

Supporting Information

Combining Flavin Photocatalysis with Parallel Synthesis: A General Platform to Optimize Peptides with Non-Proteinogenic Amino Acids

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General Information

Unless stated otherwise, all reactions were performed under an atmosphere of N₂ with magnetic stirring. Materials received from commercial suppliers were used directly without further purification. Boronic acids and potassium trifluoroborates were purchased from Sigma-Aldrich, Alfa Aesar, Matrix Scientific, Frontier Scientific, A.K. Scientific, Combi-Blocks, and Oakwood Chemicals. Methyl 2-Acetamidoacrylate was purchased from Combi-Blocks and Oakwood Chemicals. Fmoc-protected amino acids were purchased from Chem-Impex. The organic solvents (reagent grade) used for reaction optimization and purification were purchased from Sigma Aldrich and Fisher Scientific. Aqueous buffers were freshly prepared using Millipore Grade I water (Resistivity > 5 MΩ cm, Conductivity < 0.2 μS/cm, TOC <30 ppb) and the pH adjusted using a Mettler Toledo FiveEasy pH meter. Thin-layer chromatography (TLC) was performed on silica gel coated aluminum TLC plates (Merck, TLC Silica gel 60 F₂₅₄) and visualized using a UV lamp (254 nm) in combination with KMnO₄, Iodine, and Curcumin stains. The amino acid reactions were performed with Blue LED lights from Kessil (PR160L, 440 nm, 40 W). The peptide reactions were performed with the Lumidox® Generation II Controller and the 96-well LED array (445 nm, 295 mW per well) with a Lens Mat Active Cooling Base from Analytical Sales and Services Inc. The amino acid reactions were stirred on a Corning PC-620D stir plate at 1150 rpm. The plate reactions were stirred with a VP 710-C5 from V&P Scientific Inc. The products were purified by flash column chromatography on an automated Buchi Reveleris® X2 instrument or on a Isolera 1 Biotage® instrument. ¹H NMR, ¹³C NMR, and ¹⁹F NMR spectra were recorded on a Bruker Ascend™ 400 NMR spectrometer. ¹³C NMR spectra were also recorded on a Bruker Ultrashield™ Plus500 NMR spectrometer for some compounds. ¹H NMR and ¹³C NMR spectra were referenced to CDCl₃ (7.26 ppm and 77.16 ppm) or D₂O (4.90 ppm). Peak multiplicities are designated by the following abbreviations: s, singlet; d, doublet; t, triplet; p, pentet, q, quartet; hept, heptet m, multiplet; dd, doublet of doublet; dt, doublet of triplet; and ddd, doublet of doublet of doublet. NMR Spectra were recorded at 298 K. Electrospray ionization spectra were acquired on a LCT Premier (Waters Corp.) time of flight mass spectrometer (HRMS). UV spectrum for peptides were determined on a Waters Acquity UPLC H-Class, and LC-MS/MS data was acquired on a Quatro Ultima (Waters Corp.) triple quadrupole mass analyzer with an electrospray ion source. Extinction coefficients were determined on Varian Cary 100 Bio UV-Vis Spectrophotometer.

Reaction Optimization

The reaction was optimized using 2-methoxypyridine-4-boronic acid **1A** at physiological conditions (pH 6-8). Methyl 2-acetamidoacrylate was used as a surrogate for an internal dehydroalanine. Our optimization primarily focused on finding a suitable photocatalyst, organic cosolvent, buffer, and solution concentration. A summary of our optimization results is presented in Supplementary Tables 1-5 below.

Effect of photocatalyst identity on the amino acid reaction

Procedure: To a 1-dram vial charged with a stir bar, methyl 2-acetamidoacrylate (5 mg, 0.035 mmol, 1 equiv.), 2-methoxypyridine-4-boronic acid **1A** (6.42 mg, 0.042 mmol, 1.2 equiv.), photocatalyst (0.0017 mmol, 0.05 equiv.), 525 μ L of *N,N*-dimethylformamide (N_2 sparged for 5 minutes), 350 μ L of a 0.1 M solution of phosphate buffer pH 7 (N_2 sparged for 5 minutes), and 2.625 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 1150 rpm and fan cooling. After 15 hours, the sample was removed and extracted using ethyl acetate and washed with sodium bicarbonate (saturated aq.) and brine (saturated aq.) solutions. The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ¹H NMR yield was obtained by the addition of dibromomethane as an internal standard (1 equiv., 0.035 mmol, 4.93 ppm).

Table S1. Effect of photocatalyst identity on the reaction

Entry	Photocatalyst identity	% Yield ^a
1	Eosin Y	0%
2	Tris(2,2'-bipyridyl)dichlororuthenium(II) hexahydrate	0%
3	(Ir[dF(CF ₃)ppy] ₂ (dtbpy)) hexafluorophosphate	0%
4	9-Mesityl-10-methylacridinium tetrafluoroborate	<1%
5	Riboflavin	1%
6	Lumichrome	2%
7	Riboflavin 5'-monophosphate sodium salt	3%
8	Riboflavin tetrabutyrate	4%
9	Lumiflavin	8%

^a % Yield determined by ¹H NMR spectra using dibromomethane (1 equiv.) as an internal standard.

Effect of cosolvent on the amino acid reaction

Procedure: To a 1-dram vial charged with a stir bar, methyl 2-acetamidoacrylate (5 mg, 0.035 mmol, 1 equiv.), 2-methoxypyridine-4-boronic acid **1A** (6.42 mg, 0.042 mmol, 1.2 equiv.), lumiflavin (0.45 mg, 0.0017 mmol, 0.05 equiv.), 525 μ L of cosolvent (N_2 sparged for 5 minutes), 350 μ L of a 0.1 M solution of phosphate buffer pH 7 (N_2 sparged for 5 minutes), and 2.625 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 1150 rpm and fan cooling. After 15 hours, the sample was removed and extracted using ethyl acetate and washed with sodium bicarbonate (saturated aq.) and brine (saturated aq.) solutions. The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ¹H NMR yield was obtained by the addition of dibromomethane as an internal standard (1 equiv., 0.035 mmol, 4.93 ppm).

Table S2. Effect of cosolvent on the reaction

<p>S1 (1 equiv.) + 1A (1.2 equiv.) Lumiflavin (5 mol%) Phosphate Buffer pH 7 : Cosolvent (85:15) 10 mM solution, 15 hr</p>		
Entry	Cosolvent Identity	% Yield ^a
1	Cyclohexyl- <i>N</i> -methylpyrrolidine	<1%
2	Pyridine	2%
3	<i>N</i> -Methyl-2-Pyrrolidone	2%
4	Tetrahydrofuran	2%
5	Isopropanol	4%
6	<i>N,N</i> -Dimethylacetamide	5%
7	1,2-Dimethoxyethane	5%
8	Ethanol	8%
9	Diethylene glycol dimethyl ether	8%
10	Dimethylsulfoxide	9%
11	Acetonitrile	9%
12	Acetone	10%
13	Dioxane	10%
14	Sulfolane	10%
15	Propylene carbonate	10%
16	Methyl Ethyl Ketone	11%
17	Methanol	11%
18	<i>tert</i> -Butanol	12%
19	Methyl Acetate	12%
20	Glycerol	16%
21	Hexafluoro-2-propanol	17%
22	2,2,2-Trifluoroethanol	20%
23	Water (no cosolvent)	28%

^a % Yield determined by ¹H NMR spectra using dibromomethane (1 equiv.) as an internal standard.

Effect of buffer identity on the amino acid reaction

Procedure: To a 1-dram vial charged with a stir bar, methyl 2-acetamidoacrylate (5 mg, 0.035 mmol, 1 equiv.), 2-methoxypyridine-4-boronic acid **1A** (6.42 mg, 0.042 mmol, 1.2 equiv.), lumiflavin (0.45 mg, 0.0017 mmol, 0.05 equiv.), buffer solution (N_2 sparged for 5 minutes), and 3.15 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 1150 rpm and fan cooling. After 15 hours, the sample was removed and extracted using ethyl acetate and washed with sodium bicarbonate (saturated aq.) and brine (saturated aq.) solutions. The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ¹H NMR yield was obtained by the addition of dibromomethane as an internal standard (1 equiv., 0.035 mmol, 4.93 ppm).

Table S3. Effect of buffer identity on the reaction

Entry	Buffer Identity (pH)	Buffer concentration	% Yield ^a	Reaction Conditions	
				Lumiflavin (5 mol%)	Buffer (pH) 10 mM solution, 15 hr
1	Ammonium carbonate (8.4)	10 mM	8%		
2	Sodium bicarbonate (8.2)	10 mM	10%		
3	MES (6.5)	10 mM	11%		
4	HEPES (7)	10 mM	26%		
5	Phosphate (8)	10 mM	13%		
6	Phosphate (7)	10 mM	28%		
7	Phosphate (7)	25 mM	21%		
8	Phosphate (7)	50 mM	15%		
9	Phosphate (7)	100 mM	19%		

^a % Yield determined by ¹H NMR spectra using dibromomethane (1 equiv.) as an internal standard.

Effect of concentration, lumiflavin loading, and boronic acid equivalents on the amino acid reaction

Procedure: To a 1-dram vial charged with a stir bar, methyl 2-acetamidoacrylate (5 mg, 0.035 mmol, 1 equiv.), 2-methoxypyridine-4-boronic acid **1A**, lumiflavin, phosphate buffer pH 7 (N_2 sparged for 5 minutes), and ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 1150 rpm

and fan cooling. After 15 hours, the sample was removed and extracted using ethyl acetate and washed with sodium bicarbonate (saturated aq.) and brine (saturated aq.) solutions. The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ^1H NMR yield was obtained by the addition of dibromomethane as an internal standard (1 equiv., 0.035 mmol, 4.93 ppm).

Table S4. Effect of concentration, lumiflavin loading, and boronic acid equivalents on the reaction

		Lumiflavin (Y mol%)		
		Phosphate Buffer pH 7 Z mM solution, 15 hr		
Entry	Boronic acid	mol% Lumiflavin	Concentration of solution	% Yield ^a
1	1.2 equiv.	5	1 mM	2%
2	1.2 equiv.	5	5 mM	27%
3	1.2 equiv.	5	10 mM	29%
4	1.2 equiv.	5	15 mM	15%
5	3 equiv.	5	10 mM	37%
6	3 equiv.	10	10 mM	39%
7	3 equiv.	10	10 mM	b55% (c49%)

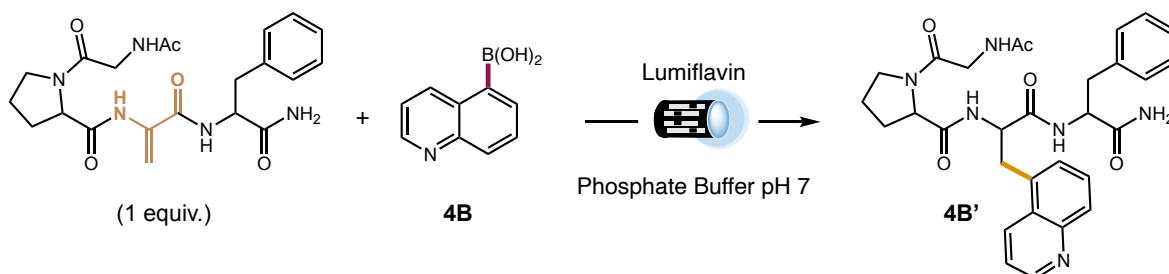
^a % Yield determined by ^1H NMR spectra using dibromomethane (1 equiv.) as an internal standard. ^b 50 mg scale of the acrylate. ^c Isolated yield.

Procedure for 50 mg scale: To a 100-mL round-bottom flask charged with a stir bar, methyl 2-acetamidoacrylate (50 mg, 0.35 mmol, 1 equiv.), 2-methoxypyridine-4-boronic acid **1A** (160.6 mg, 1.05 mmol, 3 equiv.), lumiflavin (8.97 mg, 0.035 mmol, 0.10 equiv.), 3.5 mL of a 0.1 M solution of phosphate buffer pH 7 (N_2 sparged for 5 minutes), and 31.5 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 550 rpm and fan cooling. After 15 hours, 10 equiv. of L-Cysteine and 20 mL of sodium bicarbonate (saturated aq.) were added to the stirring solution at room temperature. After being stirred for 2 hours, the sample was extracted using ethyl acetate and washed with brine (saturated aq.). The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ^1H NMR yield was obtained by the addition of nitromethane as an internal standard (1 equiv., 0.35 mmol, 4.33 ppm). The product was then purified by reverse phase flash chromatography using acetonitrile:water to yield 43.4 mg (49.2% isolated yield) of pure product.

Reaction optimization for the peptide

Procedure: To a 1 mL clear glass shell vial (purchased from Analytical Sales & Services Inc.) charged with a small stir bar, Ac-Gly-Pro-Dha-Phe-NH₂(1 mg, 2.33 μmol, 1 equiv.), quinoline-5-boronic acid **4B**, lumiflavin, phosphate buffer pH 7 (N₂ sparged for 5 minutes), and ddH₂O (N₂ sparged for 5 minutes) were added. The vial was purged under N₂ for 3 minutes, parafilmed, and irradiated with blue light with stirring at 1150 rpm on the Corning PC-620D stir plate or at ~700 rpm on the VP 710-C5 for the specified amount of time. After that time, the sample was filtered and subjected to UPLC/MS analysis.

Table S5. Optimization of the peptide reaction



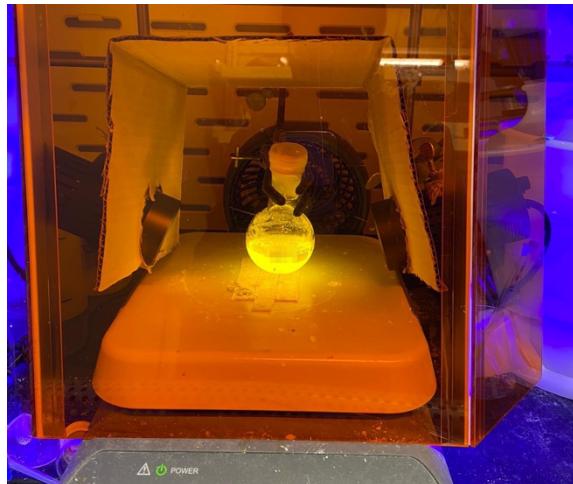
Entry	Lumiflavin	Light source	Boronic acid	Conc.	Time	% Conversion. ^a
1	5 mol%	Kessil	1 equiv.	10 mM	15 hr	19%
2	5 mol%	Kessil	3 equiv.	10 mM	15 hr	33%
3	5 mol%	Kessil	5 equiv.	10 mM	15 hr	37%
4	5 mol%	Kessil	10 equiv.	10 mM	15 hr	40%
5	5 mol%	Kessil	5 equiv.	5 mM	15 hr	36%
6	5 mol%	Kessil	5 equiv.	3 mM	15 hr	60%
7	5 mol%	Lumidox® Gen. II	5 equiv.	10 mM	15 hr	49%
8	5 mol%	Lumidox® Gen. II	5 equiv.	3 mM	8 hr	56%
9	10 mol%	Lumidox® Gen. II	5 equiv.	3 mM	8 hr	70%
10	10 mol%	Lumidox® Gen. II	5 equiv.	3 mM	6 hr	75%

^a% Product determined by integrating the product peaks on LC as compared to all other peptide-related peaks.

Experimental Procedures

Picture of reaction setups

(A)



(B)



Figure S1. Reaction setup photographs. (A) Amino acid reactions. (B) 96-well plate reactions.

General procedure for amino acid synthesis

To a 100-mL round-bottom flask charged with a stir bar, methyl 2-acetamidoacrylate (50 mg, 0.35 mmol, 1 equiv.), boronic acid (1.05 mmol, 3 equiv.), lumiflavin (8.97 mg, 0.035 mmol, 0.10 equiv.), 3.5 mL of a 0.1 M solution of phosphate buffer pH 7 (N_2 sparged for 5 minutes), and 31.5 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 550 rpm and fan cooling. After 15 hours, 10 equiv. of L-Cysteine and 20 mL of sodium bicarbonate (saturated aq.) were added to the stirring solution at room temperature. After being stirred for 2 hours, the sample was extracted using ethyl acetate and washed with brine (saturated aq.). The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. A crude ¹H NMR yield was obtained by the addition of nitromethane as an internal standard (1 equiv., 0.35 mmol, 4.33 ppm). The product was then purified by reverse phase flash chromatography using acetonitrile:water.

General procedure for setting up the 96-well reactions

To 96 individual 1 mL clear glass shell vials (purchased from Analytical Sales & Services Inc.) each charged with a small stir bar, 96 different boronic acids or potassium trifluoroborates (11.65 μ mol, 5 equiv.) were added. Stock solutions of peptide (100 mg of Ac-G-P-Dha-F-NH₂ in 50 mL of ddH₂O), phosphate buffer pH 7 (10 mL of a 0.1 M solution), and lumiflavin (9 mg in 30 mL of ddH₂O) were purged under N_2 for 5 minutes each. The 96 vials containing boronic acids and potassium trifluoroborates, the stock solutions, 100-1000 μ L and 20-200 μ L channel micropipettes (can use multichannel), and a Philips head screwdriver were placed in a N_2 glove bag (purchased from Cole-Palmer). The glove bag was filled once with N_2 , emptied, refilled, and sealed. To each of the 96 vials was added 500 μ L of the peptide solution, 77 μ L of the buffered solution, and 200 μ L of the lumiflavin solution. The screwdriver was used to screw down the cap onto the vials. The

vials were then removed from the glove bag and placed on the Lumidox® 96-well LED array (445 nm) and illuminated with stirring at ~700 rpm for 6 hours.

General procedure for purifying the 96-well reactions

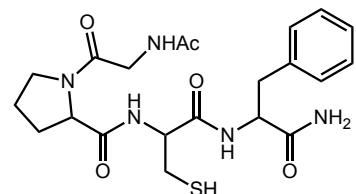
Upon completion of the reaction, the cap was removed, and the stirring was stopped. From a 0.24 mM stock solution of TCEP in ddH₂O, 100 µL were added to each vial. 10 equiv. of 2-mercaptoethylamine, polymer-bound resin was also added to each vial and the mixture stirred at ~700 rpm overnight at room temperature to remove any unreacted Dha-containing peptide. The vials were then filtered using a 96-well filter plate. Utilizing Waters Oasis HLB 96-well Plate (60 mg sorbent per well), the sorbent was first washed and filtered with 1 mL of methanol and 1 mL of water. The sample solutions were then loaded onto the sorbent and filtered. The boronic acid and potassium trifluoroborates were removed by filtering 1 mL of a 5% NaOH solution through each well. The lumiflavin was removed by carefully filtering (small amount of vacuum) 800 µL of a 25% isopropanol solution in water. Finally, the products were eluted with 1 mL of a 50% trifluoroethanol solution in water. The products were then submitted to a UPLC for analysis of purity and yield.

In instances where exogenous boronic acid remained in the eluents, diethyl ether extraction was performed. The samples of interest were first lyophilized to powder. Then, 500 µL of diethyl ether was added to each sample followed by 500 µL of ddH₂O. The aqueous solution was extracted in a glass pipette. The products were then diluted with another 500 µL of ddH₂O and submitted to a UPLC for analysis of purity and yield.

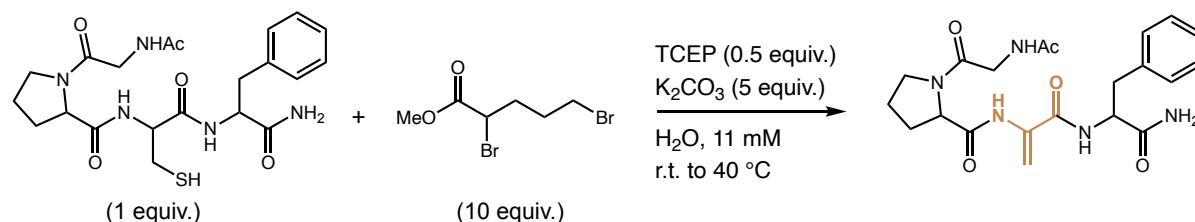
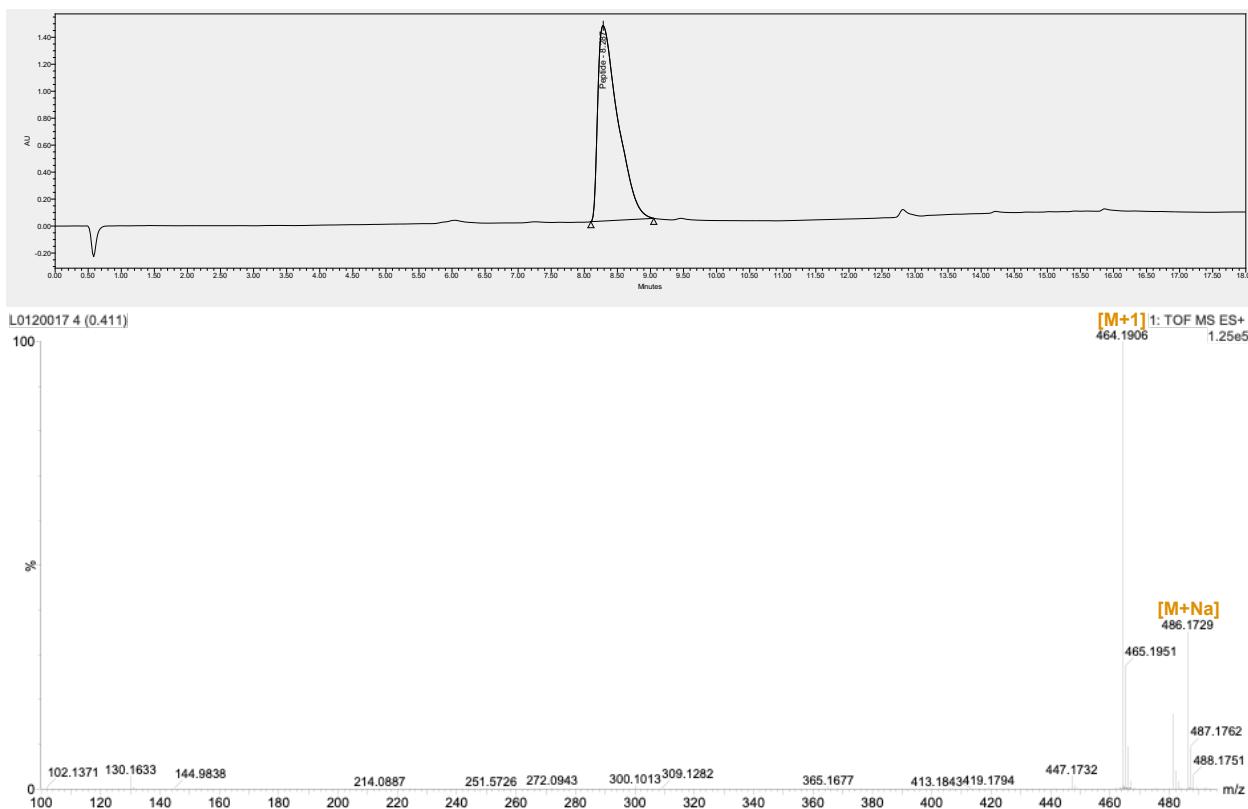
General procedure for addition into the selectivity peptide

To a 1 mL clear glass shell vial (purchased from Analytical Sales & Services Inc.) charged with a small stir bar, H₂N-Gly-Dha-His-Trp-Ser-Tyr-Gly-Met-Arg-Pro-Lys-CO₂H (1 mg, 0.77 µmol, 1 equiv.), 2-methyl-8-methoxyquinoline-5-boronic acid **6B** (0.5 mg, 2.31 µmol 3 equiv.), lumiflavin (0.020 mg, 0.077 µmol, 0.1 equiv.), 25.6 µL 0.1 M phosphate buffer pH 7 (N₂ sparged for 5 minutes), and 230.4 µL ddH₂O (N₂ sparged for 5 minutes) were added. The vial was purged under N₂ for 3 minutes, parafilmed, and irradiated with blue light with stirring at ~700 rpm on the VP 710-C5 for 6 hours. After 6 hours, the sample was filtered and subjected to UPLC/MS analysis.

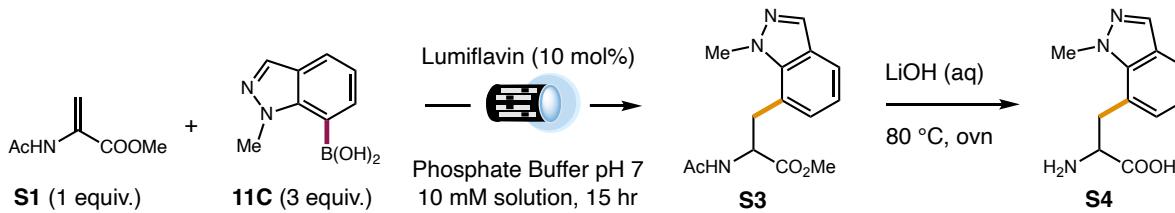
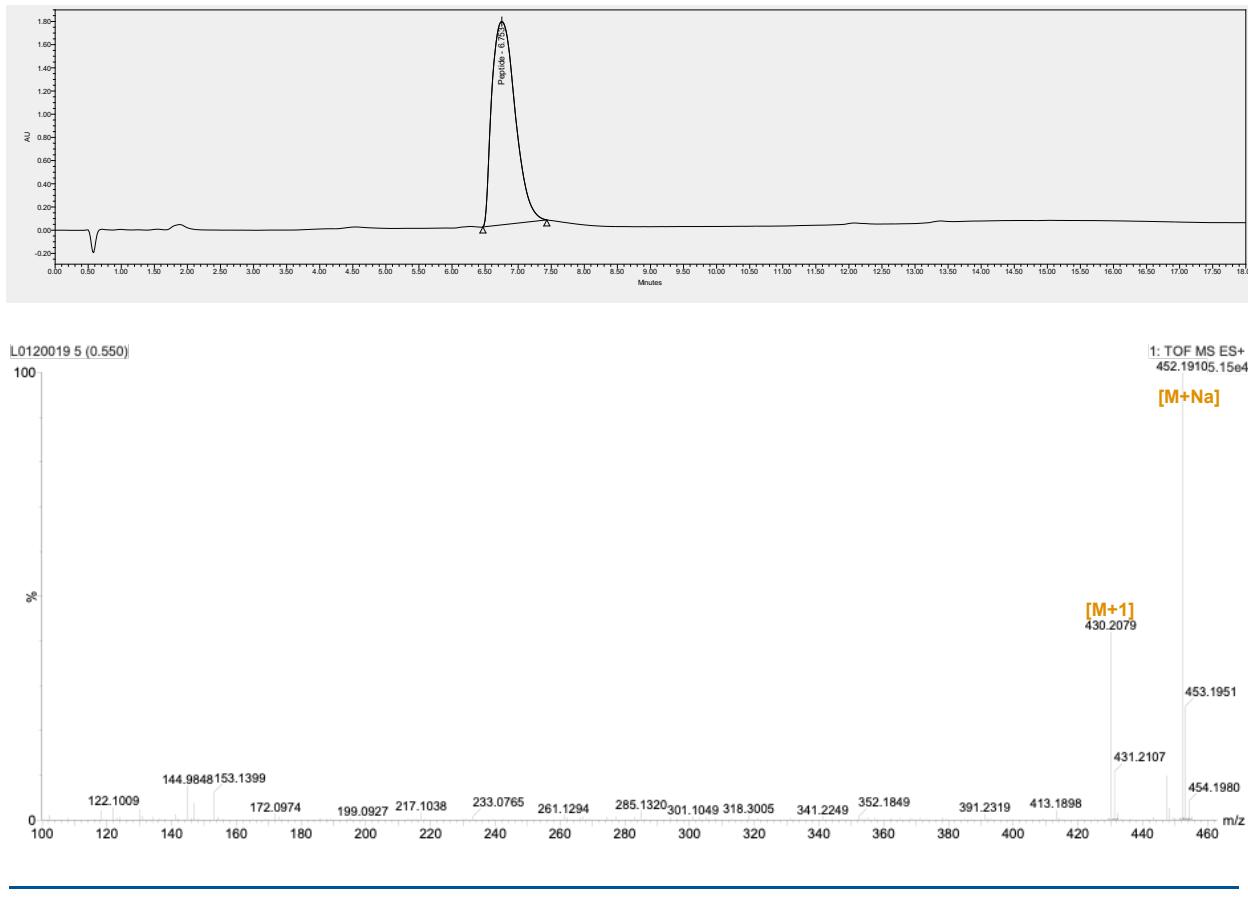
Synthesis of starting materials and other compounds



Ac-Gly-Pro-Cys-Phe-NH₂ was synthesized by solid phase peptide synthesis using Fmoc-protected amino acids on rink amid resin, cleaved with 95:2.5:2.5 TFA:TIPS:H₂O solution, and purified by reverse phase flash chromatography.

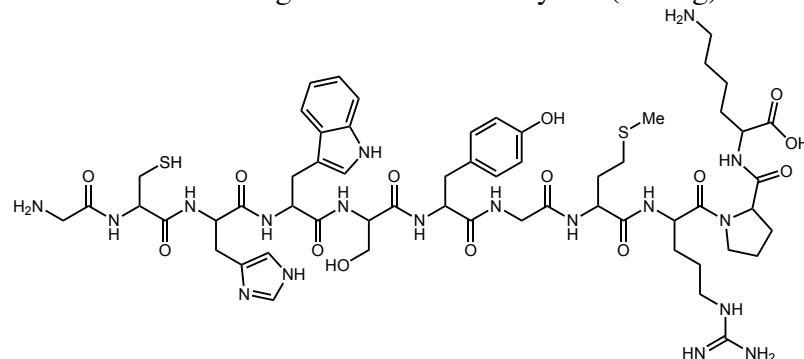


Ac-Gly-Pro-Dha-Phe-NH₂: To a 50 mL round-bottom flask charged with a stir bar, Ac-Gly-Pro-Cys-Phe-NH₂ (100 mg, 0.22 mmol, 1 equiv.), TCEP (31.5 mg, 0.11 mmol, 0.5 equiv.), and 20 mL of ddH₂O were added. The solution was stirred for 1 hour at room temperature; after which, methyl 2,5-dibromovalerate (591.0 mg, 2.2 mmol, 10 equiv.) and potassium carbonate (149 mg, 1.08 mmol, 5 equiv.) were added. The heterogenous mixture was stirred for another hour at room temperature before being heating to 40 °C for 3 hour. The reaction was monitored by UPLC/MS. Upon completion, the mixture was washed with 20 mL of ethyl acetate to remove excess methyl 2,5-dibromovalerate before being concentrated under reduced pressure. The residual solid was then redissolved in minimal methanol, filtered to remove impure solid, and purified by normal phase flash chromatography (0 – 10% methanol in methylene chloride, product elutes at 10% methanol in methylene chloride). Isolated yield = 51.2% (52.2 mg).

