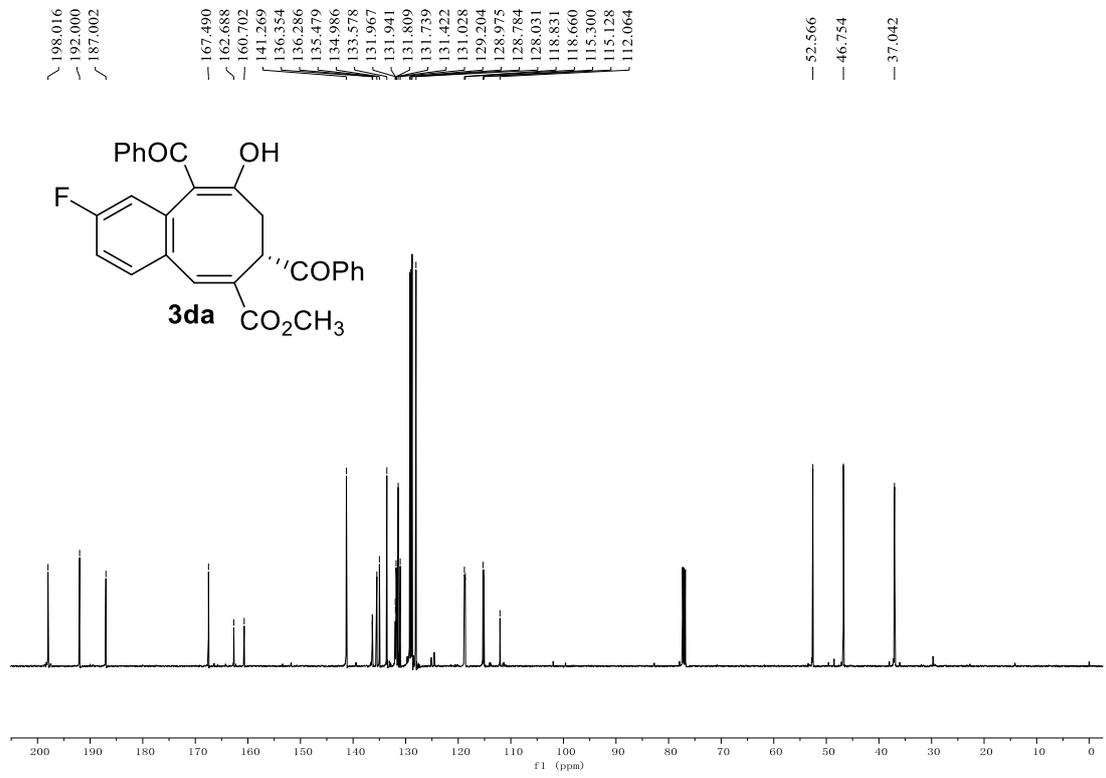
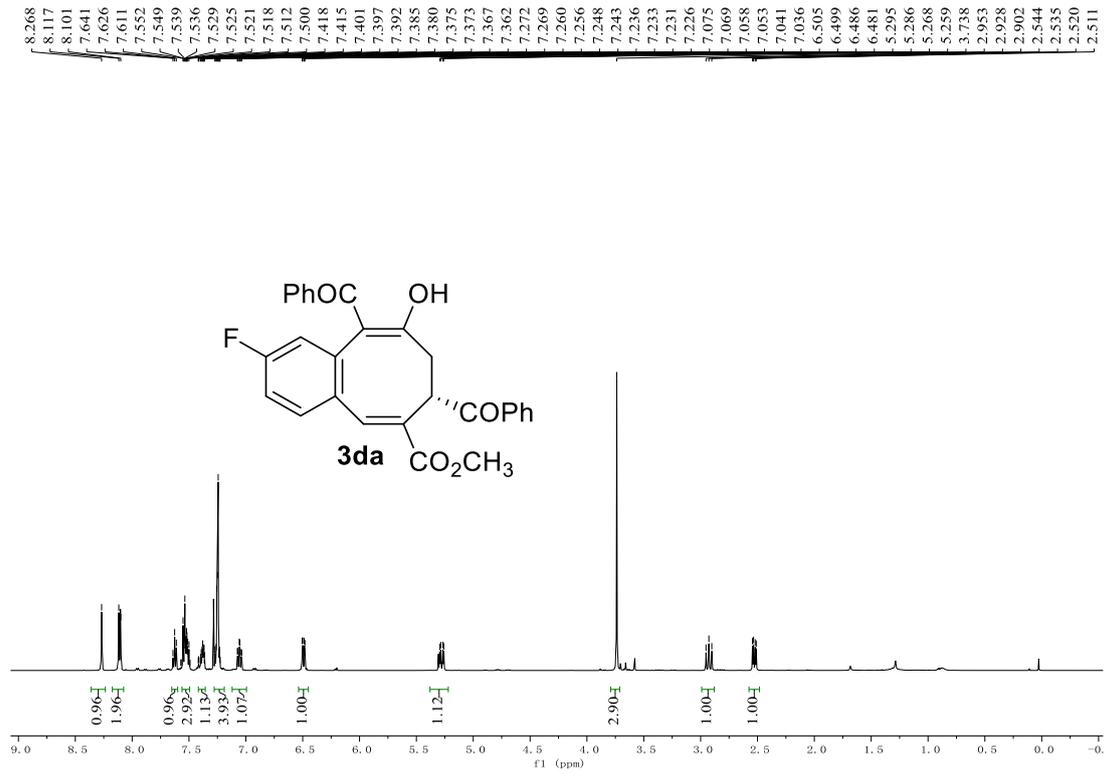
1. (a) N. T. Patil, Y. Yamamoto, *J. Org. Chem.* 2004, **69**, 5139. (b) H. Qian, W. X. Zhao, Z. B. Wang, J. W. Sun, *J. Am. Chem. Soc.* 2015, **137**, 560.
2. (a) Y. M. Deng, L. A. Massey, P. Y. Zavalij, M. P. Doyle, *Angew. Chem. Int. Ed.* 2017, **56**, 7479. (b) K. Dong, K. O. Marichev, X. Xu, M. P. Doyle, *Synlett* 2019, **30**, 1457. (c) R. Wang, K. O. Marichev, K. Y. Dong, J. A. Jensen, M. P. Doyle, *J. Org. Chem.* 2020, **85**, 9475.
3. Y. M. Deng, C. Jing, M. P. Doyle, *Chem. Commun.* 2015, **51**, 12924.
4. (a) R. F. Sweis, M. P. Schramm, S. A. Kozmin, *J. Am. Chem. Soc.* 2004, **126**, 7442. (b) Y. E. Türkmen, T. J. Montavon, S. A. Kozmin, V. H. Rawal, *J. Am. Chem. Soc.* 2012, **134**, 9062.
5. (a) K. O. Marichev, K. Wang, K. Dong, N. Greco, L. A. Massey, Y. Deng, H. Arman, M. P. Doyle, *Angew. Chem. Int. Ed.* 2019, **58**, 16188. (b) K. O. Marichev, K. Dong, L. A. Massey, Y. Deng, L.; Wang, K. De Angelis, H. Arman, M. P. Doyle, *Nature Communications* 2019, **10**, 5328.
6. Q. Q. Chen, Y. M. Deng, M. Lankelma, M. P. Doyle, *Chem. Soc. Rev.* 2017, **46**, 5425.

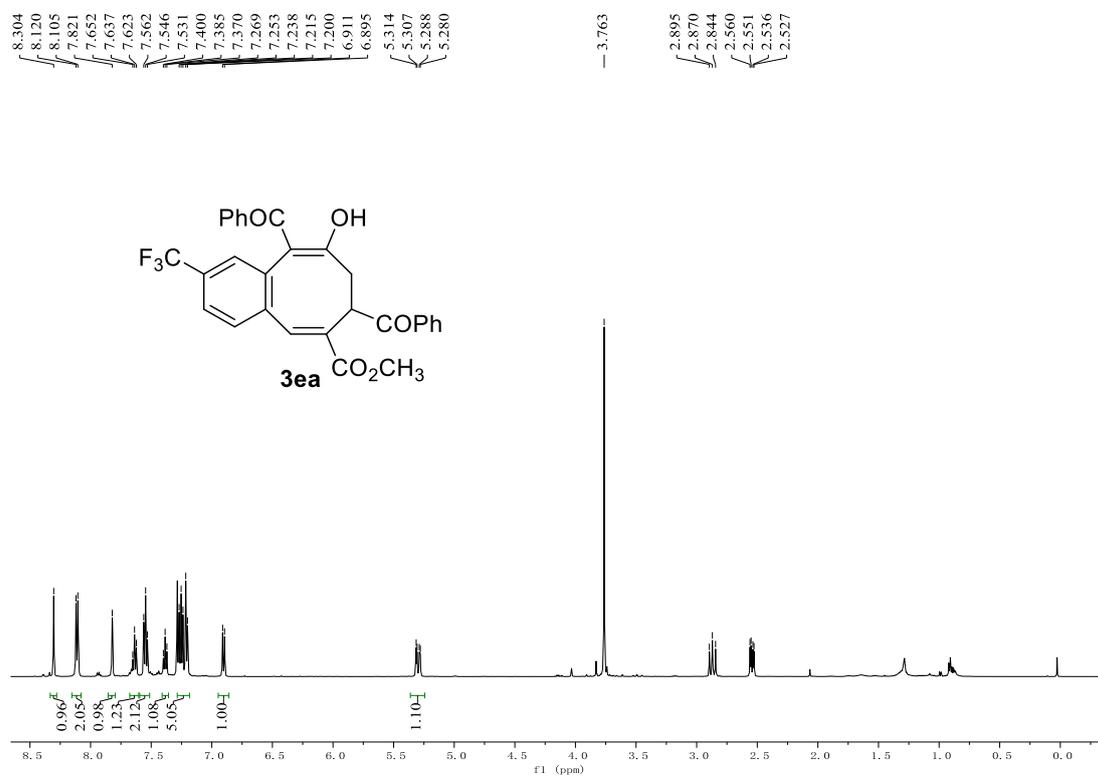
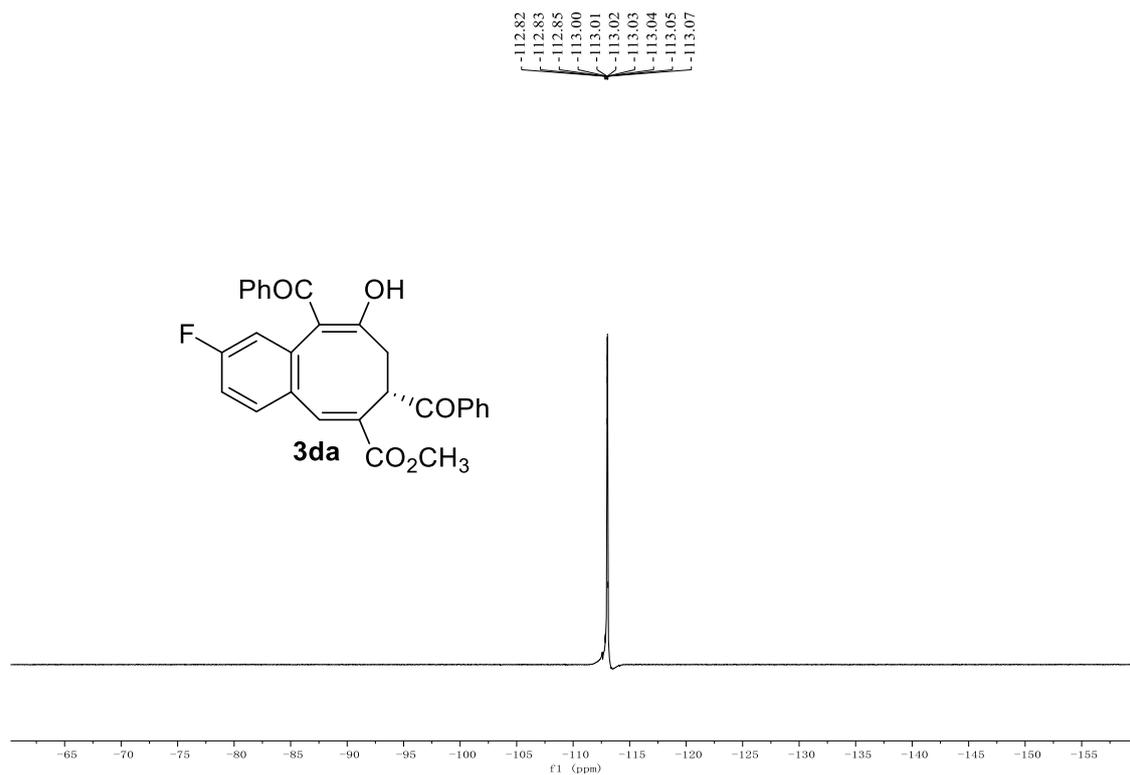
10. NMR spectrum of products

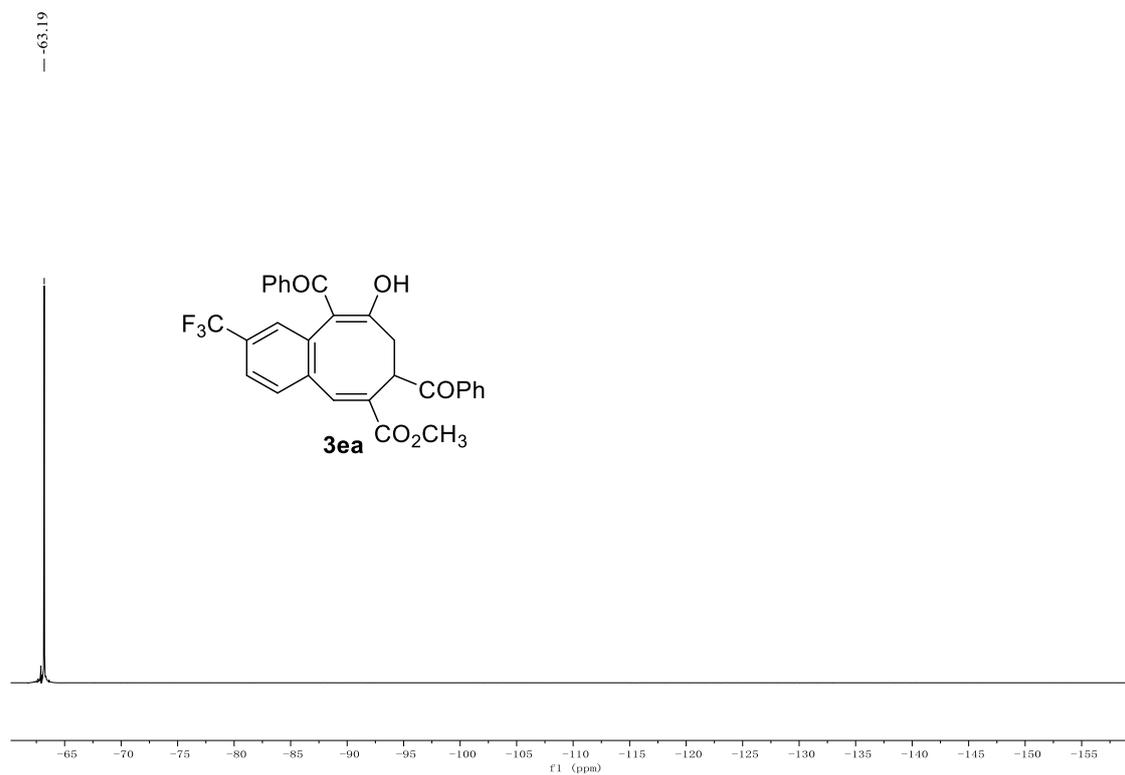
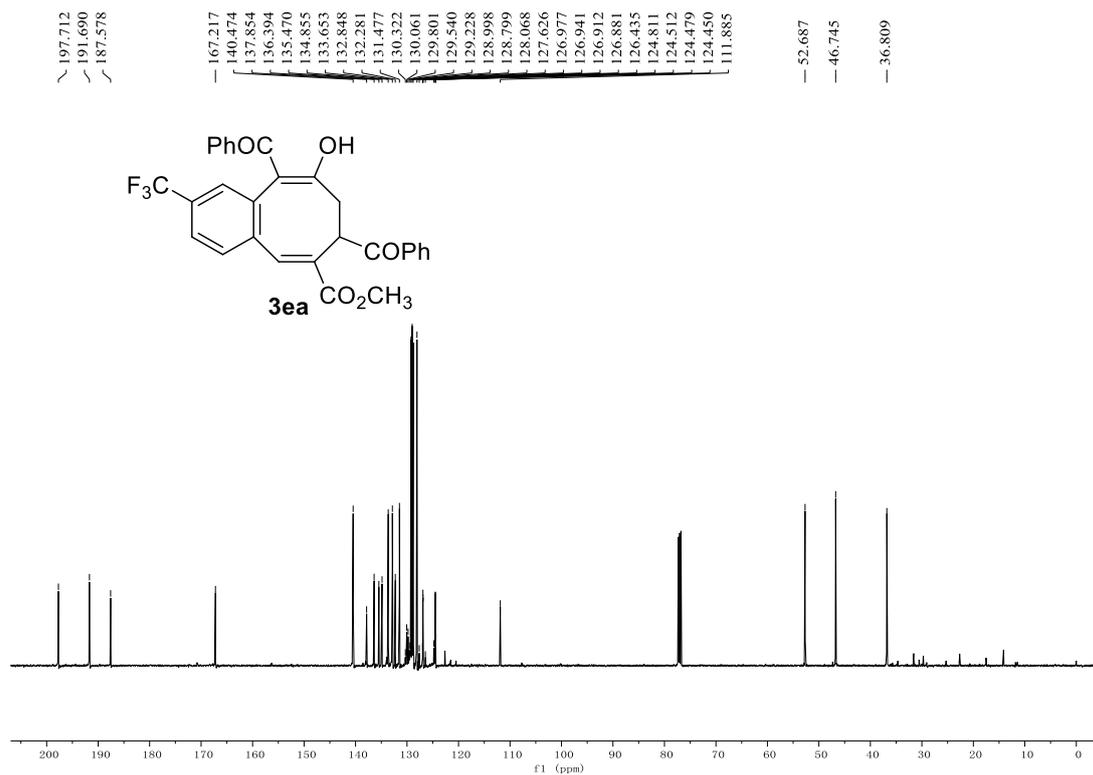


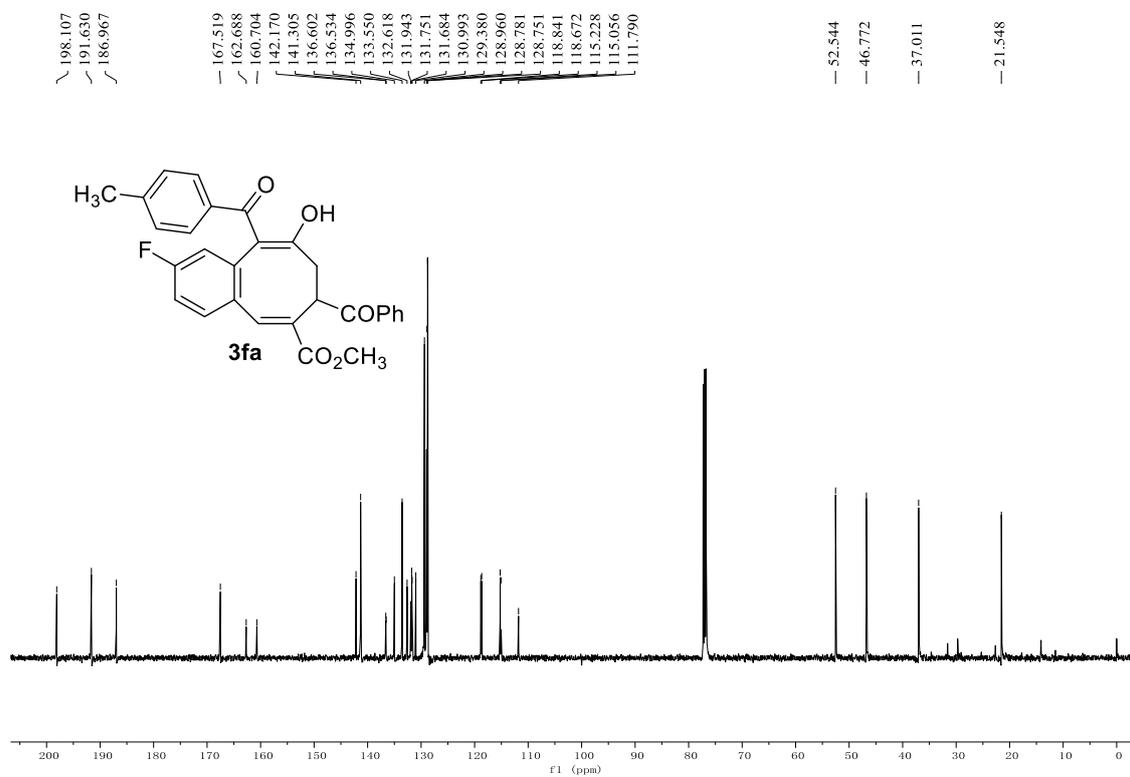
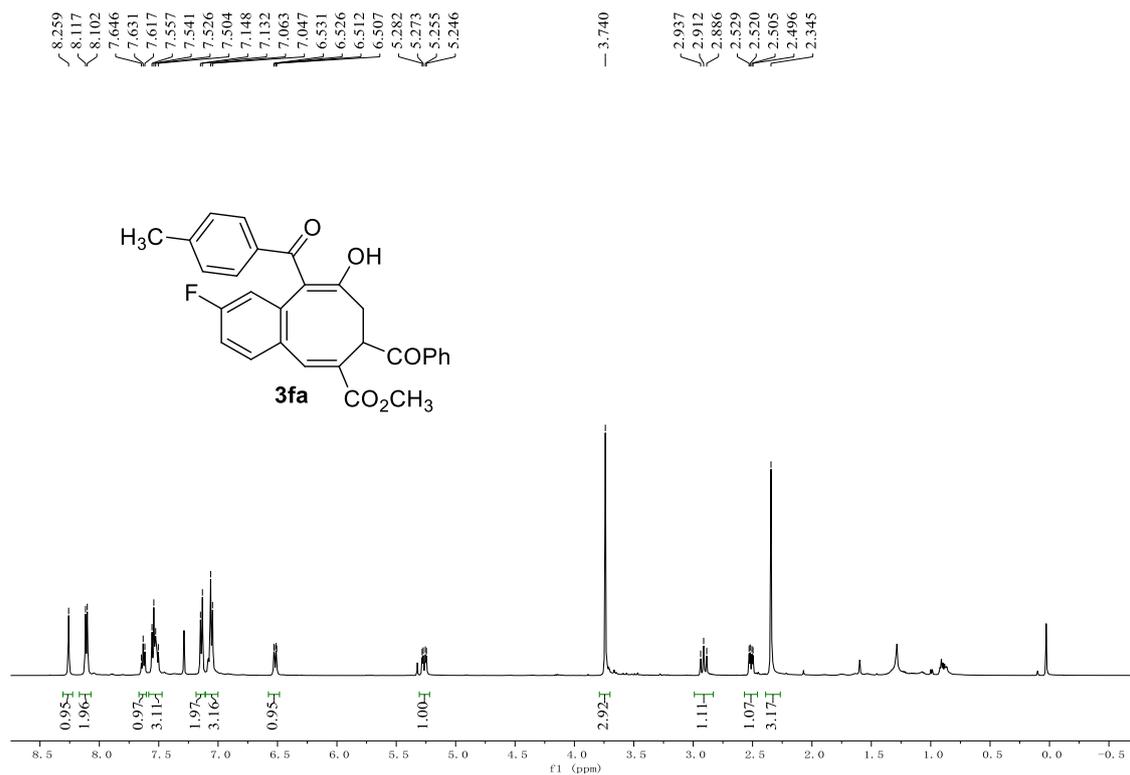


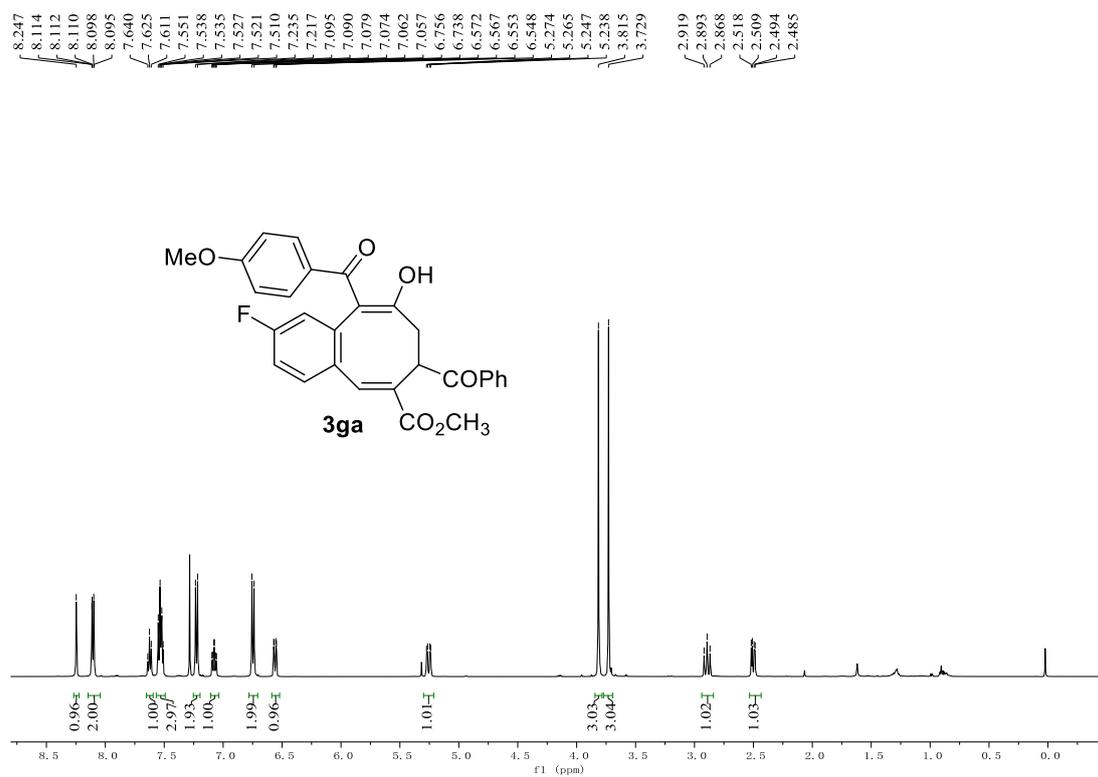
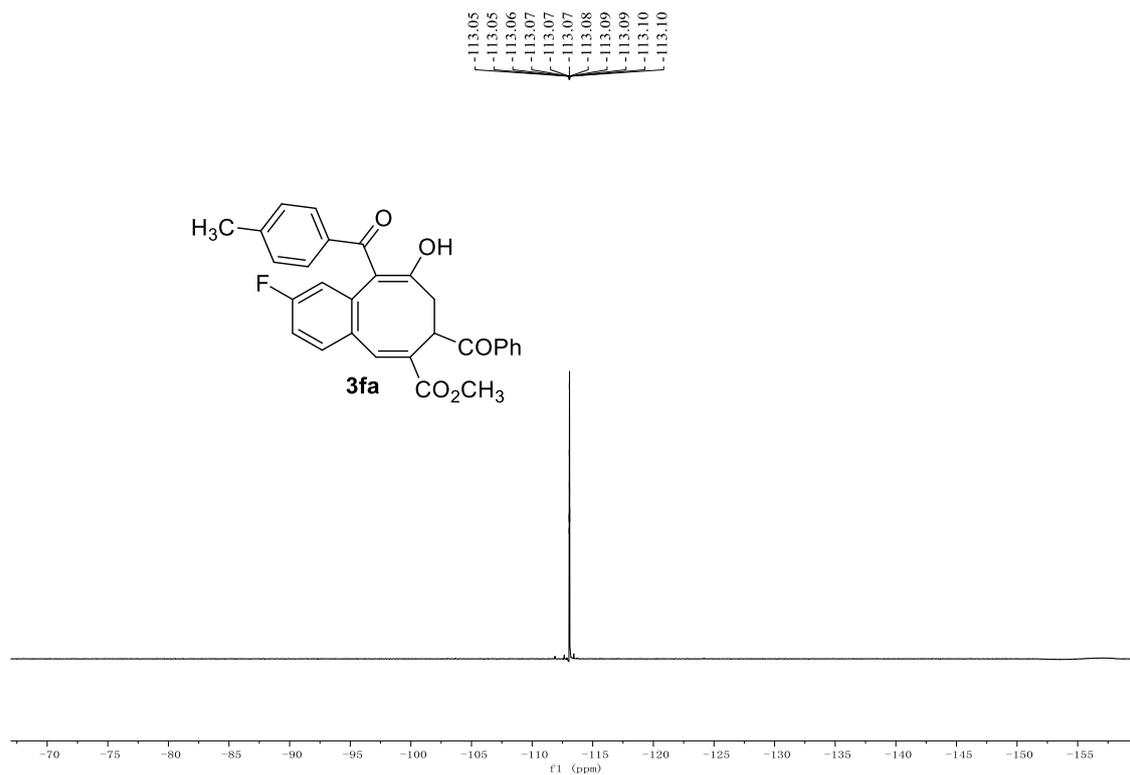


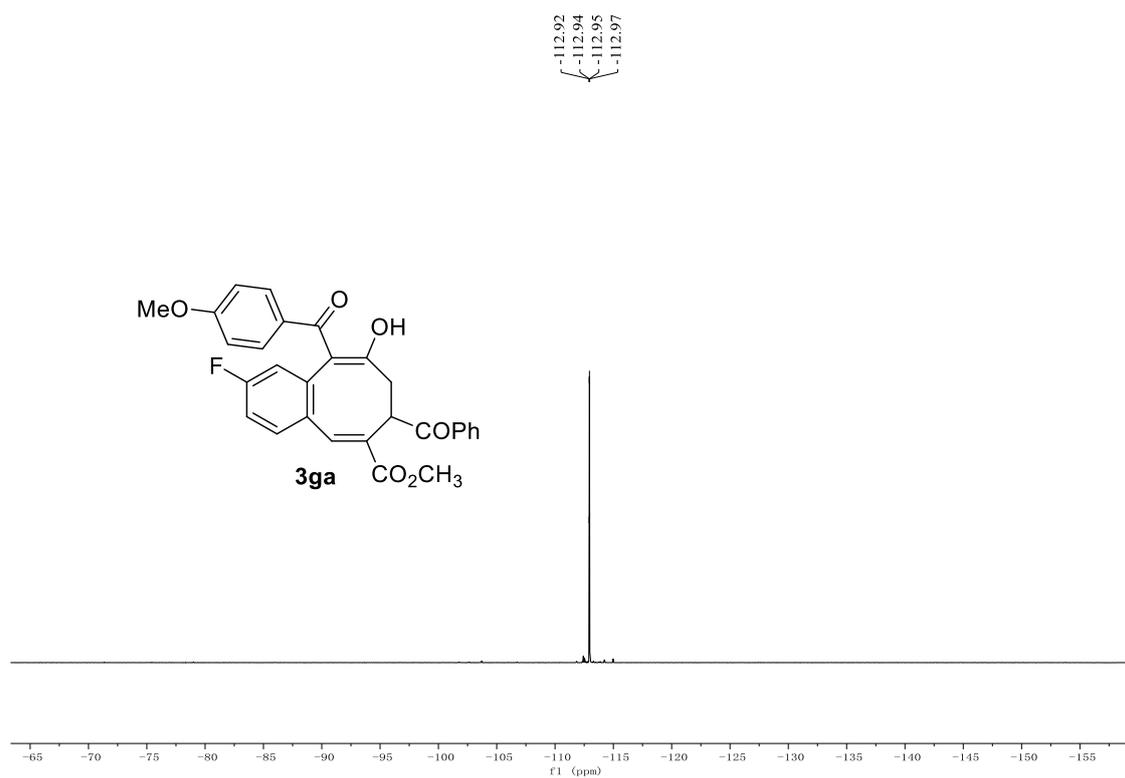
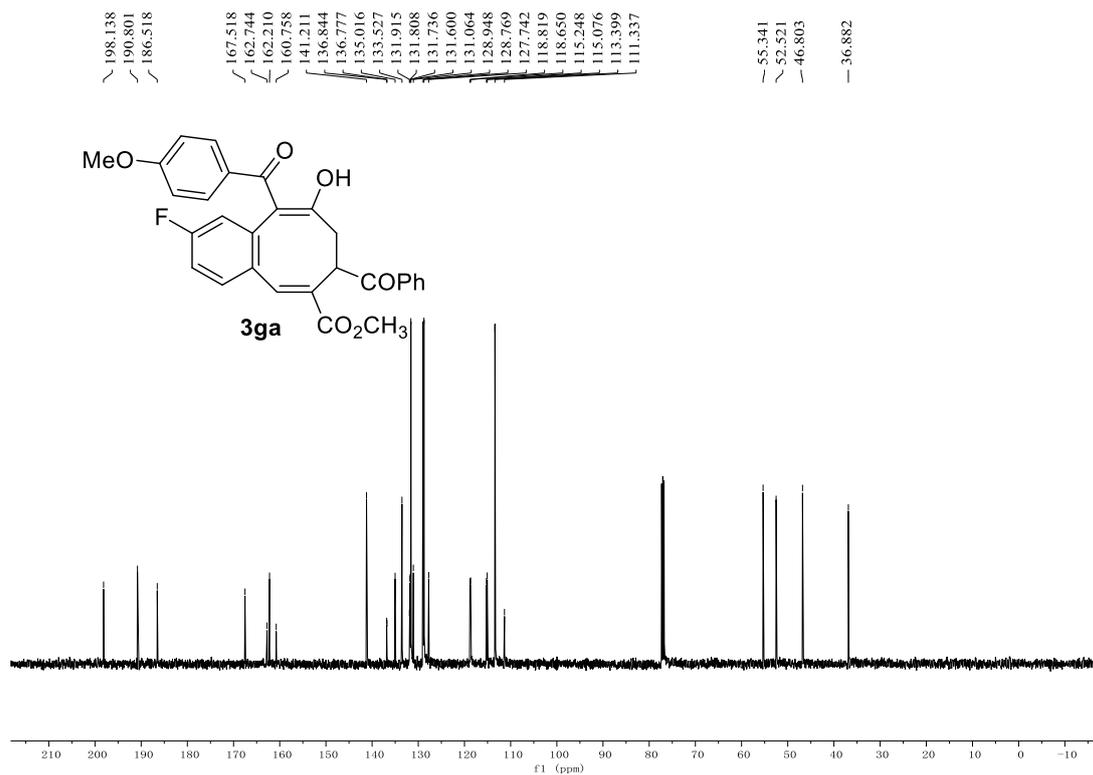


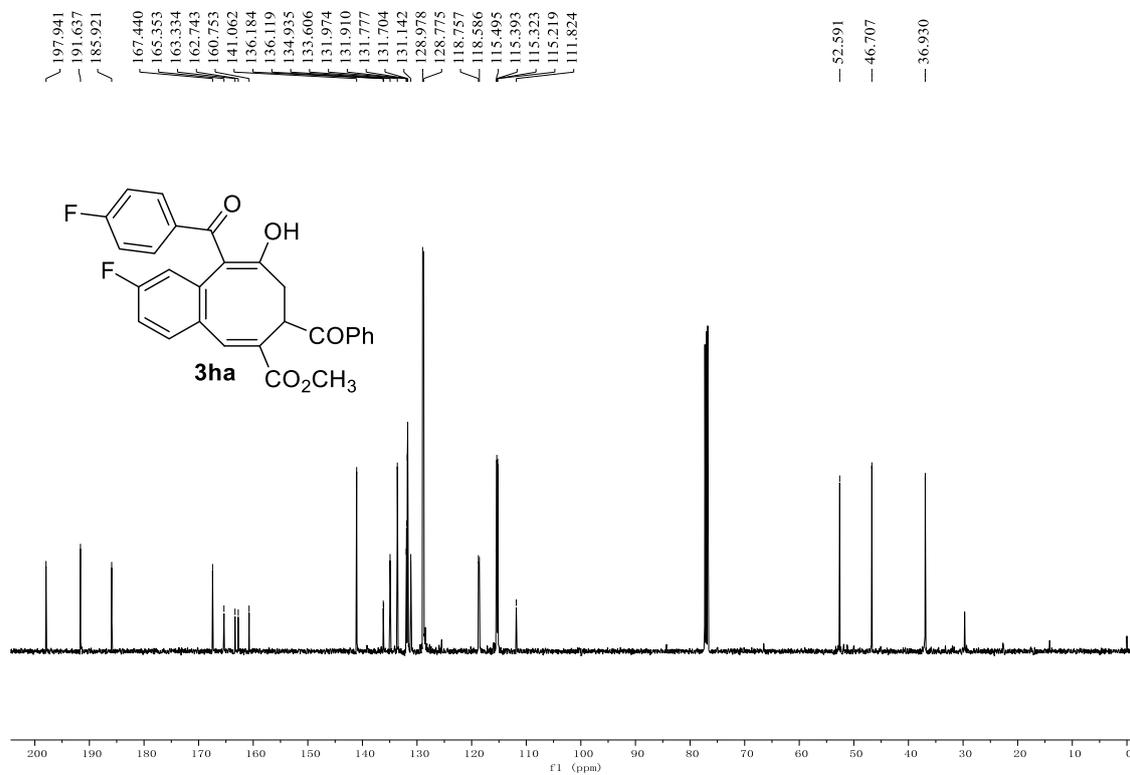
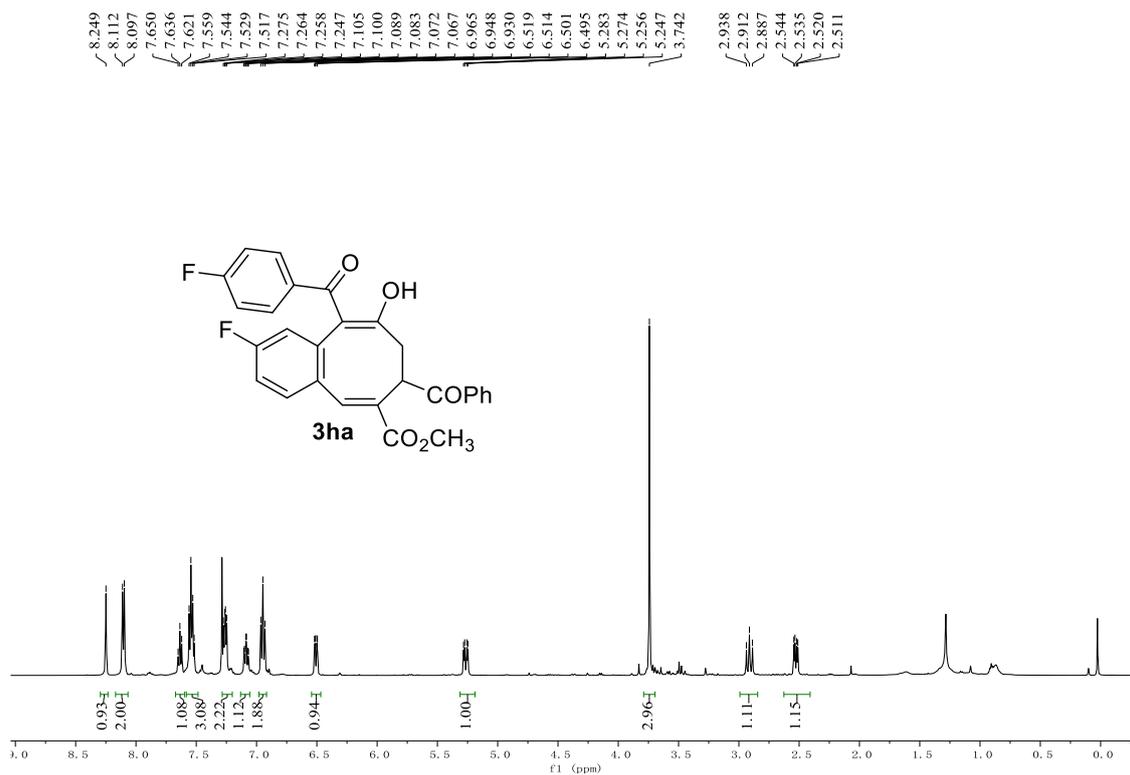


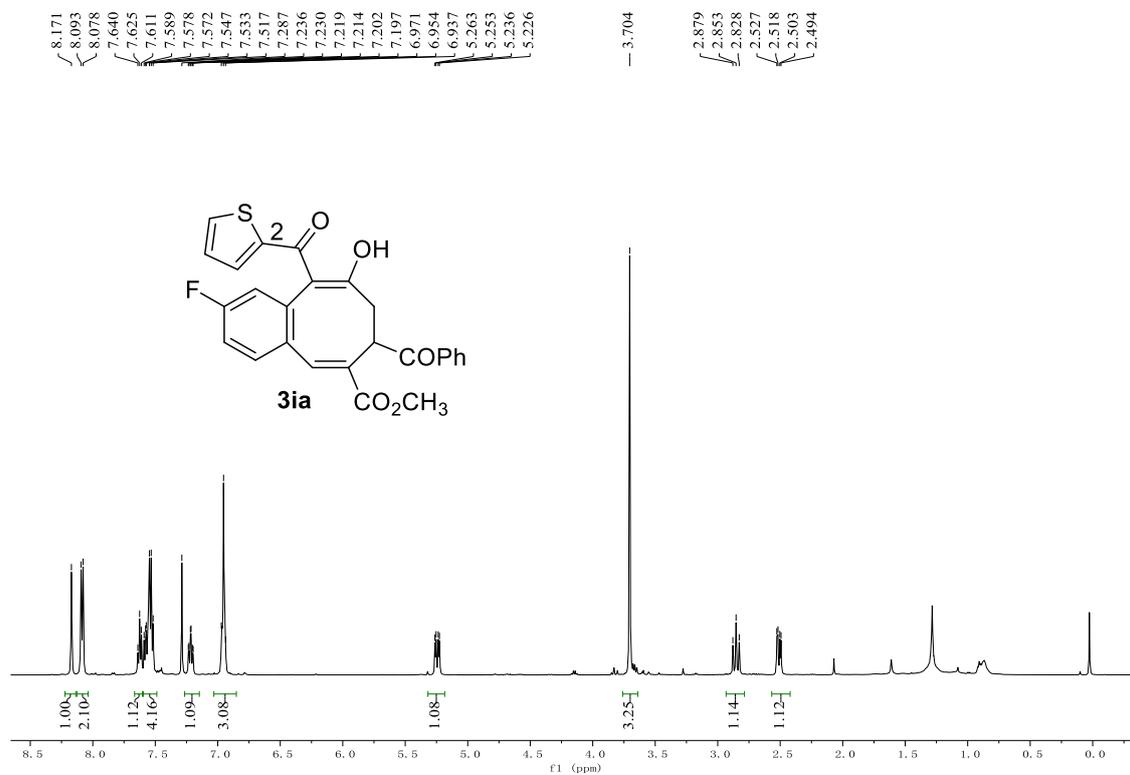
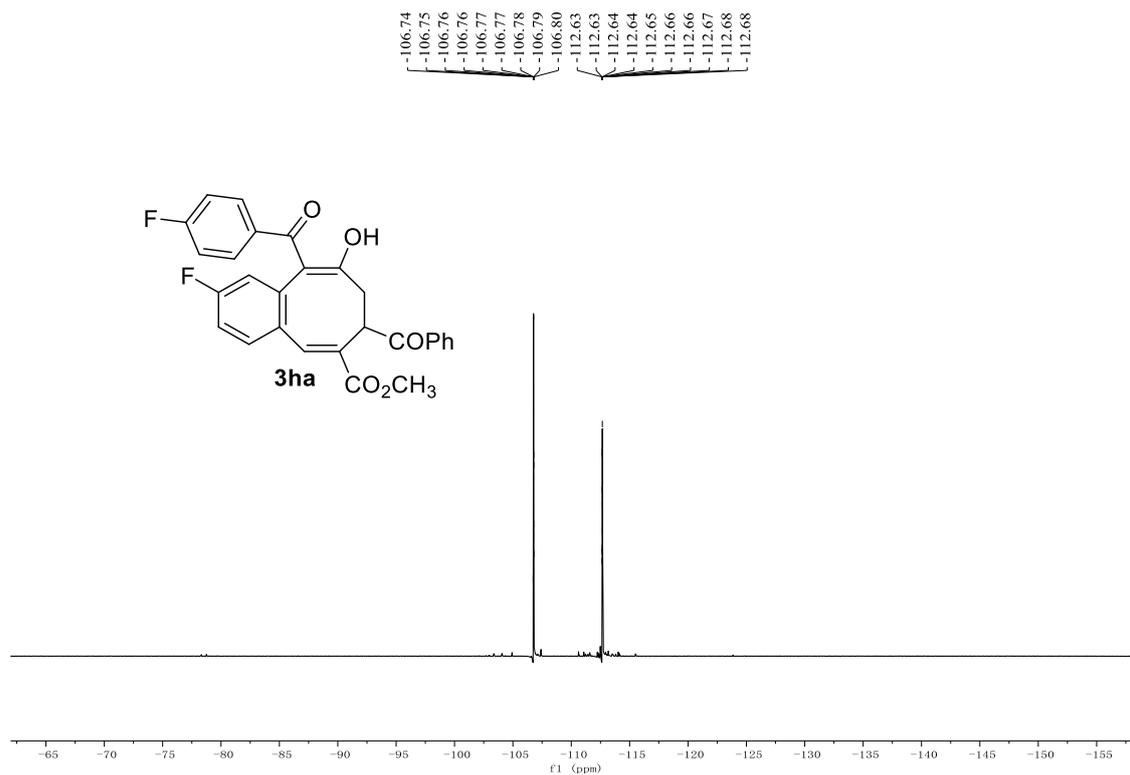


