Supporting Information for:

Machine Learning Designs Non-Hemolytic Antimicrobial Peptides

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Supporting information Note 1: Tables

Classifier	ROC AUC	Accuracy ^{a)}	Precision ^{a)}	Recall ^{a)}	F1 score ^{a)}	MCC ^{a)}
NB act.	0.55	0.55	0.59	0.32	0.42	0.11
SVM act.	0.75	0.68	0.68	0.68	0.68	0.36
RF act.	0.81	0.71	0.70	0.75	0.73	0.44
RNN act.	0.84	0.76	0.74	0.80	0.77	0.53
RNN scr. act.	0.51	0.49	0.35	0.03	0.05	-0.06
NB hem.	0.58	0.56	0.48	0.76	0.59	0.19
SVM hem.	0.69	0.73	0.72	0.58	0.65	0.44
RF hem.	0.80	0.77	0.81	0.60	0.69	0.53
RNN hem.	0.87	0.76	0.70	0.76	0.73	0.52
RNN scr. hem.	0.45	0.61	0.41	0.05	0.10	0.01

Table S1. Performance on the test of the NB, RF, SVM, RNN, and RNN with scrambled labels(RNN scr.) models for the AMP activity (act.) and hemolysis (hem.) classification tasks.

a) The probabilistic prediction values were converted into binary classification values using a threshold of 0.5. The best values and the selected classifiers are reported in bold.

Table S2. Classifiers optimization					
Classifier	Hyperparameters optimization ^{a)}				

SMV AMP activity	C = 0.1, 1, 10 , 100
	$\gamma = 0.1, 0.01, 0.001$
RF AMP activity	maximum depth = 10 , 30, 50, 70, 90, None
	no. estimators = 10, 100, 250, 500, 750, 1000 , 1500, 2000
RNN AMP activity	embedding dimensions = 2, 21, 42, 100
	GRU dimensions = 50, 100, 200, 300, 400
	no. layers = 1, 2 , 3
	epoch = [1, 2, 3,, 150]; best epoch = 38
RNN AMP activity	embedding dimensions 2 , 21, 42, 100
scrambled labels	GRU dimensions = 50 , 100, 200, 300, 400
	no. layers = 1 , 2, 3
	epoch = [1, 2, 3,, 150]; best epoch = 1
SMV Hemolysis	C = 0.1, 1 , 10, 100
	$\gamma = 0.1, 0.01, 0.001$
RF Hemolysis	maximum depth = 10 , 30, 50, 70, 90, None
	no. estimators = 10, 100, 250, 500 , 750, 1000, 1500, 2000
RNN Hemolysis	embedding dimensions = $2, 21, 42, 100$
	GRU dimensions = 50, 100, 200, 300, 400

no. layers = 1, 2, 3

epoch = [1, 2, 3, ..., 150]; best epoch = **95**

RNN Hemolysis	embedding dimensions 2, 21, 42, 100		
scrambled labels	GRU dimensions = 50 , 100, 200, 300, 400		
	no. layers = 1, 2 , 3		
	epoch = [1, 2, 3,, 150]; best epoch = 150		

a) The selected hyperparameters are highlighted in bold. All hyperparameters that have not been discussed have been used in their default values.

Supporting information Note 2: Figures



Figure S1. Properties distribution and filters. (a) length, (b-e) minimum Levenshtein distance (minLD) from training and test sets, (f) presence of D-amino acids (D-AA), (g, h) Amphiphilic helix estimation, of the 3,046 predicted active and non-hemolytic sequences derived from the model fined tuned for *A. baumannii* and *P. aeruginosa* (Generated GN) and the 2,717 predicted active and non-hemolytic sequences derived from the model finetuned for *S. aureus*. (a-d) Solid vertical lines indicate that the threshold values were included. (e, g, h) Dashed vertical lines indicated that the threshold lines were excluded.



Figure S2. RNN AMP activity classifier architecture. The tokenized and "one-hot" encoded sequences enter an 100 dimensions (dim) embedding layer (a), then they are processed through two layers of 400 dimensions GRU cells (b), and finally, a linear transformation layer shapes the last GRU output into two dimensions (c), followed by a softmax function that normalizes it into a probability (d). The architecture of the hemolytic classifier differs only by having one layer of GRU cells.