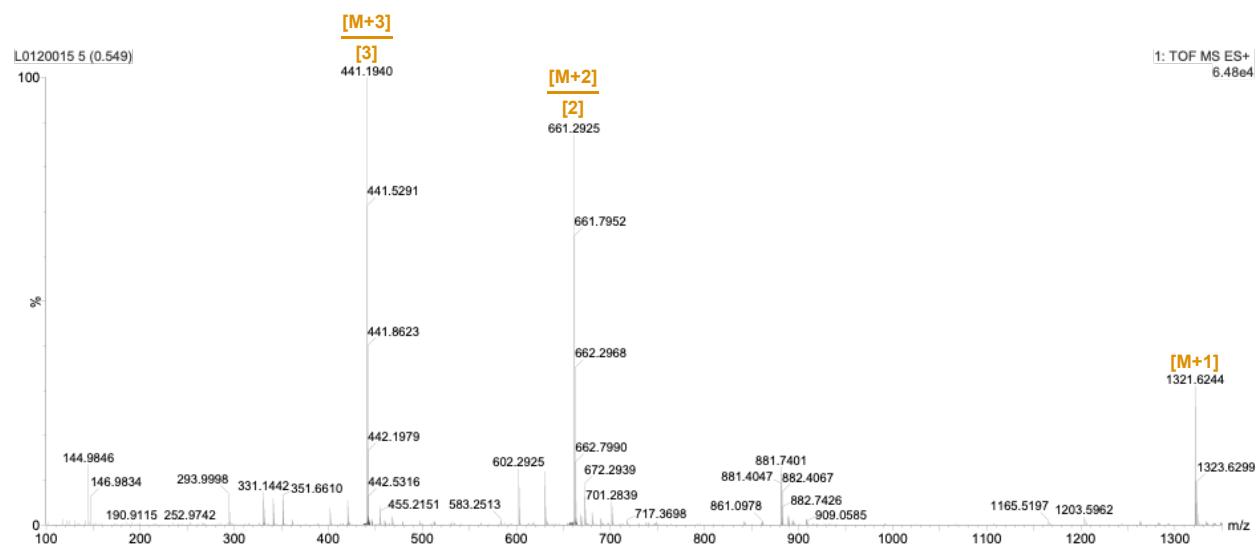
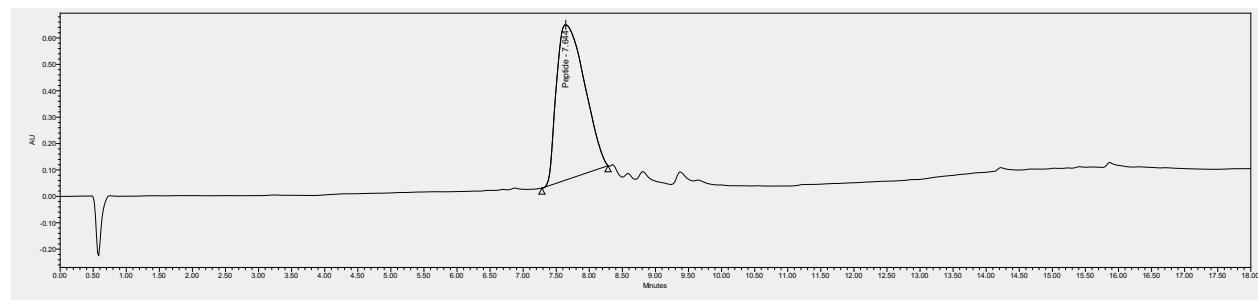


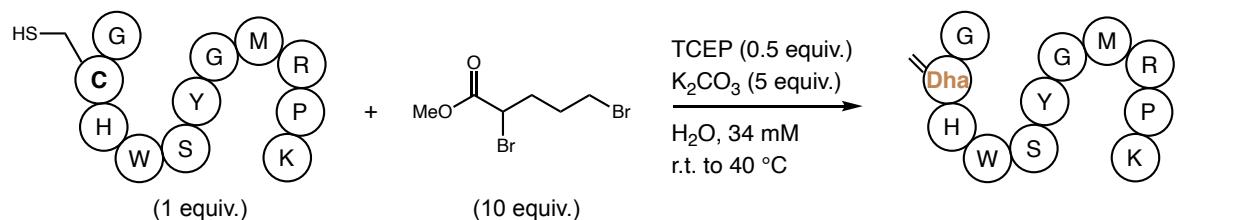
Methyl 2-acetamido-3-(1-methyl-1*H*-indazol-7-yl)propanoate (S3**):** To a 100-mL round-bottom flask charged with a stir bar, methyl 2-acetamidoacrylate (50 mg, 0.35 mmol, 1 equiv.), 1-methyl-1*H*-indazole-7-boronic acid **11C** (184.8 mg, 1.05 mmol, 3 equiv.), lumiflavin (8.97 mg, 0.035 mmol, 0.10 equiv.), 3.5 mL of a 0.1 M solution of phosphate buffer pH 7 (N_2 sparged for 5 minutes), and 31.5 mL ddH₂O (N_2 sparged for 5 minutes) were added. Then, the resulting solution was degassed by sparging with nitrogen for 5 min, parafilmed, and placed 5 cm away from two 40 W blue LED lamps. The sample was irradiated for 15 hours with stirring at 550 rpm and fan cooling. After 15 hours, 10 equiv. of L-cysteine and 20 mL of sodium bicarbonate (saturated aq.) were added to the stirring solution at room temperature. After being stirred for 2 hours, the sample was extracted using ethyl acetate and washed with brine (saturated aq.). The organic layers were combined and concentrated by rotary evaporation followed by high vacuum. The product was then purified by reverse phase flash chromatography using acetonitrile:water to give 29.6% isolated yield (28.5 mg).

2-amino-3-(1-methyl-1*H*-indazol-7-yl)propanoic acid (S4): To a 2-dram vial charged with a stir bar, methyl 2-acetamido-3-(1-methyl-1*H*-indazol-7-yl)propanoate (25 mg, 0.091 mmol, 1 equiv.), LiOH (217.5 mg, 9.1 mmol, 100 equiv.), and 5 mL of ddH₂O were added. The reaction was stirred at 80 °C overnight and monitored by TLC. Upon consumption of the starting material, the solution was concentrated, redissolved in minimal ddH₂O, and purified by reverse phase HPLC using acetonitrile:water to give 24.6% isolated yield (4.9 mg).

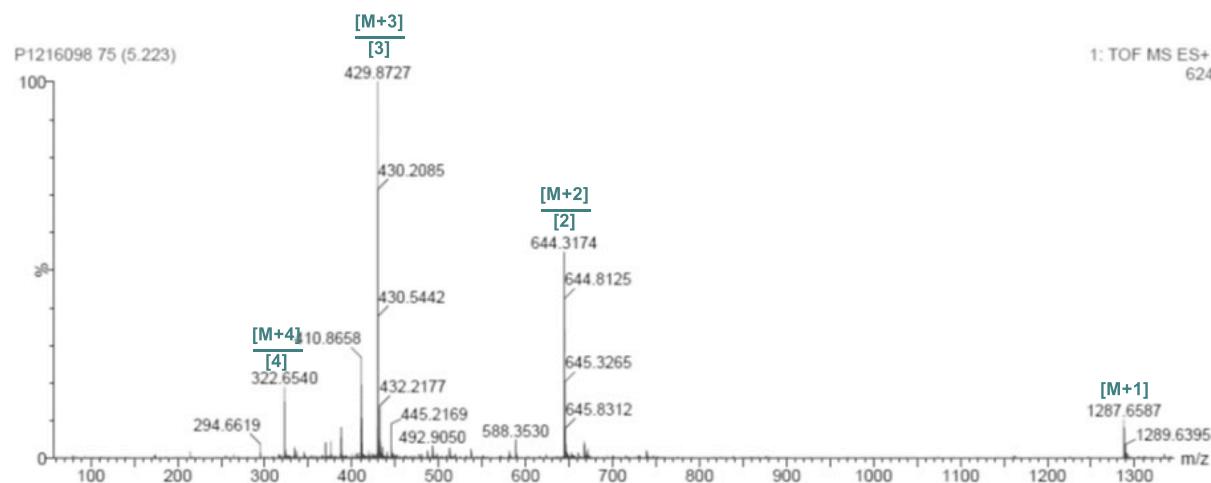
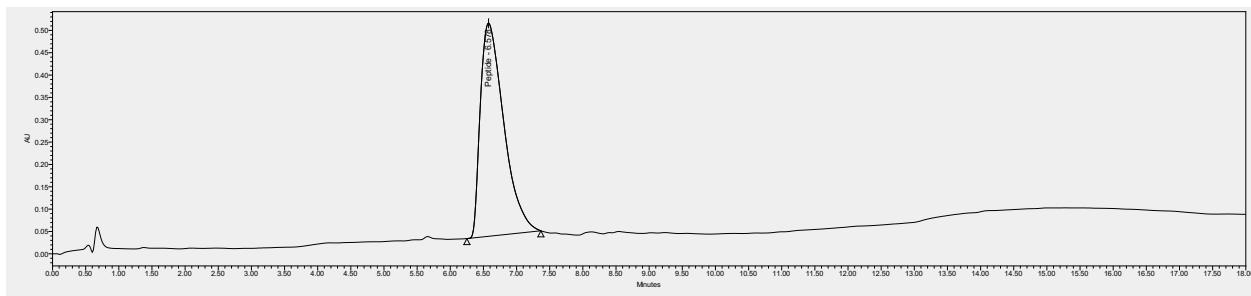


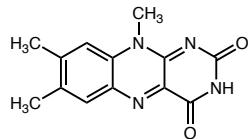
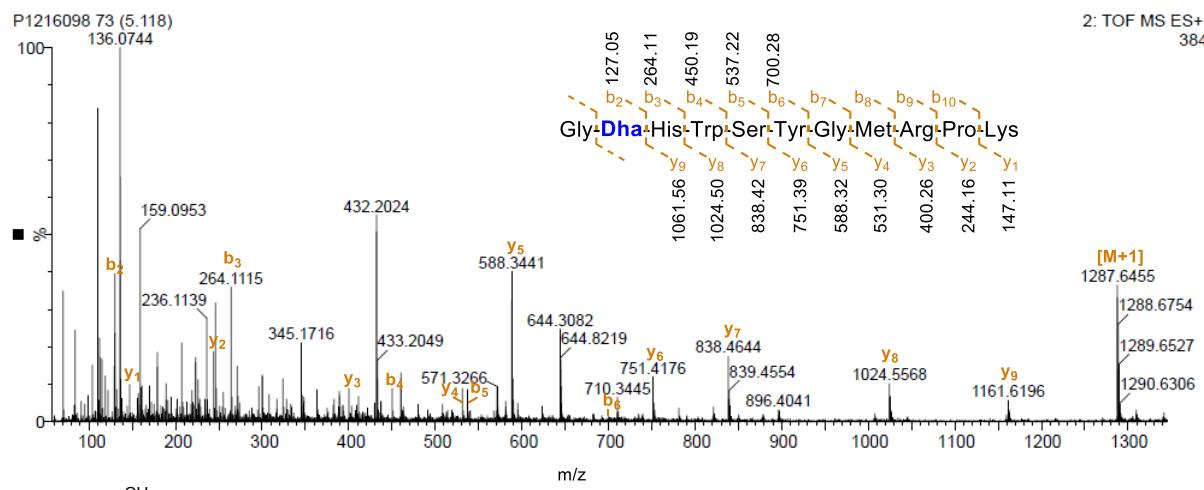
H₂N-Gly-Cys-His-Trp-Ser-Tyr-Gly-Met-Arg-Pro-Lys-CO₂H was synthesized by solid phase peptide synthesis using Fmoc-protected amino acids on 2-chlorotriptyl chloride resin, cleaved with 95:2.5:2.5 TFA:TIPS:H₂O solution, and purified by reverse phase flash chromatography.



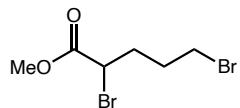


H₂N-Gly-Dha-His-Trp-Ser-Tyr-Gly-Met-Arg-Pro-Lys-CO₂H: To a 2-dram vial charged with a stir bar, H₂N-Gly-Cys-His-Trp-Ser-Tyr-Gly-Met-Arg-Pro-Lys-CO₂H (91 mg, 0.068 mmol, 1 equiv.), TCEP (9.74 mg, 0.034 mmol, 0.5 equiv.) and 2 mL of ddH₂O was added. The solution was stirred for 1 hour at room temperature; after which, methyl 2,5-dibromo-3-methoxyvalerate (184.28 mg, 0.68 mmol, 10 equiv.), and potassium carbonate (46.9 mg, 0.34 mmol, 5 equiv.) were added. The heterogenous mixture was stirred for another hour at room temperature before being heating to 40 °C for 3 hours. The reaction was monitored by UPLC/MS. Upon completion, the mixture was concentrated under reduced pressure. The residual solid was then redissolved in minimal water and purified by reverse phase flash chromatography using acetonitrile:water mix. Isolated yield = 22.7% yield (21 mg).





Lumiflavin was synthesized according to the reported procedure.¹ ¹**H NMR** (400 MHz, DMSO-*d*₆) δ 11.31 (s, 1H), 7.90 (s, 1H), 7.78 (s, 1H), 3.96 (s, 3H), 2.50 (s, 3H), 2.40 (s, 3H). ¹³**C NMR** (126 MHz, DMSO) δ 160.01, 155.59, 150.57, 146.59, 137.11, 135.87, 133.56, 131.63, 130.85, 116.45, 31.92, 20.66, 18.85.



Methyl 2,5-dibromo-3-valerate was synthesized according to the reported procedure.² ¹**H NMR** (400 MHz, Chloroform-*d*) δ 4.26 (dd, *J* = 8.2, 6.1 Hz, 1H), 3.79 (s, 3H), 3.43 (t, *J* = 6.4 Hz, 2H), 2.43 – 1.79 (m, 4H).

Extinction Coefficients

All extinction coefficients were taken of the completely unprotected amino acids. Extinction coefficients were calculated by measuring the absorbance at 214 nm with a UV-Vis spectrophotometer at 4-5 different concentrations of each amino acid. The concentrations were then plotted with the measured absorbances as a line using Beer-Lambert law ($A = \epsilon c$) where ϵ is equal to the slope of the line. All R^2 values of the lines measured were greater than 0.95 except for glycine (0.935). We used our own values measured for all amino acids but used the extinction coefficient for the peptide bond that had been reported.³

Measured extinction coefficients of proteinogenic amino acids

Table S6. Measured versus reported³ extinction coefficients for proteinogenic AAs

Measured:	ϵ	Reported:	ϵ
L-Glycine =	20.7	L-Glycine =	21
L-Lysine =	32.5	L-Lysine =	41
L-Cysteine =	223.0	L-Cysteine =	225
L-Phenylalanine =	4,406.4	L-Phenylalanine =	5,200
L-Methionine =	891.2	L-Methionine =	980
L-Tryptophan =	28,582	L-Tryptophan =	29,050
L-Histidine =	4,363.5	L-Histidine =	5,125
L-Tyrosine =	4,640.8	L-Tyrosine =	5,375
L-Proline =	652.0	Peptide bond =	923

The contributions from the acetyl group on the n-terminus plus the c-terminal amide of our peptide were determined by subtracting the extinction coefficient of the peptide Ac-G-P-C-F-NH₂ from the extinction coefficient of the peptide G-P-C-F. The extinction coefficient for proline has been shown to vary depending on its environment.^[3] Therefore, we determined the extinction coefficient for the proline in our specific peptide, Ac-G-P-C-F-NH₂, by subtracting the extinction coefficient of the whole peptide minus all the components except proline.

$$\text{Ac-G-P-C-F-NH}_2 = 9,121.4 - [20.7 \text{ (Gly)} + 223.0 \text{ (Cys)} + 4,406.4 \text{ (Phe)} + 2,769 \text{ (3 Peptide Bonds)} + 1,050.3 \text{ (Acetyl + C-terminal Amide)} = \underline{652.0} = \text{Proline.}$$

Measured extinction coefficients of non-proteinogenic amino acids

Table S7. Measured extinction coefficients for non-proteinogenic AAs

NPAAs	ϵ	NPAAs	ϵ
L-2-Naphthylalanine =	20,986	L-2-Thienylalanine =	2,809.6
H-p-Phenyl-L-Phenylalanine =	15,956	L-3-Pyridylalanine =	1,060.6
3-Benzothienyl-L-alanine =	14,625	D-3-Cyclohexylalanine =	61.3
(S)-2-Amino-3-quinolin-2-yl-propionic acid =	11,676	Acetyl + C-terminal Amide =	1,050.3
1-Methyl-1H-indazol-7-alanine =	15,311		

Measured extinction coefficients of substituted phenylalanine derivatives

Due to the inaccessibility of all 96 amino acids from our plate, we measured the extinction coefficients of four derivatives of phenylalanine and determined the average to be similar to the value for phenylalanine itself. As a result, we made the assumption that the substitutions would be approximately the same as the core amino acid itself.

AA:	L-Phenylalanine	Average = 5,071.9	4-Me-L-Phenylalanine	4-MeO-L-Phenylalanine	4-F-L-Phenylalanine	4-t-Bu-L-Phenylalanine
	<chem>C[C@@H](C(=O)N)Cc1ccccc1</chem>		<chem>C[C@@H](C(=O)N)Cc1ccc(C)c1</chem>	<chem>C[C@@H](C(=O)N)Cc1ccc([O]c)c1</chem>	<chem>C[C@@H](C(=O)N)Cc1ccc(F)c1</chem>	<chem>C[C@@H](C(=O)N)Cc1ccc(C(C)(C)C)c1</chem>
ϵ	4,406.4		6,845	2,830	2,334.2	8,278.3

Figure S2. Extinction coefficients of substituted phenylalanine derivatives

Assumptions to determine yields

Table S8. Assumptions made to determine the yields of all 96 peptides by LC analysis

Molecule	Assumption	Molecule	Assumption
Ac-GPXF-NH ₂ =	3 Peptide bonds	Aromatics =	L-Phenylalanine
Pyridines =	L-3-Pyridylalanine	Naphthyls =	L-2-Naphthylalanine
Quinolines/Isoquinolines =	(S)-2-Amino-3-quinolin-2-yl-propionic acid	Benzyl + Phenylalanine =	L-Phenylalanine + L-Phenylalanine
Pyrimidines =	L-3-Pyridylalanine	Biphenyl =	H-p-Phenyl-L-Phenylalanine
Indoles =	L-Tryptophan	Pyridine + Phenylalanine =	L-3-Pyridylalanine + L-Phenylalanine
Indazole/Azaindazoles =	1-Methyl-1H-indazol-7-alanine	Pyrazole + Phenylalanine =	L-Histidine + L-Phenylalanine
Benzothiophenes =	3-Benzothienyl-L-alanine	Aliphatics =	Average of natural alkyls (37.8)
Pyrazoles =	L-Histidine	Aliphatic + Aryl group =	Average alkyl + L-Phenylalanine
Thiophene =	L-2-Thienylalanine	Methionine =	L-Methionine

Example calculation to determine yield

Note: The flow rate used for the LC analysis was 0.5 mL/min, and the path length of the Waters Acquity UPLC H-Class was 1 cm. The UPLC reported all absorbance measurements as microAU×sec. All injections volumes were 10 μ L, and the reactions were run on a 0.00233 mmol scale.

Example calculation to determine the amount and yield of product 1A':

Beer-Lambert's Law Modified for HPLC⁴: $A \times F = 10^3 \times \epsilon \times b \times N_0$

Where: A = absorbance (AU×min), F = flow rate (mL/min), ε = extinction coefficient (AU×L/mol×cm), b = path length (cm), N₀ = mol injected (mol)

Extinction Coefficient for **1A'** = [20.7 (Gly) + 652.0 (Pro) + 1060.6 (Pyr) + 4,406.4 (Phe) + [923 × 3] (3 Peptide bonds) + 1050.3 (Acetyl + C-terminal Amide)] = 9,959 AU×L/mol×cm

Absorbance for **1A'** = 708,389 + 1,432,789 = 2,141,178 microAU×sec

$$N_0 = A \times F / 10^3 \times \epsilon \times b$$

$$N_0 = [(2,141,178 \mu\text{AU} \times \text{sec}) \times (1 \text{ min}/60 \text{ sec}) \times (1 \text{ AU}/10^6 \mu\text{AU})] \times (0.5 \text{ mL}/\text{min}) / (10^3 \text{ mL/L}) \times (9,959 \text{ AU} \times \text{L/mol} \times \text{cm}) \times (1 \text{ cm}) = \underline{1.79 \times 10^{-9} \text{ mol}} \text{ (injected mol)}$$

Concentration of product (M) = injected mol / injected L

$$M = 1.79 \times 10^{-9} \text{ mol} / 0.00001 \text{ L} = \underline{1.79 \times 10^{-4} \text{ M}}$$

Amount of product (mg) = concentration of product (M) × volume of sample (M) × MW of product

$$\text{Amount} = (1.79 \times 10^{-4} \text{ M}) \times (0.001 \text{ L}) \times (538.6 \text{ g/mol}) \times (1000 \text{ mg}/1 \text{ g}) = \underline{0.096 \text{ mg of product } \mathbf{1A}'}$$

Yield = (amount of pdt (mg) / theoretical amount of pdt (mg)) × 100%

$$\text{Yield} = (0.096 \text{ mg}) / [(0.00233 \text{ mmol}) \times (538.6 \text{ mg/mmole})] = 0.0768 \times 100\% = \underline{7.7\% \text{ yield of product } \mathbf{1A}'}$$

Thrombin Inhibition Studies

Procedure: The ability of peptides to inhibit Thrombin was tested using the Sigma Thrombin Inhibitor Screening Kit (MAK243). The test peptides were solvated in water and transferred to the assay plates using ECHO550 (Beckman Inc.). The assay was miniaturized further in 384 well plate format. First, Thrombin (12.5 µL) was preincubated with 80 µM of peptides (0.5 µL) at room temperature for 15 minutes, followed by the addition of the synthetic AMC-based peptide substrate (12 µL). The released fluorophore AMC from proteolytic cleavage of the substrate by Thrombin at 37 °C was then quantified by a BioTek Neo fluorescence reader at Ex/Em = 350/450 nm. PPACK dichloride was used as a positive control, and buffer (no thrombin) was used as a negative control. Percent inhibition at 30 minutes of incubation with the 72 screening grade peptides was calculated as an average of two independent experiments. For the Ac-Gly-Pro-Phe-Phe-NH₂ inhibitor and the resynthesized (L)- and (D)-diastereomer of Ac-Gly-Pro-[N(Me)-2-Ind]-Phe-NH₂ **3C'**, percent inhibition at 30 minutes incubation was calculated as an average of three independent experiments.

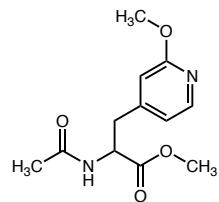
Table S9. Thrombin inhibition data

Peptide:	1st run:	2nd run:	Average:	Standard Deviation:
PPACK	100	-	-	-
Buffer	0	-	-	-
1A'	92.3	92.0	92.1	0.1
2A'	8.5	8.8	8.7	0.1
5A'	-10.7	-6.6	-8.6	1.4
6A'	-6.0	0.8	-2.6	2.4
7A'	-11.3	-2.6	-6.9	3.1
8A'	22.4	23.5	22.9	0.4
10A'	18.6	20.1	19.4	0.5
11A'	16.7	19.3	18.0	0.9
12A'	5.2	9.5	7.4	1.5
1B'	-16.8	-13.7	-15.3	1.1
3B'	70.1	69.4	69.8	0.2
4B'	-2.1	-2.9	-2.5	0.3
5B'	12.1	14.5	13.3	0.8
6B'	14.0	12.9	13.4	0.4
7B'	-14.6	-12.1	-13.3	0.9
8B'	-11.3	-5.9	-8.6	1.9
9B'	6.0	8.5	7.2	0.9
10B'	4.3	5.5	4.9	0.4
1C'	23.0	22.6	22.8	0.1
3C'	96.7	97.0	96.9	0.1
4C'	82.5	81.5	82.0	0.4
5C'	29.7	29.8	29.7	0.0
6C'	79.4	79.0	79.2	0.1
7C'	-14.2	-10.6	-12.4	1.3
8C'	-18.3	-15.8	-17.1	0.9
9C'	-10.4	-7.0	-8.7	1.2
10C'	12.7	12.6	12.7	0.0
11C'	5.0	8.9	7.0	1.4
12C'	-2.5	2.7	0.1	1.8
2D'	79.1	77.7	78.4	0.5
3D'	77.8	76.9	77.3	0.3
5D'	51.5	49.8	50.7	0.6

6D'	-5.9	-3.1	-4.5	1.0
7D'	2.4	6.1	4.2	1.3
11D'	-9.4	-5.2	-7.3	1.5
1E'	80.2	79.9	80.1	0.1
3E'	48.5	49.1	48.8	0.2
5E'	-1.3	2.7	0.7	1.4
6E'	-13.7	-9.9	-11.8	1.3
7E'	-6.8	-0.5	-3.6	2.2
8E'	-21.5	-16.9	-19.2	1.6
9E'	2.6	8.1	5.4	2.0
10E'	3.5	6.0	4.7	0.9
11E'	8.9	15.7	12.3	2.4
12E'	71.5	70.6	71.0	0.3
1F'	40.3	39.3	39.8	0.4
3F'	23.1	28.3	25.7	1.9
7F'	-4.6	-0.7	-2.7	1.4
8F'	-29.3	-24.7	-27.0	1.6
9F'	37.8	36.4	37.1	0.5
10F'	0.1	6.5	3.3	2.3
11F'	2.3	5.9	4.1	1.3
12F'	11.6	14.6	13.1	1.1
1G'	11.0	13.4	12.2	0.8
2G'	87.1	86.7	86.9	0.1
3G'	67.4	67.6	67.5	0.0
4G'	34.1	33.6	33.9	0.2
5G'	-5.5	0.2	-2.6	2.0
6G'	-11.0	-6.7	-8.8	1.5
7G'	21.1	21.1	21.1	0.0
8G'	12.8	16.8	14.8	1.4
9G'	-15.8	-11.6	-13.7	1.5
10G'	-3.9	-1.8	-2.9	0.8
11G'	-16.4	-14.6	-15.5	0.7
12G'	18.3	18.0	18.2	0.1
1H'	-3.1	0.9	-1.1	1.4
4H'	1.4	3.4	2.4	0.7
6H'	0.3	5.1	2.7	1.7
8H'	10	10.4	10.2	0.1
10H'	2.5	4.1	3.3	0.6
11H'	18.6	17.0	17.8	0.6
12H'	15.4	15.2	15.3	0.1
Peptide:	1st run:	2nd run:	3rd run:	Average:
L-N-methyl-2-indole 3C'	64.0	68.7	67.7	66.8
D-N-methyl-2-indole 3C'	27.0	29.2	33.5	29.9
Ac-G-P-F-F-NH ₂	9.4	5.5	6.5	7.1

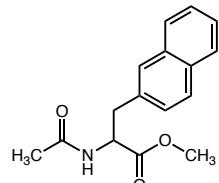
Analytical Data of Products

Amino Acids



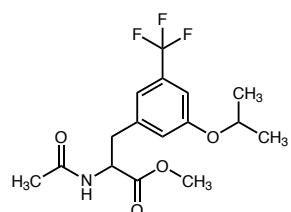
Methyl 2-acetamido-3-(2-methoxypyridin-4-yl)propanoate

Isolated Yield 49.0% (43 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.12 (d, *J* = 5.0 Hz, 1H), 6.68 (d, *J* = 5.1 Hz, 1H), 6.55 (s, 1H), 6.04 (d, *J* = 7.8 Hz, 1H), 4.91 (q, *J* = 6.1 Hz, 1H), 3.95 (s, 3H), 3.76 (s, 3H), 3.40 – 2.77 (m, 2H), 2.02 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 171.57, 170.12, 164.26, 149.07, 146.25, 118.07, 111.38, 54.10, 52.76, 52.49, 37.25, 23.18. **HRMS (ESI-ToF)**: m/z calculated for C₁₂H₁₇N₂O₄ [MH]⁺: 253.1188. Found 253.1183.



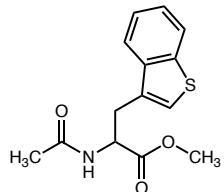
Methyl 2-acetamido-3-(naphthalen-2-yl)propanoate

Isolated Yield 52.7% (50 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.89 – 7.67 (m, 3H), 7.64 – 7.52 (m, 1H), 7.53 – 7.33 (m, 2H), 7.23 (dd, *J* = 8.4, 1.8 Hz, 1H), 6.10 (d, *J* = 7.3 Hz, 1H), 4.97 (dt, *J* = 7.8, 5.9 Hz, 1H), 3.72 (s, 3H), 3.27 (qd, *J* = 13.9, 5.9 Hz, 2H), 1.96 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 172.26, 169.86, 133.49, 132.55, 128.35, 128.09, 127.75, 127.75, 127.63, 127.30, 126.30, 125.88, 53.27, 52.44, 38.06, 23.16. **HRMS (ESI-ToF)**: m/z calculated for C₁₆H₁₈NO₃ [MH]⁺: 272.1287. Found 272.1274.



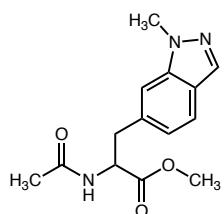
Methyl 2-acetamido-3-(3-isopropoxy-5-(trifluoromethyl)phenyl)propanoate

Isolated Yield 43.6% (53 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 6.99 (t, *J* = 2.0 Hz, 1H), 6.87 (dq, *J* = 1.6, 0.8 Hz, 1H), 6.78 (t, *J* = 1.9 Hz, 1H), 5.99 (d, *J* = 7.6 Hz, 1H), 4.88 (dt, *J* = 7.6, 5.7 Hz, 1H), 4.54 (hept, *J* = 6.1 Hz, 1H), 3.74 (s, 3H), 3.14 (qd, *J* = 13.8, 5.6 Hz, 2H), 2.01 (s, 3H), 1.34 (s, 3H), 1.33 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 171.86, 169.81, 158.36, 138.52, 132.19, 131.88, 120.11, 118.08 (d, *J* = 3.8 Hz), 111.44 (d, *J* = 4.0 Hz), 70.52, 53.18, 52.60, 37.89, 23.26, 22.02. **¹⁹F NMR** (376 MHz, Chloroform-*d*) δ -62.75. **HRMS (ESI-ToF)**: m/z calculated for C₁₆H₂₁F₃NO₄ [MH]⁺: 348.1423. Found 348.1403.



