

Supporting Information for: Accessing Lanthanide-based, In Situ Illuminated Optical Turn-On Probes by Modulation of the Antenna Triplet State Energy

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Table S1. Summary of relevant discrete lanthanide turn-on probes								
"Off/On"	Probe	Φ_{Ln}	Φ_{Ln} x-fold increase	Em. x-fold increase	[Probe]	Analyte/Concentration	Time-gated? (Y/N)	Ref
Off	EuBn	0.0012	--	--	--	--		
On	EuCou	0.0108	9	ND	--	--		
Off	TbBn	0.0041	--	--	--	--		
On	TbCou	0.0163	4	ND	50 μ M	Various enzymes/0.1-5 U	Y	[1]
Off	EuL1	0.07	--	--	--	--		
On	EuL1-F	ND	ND	9	20 μ M	F ⁻ /20-260 μ M	N	[2]
Off	Tb-DO3A	ND	--	--	--	--		
On	Tb-DO3A-trimesate	ND	ND	11	50.9 μ M	HO [*] /0.37 fM	Y	[3]
Off	Eu-Lys-HOPO	ND	--	--	--	--		
On	Eu-Lys-HOPO + CN	ND	ND	9	16 μ M	CN ⁻ /1.6 mM	Y	[4]
Off	EuL	0.002	--	--	--	--		
On	F-(Eu)L2	0.047	24	22	49 μ M	F ⁻ /24 nM	N	[5]
Off	Eu-Lys-HOPO	ND	--	--	--	--		
Off	Eu-Ser-HOPO	ND	--	--	--	--		
Off	Eu-Gly-HOPO	ND	--	--	--	--		
On	Eu-Lys-HOPO + HPO ₄ ²⁻	ND	ND	5	--	--		
On	Eu-Ser-HOPO + HPO ₄ ²⁻	ND	ND	36	--	--		
On	Eu-Gly-HOPO + HPO ₄ ²⁻	ND	ND	20	7.6 μ M	HPO ₄ ²⁻ /76 μ M	Y	[6]
Off	Mito-NSTTA-Eu	0.036	--	--	--	--		
Off	Mito-NSTTA-Tb	0.0043	--	--	--	--		
On	Mito-HTTA-Eu	0.035	1	--	--	--		
On	Mito-HTTA-Tb	0.147	34	ND	3.0 μ M	glutathione / Cys // 5 uM	Y	[7]
Off	ANMTTA-Eu	0.0023	--	--	--	--		
Off	ANMTTA-Tb	0.0007	--	--	--	--		
On	HTTA-Eu	0.163	71	100-600	--	--		
On	HTTA-Tb	0.115	164	100-600	5 μ M	HOCl / 70 μ M	Y	[8]
Off	[Tb-1*Zn ₂](NO ₃) ₄	0.01	--	--	--	--		
On	[Tb-1*Zn ₂](NO ₃) ₄ +GMP	0.48	48	87	20 μ M	GMP / 20 μ M	Y	[9]
Off	Eu.2	0.0045	--	--	--	--		
On	Eu.1	0.0056	1.24	20	10 μ M	HS ⁻ / 250 μ M	Y	[10]
Off	ATTA-Eu	0.0058	--	--	--	--		
On	EP-ATTA-Eu	0.10	17.2	ND	100 nM	¹ O ₂ / 2.8 nM	Y	[11]

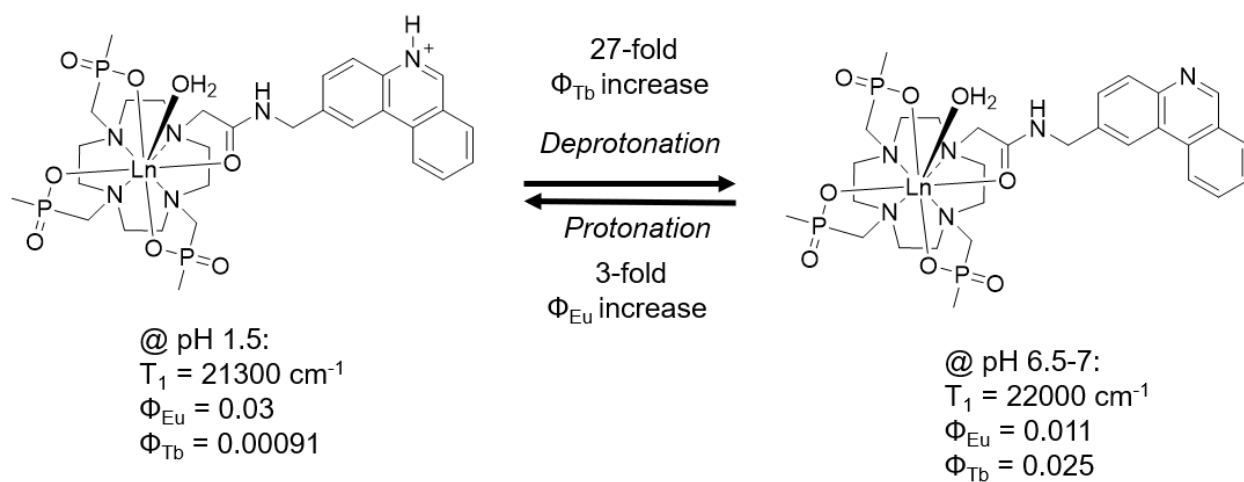


Figure S1. Structures and reactivity of pH-sensors developed by Parker and coworkers.¹²

3. Synthesis protocols, characterization, and HPLC traces

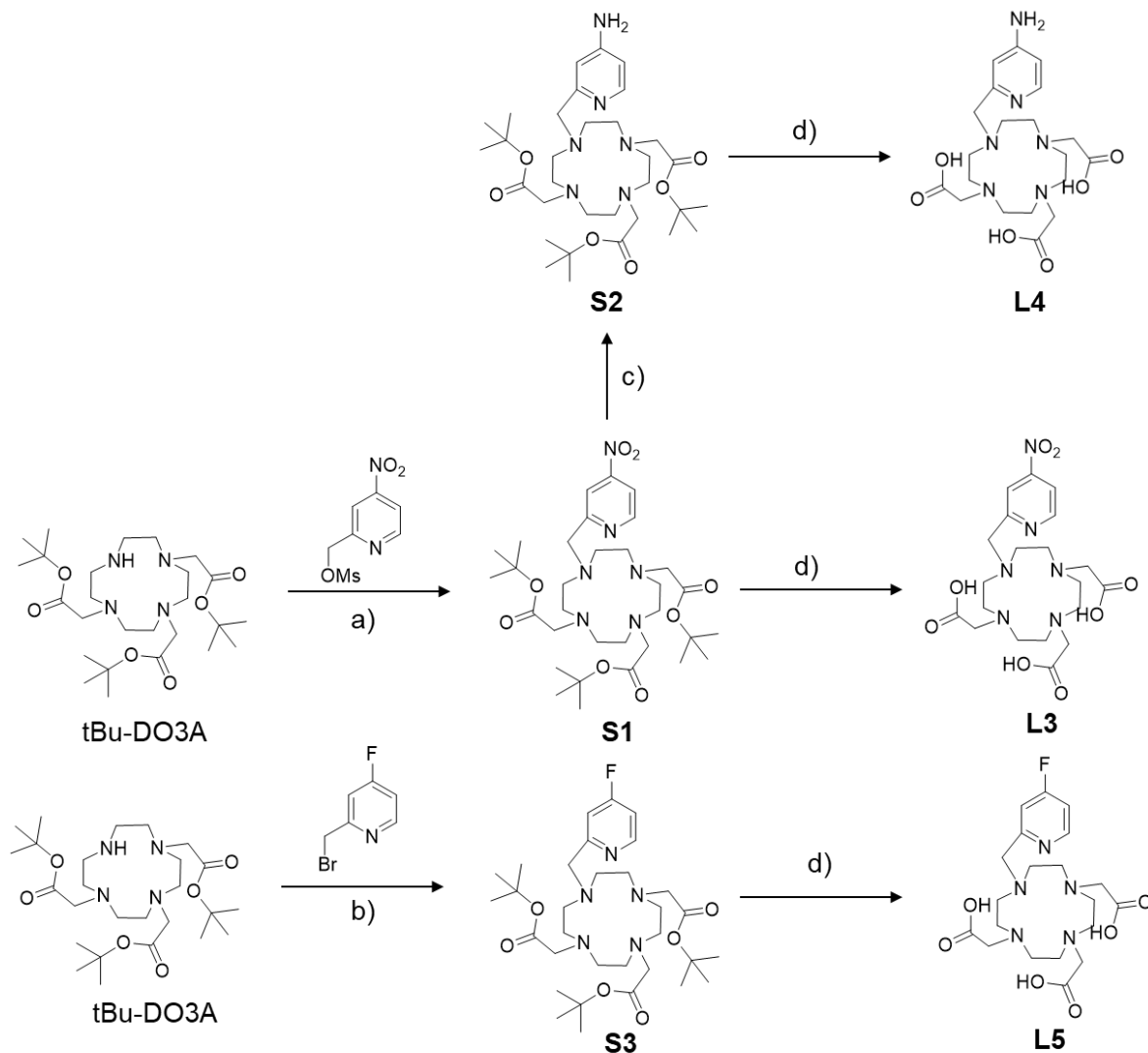


Figure S2. Synthetic scheme of ligands. a) dry MeCN, 8 eq. K₂CO₃, 90 °C, 12 hr. b) dry MeCN, 8 eq. K₂CO₃, 90 °C, 4 hr. c) 40% Pd/C, H₂, dry MeOH, 2 hr. d) 2:1 TFA:DCM, RT, 12 hr.

General protocols and methods featured in main manuscript.

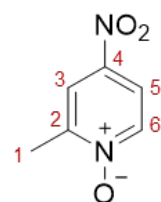
Cryogenic phosphorescence measurements were recorded on an Cary Eclipse fluorescence spectrometer from Agilent Technologies. The spectrometer was equipped with a photomultiplier tube and a quartz cold finger dewar fitted for measurements on NMR tubes in liquid nitrogen temperatures. All complexes were dissolved in DPBS and flash frozen in an NMR tube. Phosphorescence spectra were recorded with a 2 nm data intervals and time-gated in the range 0.5 – 60 ms unless otherwise indicated. Due to large differences in quantum yields between samples

both slit sizes and detector voltage varied with the sample. The T_1 energy levels were determined from the gadolinium samples at 3 % of the maximum peak intensity on the high energy edge of the transition band. The Gd(**L3**) exhibited the presence of several emitting species and therefore a convolution on the high energy side of the band. Due to the greater error the T_1 energy level was determined at the 10 % intensity of the maxima.

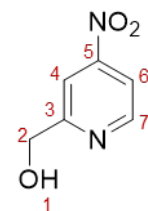
Phosphorescence decays were collected in similar fashion as the phosphorescence spectra in the 0.5 – 80 ms range and fitted to single, double or tripple component exponential decay funtions using the Origin software.

3. Synthesis and characterization

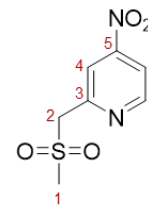
2-methyl-4-nitropyridine 1-oxide, 1. Concentrated H_2SO_4 (9.19 g) was massed and put into a 100 ml RBF on a salt-ice bath, to which 2-methylpyridine N-oxide (1.45 g, 1.31 mL) was added dropwise with stirring. Concentrated HNO_3 (9.02 g) was massed and added slowly over 10 minutes. After an additional 10 minutes at $0^\circ C$, the flask was moved to a oil bath preheated to $45^\circ C$. The temperature was slowly increased to $110^\circ C$ over 1 hour, and then refluxed for 2 hours. After cooling to room temperature and allowing excess gases to dissipate, the mixture was poured over ice water (100 g). The aqueous phase was extracted with DCM (3x 50 mL), washed with saturated K_2CO_3 (120 mL) and extracted again with DCM (50 mL). After drying over Na_2SO_4 , the final solution was concentrated in vacuo to yield a yellow crystalline solid (0.944g, 47%). 1H NMR ($CDCl_3$, 500 MHz): δ 8.31 (d, 1H, H^5), 8.14 (d, 1H, H^3), 7.99 (d, 1H, H^6), 2.57 (s, 3H, H^1). ^{13}C NMR ($CDCl_3$, 125 MHz): δ 150.61 (C^2), 141.64 (C^4), 140.02(C^6), 120.64 (C^5), 118.07 (C^3), 18.06 (C^1). ESI-MS: calcd. for $C_6H_6N_2O_3$: 154.04.



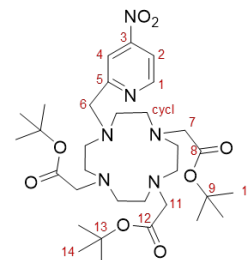
2-(Hydroxymethyl)-4-nitropyridine, 2. Trifluoroacetic acid anhydride (3 mL, 21.2 mmol, 3.5 eq) in 2 mL DCM was added dropwise, to a solution of **1** (0.944g, 6.10 mmol, 1 eq) in 10 mL dry DCM and refluxed overnight. The mixture was concentrated and then redissolved in 40 mL H_2O . The pH was brought up to 8 with K_2CO_3 and stirred at RT for 5 hours, after which KOH was added to increase the pH to 11. The aqueous phase was extracted with DCM (3x 50 mL), dried over Na_2SO_4 , and concentrated to dryness in vacuo. The resutling orange crystalline solid was used without further purification (0.555 g, 59%). 1H NMR ($CDCl_3$, 500 MHz): δ 8.86 (d, 1H, H^7), 8.08 (s, 1H, H^4), 7.95 (d, 1H, H^6), 4.95 (s, 2H, H^2), 3.20 (br s, 1H, H^1). ^{13}C NMR ($CDCl_3$, 125 MHz): δ 163.15 (C^3), 154.41 (C^7), 150.99 (C^5), 114.95 (C^4), 113.13 (C^6), 64.39 (C^2). ESI-MS: calcd. for $C_6H_6N_2O_3$: 154.02.



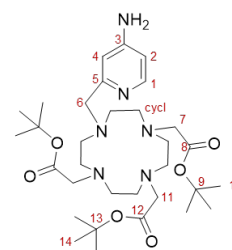
2-[(Methylsulfonyloxy)methyl]-4-nitropyridine, 3. Triethylamine (390 μ L, 2.8 mmol, 3 eq) was added to a solution of **2** (0.143 g, 0.93 mmol, 1 eq) in 10 mL dry THF. The solution was cooled to 0°C under an atmosphere of N₂. Methanesulfonyl chloride (110 μ L, 1.39 mmol, 1.5 eq) was added dropwise and the solution stirred at 0°C for 20 minutes. The residue was concentrated in vacuo, redissolved in DCM and washed with brine. The aqueous layer was further extracted with DCM, and the organic layers combined, dried over Na₂SO₄ and concentrated. The crude material was subject to silica column chromatography (10% MeOH in DCM, 8 cm x 3 cm silica plug) to afford a yellow oil (51%, 73.0 mg, 0.314 mmol, R_f [2% MeOH in DCM]: 0.76). ¹H NMR (CDCl₃, 400 MHz): δ 8.90 (d, 1H, H⁷), 8.17 (s, 1H, H⁴), 8.00 (d, 1H, H⁶), 5.43 (s, 2H, H²), 3.16 (s, 3H, H¹). ¹³C NMR (CDCl₃, 100 MHz): δ 157.33 (C³), 154.51 (C⁷), 151.82 (C⁵), 116.06 (C⁴), 114.49 (C⁶), 69.66 (C¹), 37.98 (C²).



***tert*-Butyl {4,10-bis(*tert*-butoxycarbonylmethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetate, S1.** **3** (74.5 mg, 0.32 mmol) was dissolved in anhydrous acetonitrile and added dropwise to a solution of tBu-DO3A (156 mg, 0.30 mmol) and K₂CO₃ (491 mg, 2.1 mmol) in 4 mL anhydrous acetonitrile. The mixture refluxed for 2 h and was then filtered and concentrated. The filtrate was purified using reverse-phase chromatography (method D), with the product eluting from 13-14 column volumes (CV). The fractions containing product were pooled and the solvent was removed in vacuo to afford **S1** as a yellow oil (23%, 47.8 mg, 0.074 mmol, R_{CV} (method D) = 13-15 CV). ¹H NMR (CD₃OD, 500 MHz): δ 8.86 (s, 1H, H¹), 8.23 (m, 2H, H^{2,4}), 4.21 (br s, 2H, H⁶), 3.65-3.51 (m, 14H, H^{yc}), 3.25-3.02 (m, 8H, H^{7,11,cycl}), 1.55 (s, 9H, H¹⁴), 1.33 (s, 18H, H¹⁰). ¹³C NMR (CD₃OD, 125 MHz): δ 172.51 (C⁸), 166.95 (C¹²), 156.15 (C⁵), 152.16 (C¹), 117.11 (C³), 116.06 (C⁴), 114.20 (C²), 86.08 (C⁹), 83.40 (C¹³), 58.17 (C⁷), 56.14 (C^{cycl}), 55.18 (C⁶), 53.22 (C^{cycl}), 50.05 (C^{cycl}), 28.51 (C¹⁰), 28.39 (C¹⁴). ESI-MS: calcd. for C₃₂H₅₄N₆O₈: 650.40. Found: 651.2 [M+H]⁺.