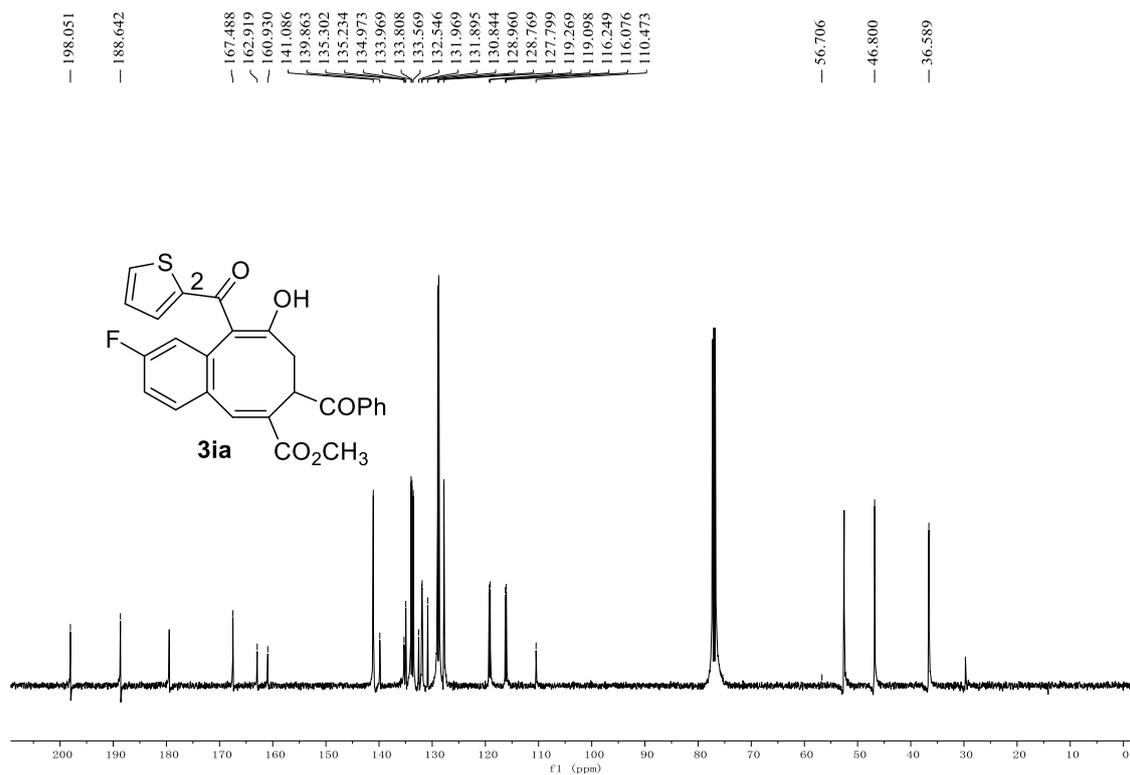


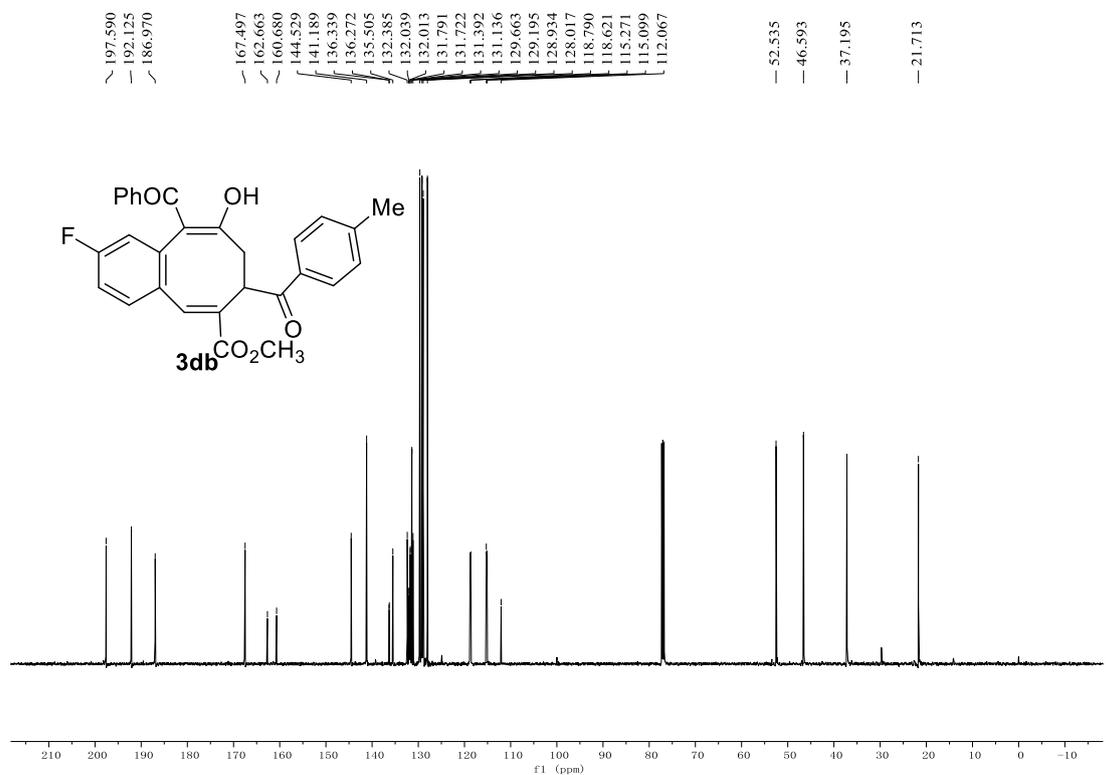
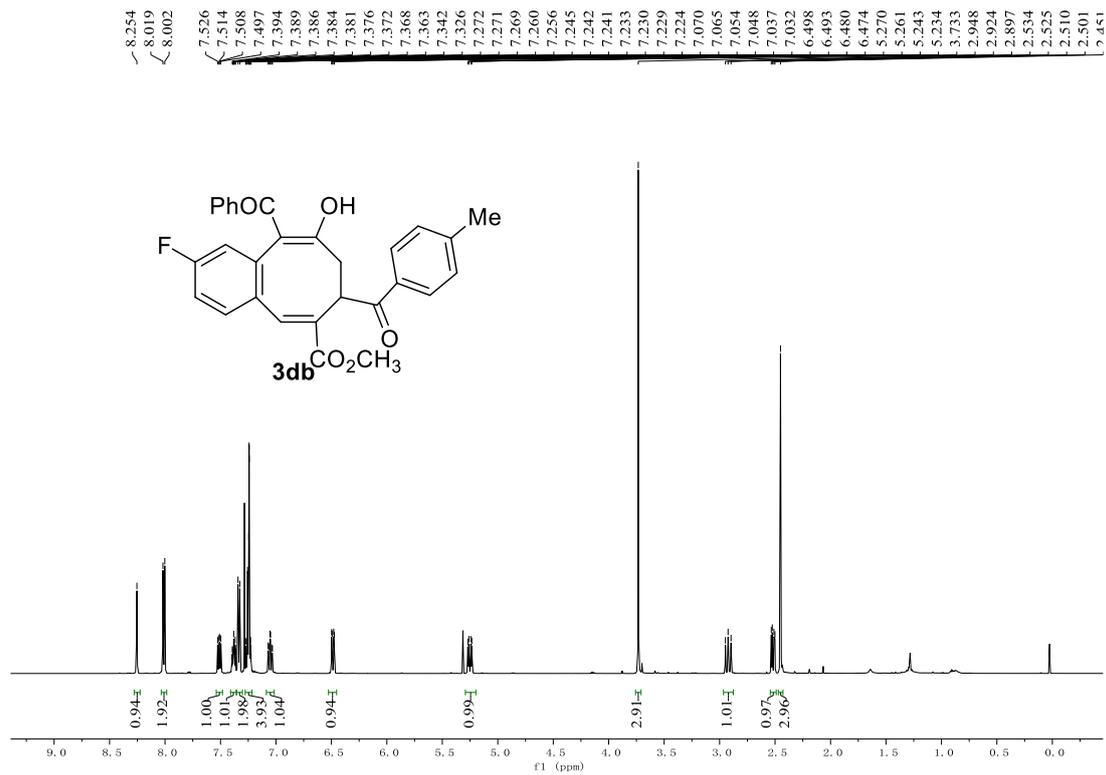


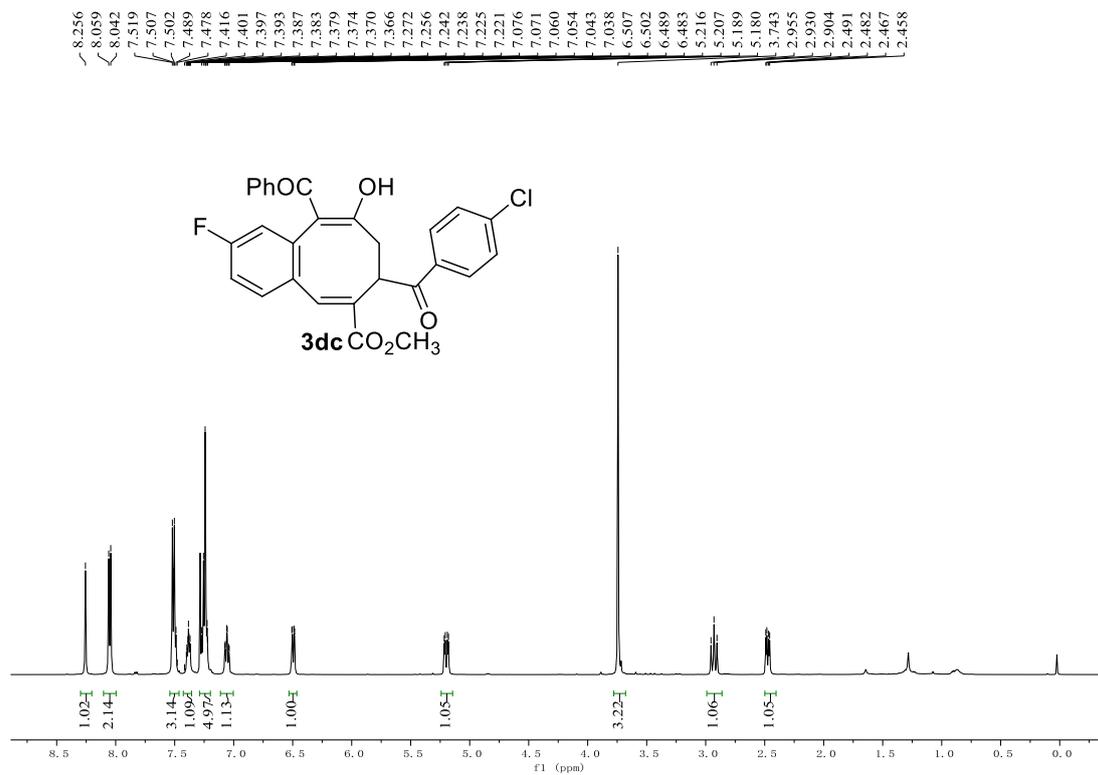
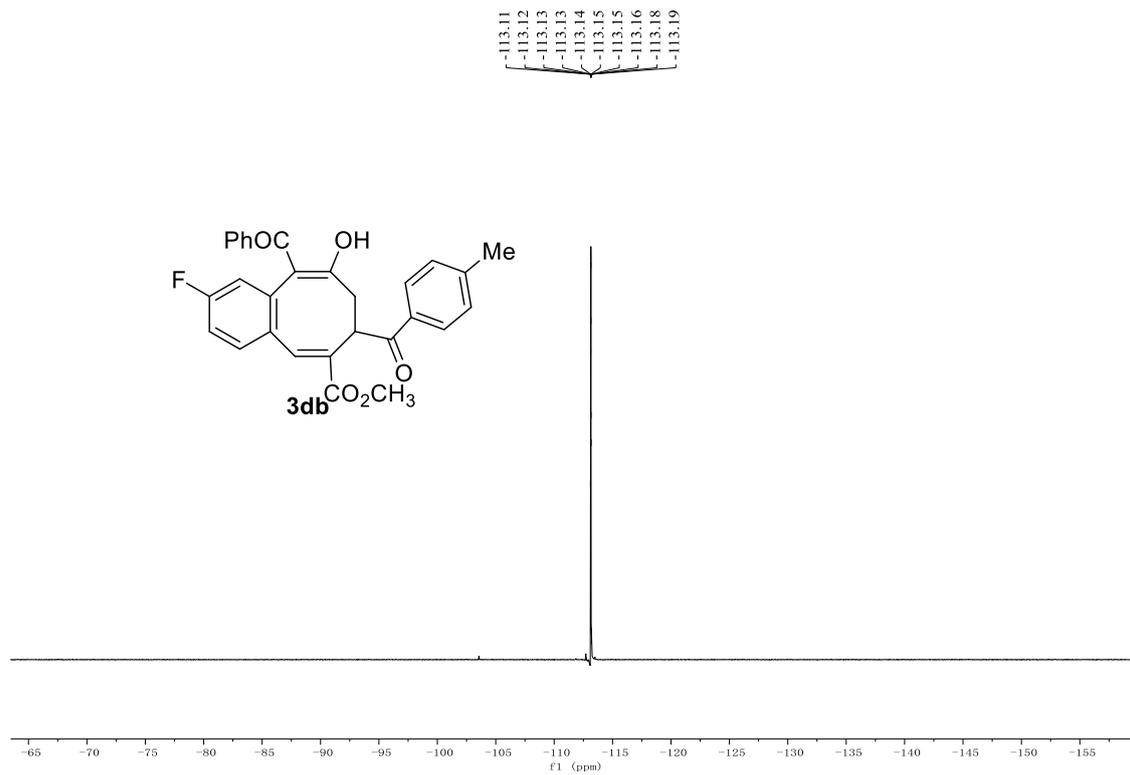


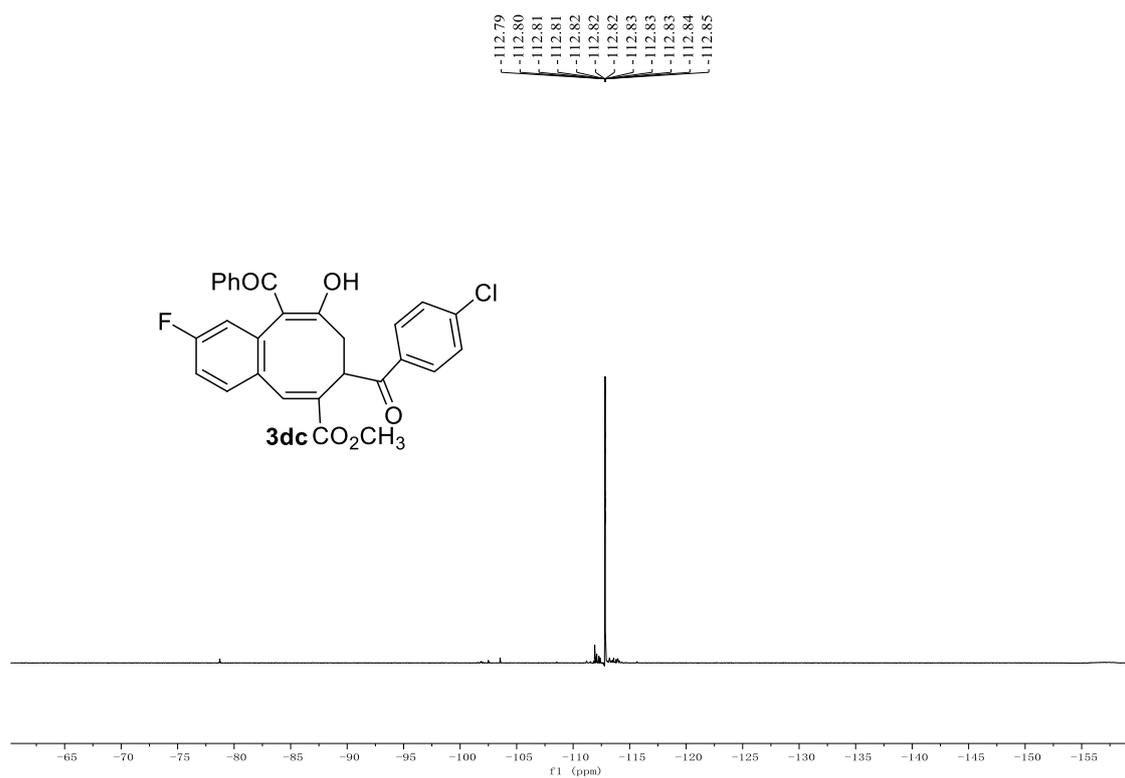
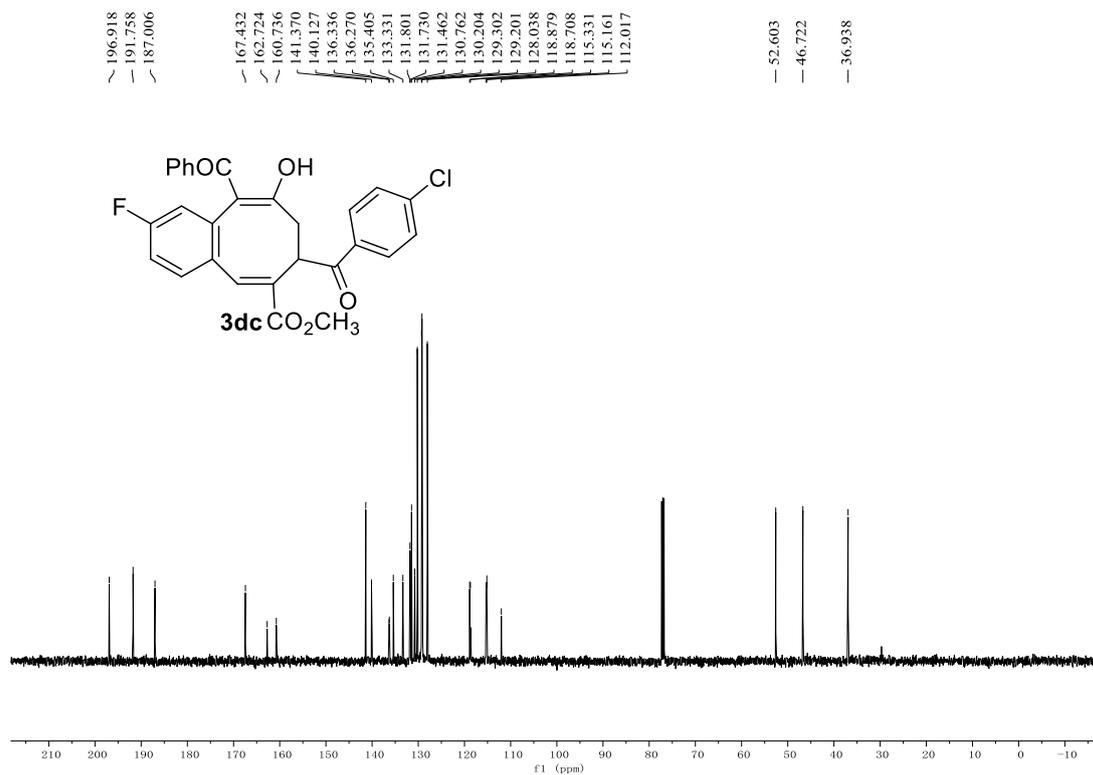


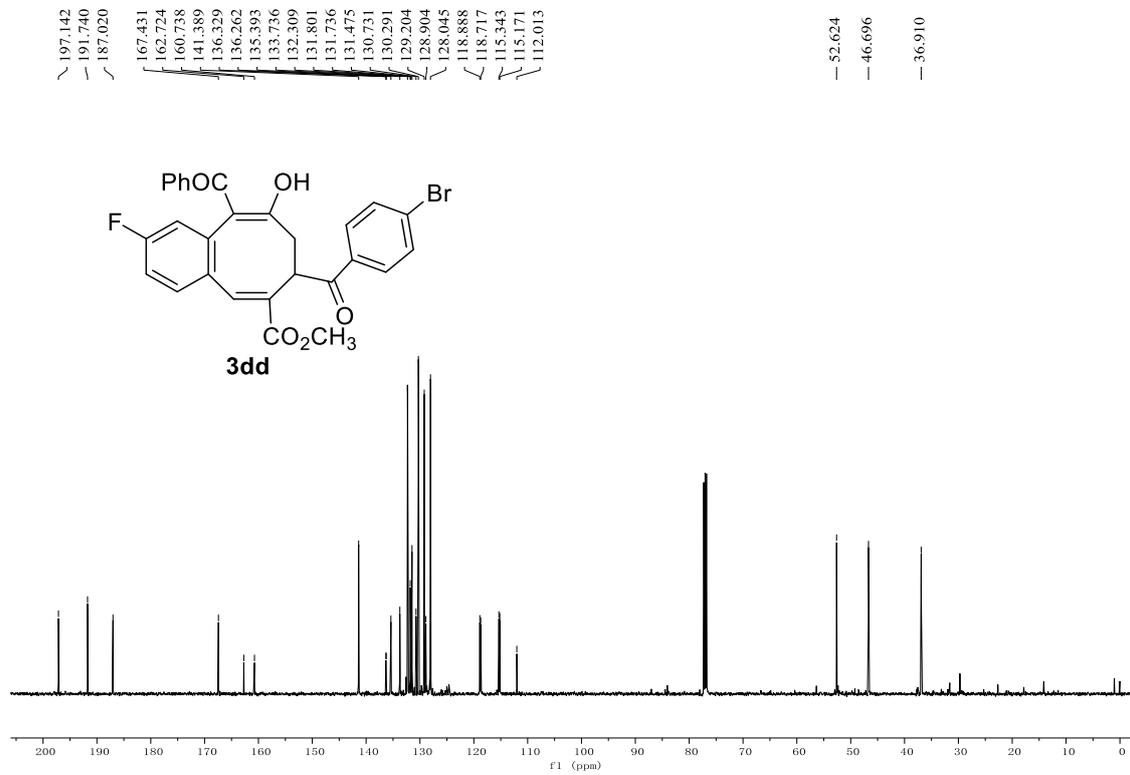
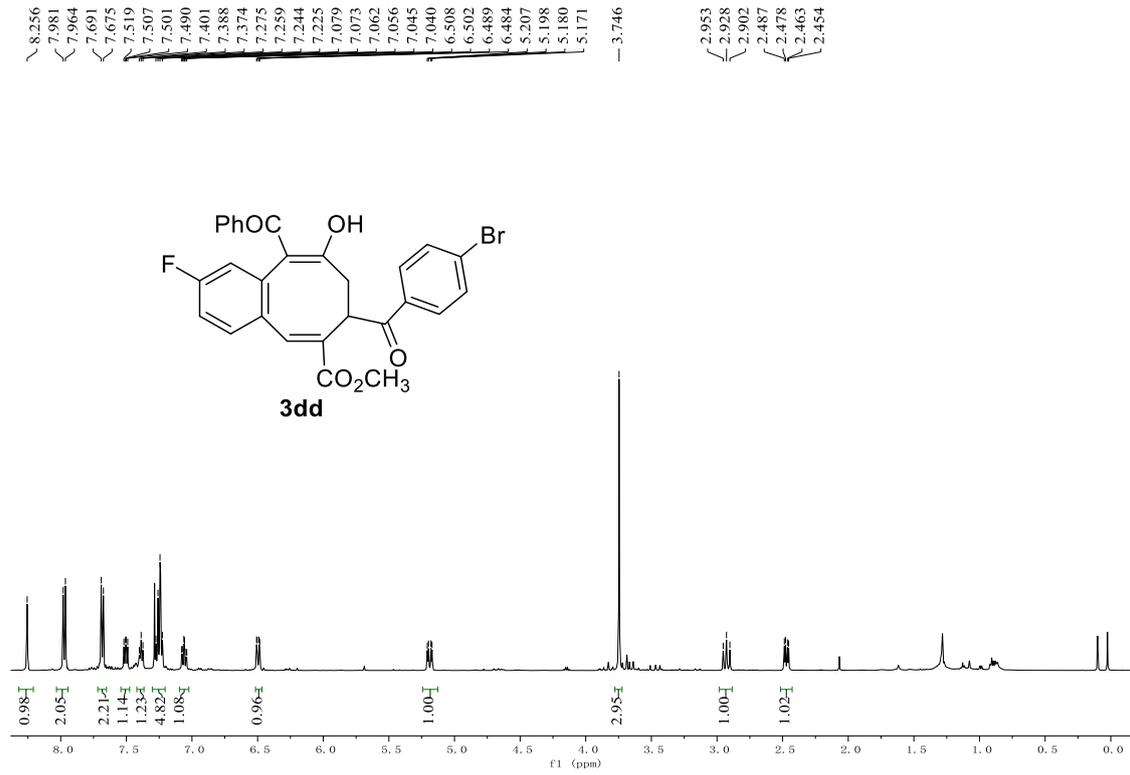


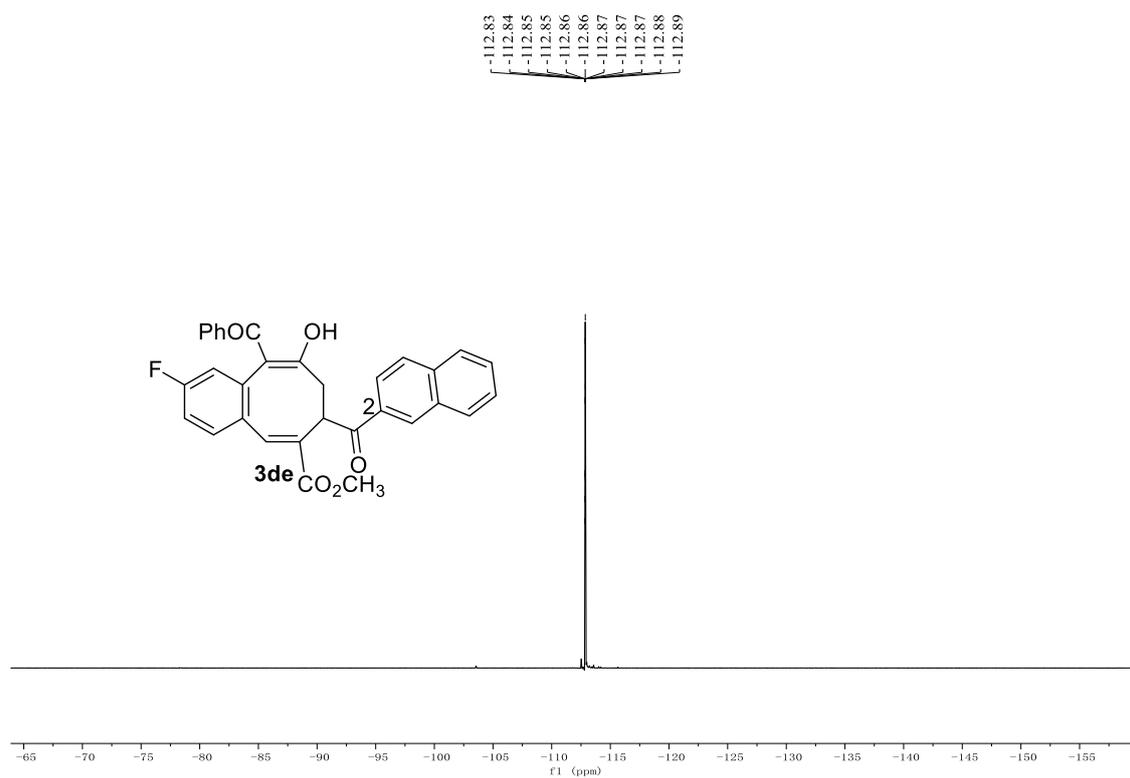
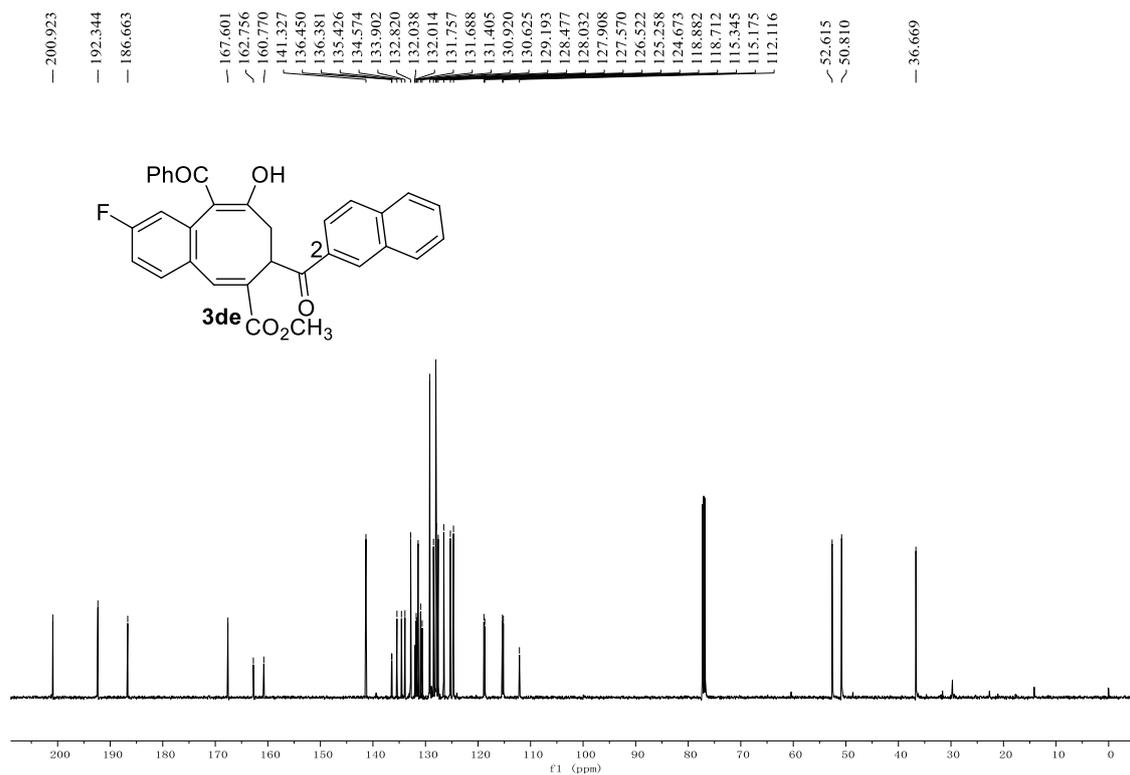


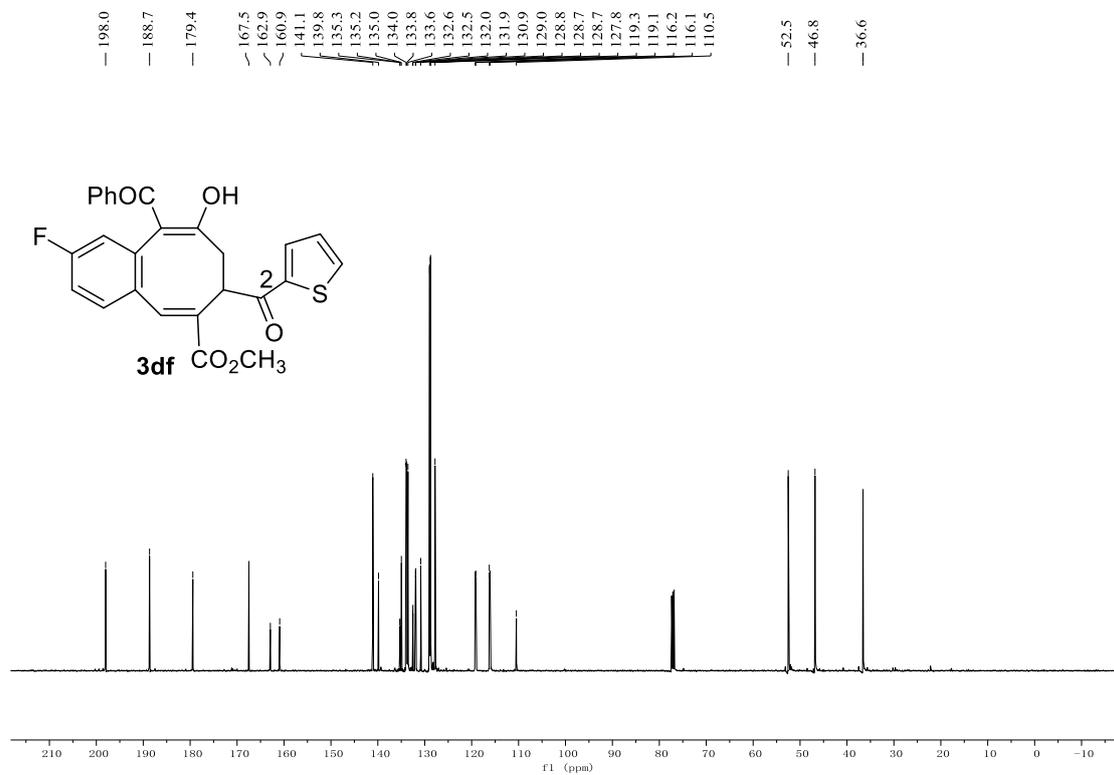
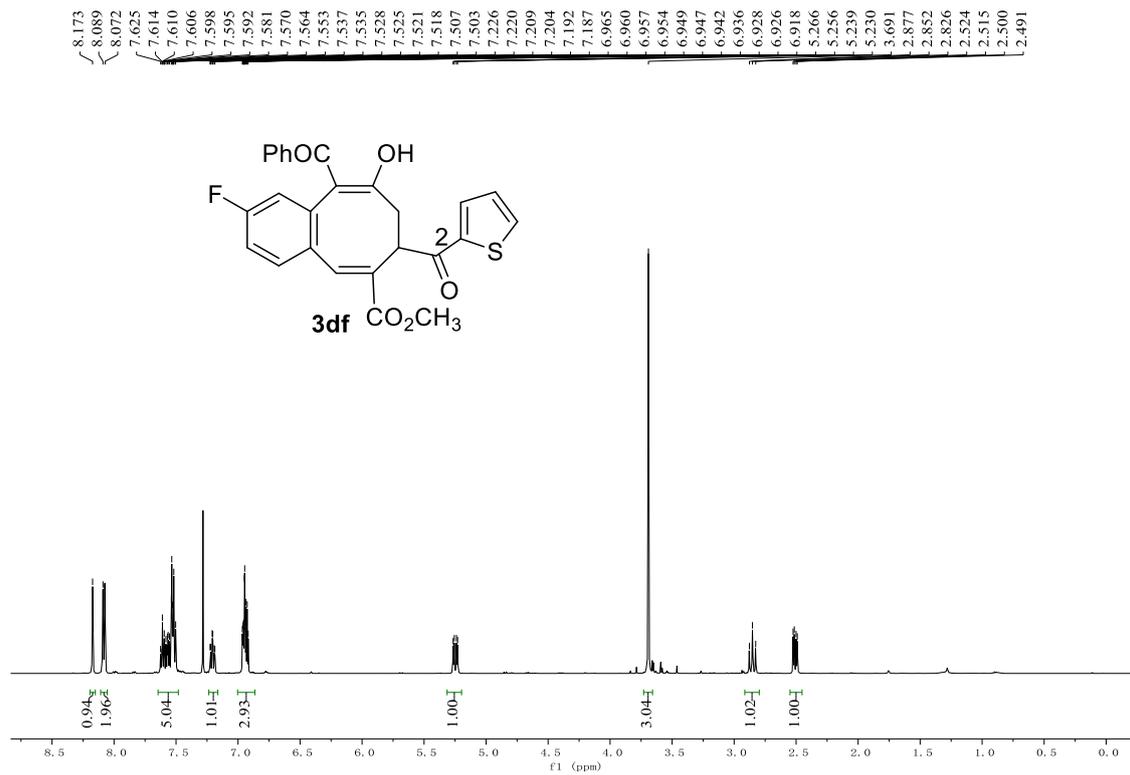


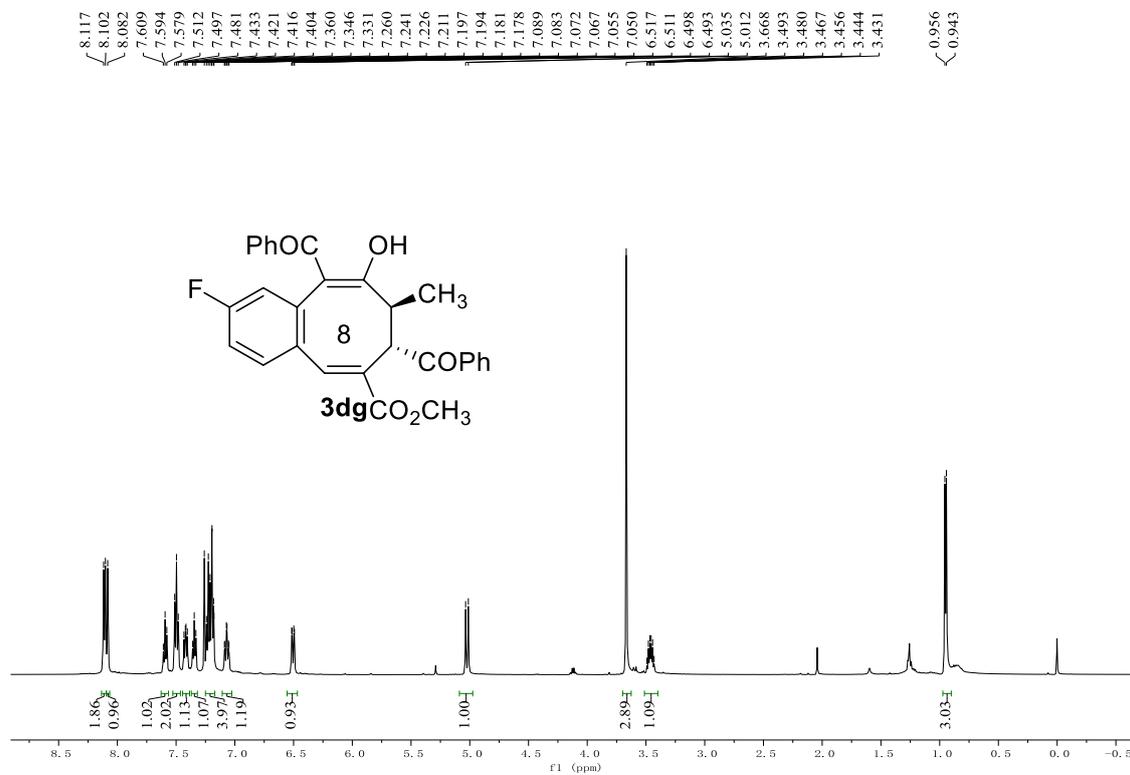
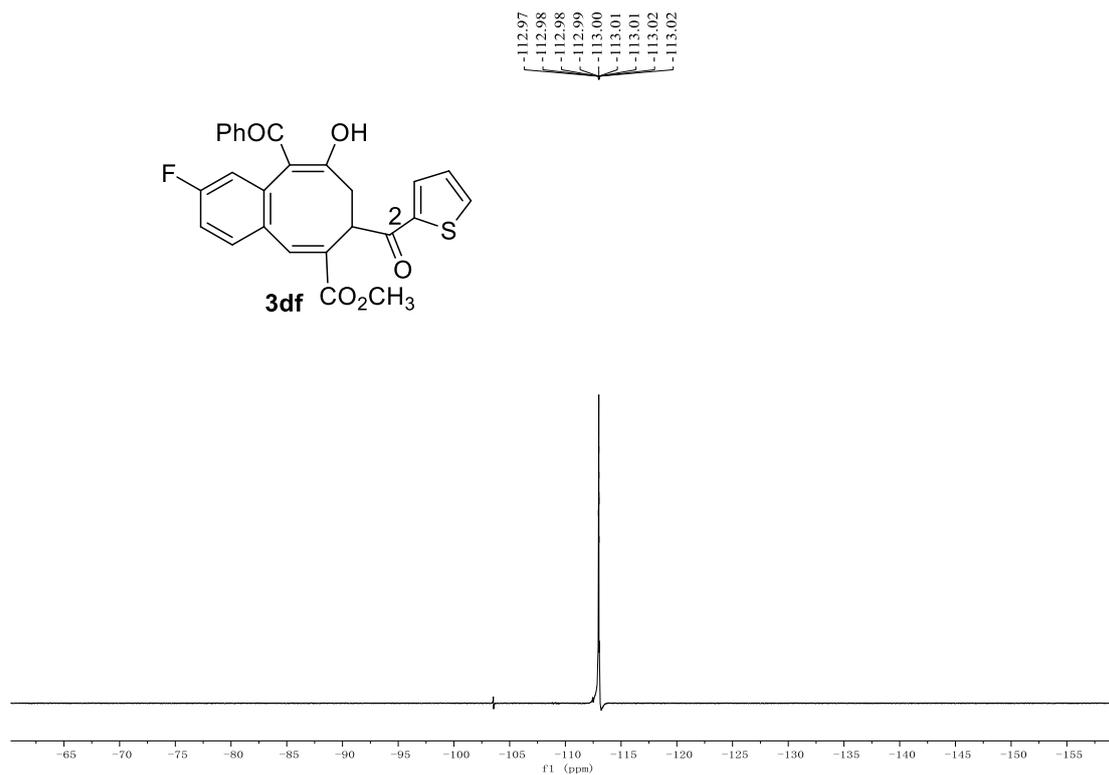