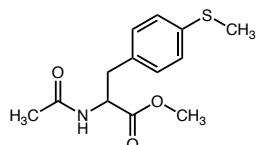
Methyl 2-acetamido-3-(benzo[*b*]thiophen-3-yl)propanoate

Isolated Yield 25.8% (25 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.06 – 7.80 (m, 1H), 7.81 – 7.63 (m, 1H), 7.64 – 7.30 (m, 2H), 7.13 (s, 1H), 6.10 (d, *J* = 7.7 Hz, 1H), 5.00 (dt, *J* = 7.7, 5.7 Hz, 1H), 3.69 (s, 3H), 3.56 – 2.99 (m, 2H), 1.96 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 172.20, 169.94, 140.37, 139.05, 130.86, 128.19, 124.57, 124.27, 123.91, 123.02, 121.63, 52.60, 30.80, 23.28. **HRMS (ESI-ToF)**: m/z calculated for C₁₄H₁₆NO₃S [MH]⁺: 278.0851. Found 278.0831.



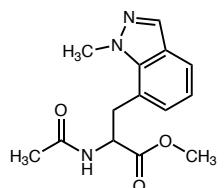
Methyl 2-acetamido-3-(1-methyl-1*H*-indazol-6-yl)propanoate

Isolated Yield 46.7% (45 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 8.07 – 7.77 (m, 1H), 7.61 (dd, *J* = 8.2, 0.8 Hz, 1H), 7.11 (d, *J* = 1.3 Hz, 1H), 6.87 (dd, *J* = 8.3, 1.3 Hz, 1H), 6.18 (d, *J* = 7.8 Hz, 1H), 4.93 (dt, *J* = 7.8, 6.0 Hz, 1H), 4.00 (s, 3H), 3.70 (s, 3H), 3.24 (qd, *J* = 13.9, 6.0 Hz, 2H), 1.96 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 172.18, 169.83, 140.24, 134.64, 132.60, 123.19, 122.25, 121.22, 109.22, 53.44, 52.43, 38.38, 35.47, 23.17. **HRMS (ESI-ToF)**: m/z calculated for C₁₄H₁₈N₃O₃ [MH]⁺: 276.1348. Found 276.1345.



Methyl 2-acetamido-3-(4-(methylthio)phenyl)propanoate

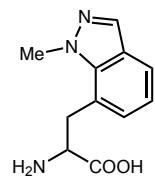
Isolated Yield 60.9% (57 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.23 – 7.07 (m, 2H), 7.04 – 6.89 (m, 2H), 6.07 (d, *J* = 7.6 Hz, 1H), 4.83 (dt, *J* = 7.8, 5.8 Hz, 1H), 3.70 (s, 3H), 3.05 (qd, *J* = 13.9, 5.8 Hz, 2H), 2.44 (s, 3H), 1.96 (s, 3H). **¹³C NMR** (101 MHz, Chloroform-*d*) δ 172.10, 169.76, 137.24, 132.67, 129.75, 126.73, 53.15, 52.40, 37.29, 23.13, 15.81. **HRMS (ESI-ToF)**: m/z calculated for C₁₃H₁₇NO₃SNa [MNa]⁺: 290.0827. Found 290.0805.



Methyl 2-acetamido-3-(1-methyl-1*H*-indazol-7-yl)propanoate

Isolated Yield 29.6% (28.5 mg). **¹H NMR** (400 MHz, Chloroform-*d*) δ 7.97 (s, 1H), 7.63 (dd, *J* = 6.6, 2.6 Hz, 1H), 7.12 – 6.95 (m, 2H), 6.02 (d, *J* = 8.0 Hz, 1H), 5.16 – 4.79 (m, 1H), 4.34 (s, 3H), 3.58 (s, 3H), 3.57 – 3.46 (m, 2H), 1.98 (s, 3H). **¹³C NMR** (126 MHz, Chloroform-*d*) δ 172.23,

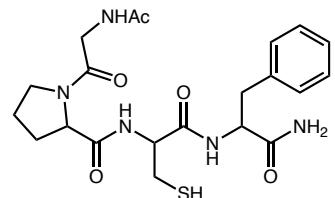
169.87, 138.86, 133.17, 128.47, 125.91, 120.75, 120.70, 118.76, 53.78, 52.49, 39.45, 35.19, 23.27.
HRMS (ESI-ToF): m/z calculated for C₁₄H₁₈N₃O₃ [MH]⁺: 276.1348. Found 276.1317.



2-amino-3-(1-methyl-1*H*-indazol-7-yl)propanoic acid

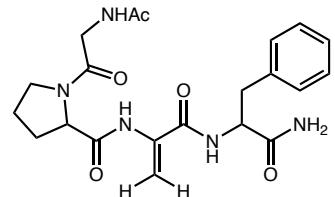
Isolated Yield 24.6% (4.9 mg). **¹H NMR** (400 MHz, Deuterium Oxide) δ 8.02 (s, 1H), 7.71 (dd, *J* = 8.1, 1.2 Hz, 1H), 7.22 (dd, *J* = 7.1, 1.1 Hz, 1H), 7.12 (dd, *J* = 8.1, 7.0 Hz, 1H), 4.22 (s, 3H), 3.82 (dd, *J* = 9.2, 5.9 Hz, 1H), 3.73 (dd, *J* = 14.9, 5.8 Hz, 1H), 3.24 (dd, *J* = 14.9, 9.2 Hz, 1H).

Peptides



Ac-Gly-Pro-Cys-Phe-NH₂

¹H NMR (400 MHz, Deuterium Oxide) δ 7.59 – 7.04 (m, 5H), 4.59 (dd, *J* = 8.9, 6.1 Hz, 1H), 4.55 – 4.29 (m, 2H), 4.06 (s, 2H), 3.70 – 3.49 (m, 2H), 3.17 (dd, *J* = 13.9, 6.0 Hz, 1H), 2.98 (dd, *J* = 14.1, 8.9 Hz, 1H), 2.90 – 2.63 (m, 2H), 2.19 (dq, *J* = 12.7, 7.8 Hz, 1H), 2.06 – 1.90 (m, 5H), 1.77 (dq, *J* = 12.3, 6.2 Hz, 1H).

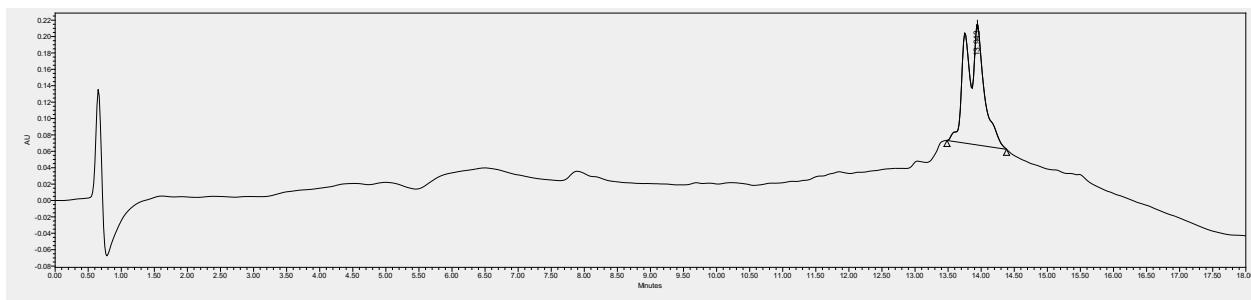


Ac-Gly-Pro-Dha-Phe-NH₂

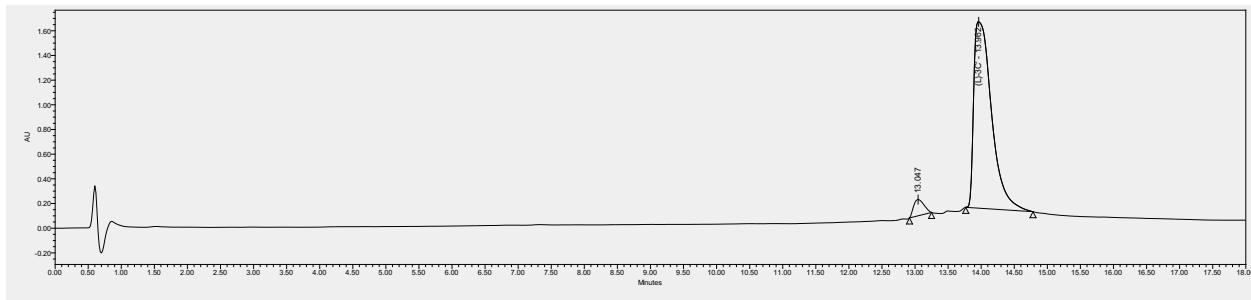
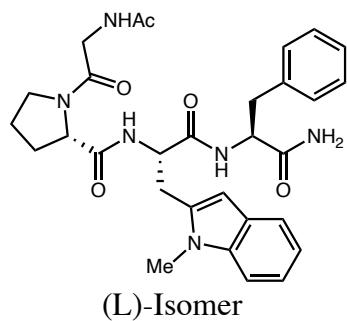
¹H NMR (400 MHz, Deuterium Oxide) δ 7.39 – 7.18 (m, 5H), 5.54 (dd, *J* = 11.4, 1.3 Hz, 2H), 4.59 (dd, *J* = 8.8, 6.1 Hz, 1H), 4.41 (dd, *J* = 8.5, 5.2 Hz, 1H), 4.05 (d, *J* = 2.2 Hz, 2H), 3.79 – 3.41 (m, 2H), 3.21 (dd, *J* = 13.8, 6.1 Hz, 1H), 3.02 (dd, *J* = 13.9, 8.9 Hz, 1H), 2.25 (ddd, *J* = 12.4, 8.2, 6.0 Hz, 1H), 2.10 – 1.76 (m, 6H).

3C' Isomers

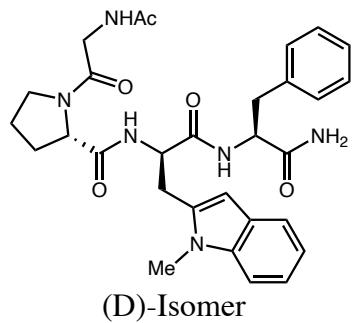
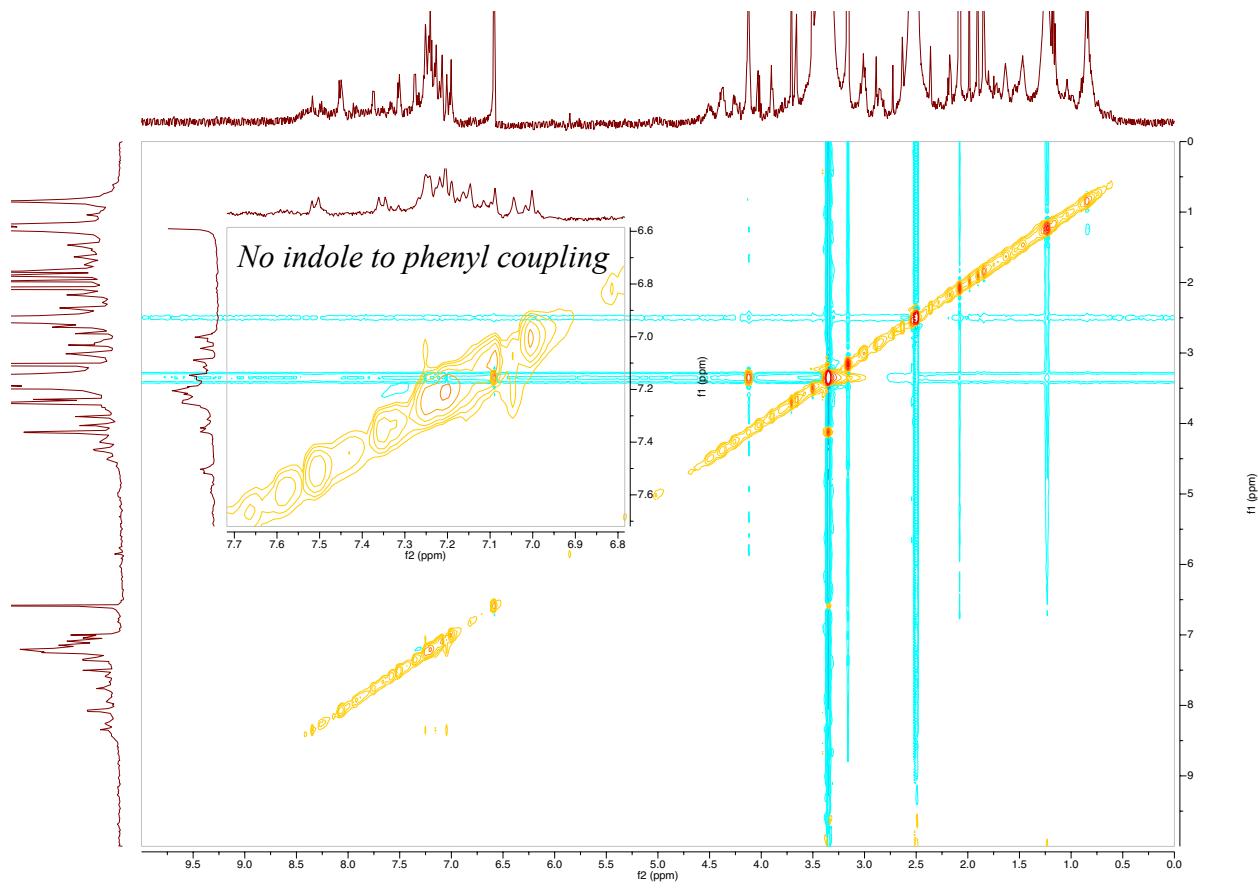
Crude mixture of both diastereomers:



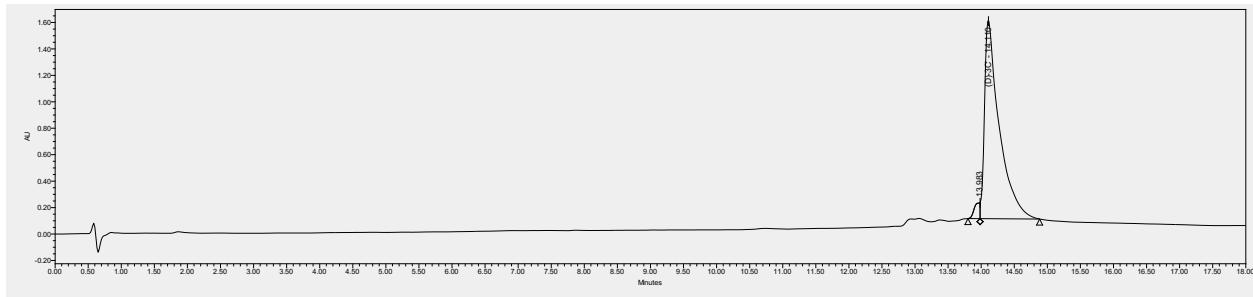
Pure products:



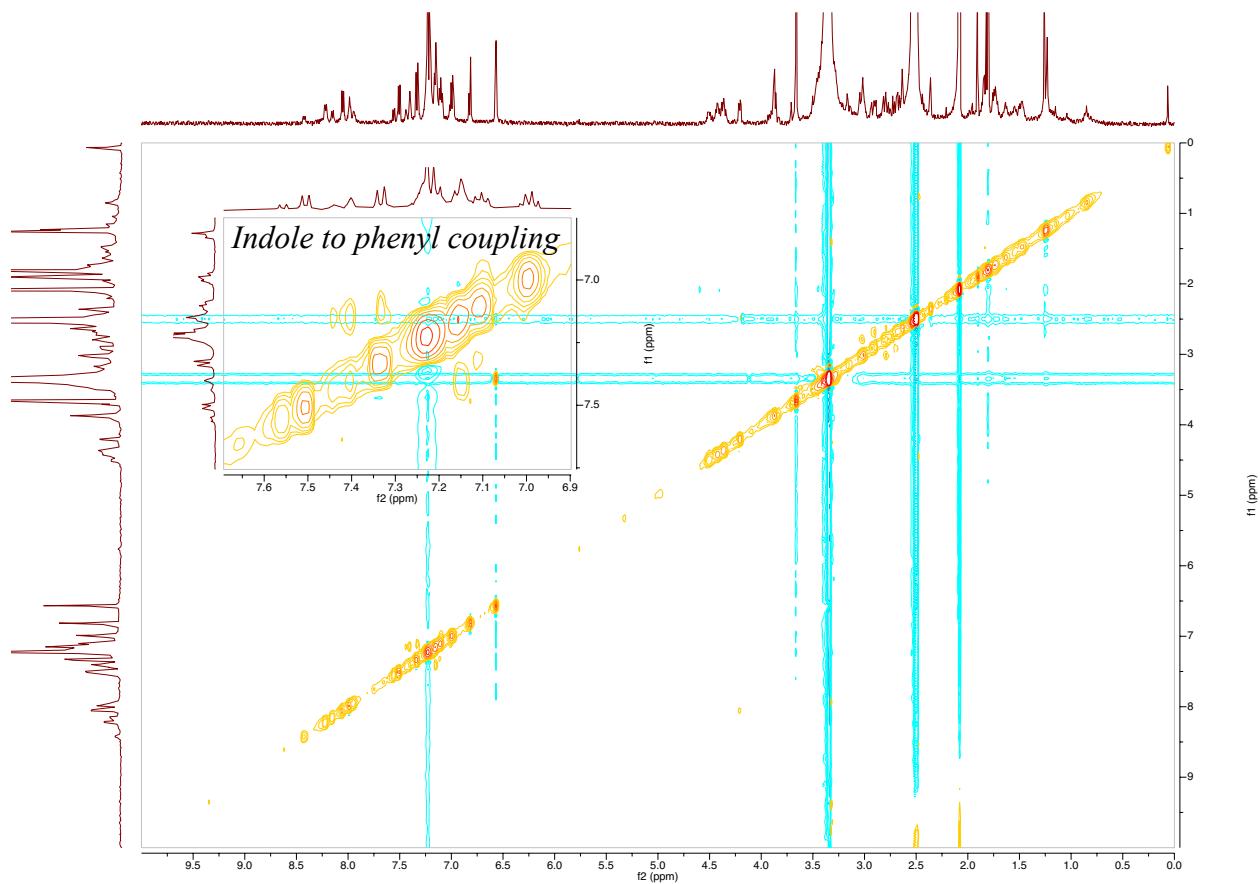
	Name	Retention Time	Area	% Area
1		13.047	1443948	4.84
2	(L)-3C'	13.962	28365465	95.16



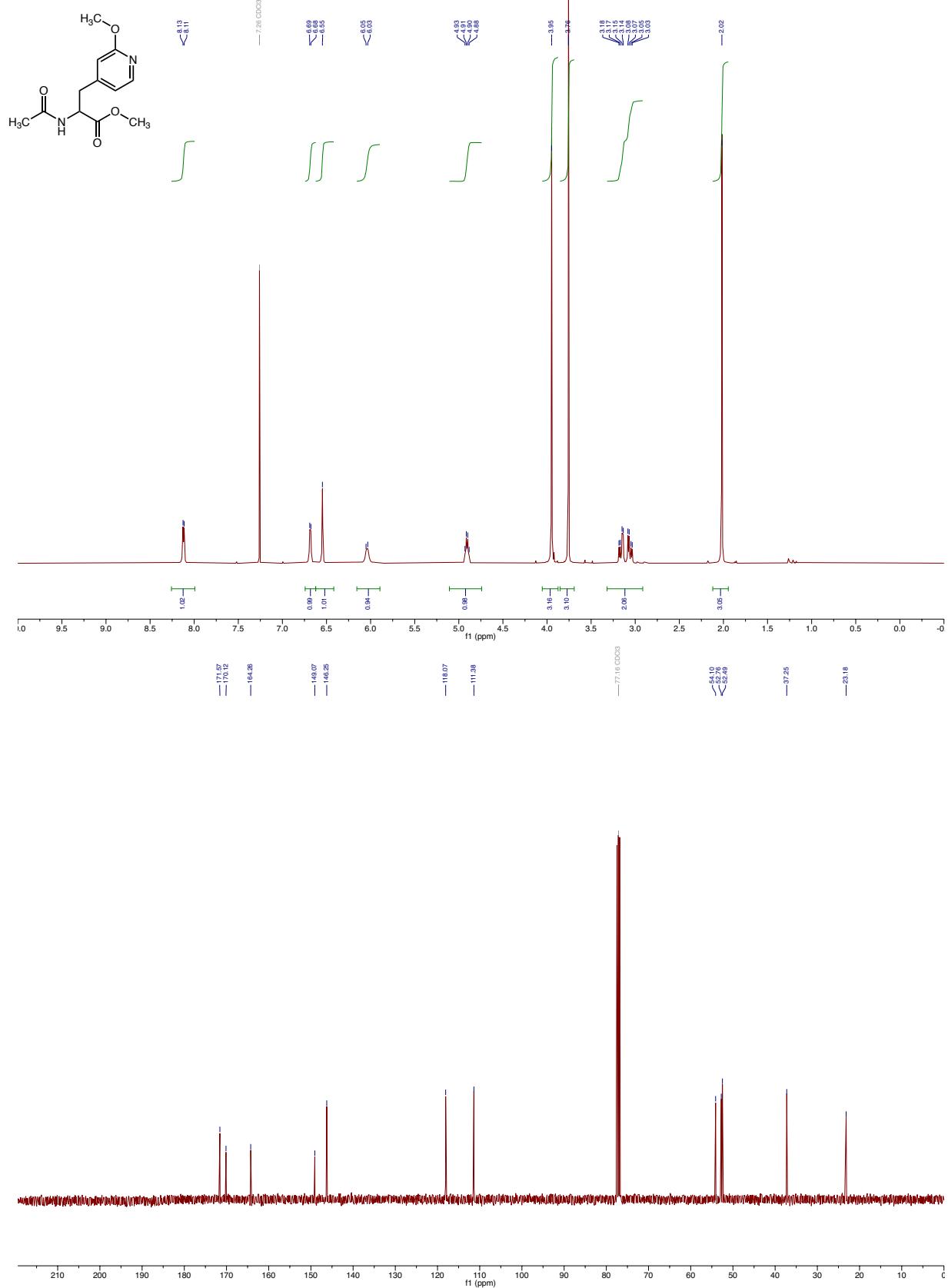
Indole and Phenyl on same side of peptide

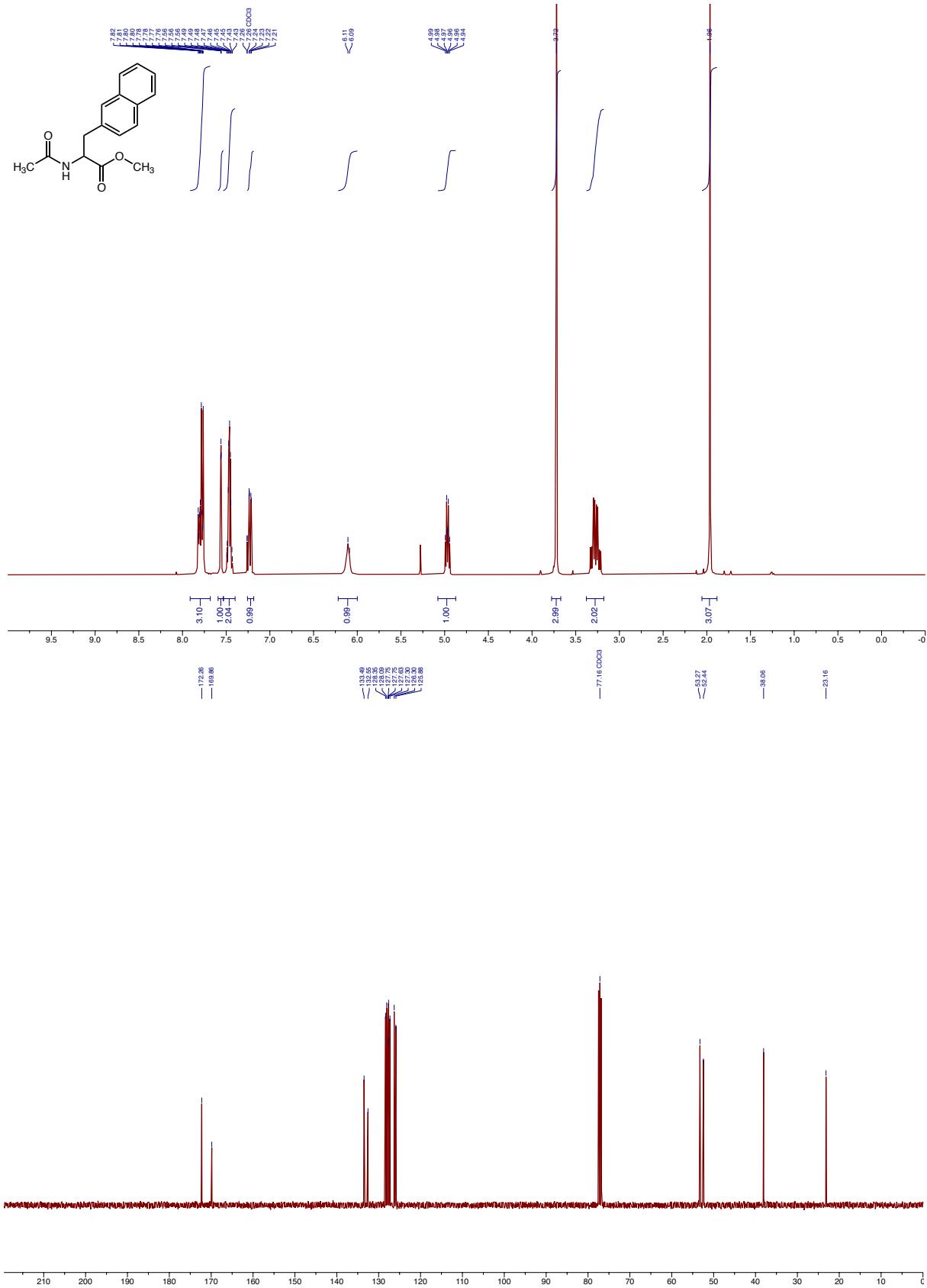


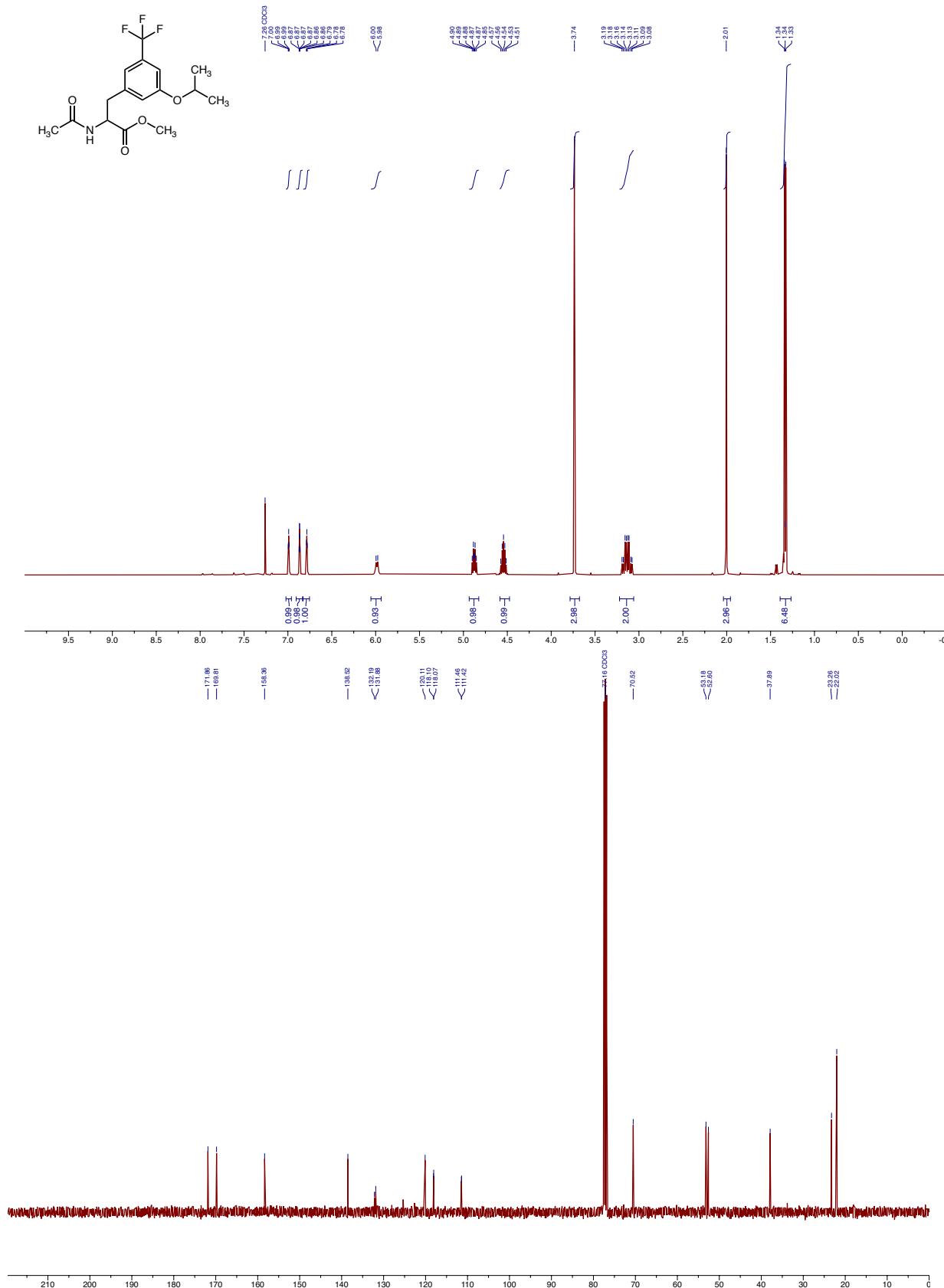
	Name	Retention Time	Area	% Area
1		13.983	707558	2.96
2	(D)-3C'	14.110	23201032	97.04