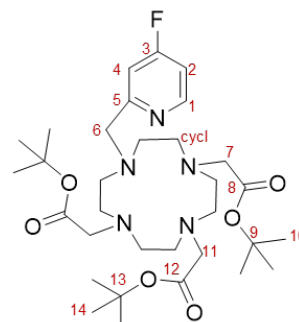


***tert*-Butyl {10-[(4-amino-2-pyridyl)methyl]-4,7-bis(*tert*-butoxycarbonylmethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetate, S2.** 40 mol % Pd/C (10.8 mg) was added to **S1** (27.4mg, 0.042 mmol) in 2.5 mL dry methanol and placed under H₂. Upon completion of the reaction after 2 hours as confirmed by LRMS, the mixture was filtered, and the filtrate was purified using semi-preparative HPLC (method A), with the product eluting at 10.1 minutes. The fractions containing product were pooled and the solvent was removed in vacuo to afford **S2** as a white solid (45%, 11.6 mg, 0.019 mmol, R_t (method A) = 10.1 minutes). ¹H NMR (CD₃OD, 500 MHz): 7.90 (d, 1H, H¹), 6.84 (m, 1H, H⁴), 6.74 (s, 1H, H²), 4.05 (s, 2H, H⁶), 3.64 (m, 14H, H^{yc}), 3.11-3.01 (m, 8H, H^{7,11,cycl}), 1.55 (s, 9H, H¹⁴), 1.49 (s, 18H, H¹⁰). ¹³C NMR (CD₃OD, 125 MHz): δ 170.38 (C^{8,12}), 161.80 (C⁵), 141.19 (C¹), 140.93 (C³), 110.05 (C⁴), 109.34 (C²), 84.48 (C⁹), 84.02 (C¹³), 56.39 (C⁷), 55.54

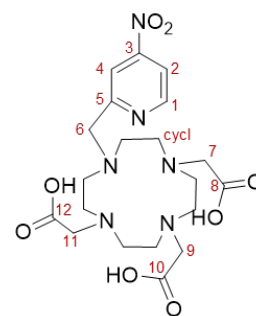


(C^{cycl}), 55.35 (C^{cycl}), 54.72 (C¹¹), 52.22 (C^{cycl}), 51.40 (C^{cycl}), 49.62 (C⁶), 28.51 (C¹⁰), 28.39 (C¹⁴). ESI-MS: calcd. for C₃₂H₅₆N₆O₆: 620.43. Found: 621.2 [M+H]⁺.

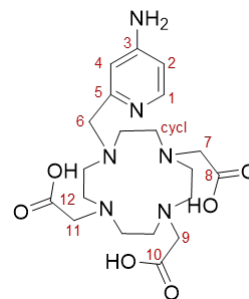
tert-Butyl {4,10-bis(tert-butoxycarbonylmethyl)-7-[(4-fluoro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetate, S3. 2-methyl-4-fluoropyridine (19.2 mg, 0.102 mmol) and K₂CO₃ (67.3 mg, 0.510 mmol) was added dropwise to tBu-DO3A (52.5 mg, 0.102 mmol) in anhydrous acetonitrile and heat at reflux for 4 hours. Upon completion of the reaction as confirmed by LRMS, the mixture was filtered, and the filtrate was purified using semi-preparative HPLC (method A), with the product eluting at 14.1 minutes. The fractions containing product were pooled and the solvent was removed in vacuo to afford **6** as a yellow oil (24%, 15.2 mg, 0.023 mmol, R_t (method A) = 14.1 minutes). ¹H NMR (CD₃OD, 700 MHz): 8.56 (m, 1H, H¹), 7.29 (d, 2H, H^{2,4}), 4.76 (s, 2H, H⁶), 4.22 (s, 2H, H¹¹), 3.75-3.49 (m, 12H, H^{cycl}), 3.23-3.03 (m, 8H, H^{7,cycl}), 1.57 (s, 9H, H¹⁴), 1.35 (s, 18H, H¹⁰). ¹³C NMR (CD₃OD, 175 MHz): δ 172.25 (C⁸), 171.49 (C³), 170.00 (C⁵), 166.96 (C¹²), 52.47 (C¹), 112.63 (C⁴), 111.58 (C²), 86.07 (C⁹), 83.41 (C¹³), 58.12 (C⁷), 56.23 (C^{cycl}), 55.10 (C¹¹), 53.05 (C^{cycl}), 50.09 (C^{cycl}), 49.52 (C⁶), 28.52 (C¹⁰), 28.42 (C¹⁴). ¹⁹F NMR: (CD₃OD, 376 MHz): -101.24. ESI-MS: calcd. for C₃₂H₅₄FN₅O₆: 623.41. Found: 624.1 [M+H]⁺.



{4,10-Bis(carboxymethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L3. **S1** (34.1 mg, 0.023 mmol) was added to a 2:1 solution of TFA and DCM and stirred overnight at room temperature. The solvent was removed in vacuo, and the product was dissolved in H₂O. The solution was then lyophilized to yield **L3** as an off-white solid (11.1 mg, 0.022 mmol, R_t (method C) = 3.0 minutes). ¹H NMR (CD₃OD, 500 MHz): δ 8.88 (d, 1H, H¹), 8.23 (s, 1H, H⁴), 8.10 (s, 1H, H²), 4.13 (br s, 2H, H⁶), 3.76-3.57 (m, 16H, H^{9,cycl}), 3.23-3.21 (m, 6H, H^{7,11,cycl}). ¹³C NMR (CD₃OD, 125 MHz): δ 173.12 (C^{8,11}), 169.36 (C¹⁰), 159.54 (C⁵), 156.12 (C¹), 152.71 (C³), 117.43 (C⁴), 116.77 (C²), 58.33 (C⁶), 55.34 (C^{cycl}), 54.12 (C^{7,11}), 52.25 (C^{cycl}), 50.48 (C^{cycl}), 49.62 (C⁹). ESI-MS calcd. for C₂₀H₃₀N₆O₈: 482.2125. Found: 483.2198 [M+H]⁺.



{10-[(4-Amino-2-pyridyl)methyl]-4,7-bis(carboxymethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L4. **S2** (7.1 mg, 0.011 mmol) was added to a 2:1 solution of TFA and DCM and stirred overnight at room temperature. The solvent was removed in vacuo, and the product was dissolved in H₂O. The solution was then lyophilized to yield **L4** as an off-white solid (4.7 mg, 0.010 mmol, (method C) = 1.89 minutes). ¹H NMR (CD₃OD, 400 MHz): 7.90 (d, 1H, H¹), 6.86 (s, 1H, H⁴), 6.72 (d, 2H, H²), 4.07 (br s, 2H, H⁶), 3.96-3.66 (m, 4H, H^{7,11}), 3.26-2.93 (m, 18H, H^{cycl,9}). ¹³C NMR (CD₃OD, 100 MHz): δ 177.27 (C⁸), 176.46 (C¹²), 175.85 (C¹⁰), 161.89 (C⁵), 149.85 (C¹), 142.07 (C³), 111.23 (C⁴), 109.06 (C²), 57.47 (C⁹), 56.14 (C^{cycl}), 56.13 (C⁷), 55.44

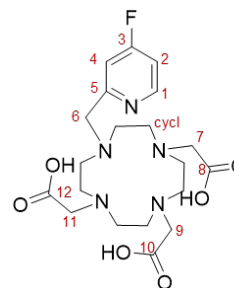


(C^{cycl}), 54.34 (C^{cycl}), 51.785 (C⁶). ESI-MS calcd. for C₂₀H₃₂N₆O₆: 452.2383. Found: 453.2457 [M+H]⁺.

{4,10-Bis(carboxymethyl)-7-[(4-fluoro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L5. S3 (15.2 mg, 0.023 mmol) was added to a

2:1 solution of TFA and DCM and stirred overnight at room temperature. The solvent was removed in vacuo, and the product was dissolved in H₂O.

The solution was then lyophilized to yield **L5** as an off-white solid (97%, 10.3 mg, 0.023 mmol, (method C) = 2.01 minutes). ¹H NMR (CD₃OD, 400 MHz): δ 8.58 (m, 1H, H¹), 7.35 (d, 1H, H⁴), 7.22 (m, 1H, H²), 4.62 (br s, 2H, H⁶), 4.11 (br s, 2H, H⁹), 3.69-3.49 (m, 14H, H^{cycl}), 3.22 (m, 6H, H^{cycl,7}). ¹³C NMR (CD₃OD, 125 MHz): δ 173.05 (C⁸), 172.01 (C¹²), 169.91 (C¹⁰), 158.79 (C³), 153.12 (C⁵), 120.96 (C¹), 118.64 (C⁴), 116.34 (C²), 58.17 (C⁷), 55.77 (C^{cycl}), 53.82 (C⁹), 52.61 (C^{cycl}), 51.37 (C^{cycl}), 50.75 (C⁶), 50.17 (C^{cycl}). ¹⁹F NMR: (CD₃OD, 376 MHz): -100.16. (CD₃OD, 376 MHz): ESI-MS calcd. for C₂₀H₃₀FN₅O₆: 455.2180. Found: 456.2256 [M+H]⁺.



HPLC traces

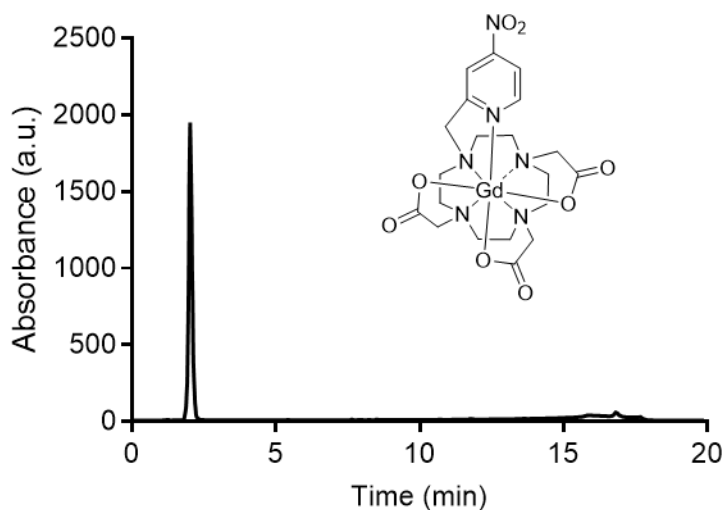


Figure S3. HPLC trace of Gd(**L3**). τ_r = 2.13 min.

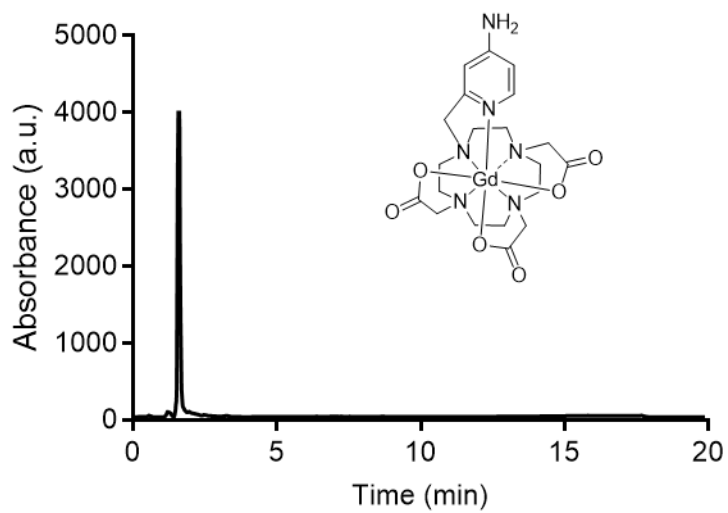


Figure S4. HPLC trace of Gd(L4). $\tau_r = 1.58$ min.

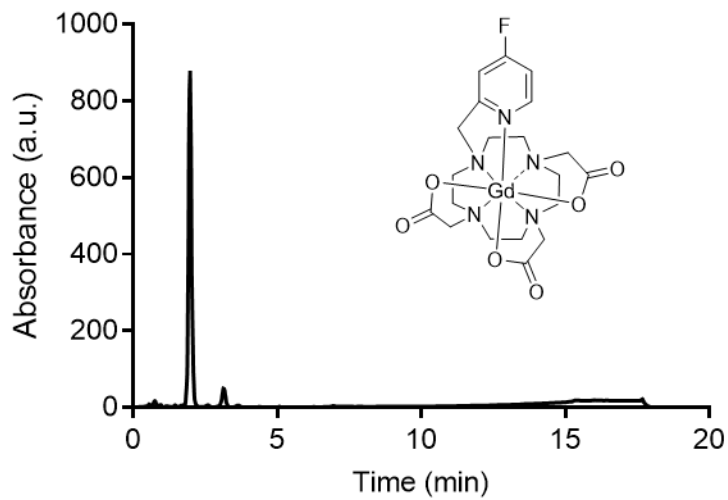


Figure S5. HPLC trace of Gd(L5). $\tau_r = 1.97$ min.

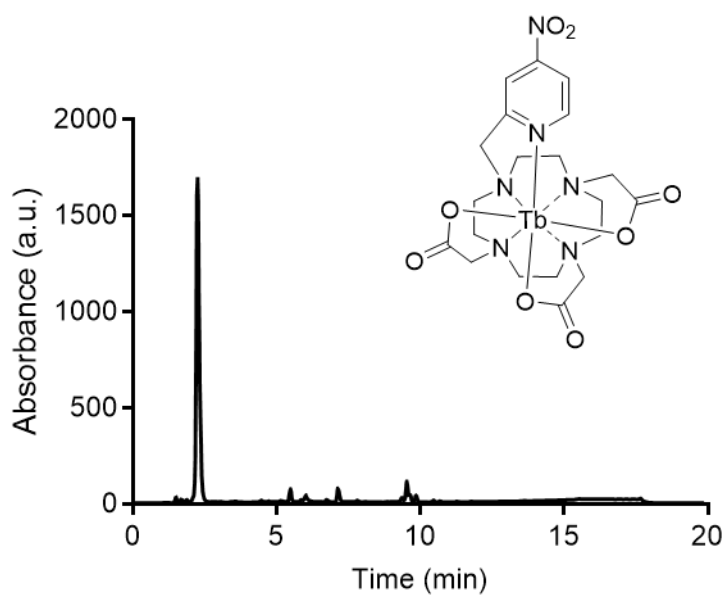


Figure S6. HPLC trace of Tb(L3). $\tau_r = 2.13$ min.

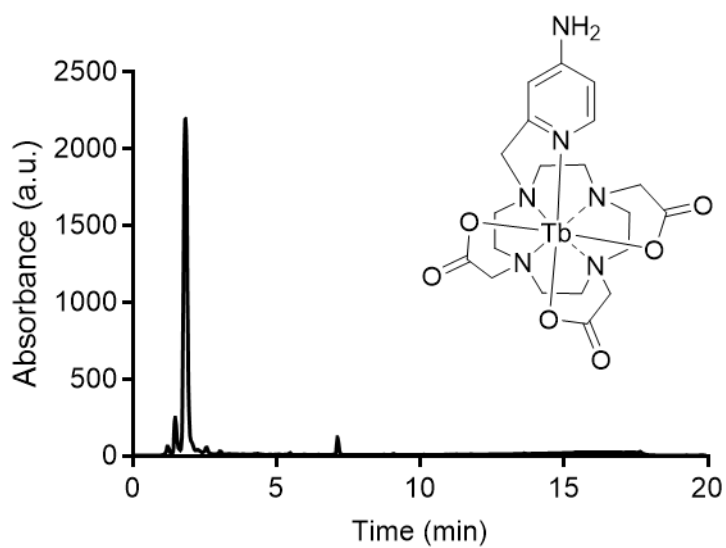


Figure S7. HPLC trace of Tb(L4). $\tau_r = 1.82$ min.