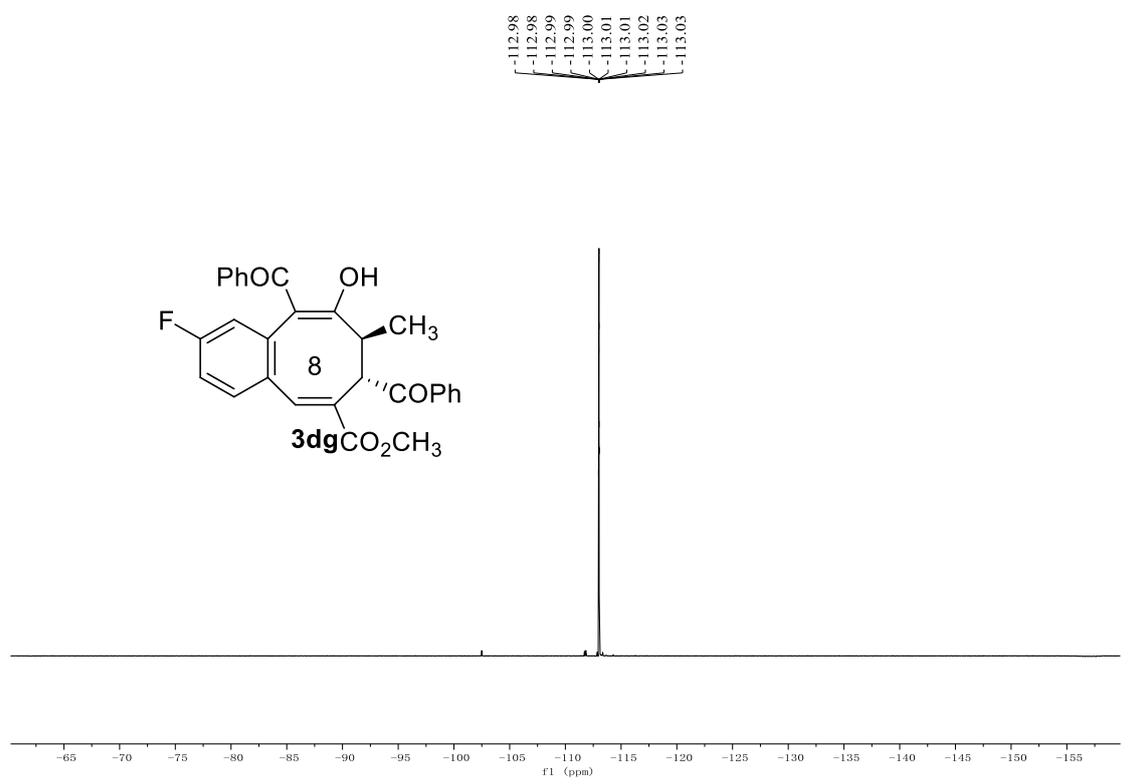
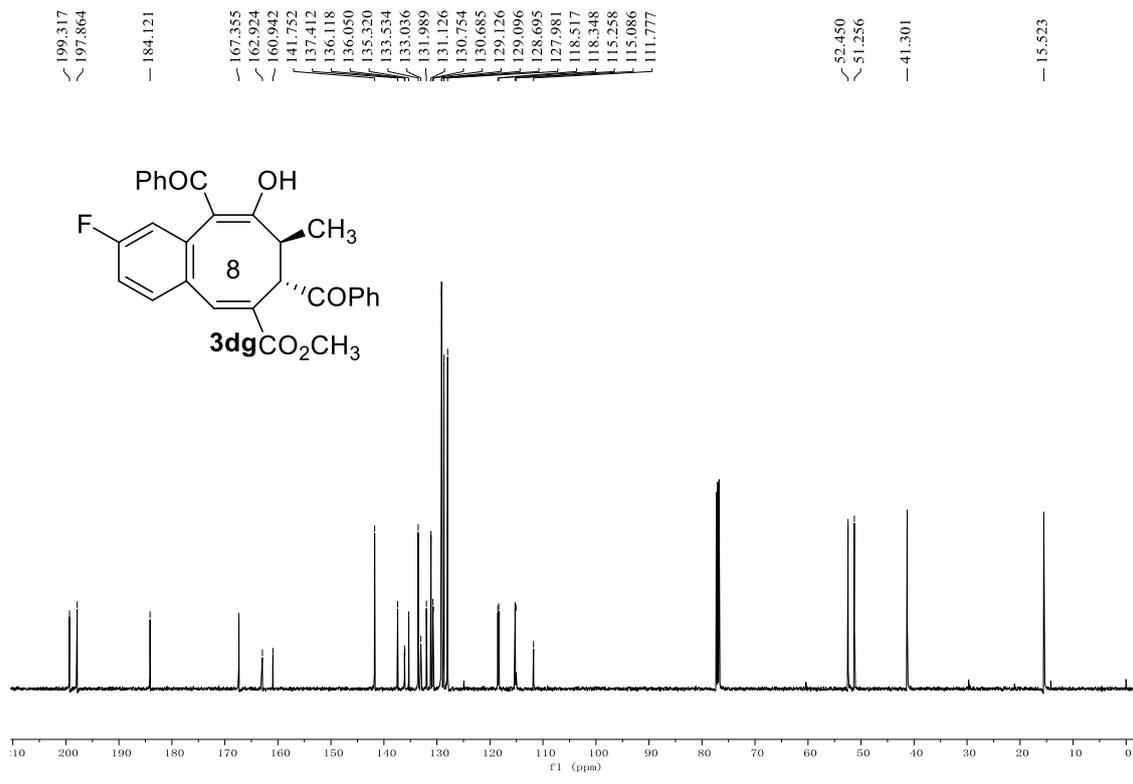


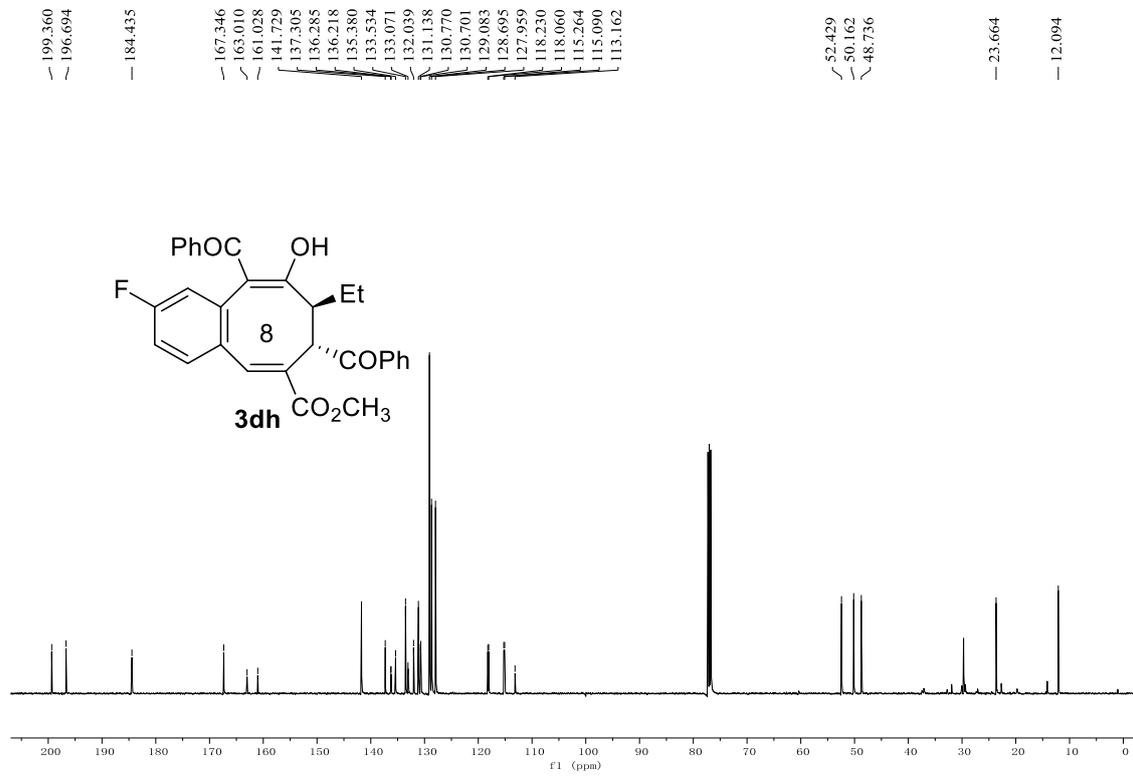
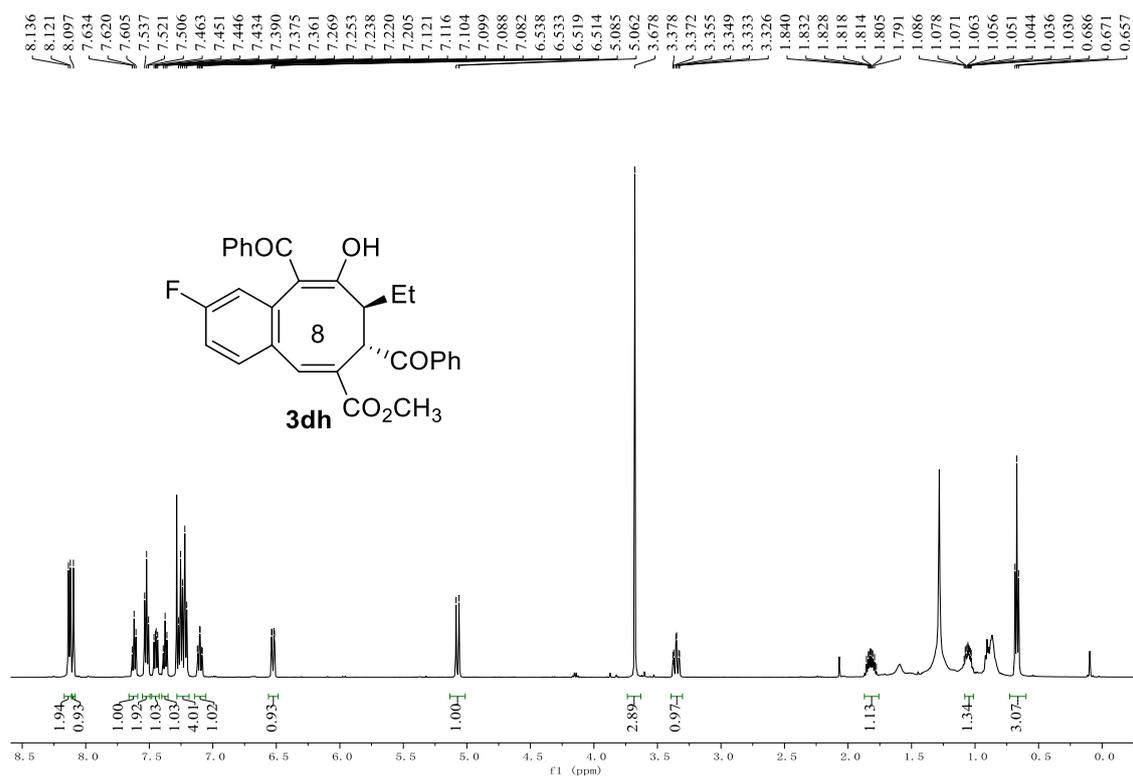


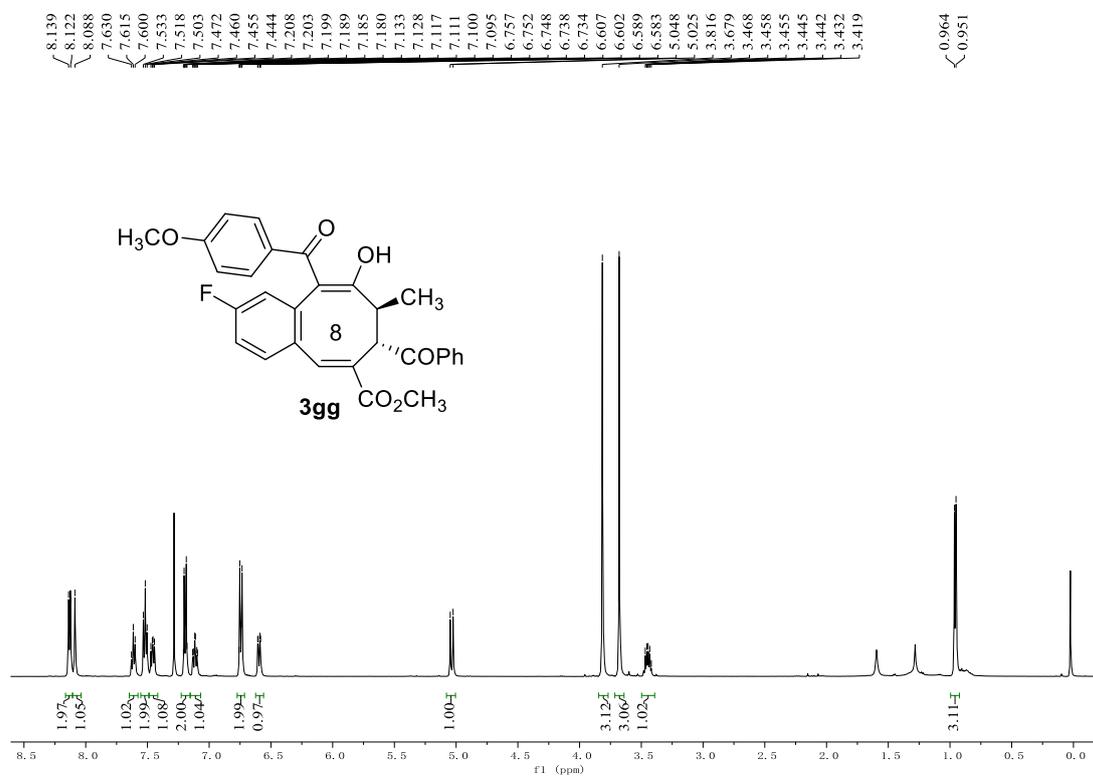
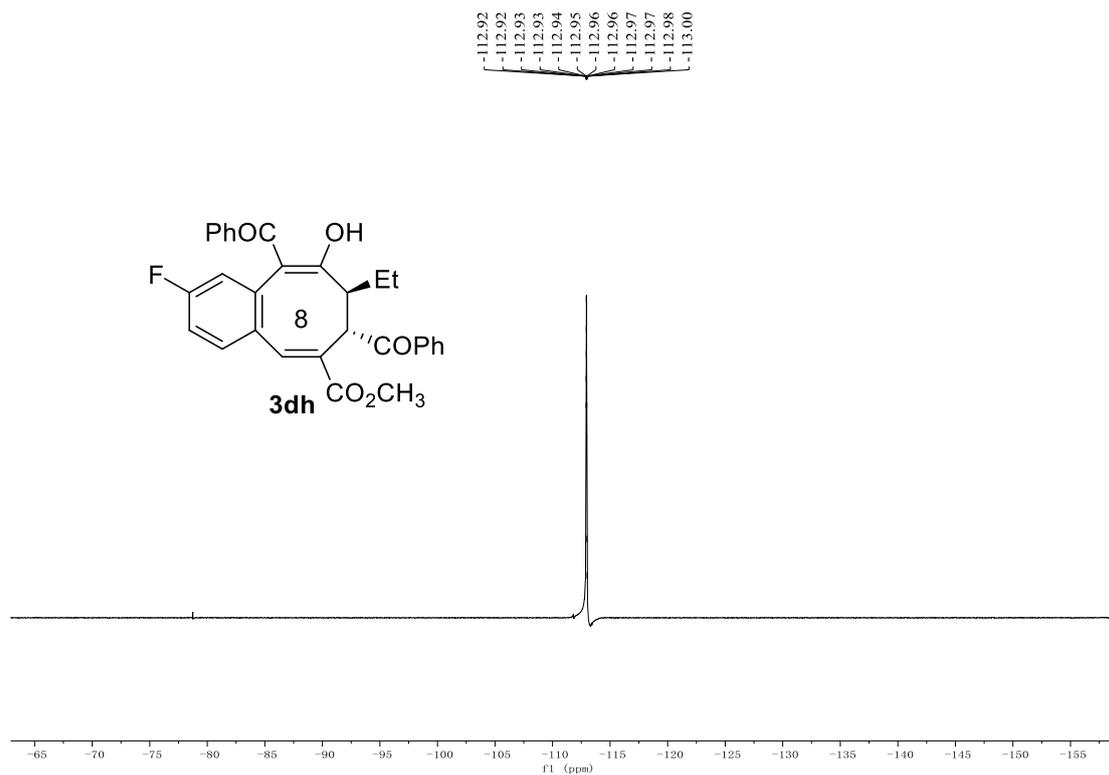


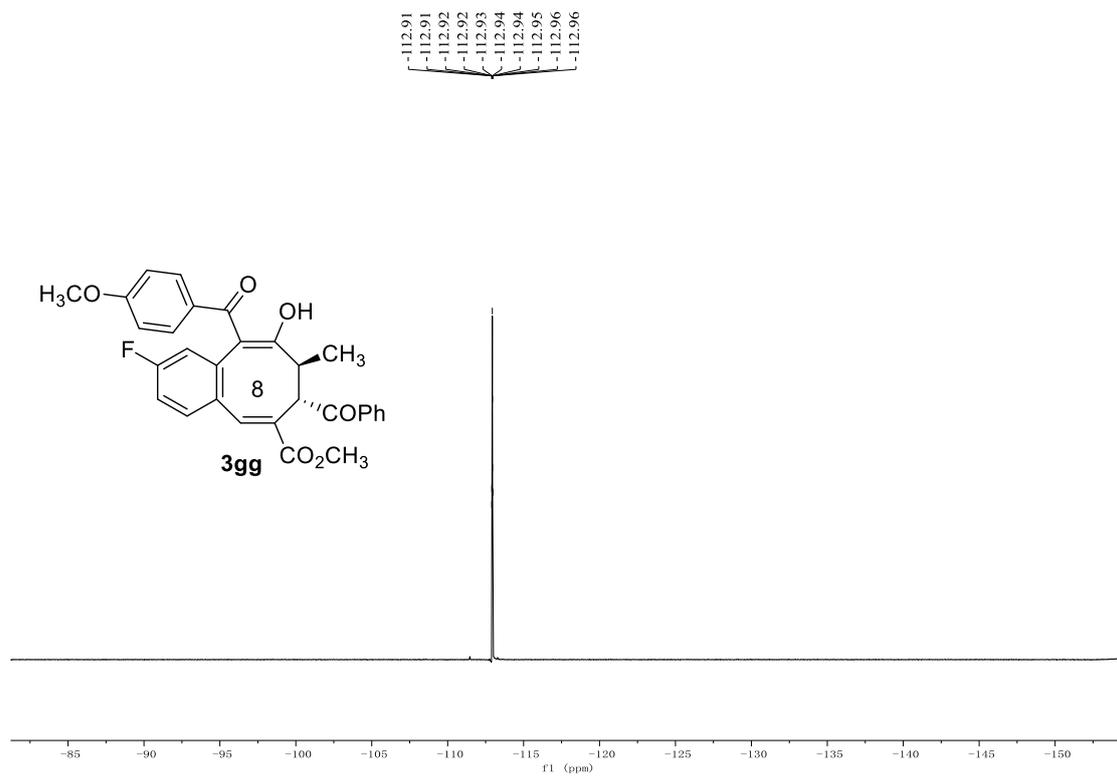
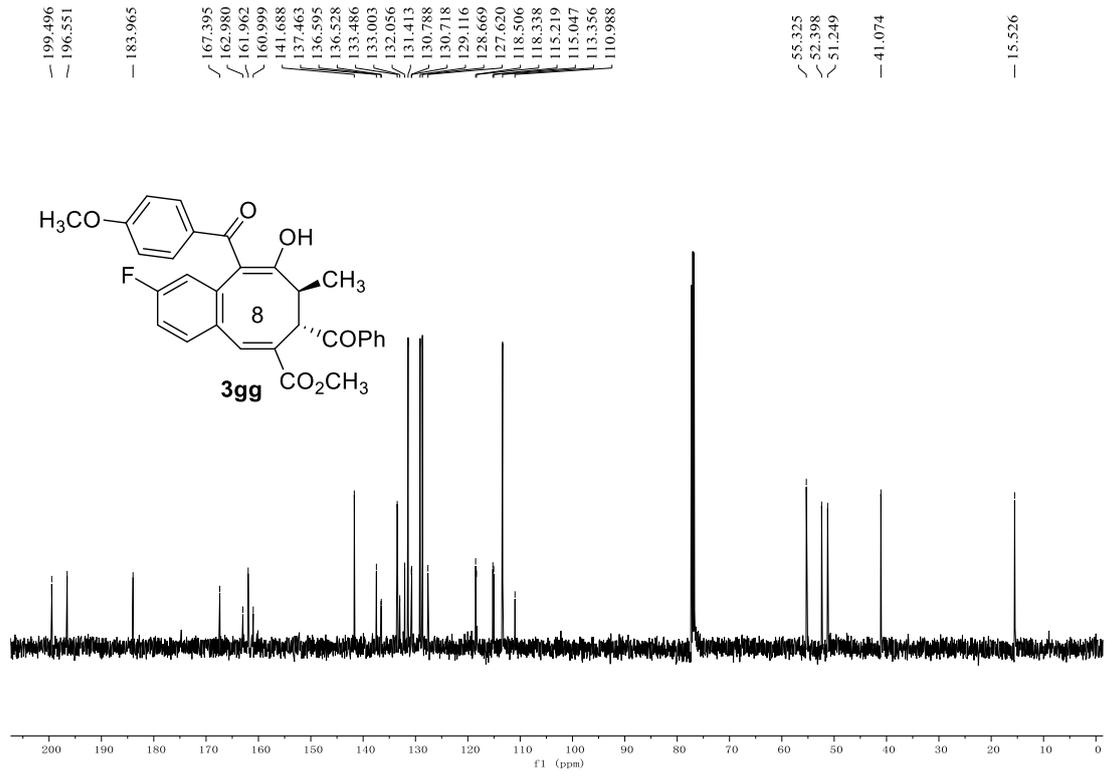


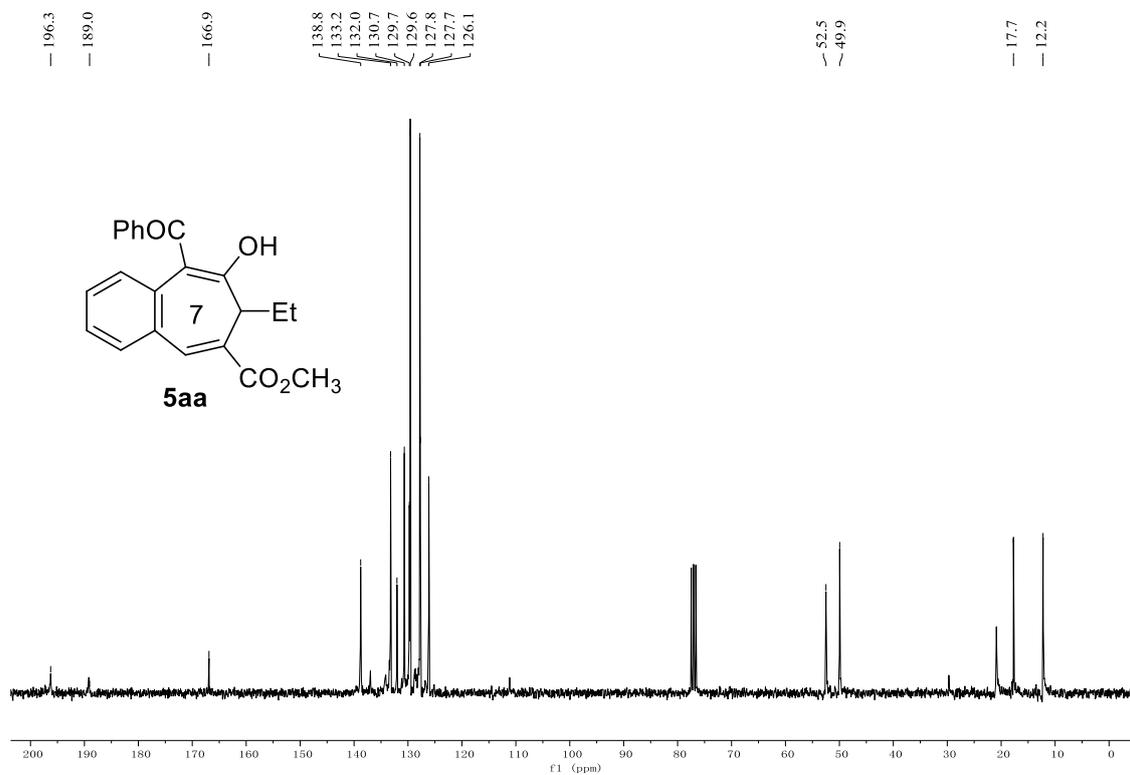
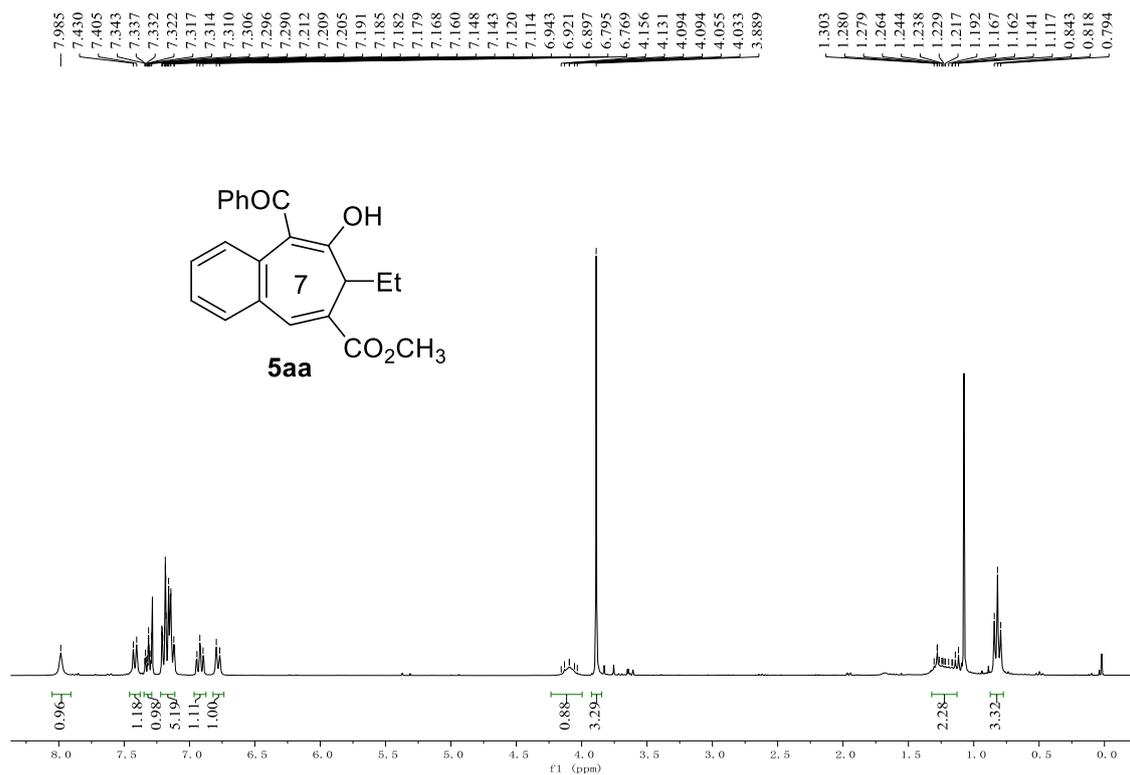


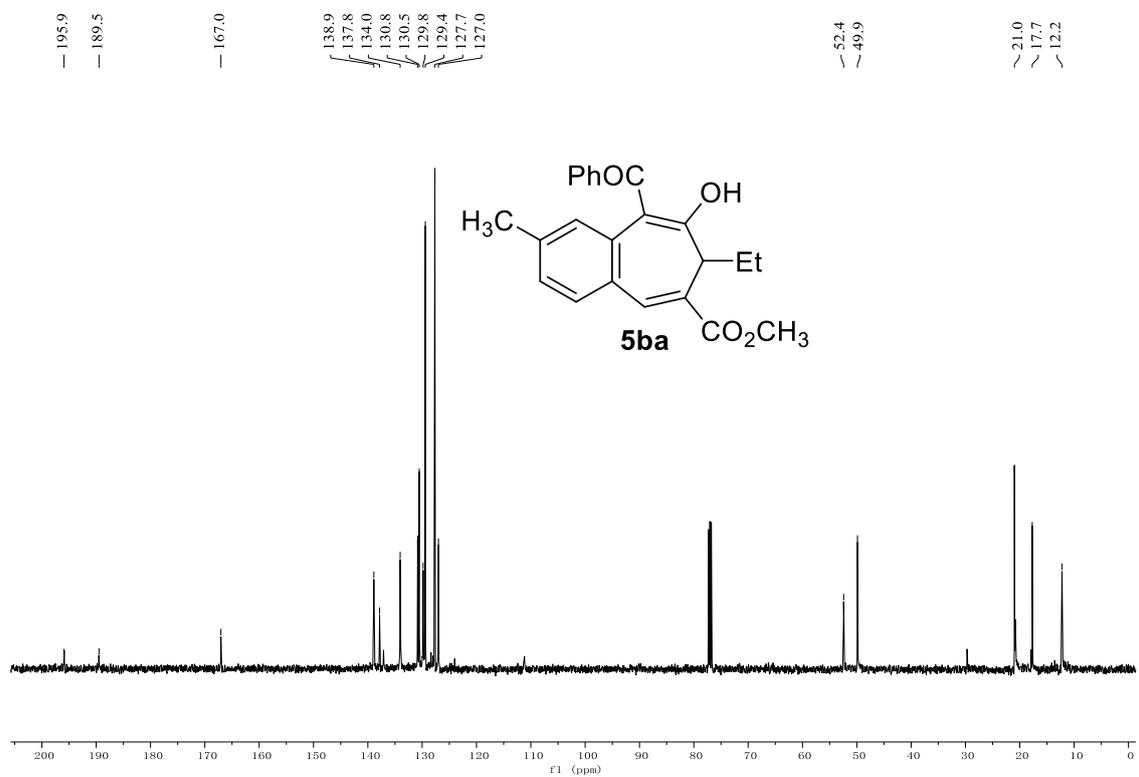
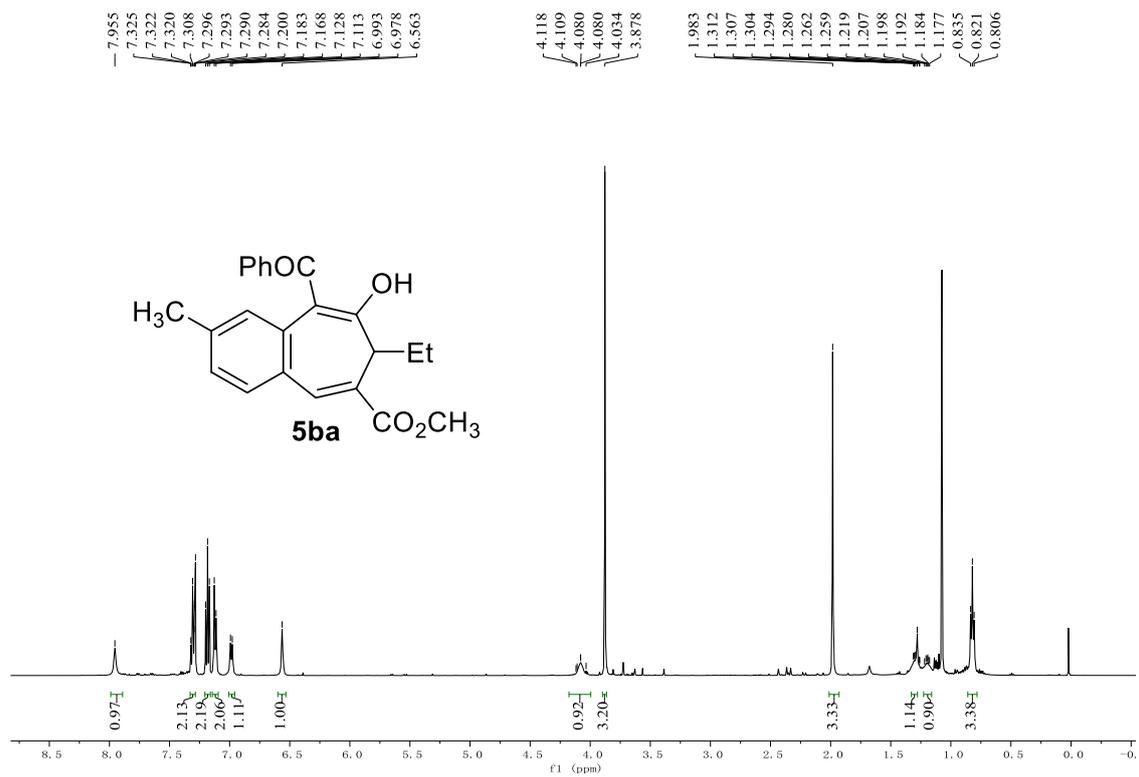


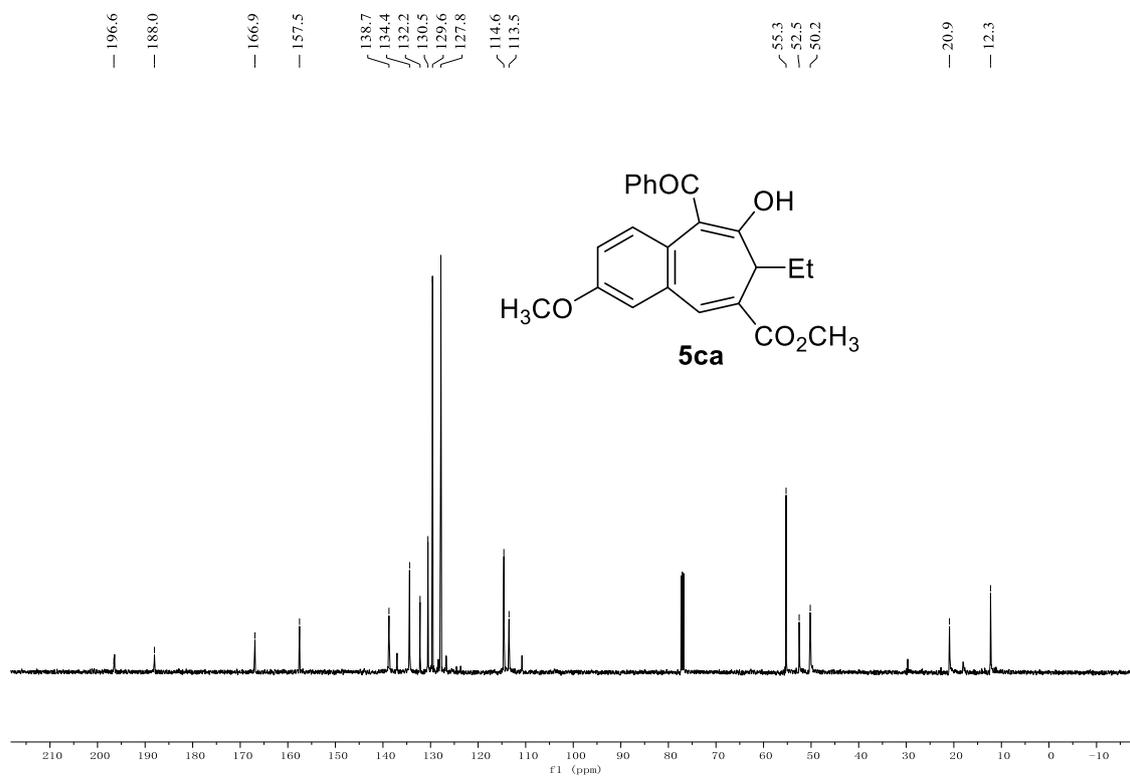
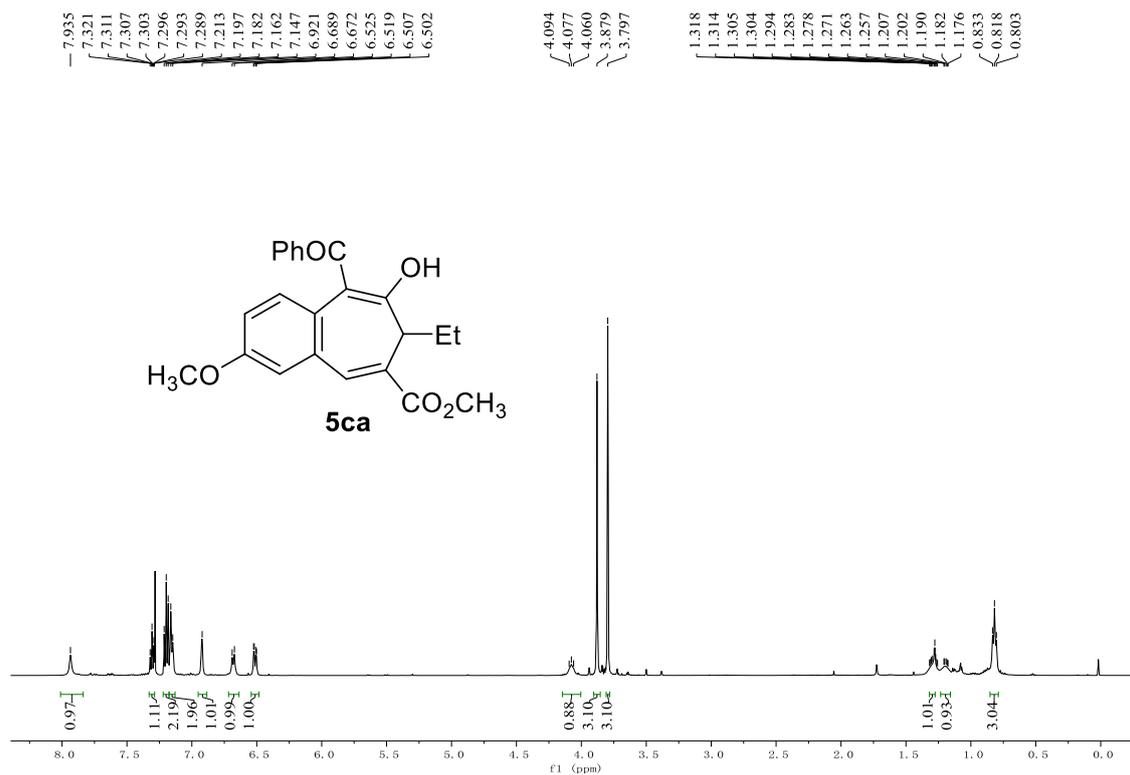


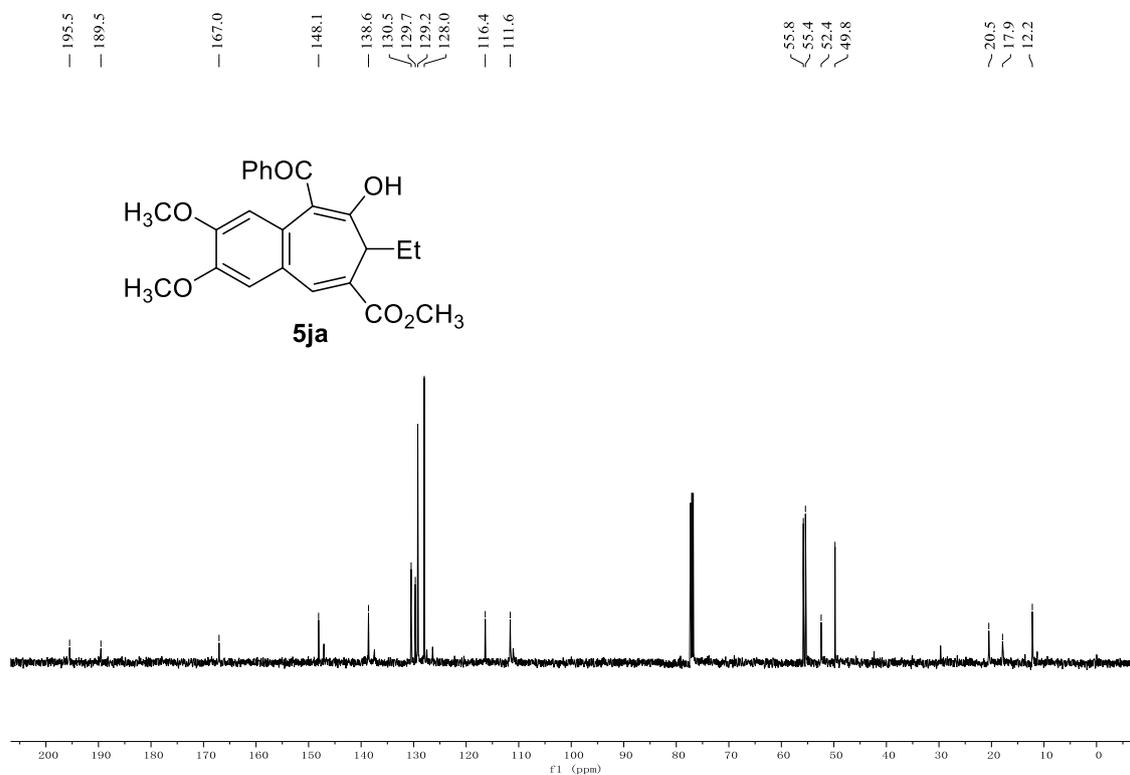
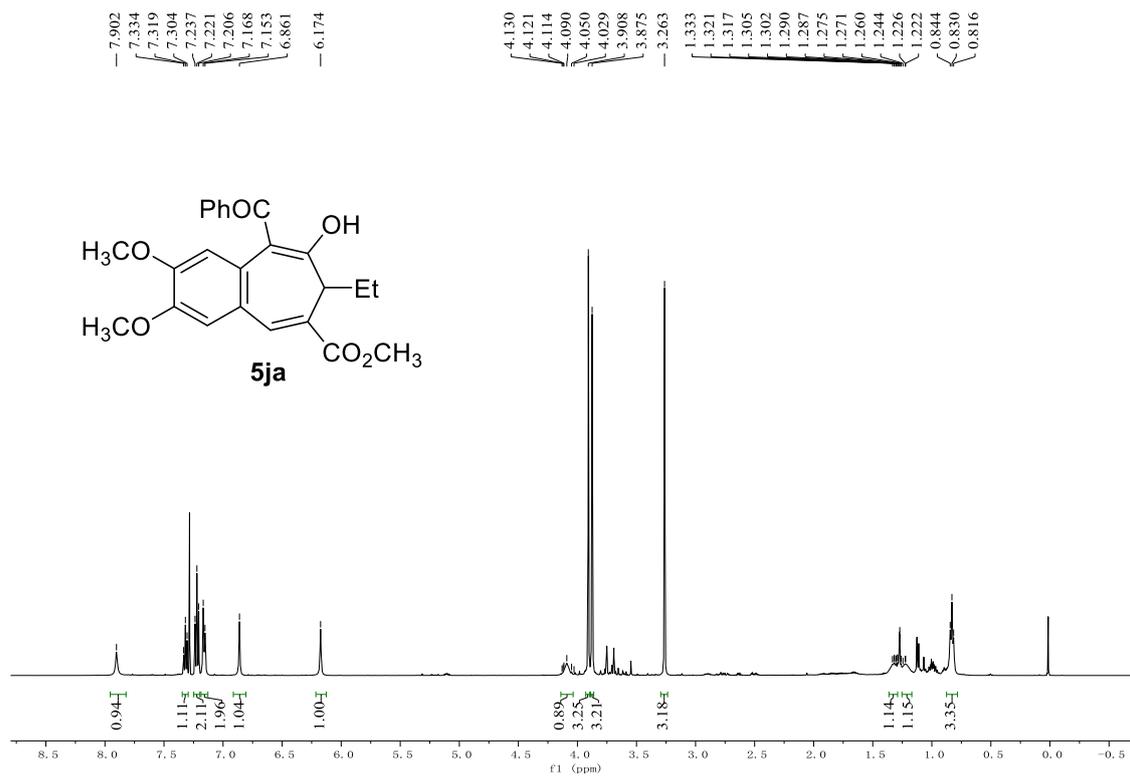