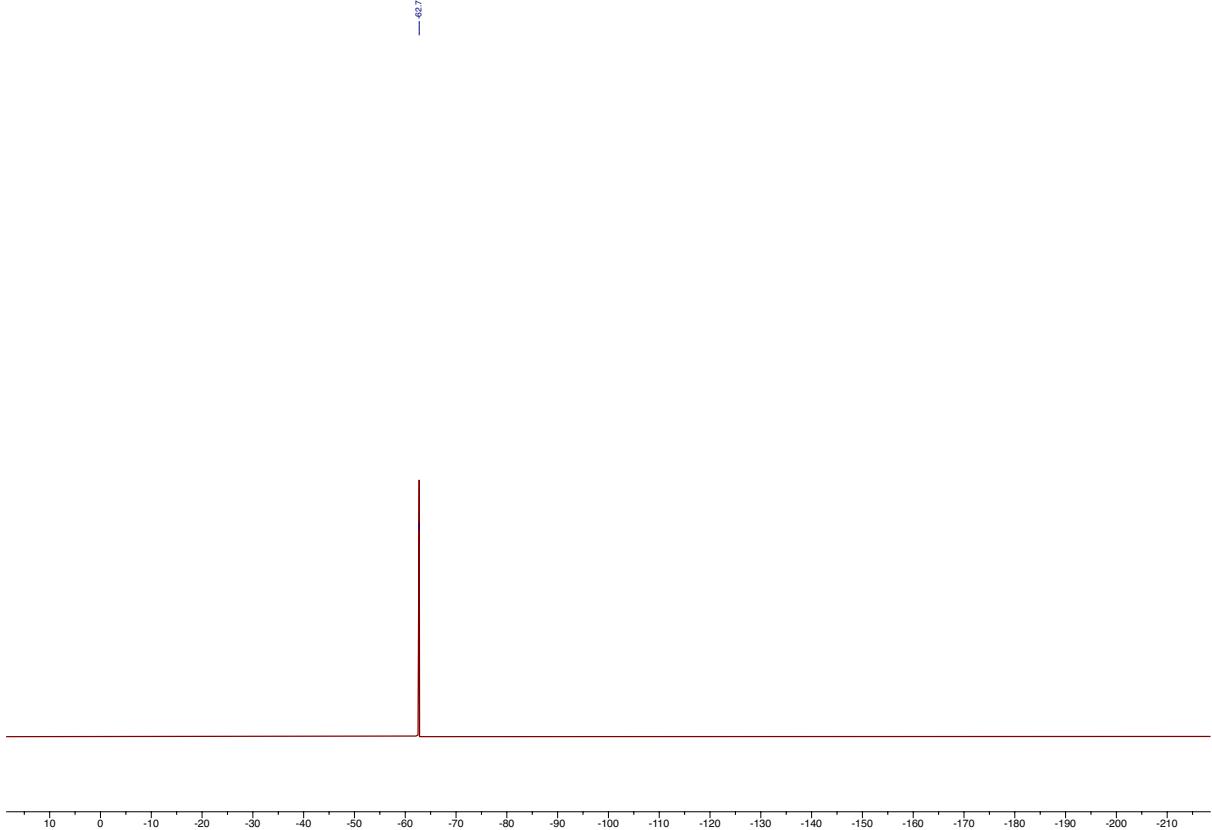


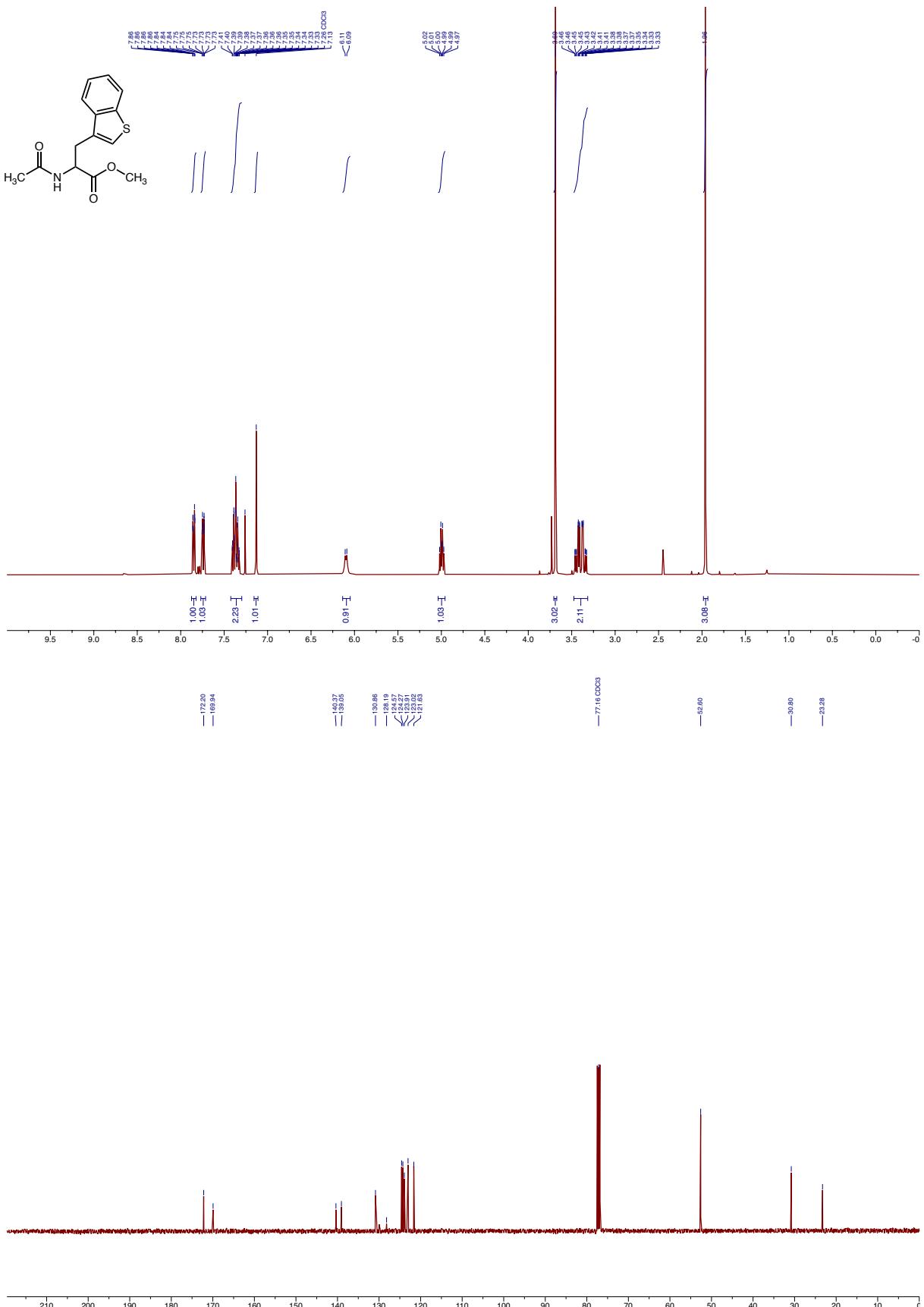
NMR Spectra

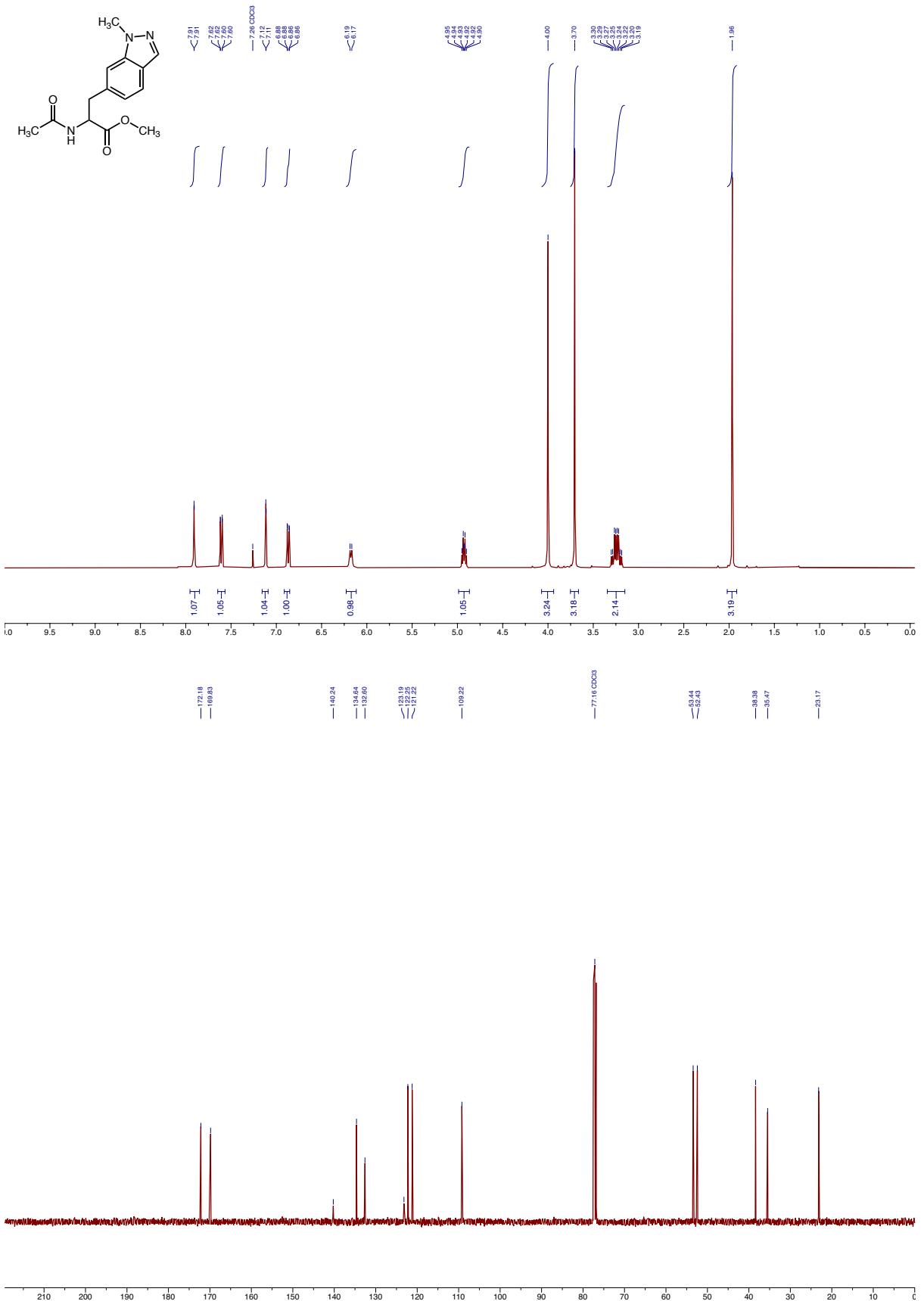


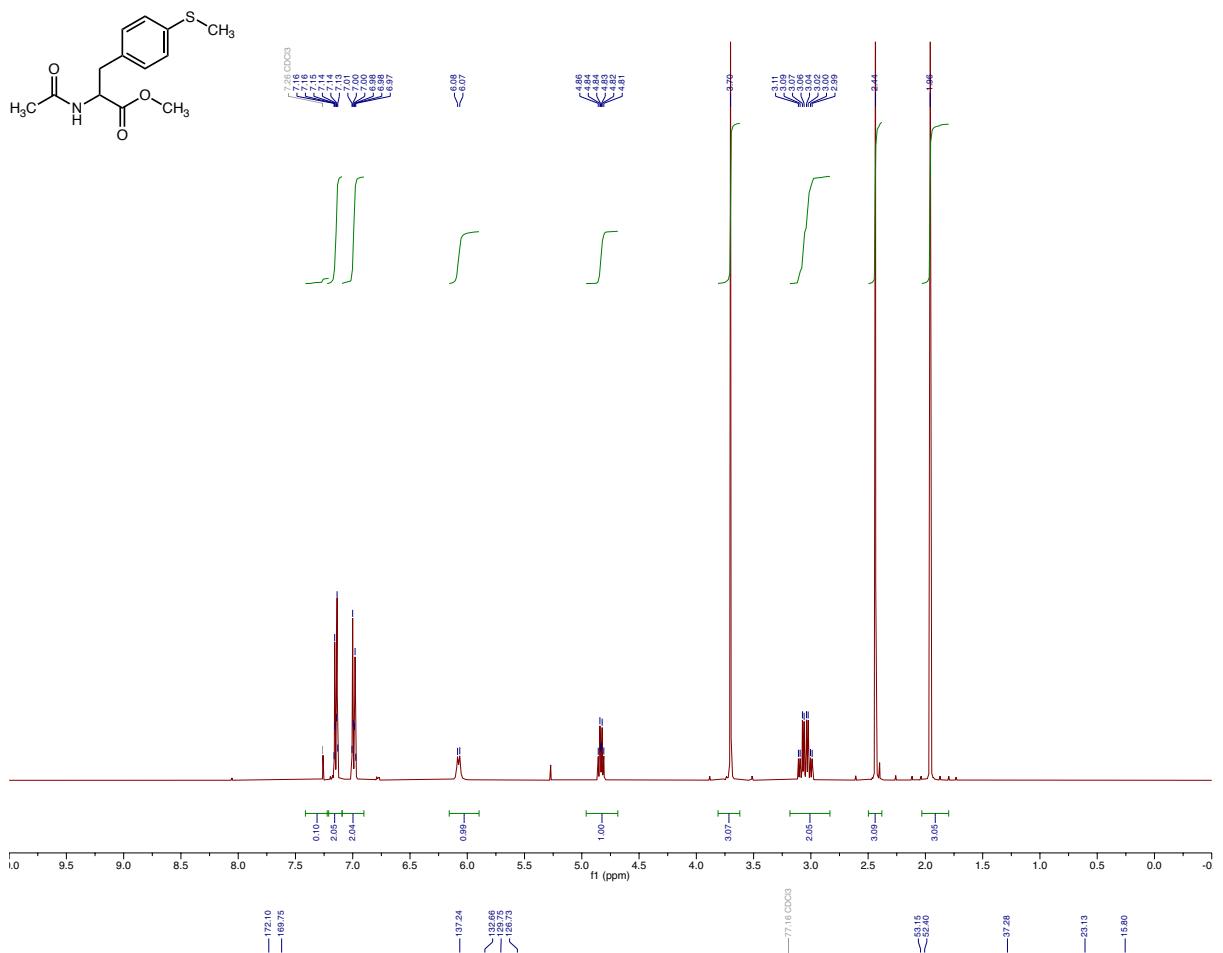


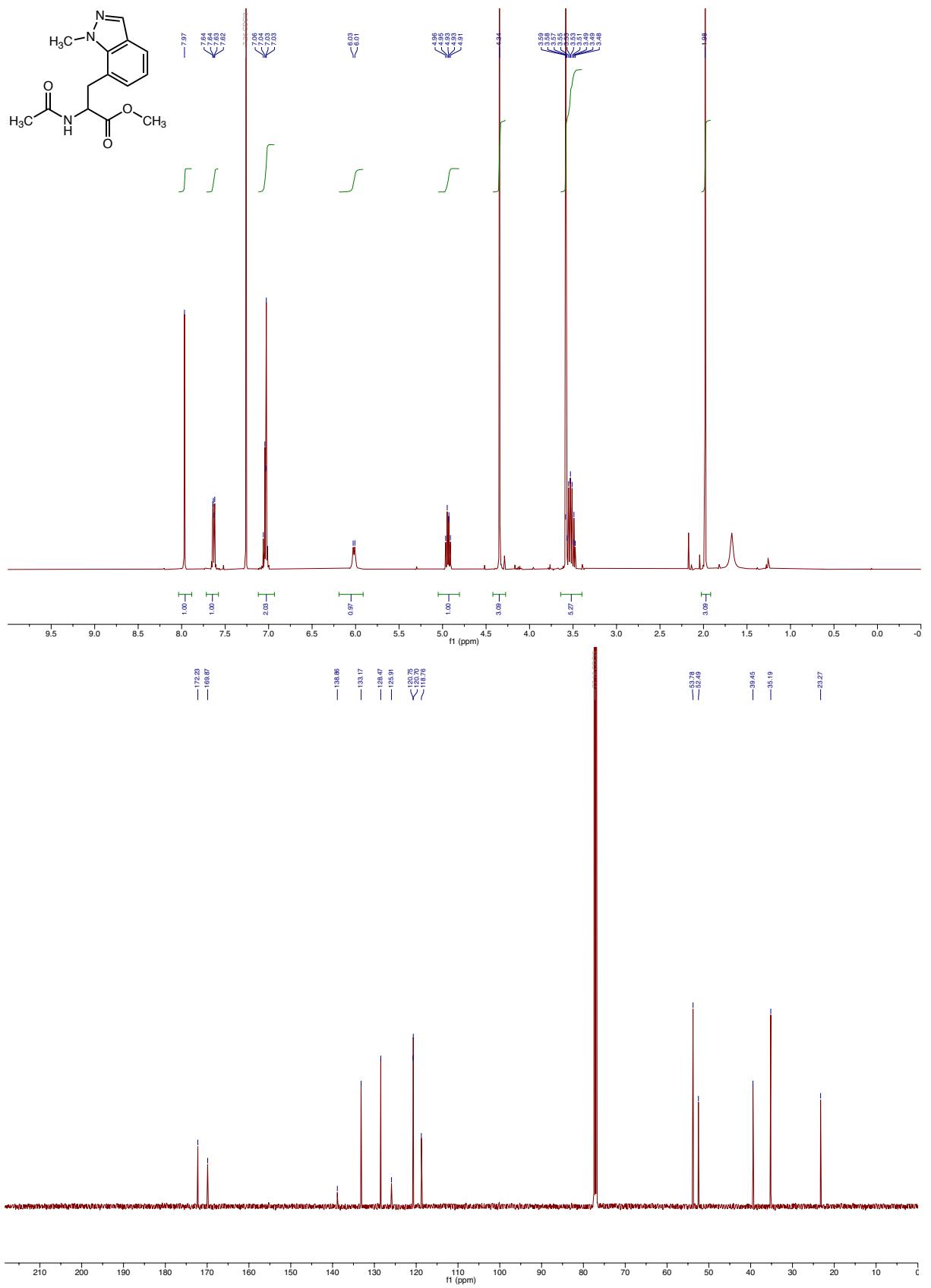


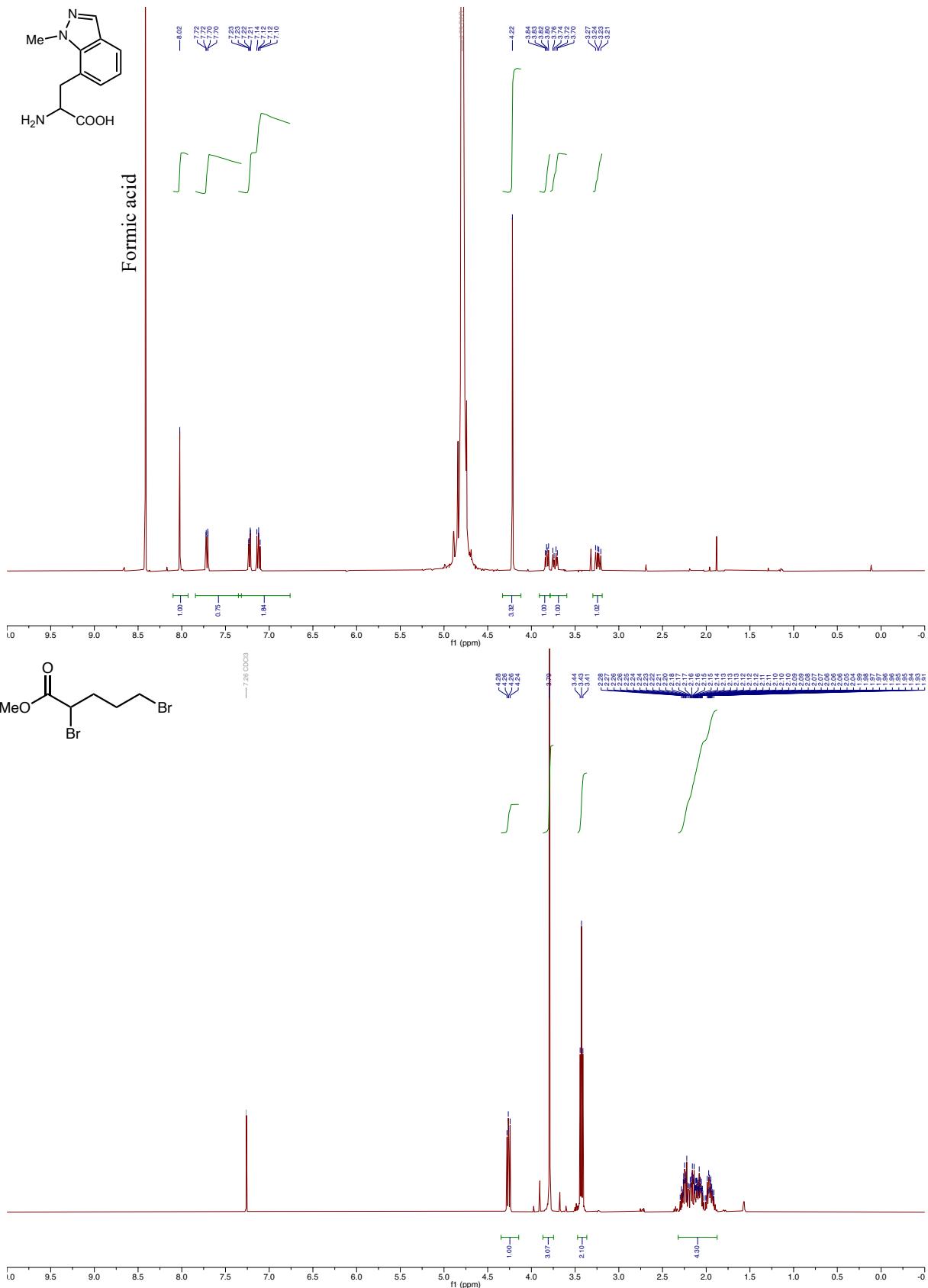


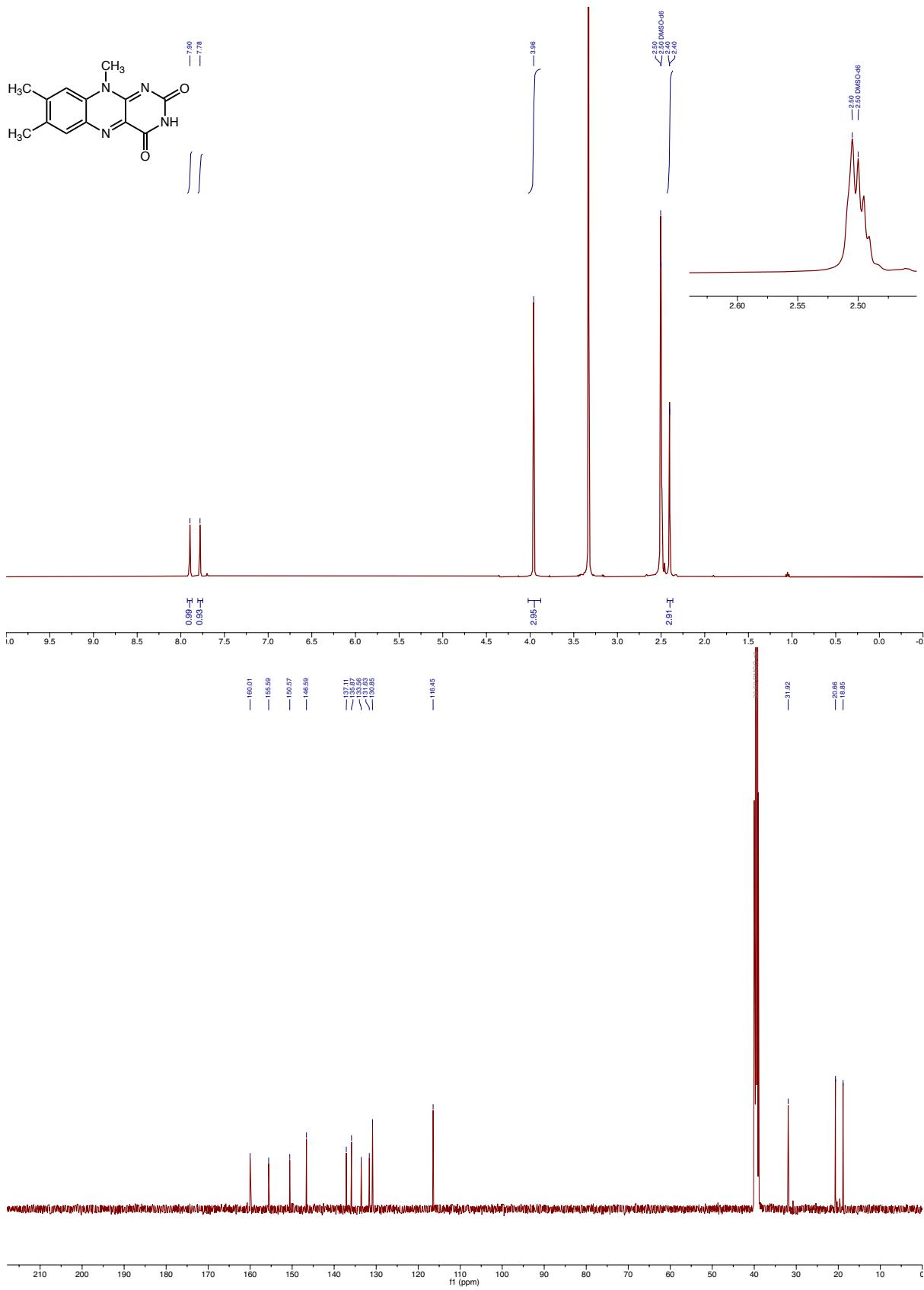


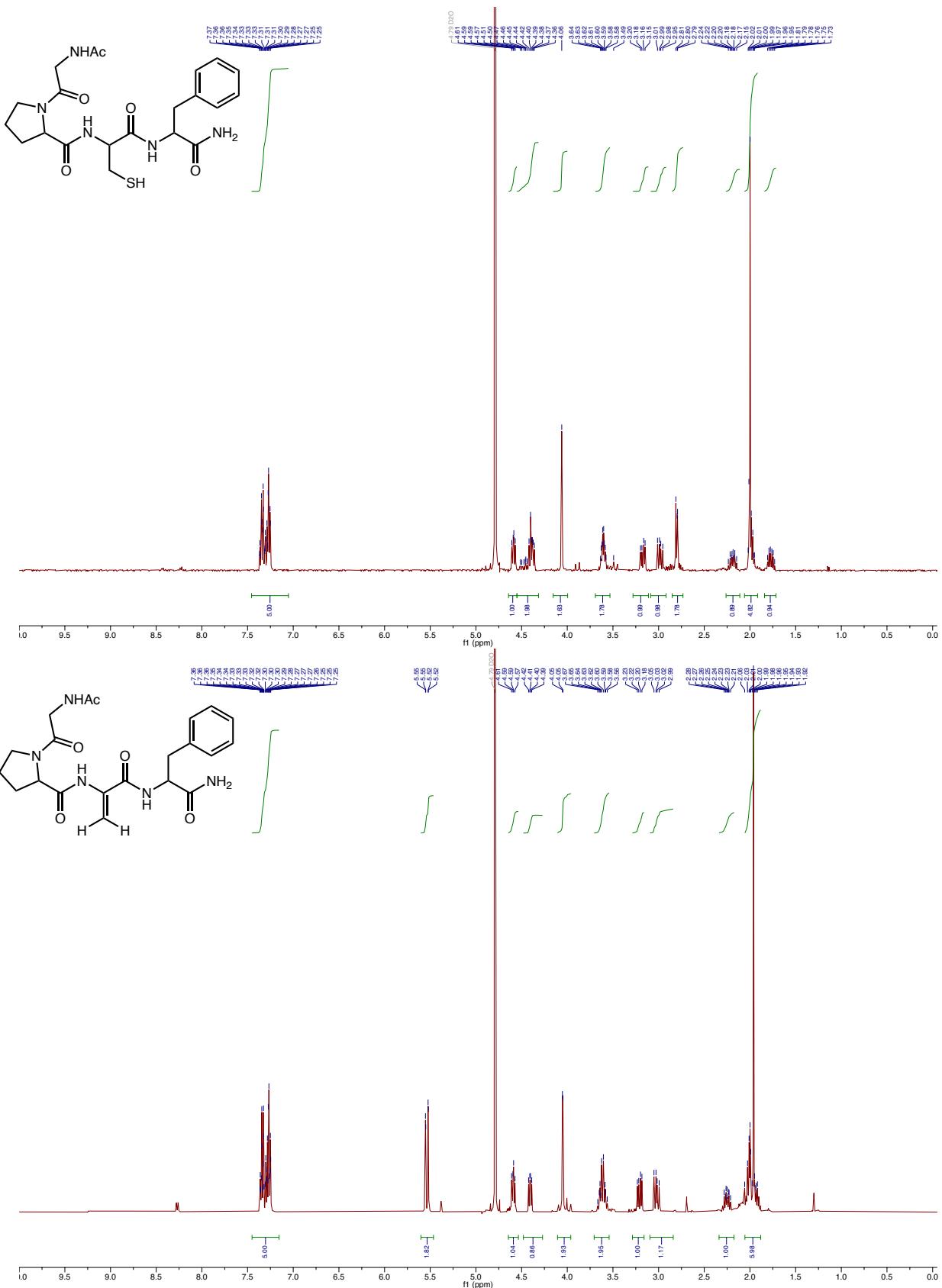




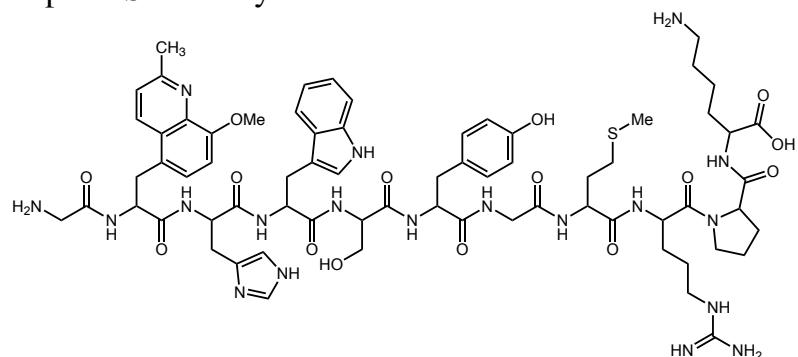




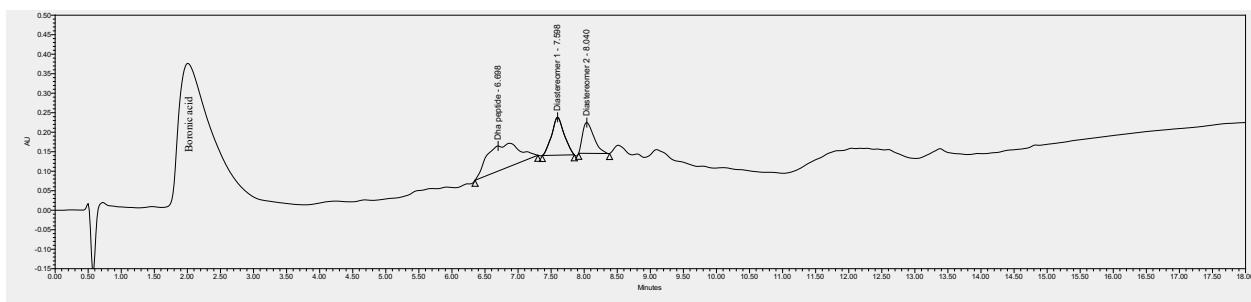




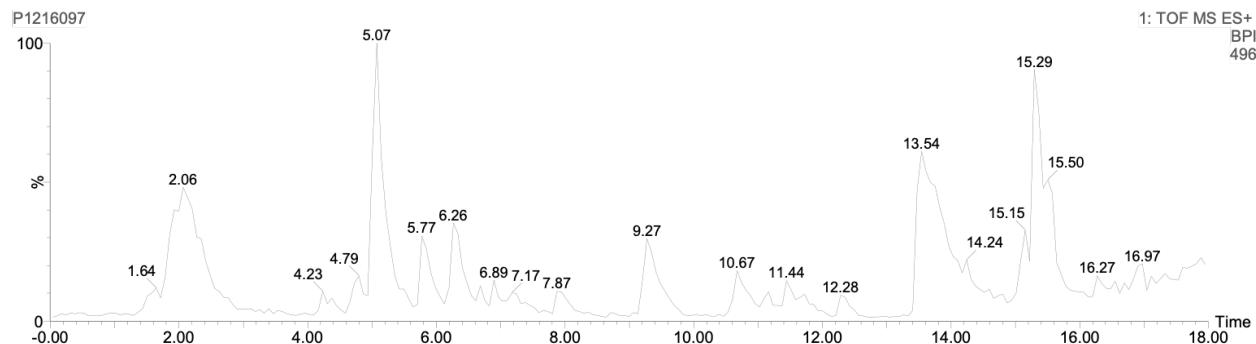
Peptide Selectivity

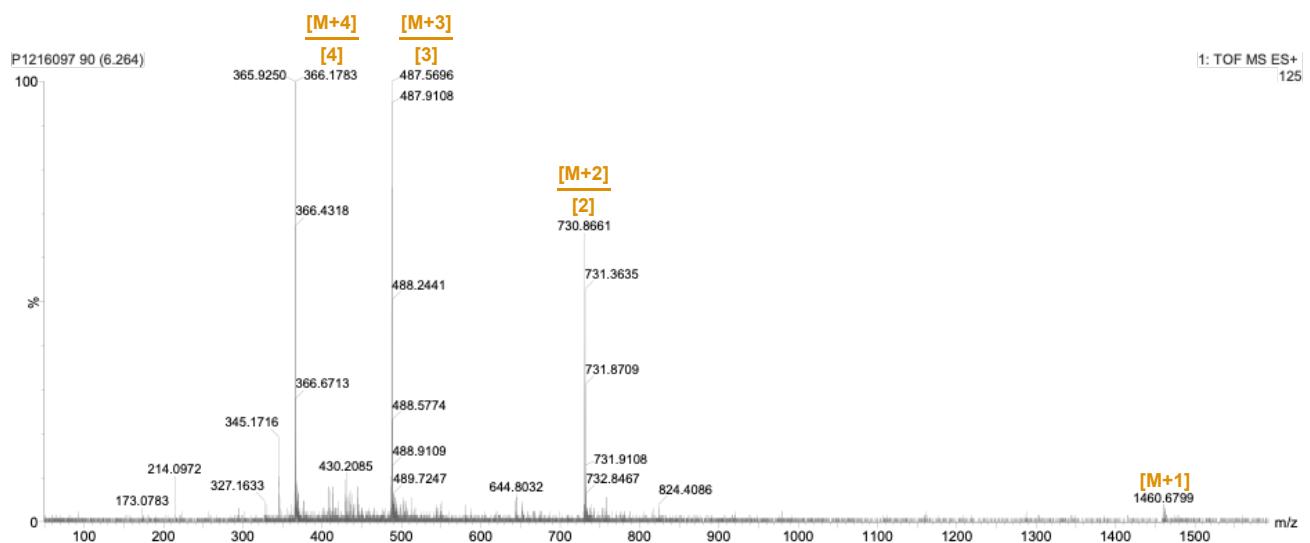
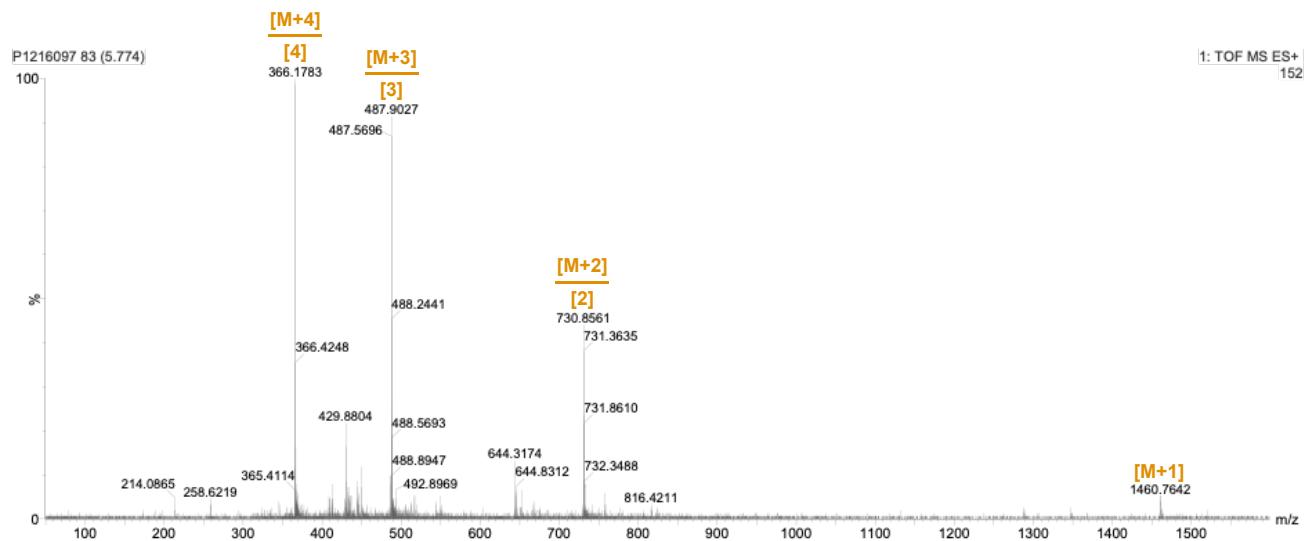


$\text{H}_2\text{N-Gly-[(8-MeO-2-Me)-5-Qin]-His-Trp-Ser-Tyr-Gly-Met-Arg-Pro-Lys-CO}_2\text{H}$



	Name	Retention Time	Area	% Area
1	Dha peptide	6.698	2168823	49.35
2	Diastereomer 1	7.598	1287871	29.31
3	Diastereomer 2	8.040	937734	21.34





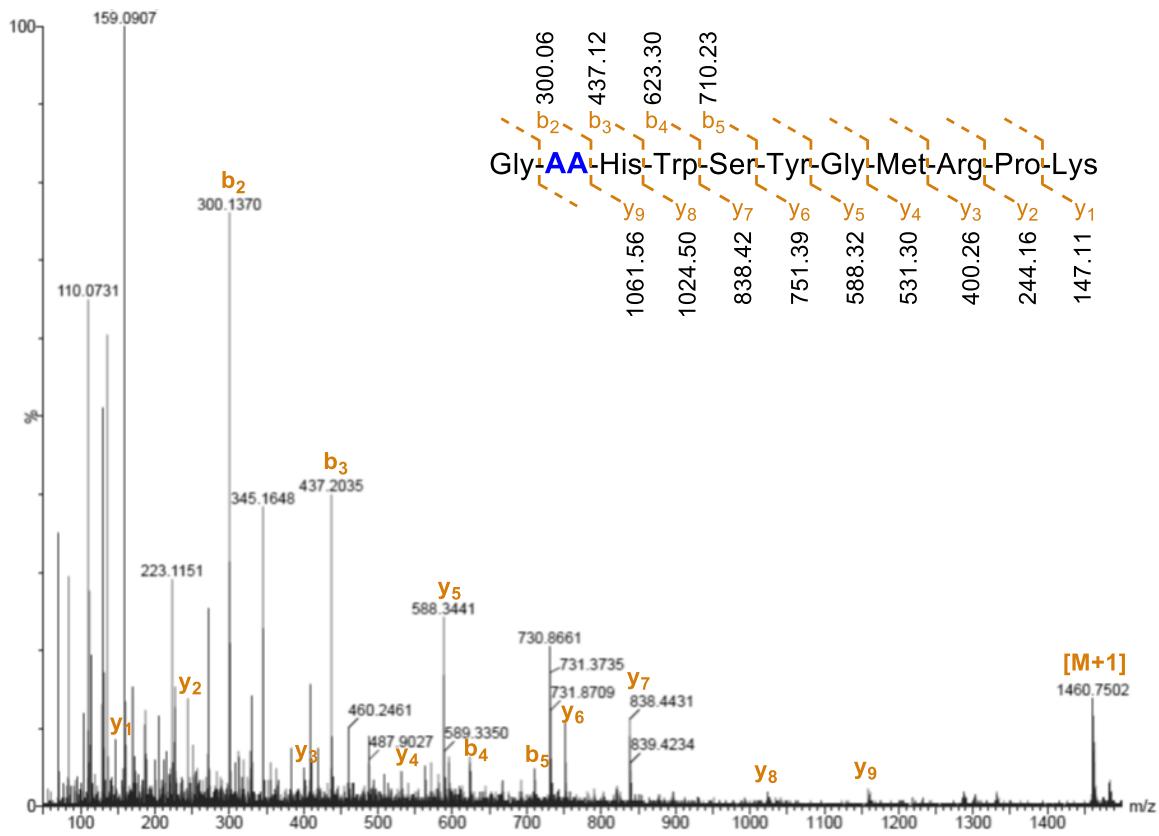


Plate Data

Liquid Chromatography analyses were performed on a Waters Acquity H-Class UPLC. Mass Spectrometry analyses were performed on a Quattro Ultima triple quadrupole mass analyzer with an electrospray ion source connected through column (without UV). The same column was used on both instruments, Waters Zorbax StableBondC18 3.5 μ m, 2.1 x 100 mm column, but the elution times did not match exactly.

Method: 0.5 mL/min flow rate. A = Acetonitrile with 1% formic acid modifier, B = ddH₂O with 1% formic acid modifier.

Time:	%A:	%B:
0.0	5.0	95.0
2.0	5.0	95.0
3.0	10.0	90.0
10.0	20.0	80.0
12.0	30.0	70.0
16.5	95.0	5.0
18.0	95.0	5.0

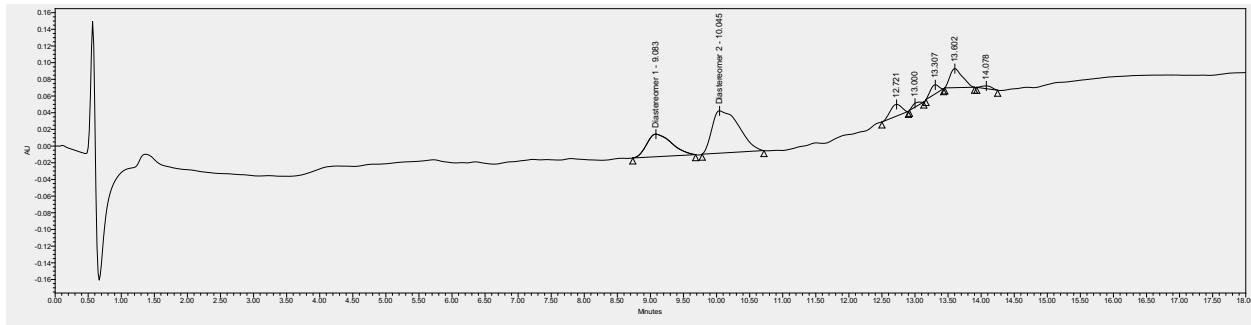
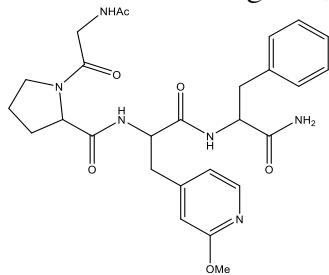
Table S10. 96-well plate results

Peptide:	MW:	Purity:	Amount:	Yield:
1A'	538.6	77.0%	0.096 mg	7.7%
2A'	556.6	51.6%	0.130 mg	10.0%
3A'	556.6	36.3%	0.107 mg	8.3%
4A'	614.7	36.3%	0.060 mg	4.2%
5A'	572.7	95.0%	0.206 mg	15.4%
6A'	566.7	91.7%	0.319 mg	24.2%
7A'	538.6	64.8%	0.217 mg	17.3%
8A'	568.6	81.1%	0.237 mg	17.9%
9A'	538.6	17.4%	0.035 mg	2.8%
10A'	556.6	61.9%	0.197 mg	15.2%
11A'	623.7	96.2%	0.163 mg	11.2%
12A'	554.7	85.5%	0.280 mg	21.6%
1B'	552.6	68.8%	0.226 mg	17.5%
2B'	552.6	46.1%	0.184 mg	14.3%
3B'	558.6	56.9%	0.043 mg	3.3%
4B'	558.6	91.1%	0.126 mg	9.7%

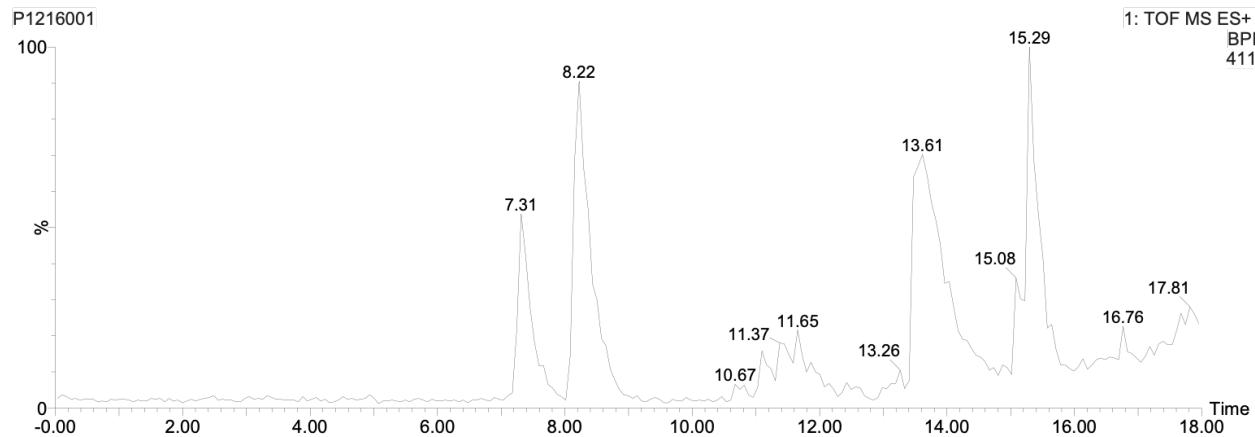
5B'	572.7	89.0%	0.193 mg	14.4%
6B'	602.7	78.2%	0.208 mg	14.8%
7B'	558.6	92.0%	0.217 mg	16.7%
8B'	558.6	92.3%	0.375 mg	28.8%
9B'	558.6	90.4%	0.254 mg	19.5%
10B'	558.6	73.6%	0.078 mg	6.0%
11B'	552.6	46.0%	0.059 mg	4.6%
12B'	555.6	42.8%	0.052 mg	4.0%
1C'	601.7	86.1%	0.078 mg	5.5%
2C'	569.6	41.7%	0.036 mg	2.7%
3C'	560.7	89.0%	0.047 mg	3.6%
4C'	546.6	77.8%	0.083 mg	6.5%
5C'	546.6	54.2%	0.123 mg	9.6%