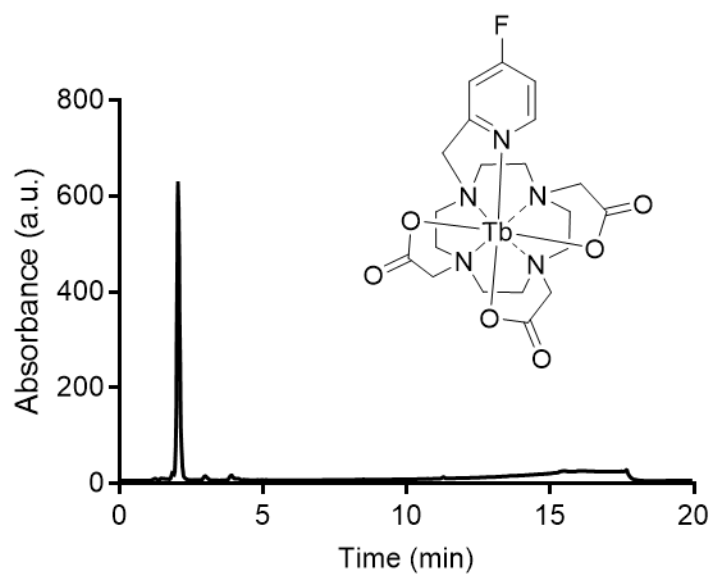


Figure S8. HPLC trace of Tb(L5). $\tau_r = 2.03$ min.

4. Phosphorescence spectra and data analysis

Phosphorescence data analysis

The T_1 energy levels of the pyridyl-functionalized groups were determined from the Gd(III) complexes to ensure that the observed transitions are ligand-based and unaffected by lanthanide(III) energy levels. The emission and excitation spectra of Gd(L4) and Gd(L5) complexes are unaffected by the selected excitation and emission wavelengths. The T_1 energy level can be directly determined at the 3 % intensity maxima at the onset of the high-energy side of the transition band.

Several features can be seen from the Gd(L3) emission spectra depending on the excitation wavelength and the selected time-gating of the measurement, indicating the presence of additional emitting species. However, the excitation spectra of the L3 complexes seen in Figure S8 also exhibit the low intensity peak at ~330 nm indicating a charge transfer band that is also observed in the absorption spectrum. In order to correctly assign the L3 T_1 state additional insight into the possible emitting species must be obtained. For this purpose, cryogenic spectra were recorded for Y-DOTA and the Tb(III) complexes along with phosphorescence decay measurements of all complexes.

From the phosphorescence measurements of Y-DOTA the emission of the carboxylate arms of the DOTA molecule is determined with a transition maxima at 430 nm, (Figure S11). The Tb(III)-based emission is characterized by high intensity and shorter lifetimes compared to their organic counterparts. A 15 ms time-gate can exclude the majority of Tb(III)-based emission and reveal the presence of underlying DOTA carboxylate emission.

The Tb(III) complexes reveal that the excitation spectra of the Gd(III) complexes are made of two distinct excitation spectra. Excitation spectra recorded by monitoring the emission of Tb(III) is blue-shifted compared with the pyridyl-based emission. These excitation profiles are consistent with those of pyridine: the blue-shifted excitation spectrum is consistent with the neutral form of pyridine and the red-shifted excitation spectrum consistent with the protonated version of pyridine, forming the pyridinium moiety and breaking the conjugated system. At the PBS moderated pH the neutral and protonated forms of the pyridyl exist in equilibrium.

By analyzing these spectra, the four key local excited states of Tb(III), DOTA, pyridine and pyridinium is determined. **DOTA-based emission is present in all complexes and should not be mistaken as the pyridyl T_1 state.** By exciting the neutral pyridyl group energy transfer to Tb(III) is favored and excitation of the pyridinium will favor emission via the DOTA carboxylic arms for Tb(L3) and Tb(L4). Tb(L5) exhibit both DOTA and pyridyl-based emission.

The excited state lifetimes of all complexes derived from the phosphorescent decays are given in Tables S3 to S6. From the Tb(L3) complex the lifetimes derived by exciting the pyridyl group at 275 nm (1.8 ms) is similar to the lifetime derived from exciting the charge transfer band (1.75 ms), when emission is observed through Tb(III). This indicates that the emission observed via the charge-transfer band (S_1) is representative of the L3 triplet state. The T_1 energy level can therefore be obtained from Gd(L3) with 330 nm excitation. Even through with excitation through the charge transfer band contributions to the emission of other phosphorescent species could not be avoided.

This produce a partial convolution and an increased error on the energy level determination. The T_1 state is therefore determined at the 10 % onset instead.

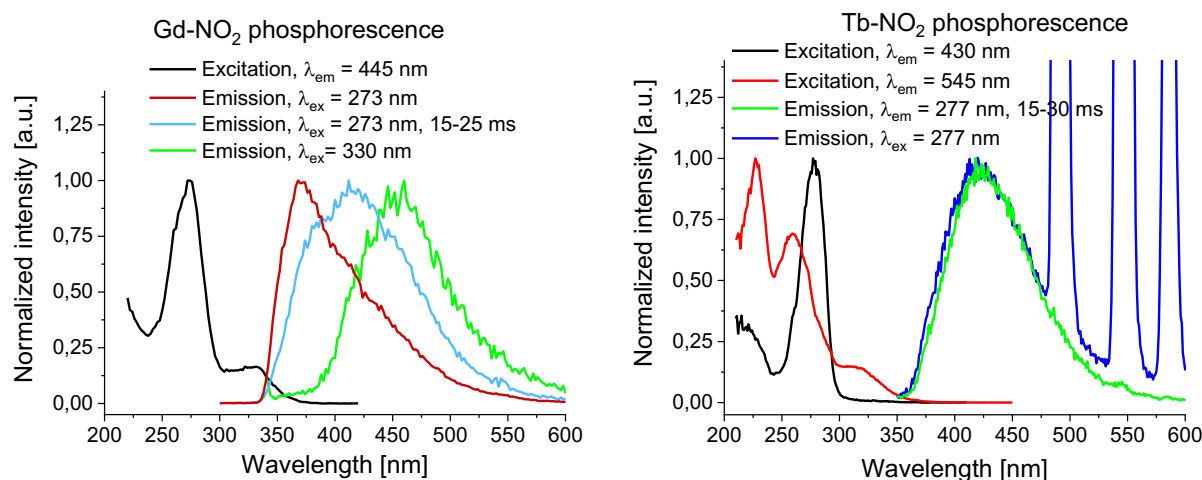


Figure S9. Phosphorescence spectra of Gd(L3) (left) and Tb(L3) (right) in PBS buffer at 77 K.

Table S2. Ln(L3), Ln = Gd(III), Tb(III) fitted lifetimes and amplitudes

NO ₂	λ_{ex} (nm)	λ_{em} (nm)	A ₁	τ_1	A ₂	τ_2	A ₃	τ_3
Gd(III)	273	370	9	1.3 ms	2	8.3 ms	-	-
	273	420	4	2.5 ms	2	21.6 ms	-	-
	273	455	8	2.7 ms	5	22.0 ms	-	-
	330	370	6	0.1 ms	0.01	5.9 ms	-	-
	330	420	8	0.2 ms	4	2.7 ms	1	14.4 ms
	330	455	3	2.2 ms	1	13.8 ms	-	-
Tb(III)	277	420	5	4.4 ms	11	51.0 ms	-	-
	317	420	3	0.2 ms	0.3	22.4 ms	-	-
	277	545	118	1.8 ms	5.7	0.2 ms	0.5	14.6 ms
	317	545	-	1.75 ms	-	-	-	-

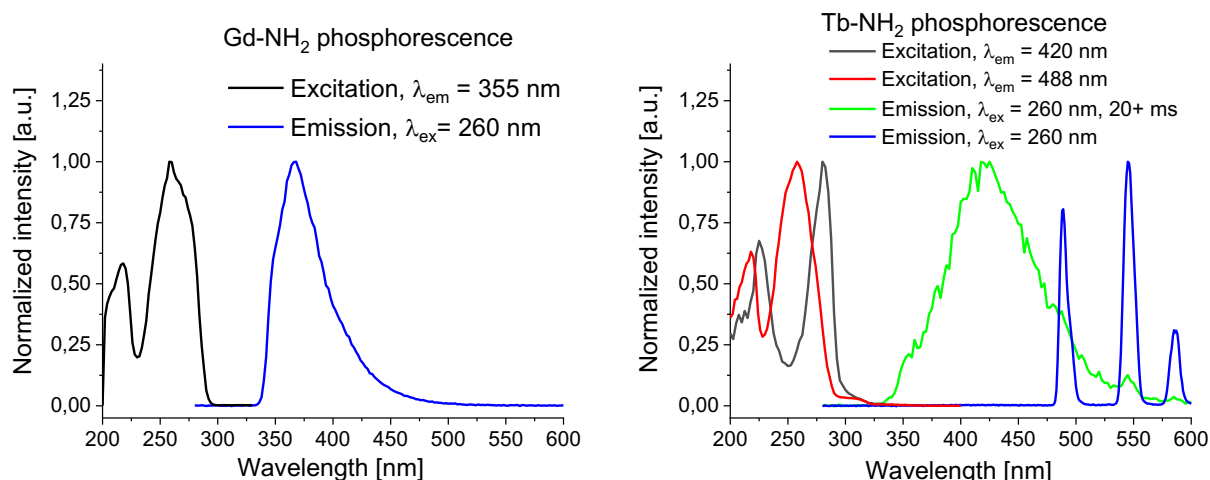


Figure S10. Phosphorescence spectra of Gd(L4) (left) and Tb(L4) (right) in PBS buffer at 77 K.

Table S3. Ln(L4), Ln = Gd(III), Tb(III) fitted lifetimes and amplitudes

NH ₂	λ _{ex} (nm)	λ _{em} (nm)	A ₁	τ ₁	A ₂	τ ₂	A ₃	τ ₃
Gd(III)	260	330	5	0.9 ms	3	3.5 ms	-	-
Tb(III)	250	420	19	0.1 ms	2	2.3 ms	0.5	36.7 ms
	285	420	6	3.1 ms	13	55.5 ms	-	-
	285	545	-	1.74 ms	-	-	-	-

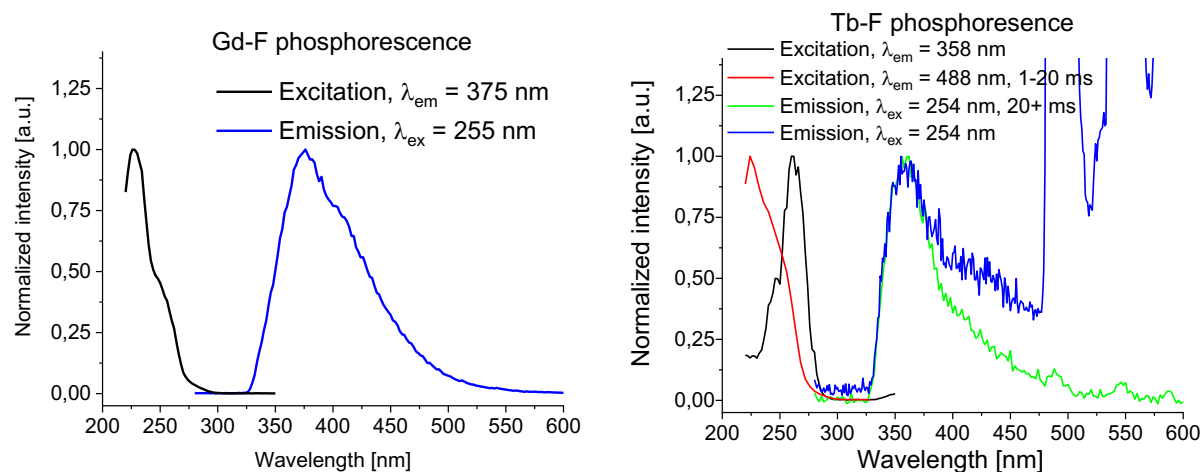
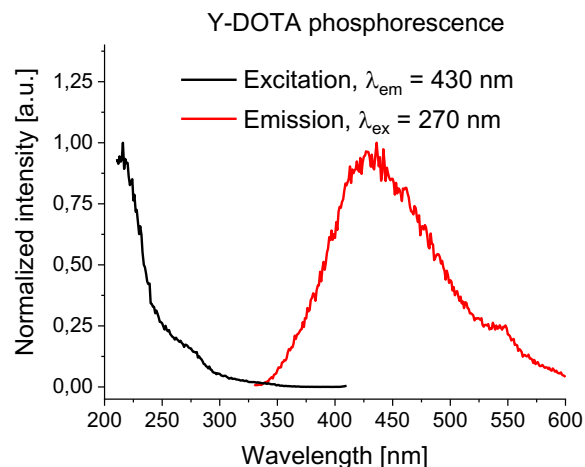


Figure S11. Phosphorescence spectra of Gd(L5) (left) and Tb(L5) (right) in PBS buffer at 77 K.

Table S4. Ln(L5), Ln = Gd(III), Tb(III) fitted lifetimes and amplitudes

F	λ_{ex} (nm)	λ_{em} (nm)	A ₁	τ_1	A ₂	τ_2	A ₃	τ_3
Gd(III)	255	340	25	1.8 ms	25	24.1 ms	-	-
	255	435	21	1.2 ms	19	17.3 ms	-	-
Tb(III)	255	435	118	0.2 ms	19	2.4 ms	5	24 ms
	275	358	5	1.9 ms	590	0.1 ms	6	62 ms

**Figure S12.** Phosphorescence spectra of Y-DOTA in DPBS buffer at 77 K. The small shoulder at 550 nm is likely a Tb(III) contamination.**Table S5.** Y-DOTA fitted lifetimes and amplitudes

Y-DOTA	λ_{ex} (nm)	λ_{em} (nm)	A ₁	τ_1	A ₂	τ_2	A ₃	τ_3
Y(III)	270	430	1.3	0.5 ms	1.6	5.4 ms	0.6	30.9 ms

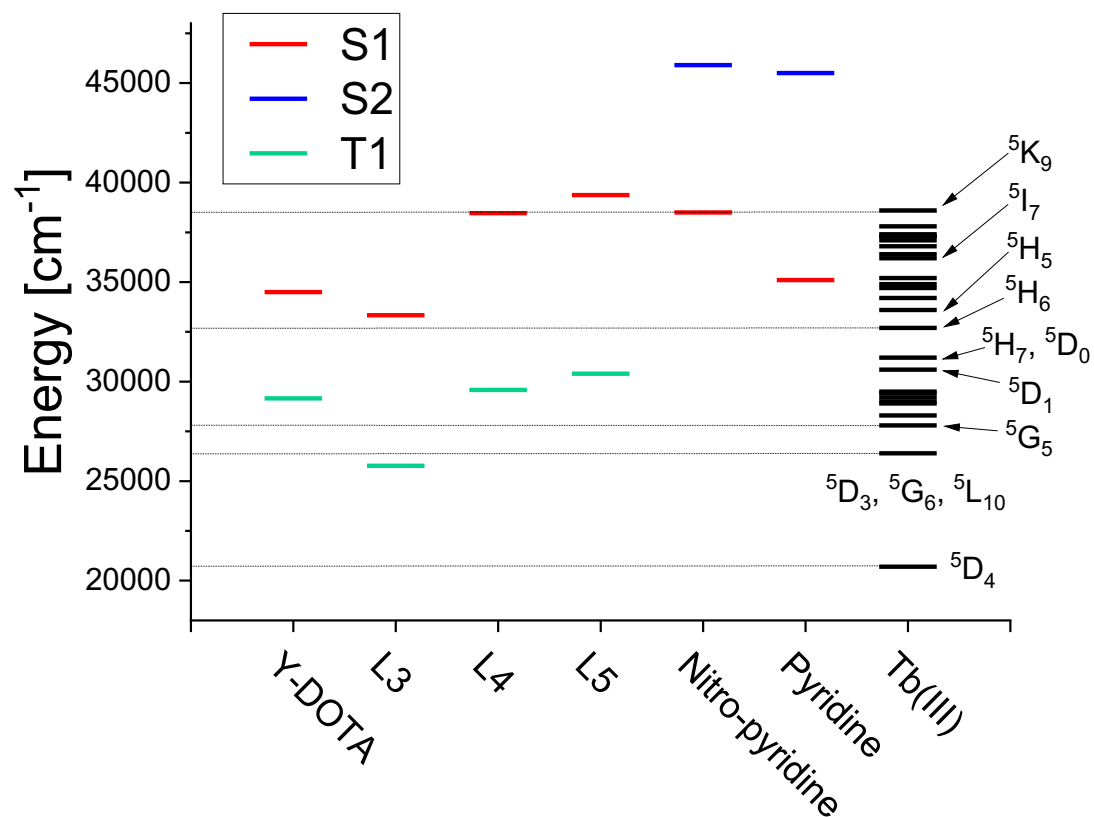


Figure S13. Energy level diagram of the obtained S₁ and T₁ energy levels of **L3**, **L4** and **L5** compared with the carboxylic DOTA energy levels, the pyridine and nitro-pyridine S₁ and S₂ levels and many of the Tb(III) excited energy levels.