Figure S3. RNN generative models architecture. The tokenized and "one-hot" encoded sequences enter an 100 dimensions (dim) embedding layer (a), then they are processed through two layers of 400 dimensions GRU cells (b), and finally, a linear transformation layer shapes the last GRU output into 41 dimensions (c), followed by a softmax function that normalizes it into a probability (d). The architecture of the hemolytic classifier differs only by having one layer of GRU cells.



Figure S4: MD simulations of **GP1** in water and in presence of a DPC micelle over 250 ns using GROMACS. (a) Average structure (stick model) in water over 100 structures sampled over the last 100 ns (thin lines). Hydrophobic side chains are colored in red and cationic side chains in blue. (b) Average structure (cartoon model for backbone and stick model for side chains) with DPC micelle over 100 structures sampled over the last 100 ns (thin lines). (c) RMSD (root mean square deviation) of the peptide backbone atoms relative to the starting α -helical conformation. (d) Number of intramolecular hydrogen bonds. The DPC micelle was omitted for clarity.



Figure S5: MD simulations of **GN2** in water and in presence of a DPC micelle over 250 ns using GROMACS. (a) Average structure (stick model) in water over 100 structures sampled over the last 100 ns (thin lines). Hydrophobic side chains are colored in red and cationic side chains in blue. (b) Average structure (cartoon model for backbone and stick model for side chains) with DPC micelle over 100 structures sampled over the last 100 ns (thin lines). (c) RMSD (root mean square deviation) of the peptide backbone atoms relative to the starting α -helical conformation. (d) Number of intramolecular hydrogen bonds. The DPC micelle was omitted for clarity.

Supporting information Note 3: HPLC/MS and HRMS spectra

GN1 (AKRIRKLIKKIFKKI-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (302.5 mg, 9.6%). Analytical RP-HPLC: $t_R = 1.35 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): C₉₀H₁₆₈N₂₈O₁₅ calc./obs. 1881.32/1881.33 [M]⁺.





GN2 (RRWKWRRKIKKWL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (308.4 mg, 39.0%). Analytical RP-HPLC: $t_R = 1.29 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{93}H_{151}N_{33}O_{13}$ calc./obs. 1938.22/1938.22 [M]⁺.





GN3 (IDKWKAAFKKIKNLF-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (263.4 mg, 4.9%). Analytical RP-HPLC: $t_R = 1.48 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{91}H_{145}N_{23}O_{18}$ calc./obs. 1848.11/1848.11 [M]⁺.





GN4 (LNALKKVFQKIRQGL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (241.8 mg, 28.8%). Analytical RP-HPLC: $t_R = 1.53 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{81}H_{143}N_{25}O_{18}$ calc./obs. 1745.10/1745.11 [M]⁺.





GN5 (KFFRKLKKLVKK-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (257.3 mg, 33.5%). Analytical RP-HPLC: $t_R = 1.24 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{77}H_{136}N_{22}O_{12}$ calc./obs. 1561.07/1561.07 [M]⁺.





GN6 (RLRKKWRKLKKLL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (290.3 mg, 31.4%). Analytical RP-HPLC: $t_R = 1.30 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{83}H_{153}N_{29}O_{13}$ calc./obs. 1764.22/1764.22 [M]⁺.





GN7 (KRIRKWVRRILKKL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (291.7 mg, 19.4%). Analytical RP-HPLC: $t_R = 1.37 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{88}H_{162}N_{32}O_{14}$ calc./obs. 1891.29/1891.30 [M]⁺.





GN8 (LRKFWKKIRKFLKKI-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (372.0 mg, 17.2%). Analytical RP-HPLC: $t_R = 1.46 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{101}H_{171}N_{29}O_{15}$ calc./obs. 2030.35/2030.35 [M]⁺.





GN9 (KRLWKRIYRLLKK-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (282.0 mg, 8.8%). Analytical RP-HPLC: $t_R = 1.36 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{86}H_{150}N_{28}O_{14}$ calc./obs. 1799.19/1799.18 [M]⁺.





GN10 (IRRIRKKIKKIFKKI-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (335.0 mg, 8.2%). Analytical RP-HPLC: $t_R = 1.26 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{93}H_{175}N_{31}O_{15}$ calc./obs. 1966.39/1966.39 [M]⁺.