6C'	547.6	62.0%	0.080 mg	6.2%
7C'	547.6	77.9%	0.162 mg	12.7%
8C'	561.6	69.2%	0.433 mg	33.1%
9C'	561.6	84.6%	0.311 mg	23.8%
10C'	561.6	74.9%	0.186 mg	14.2%
11C'	561.6	88.2%	0.230 mg	17.5%
12C'	548.6	57.3%	0.038 mg	3.0%
1D'	548.6	11.5%	0.003 mg	0.2%
2D'	561.6	66.5%	0.039 mg	3.0%
3D'	564.7	55.7%	0.066 mg	5.0%
4D'	563.7	17.1%	0.010 mg	0.7%
5D'	563.7	59.6%	0.021 mg	1.6%
6D'	565.6	75.0%	0.246 mg	18.7%
7D'	551.6	82.8%	0.216 mg	16.8%
8D'	497.6	35.3%	0.015 mg	1.3%
9D'	497.6	28.3%	0.025 mg	2.1%
10D'	511.6	5.2%	0.004 mg	0.3%
11D'	513.6	68.8%	0.090 mg	7.5%
12D'	513.6	48.4%	0.038 mg	3.2%
1E'	521.6	58.4%	0.072 mg	5.9%
2E'	535.7	45.8%	0.196 mg	15.7%
3E'	547.6	63.9%	0.042 mg	3.3%
4E'	563.7	46.7%	0.018 mg	1.4%
5E'	553.7	78.7%	0.228 mg	17.7%
6E'	537.6	84.9%	0.281 mg	22.4%
7E'	537.6	79.6%	0.178 mg	14.2%
8E'	537.6	79.5%	0.222 mg	17.7%

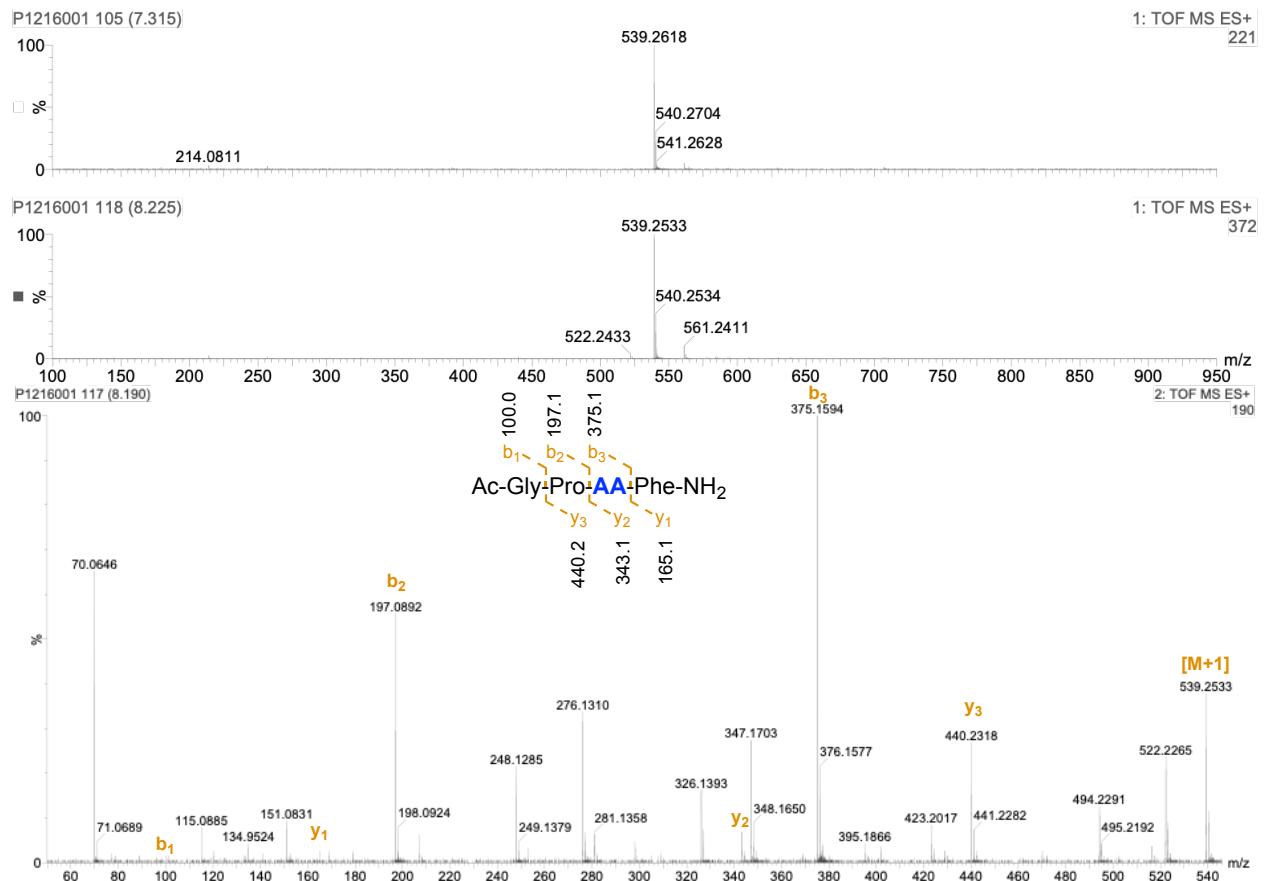
9E'	567.6	79.2%	0.296 mg	22.4%
10E'	567.6	88.7%	0.308 mg	23.3%
11E'	567.6	65.2%	0.086 mg	6.5%
12E'	621.7	59.1%	0.087 mg	6.0%
1F'	555.6	76.0%	0.189 mg	14.6%
2F'	613.7	27.0%	0.010 mg	0.7%
3F'	597.7	81.7%	0.614 mg	44.1%
4F'	599.7	37.2%	0.012 mg	0.8%
5F'	597.7	24.9%	0.019 mg	1.3%
6F'	535.7	21.0%	0.087 mg	6.9%
7F'	564.6	65.7%	0.186 mg	14.1%
8F'	622.7	82.8%	0.305 mg	21.0%
9F'	583.7	53.4%	0.017 mg	1.2%
10F'	584.7	78.8%	0.218 mg	16.0%
11F'	557.7	72.8%	0.044 mg	3.4%
12F'	557.7	86.1%	0.084 mg	6.5%
1G'	551.6	87.9%	0.297 mg	23.1%
2G'	521.6	73.5%	0.057 mg	4.7%
3G'	573.6	76.9%	0.134 mg	10.0%
4G'	555.6	71.7%	0.149 mg	11.5%
5G'	553.7	82.4%	0.454 mg	35.2%
6G'	633.7	86.7%	0.262 mg	17.8%
7G'	581.6	71.2%	0.056 mg	4.2%
8G'	535.6	80.6%	0.158 mg	12.7%
9G'	551.6	67.7%	0.133 mg	10.4%
10G'	572.7	66.2%	0.095 mg	7.1%
11G'	565.7	78.9%	0.228 mg	17.3%
12G'	581.7	69.2%	0.376 mg	27.7%
1H'	529.6	81.1%	0.192 mg	15.6%
2H'	551.6	49.7%	0.112 mg	8.7%
3H'	517.6	38.1%	0.124 mg	10.3%
4H'	600.7	69.7%	0.178 mg	12.7%
5H'	543.7	12.4%	0.029 mg	2.3%
6H'	491.6	51.1%	0.193 mg	16.8%
7H'	600.7	35.2%	0.042 mg	3.0%
8H'	485.6	50.9%	0.161 mg	14.2%
9H'	586.7	28.5%	0.112 mg	8.2%
10H'	473.6	68.9%	0.316 mg	28.7%
11H'	513.6	75.1%	0.156 mg	13.1%
12H'	487.6	53.6%	0.090 mg	7.9%

1A': MW = 538.6 g/mol, Purity = 77.0%, Yield = 7.7% [0.096 mg]

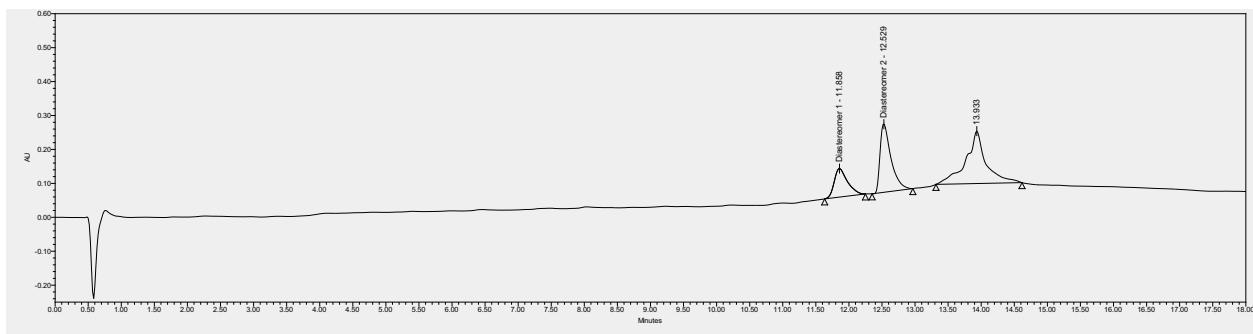
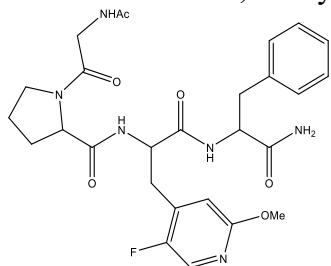


	Name	Retention Time	Area	% Area
1	Diastereomer 1	9.083	708389	25.47
2	Diastereomer 2	10.045	1432789	51.51
3		12.721	170064	6.11
4		13.000	35460	1.27
5		13.307	97753	3.51
6		13.602	300746	10.81
7		14.078	36556	1.31

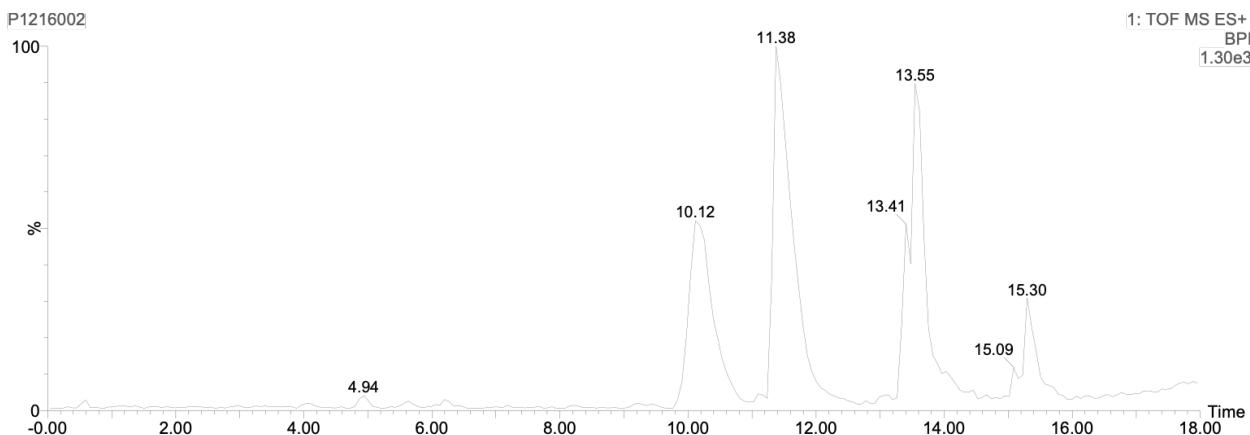


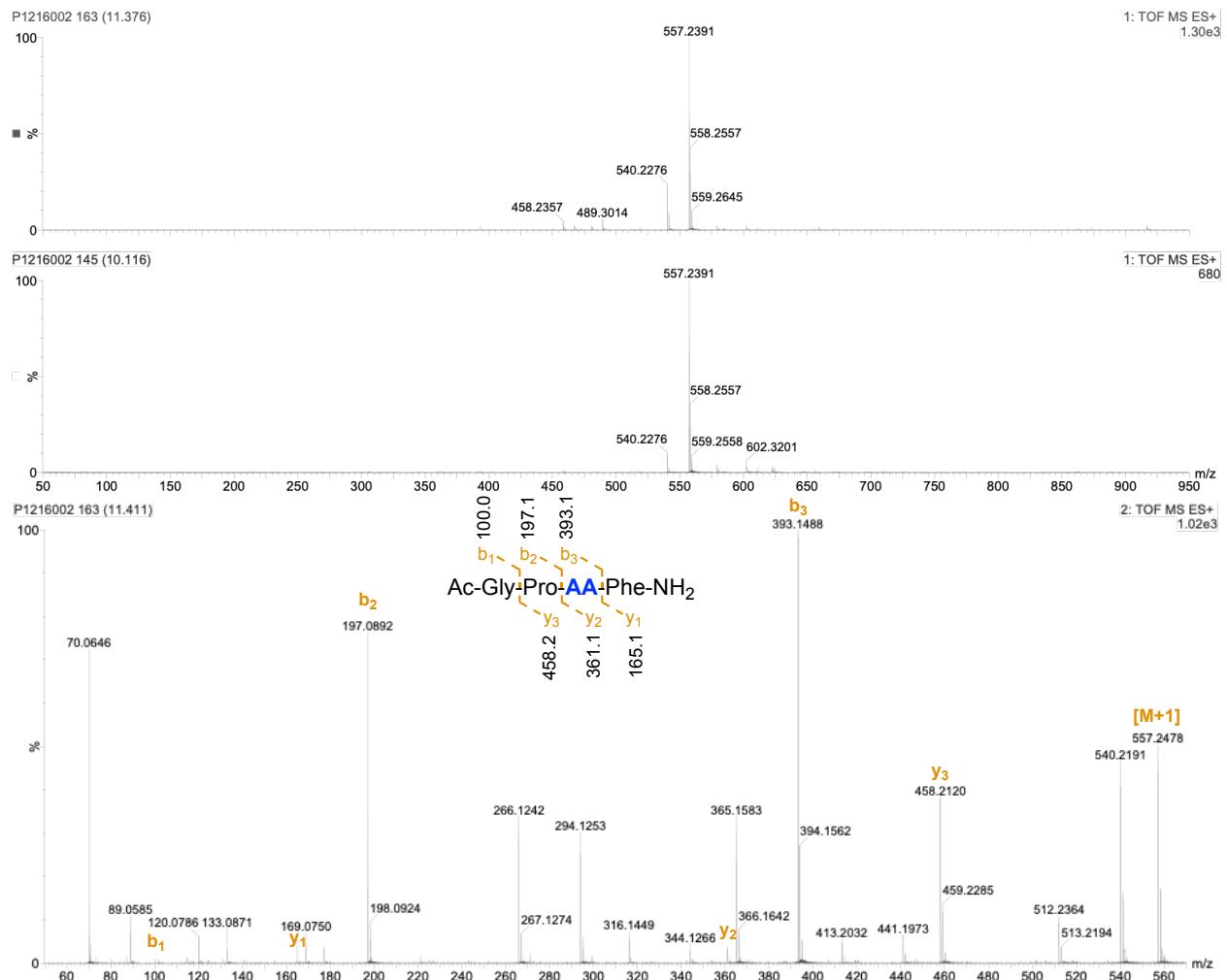


2A': MW = 556.6, Purity = 51.6% , Yield = 10.0% [0.13 mg]