5. Quantum yield, lifetime, and q measurements

Quantum yield for each complex was determined using the following equation:

$$QY_X = QY_S * \frac{\text{Gradient}_X}{\text{Gradient}_S} \quad (\text{S1})$$

where “S” refers to the $[\text{Tb}(\text{L1})]^-$ ($\Phi = 0.47$) and “X” is the unknown. The gradient is the slope of the graph of integrated emission intensity versus the peak absorption value for a range of concentrations with $\text{abs} < 0.1$.

Quantum yield gradients

Gradients for quantum yield determination were measured using the following procedure: Complex was diluted in 1X DPBS and measuring absorbances ranging 0.01-0.10, followed by measurement of fluorescence emission. Total emission integrals were taken between 450-650 nm (only Tb-based emission), assuming a Gaussian distribution, and the integral of the second-order scattering peak was subtracted.

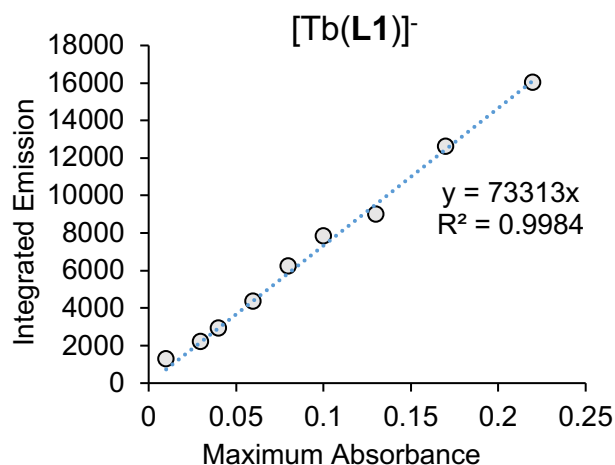


Figure S14. Quantum yield gradient of reference compound $[\text{Tb}(\text{L1})]^-$.

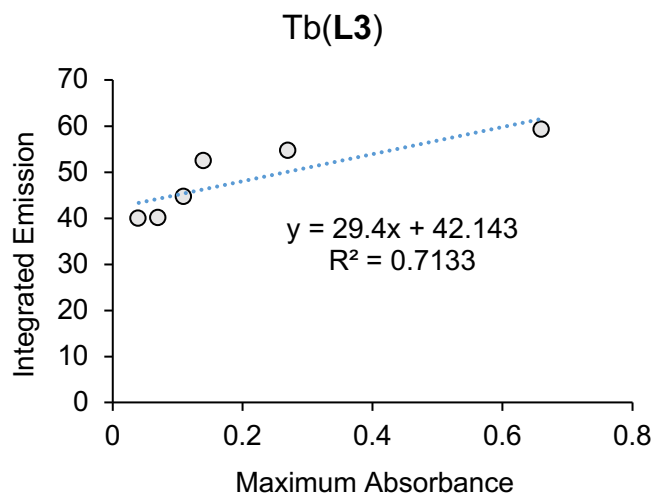


Figure S15. Quantum yield gradient of Tb(L3).

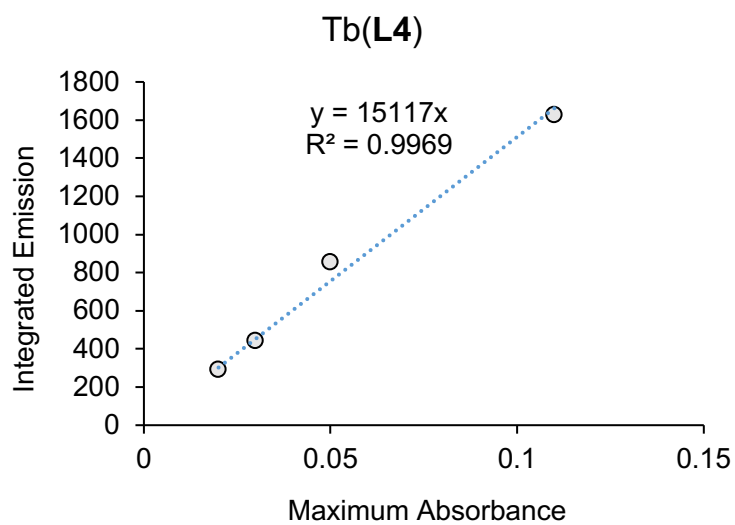


Figure S16. Quantum yield gradient of Tb(L4).

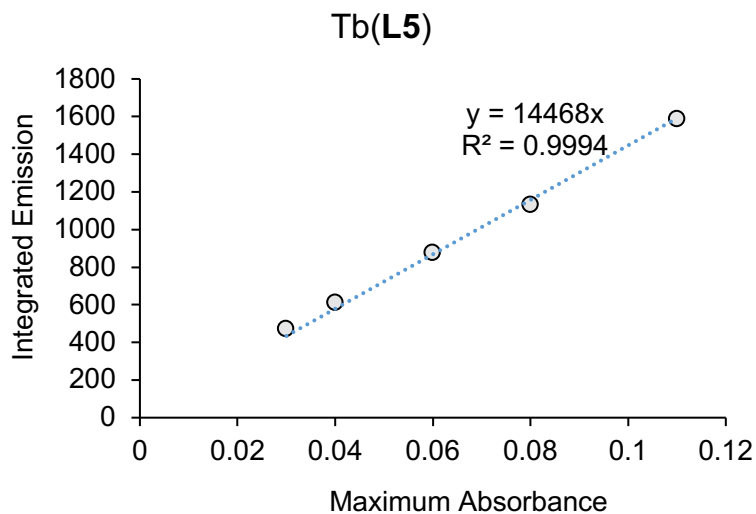


Figure S17. Quantum yield gradient of Tb(L5).

Table S6. Gradient QY summary			
	λ_{ex}	Gradient	Φ_{Tb} (%)
Tb(L3)	300	29.4	0.02
Tb(L4)	260	15117	10
Tb(L5)	254	14468	9.3
[Tb(L1)] ⁻	282	73315	47

Lifetime and q measurements

Lifetime values were extracted by fitting the luminescent decay curves with the equation:

$$I_t = I_0 * \exp\left(-\frac{x}{\tau}\right) \quad (\text{S2})$$

where I_t is the initial luminescent emission intensity, I_0 is the intensity at time $x = 0$, and τ is the luminescence lifetime. Data was fit using Prism. Q was calculated using Horrocks' method, eq. 3.

$$q = 5.0 \left(\frac{1}{\tau_{(\text{H}_2\text{O})}} - \frac{1}{\tau_{(\text{D}_2\text{O})}} - 0.06 \right) \quad (\text{S3})$$

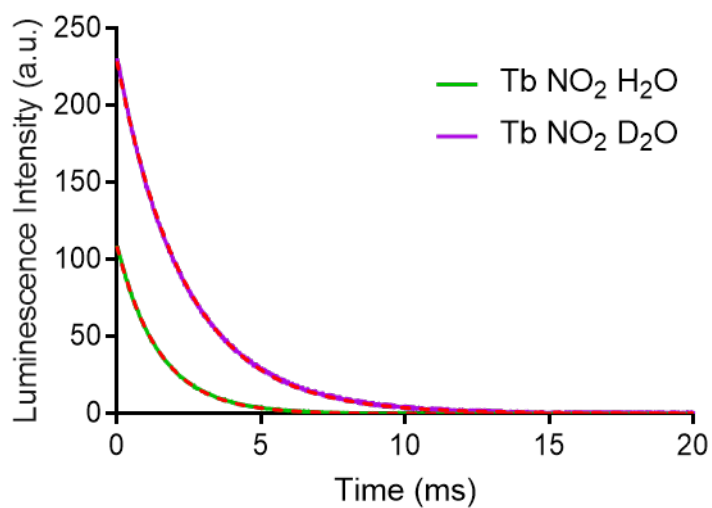


Figure S18. Luminescent lifetime decay curves of Tb(L3) in H₂O (buffer) and D₂O. Fits of Equation S2 indicated as dashed red overlay.

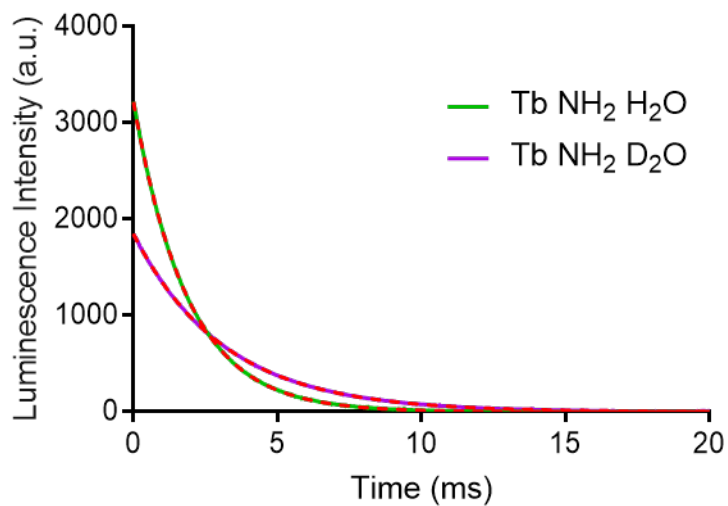


Figure S19. Luminescent lifetime decay curves of Tb(L4) in H₂O (buffer) and D₂O. Fits of Equation S2 indicated as dashed red overlay.

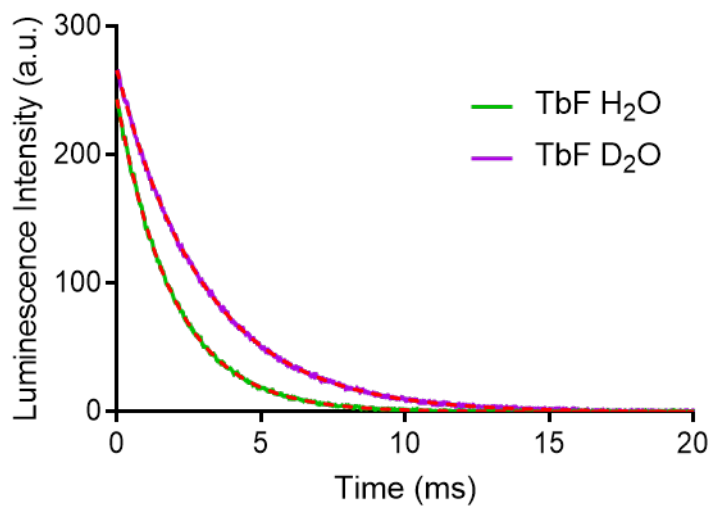


Figure S20. Luminescent lifetime decay curves of Tb(**L5**) in H₂O (buffer) and D₂O. Fits of Equation S2 indicated as dashed red overlay.

Table S7. Luminescent Lifetimes and q values.					
Complex	Buffer lifetime (ms)	D ₂ O lifetime (ms)	Buffer τ (ms ⁻¹)	D ₂ O τ (ms ⁻¹)	q
Tb(L3)	1.47	2.38	0.68	0.42	1.30
Tb(L4)	1.87	3.15	0.53	0.32	1.08
Tb(L5)	1.96	3.05	0.51	0.33	0.91

6. Characterization of hydrogen sulfide sensing

HPLC:

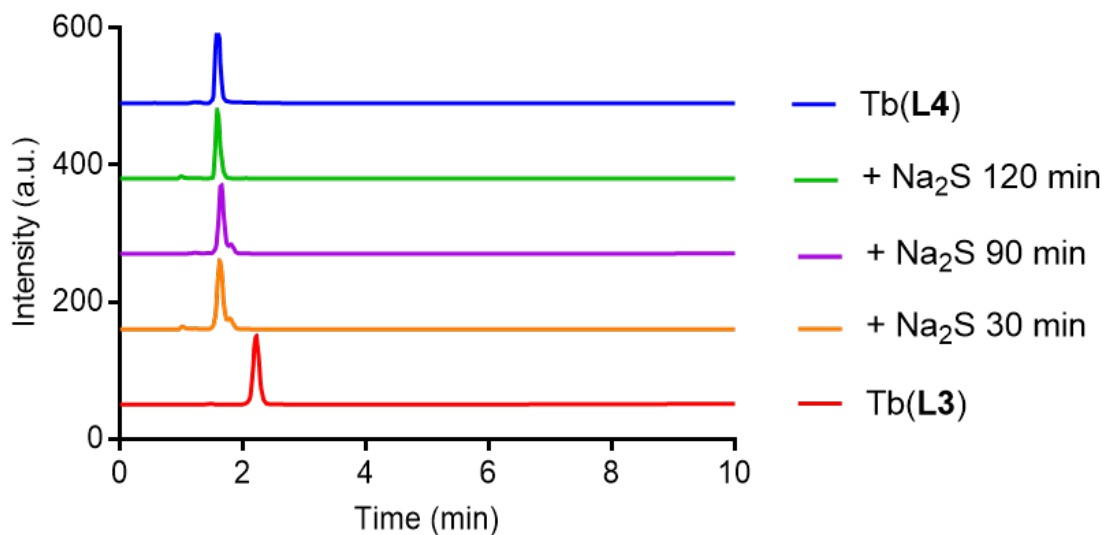


Figure S21. Normalized HPLC traces of Tb(L3) (red); Tb(L3) + 100 eq Na₂S after 30 min (orange), after 90 min (purple), after 120 min (green); overlay of Tb(L4) as complexed from ligand L2 (blue).

Table S8. Average fold-increase (n = 3) of Tb(L3) + analyte/salt;		
	Fold-increase	Std. deviation
Tb(L3)	1.000	1.000
NaOAc	1.621	0.567
NaI	1.619	0.230
NaHCO ₃	1.792	0.163
Na ₂ CO ₃	1.863	0.110
NaCl	1.758	0.240
Cys	0.765	0.147
Na ₂ SO ₄	1.730	0.007
Na ₂ S	31.321	5.421

7. ROI Analysis of CRET imaging

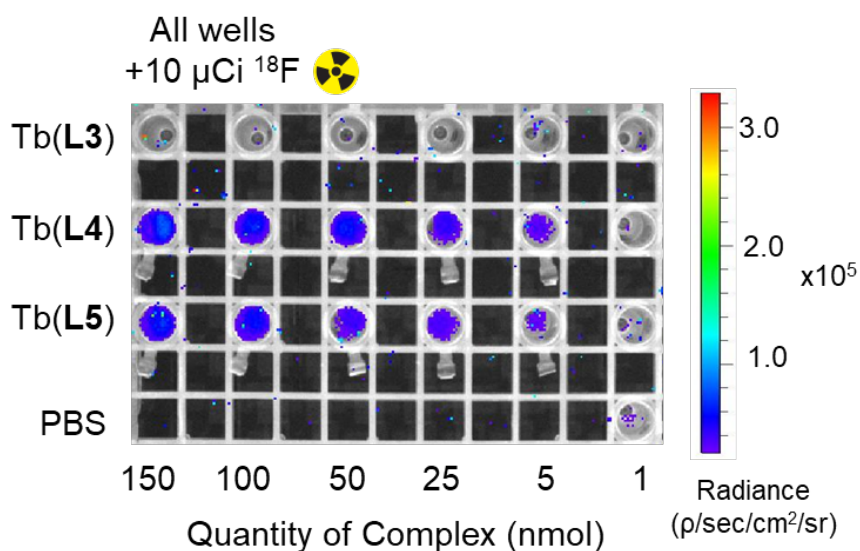


Figure S22. Plate of complexes featured in manuscript Figure 5, rewindowed to remove signal from buffer-only sample and Tb(L3) series.