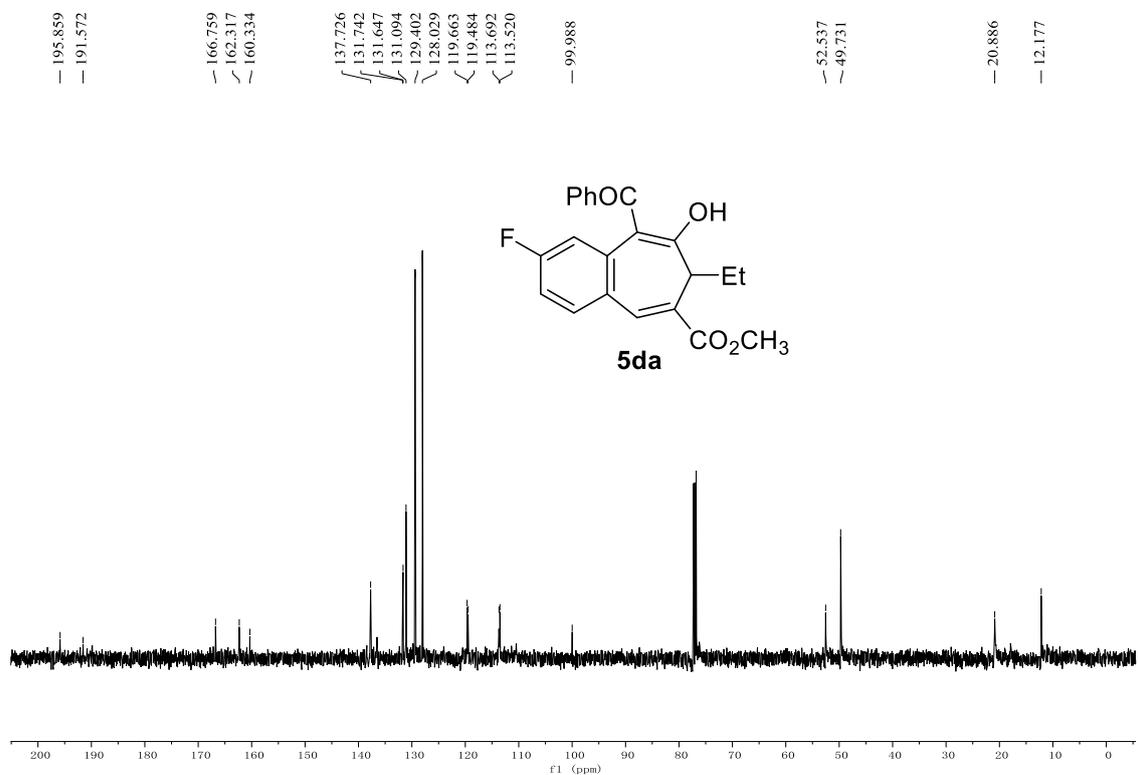
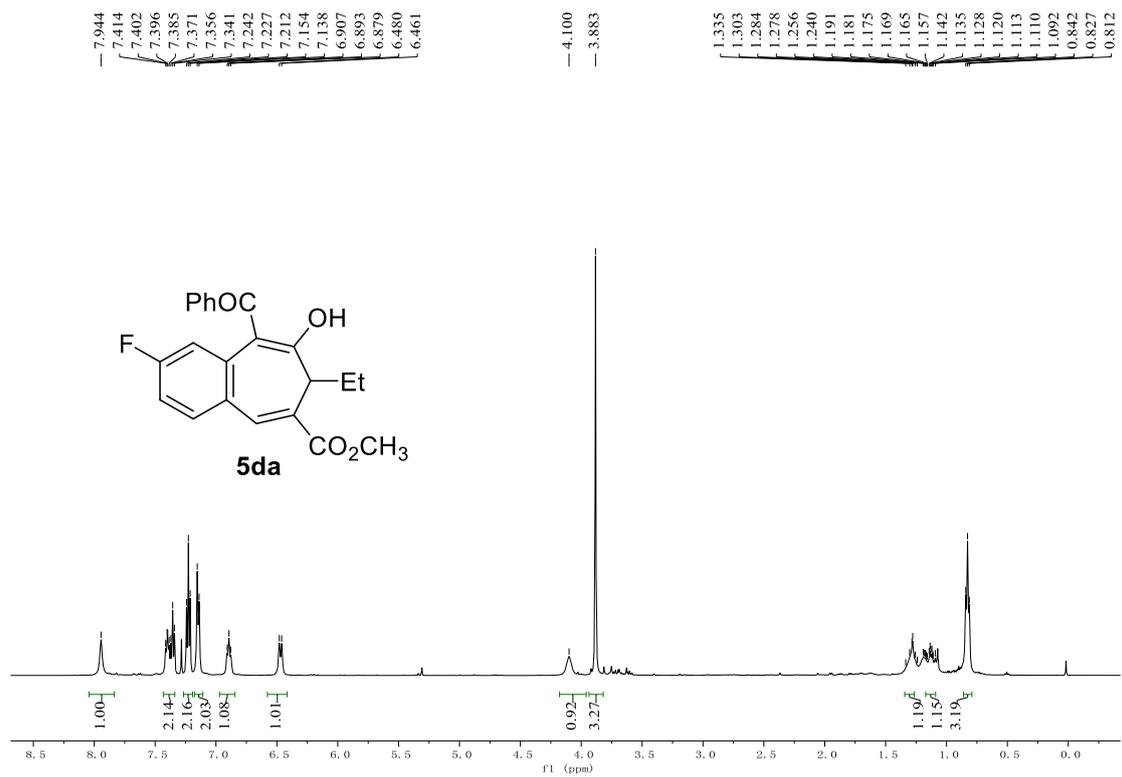


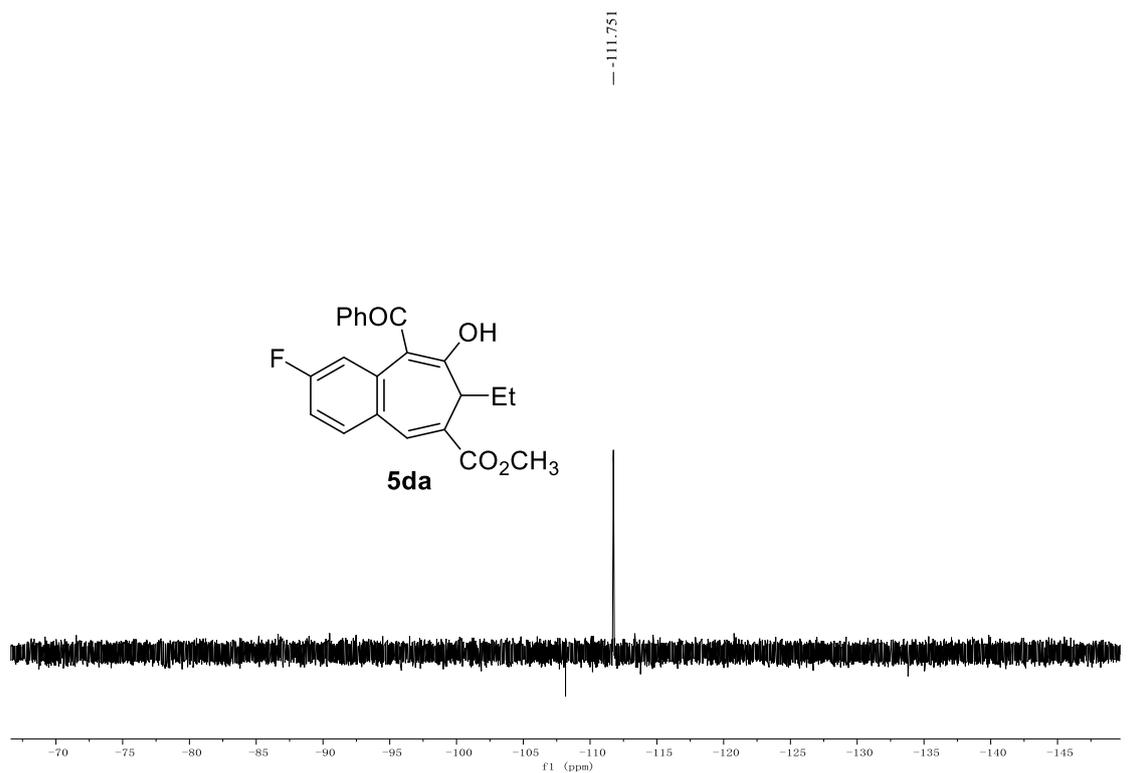








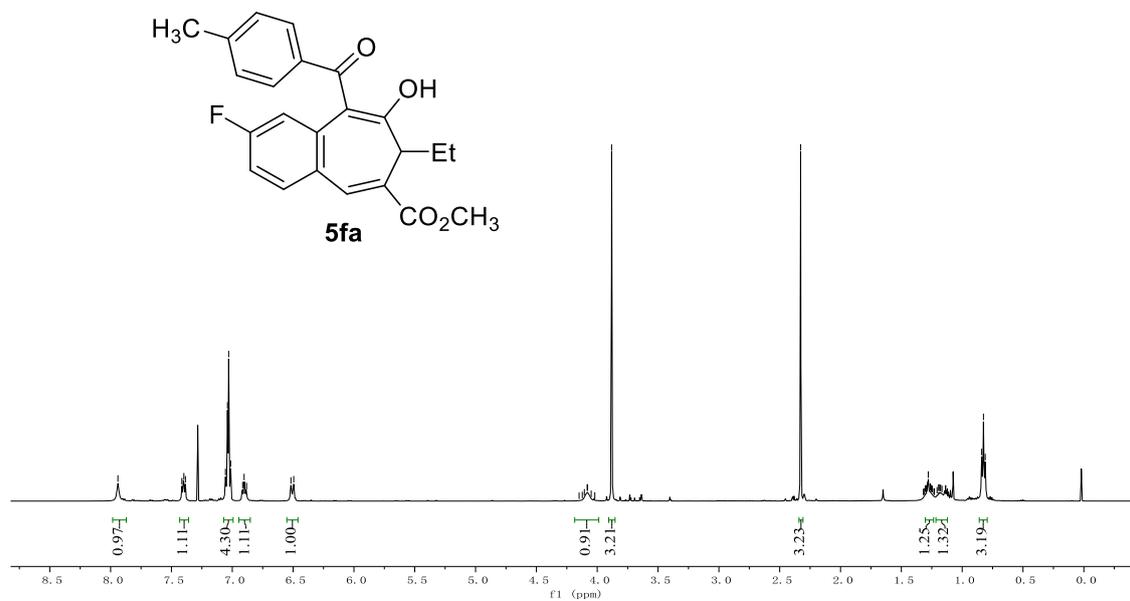


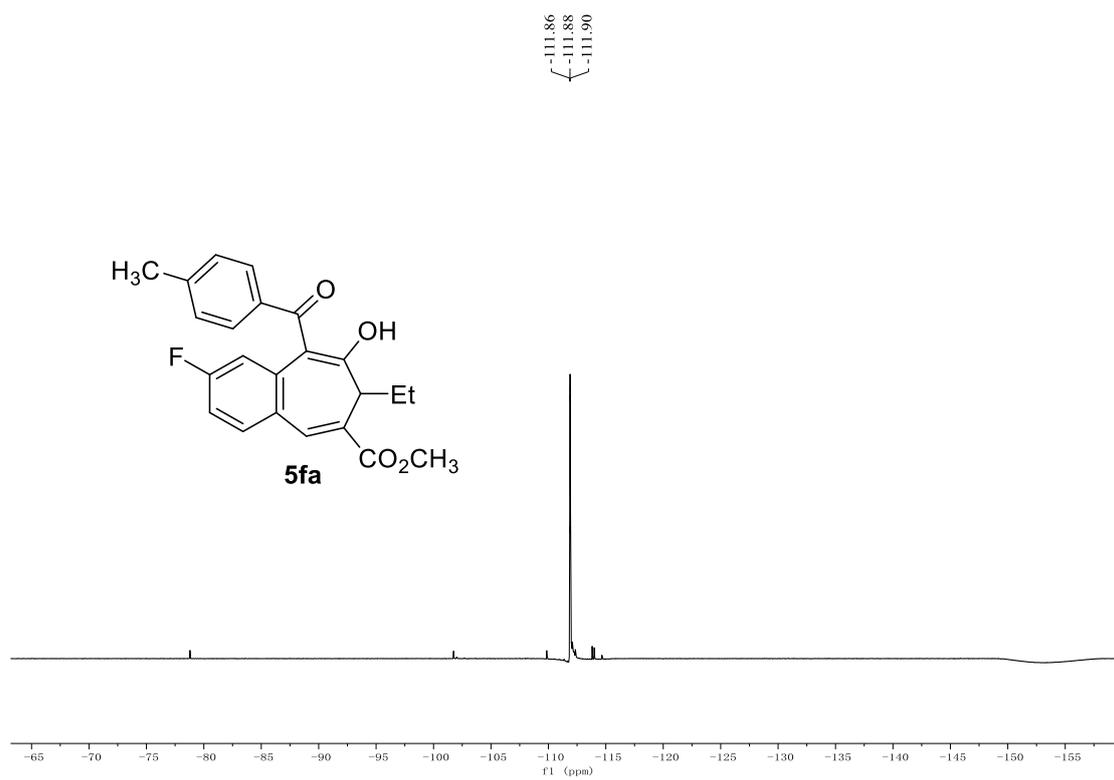
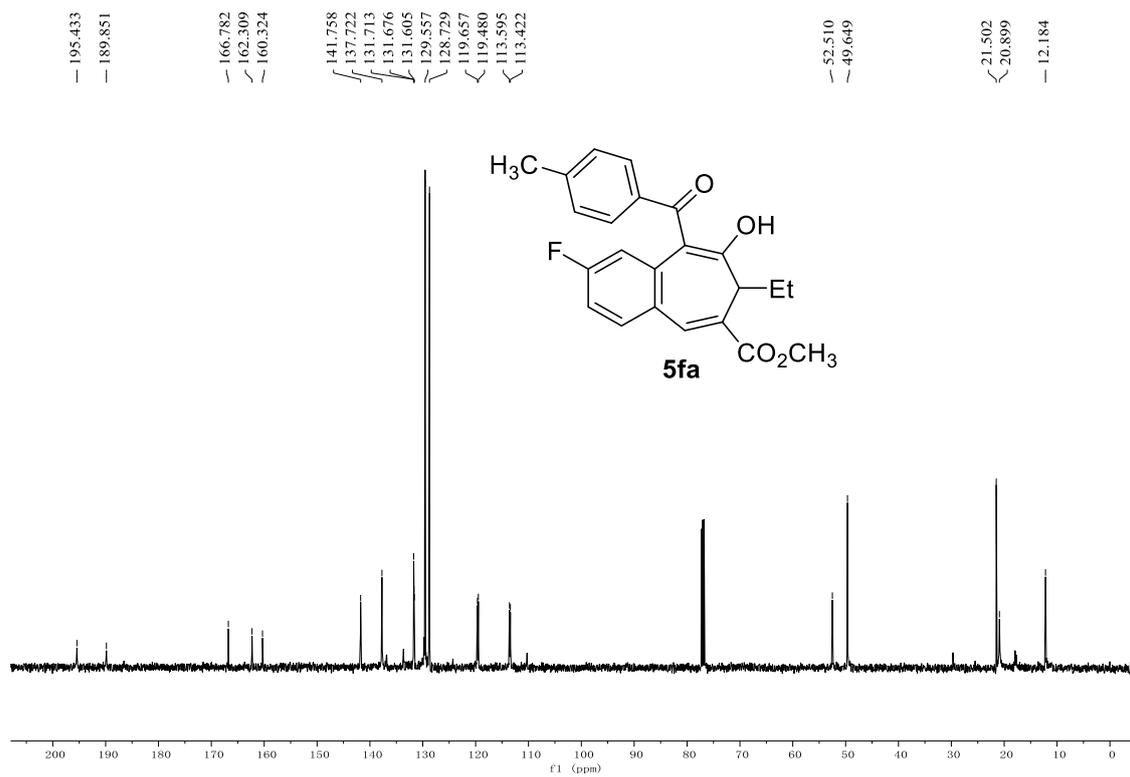


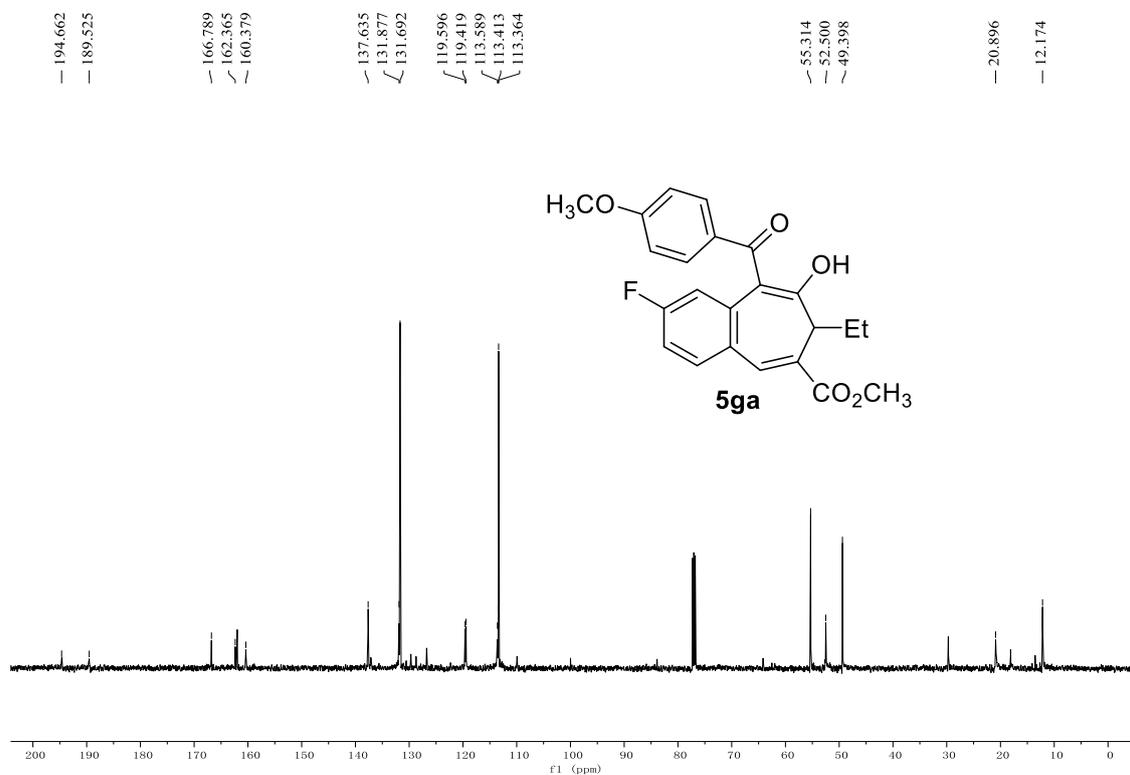
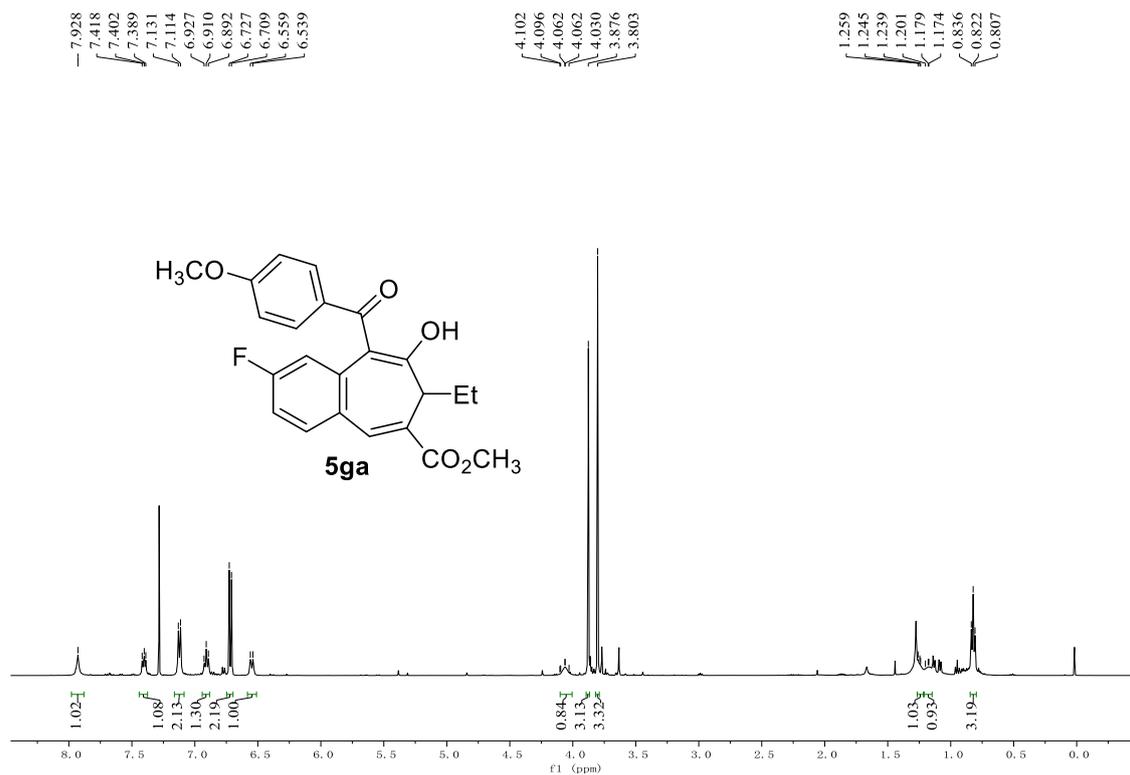
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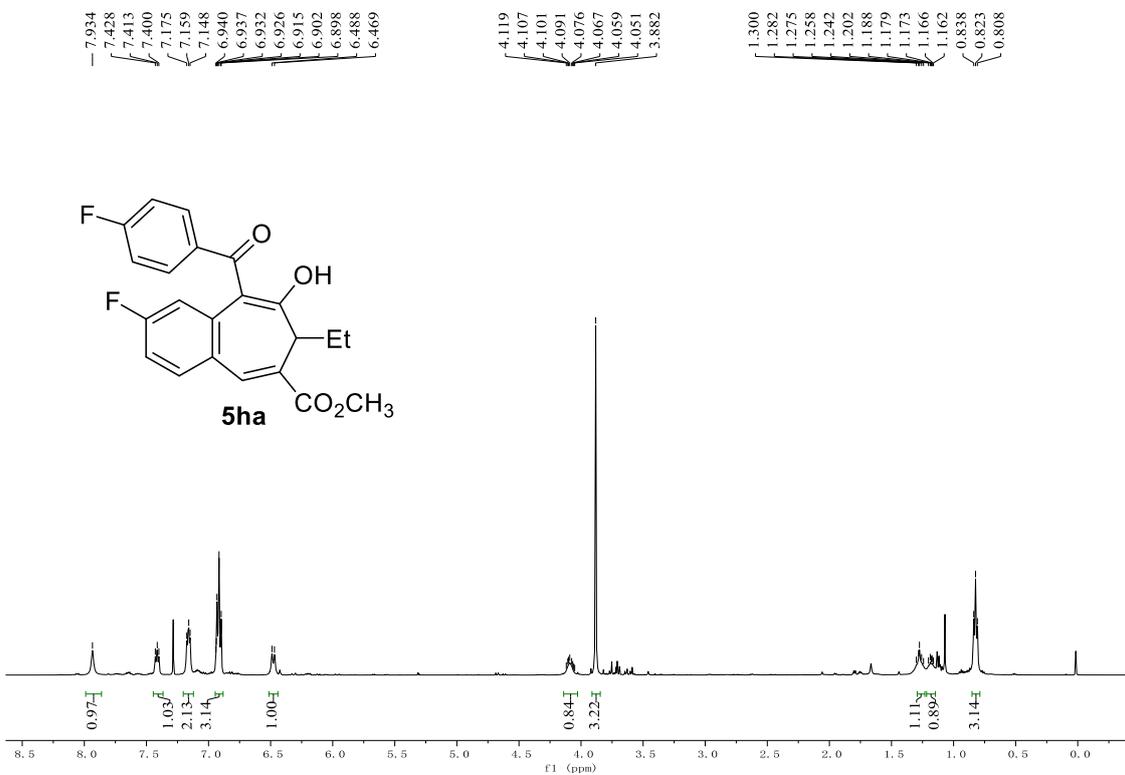
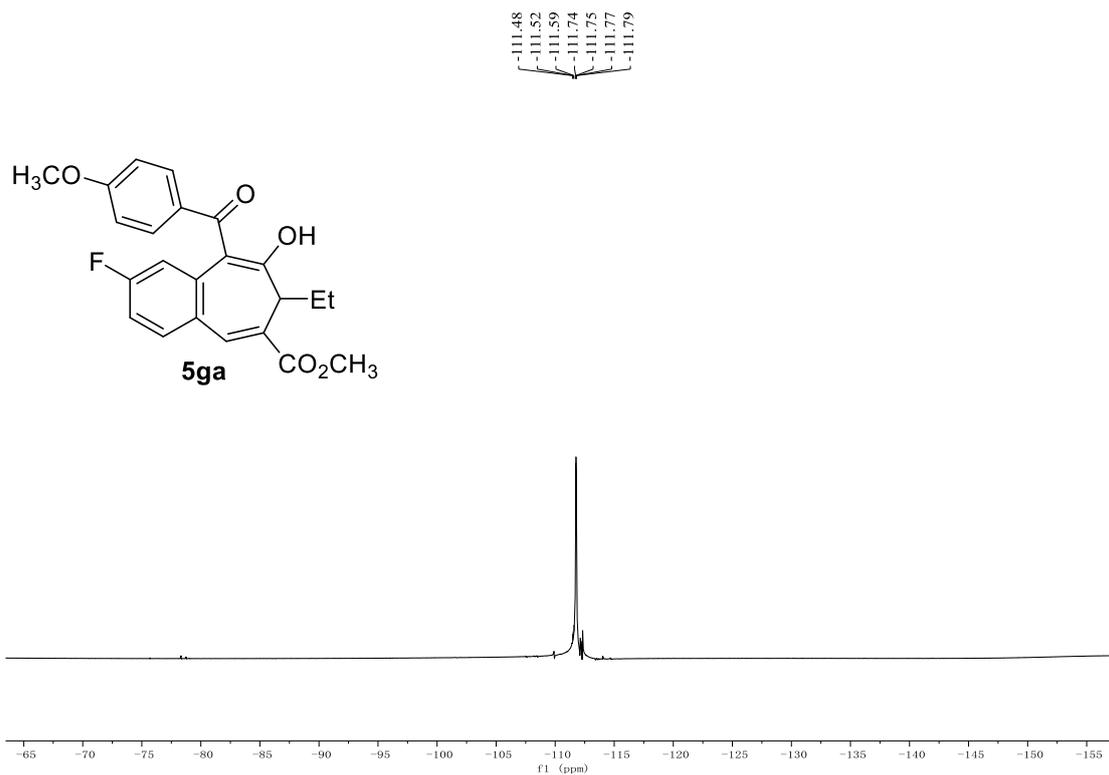
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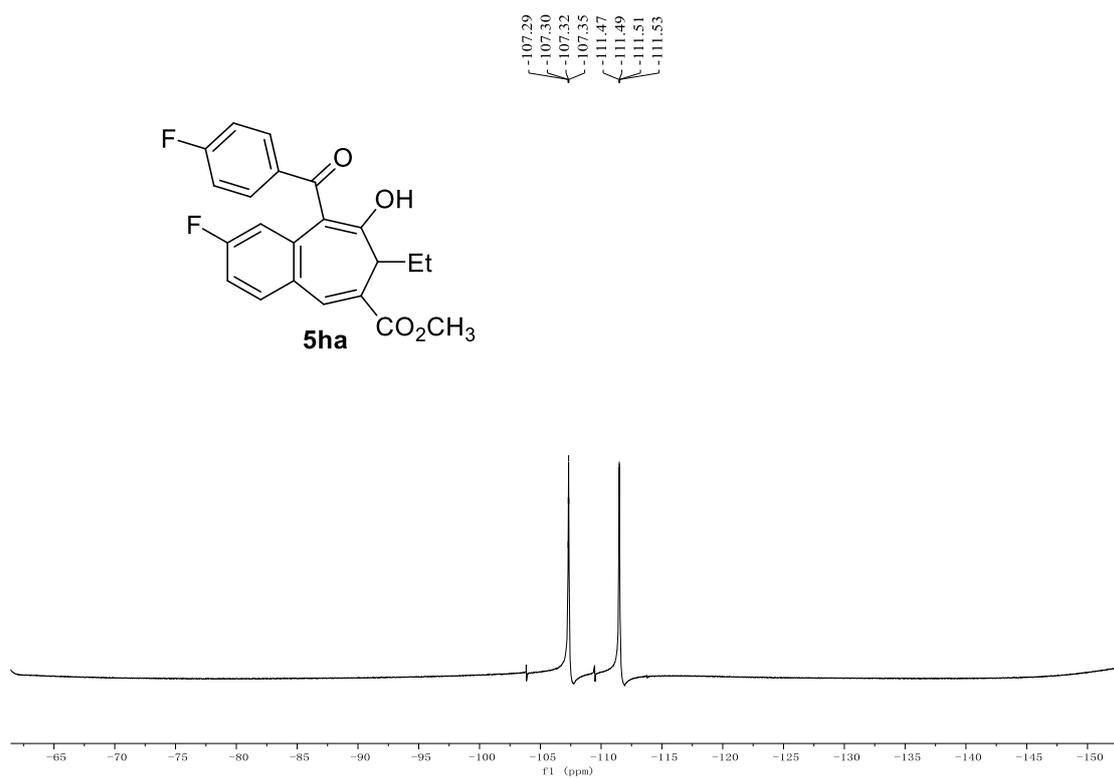
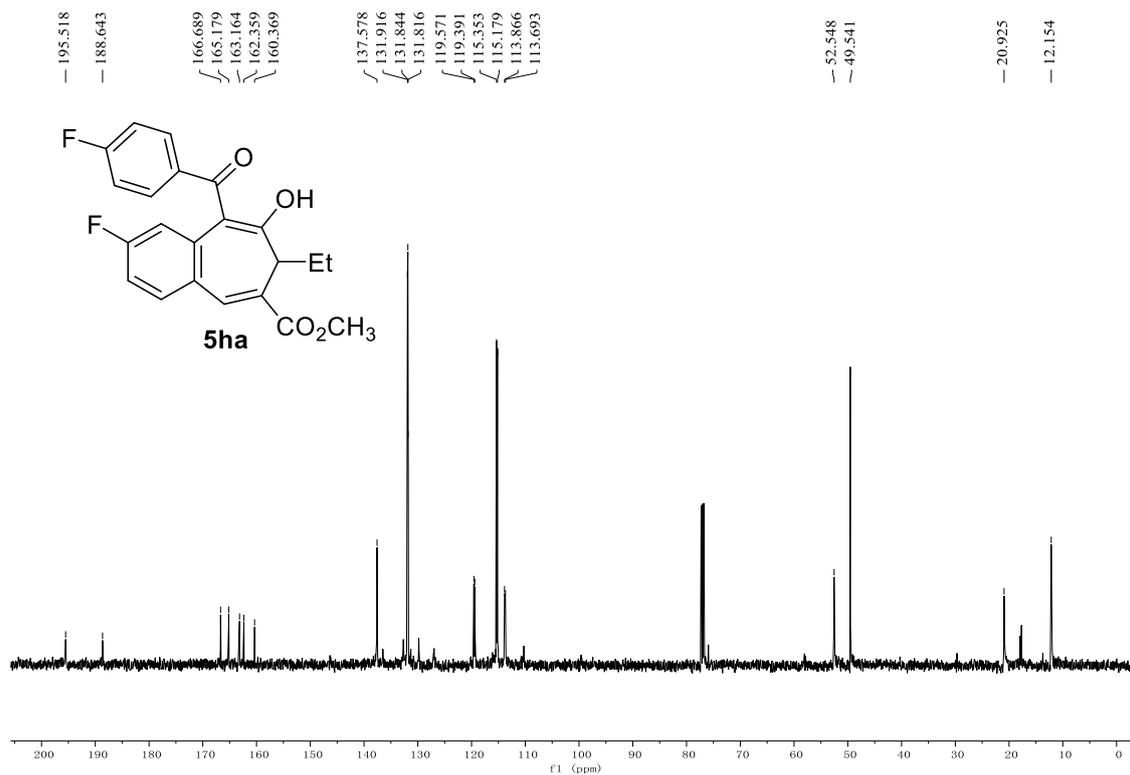
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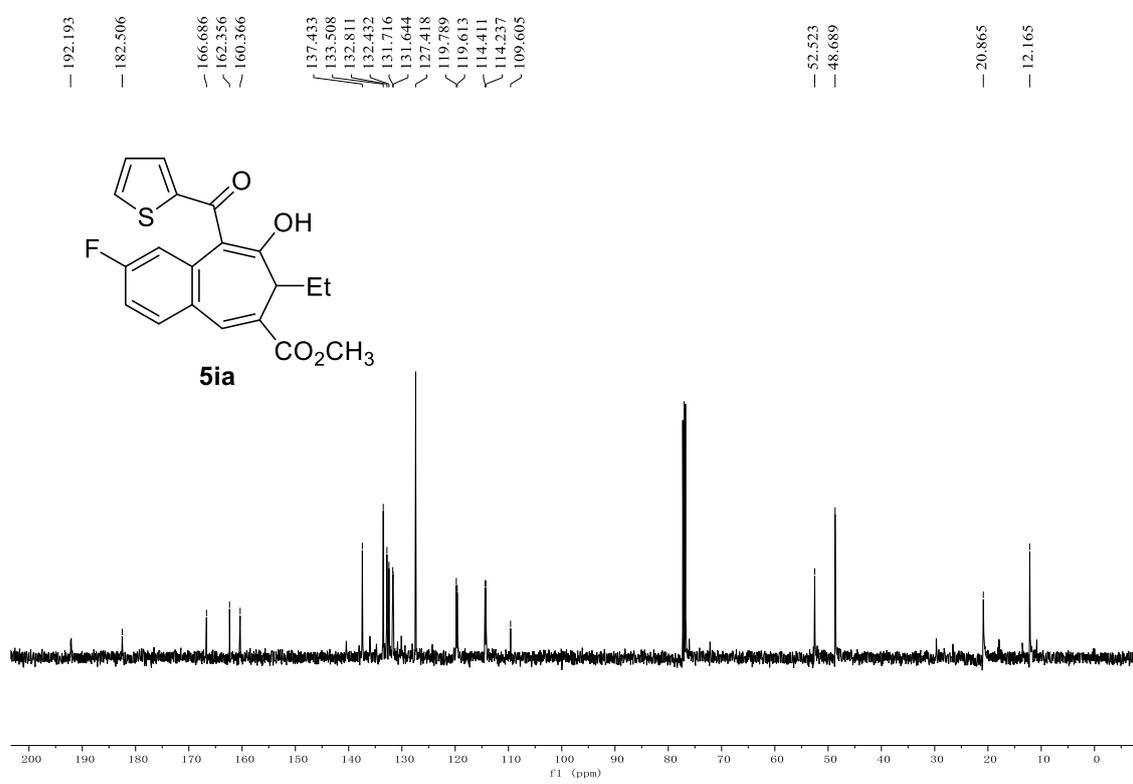
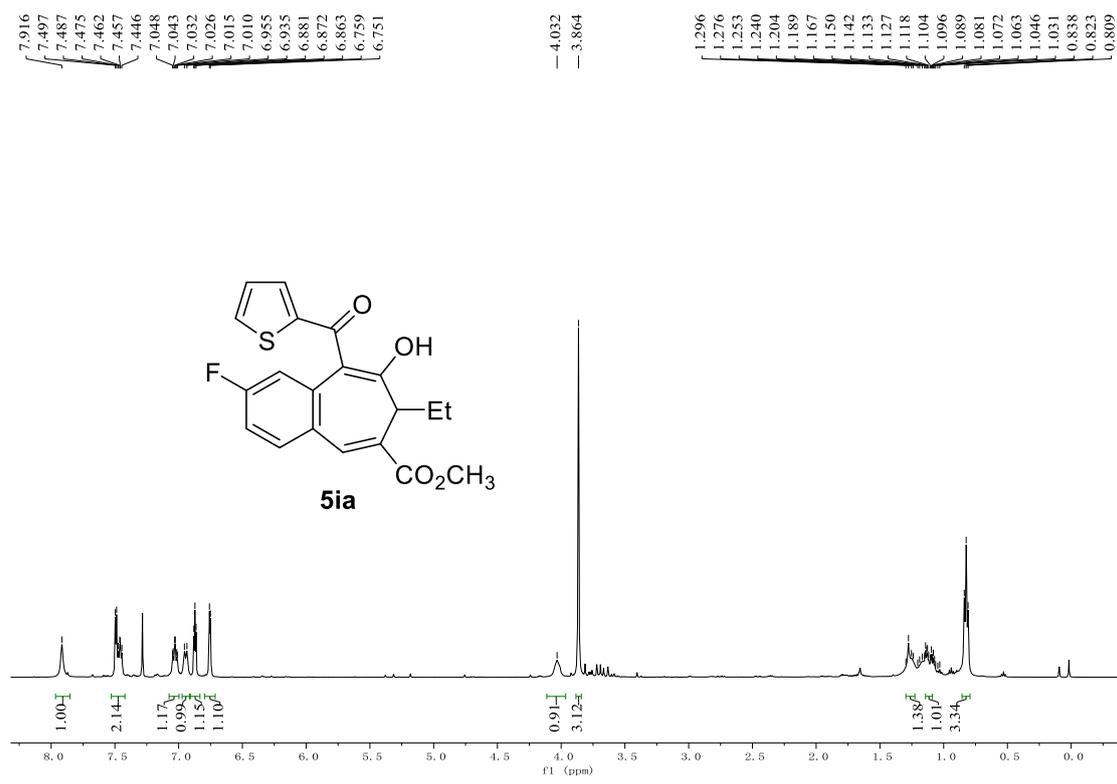




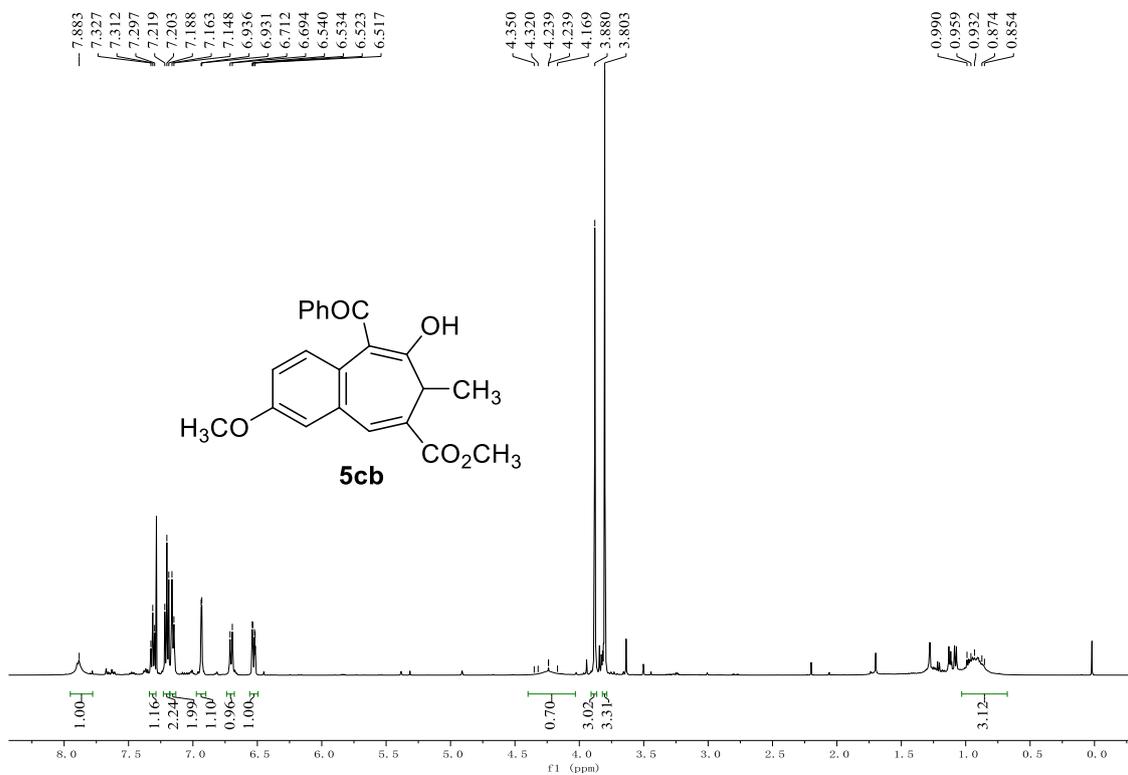
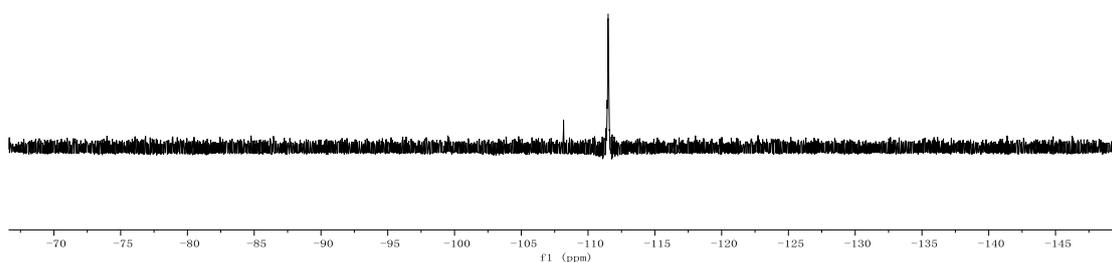
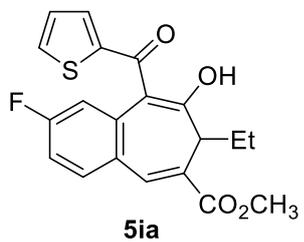


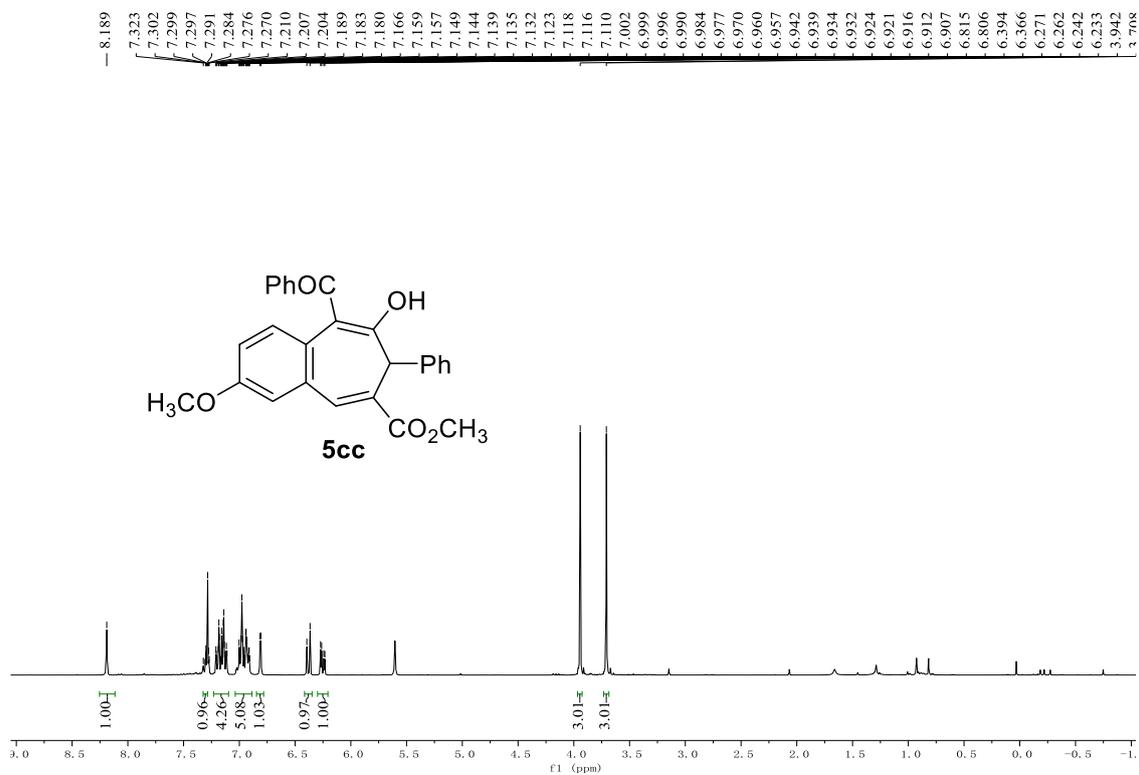
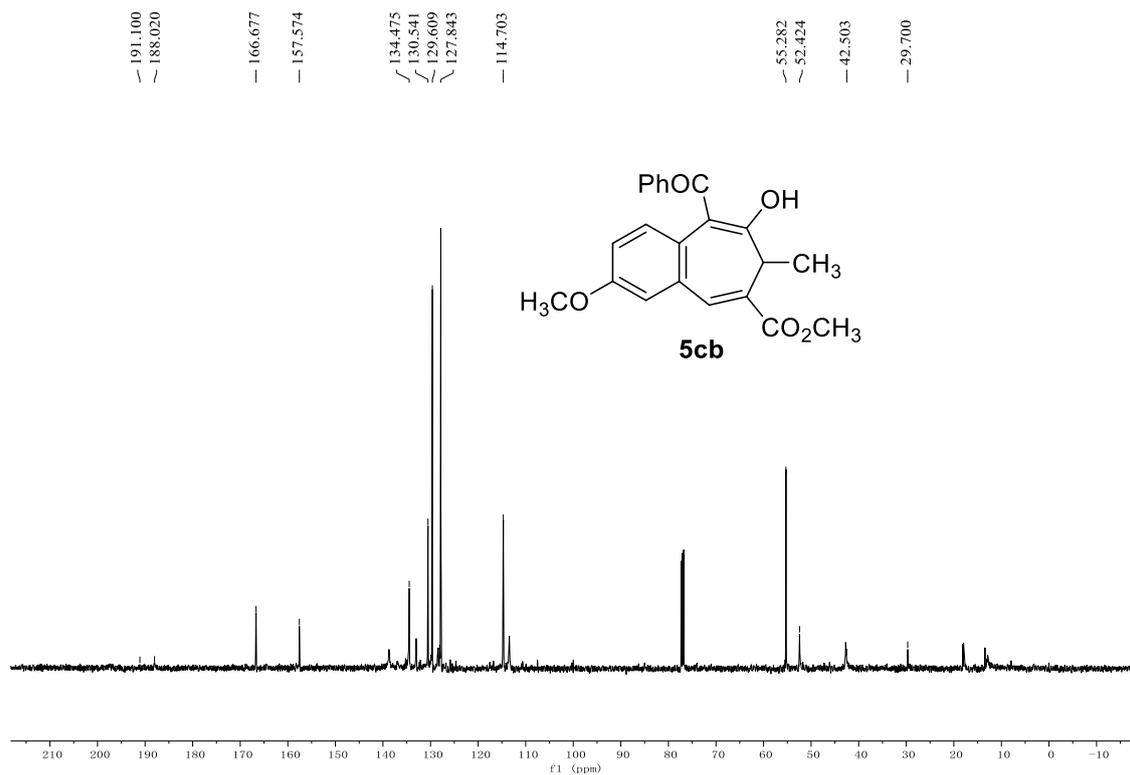