GN11 (LRKARRLLKKLRARL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (303.3 mg, 21.4%). Analytical RP-HPLC: $t_R = 1.28 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{84}H_{164}N_{34}O_{15}$ calc./obs. 1889.31/1889.31 [M]⁺.





GN12 (GNWRKIVHKIKKAG-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (252.9 mg, 36.9%). Analytical RP-HPLC: $t_R = 1.24 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{75}H_{128}N_{26}O_{15}$ calc./obs. 1633.01/1633.01 [M]⁺.





GN13 (AGRLQKVFKVIAK-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (210.8 mg, 23.7%). Analytical RP-HPLC: $t_R = 1.33 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{68}H_{121}N_{21}O_{14}$ calc./obs. 1455.94/1455.94 [M]⁺.





GN14 (IHKLAKLAKNVL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (199.4 mg, 18.0%). Analytical RP-HPLC: $t_R = 1.32 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{63}H_{115}N_{19}O_{13}$ calc./obs. 1345.89/1345.89 [M]⁺.





GP1 (FLKAVKKLIPSLF-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (203.7 mg, 6.9%). Analytical RP-HPLC: $t_R = 1.56 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{76}H_{127}N_{17}O_{14}$ calc./obs. 1501.97/1501.97 [M]⁺.





GP2 (RWRWPILGRILR-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (227.9 mg, 18.3%). Analytical RP-HPLC: $t_R = 1.54 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{77}H_{125}N_{27}O_{12}$ calc./obs. 1620.00/1620.00 [M]⁺.





GP3 (FLHSIGKAIGRLLR-NH₂ was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (223.6 mg, 9.3%). Analytical RP-HPLC: $t_R = 1.51 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{73}H_{126}N_{24}O_{15}$ calc./obs. 1578.98/1578.99 [M]⁺.





GP4 (GIGAVLNVAKKLL-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (170.2 mg, 3.3%). Analytical RP-HPLC: $t_R = 1.70 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{60}H_{111}N_{17}O_{14}$ calc./obs. 1293.85/1293.85 [M]⁺.





GP5 (KVARFLKKFFR-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (220.8 mg, 10.7%). Analytical RP-HPLC: $t_R = 1.38 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{71}H_{115}N_{21}O_{11}$ calc./obs. 1437.91/1437.91 [M]⁺.





GP6 (LKKLWKRIIKVGR-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (253.3 mg, 20.3%). Analytical RP-HPLC: $t_R = 1.31 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{78}H_{141}N_{25}O_{13}$ calc./obs. 1636.11/1636.11 [M]⁺.





GP7 (ARKWRKFLKKI-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (236.2 mg, 16.9%). Analytical RP-HPLC: $t_R = 1.28 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{71}H_{121}N_{23}O_{11}$ calc./obs. 1471.96/1471.96 [M]⁺.





GP8 (GRIKRIRKIIHKY-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (269.6 mg, 20.6%). Analytical RP-HPLC: $t_R = 1.22 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{77}H_{138}N_{28}O_{14}$ calc./obs. 1679.09/1679.10 [M]⁺





GP9 (ARKKWRKRLKKLKI-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (311.1 mg, 15.7%). Analytical RP-HPLC: $t_R = 1.22 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{86}H_{159}N_{31}O_{14}$ calc./obs. 1850.27/1850.26 [M]⁺.





GP10 (AKKVVKKIYKRFQK-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (278.2 mg, 4.9%). Analytical RP-HPLC: $t_R = 1.16 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{84}H_{147}N_{25}O_{16}$ calc./obs. 1762.15/1762.15 [M]⁺.





GP11 (ARKFRRLVKKLR-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (246.3 mg, 4.5%). Analytical RP-HPLC: $t_R = 1.21 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{71}H_{132}N_{28}O_{12}$ calc./obs. 1569.06/1569.05 [M]⁺.





GP12 (LRKARRLVKKLA-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (233.9 mg, 10.9%). Analytical RP-HPLC: $t_R = 1.18 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{65}H_{127}N_{25}O_{12}$ calc./obs. 1450.01/1450.01 [M]⁺.