Table S9. Average radiance values of all complexes. N = 3.			
	Quantity (nmol)	Average Radiance ($\text{p/sec/cm}^2/\text{sr}$)	Std. Deviation
Tb(L3)	150	2530	2254
	100	-2003	357
	50	-2427	335
	25	-3110	174
	5	-1570	1876
	1	-2840	528
Tb(L4)	150	36537	1152
	100	26970	743
	50	28160	1183
	25	14460	337
	5	8350	130
	1	-2373	178
Tb(L5)	150	31717	1685
	100	26520	573
	50	13647	464
	25	10820	650
	5	6110	316
	1	483	849

Table S10. Average radiance values of Tb(L3) in situ reduction with Na ₂ S over time		
Incubation time (hrs)	Average Radiance ($\rho/\text{sec}/\text{cm}^2/\text{sr}$)	Std. Deviation
3	80467	12150
2.5	71467	1482
2	65900	2146
1.5	46900	6871
1	31233	3311
0 (¹⁸ F in PBS)	167	3484

8. NMR spectra

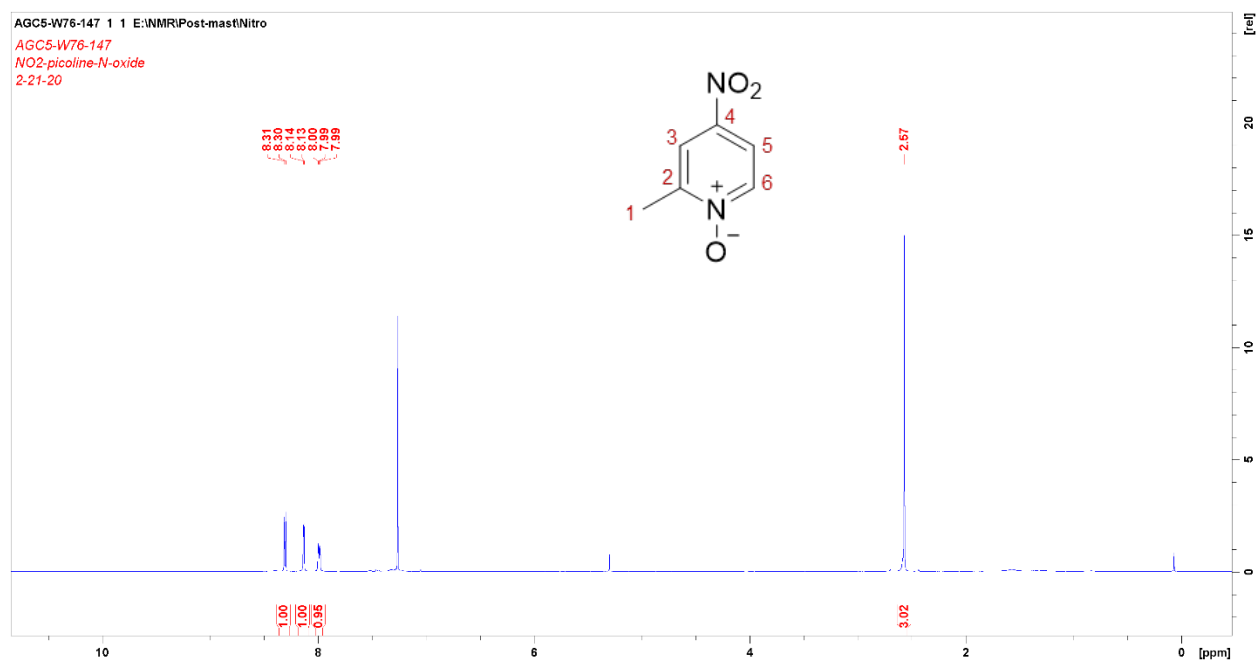


Figure S23. ^1H NMR spectra of 2-methyl-4-nitropyridine 1-oxide, 1.

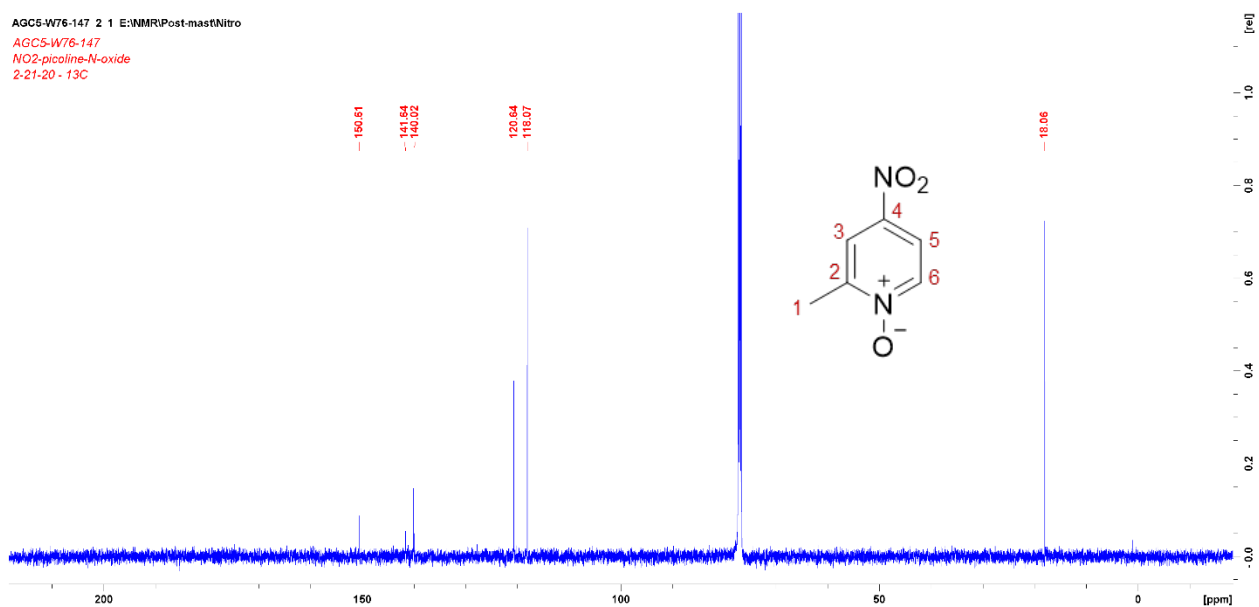


Figure S24. ^{13}C NMR spectra of 2-methyl-4-nitropyridine 1-oxide, 1.

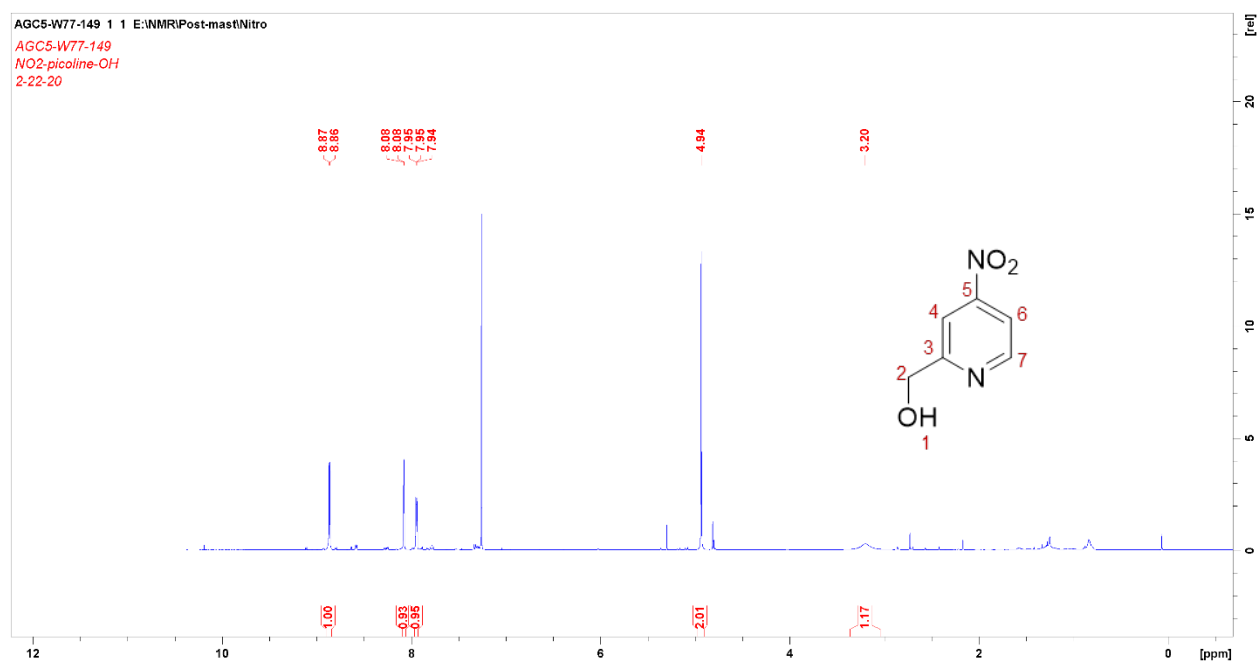


Figure S25. ^1H NMR spectra of 2-(Hydroxymethyl)-4-nitropyridine, 2.

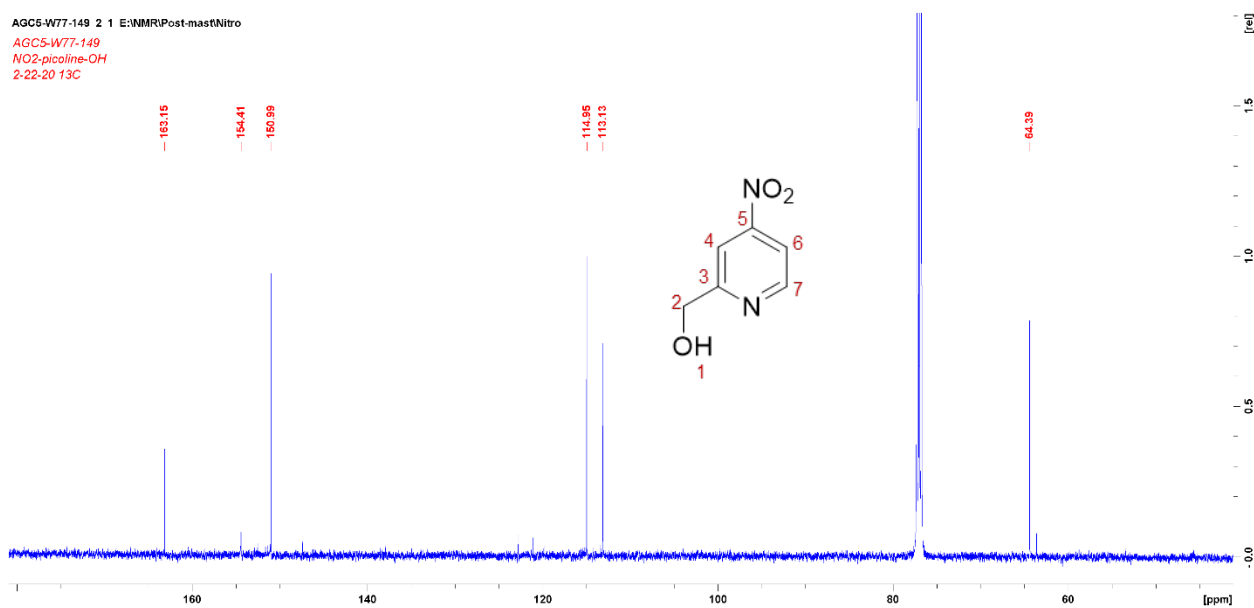


Figure S26. ^{13}C NMR spectra of 2-(Hydroxymethyl)-4-nitropyridine, 2.

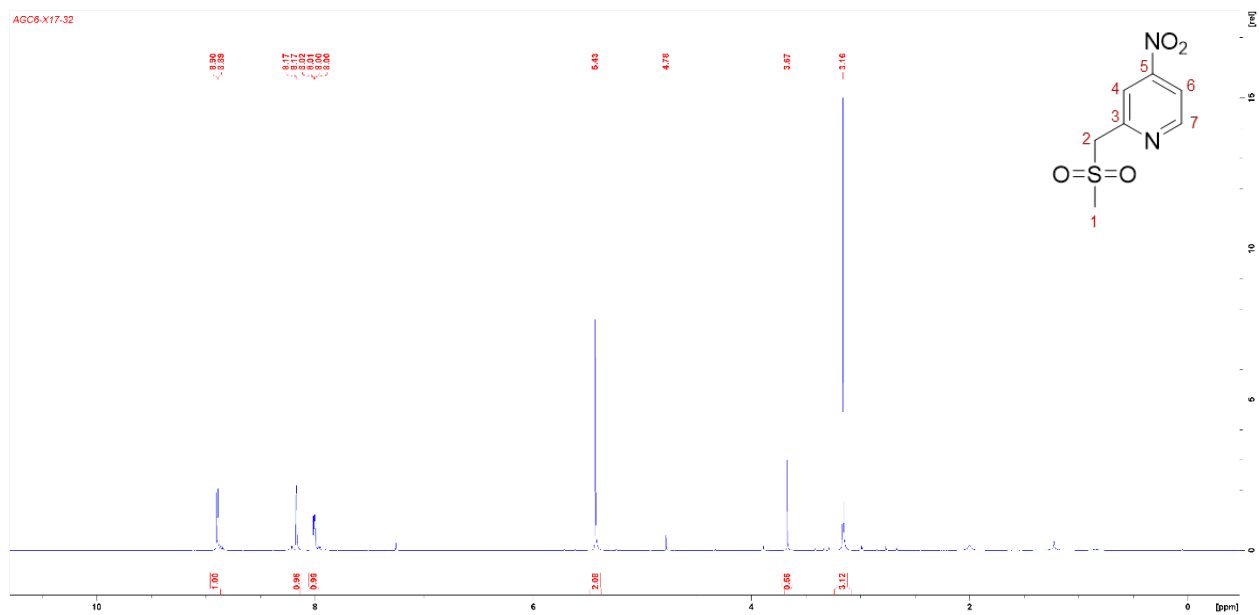


Figure S27. ^1H NMR spectra of 2-[(Methylsulfonyloxy)methyl]-4-nitropyridine, 3.

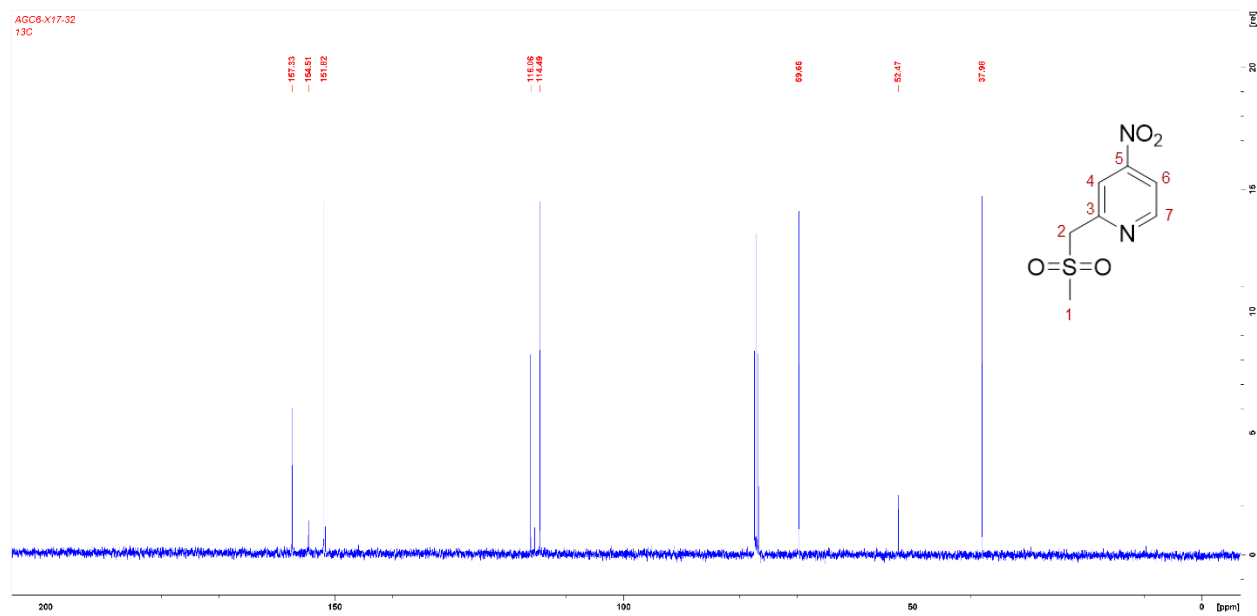


Figure S28. ^{13}C NMR spectra of 2-[(Methylsulfonyloxy)methyl]-4-nitropyridine, 3.

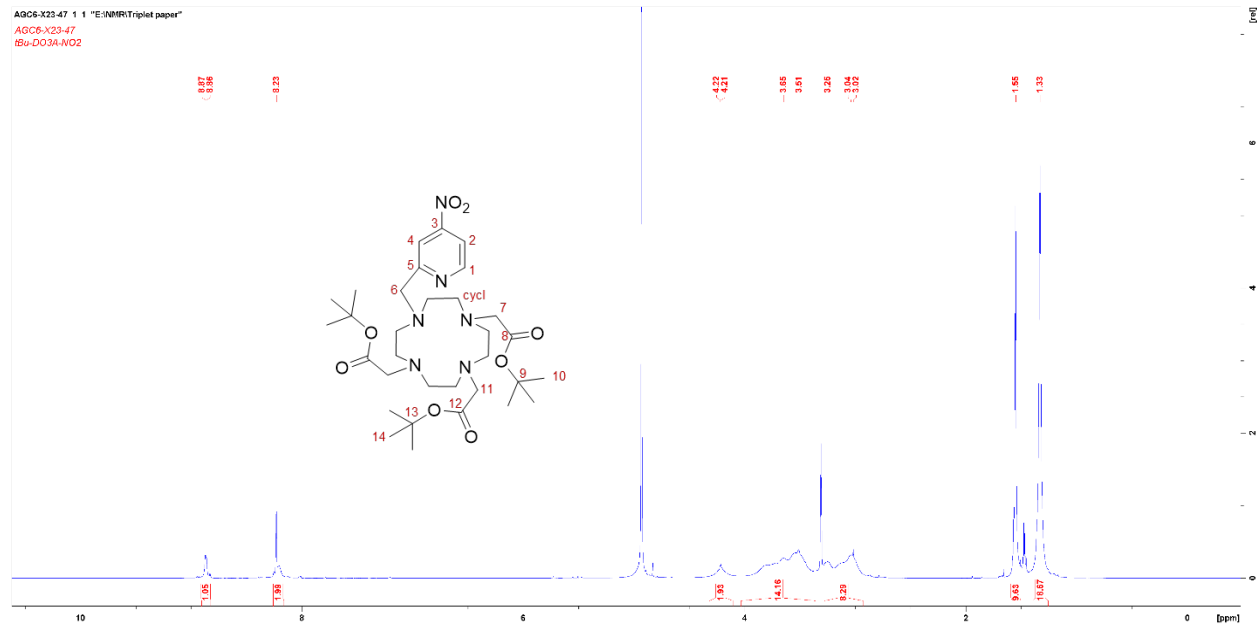


Figure S29. ^1H NMR spectra of *tert*-Butyl {4,10-bis(*tert*-butoxycarbonylmethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetate, 4.