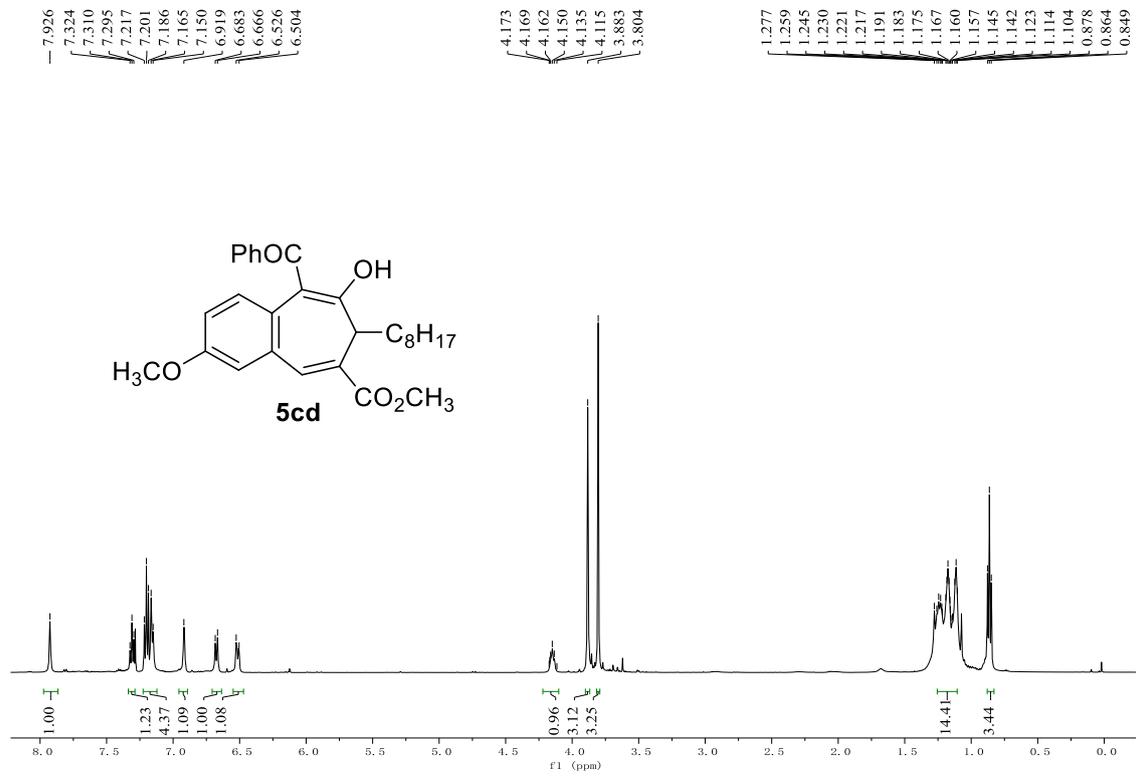
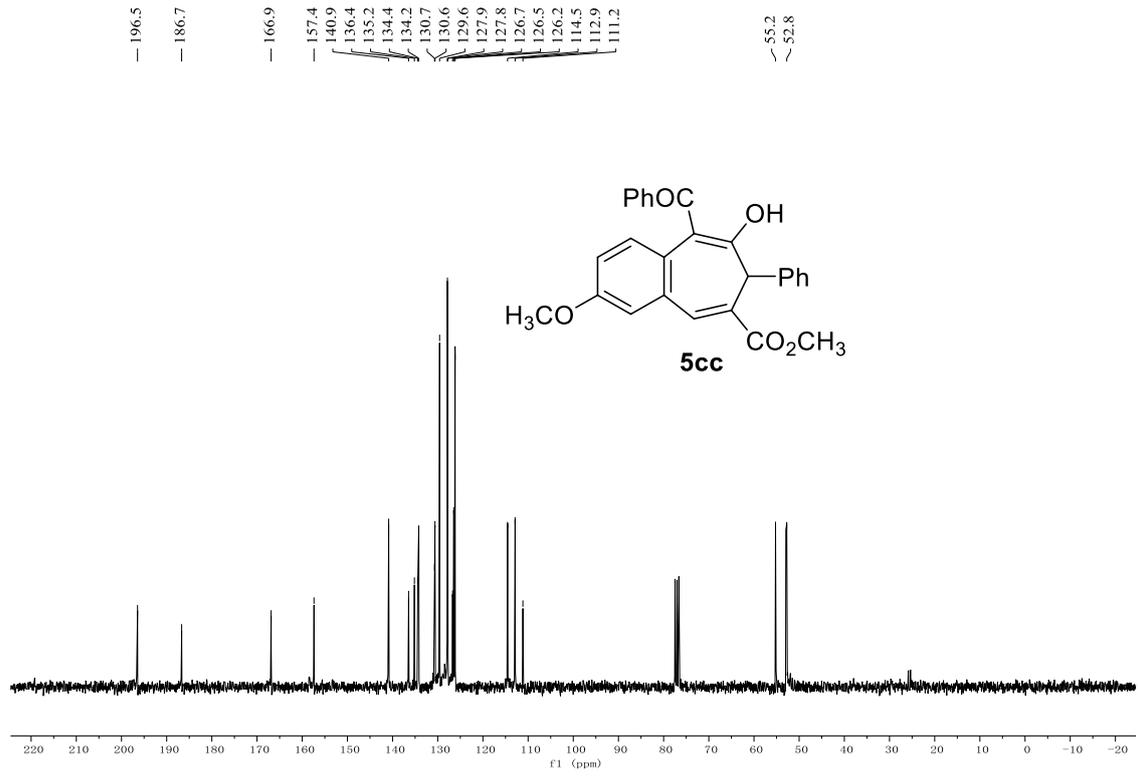


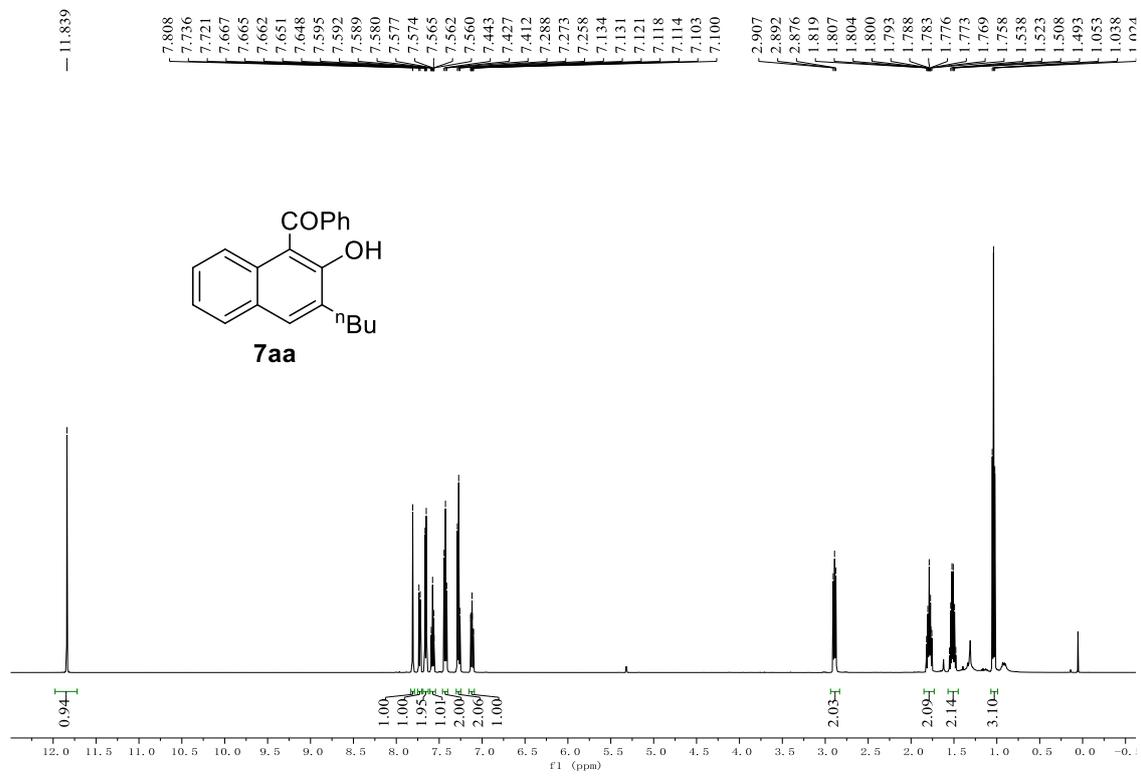
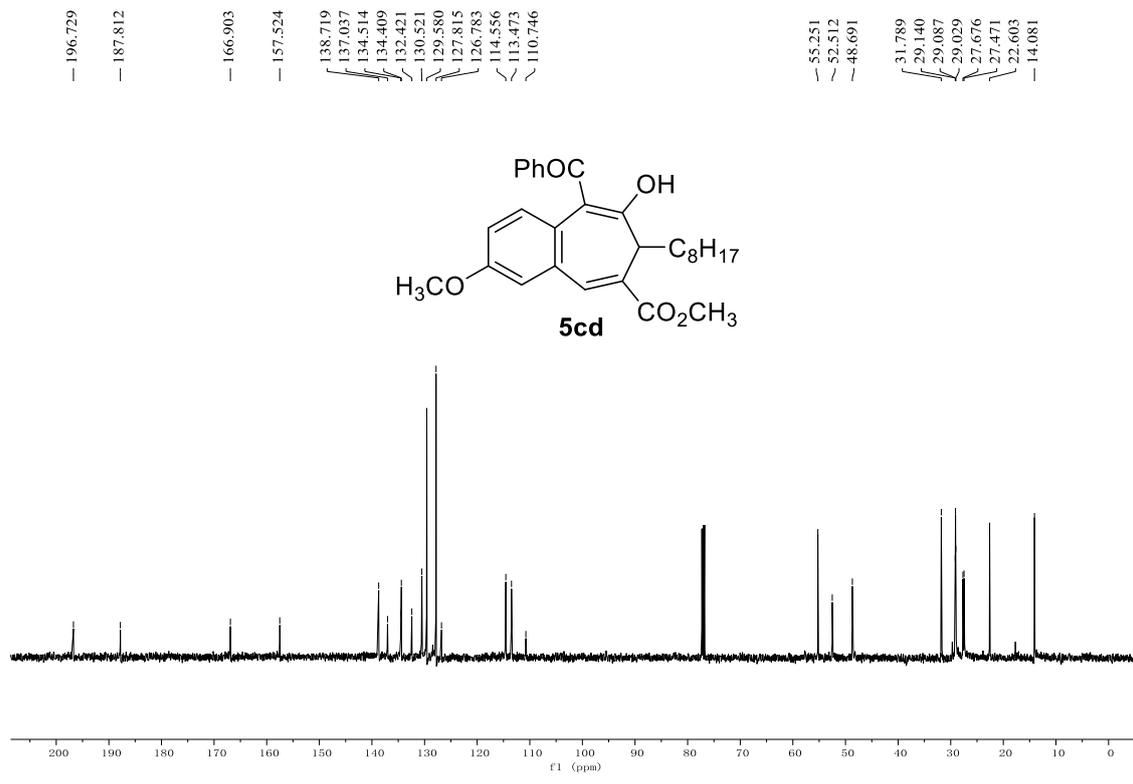


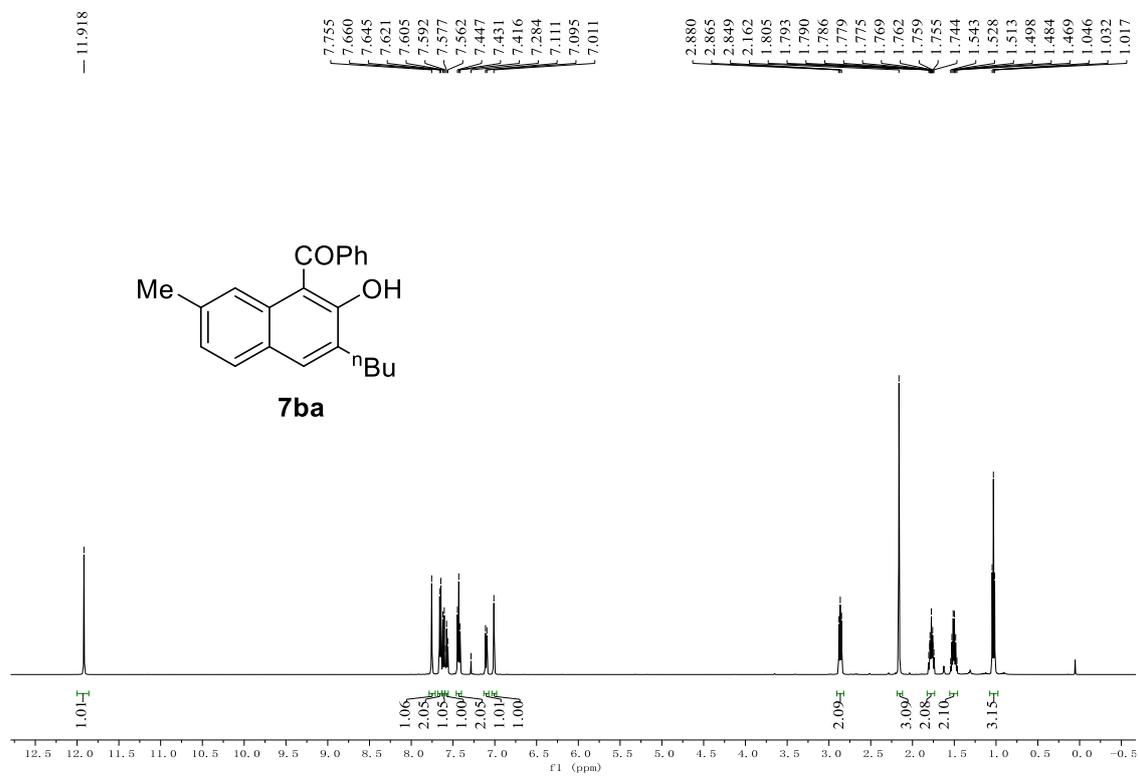
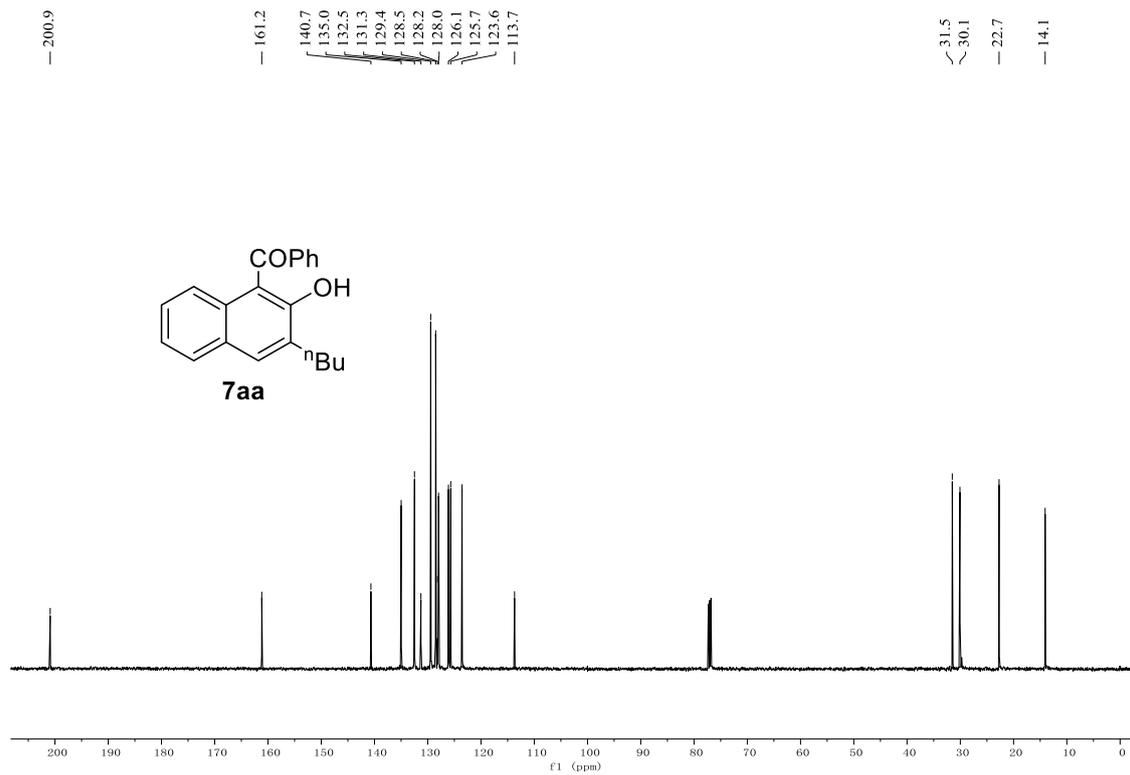
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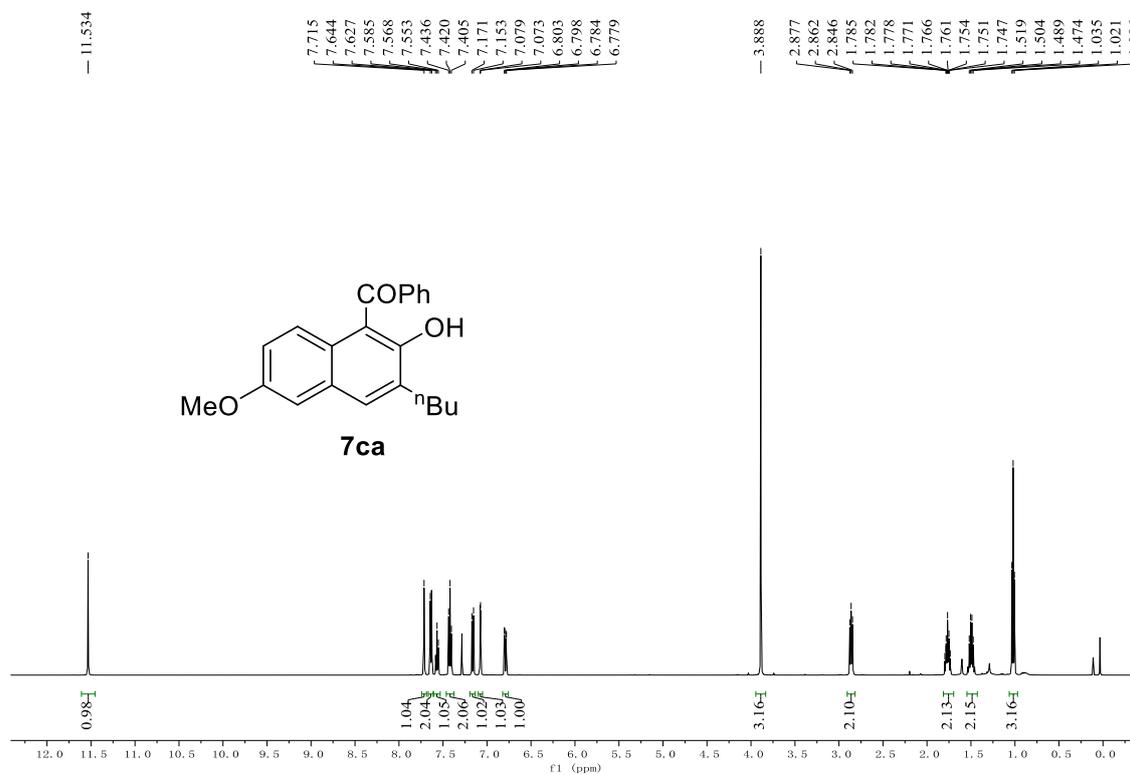
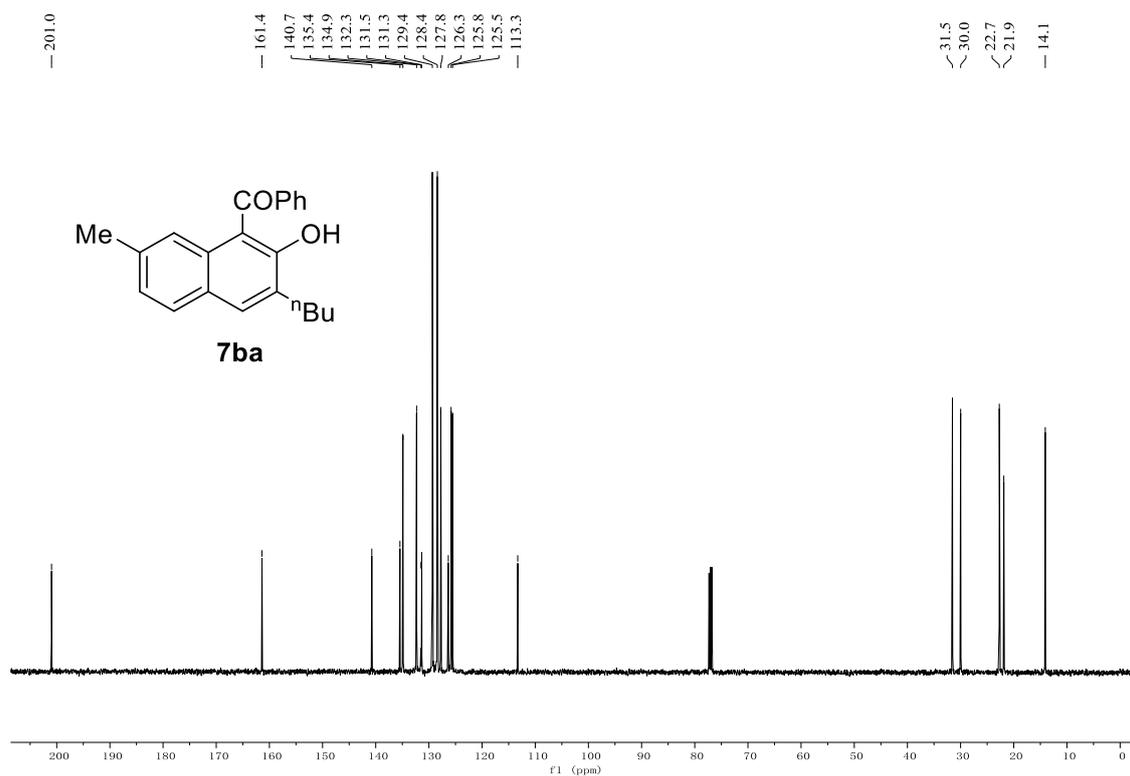


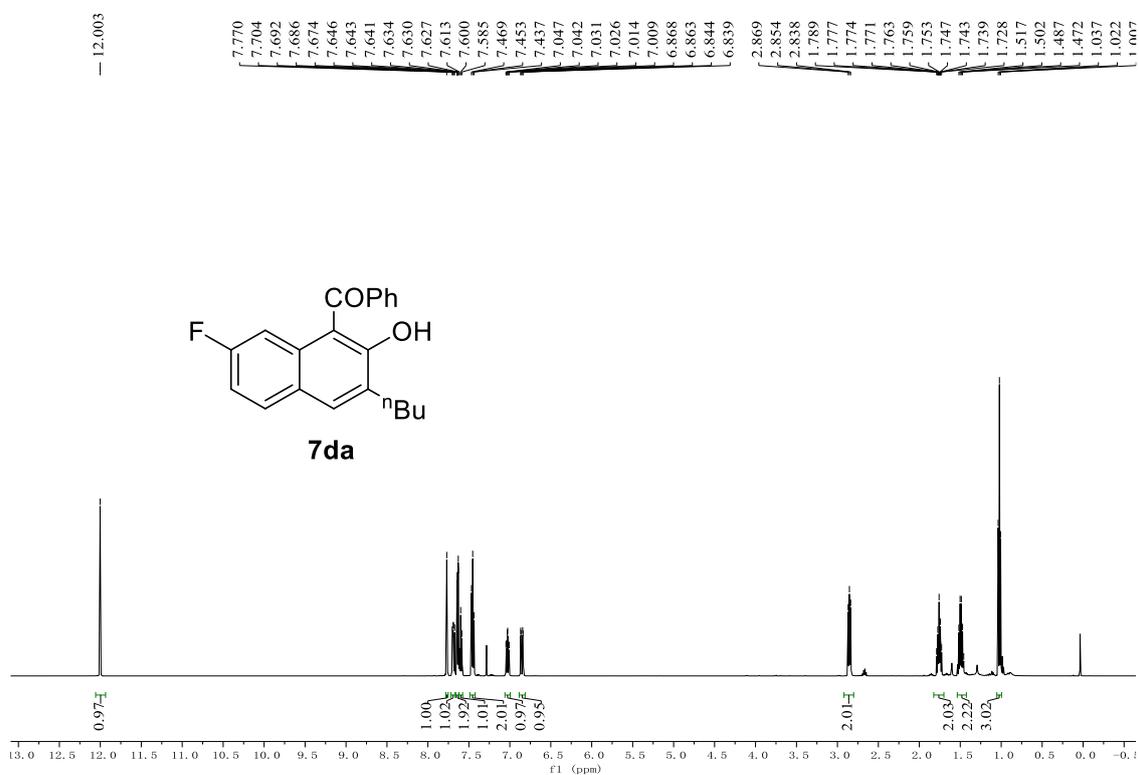
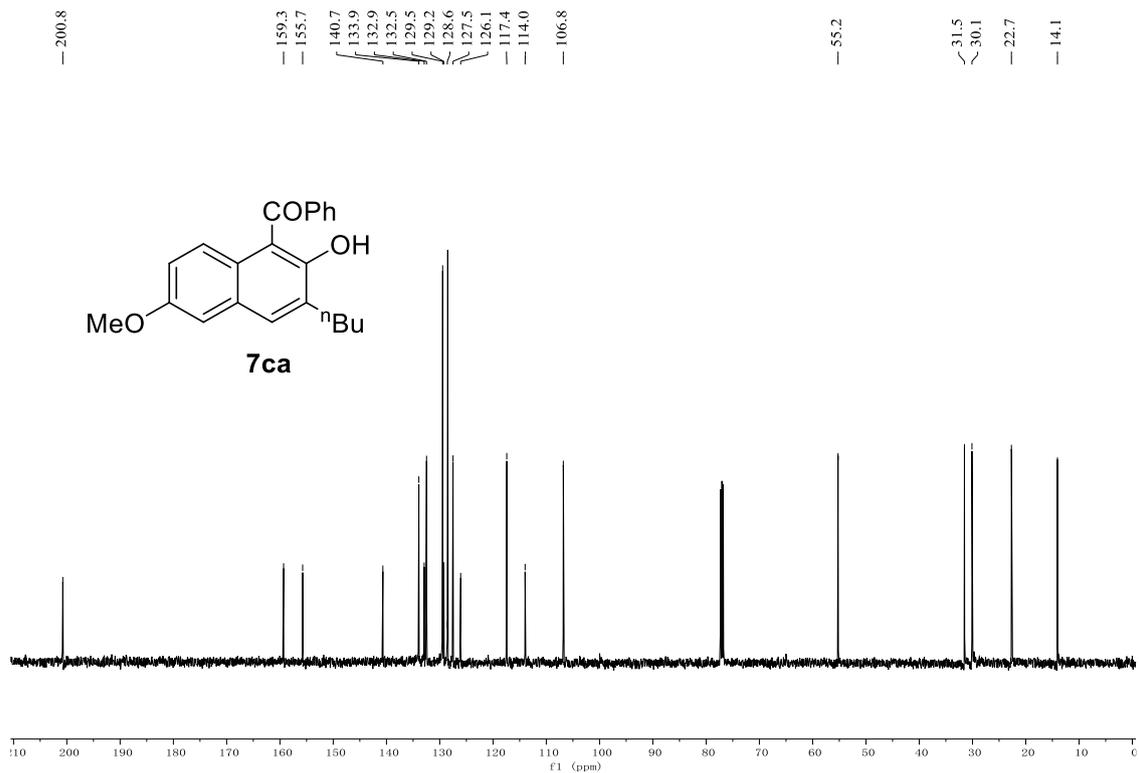








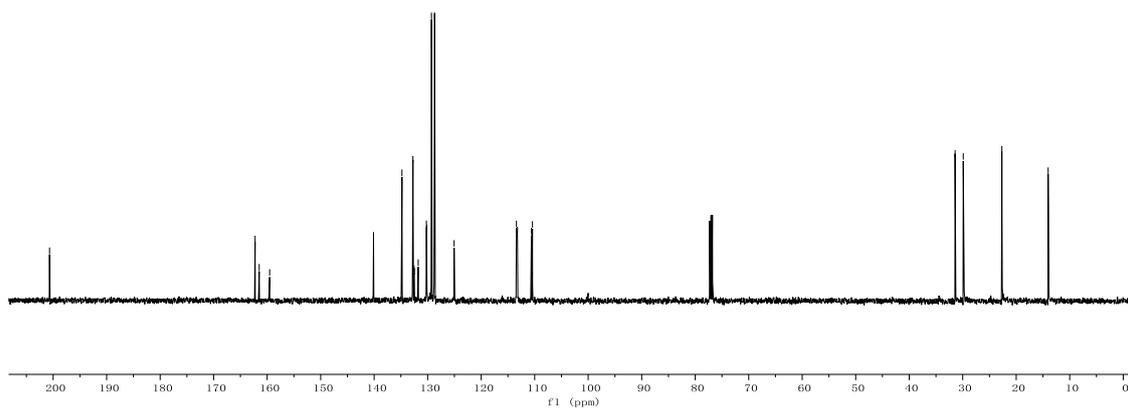
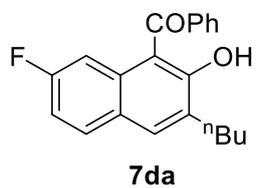




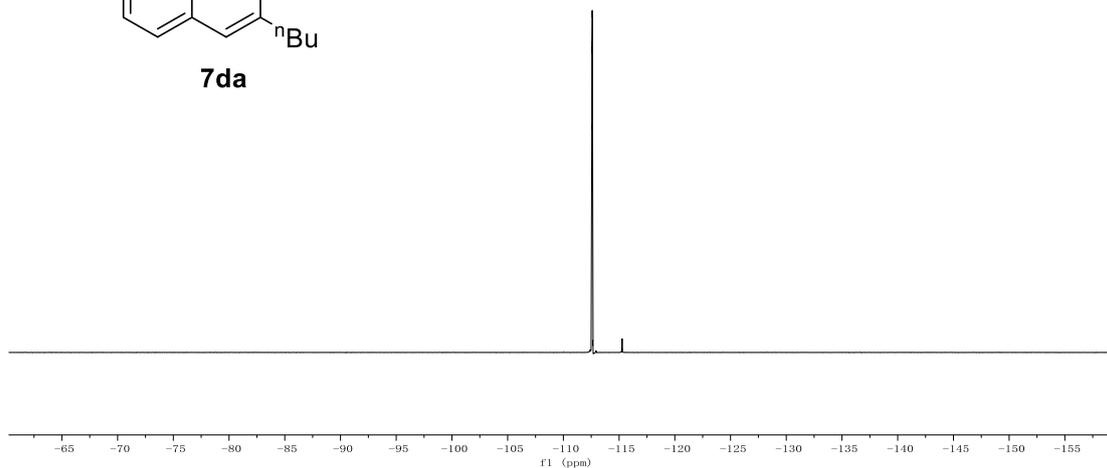
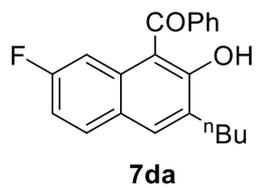
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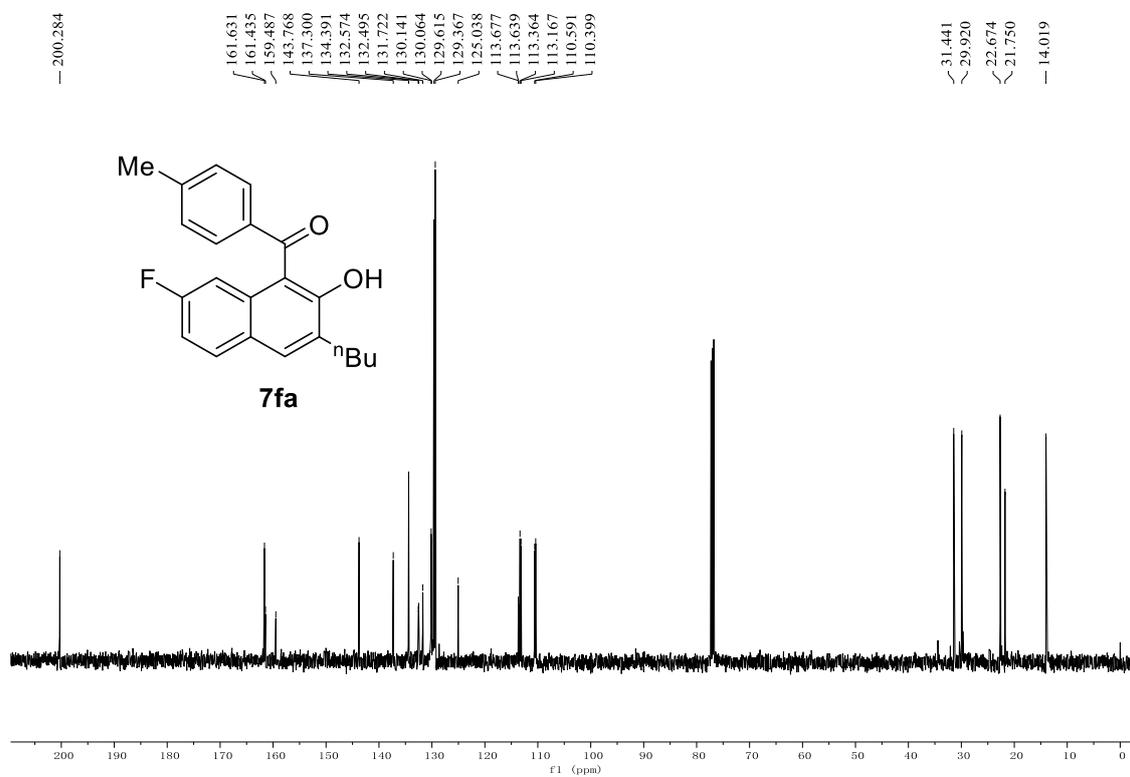
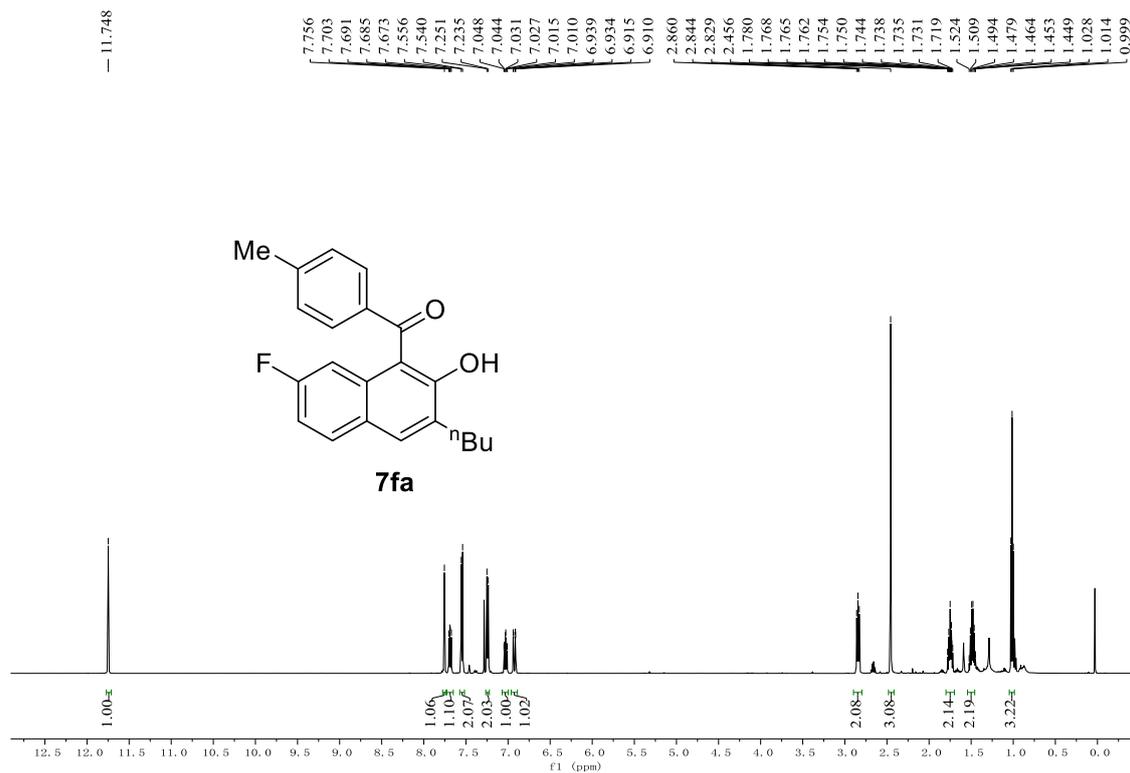
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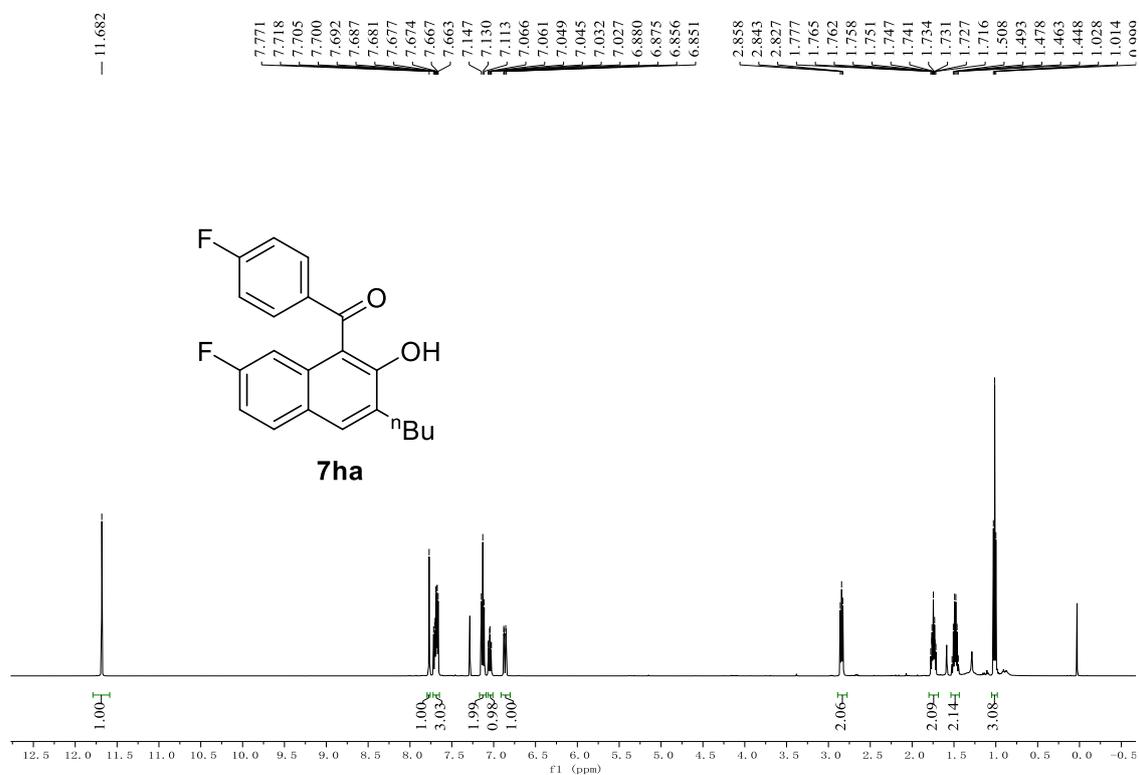
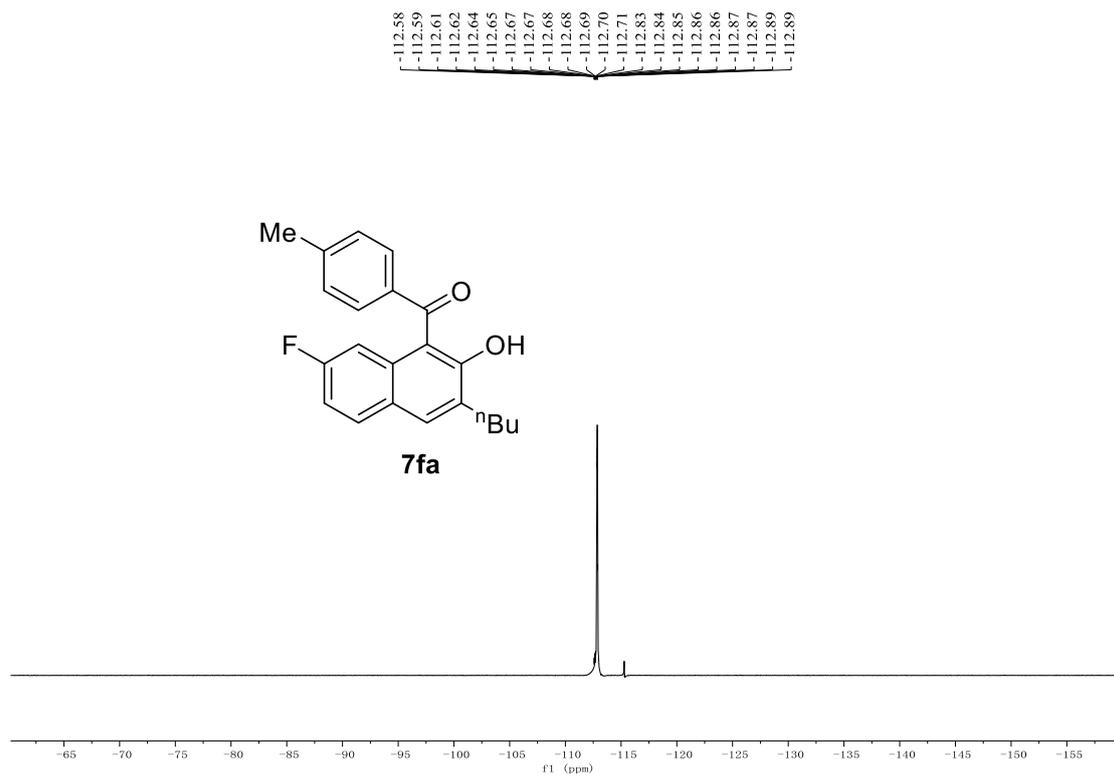
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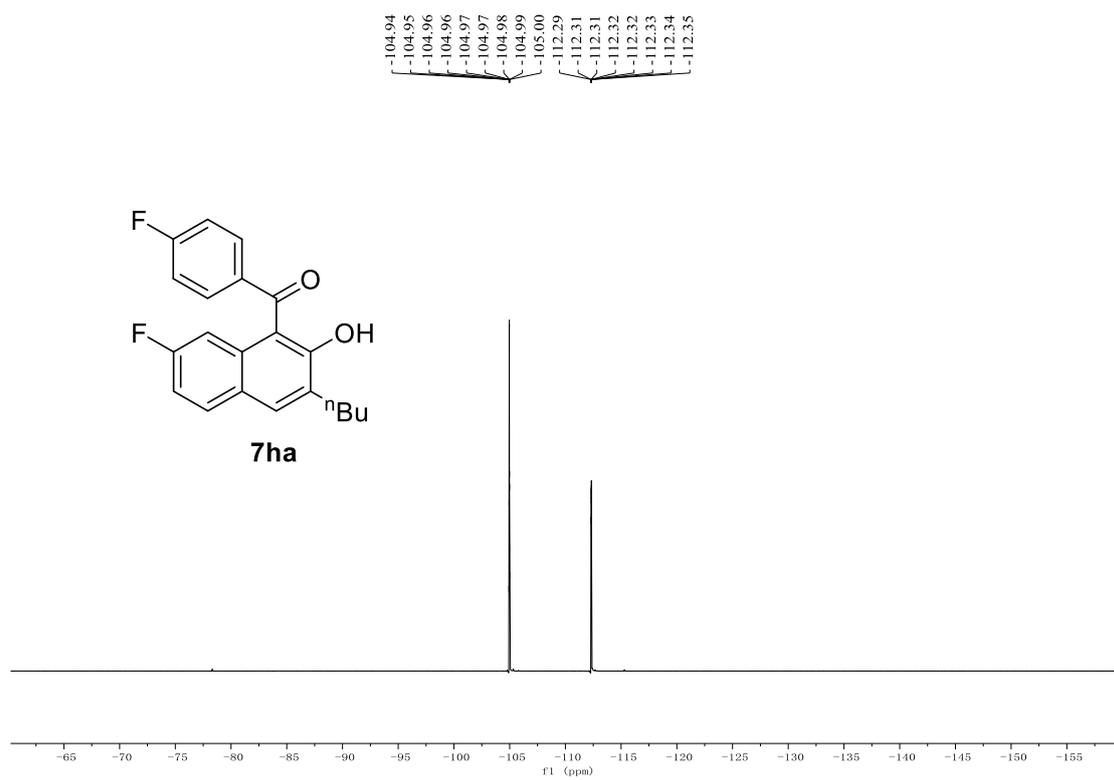
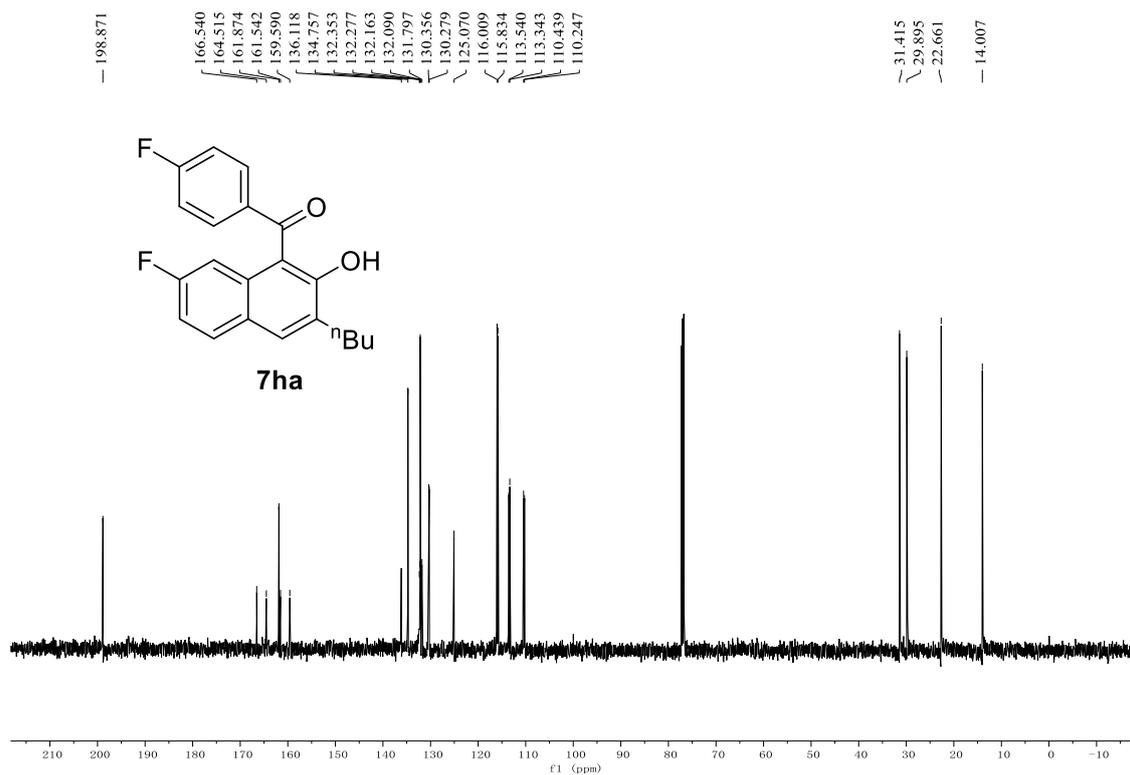