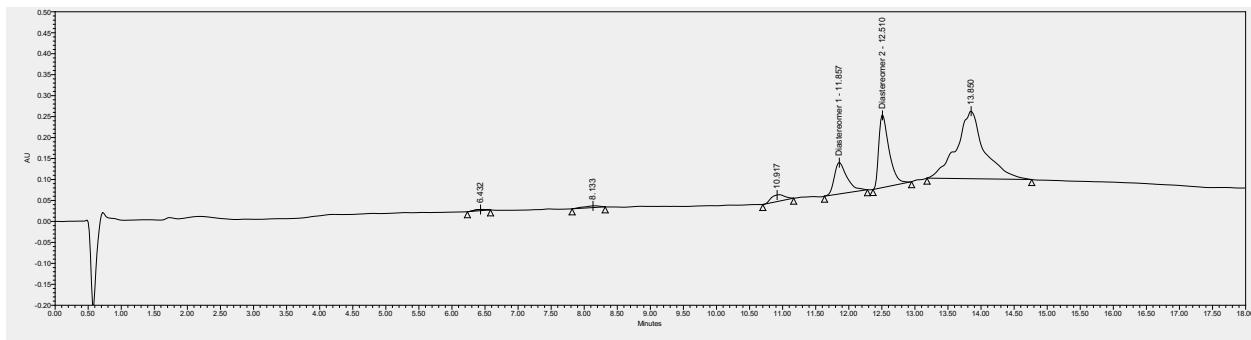
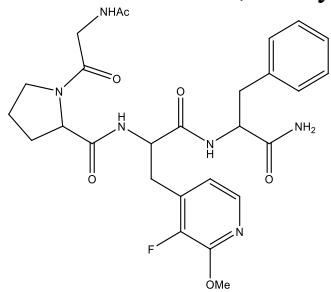


	Name	Retention Time	Area	% Area
1	Diastereomer 1	11.858	1167576	17.31
2	Diastereomer 2	12.529	2314842	34.31
3		13.933	3264156	48.38

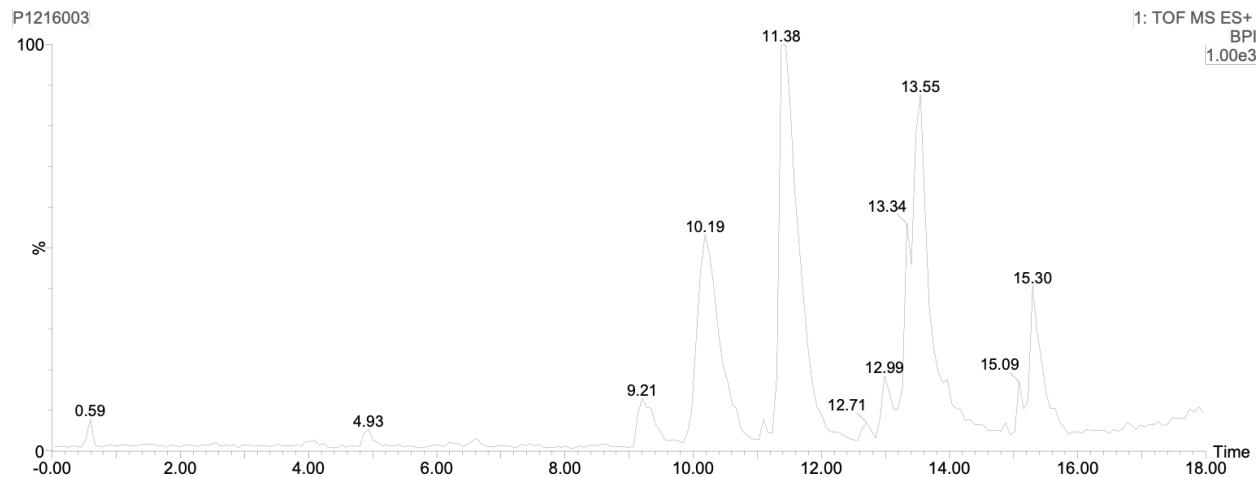


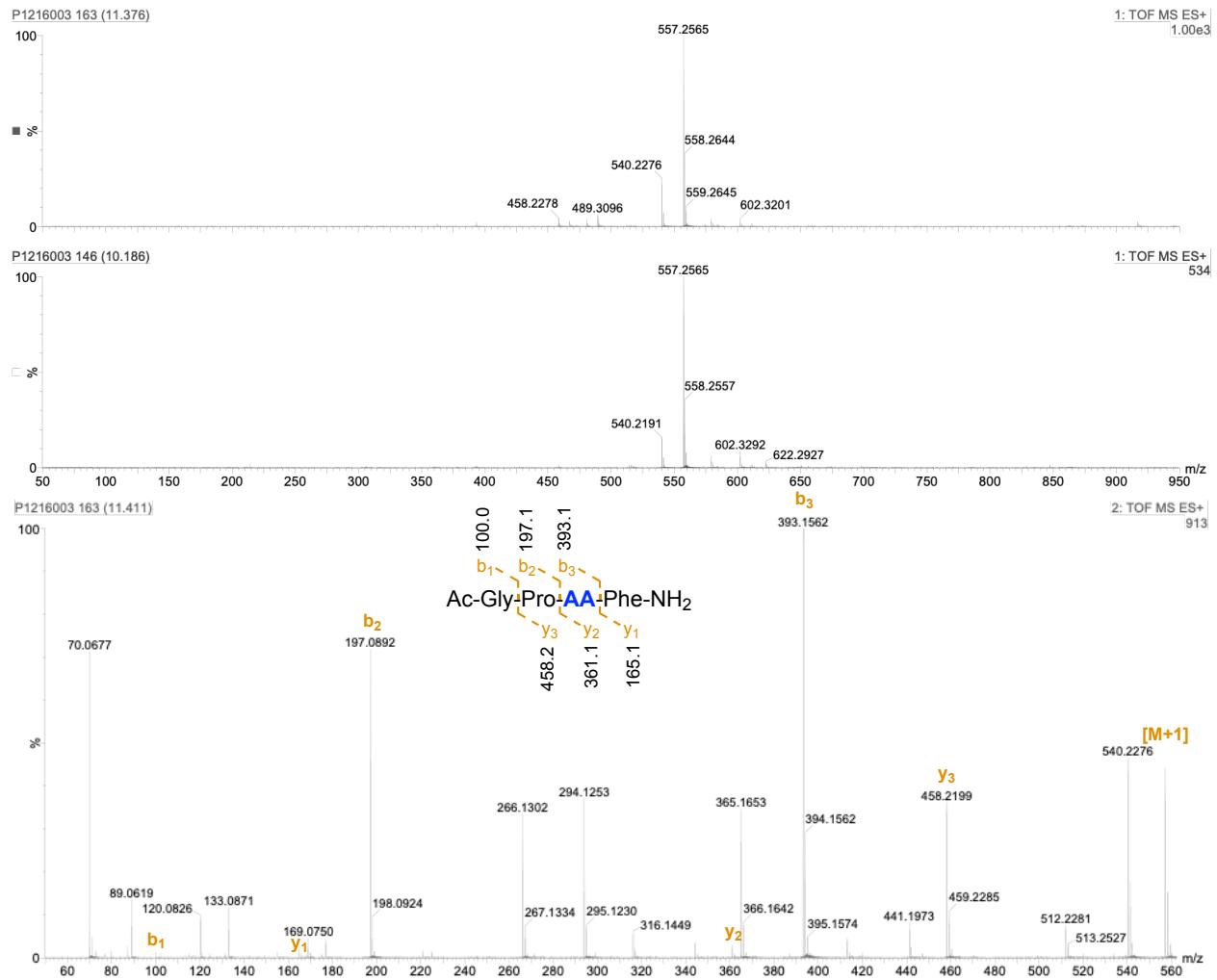


3A': MW = 556.6, Purity = 36.3%, Yield = 8.3% [0.11 mg]

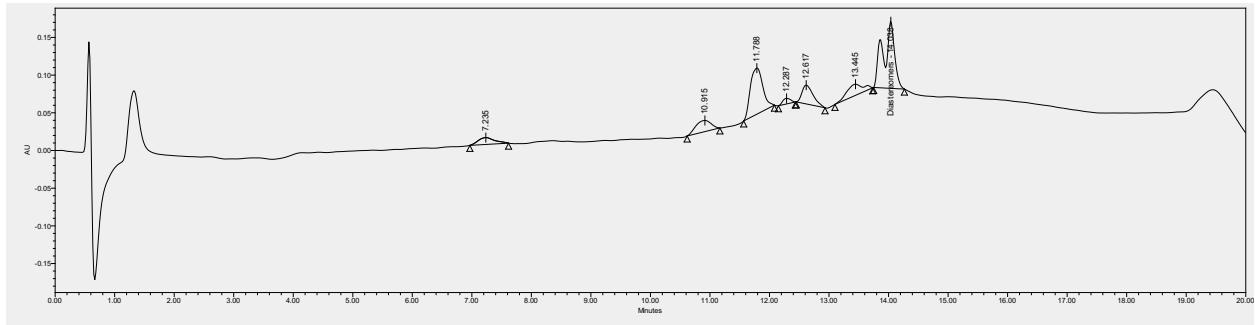
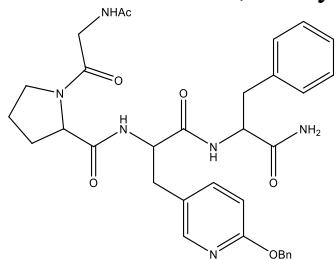


	Name	Retention Time	Area	% Area
1		6.432	35074	0.44
2		8.133	72544	0.91
3		10.917	226520	2.85
4	Diastereomer 1	11.857	1003370	12.62
5	Diastereomer 2	12.510	1878973	23.63
6		13.850	4736735	59.56

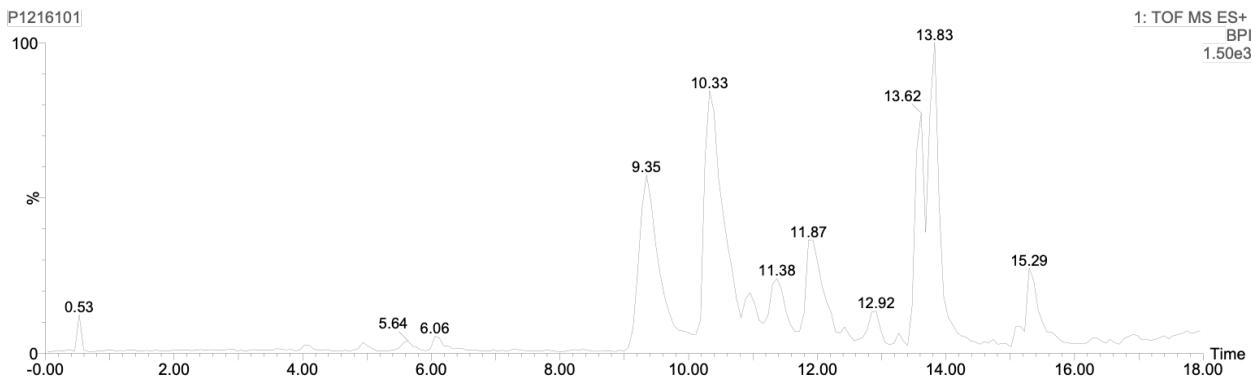


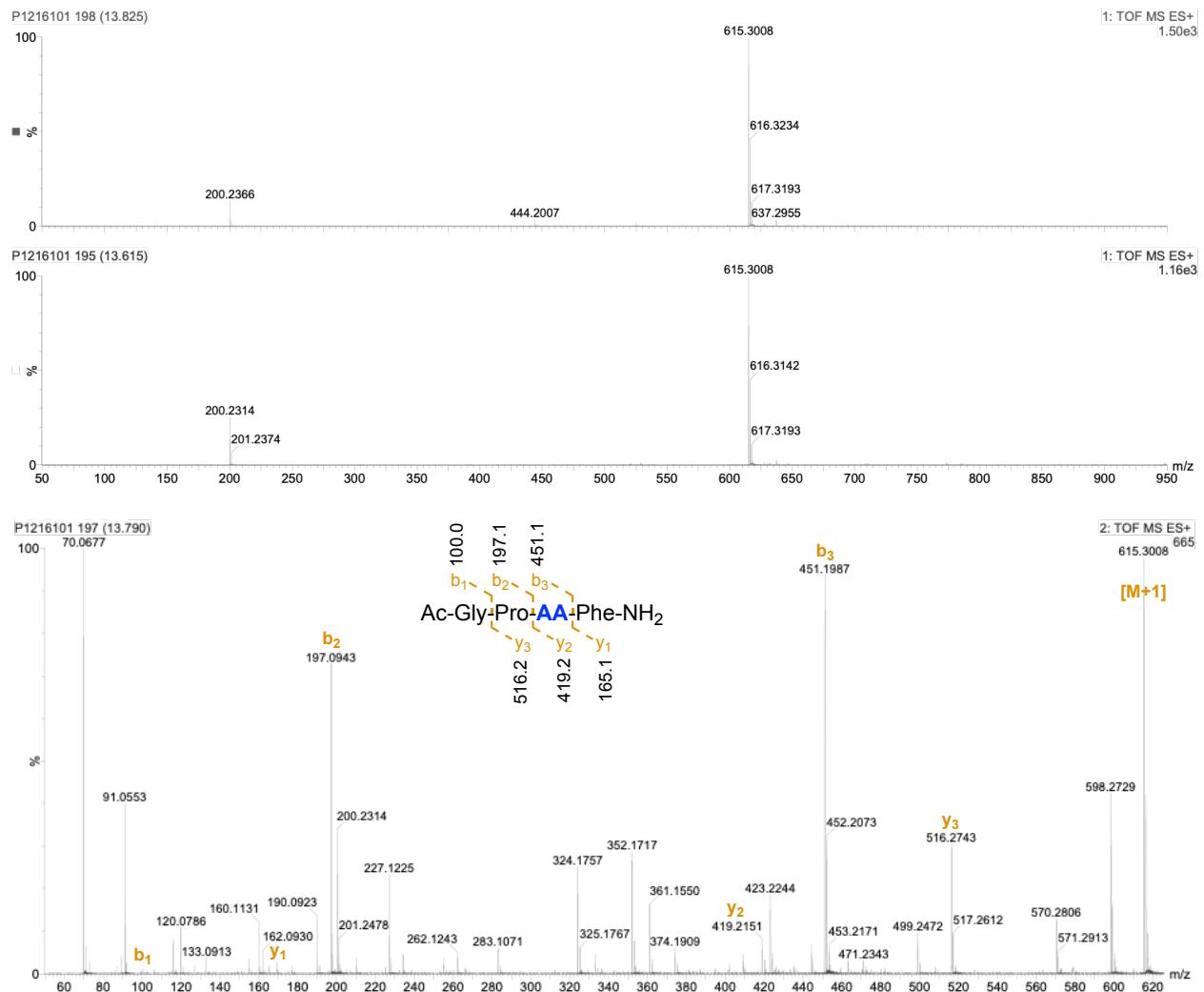


4A': MW = 614.7, Purity = 36.3%, Yield = 4.2% [0.06 mg]

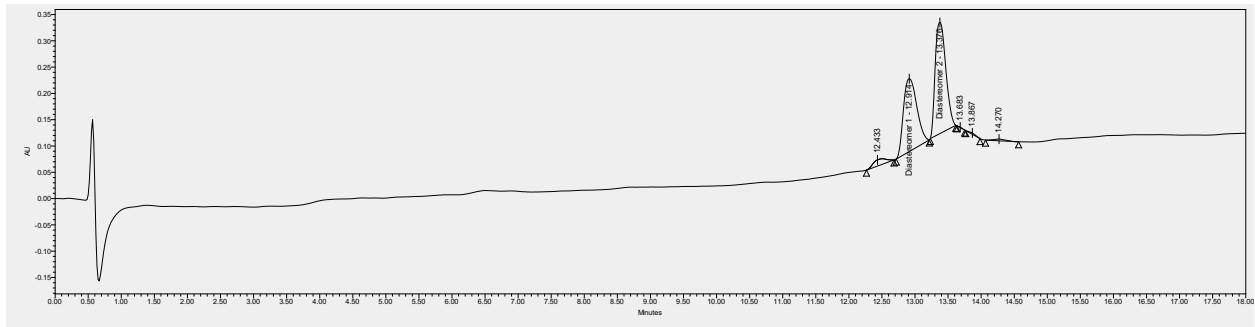
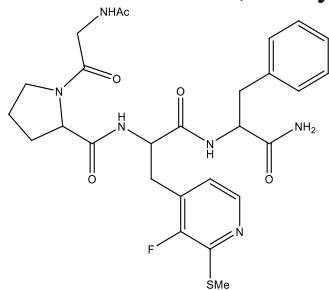


	Name	Retention Time	Area	% Area
1		7.235	176821	5.55
2		10.915	272606	8.56
3		11.788	896333	28.14
4		12.287	67824	2.13
5		12.617	326932	10.27
6		13.445	286984	9.01
7	Diastereomers	14.038	1157225	36.34

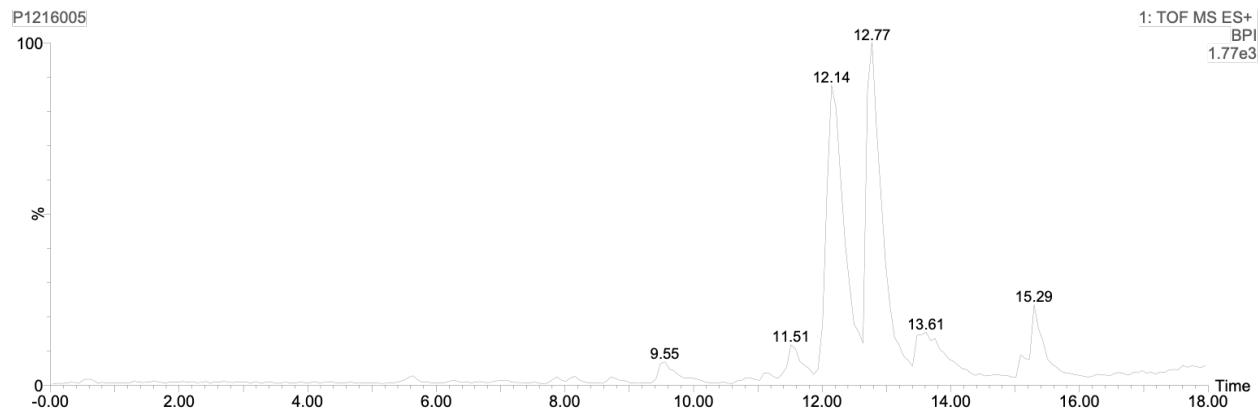


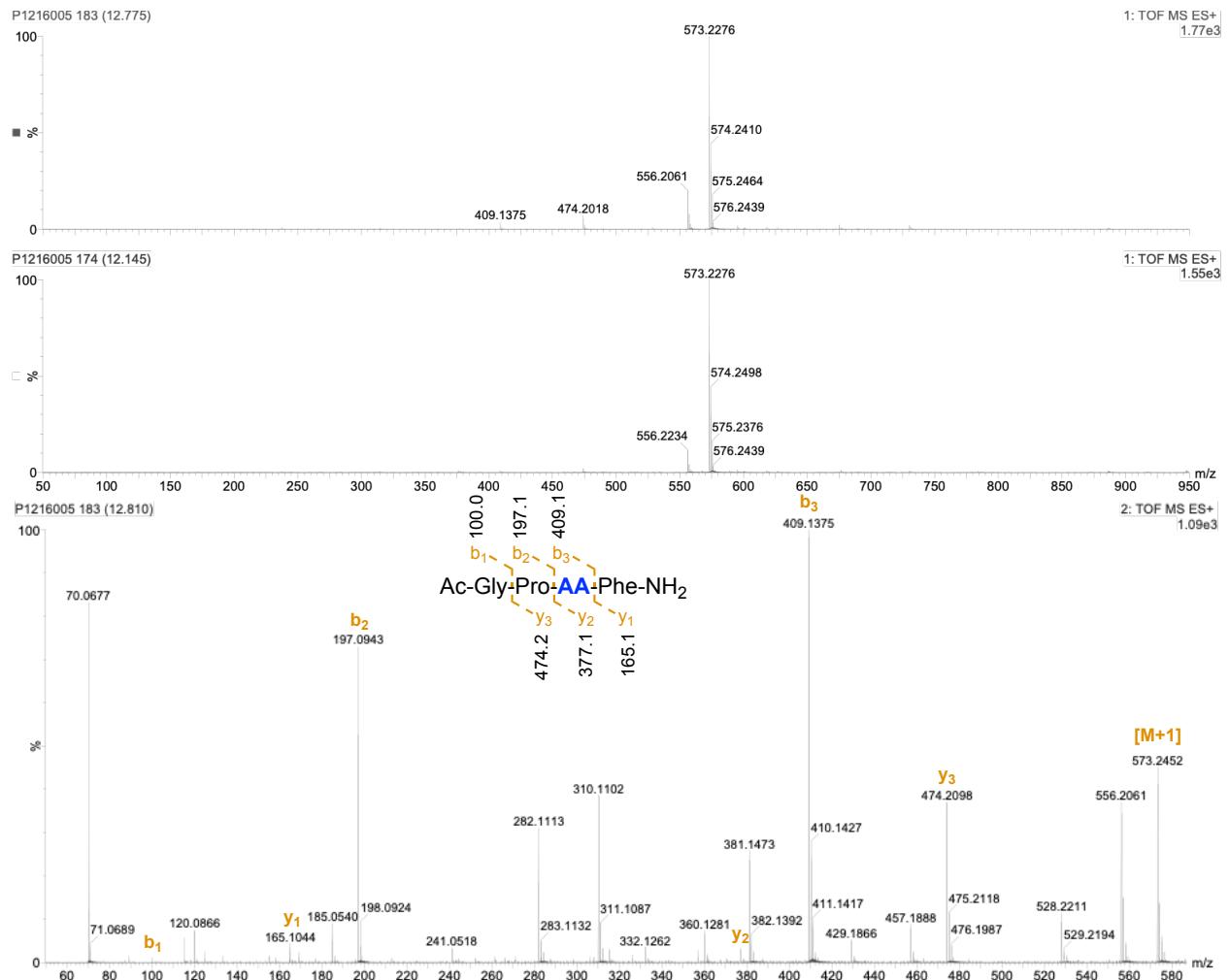


5A': MW = 572.7, Purity = 95.0% , Yield = 15.4% [0.21 mg]

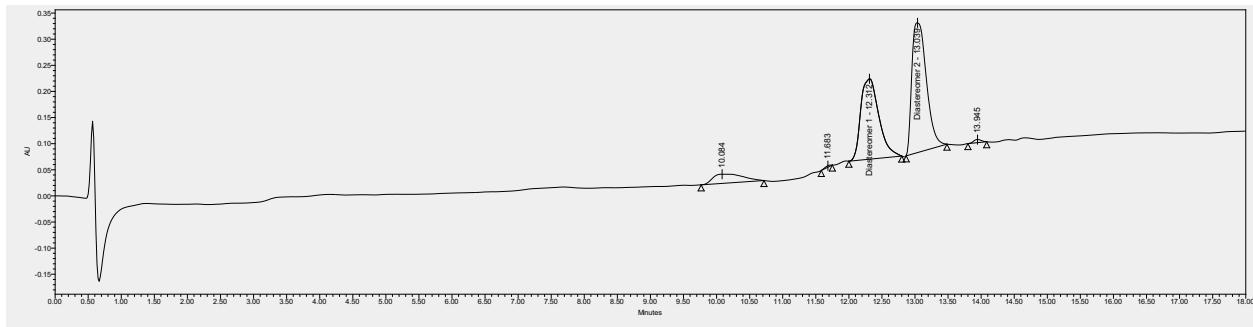
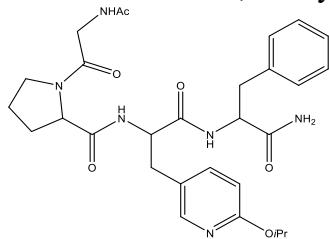


	Name	Retention Time	Area	% Area
1		12.433	162641	3.60
2	Diastereomer 1	12.914	1946297	43.08
3	Diastereomer 2	13.376	2346200	51.93
4		13.683	4790	0.11
5		13.867	16592	0.37
6		14.270	41836	0.93

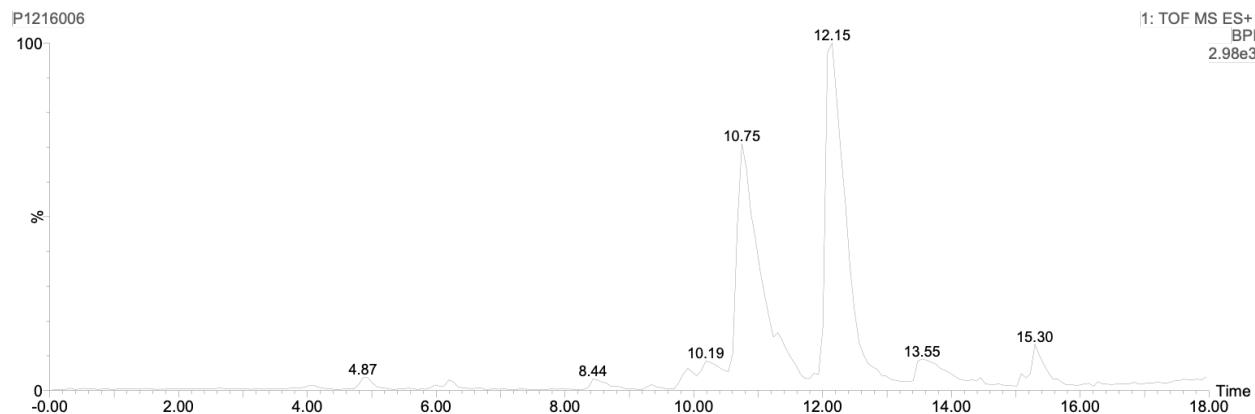


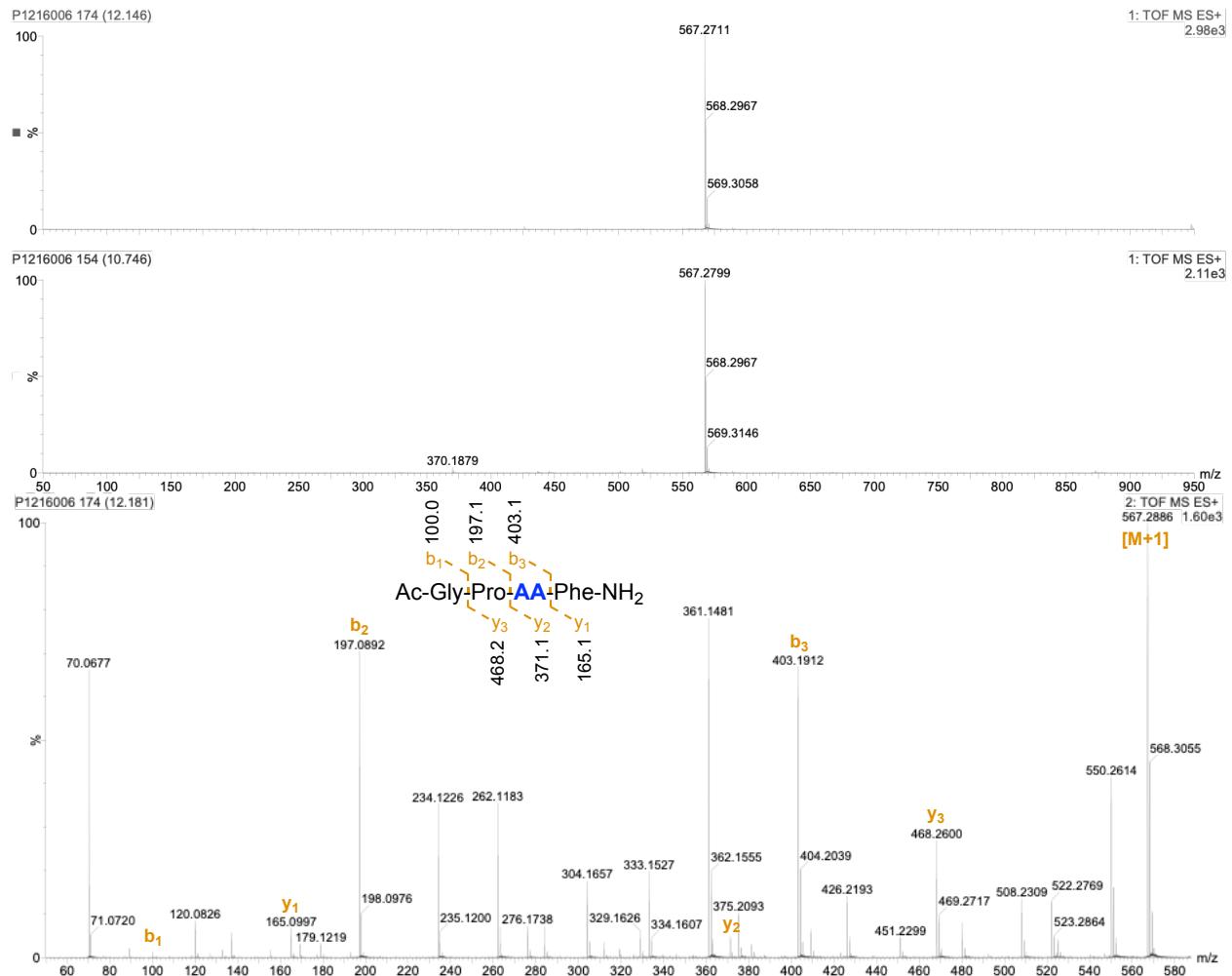


6A': MW = 566.7, Purity = 91.7%, Yield = 24.2% [0.32 mg]

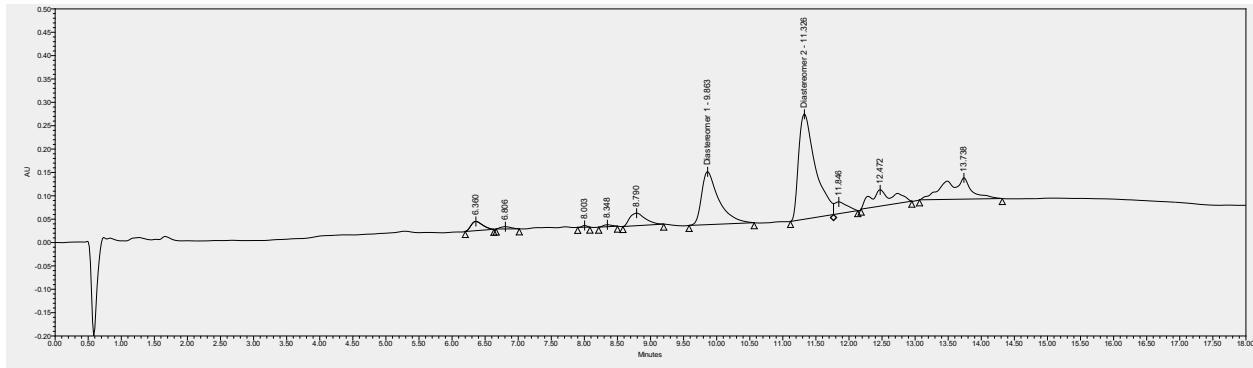
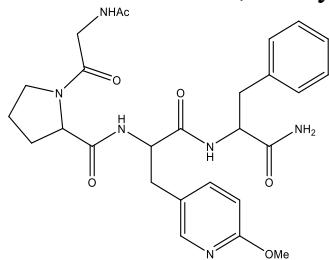