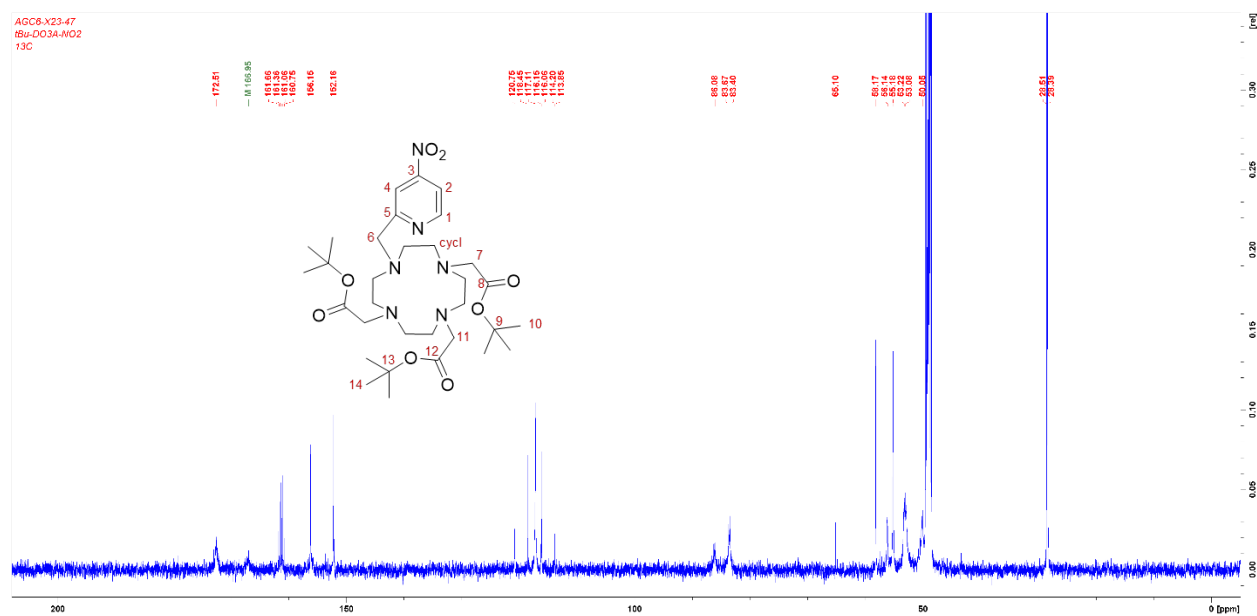


Figure S30. ^{13}C NMR spectra of *tert*-Butyl {4,10-bis(*tert*-butoxycarbonylmethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetate, 4.

AGC6-X26-49 1 1 "E:NMRI Triplet paper"

AGC6-X26-49

¹H

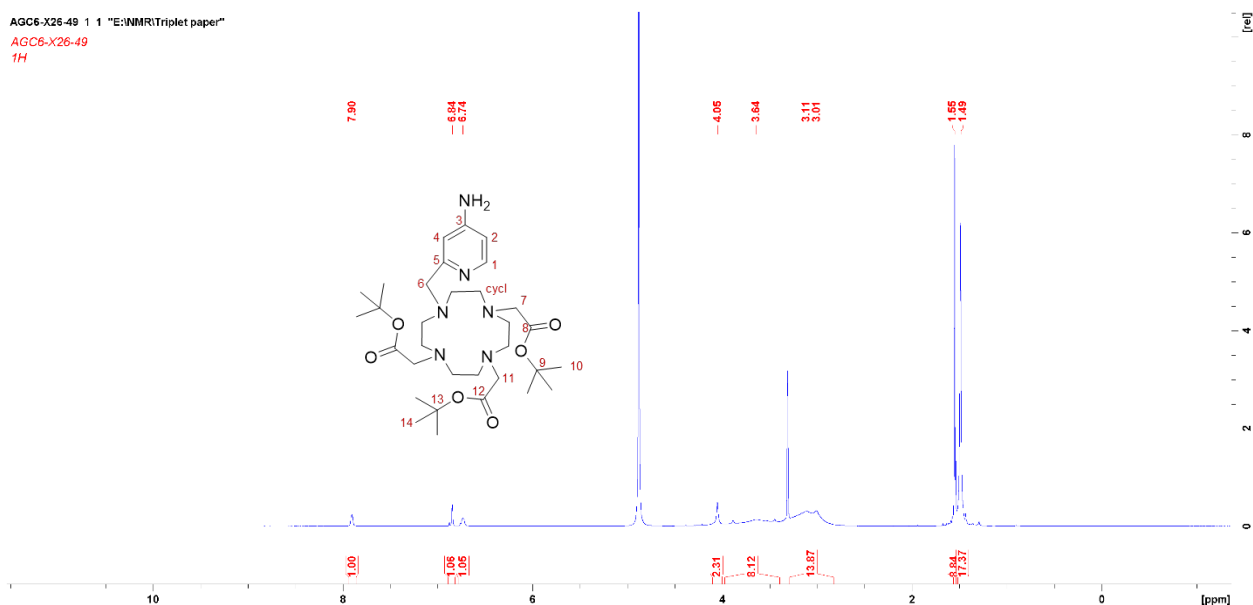


Figure S31. ¹H NMR spectra of *tert*-Butyl {10-[(4-amino-2-pyridyl)methyl]-4,7-bis(*tert*-butoxycarbonylmethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetate, 5.

AGC6-X26-49 2 1 "E:NMRI Triplet paper"

AGC6-X26-49

¹³C

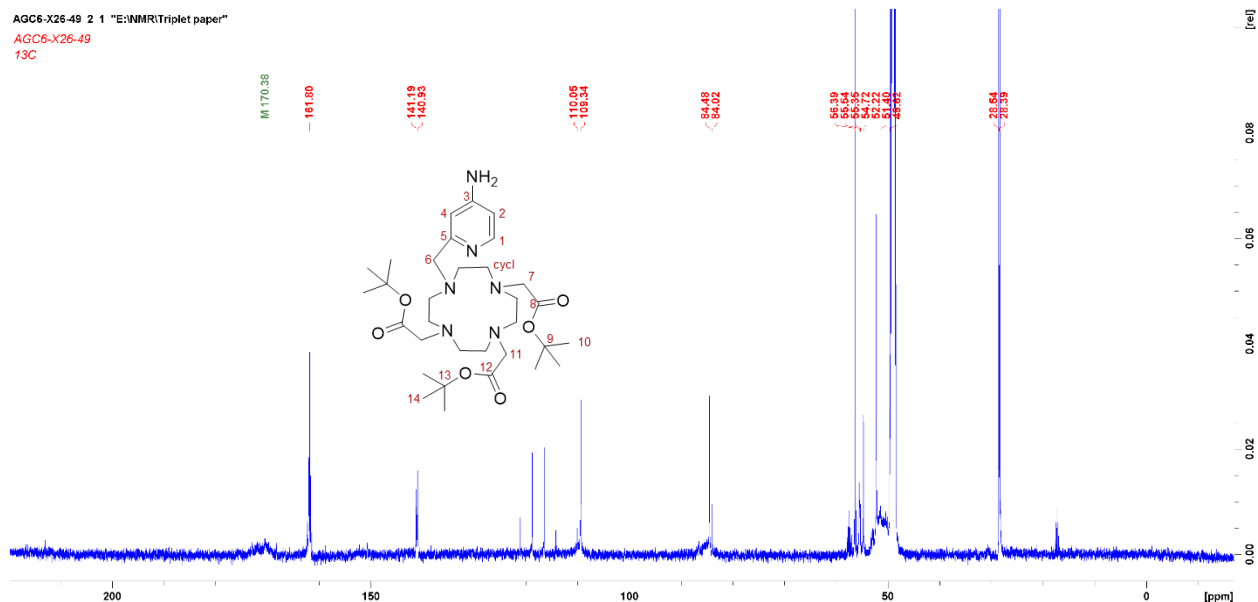
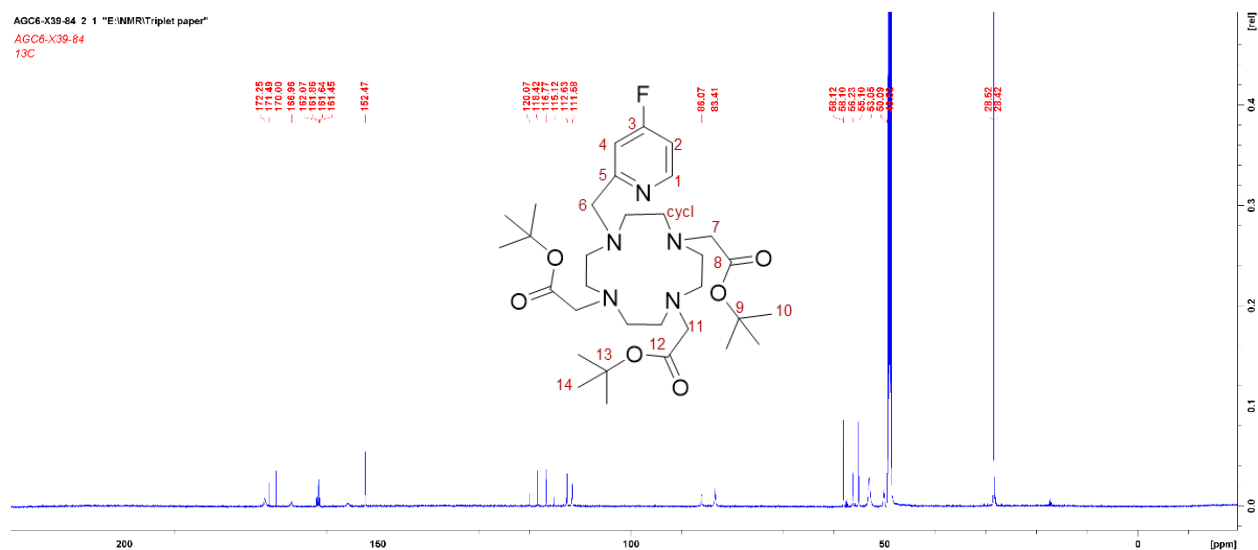
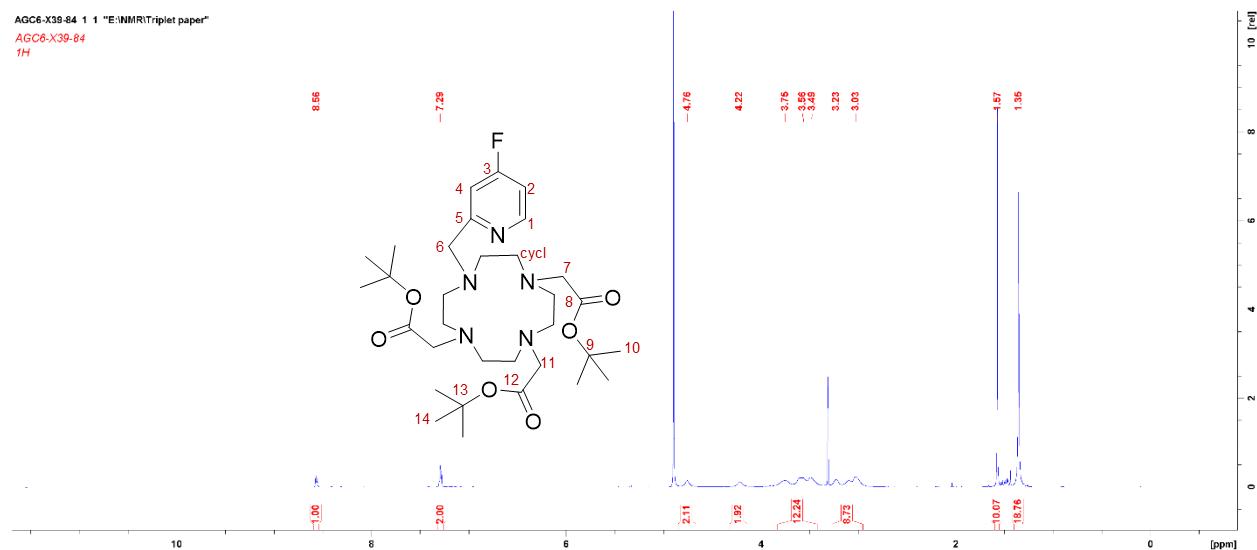


Figure S32. ¹³C NMR spectra of *tert*-Butyl {10-[(4-amino-2-pyridyl)methyl]-4,7-bis(*tert*-butoxycarbonylmethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetate, 5.



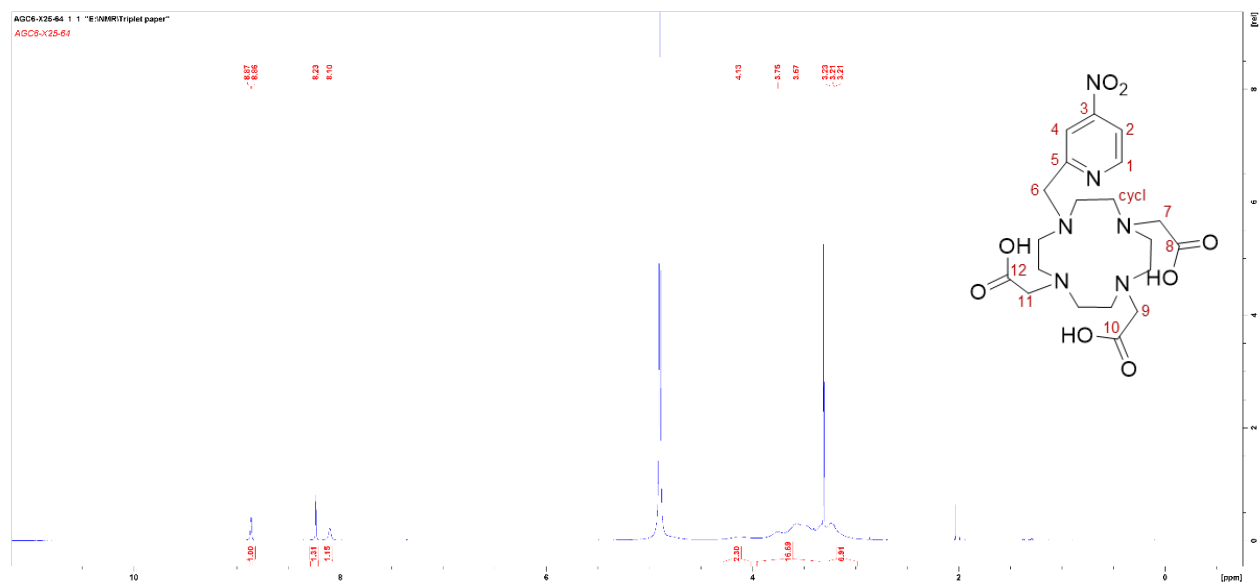


Figure S35. ^1H NMR spectra of {4,10-Bis(carboxymethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L3.