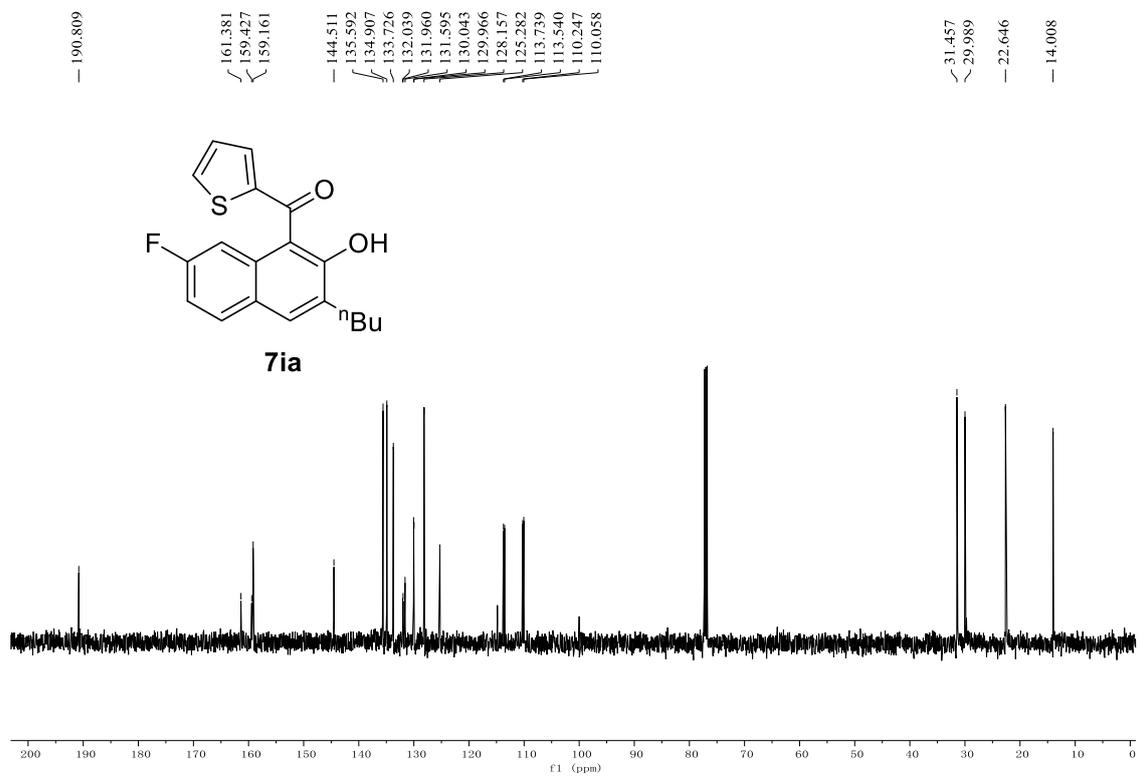
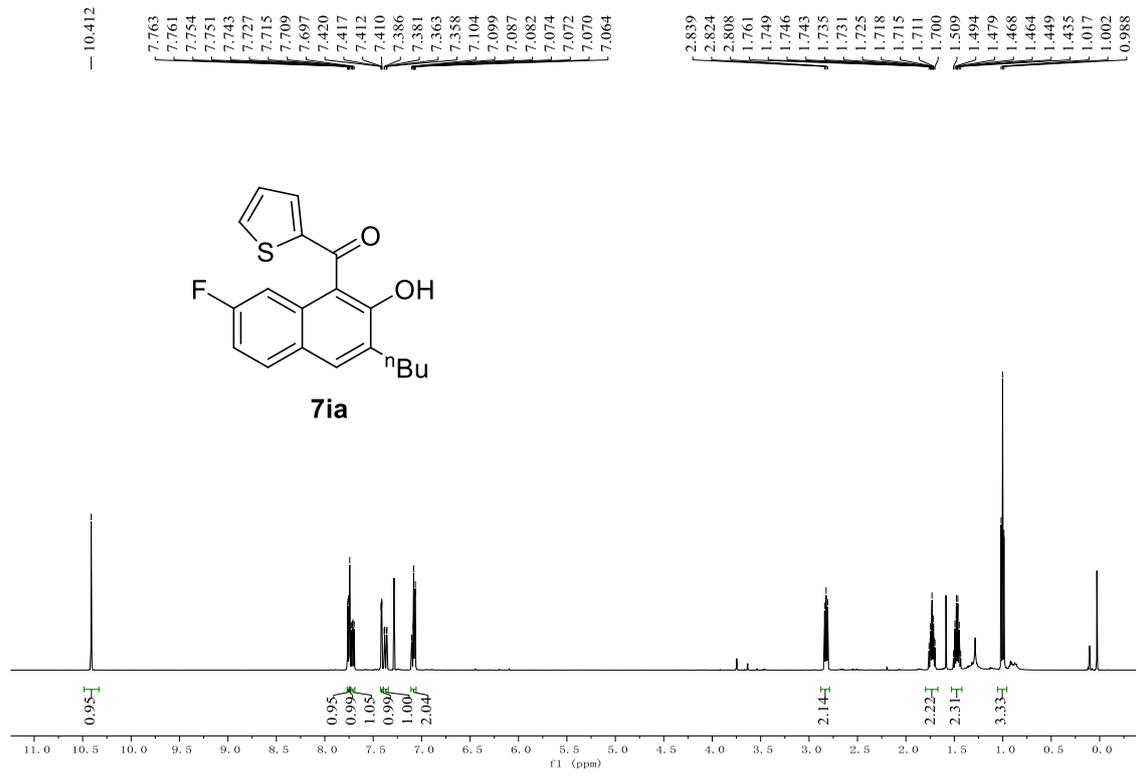
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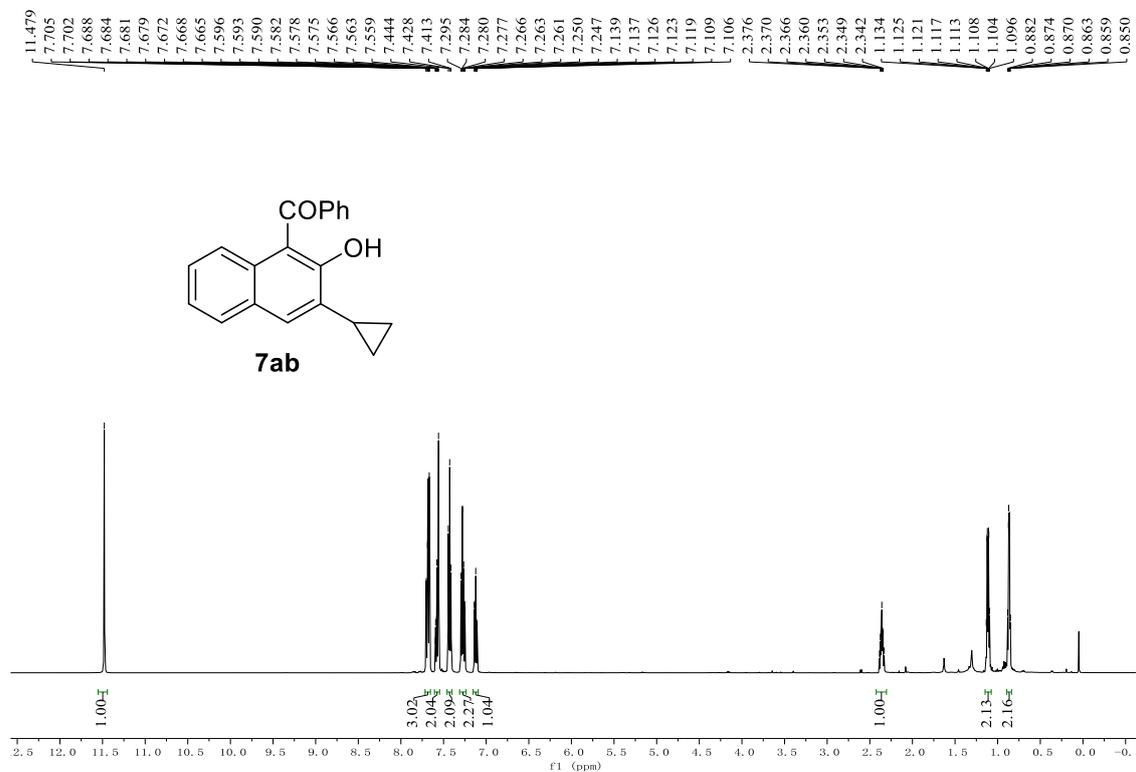
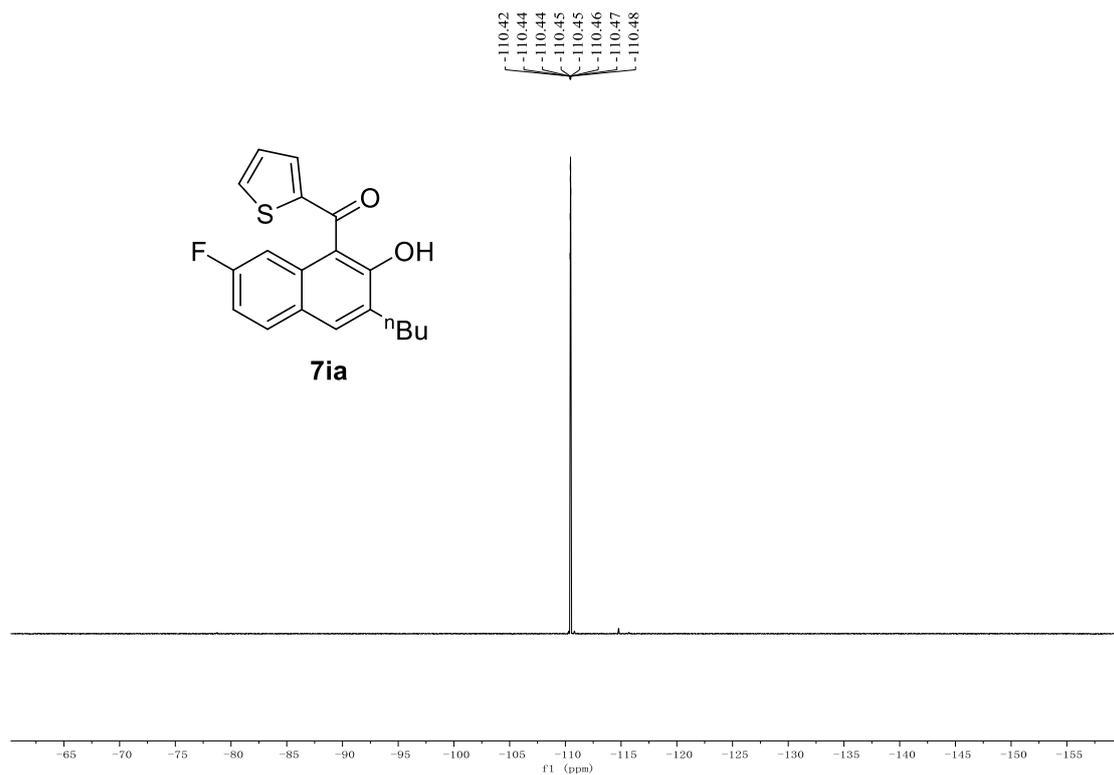


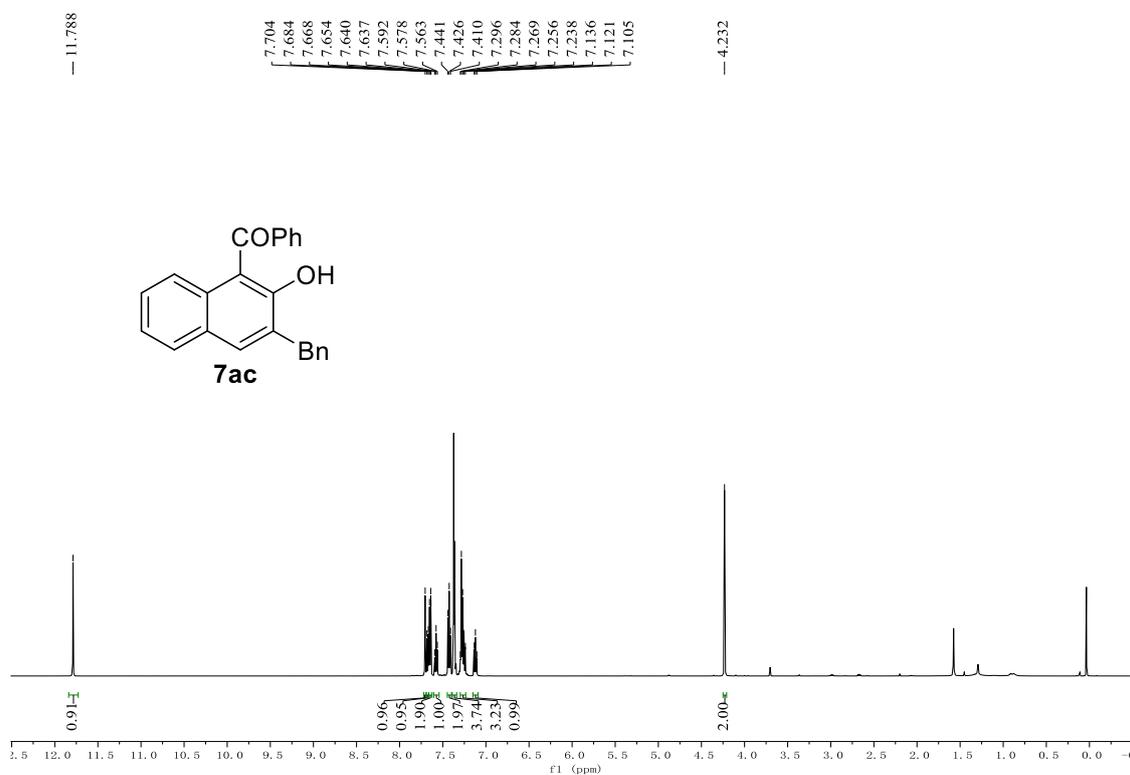
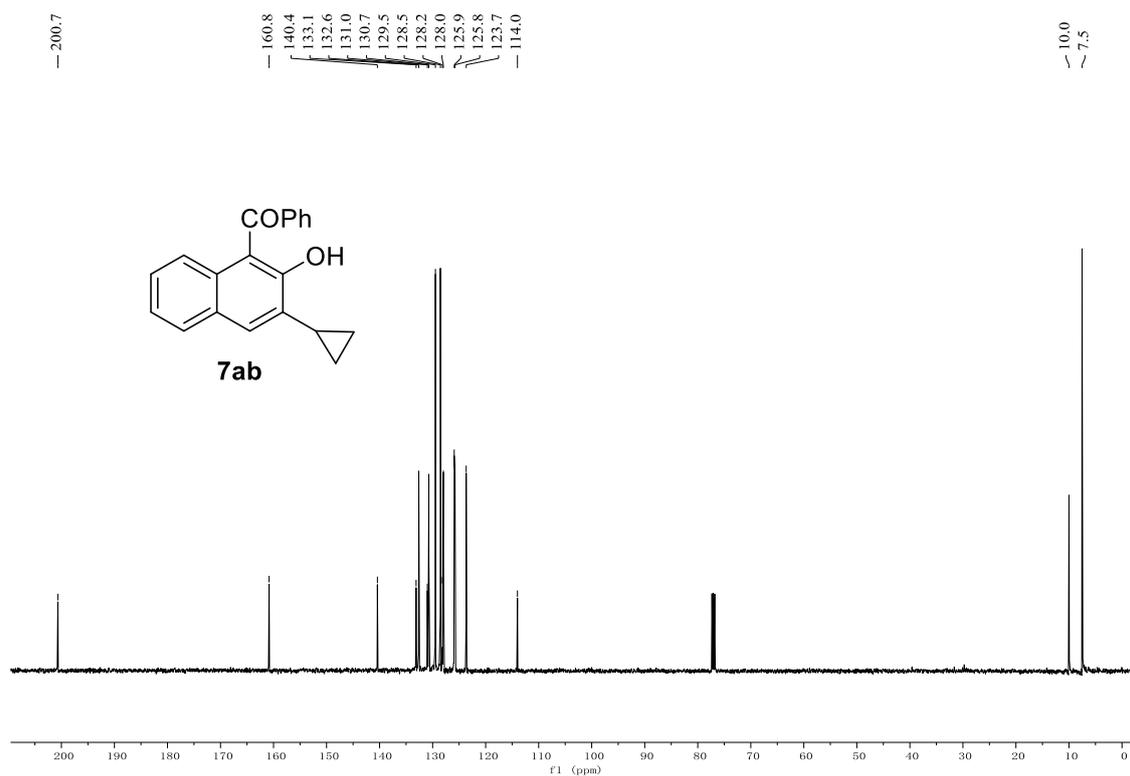


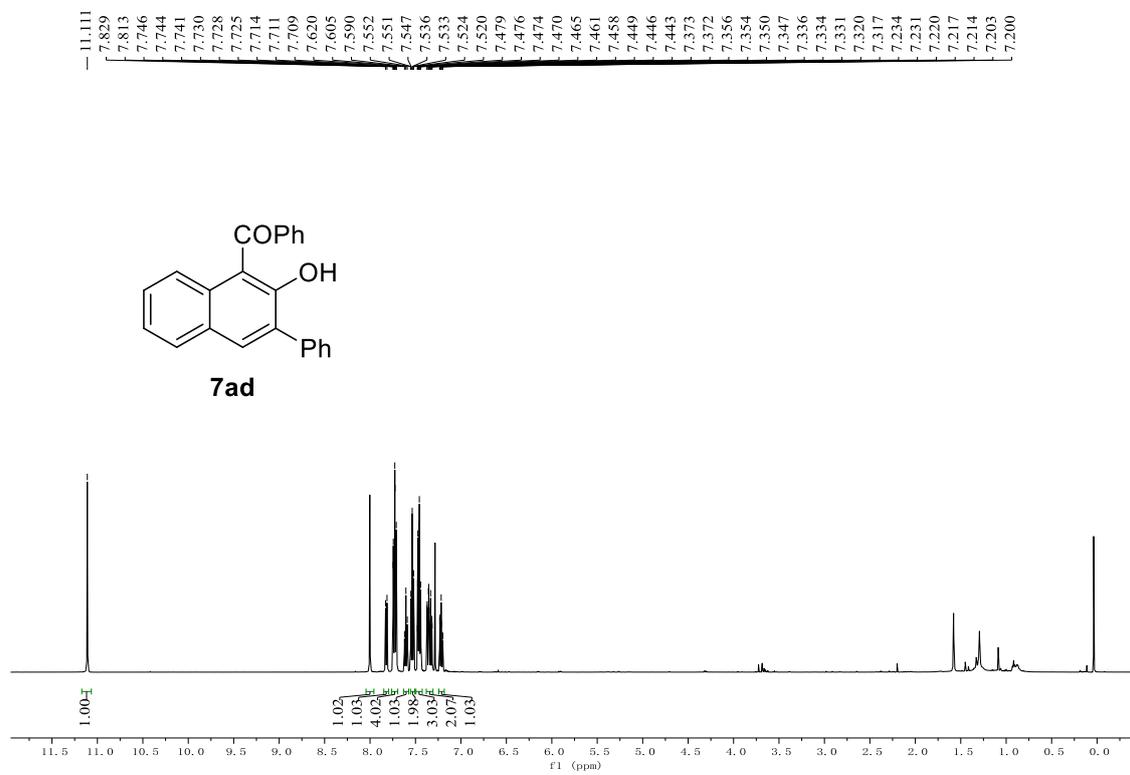
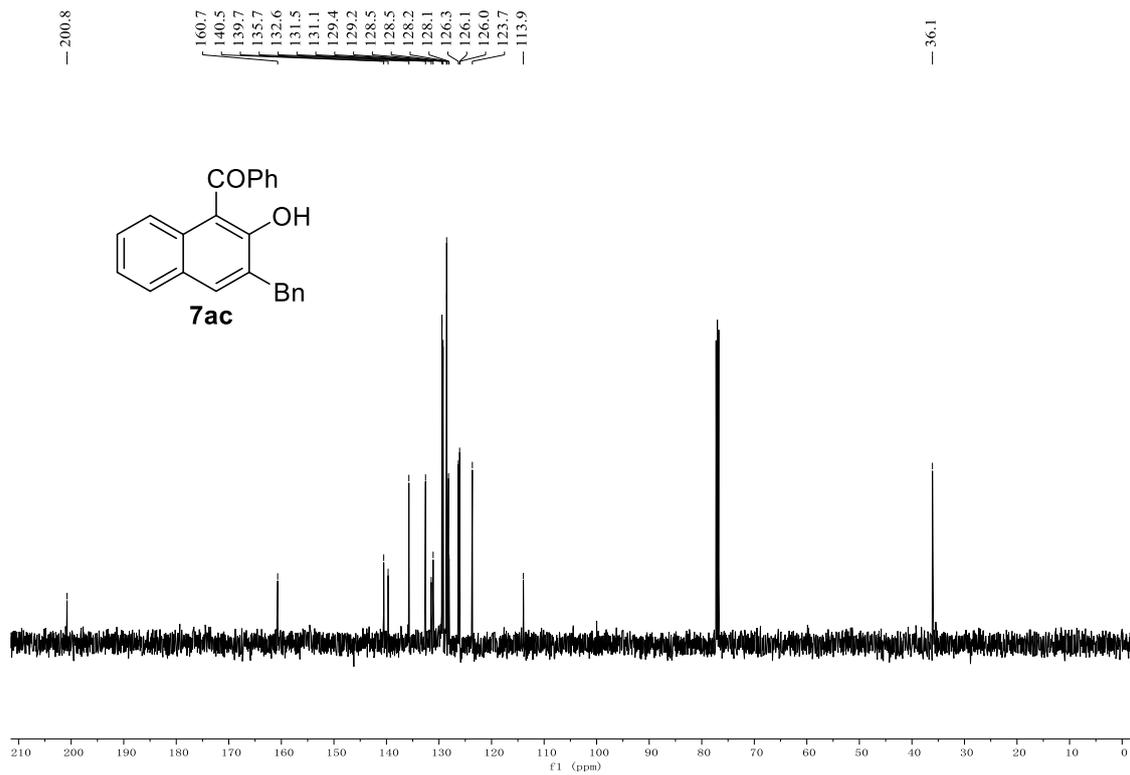


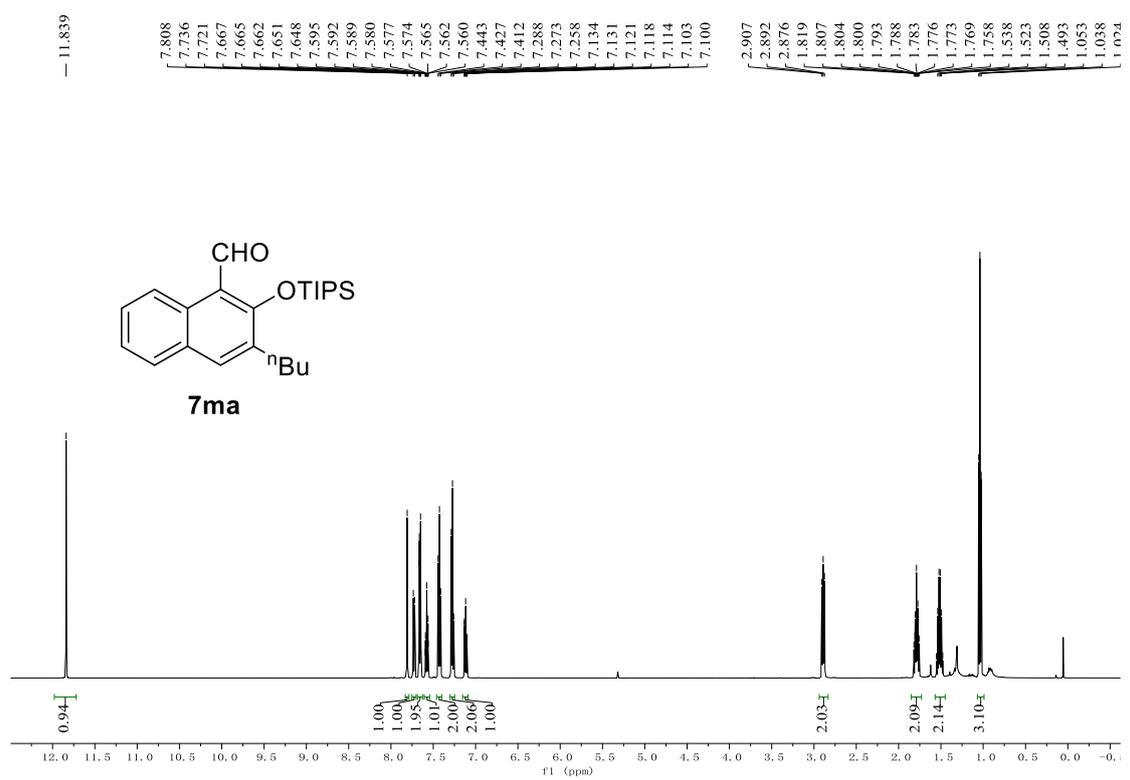
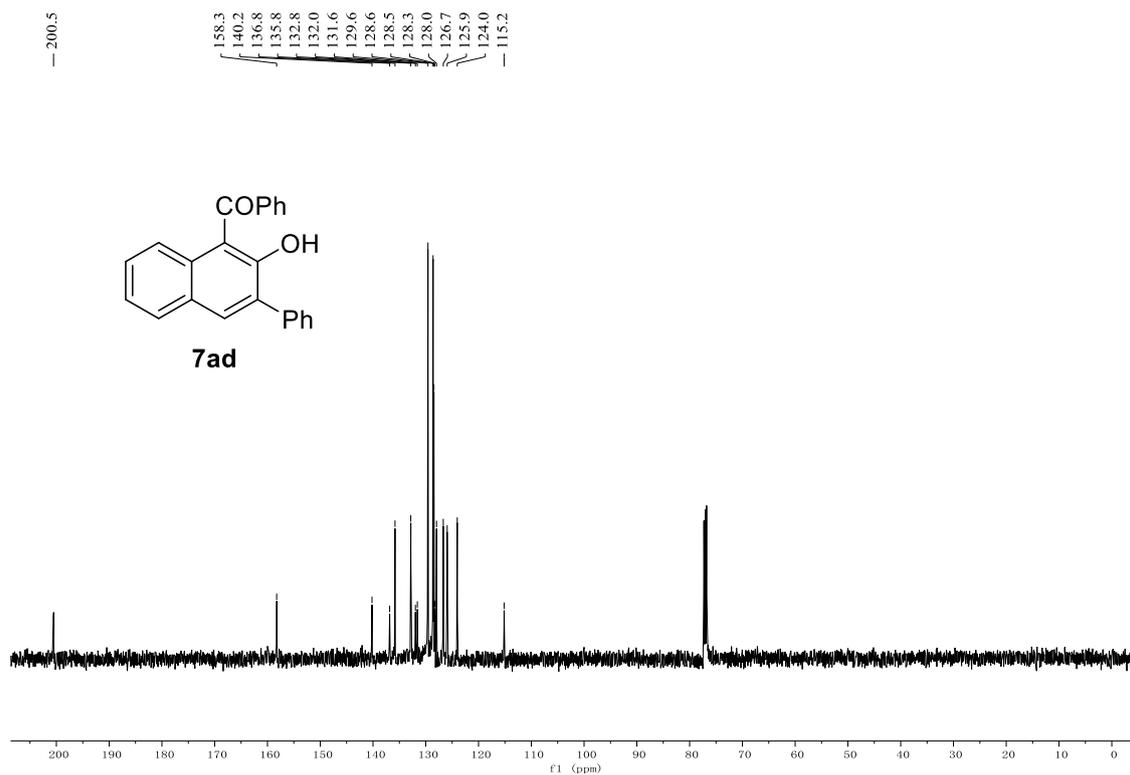


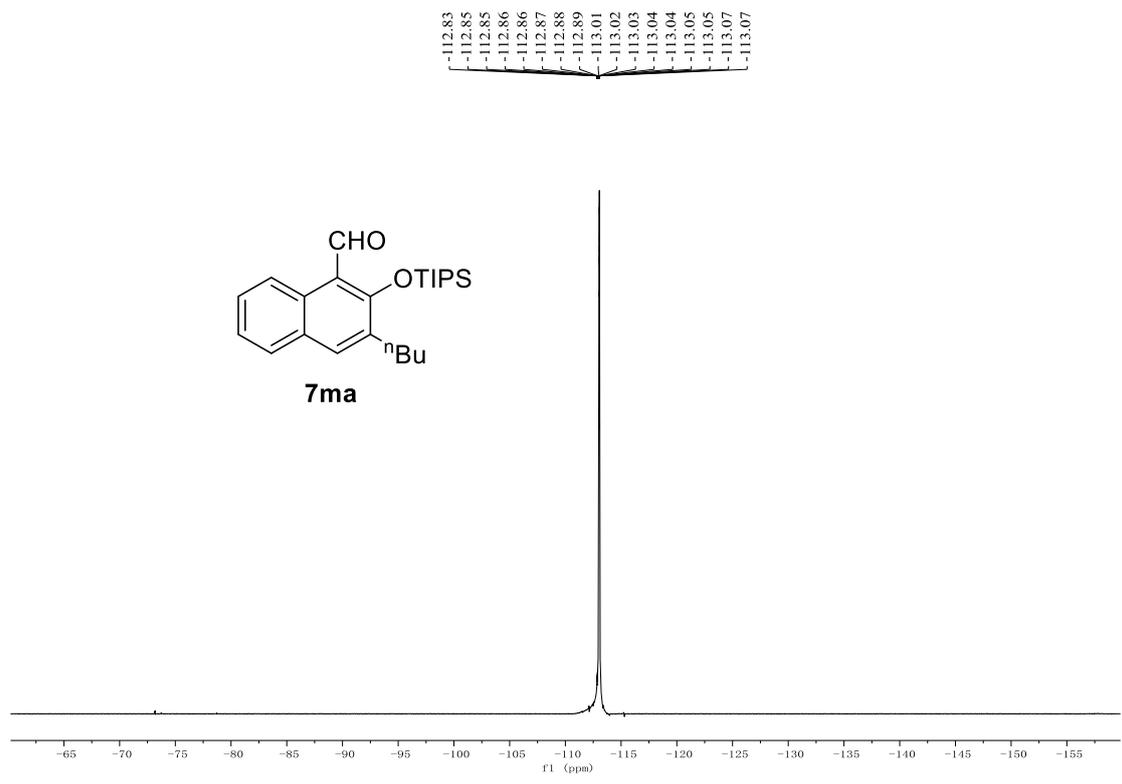
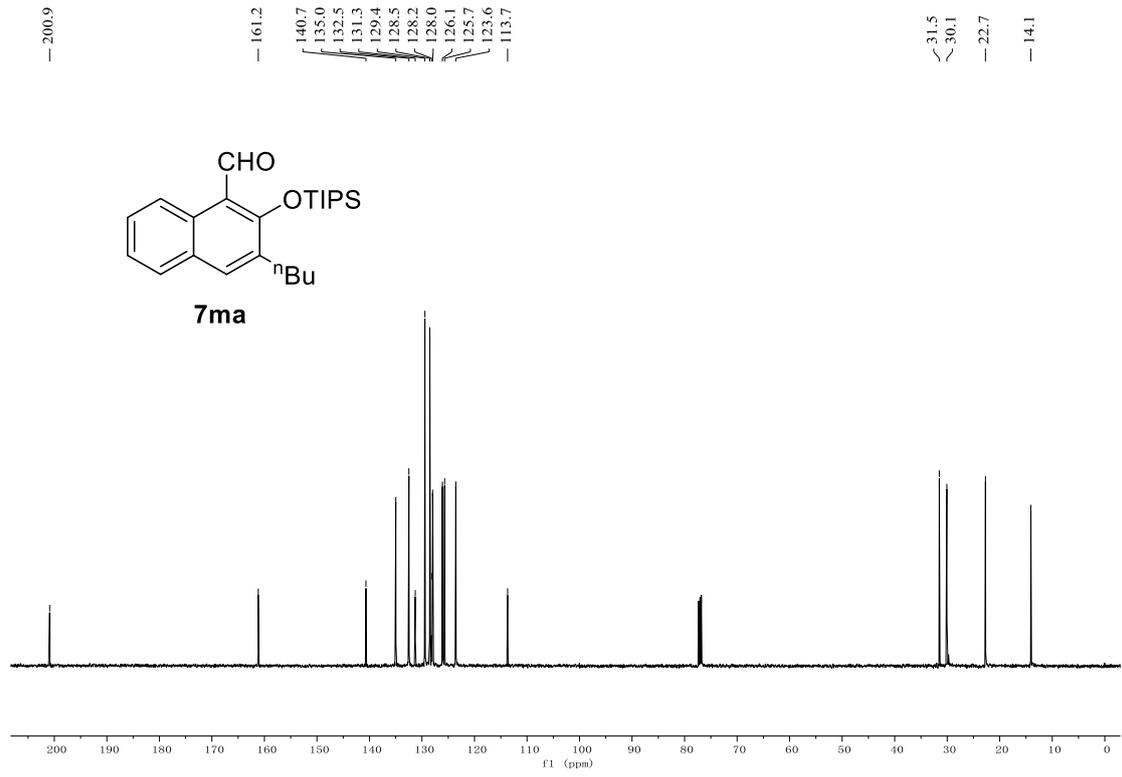


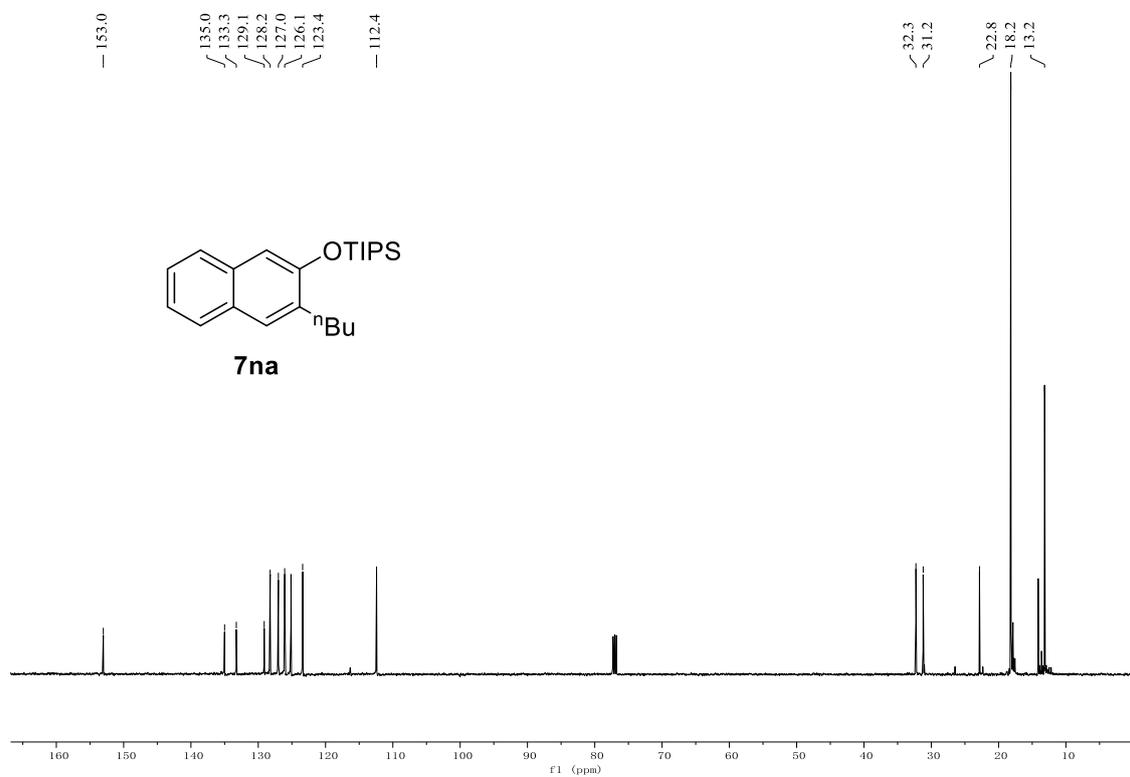
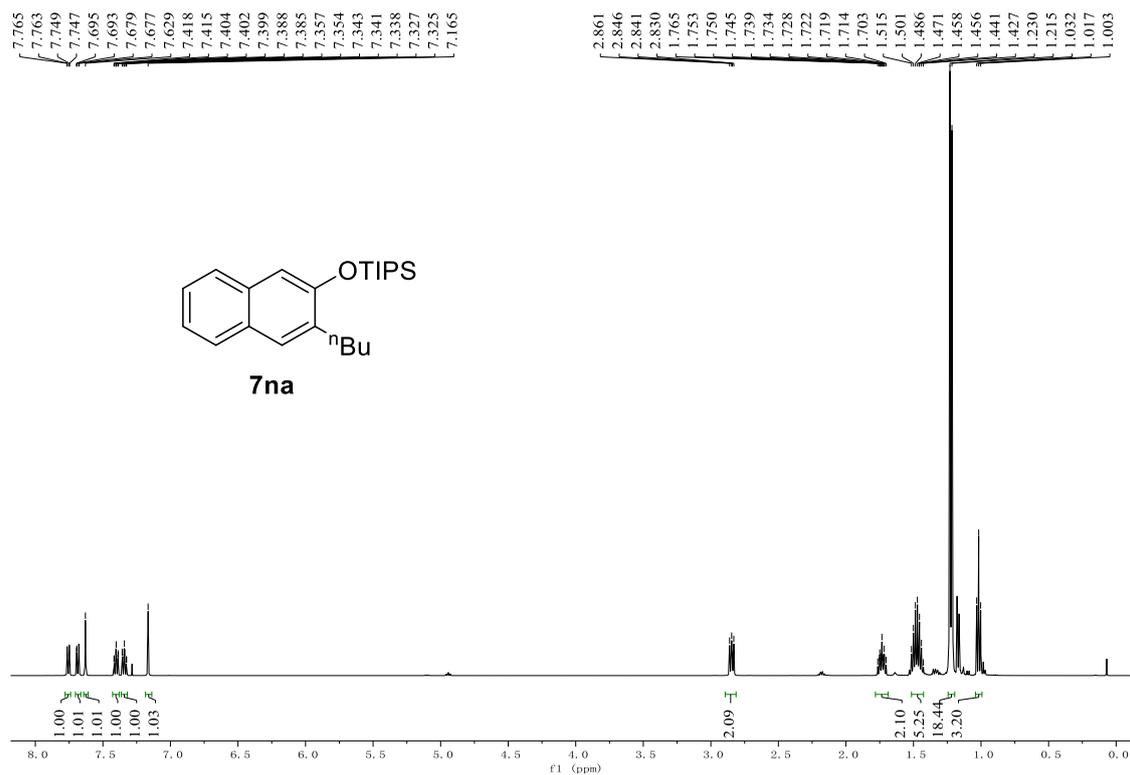