	Name	Retention Time	Area	% Area
1		10.084	547509	7.45
2		11.683	17756	0.24
3	Diastereomer 1	12.312	2935241	39.94
4	Diastereomer 2	13.039	3799736	51.71
5		13.945	48614	0.66

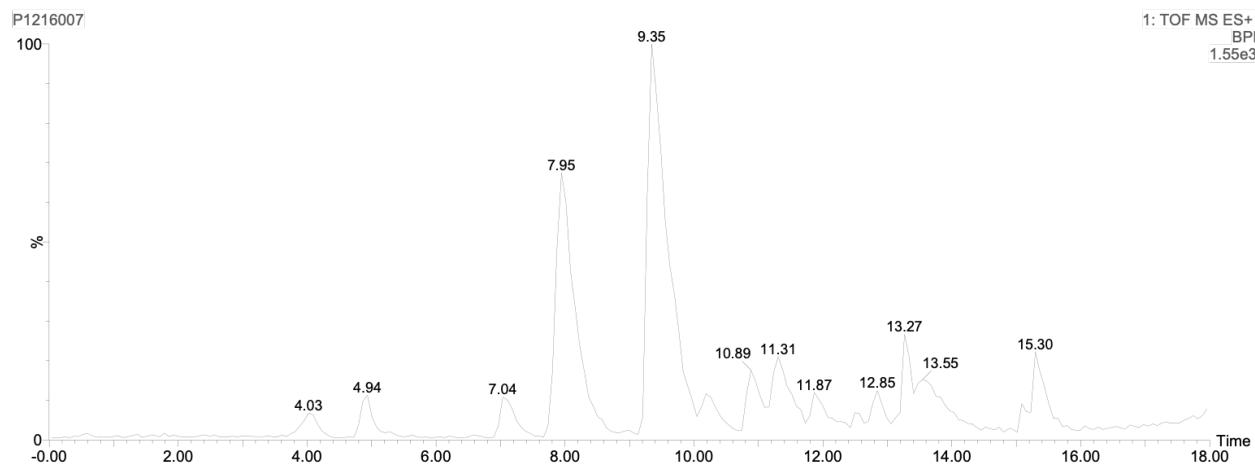


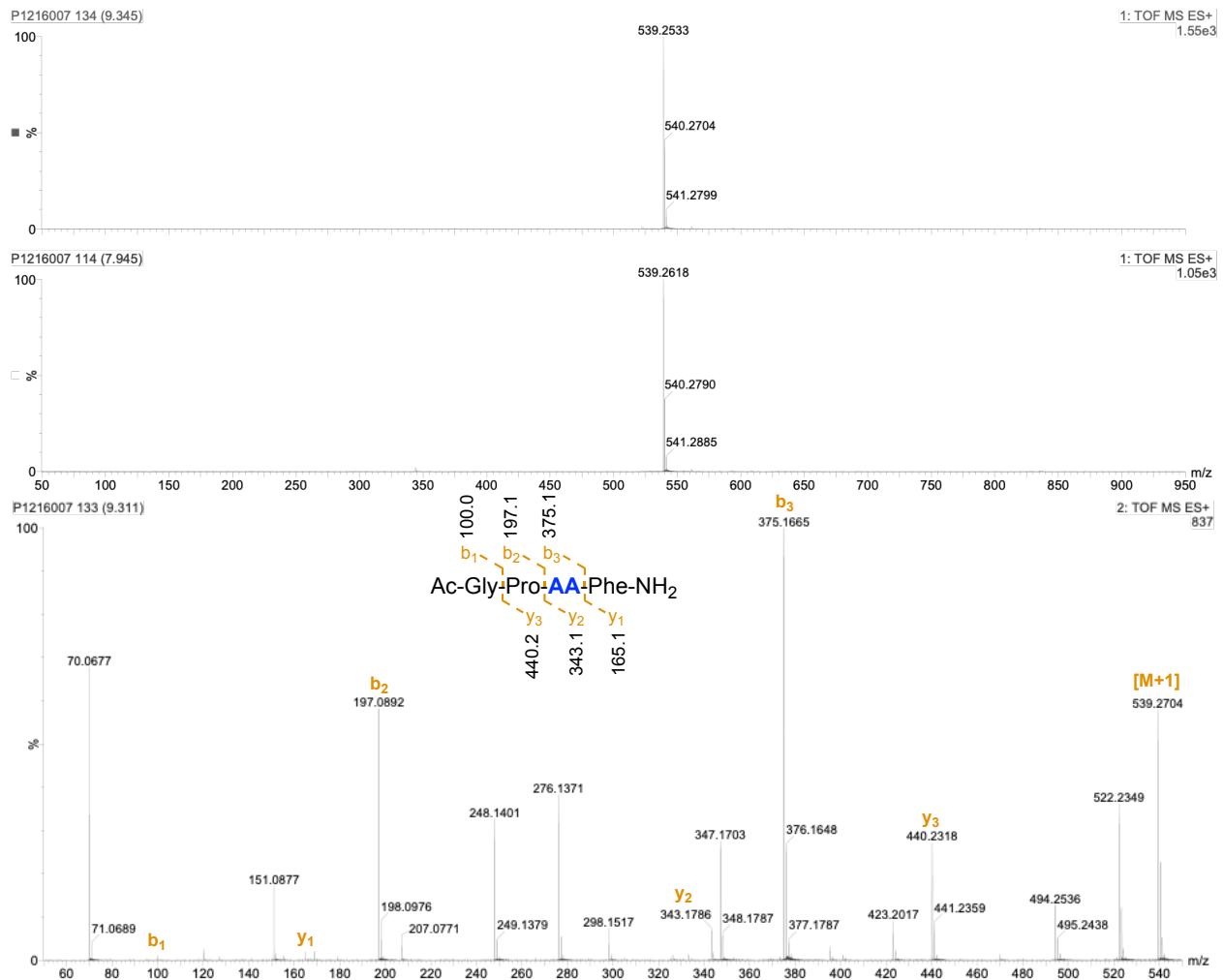


7A': MW = 538.6, Purity = 64.8% , Yield = 17.3% [0.22 mg]

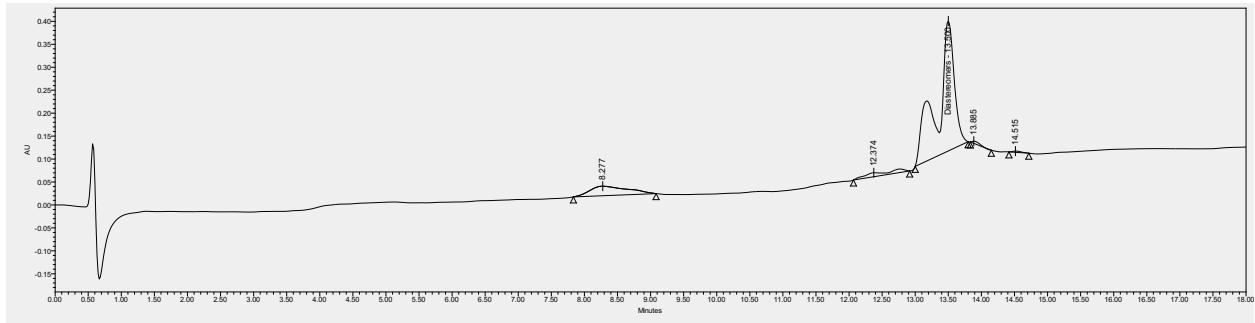
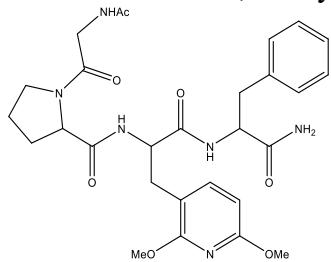


	Name	Retention Time	Area	% Area
1		6.360	243101	2.61
2		6.806	54944	0.59
3		8.003	19553	0.21
4		8.348	37482	0.40
5		8.790	441959	4.75
6	Diastereomer 1	9.863	2051406	22.05
7	Diastereomer 2	11.326	3976868	42.74
8		11.846	335546	3.61
9		12.472	839715	9.02
10		13.738	1304791	14.02

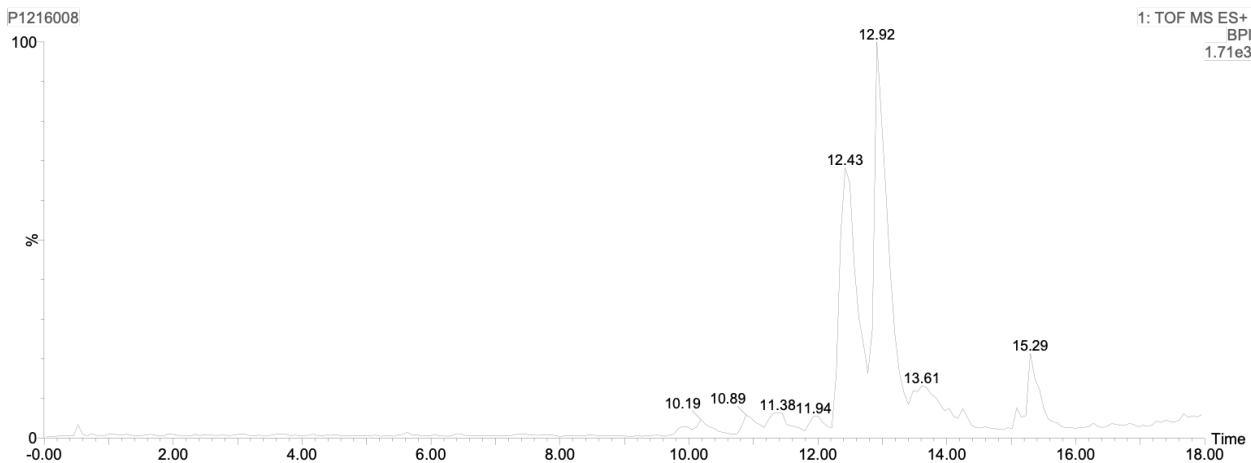


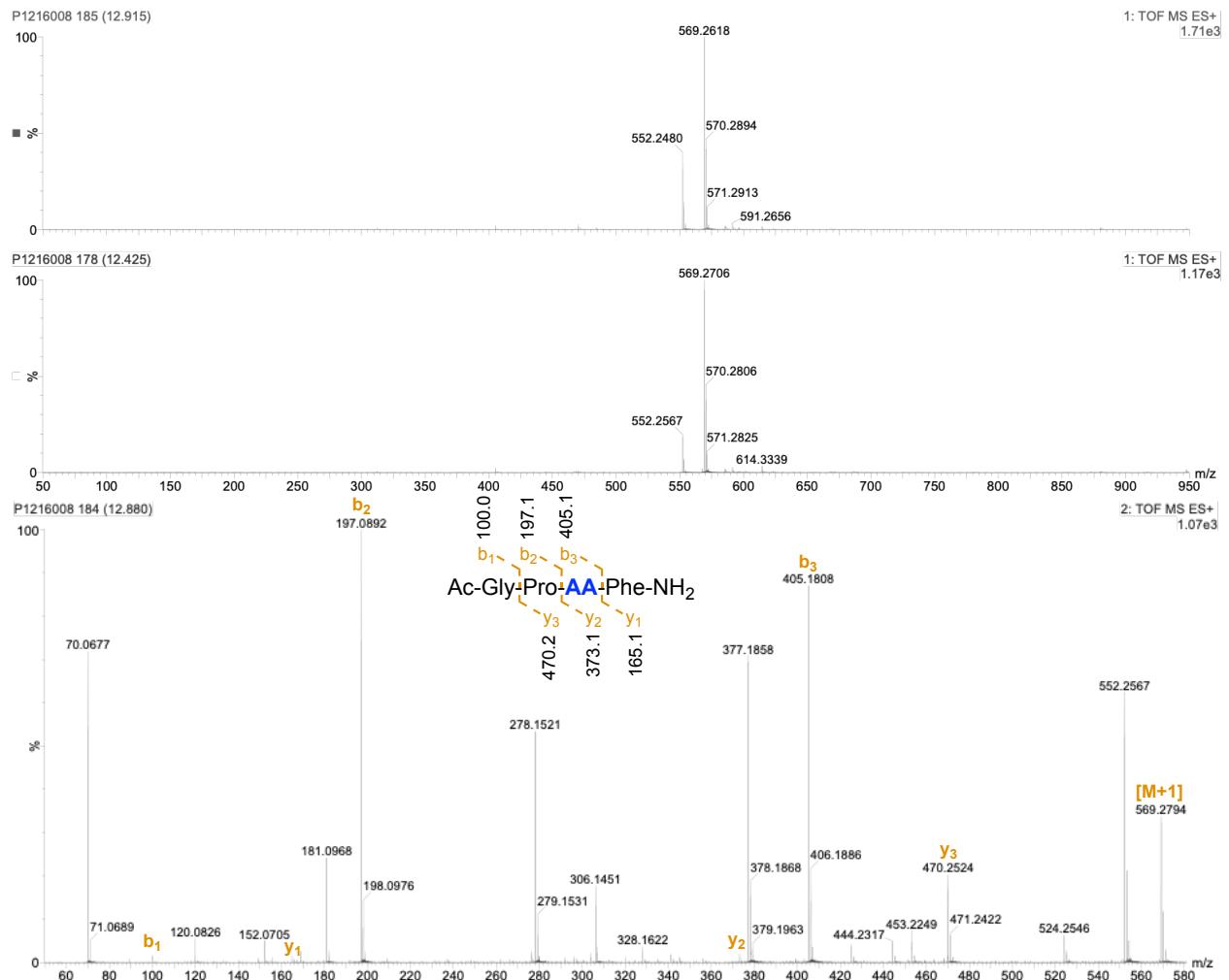


8A': MW = 568.6, Purity = 81.1%, Yield = 17.9% [0.24 mg]

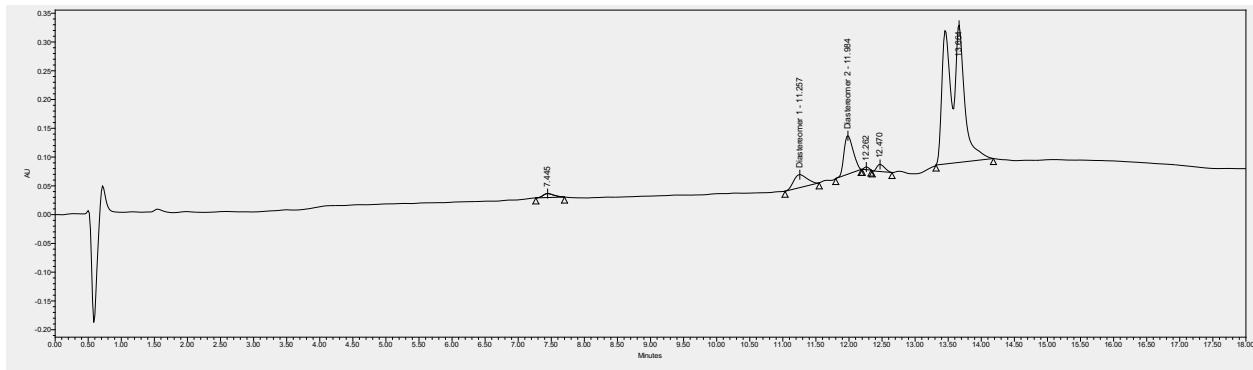
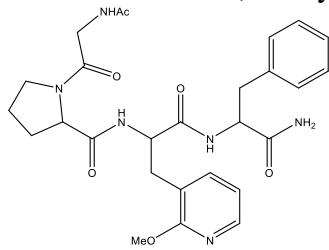


	Name	Retention Time	Area	% Area
1		8.277	818820	13.35
2		12.374	270674	4.41
3	Diastereomers	13.500	4974023	81.10
4		13.885	44795	0.73
5		14.515	24841	0.41

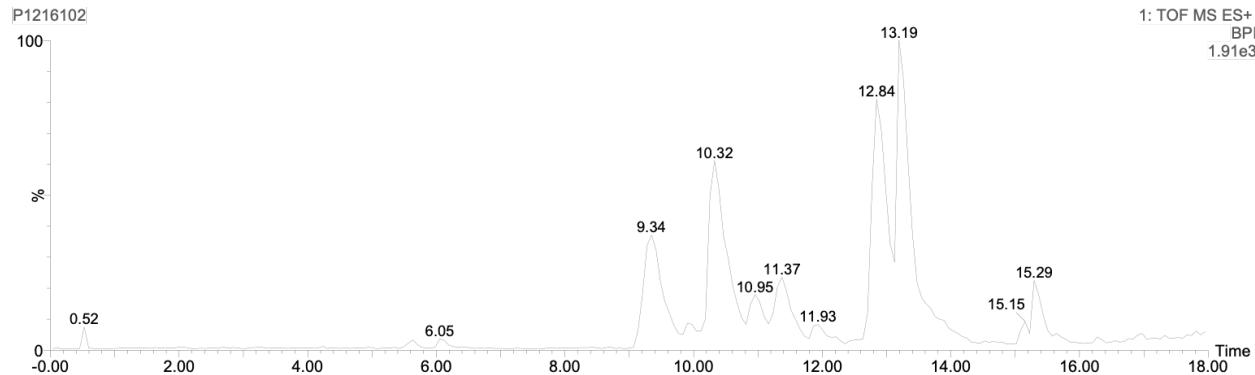


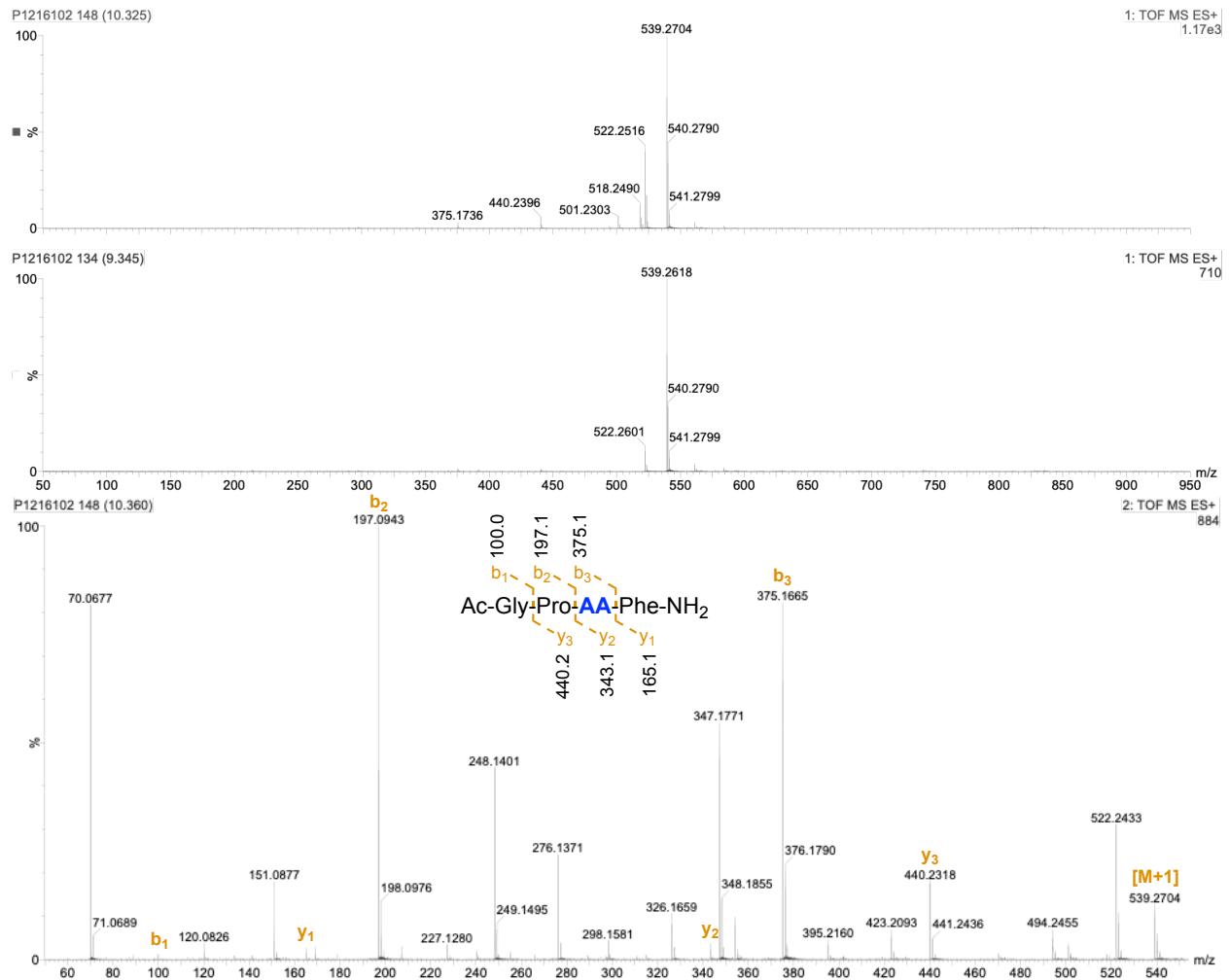


9A': MW = 538.6, Purity = 17.4%, Yield = 2.8% [0.035 mg]

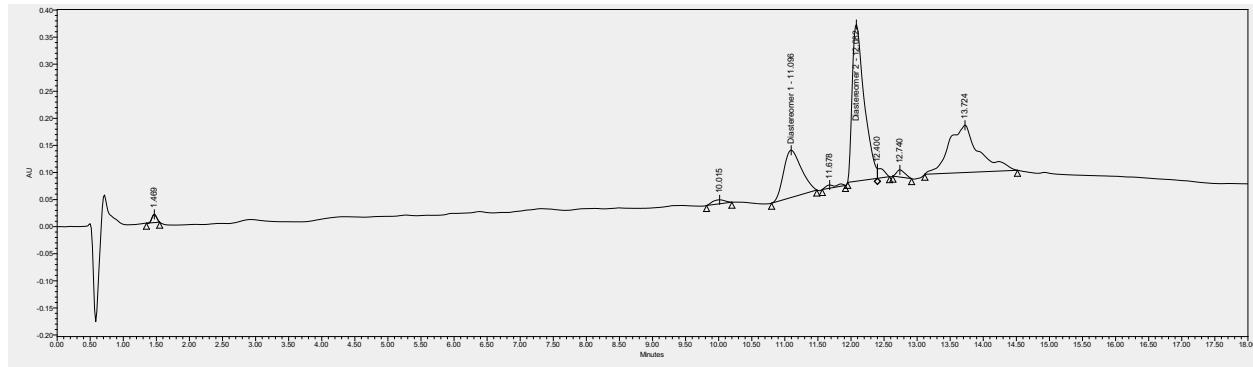
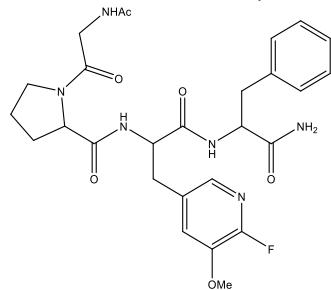


	Name	Retention Time	Area	% Area
1		7.445	79019	1.43
2	Diastereomer 1	11.257	320073	5.79
3	Diastereomer 2	11.984	639923	11.57
4		12.262	22771	0.41
5		12.470	100891	1.82
6		13.664	4367606	78.98

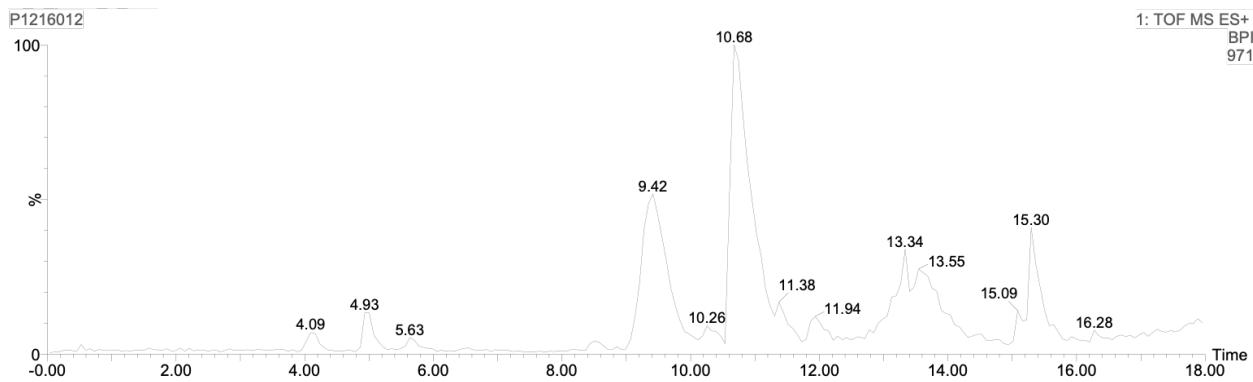


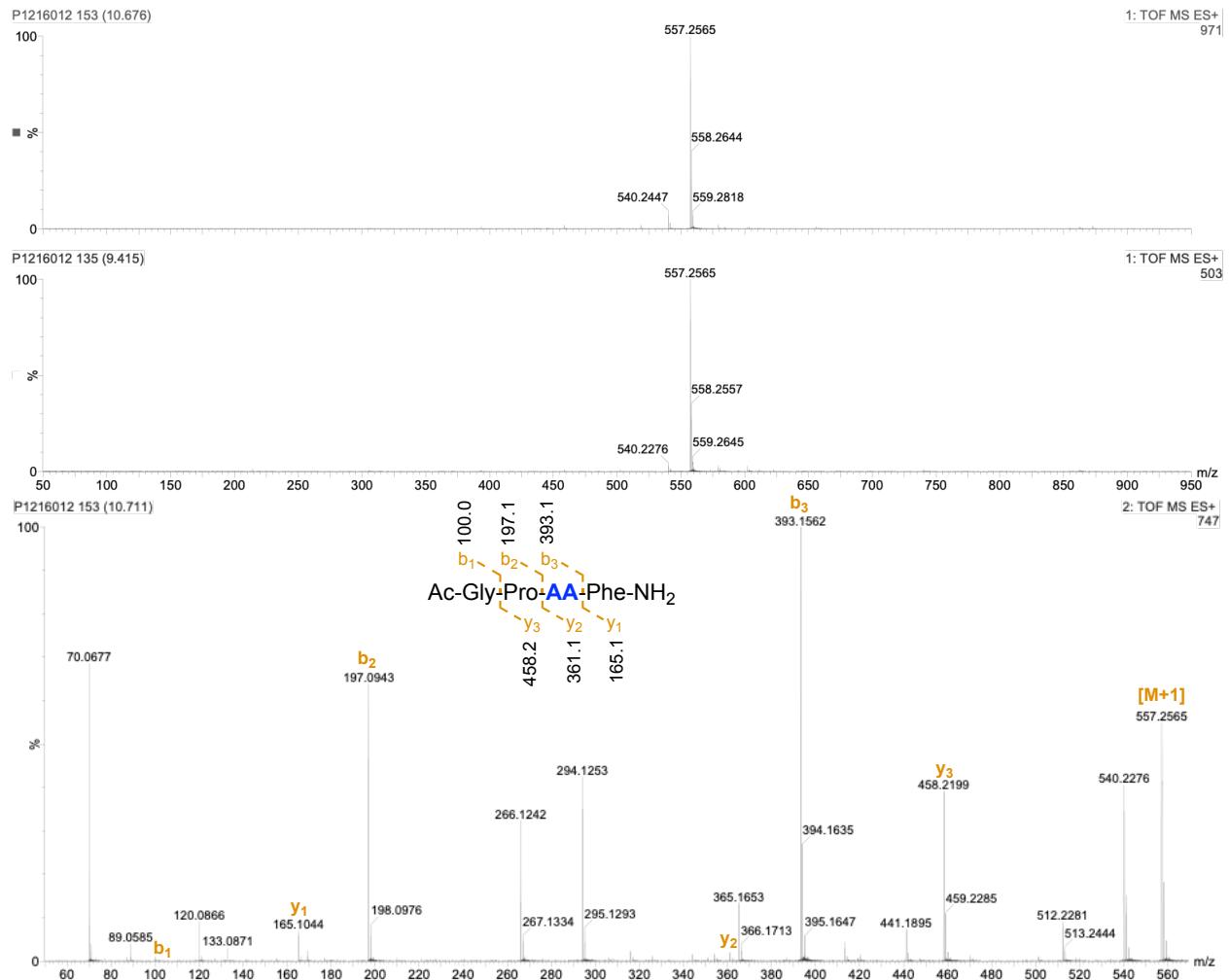


10A': MW = 556.6, Purity = 61.9%, Yield = 15.2% [0.20 mg]

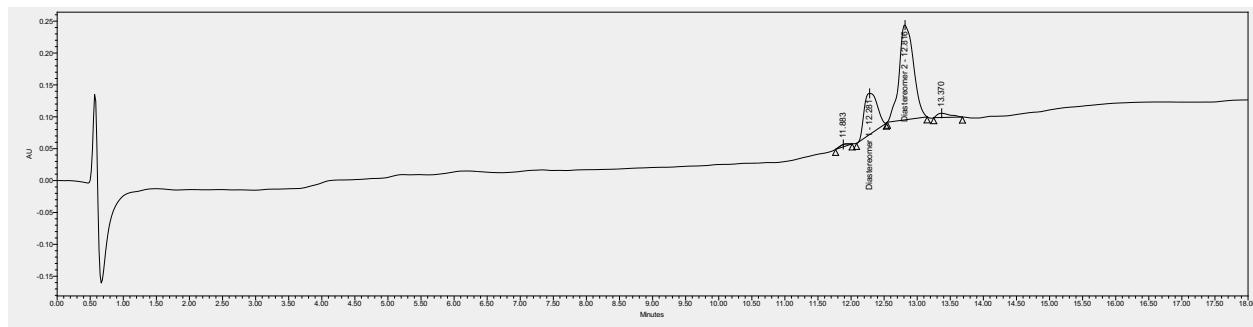
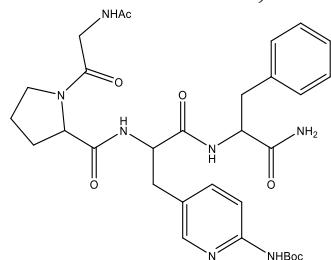


	Name	Retention Time	Area	% Area
1		1.469	75484	0.88
2		10.015	99685	1.17
3	Diastereomer 1	11.096	1629105	19.10
4		11.678	72677	0.85
5	Diastereomer 2	12.082	3650747	42.80
6		12.400	124123	1.46
7		12.740	116955	1.37
8		13.724	2761940	32.38