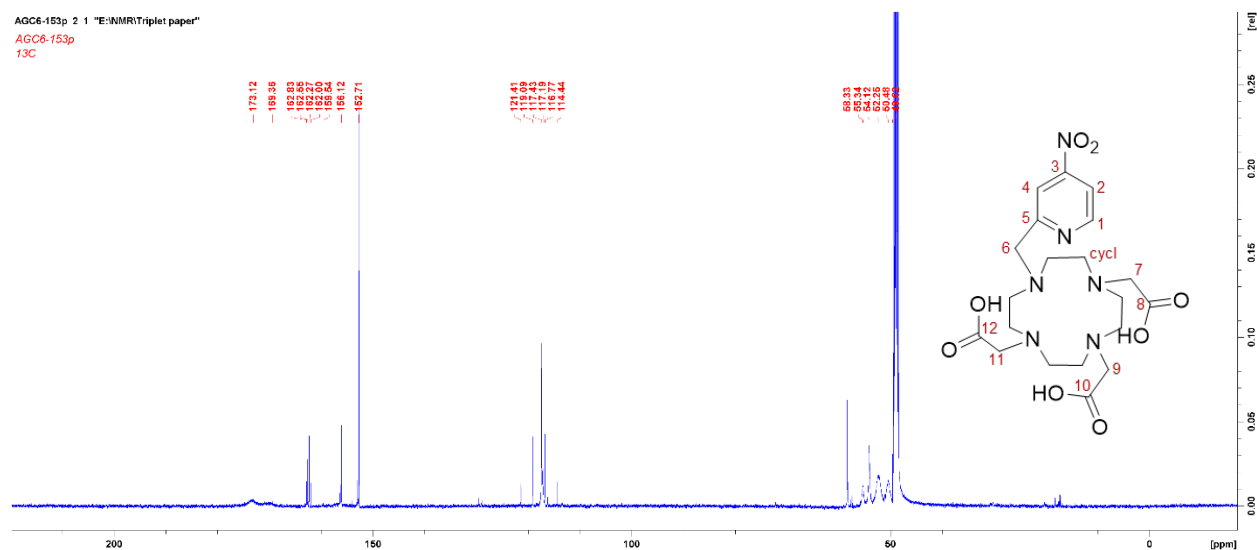


Figure S36. ^{13}C NMR spectra of {4,10-Bis(carboxymethyl)-7-[(4-nitro-2-pyridyl)methyl]-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L3.

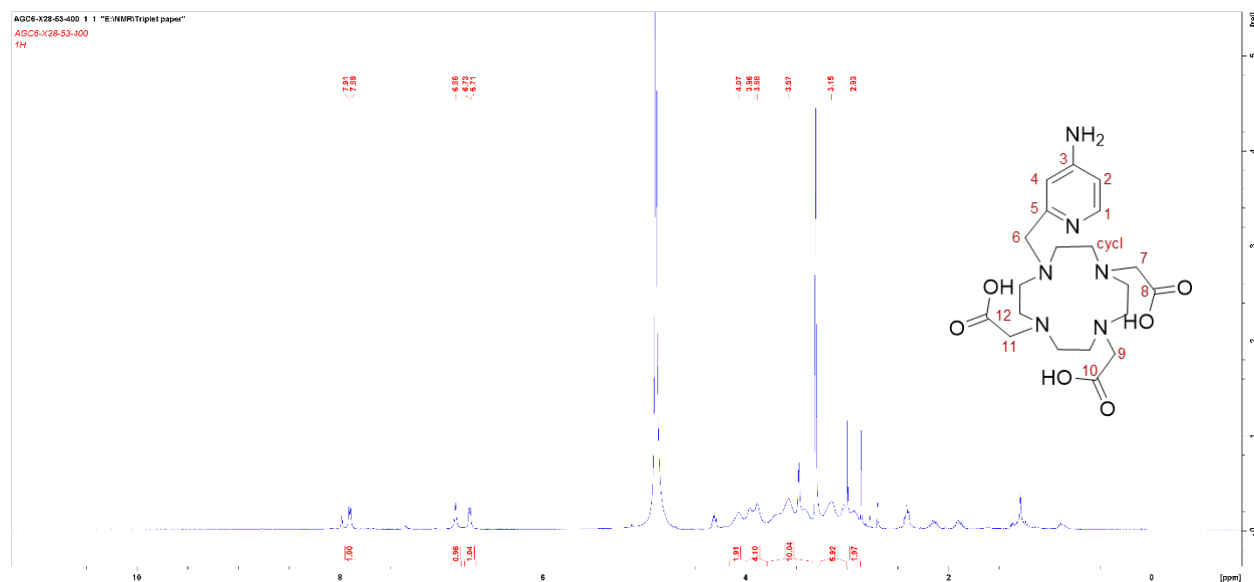


Figure S37. ^1H NMR spectra of {10-[(4-Amino-2-pyridyl)methyl]-4,7-bis(carboxymethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L4.

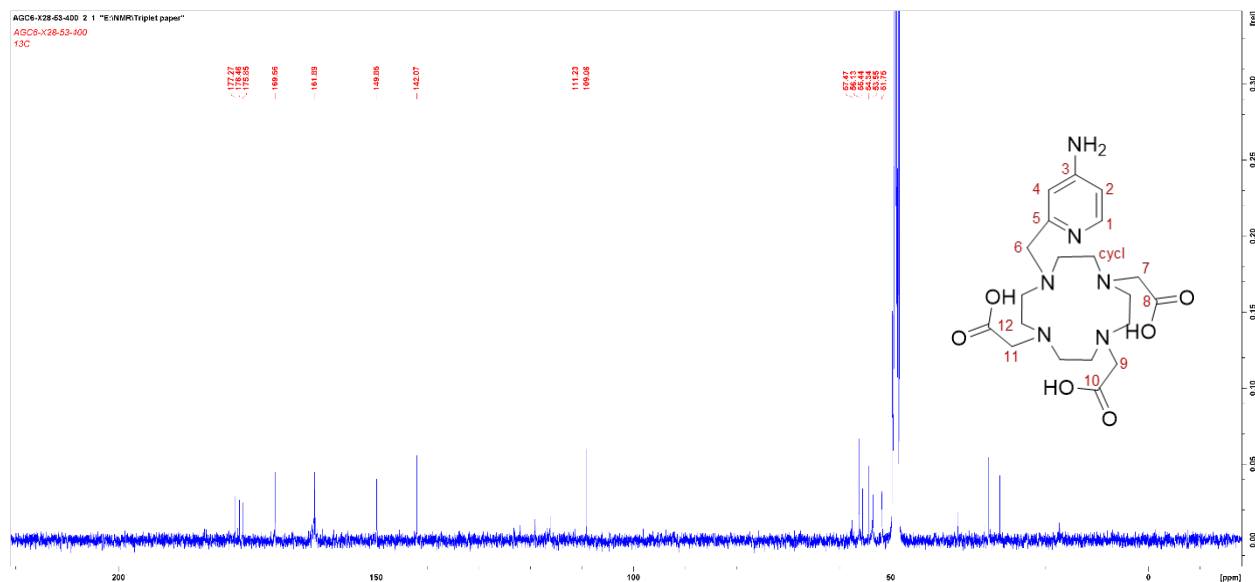
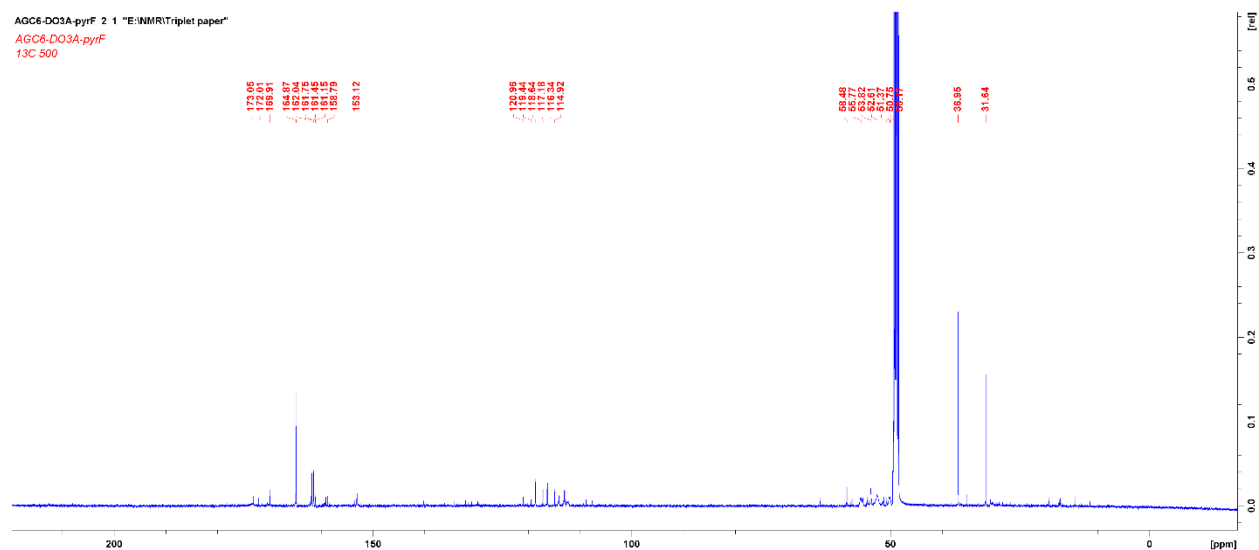
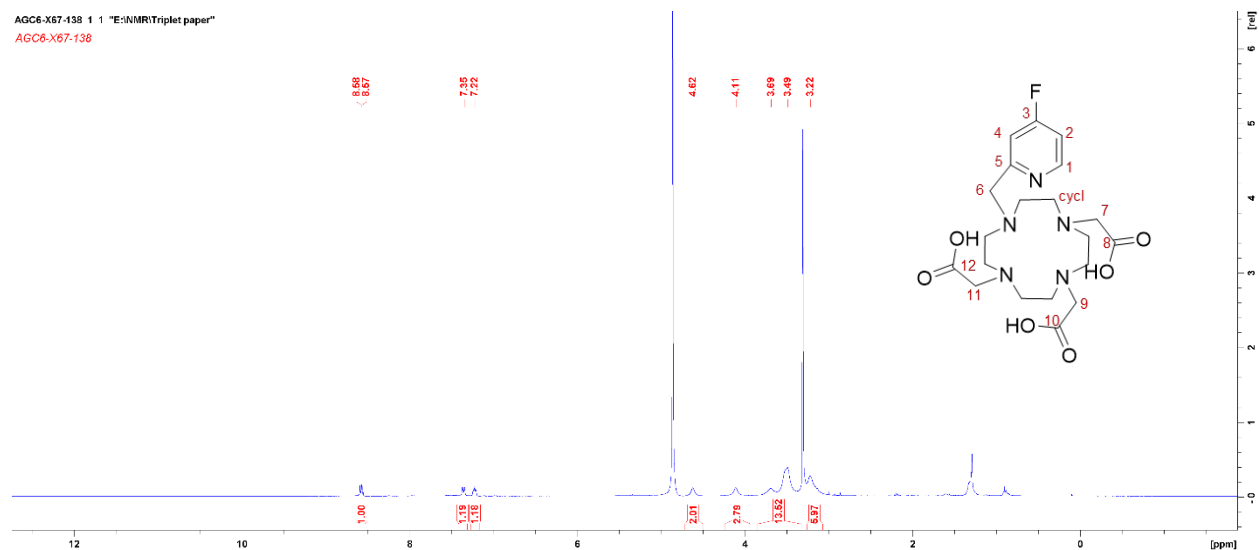


Figure S38. ^{13}C NMR spectra of {10-[(4-Amino-2-pyridyl)methyl]-4,7-bis(carboxymethyl)-1,4,7,10-tetraaza-1-cyclododecyl}acetic acid, L4.



9. Optimized Geometries

Tb(L3) GS		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.64118	-0.03474	-0.45863
2	O	0.24541	-2.2408	-0.95407
3	C	-0.0333	-3.23095	-0.15762
4	O	0.11577	-4.42966	-0.375
5	C	-0.6576	-2.7871	1.1868
6	N	-0.06154	-1.52985	1.71647
7	C	-1.02663	-0.83199	2.59814
8	C	-0.659	0.63124	2.83648
9	N	-0.49694	1.42385	1.59162
10	C	0.36634	2.60744	1.87831
11	C	1.85536	2.26629	1.84982
12	N	2.31663	1.74302	0.54367
13	C	3.66755	1.13855	0.68302
14	C	3.6209	-0.2414	1.3386
15	N	2.74391	-1.20964	0.63006
16	C	2.33355	-2.28734	1.56754
17	C	1.17972	-1.84695	2.46826
18	H	0.98072	-2.64023	3.20769
19	H	1.47444	-0.95478	3.03098
20	H	3.17771	-2.59724	2.20345
21	H	2.0409	-3.1619	0.98299
22	C	3.44537	-1.78345	-0.5555
23	C	3.51851	-0.79519	-1.73812
24	O	2.43629	-0.06792	-1.88224
25	O	4.5136	-0.76969	-2.45029
26	H	2.85477	-2.63829	-0.9008
27	H	4.45492	-2.12529	-0.2924
28	H	4.64674	-0.63669	1.40201
29	H	3.25826	-0.15021	2.36781
30	H	4.32922	1.78889	1.27787
31	H	4.10086	1.06262	-0.31483
32	C	2.3448	2.82541	-0.48131
33	C	0.93848	3.18502	-1.00205
34	O	0.16842	2.14325	-1.19531
35	O	0.63703	4.35719	-1.2039
36	H	2.90701	2.43747	-1.33647
37	H	2.84305	3.72554	-0.09719
38	H	2.42233	3.1715	2.12381
39	H	2.07708	1.51236	2.61328
40	H	0.12178	3.03065	2.86543
41	H	0.15129	3.38249	1.14165

42	C	-1.79913	1.90647	1.06748
43	C	-2.61986	0.86669	0.33799
44	N	-1.95339	-0.01004	-0.43827
45	C	-2.63569	-0.87392	-1.21183
46	C	-4.02988	-0.91353	-1.24355
47	C	-4.69831	-0.01675	-0.41911
48	N	-6.18072	-0.02554	-0.38841
49	O	-6.72928	0.76614	0.37902
50	O	-6.75114	-0.82472	-1.129
51	C	-4.01762	0.88822	0.38622
52	H	-4.56563	1.57945	1.01492
53	H	-4.57433	-1.60891	-1.86924
54	H	-2.02612	-1.5423	-1.81177
55	H	-1.56705	2.67428	0.3204
56	H	-2.40352	2.3637	1.86657
57	H	-1.42038	1.0887	3.48953
58	H	0.28726	0.68561	3.38265
59	H	-1.11301	-1.3344	3.57598
60	H	-2.00855	-0.8934	2.12406
61	H	-1.7196	-2.59937	0.99595
62	H	-0.58601	-3.60271	1.91858
63	O	-0.09087	0.10084	-2.86276
64	H	0.80525	-0.09881	-3.19884
65	H	-0.12529	1.07895	-2.80822

Tb(L3) T ₁		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.63016	-0.01655	-0.46025
2	O	0.2712	-2.24746	-0.9724
3	C	-0.15685	-3.19887	-0.21092
4	O	-0.11964	-4.40974	-0.44341
5	C	-0.81432	-2.7326	1.10829
6	N	-0.17201	-1.51663	1.67166
7	C	-1.13092	-0.77913	2.53123
8	C	-0.70172	0.66494	2.78475
9	N	-0.47675	1.4532	1.54651
10	C	0.41236	2.60937	1.85797
11	C	1.88888	2.21947	1.88594
12	N	2.37987	1.67278	0.59851
13	C	3.69454	1.00818	0.79719

14	C	3.55699	-0.36886	1.44402
15	N	2.67572	-1.29575	0.68787
16	C	2.19386	-2.37034	1.59458
17	C	1.02523	-1.90179	2.46028
18	H	0.76744	-2.70148	3.17404
19	H	1.33325	-1.03472	3.0537
20	H	3.00362	-2.7191	2.25466
21	H	1.89299	-3.22613	0.98724
22	C	3.40353	-1.88363	-0.47054
23	C	3.52964	-0.90086	-1.65289
24	O	2.49815	-0.12947	-1.83091
25	O	4.54665	-0.93113	-2.34453
26	H	2.80862	-2.72435	-0.84075
27	H	4.39777	-2.24577	-0.17869
28	H	4.56055	-0.80797	1.55852
29	H	3.14624	-0.26399	2.45334
30	H	4.35687	1.62709	1.42371
31	H	4.17122	0.91609	-0.17951
32	C	2.51019	2.75393	-0.41869
33	C	1.15569	3.17384	-1.02186
34	O	0.33153	2.18033	-1.22036
35	O	0.94969	4.35491	-1.28842
36	H	3.10087	2.34674	-1.24523
37	H	3.02566	3.63136	-0.00616
38	H	2.47829	3.10461	2.17654
39	H	2.05574	1.46178	2.6593
40	H	0.14606	3.04872	2.83227
41	H	0.24704	3.38381	1.10784
42	C	-1.75348	1.97719	0.98802
43	C	-2.5958	0.94263	0.27779
44	N	-1.92663	0.06773	-0.51411
45	C	-2.64583	-0.7893	-1.27819
46	C	-4.02942	-0.83721	-1.27691
47	C	-4.71032	0.05643	-0.42501
48	N	-6.08467	0.04196	-0.35763
49	O	-6.77585	0.81979	0.40848
50	O	-6.82993	-0.76216	-1.04272
51	C	-3.97991	0.96727	0.36657
52	H	-4.49873	1.66704	1.01108
53	H	-4.57492	-1.53496	-1.89978
54	H	-2.06025	-1.45206	-1.90703
55	H	-1.47462	2.7186	0.23162
56	H	-2.34628	2.47567	1.77024
57	H	-1.45893	1.15351	3.41924
58	H	0.23213	0.67586	3.35509