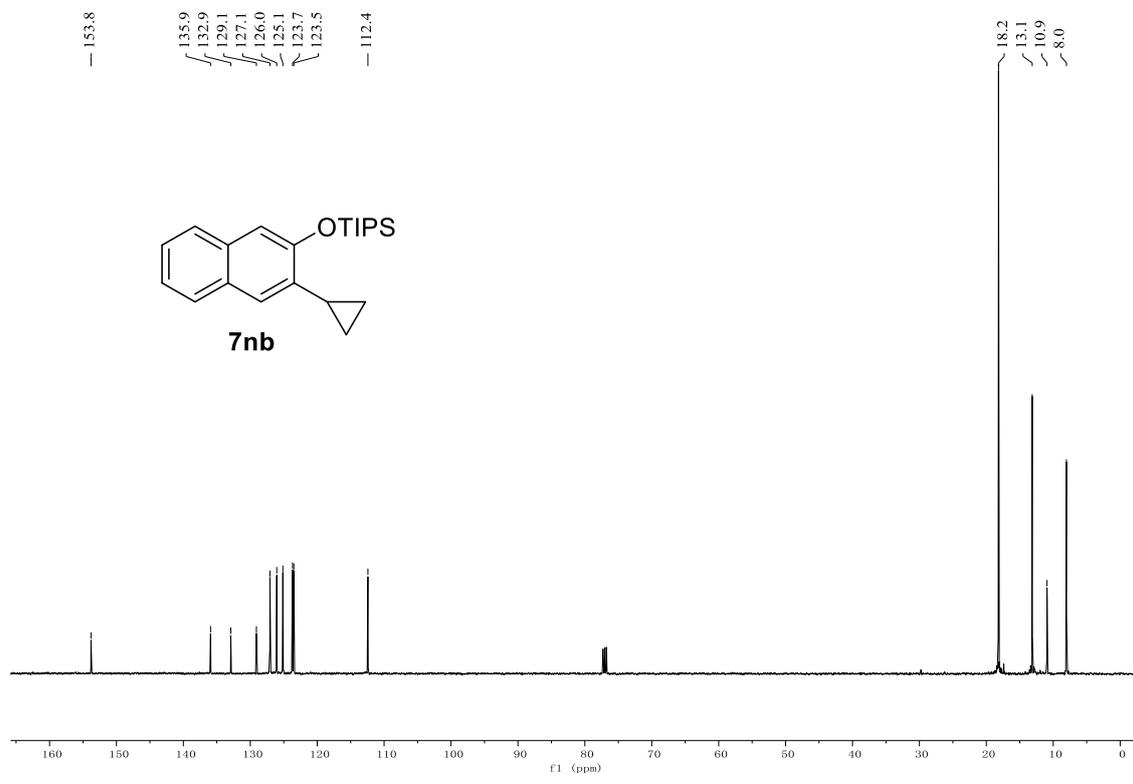
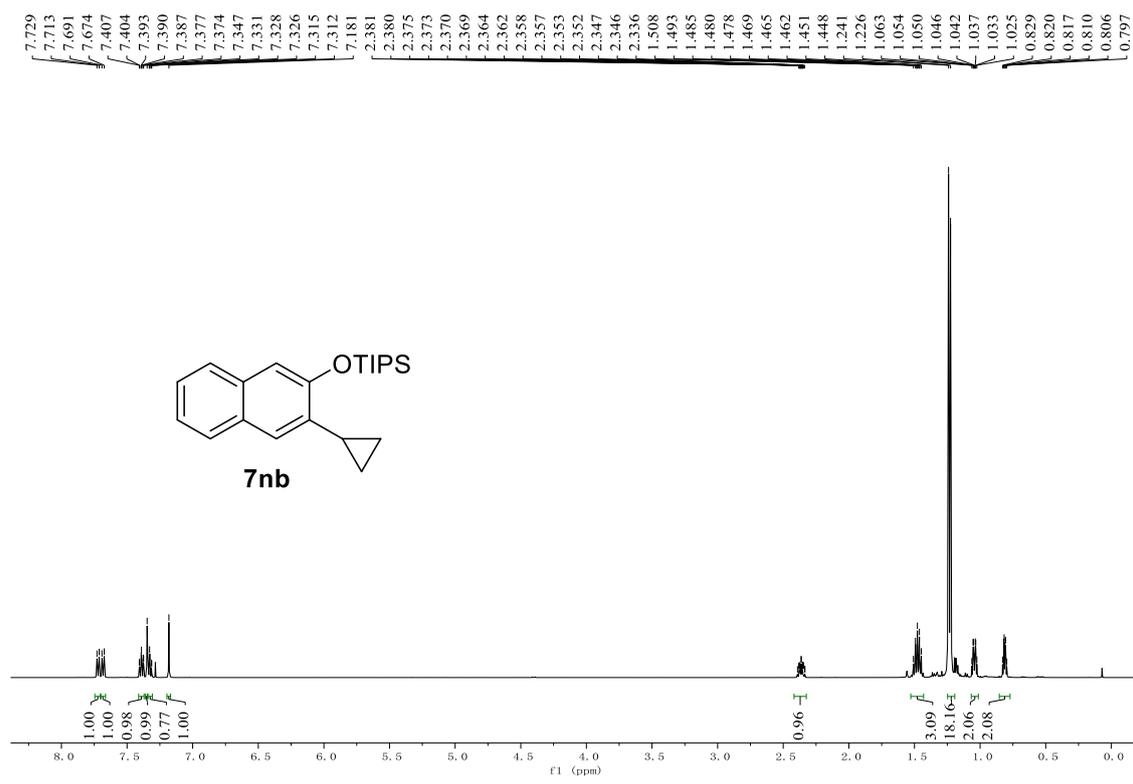


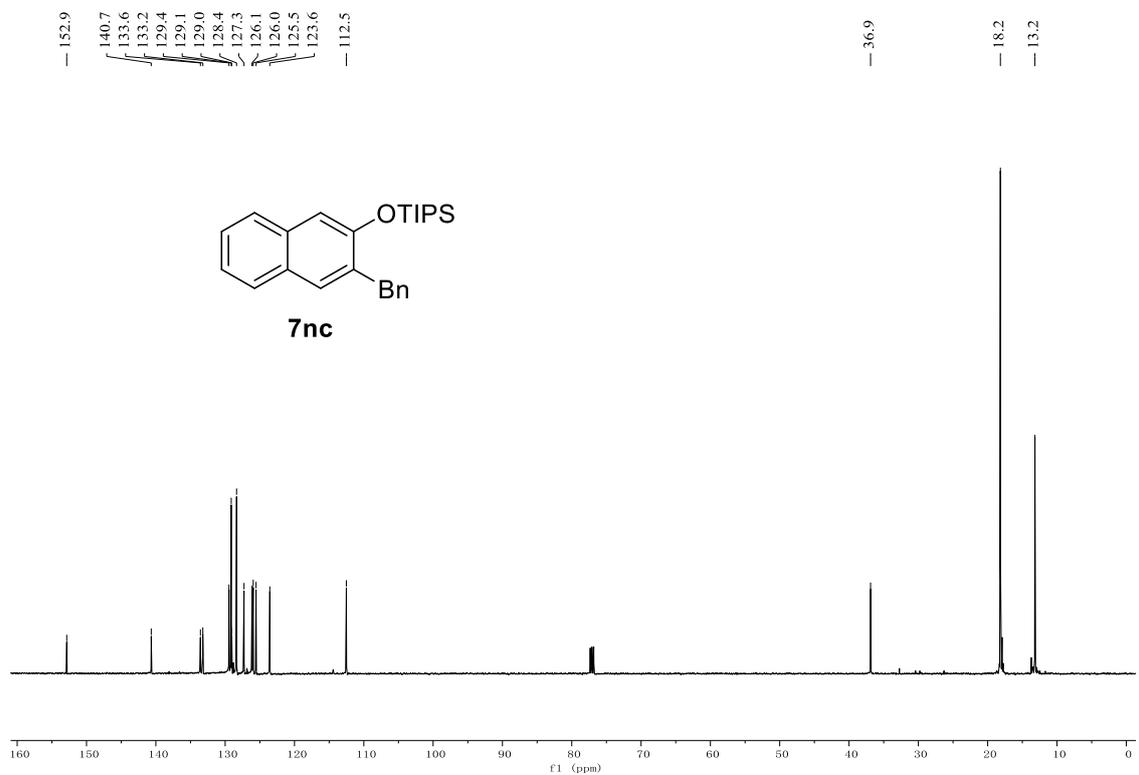
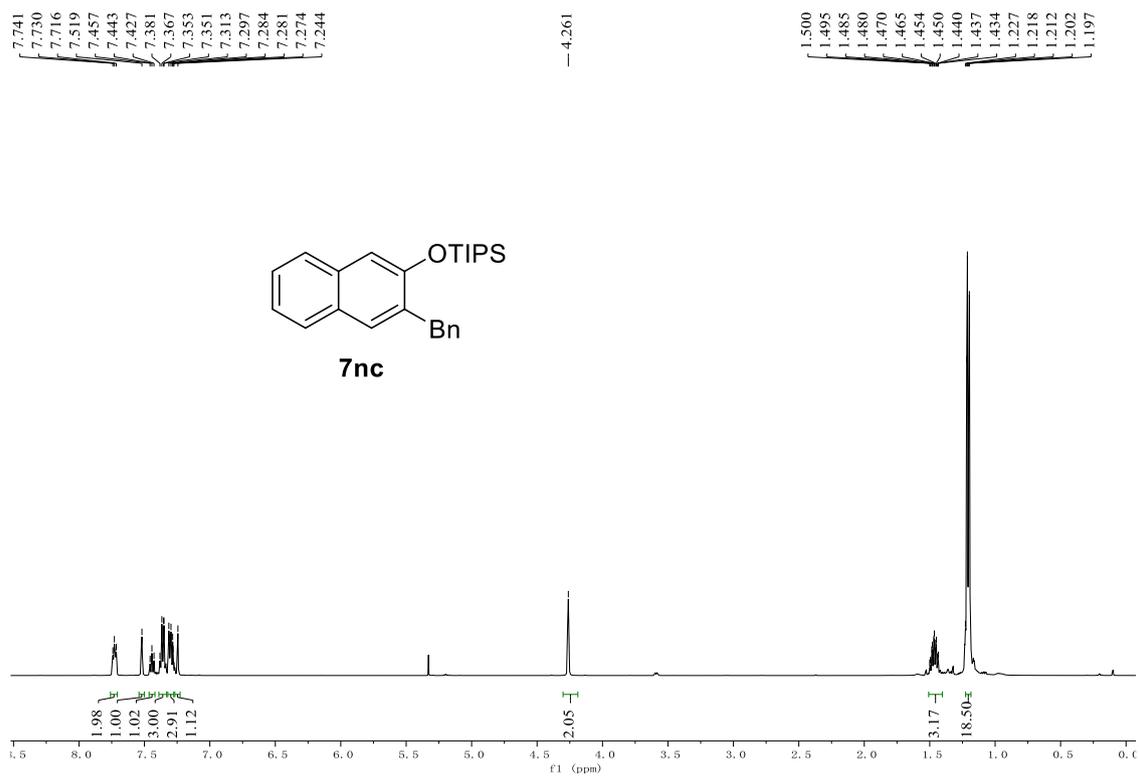


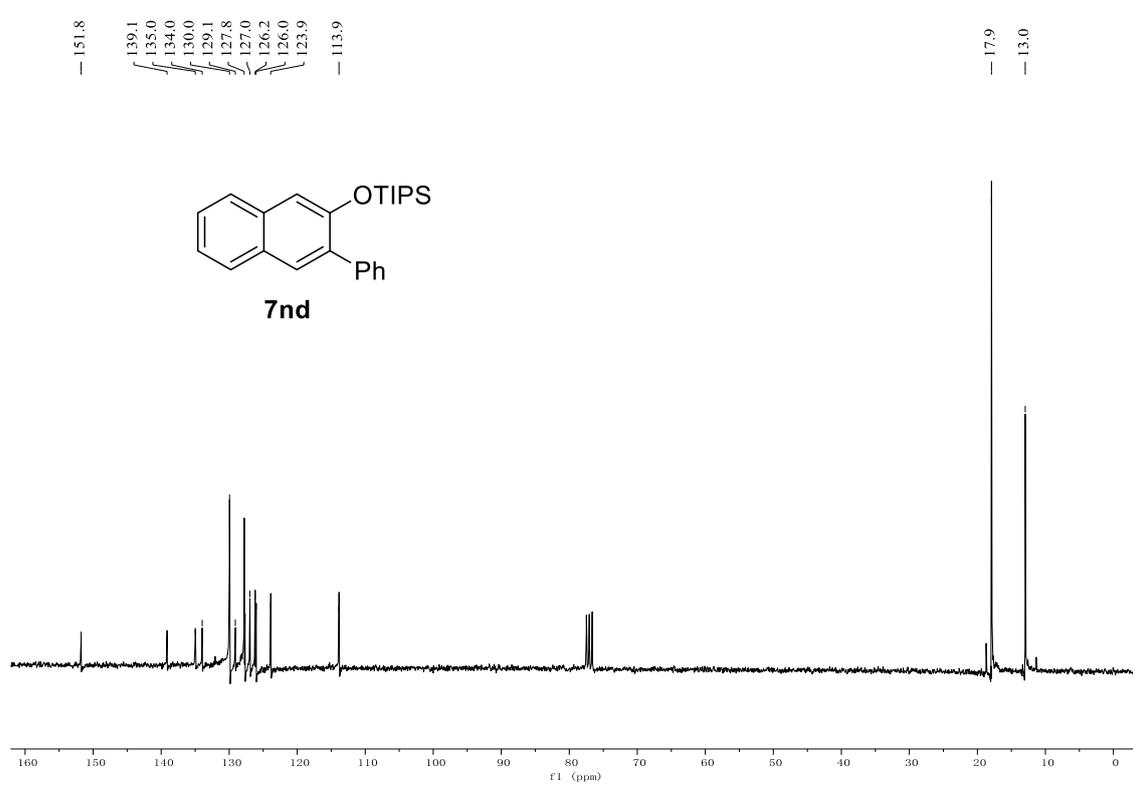
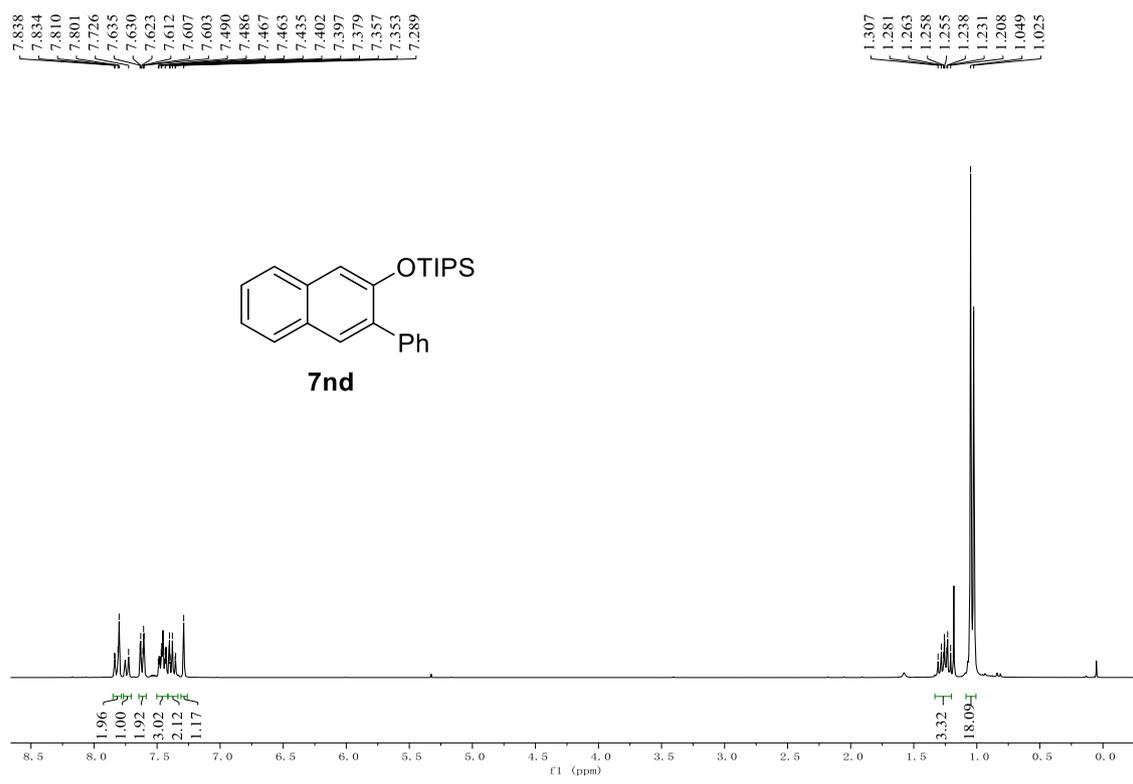


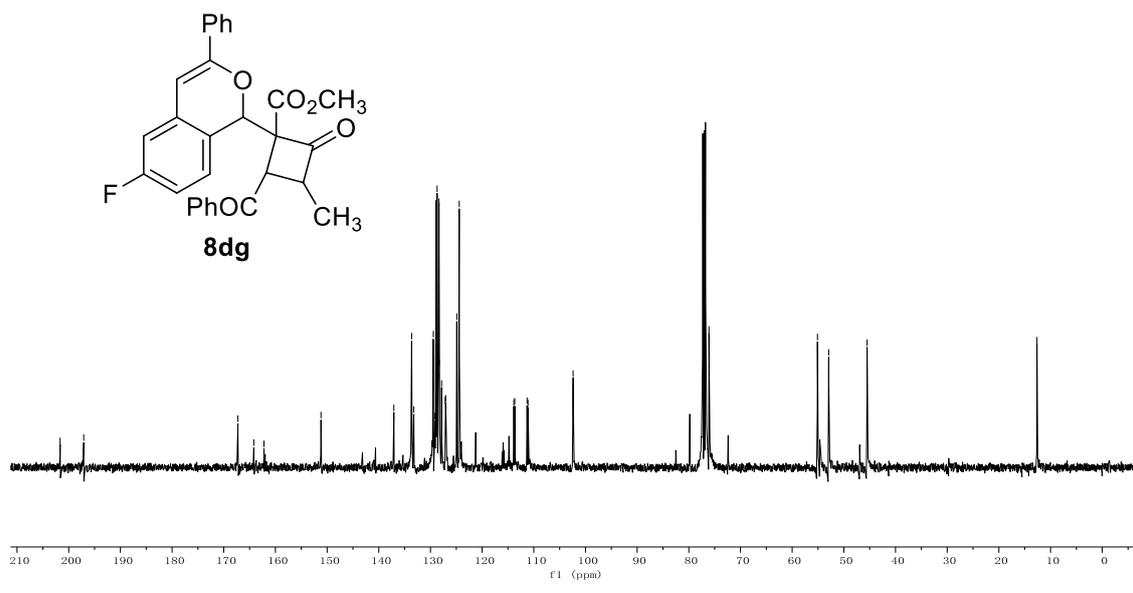
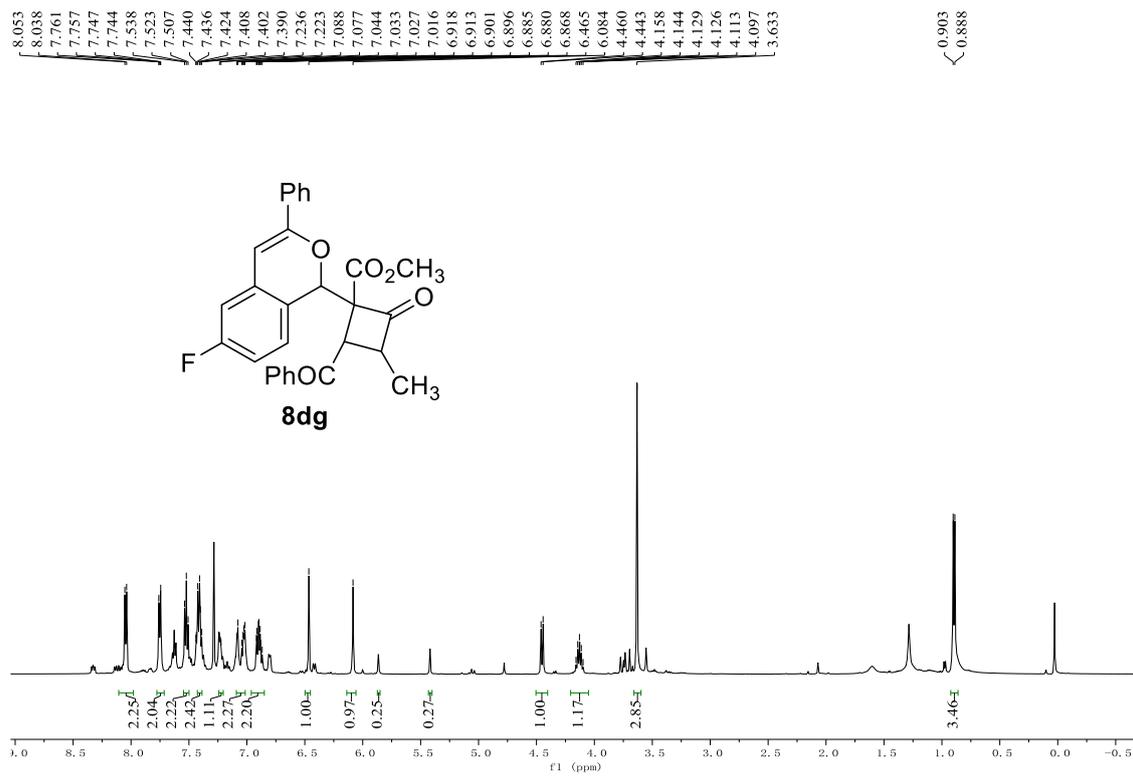


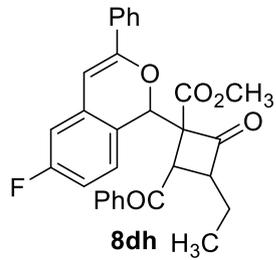
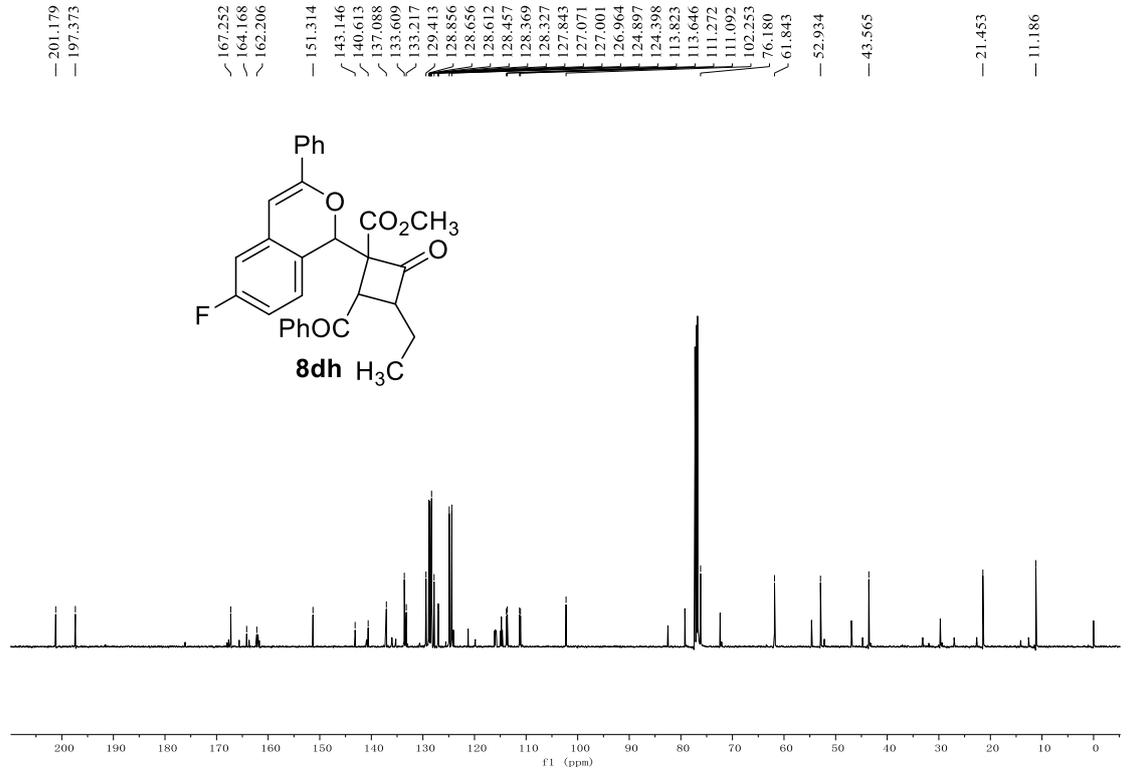
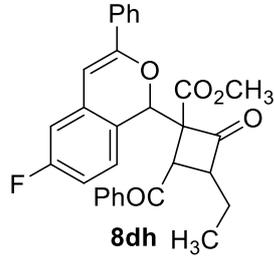
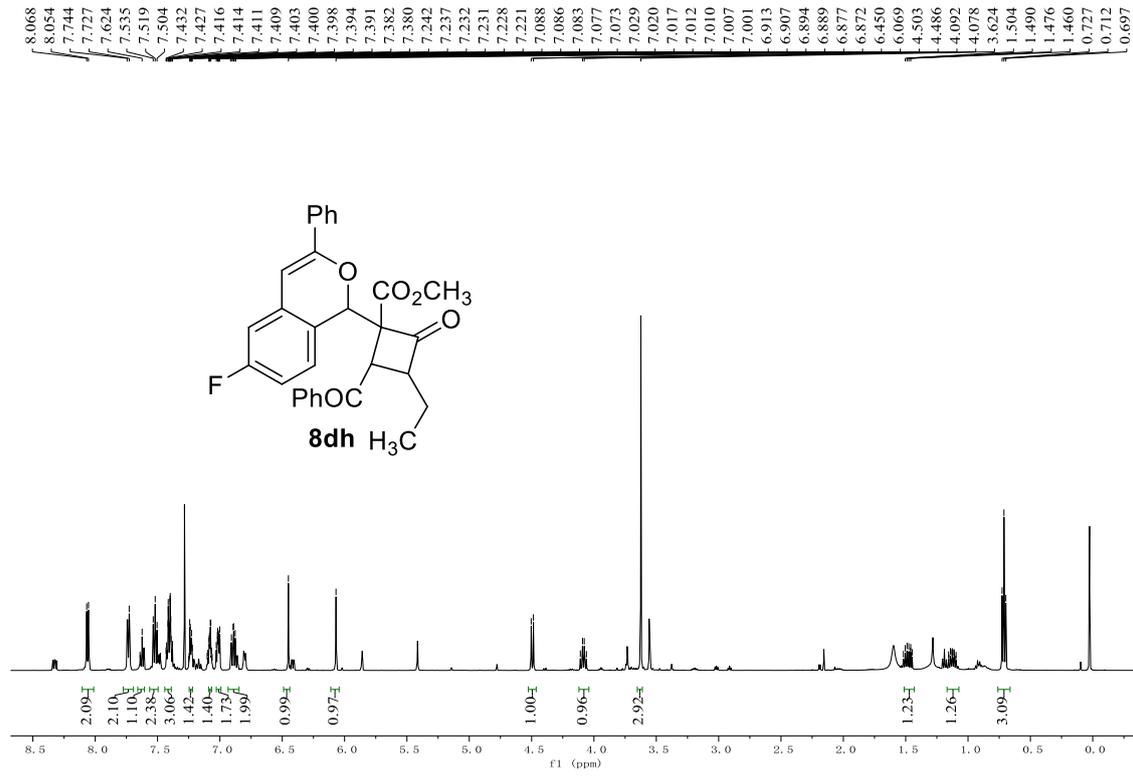


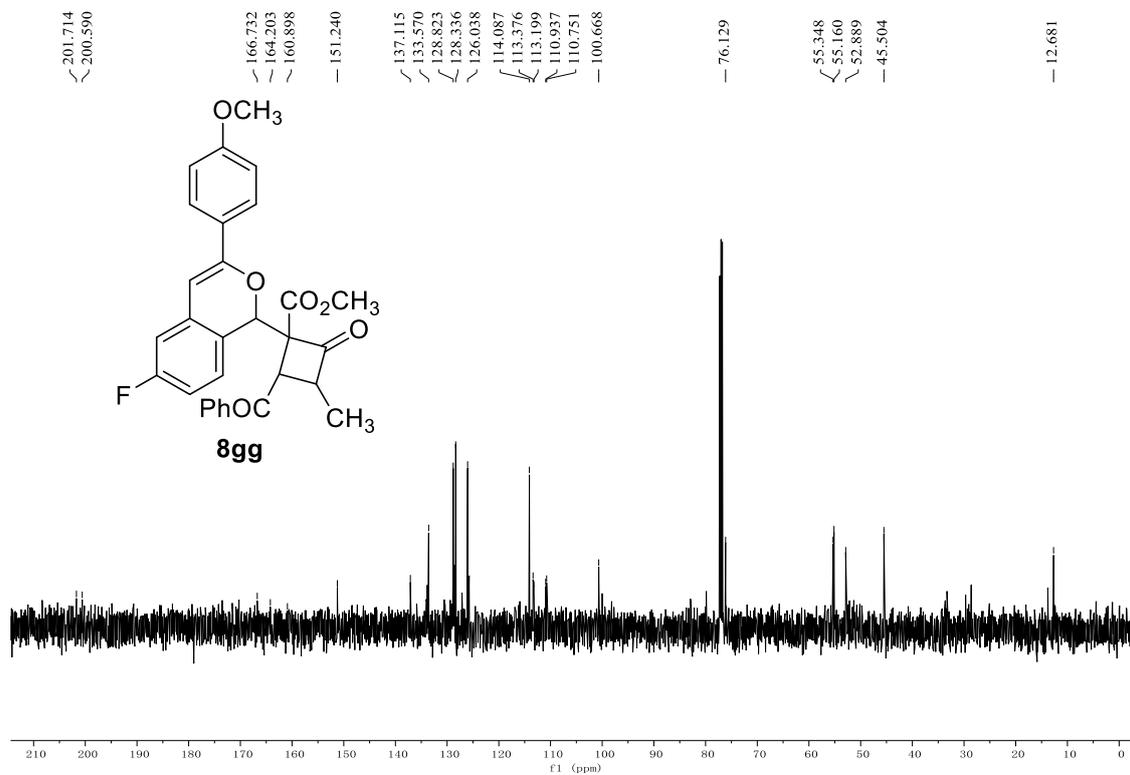
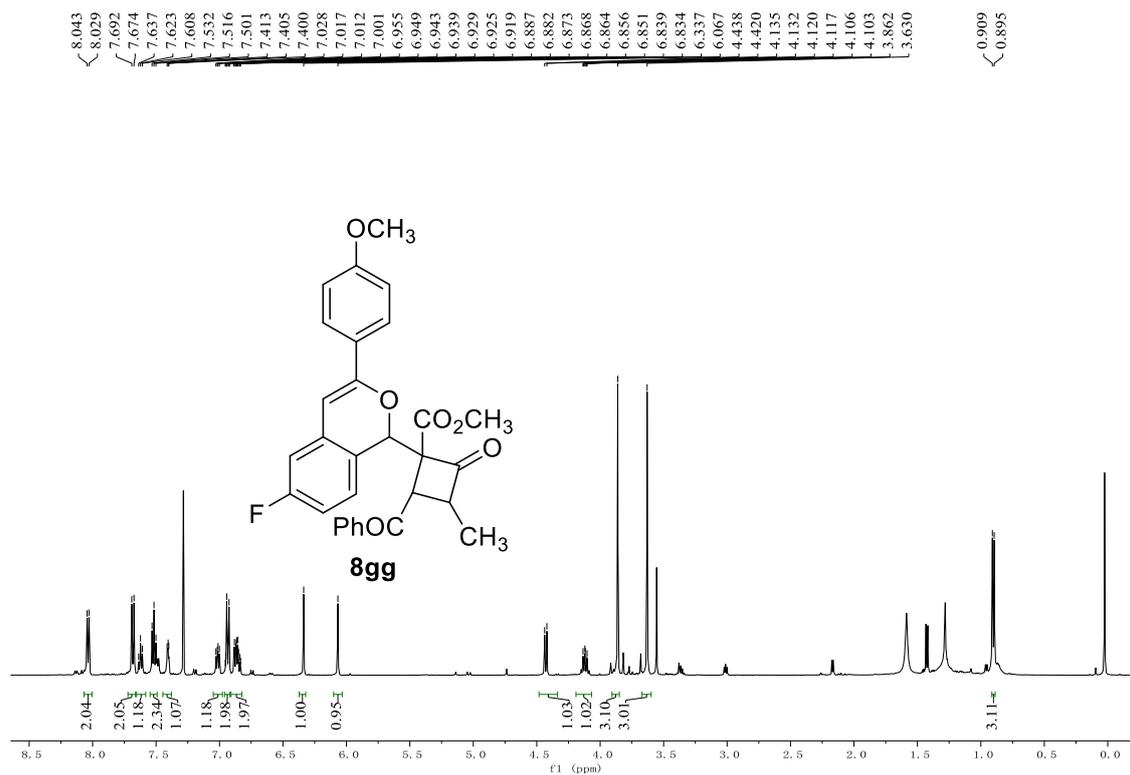


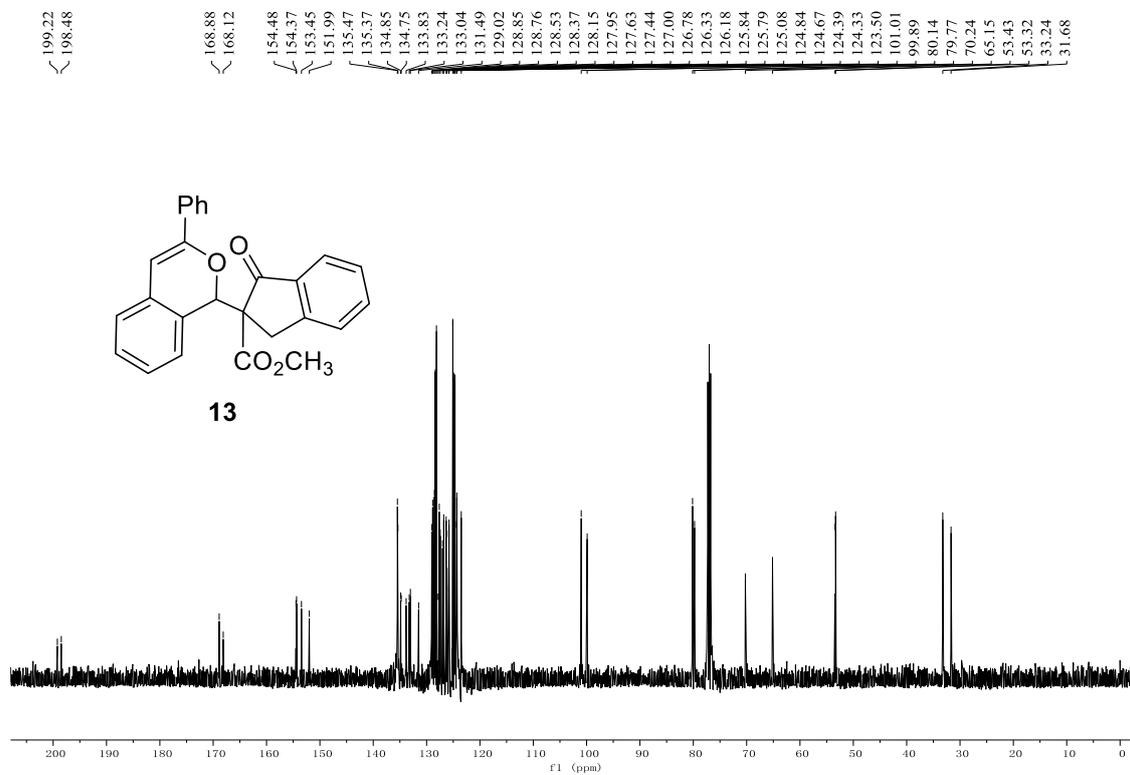
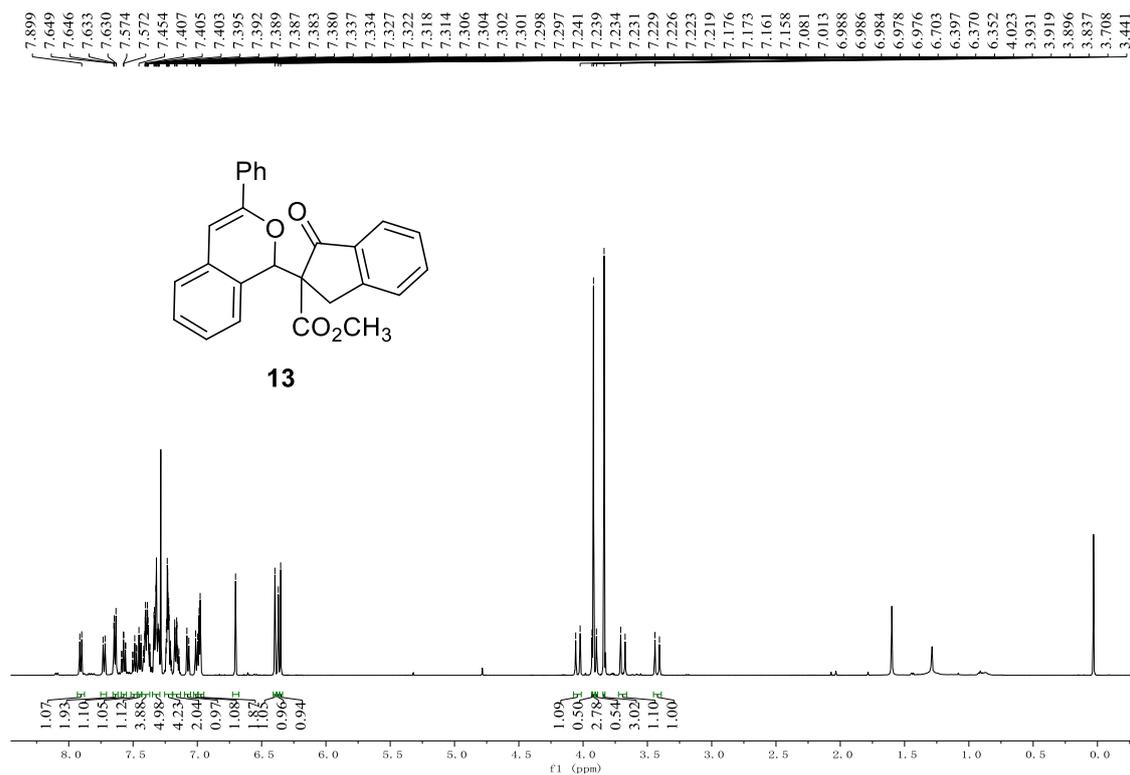


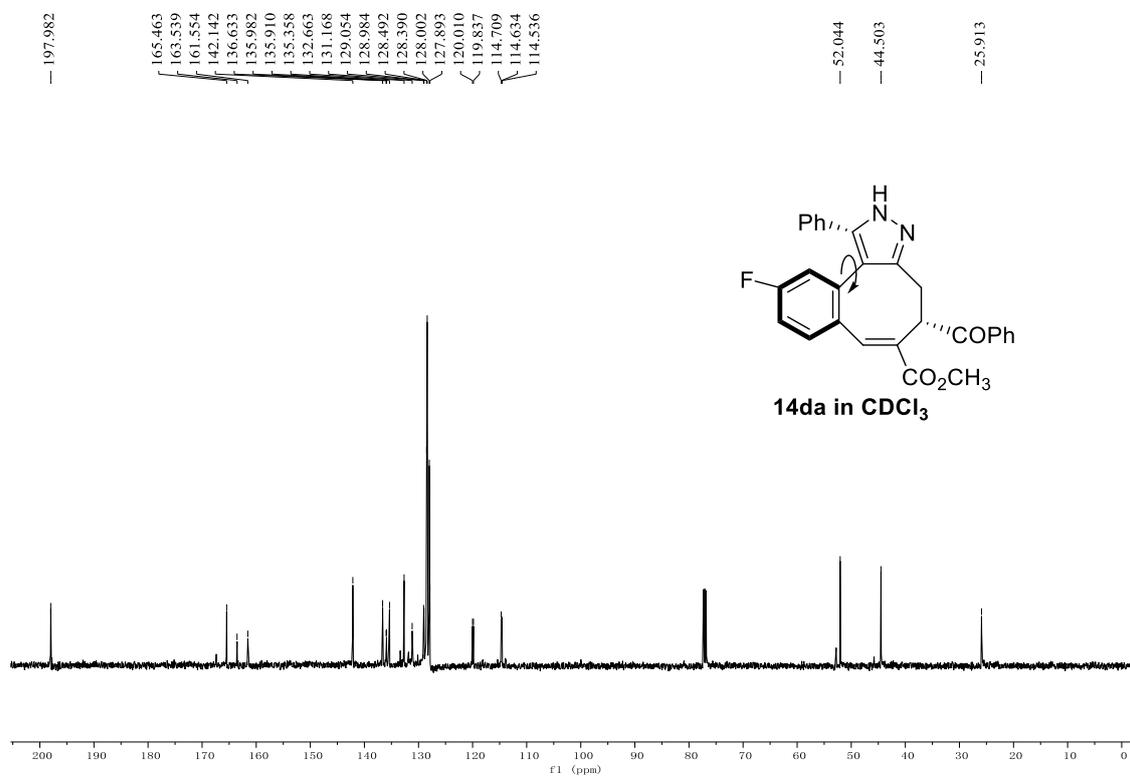
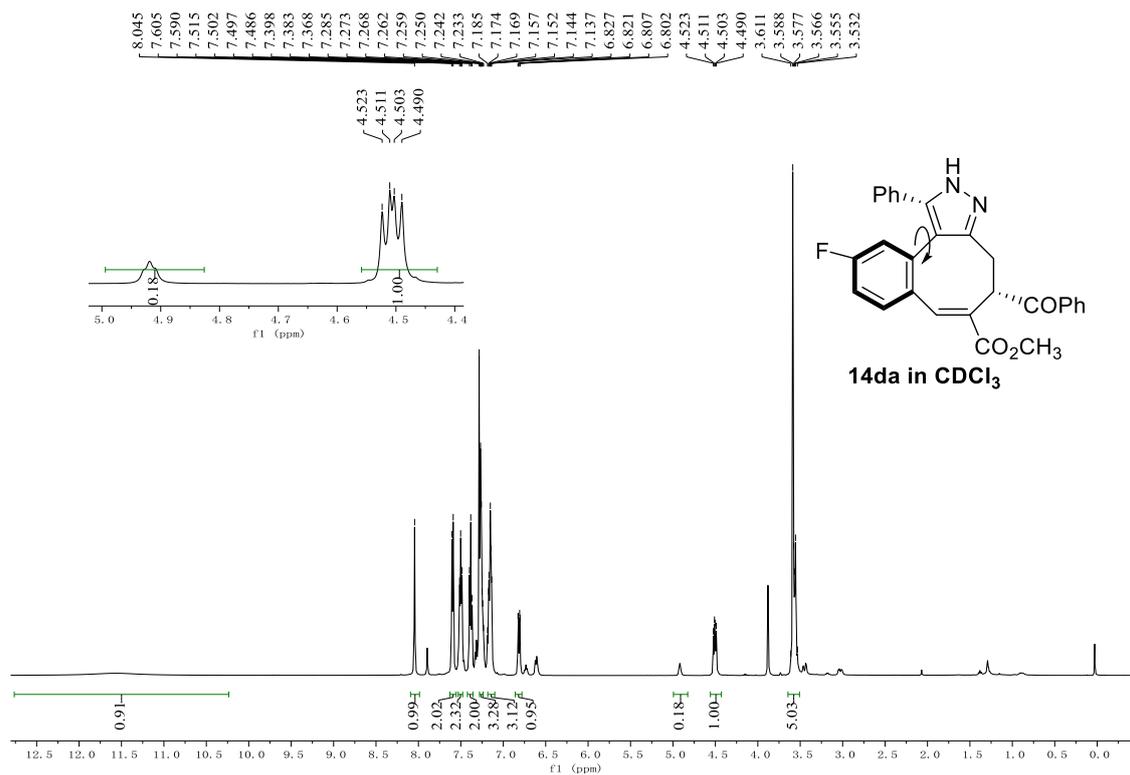