GP13 (KRLWKIRQRIAK-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (248.9 mg, 34.3%). Analytical RP-HPLC: $t_R = 1.18 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{73}H_{131}N_{27}O_{13}$ calc./obs. 1594.04/1594.04 [M]⁺.





GP14 (LNALKKVFQKIH-NH₂) was obtained from Rink Amide AM resin LL (400 mg, 0.26 mmol/g), the peptide was obtained as a white foamy solid after preparative RP-HPLC purification (208.8 mg, 7.8%). Analytical RP-HPLC: $t_R = 1.39 \text{ min}$ (100% A to 100% D in 3.5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{68}H_{116}N_{20}O_{14}$ calc./obs. 1436.9/1437.91 [M]⁺.





Supporting information Note 4: Topology of the DPC molecule

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; Charge from Chiu et al. ; Chiu, S. W.; Clark, M.; Balaji, V.; Subramaniam, S.; Scott, H. L.; Jakobsson, E. Incorporation of surface tension into molecular dynamics simulation of an interface: a fluid phase lipid bilayer membrane. Biophys. J. 1995, 69, 1230-1245. ; Atom types from GROMOS53A6 ; Oostenbrink, C.; Soares, T. A.; van der Vegt, N. F. A.; van Gunsteren, W. F. Validation of the 53A6 GROMOS force field. Eur. Biophys. J. 2005, 34, 273-284. [moleculetype] ; Name nrexcl DPC З [atoms] nr type resnr residu atom cgnr charge mass CH3 DPC C1 0.40 15.035 ; qtot: 0.25 1 1 1 0.40 15.035 ; qtot: 0.50 0.40 15.035 ; qtot: 0.75 2 CH3 1 DPC C2 2 3 CH3 1 DPC C3 3 -0.5 14.0067 ; qtot: 0.75 0.30 14.027 ; qtot: 1.0 DPC N4 4 NL 1 4 5 CH2 1 DPC C5 5 CH2 1 DPC C6 6 0.40 14.027 ; qtot: 1.0 6 -0.80 15.999; qtot: 0.64 1.7 30.973; qtot: 1.63 OA DPC 07 7 7 1 P8 DPC 8 8 Ρ 1 -0.8 15.999 ; qtot: 0.995 9 ОM 1 DPC 09 9 10 OM 1 DPC 010 10 -0.8 15.999 ; qtot: 0.36 -0.7 15.999 ; qtot: 0.0 11 OA 1 DPC 011 11 14.027 ; qtot: 0 14.027 ; qtot: 0 CH2 DPC 12 12 1 C12 0.0 13 CH2 1 DPC C13 13 0.0 14 CH2 1 DPC C14 14 0.0 14.027 ; qtot: 0 15 CH2 1 DPC C15 15 0.0 14.027 ; qtot: 0 CH2 1 DPC C16 16 14.027 ; qtot: 0 16 0.0 DPC ; qtot: 0 17 CH2 1 C17 17 0.0 14.027 ; qtot: 0 18 CH2 1 DPC C18 18 0.0 14.027 DPC C19 14.027 ; qtot: 0 19 CH2 1 19 0.0 CH2 DPC C20 20 ; qtot: 0 20 0.0 14.027 1 DPC C21 14.027 CH2 1 21 0.0 21 ; qtot: 0 22 CH2 1 DPC C22 22 0.0 14.027 ; qtot: 0 23 СНЗ 1 DPC C23 23 0.0 15.035 ; qtot: 0 [bonds] ai aj funct c0 c1 c2 c3 ; 2 gb_21 1 4 2 4 2 gb_21 3 4 2 gb 21 5 2 gb 21 4 gb_27 gb_18 5 6 2 6 7 2 7 8 2 gb 28 8 9 2 gb 24 8 10 2 gb 24 8 2 11 gb_28 11 12 2 gb 18 12 13 2 gb 27 13 14 2 gb_27 2 gb_27 14 15 gb_27 15 16 2 16 17 2 gb 27 17 2 qb 27 18 18 19 2 gb 27 2 19 20 gb_27 20 21 2 gb_27 21 22 2 gb 27 2 qb 27 22 23 [pairs] ai aj funct ; 1 6 1 2 6 1 3 6 1 4 7 1 5 8 1

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