59	H	-1.26215	-1.28171	3.50355
60	H	-2.10079	-0.79236	2.03119
61	H	-1.85278	-2.48606	0.86555
62	H	-0.82193	-3.55772	1.83256
63	O	0.0053	0.12958	-2.8829
64	H	0.89798	-0.07872	-3.21905
65	H	-0.03075	1.10783	-2.85035

Tb(L4) GS		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.34407	-0.02207	-0.47022
2	O	-0.00987	-2.22641	-1.02744
3	C	-0.4216	-3.21461	-0.29132
4	O	-0.35916	-4.41487	-0.54603
5	C	-1.10124	-2.77325	1.0269
6	N	-0.48511	-1.56018	1.62857
7	C	-1.46639	-0.85471	2.48516
8	C	-1.06371	0.58949	2.77734
9	N	-0.83184	1.40778	1.5623
10	C	0.0384	2.56589	1.91371
11	C	1.51945	2.19271	1.95499
12	N	2.03526	1.68077	0.66526
13	C	3.35654	1.03162	0.86611
14	C	3.23132	-0.36279	1.47851
15	N	2.37313	-1.28332	0.69099
16	C	1.89083	-2.3828	1.56518
17	C	0.70507	-1.94697	2.42563
18	H	0.44718	-2.76542	3.11862
19	H	0.99515	-1.08942	3.04217
20	H	2.69503	-2.73898	2.22885
21	H	1.60557	-3.22614	0.93341
22	C	3.12323	-1.833	-0.47501
23	C	3.28251	-0.81093	-1.61985
24	O	2.22835	-0.05439	-1.79999
25	O	4.31734	-0.7899	-2.27547
26	H	2.53014	-2.6593	-0.87977
27	H	4.10937	-2.20879	-0.17121
28	H	4.24033	-0.78984	1.59358

29	H	2.80828	-0.28734	2.48582
30	H	4.00103	1.64564	1.51645
31	H	3.8467	0.9682	-0.10598
32	C	2.15743	2.78271	-0.33061
33	C	0.79822	3.20381	-0.92489
34	O	0.00102	2.19808	-1.17613
35	O	0.56088	4.39112	-1.12988
36	H	2.75399	2.39337	-1.16147
37	H	2.6636	3.6563	0.10223
38	H	2.09217	3.07978	2.27381
39	H	1.68478	1.42103	2.71502
40	H	-0.24702	2.97867	2.89491
41	H	-0.12295	3.35679	1.18013
42	C	-2.10236	1.92453	0.99006
43	C	-2.91129	0.90334	0.22085
44	N	-2.21192	0.04605	-0.55304
45	C	-2.90736	-0.78937	-1.35442
46	C	-4.29035	-0.81779	-1.41259
47	C	-5.03018	0.06191	-0.59359
48	N	-6.3953	0.09217	-0.63291
49	H	-6.90596	0.60179	0.07103
50	H	-6.89581	-0.6459	-1.1031
51	C	-4.30069	0.93341	0.24377
52	H	-4.81891	1.63899	0.88734
53	H	-4.79165	-1.50884	-2.08275
54	H	-2.30354	-1.45404	-1.96451
55	H	-1.81914	2.69342	0.26296
56	H	-2.72372	2.38905	1.77265
57	H	-1.83683	1.05331	3.41282
58	H	-0.13819	0.60044	3.36119
59	H	-1.61034	-1.37904	3.44511
60	H	-2.42642	-0.8691	1.96564
61	H	-2.13948	-2.53062	0.77759
62	H	-1.1102	-3.61099	1.7376
63	O	-0.21746	0.19978	-2.91428
64	H	0.70785	0.01772	-3.17523
65	H	-0.26085	1.17453	-2.8207

Tb(L4) T ₁		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.34933	-0.02154	-0.46213
2	O	-0.07635	-2.21064	-1.0403
3	C	-0.58486	-3.16832	-0.3265
4	O	-0.61132	-4.36737	-0.59642
5	C	-1.26133	-2.6936	0.98095
6	N	-0.59118	-1.52081	1.6045
7	C	-1.55128	-0.76882	2.44661
8	C	-1.08903	0.65517	2.75186
9	N	-0.79622	1.45758	1.54106
10	C	0.11458	2.57951	1.89469
11	C	1.57757	2.14622	1.9755
12	N	2.10321	1.59878	0.70351
13	C	3.38294	0.8837	0.94616
14	C	3.16892	-0.50154	1.55435
15	N	2.2957	-1.38033	0.73699
16	C	1.74046	-2.46212	1.58975
17	C	0.55741	-1.9756	2.42518
18	H	0.24088	-2.78676	3.10233
19	H	0.87268	-1.13977	3.05883
20	H	2.51103	-2.86319	2.26766
21	H	1.42862	-3.28409	0.94254
22	C	3.05407	-1.9575	-0.40897
23	C	3.27747	-0.94544	-1.55187
24	O	2.26394	-0.14242	-1.75202
25	O	4.32332	-0.97509	-2.19071
26	H	2.44234	-2.76239	-0.82864
27	H	4.01835	-2.36852	-0.08151
28	H	4.15158	-0.97663	1.70497
29	H	2.71459	-0.40327	2.54583
30	H	4.03706	1.46551	1.61612
31	H	3.90035	0.79334	-0.00967
32	C	2.31467	2.68609	-0.29211
33	C	1.00898	3.16444	-0.9578
34	O	0.15553	2.2065	-1.20473
35	O	0.86931	4.35412	-1.22921
36	H	2.92821	2.26701	-1.09561
37	H	2.84035	3.54047	0.15545
38	H	2.18184	3.01014	2.29973
39	H	1.69089	1.37558	2.74605
40	H	-0.17319	3.02234	2.86235
41	H	-0.0007	3.36296	1.14373
42	C	-2.045	2.02975	0.93168

43	C	-2.91587	1.01227	0.28116
44	N	-2.19017	0.15751	-0.65911
45	C	-2.89112	-0.51294	-1.57605
46	C	-4.28431	-0.56656	-1.58465
47	C	-5.03019	0.15817	-0.51076
48	N	-6.40555	0.07197	-0.55301
49	H	-6.93849	0.46355	0.21238
50	H	-6.82275	-0.74174	-0.98706
51	C	-4.30316	0.96258	0.36462
52	H	-4.81707	1.57726	1.1009
53	H	-4.8184	-1.1199	-2.34973
54	H	-2.30501	-1.06498	-2.30633
55	H	-1.68309	2.72935	0.16583
56	H	-2.61827	2.59528	1.68458
57	H	-1.85926	1.15063	3.36677
58	H	-0.1792	0.62767	3.36049
59	H	-1.7418	-1.29232	3.39915
60	H	-2.49709	-0.72231	1.90342
61	H	-2.27827	-2.38979	0.71238
62	H	-1.33386	-3.53349	1.68559
63	O	-0.12872	0.21618	-2.93299
64	H	0.79467	-0.01969	-3.15594
65	H	-0.11528	1.19192	-2.83367

Tb(L5) GS		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.34383	-0.02187	-0.46558
2	O	-0.01407	-2.22021	-1.03006
3	O	-0.23038	-4.42066	-0.52575
4	O	2.20378	-0.0228	-1.81567
5	O	4.29874	-0.72263	-2.31776
6	O	-0.07147	2.18339	-1.16436
7	O	0.42333	4.39116	-1.10465
8	O	-0.27018	0.18501	-2.89784
9	N	-0.85604	1.3873	1.57687
10	N	-0.44467	-1.57274	1.63905
11	N	2.40072	-1.24161	0.67295
12	N	1.99792	1.71539	0.65143

13	N	-2.2344	0.00821	-0.53616
14	F	-6.33198	0.00545	-0.62868
15	C	-1.06701	0.56287	2.79276
16	C	-1.43711	-0.89019	2.50182
17	C	0.76268	-1.9247	2.42807
18	C	1.94829	-2.34469	1.55949
19	C	3.25262	-0.30107	1.44538
20	C	3.33684	1.09611	0.83168
21	C	1.48675	2.20979	1.95
22	C	-0.0006	2.55857	1.92639
23	C	-2.13668	1.8874	1.01593
24	C	-1.03282	-2.80591	1.04867
25	C	-0.35666	-3.22457	-0.27866
26	C	3.1464	-1.78364	-0.49997
27	C	3.2741	-0.76125	-1.64845
28	C	2.07907	2.82329	-0.34257
29	C	0.70057	3.21047	-0.91519
30	C	-2.93254	0.86472	0.23566
31	C	-4.32917	0.88946	0.23994
32	C	-4.99178	0.00687	-0.60795
33	C	-4.29016	-0.87289	-1.42622
34	C	-2.90053	-0.83462	-1.34904
35	H	0.64657	0.00382	-3.18643
36	H	-0.3155	1.16032	-2.81179
37	H	-1.84799	1.00879	3.43086
38	H	-0.14025	0.59455	3.37342
39	H	-1.56403	-1.41765	3.46209
40	H	-2.40012	-0.92891	1.98867
41	H	0.52937	-2.74006	3.13289
42	H	1.04093	-1.05361	3.03091
43	H	2.76544	-2.68006	2.21769
44	H	1.67366	-3.19847	0.93682
45	H	4.27231	-0.70599	1.54201
46	H	2.84629	-0.23528	2.46013
47	H	3.97686	1.72416	1.47269
48	H	3.81287	1.04449	-0.14807
49	H	2.04705	3.10454	2.2689
50	H	1.67317	1.43617	2.7031
51	H	-0.28283	2.95997	2.91284
52	H	-0.18217	3.3517	1.20001
53	H	-1.87302	2.67	0.29556
54	H	-2.7662	2.33048	1.80387
55	H	-2.08218	-2.59648	0.81565
56	H	-1.0052	-3.64208	1.76026
57	H	2.56403	-2.62234	-0.89504

58	H	4.14224	-2.14112	-0.20662
59	H	2.67435	2.45193	-1.18257
60	H	2.56817	3.70835	0.08614
61	H	-4.88752	1.57501	0.86704
62	H	-4.80619	-1.55664	-2.08879
63	H	-2.27737	-1.49139	-1.9481

Tb(L5) T ₁		0 imaginary frequencies		
Center Number	Atom	X	Y	Z
1	Tb	0.32922	-0.01383	-0.45083
2	O	0.0417	-2.22463	-1.0739
3	O	-0.43093	-4.39701	-0.67472
4	O	2.23472	-0.02405	-1.77468
5	O	4.31227	-0.76559	-2.27648
6	O	0.03706	2.21961	-1.14564
7	O	0.69089	4.38213	-1.19736
8	O	-0.1965	0.23986	-2.90389
9	N	-0.82821	1.36945	1.59004
10	N	-0.49191	-1.60251	1.58368
11	N	2.36723	-1.31036	0.68617
12	N	2.04229	1.65958	0.71299
13	N	-2.21373	0.03982	-0.59515
14	F	-6.39362	0.02917	-0.55364
15	C	-1.05823	0.52468	2.78727
16	C	-1.47064	-0.90782	2.45318
17	C	0.68327	-2.03166	2.38121
18	C	1.87862	-2.43837	1.52111
19	C	3.20595	-0.40763	1.51387
20	C	3.35427	0.99557	0.92873
21	C	1.51354	2.16661	2.00135
22	C	0.03068	2.52977	1.95097
23	C	-2.12003	1.8871	1.02573
24	C	-1.12769	-2.78534	0.94254
25	C	-0.44018	-3.19982	-0.37707
26	C	3.13861	-1.82455	-0.47872
27	C	3.28067	-0.78473	-1.61039

28	C	2.19721	2.77022	-0.26656
29	C	0.86796	3.19859	-0.91608
30	C	-2.94213	0.82066	0.39392
31	C	-4.38535	0.79328	0.44017
32	C	-5.05077	0.14459	-0.54246
33	C	-4.32565	-0.51141	-1.63503
34	C	-2.90709	-0.52835	-1.56004
35	H	0.72572	0.05416	-3.17071
36	H	-0.23442	1.21447	-2.80635
37	H	-1.83207	0.97234	3.43257
38	H	-0.1327	0.51847	3.37214
39	H	-1.63414	-1.46193	3.39264
40	H	-2.42482	-0.87989	1.9188
41	H	0.41001	-2.87605	3.03546
42	H	0.96706	-1.20269	3.03794
43	H	2.67706	-2.81418	2.18064
44	H	1.60246	-3.26106	0.85905
45	H	4.20956	-0.8409	1.65128
46	H	2.75333	-0.34652	2.50889
47	H	3.9907	1.59251	1.60194
48	H	3.86509	0.94516	-0.03367
49	H	2.08167	3.05417	2.32592
50	H	1.67751	1.39369	2.76011
51	H	-0.26389	2.94066	2.93014
52	H	-0.13663	3.31819	1.21516
53	H	-1.82079	2.61946	0.26015
54	H	-2.70035	2.40923	1.80353
55	H	-2.15561	-2.51404	0.68342
56	H	-1.16441	-3.64236	1.62828
57	H	2.5695	-2.65625	-0.90566
58	H	4.13106	-2.18309	-0.17574
59	H	2.82216	2.39392	-1.08229
60	H	2.68775	3.64086	0.18896
61	H	-4.91834	1.30048	1.2379
62	H	-4.8597	-0.99947	-2.4399
63	H	-2.33056	-1.05551	-2.31676

10. References

1. Pershagen, E.; Borbas, K. E., Multiplex detection of enzymatic activity with responsive lanthanide-based luminescent probes. *Angew. Chem. Int. Ed.* 2015, **54**, 1787-1790.
2. Butler, S. J., Quantitative determination of fluoride in pure water using luminescent europium complexes. *Chem. Commun.* 2015, **51**, 10879-10882.
3. Page, S. E.; Wilke, K. T.; Pierre, V. C., Sensitive and selective time-gated luminescence detection of hydroxyl radical in water. *Chem. Commun.* 2010, **46**, 2423-2425.
4. Huang, S.-Y.; Pierre, V. C., A turn-on luminescent europium probe for cyanide detection in water. *Chem. Commun.* 2018, **54**, 9210-9213.
5. Liu, T.; Nonat, A.; Beyler, M.; Regueiro-Figueroa, M.; Nchimi Nono, K.; Jeannin, O.; Camerel, F.; Debaene, F.; Cianfèrani-Sanglier, S.; Tripier, R.; Platas-Iglesias, C.; Charbonnière, L. J., Supramolecular Luminescent Lanthanide Dimers for Fluoride Sequestering and Sensing. *Angew. Chem. Int. Ed.* 2014, **53**, 7259-7263.
6. Huang, S.-Y.; Qian, M.; Pierre, V. C., A Combination of Factors: Tuning the Affinity of Europium Receptors for Phosphate in Water. *Inorg. Chem.* **2019**, **58**, 16087-16099.
7. Liu, X.; Song, B.; Ma, H.; Tang, Z.; Yuan, J., Development of a mitochondria targetable ratiometric time-gated luminescence probe for biothiols based on lanthanide complexes. *J. Mater. Chem. B.* 2018, **6**, 1844-1851.
8. Xiao, Y.; Zhang, R.; Ye, Z.; Dai, Z.; An, H.; Yuan, J., Lanthanide Complex-Based Luminescent Probes for Highly Sensitive Time-Gated Luminescence Detection of Hypochlorous Acid. *Anal. Chem.* 2012, **84**, 10785-10792.
9. Aulsebrook, M. L.; Starck, M.; Grace, M. R.; Graham, B.; Thordarson, P.; Pal, R.; Tuck, K. L., Interaction of Nucleotides with a Trinuclear Terbium(III)–Dizinc(II) Complex: Efficient Sensitization of Terbium Luminescence by Guanosine Monophosphate and Application to Real-Time Monitoring of Phosphodiesterase Activity. *Inorg. Chem.* 2019, **58**, 495-505.
10. Tropiano, M.; Faulkner, S., A lanthanide based sensor for the time-gated detection of hydrogen sulfide. *Chem. Commun.* 2014, **50**, 4696-8.
11. Song, B.; Wang, G.; Yuan, J., A new europium chelate-based phosphorescence probe specific for singlet oxygen. *Chem. Commun.* 2005, **28**, 3553-3555.
12. Parker, D.; Kanthi Senanayake, P.; A. Gareth Williams, J., Luminescent sensors for pH, pO₂, halide and hydroxide ions using phenanthridine as a photosensitiser in macrocyclic europium and terbium complexes. *J. Chem. Soc., Perkin Trans. 2* 1998, **10**, 2129-2140.