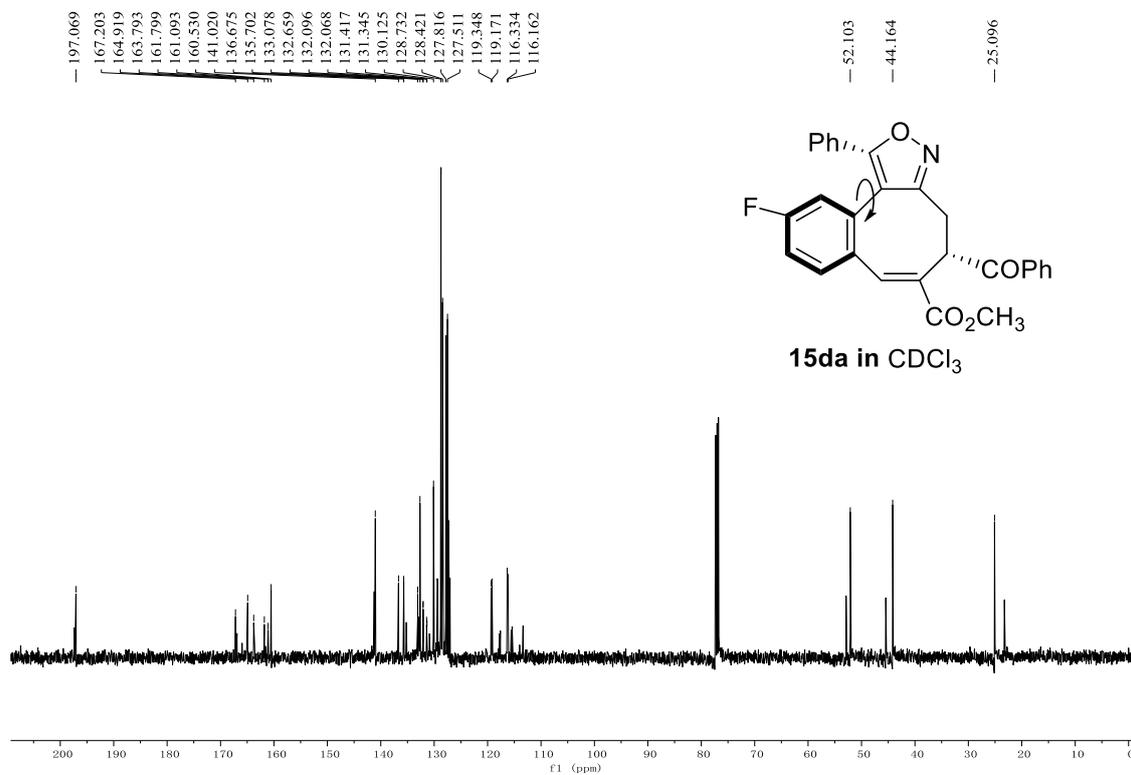
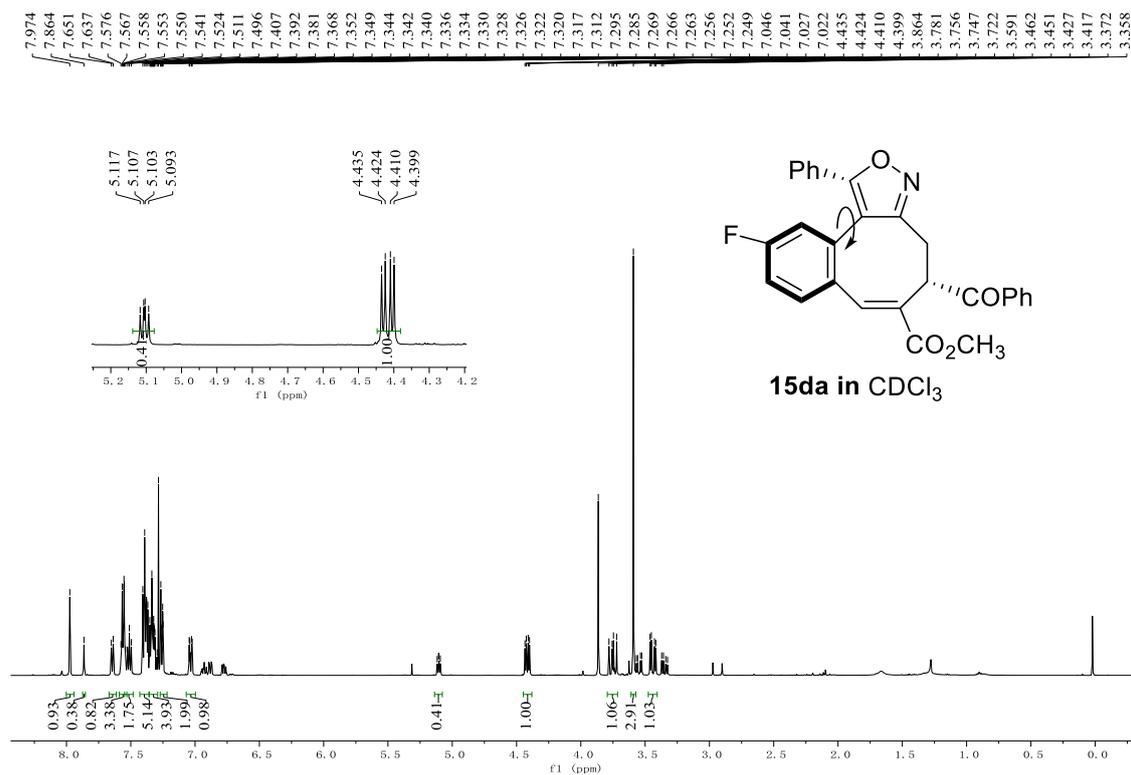


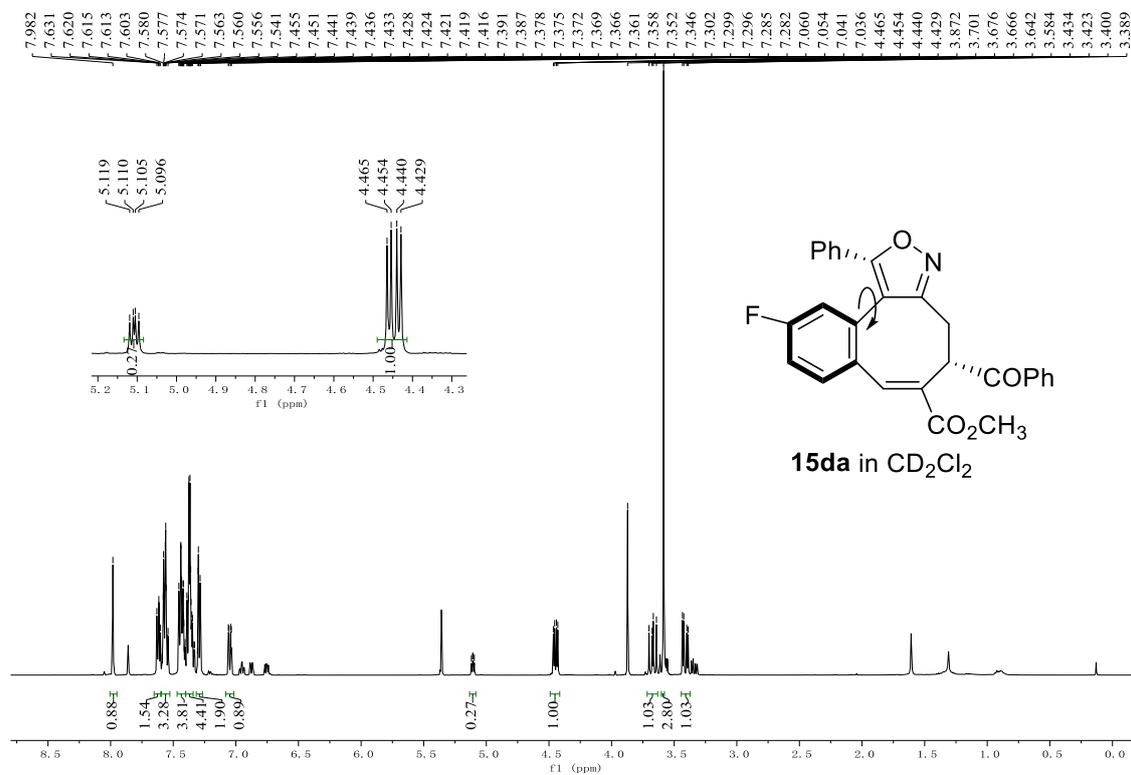
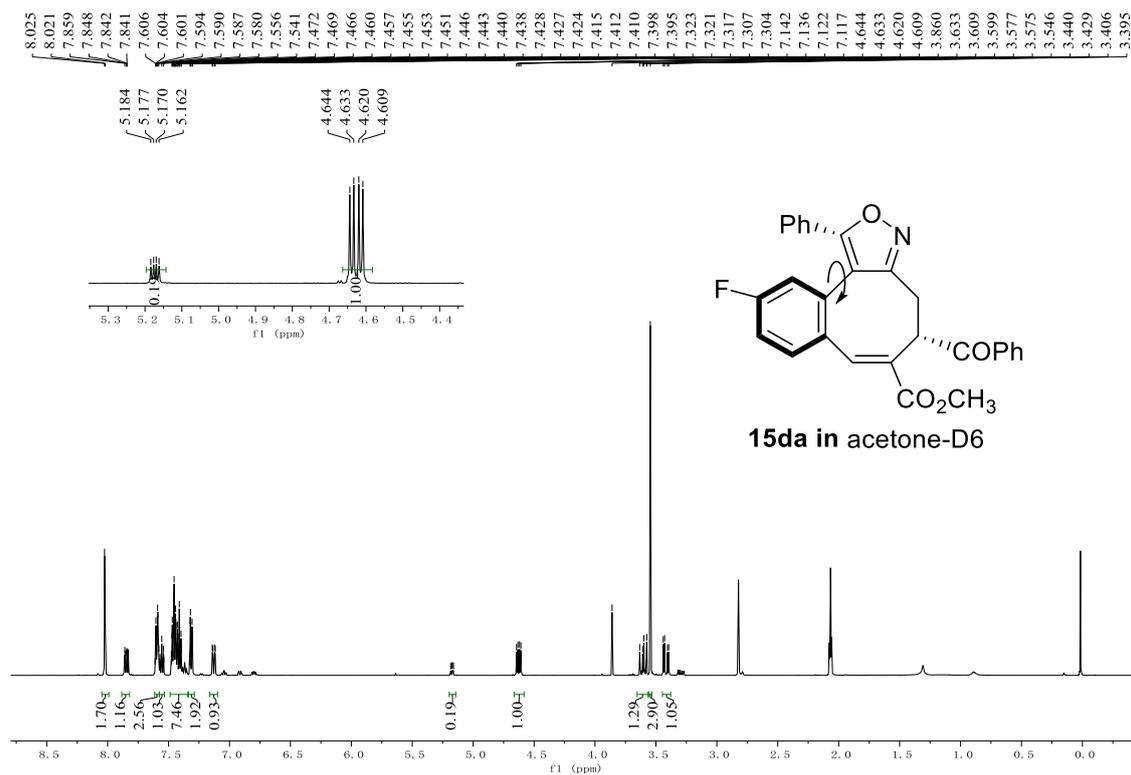


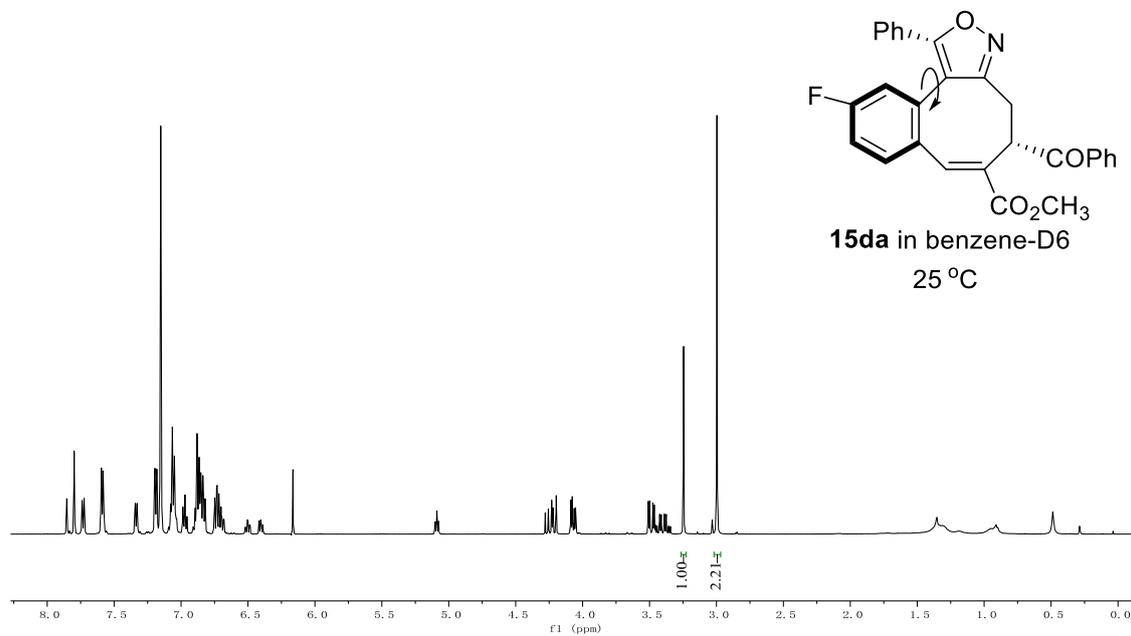
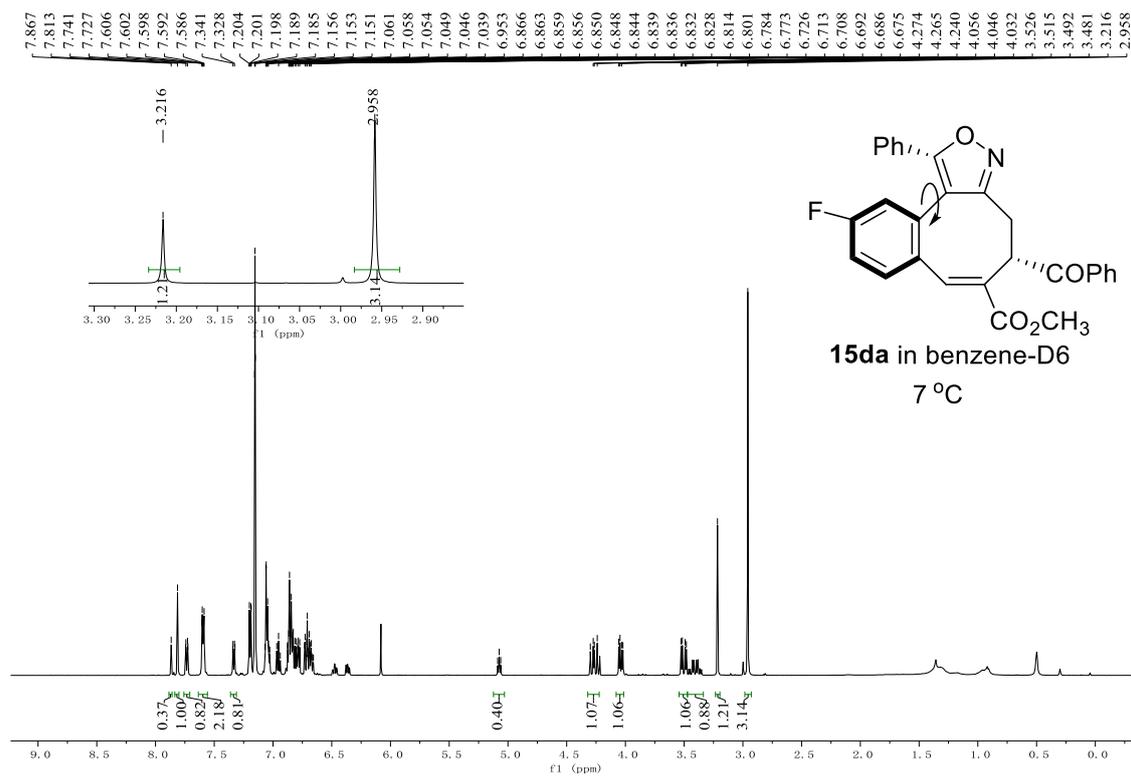


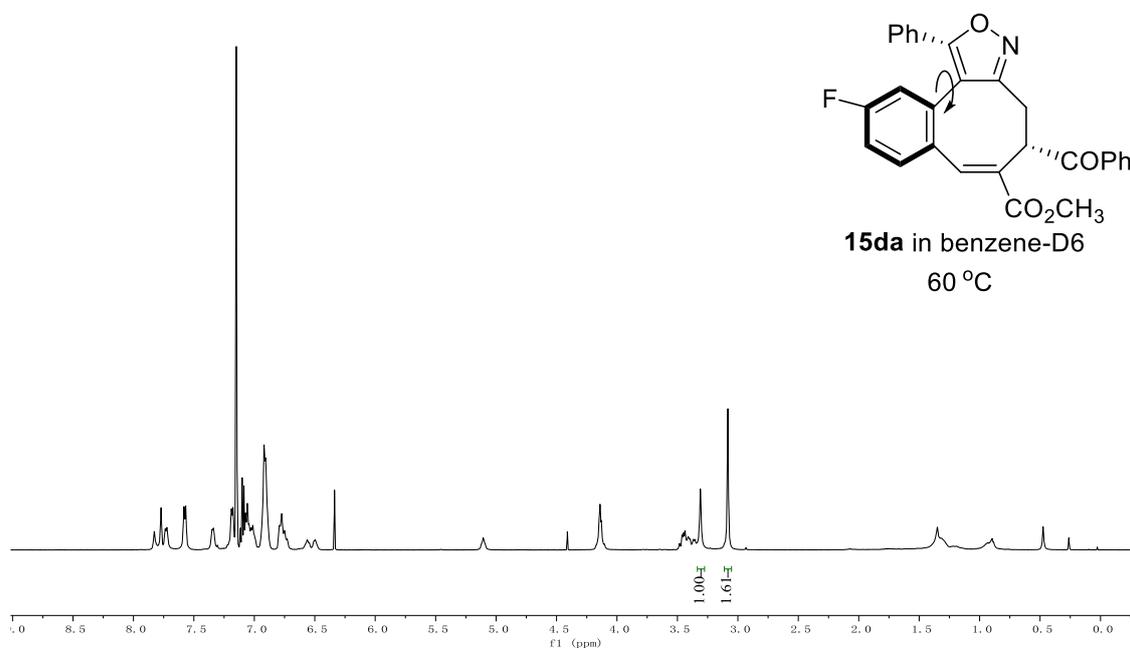
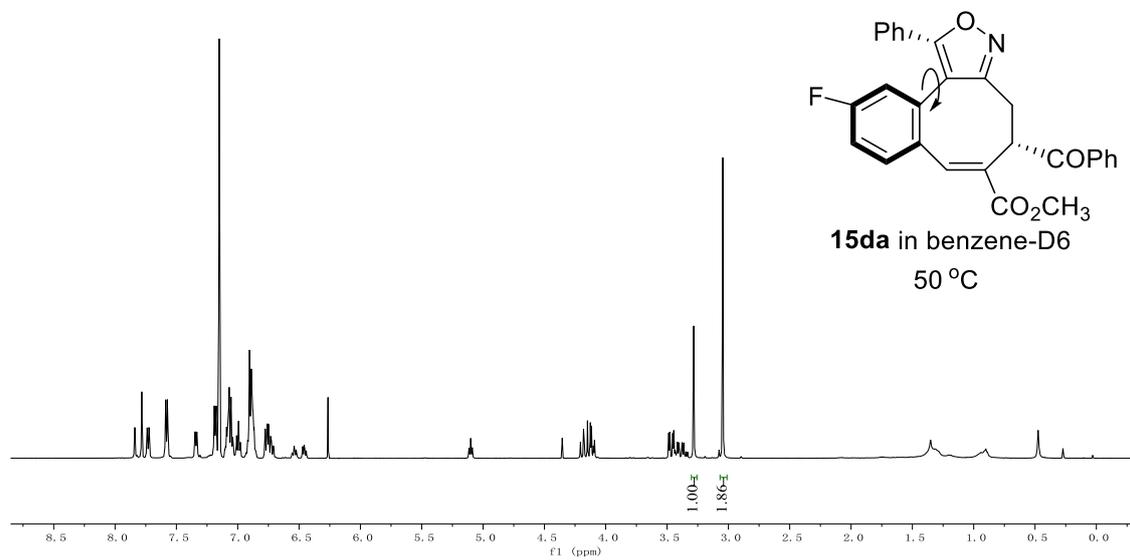


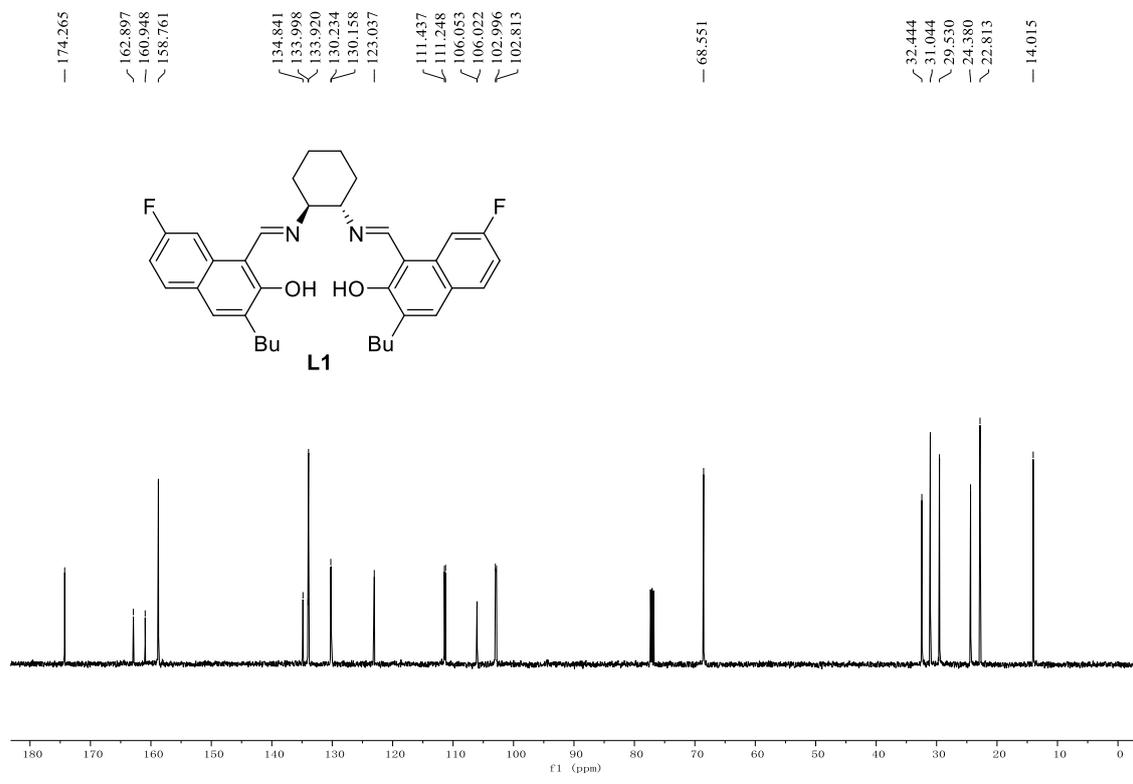
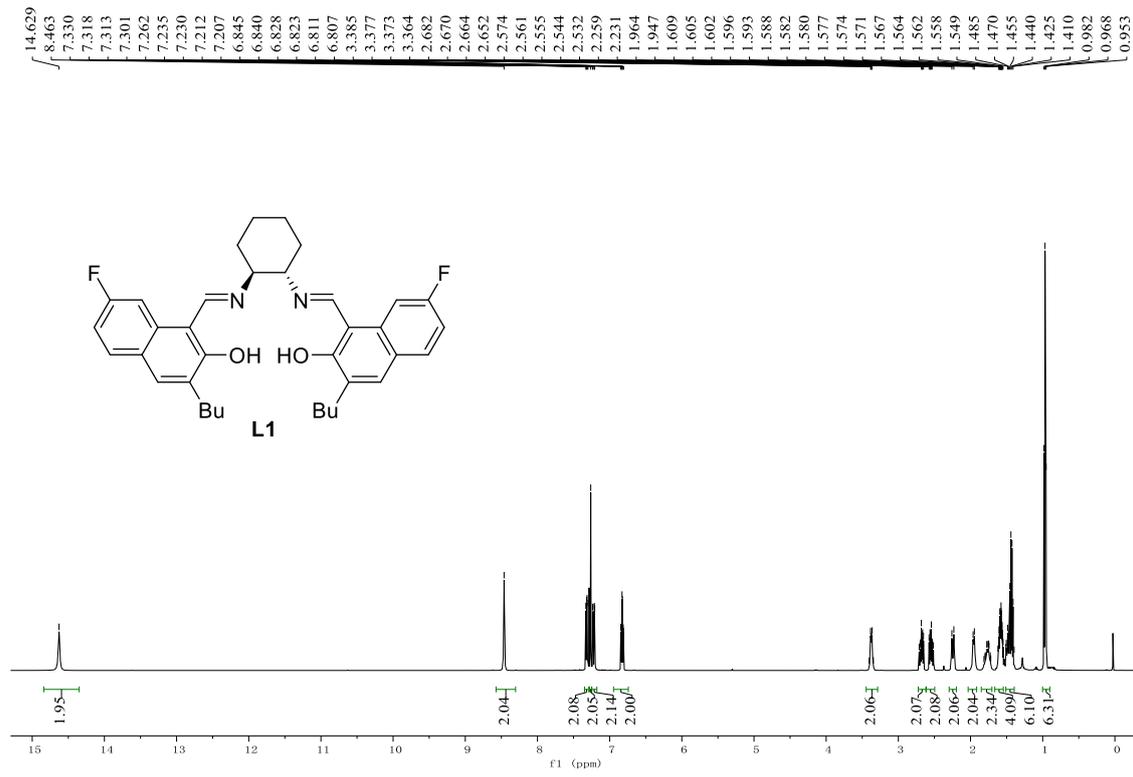


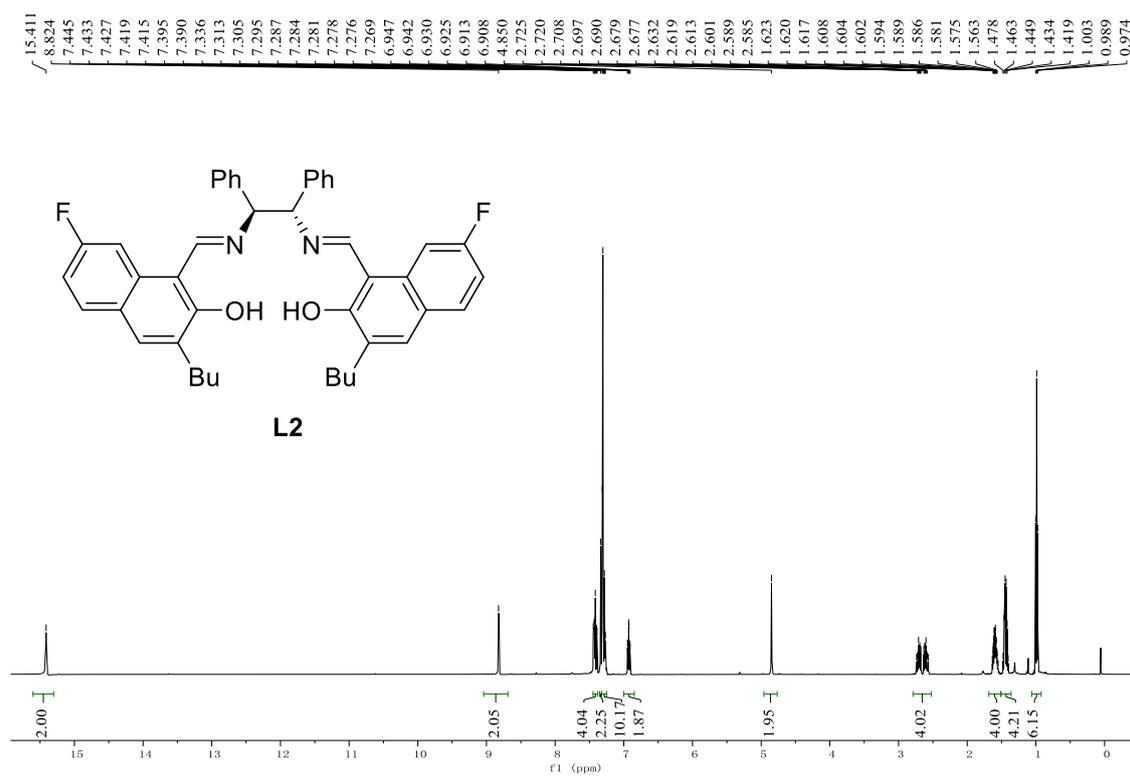
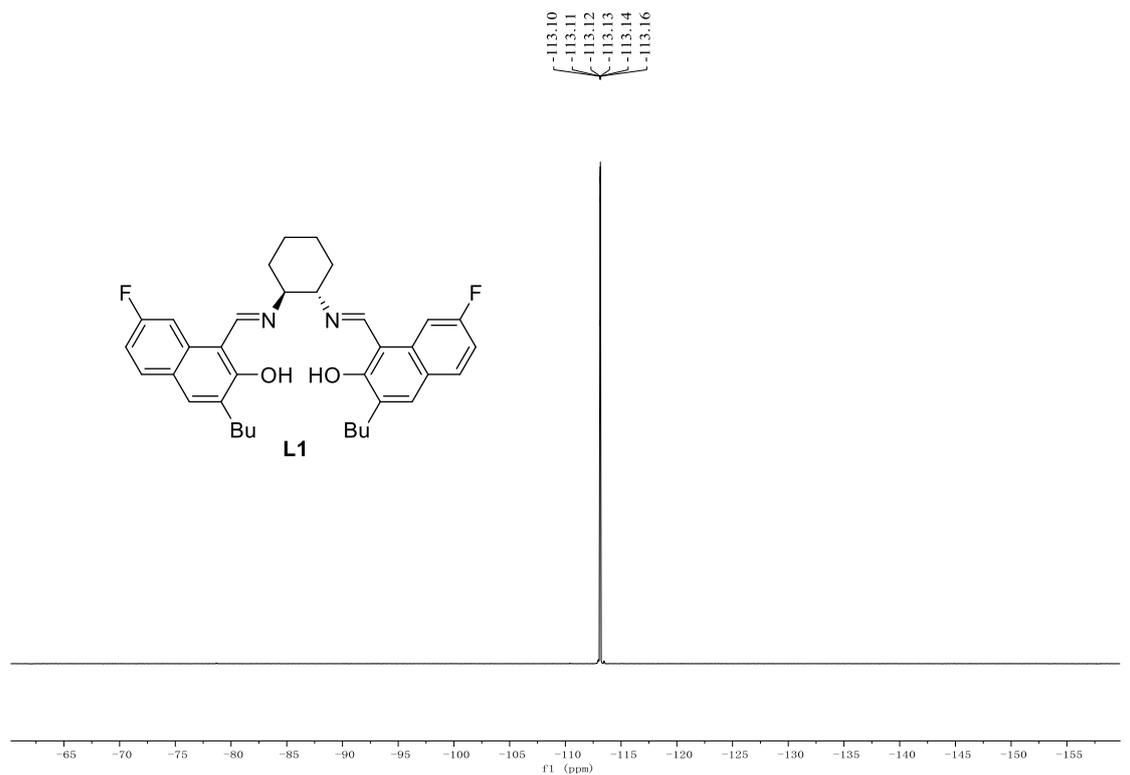


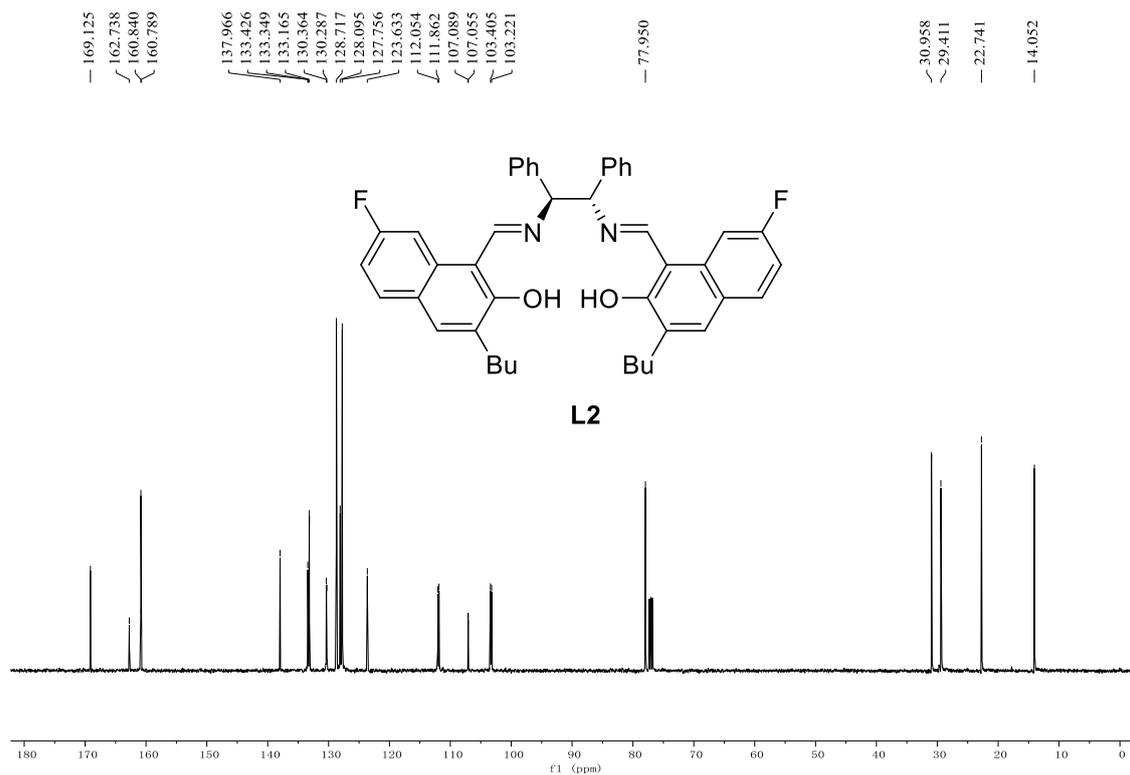






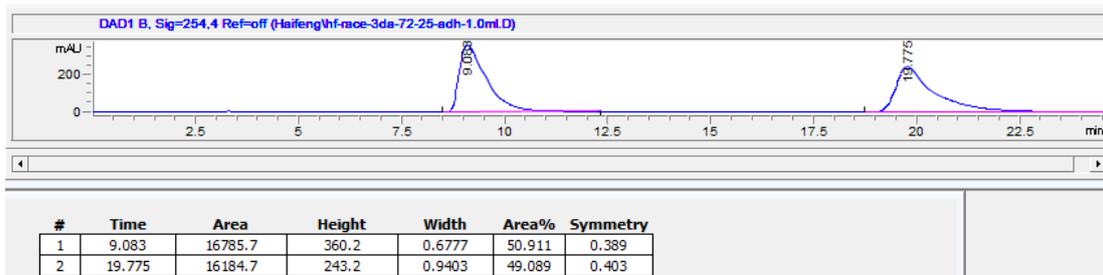
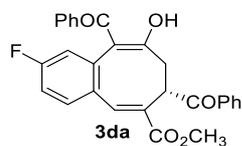




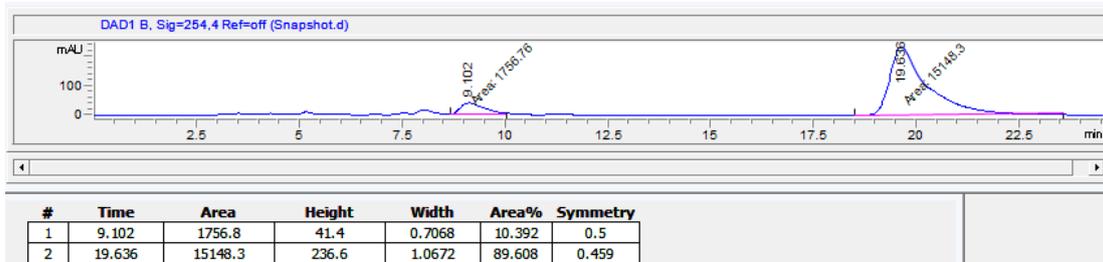


11. HPLC spectra of products

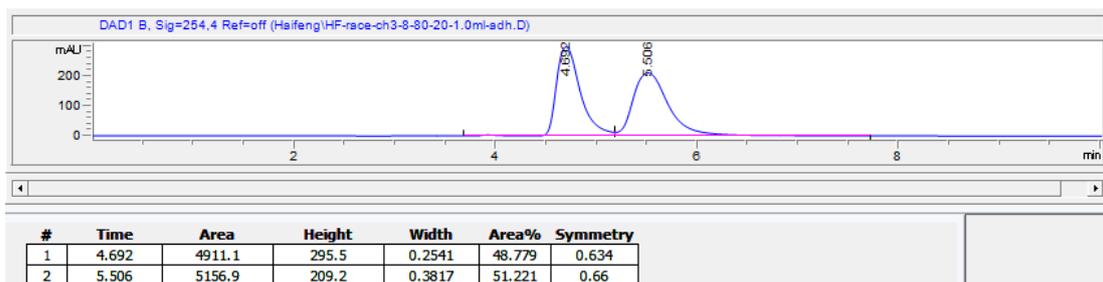
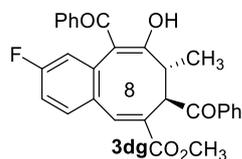
Chiral HPLC of racemic 3da



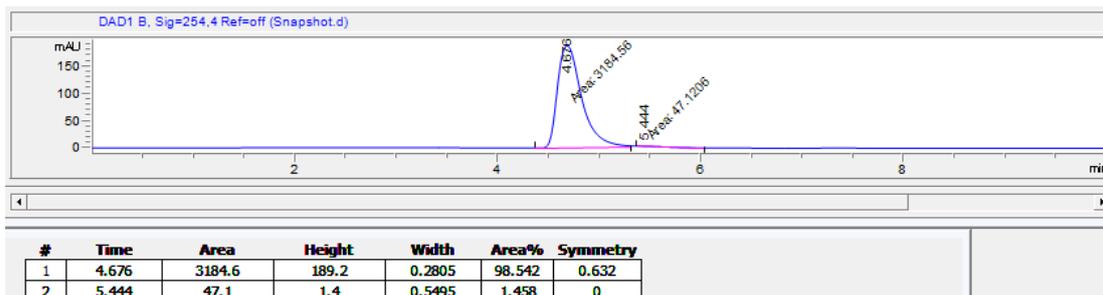
Chiral HPLC of chiral 3da



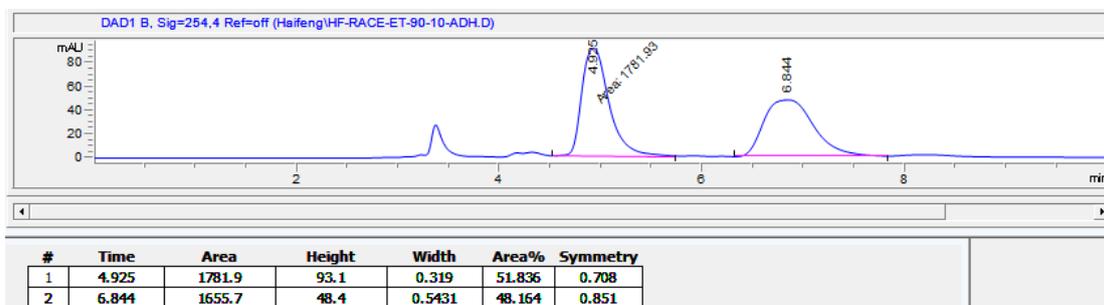
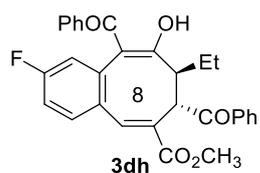
Chiral HPLC of racemic 3dg



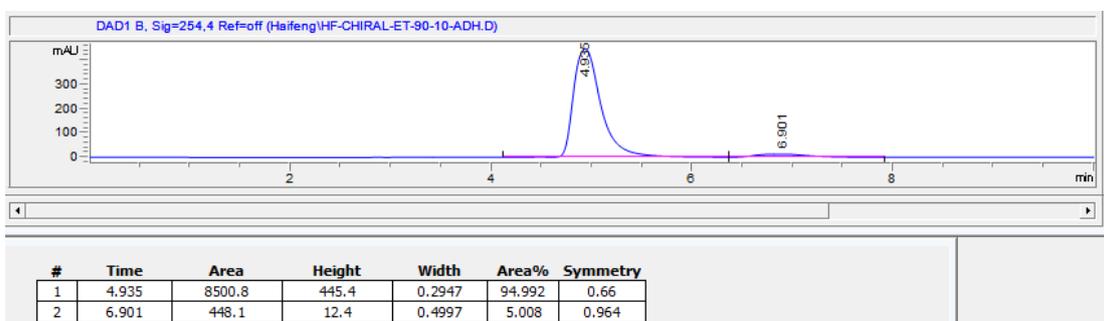
Chiral HPLC of chiral 3dg



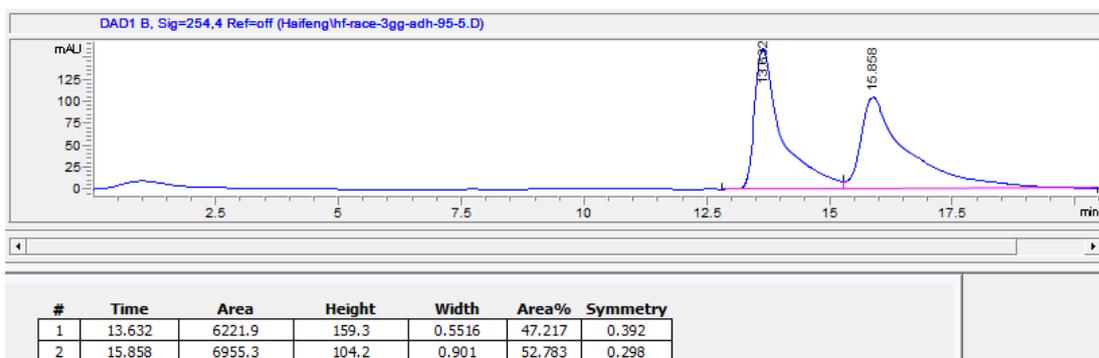
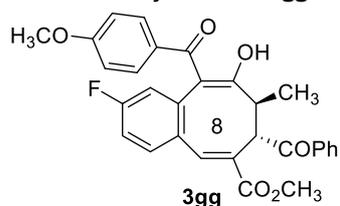
Chiral HPLC of racemic **3dh**



Chiral HPLC of chiral **3dh**



Chiral HPLC of racemic **3gg**



Chiral HPLC of chiral **3gg**