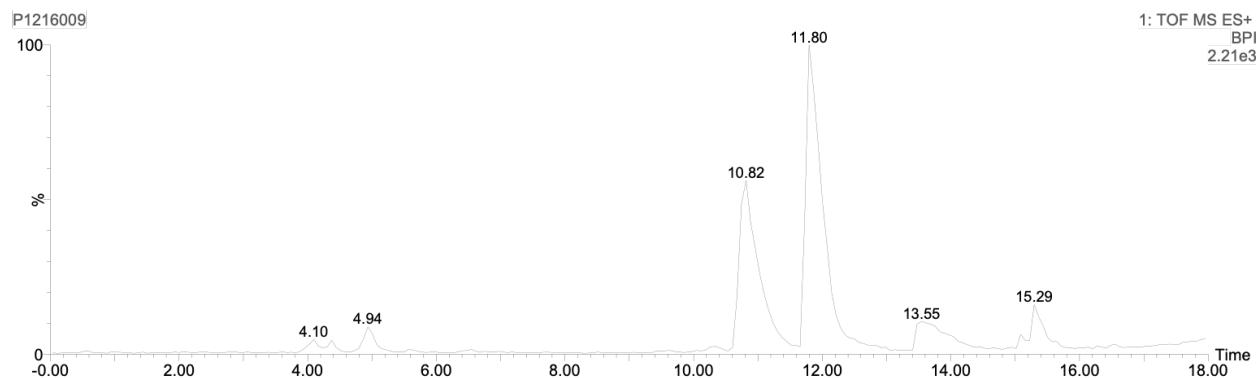


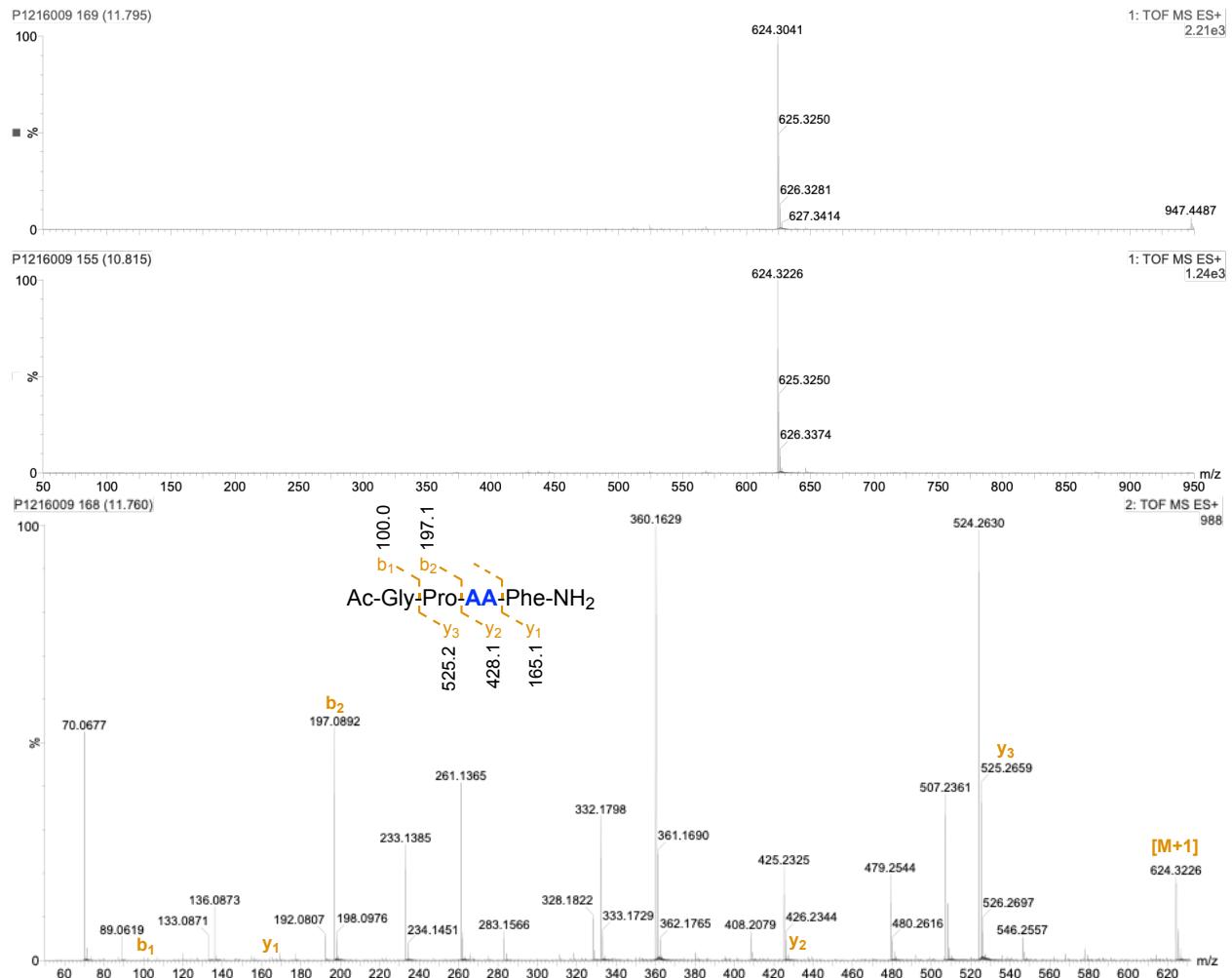


11A': MW = 623.7, Purity = 96.2%, Yield = 11.2% [0.16 mg]

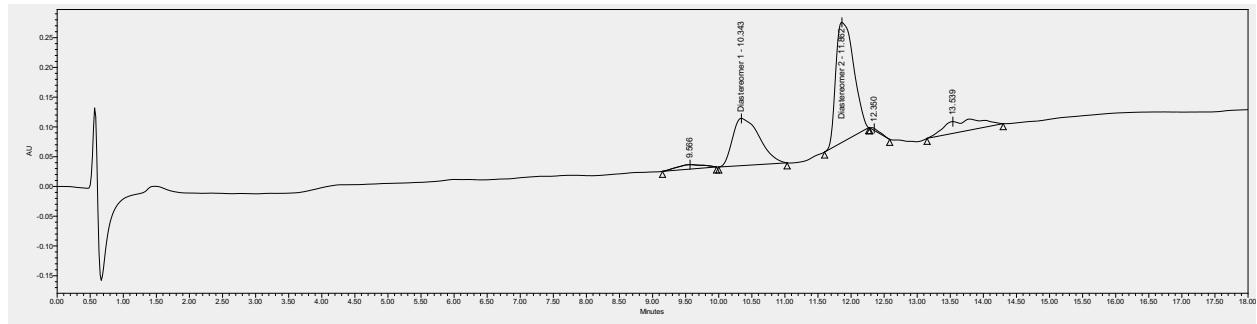
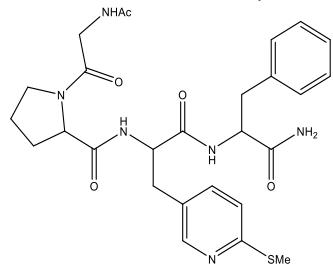


	Name	Retention Time	Area	% Area
1		11.883	32108	0.99
2	Diastereomer 1	12.281	882241	27.10
3	Diastereomer 2	12.816	2249611	69.10
4		13.370	91744	2.82

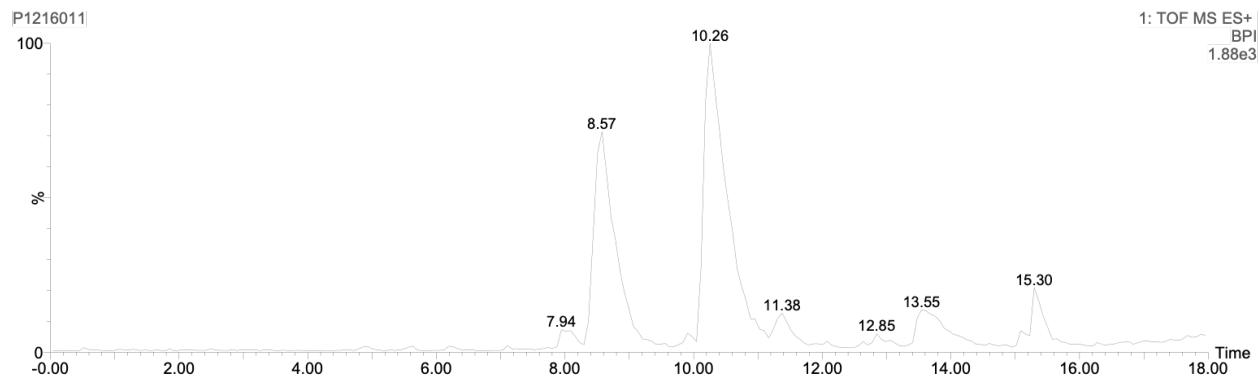


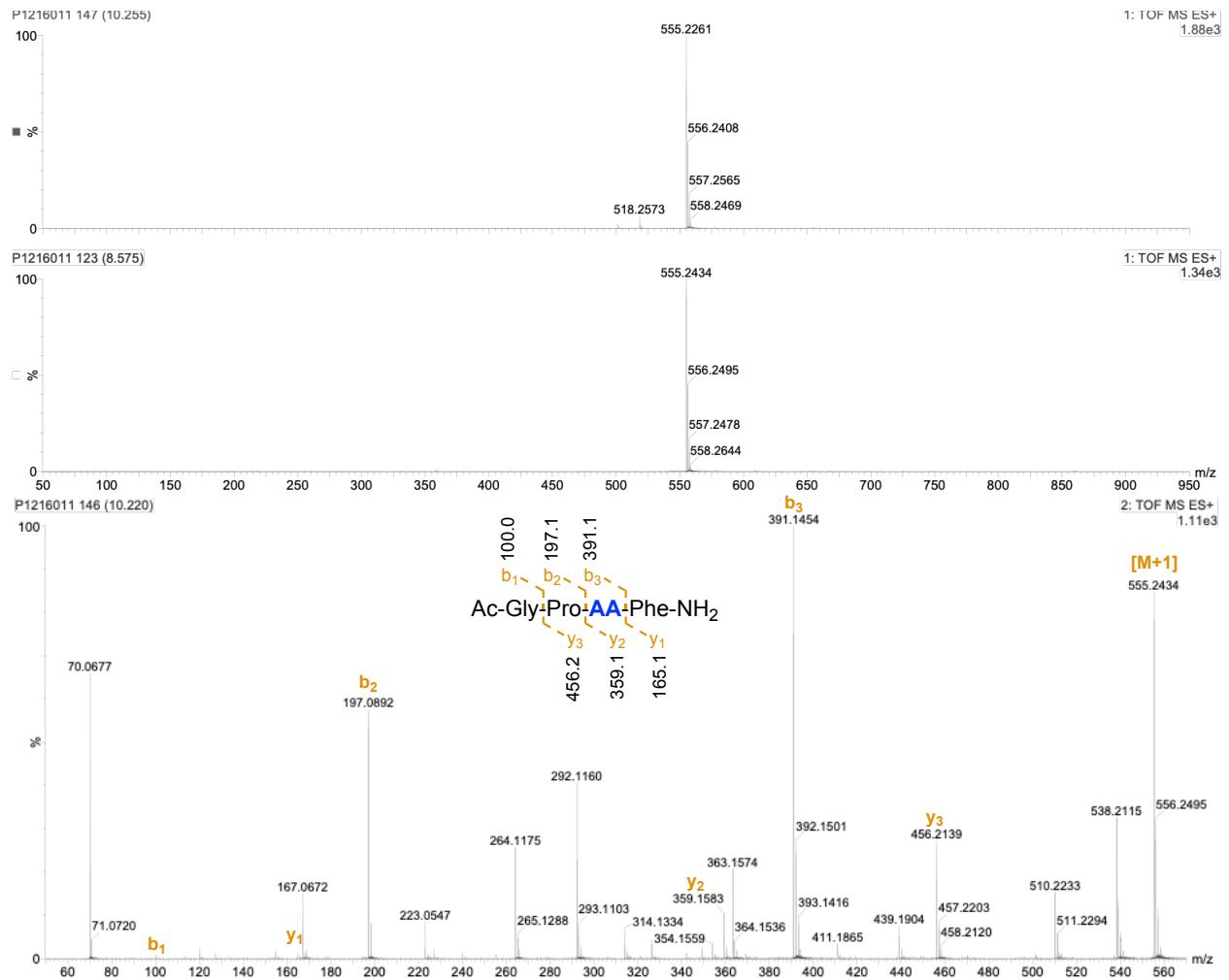


12A': MW = 554.7, Purity = 85.5% , Yield = 21.6% [0.28 mg]

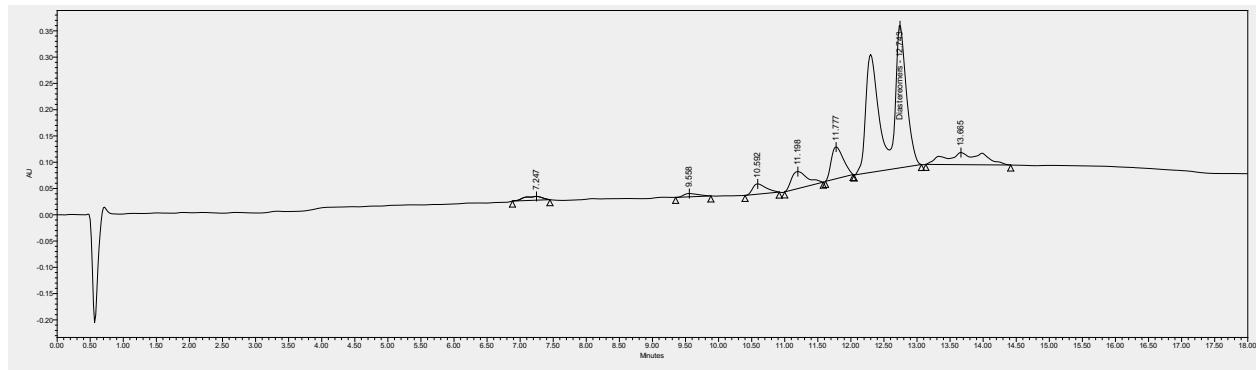
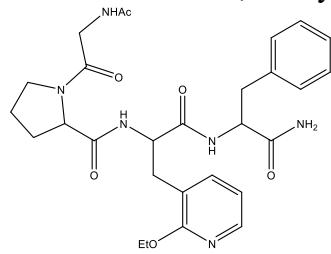


	Name	Retention Time	Area	% Area
1		9.566	210062	2.98
2	Diastereomer 1	10.343	2232302	31.70
3	Diastereomer 2	11.862	3789879	53.81
4		12.350	32135	0.46
5		13.539	778419	11.05

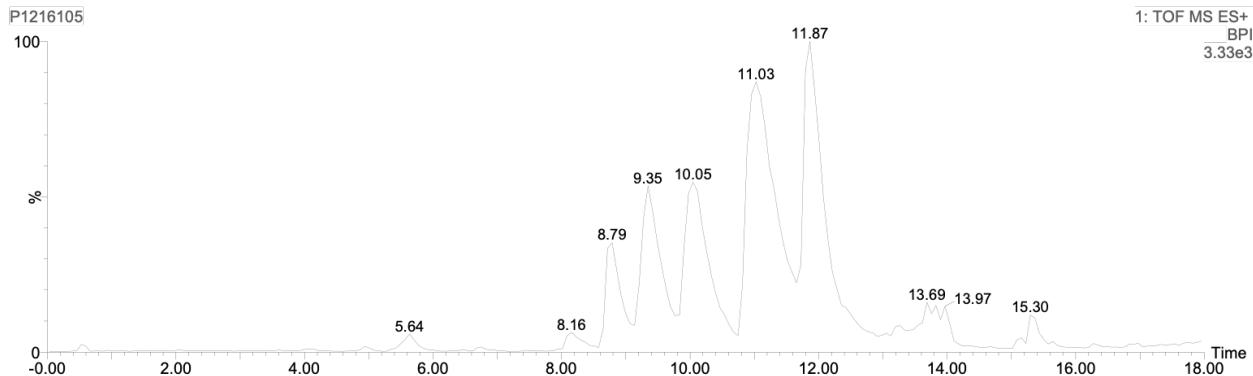


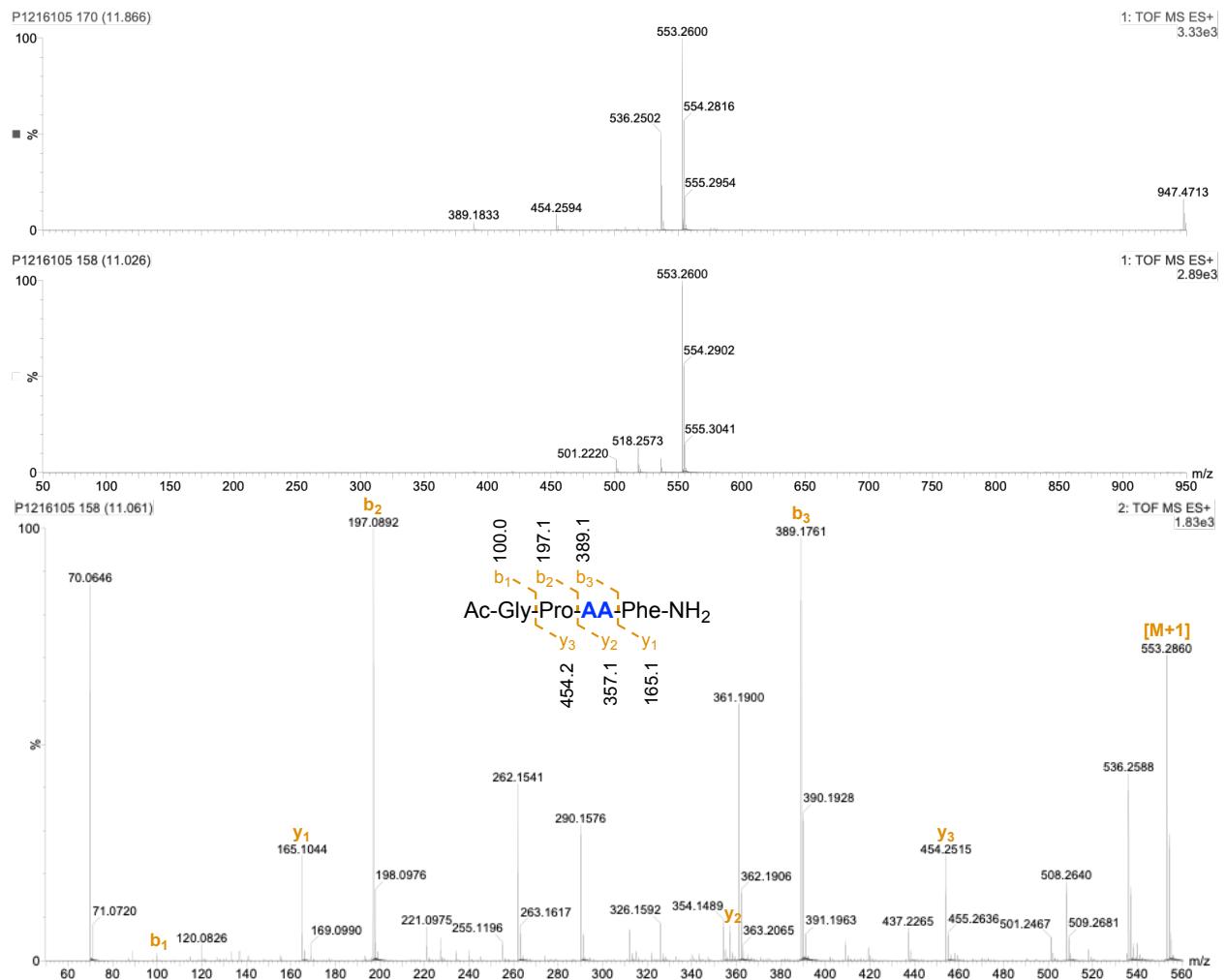


1B': MW = 552.6, Purity = 68.8%, Yield = 17.5% [0.23 mg]

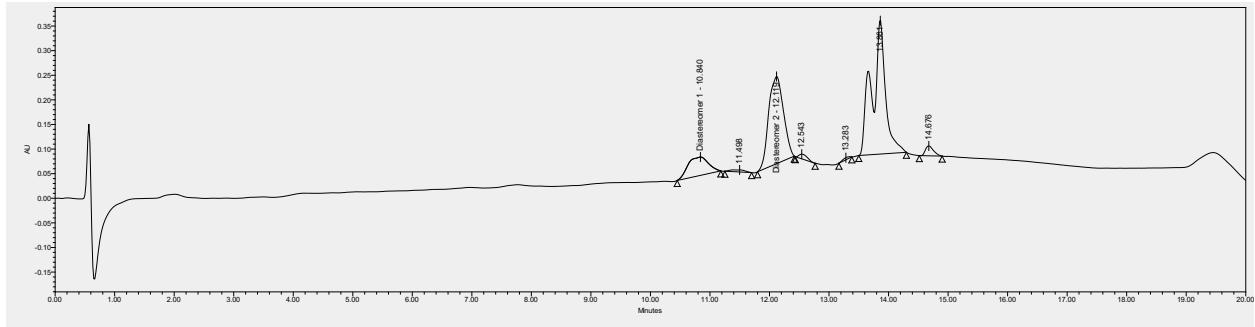
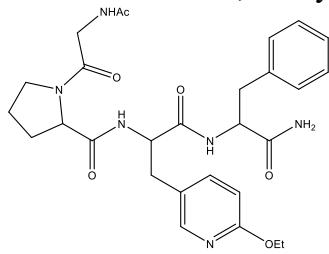


	Name	Retention Time	Area	% Area
1		7.247	139436	1.57
2		9.558	96400	1.09
3		10.592	277723	3.13
4		11.198	547354	6.18
5		11.777	765829	8.64
6	Diastereomers	12.743	6100197	68.83
7		13.665	935889	10.56

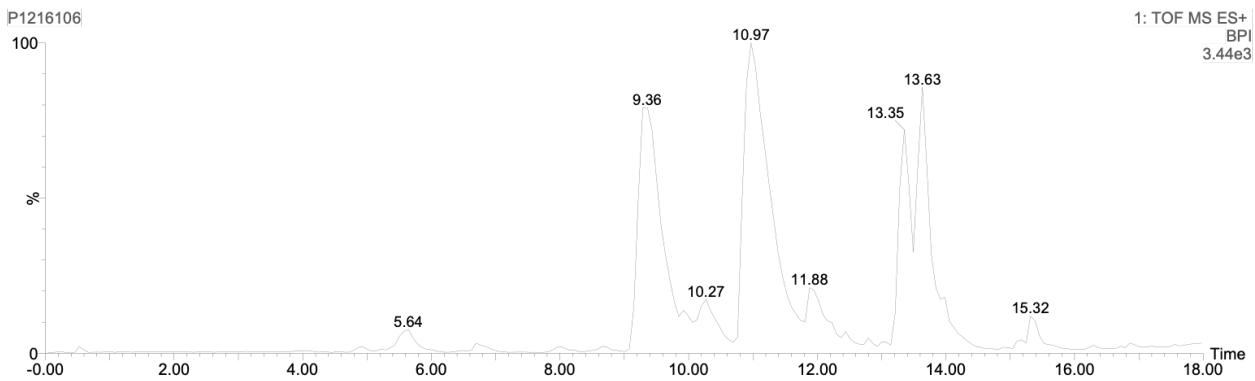


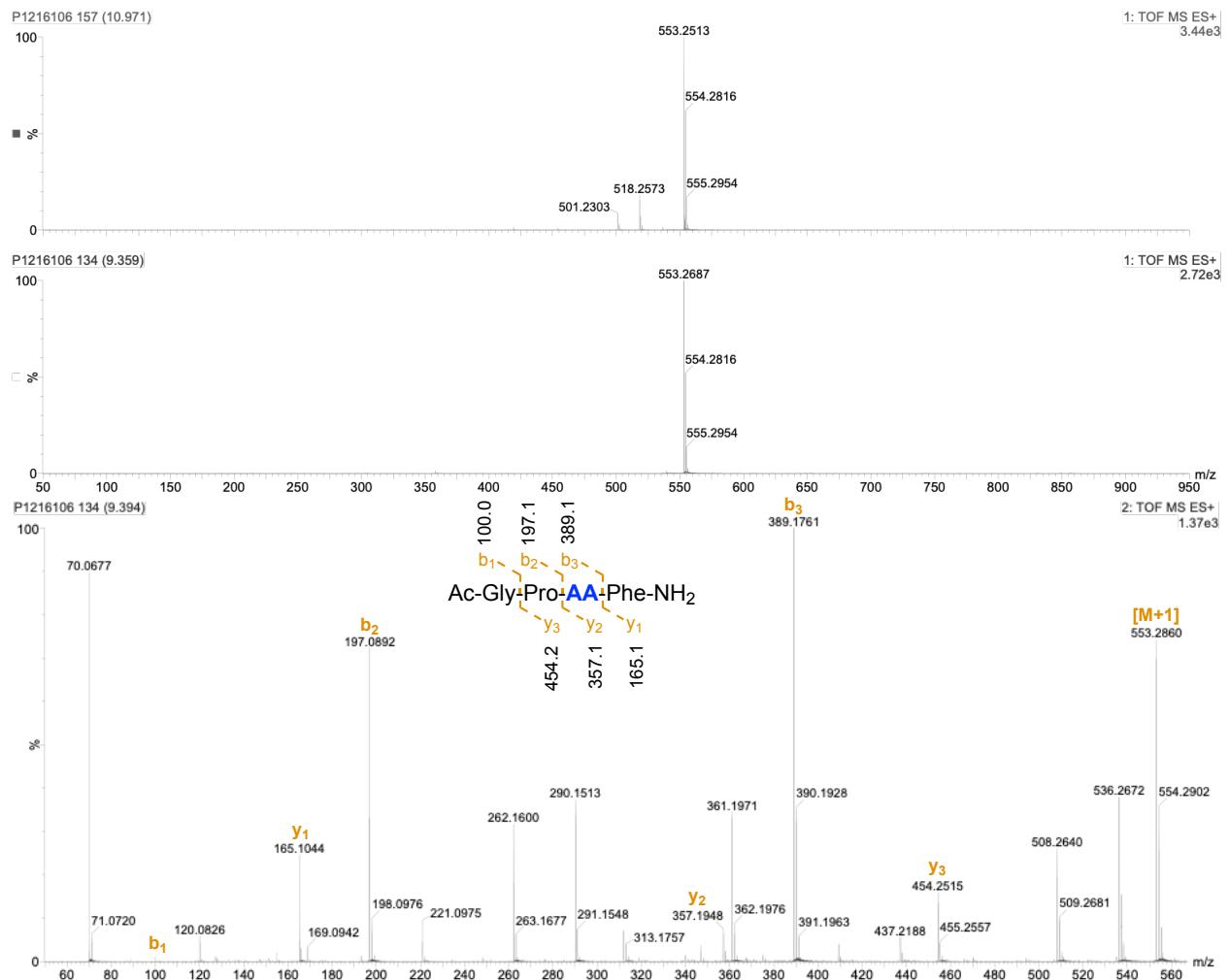


2B': MW = 552.6, Purity = 46.1%, Yield = 14.3% [0.18 mg]

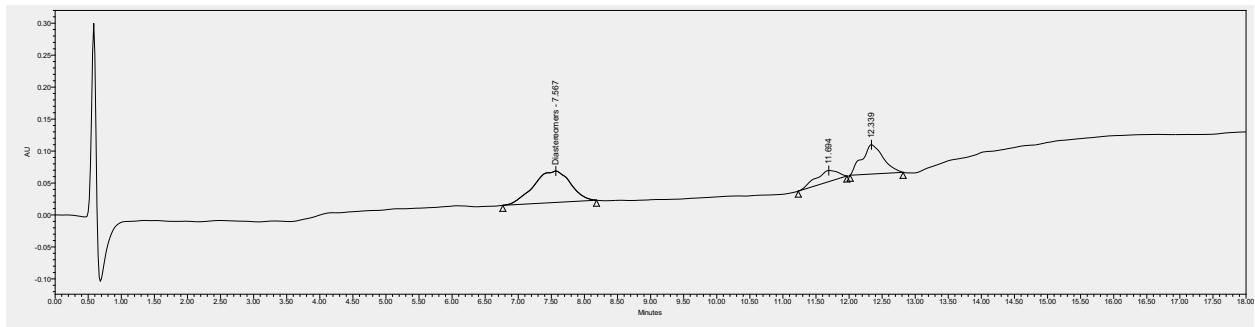
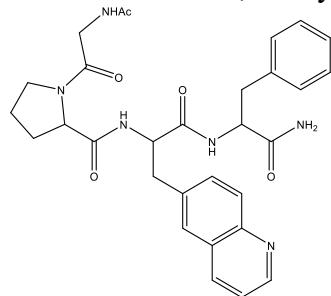


	Name	Retention Time	Area	% Area
1	Diastereomer 1	10.840	915908	10.63
2		11.498	69924	0.81
3	Diastereomer 2	12.119	3055292	35.45
4		12.543	95775	1.11
5		13.283	34527	0.40
6		13.861	4257742	49.40
7		14.676	190406	2.21

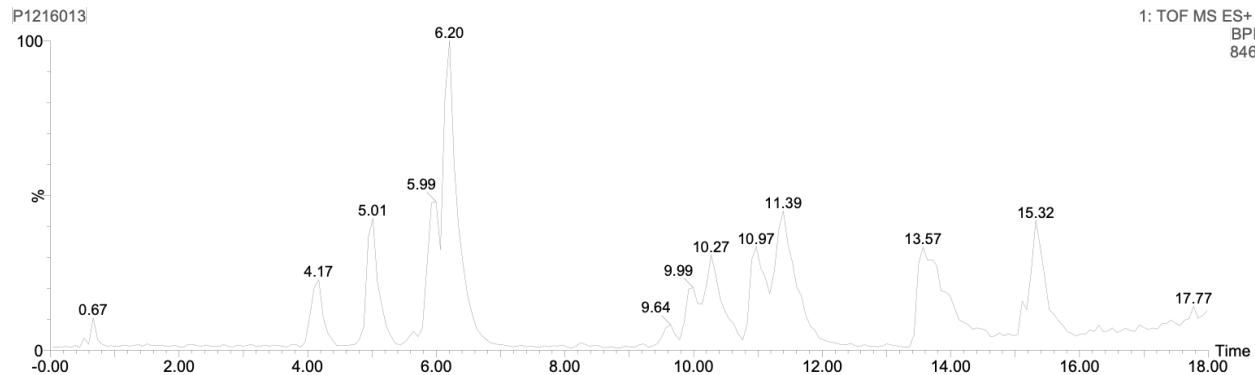


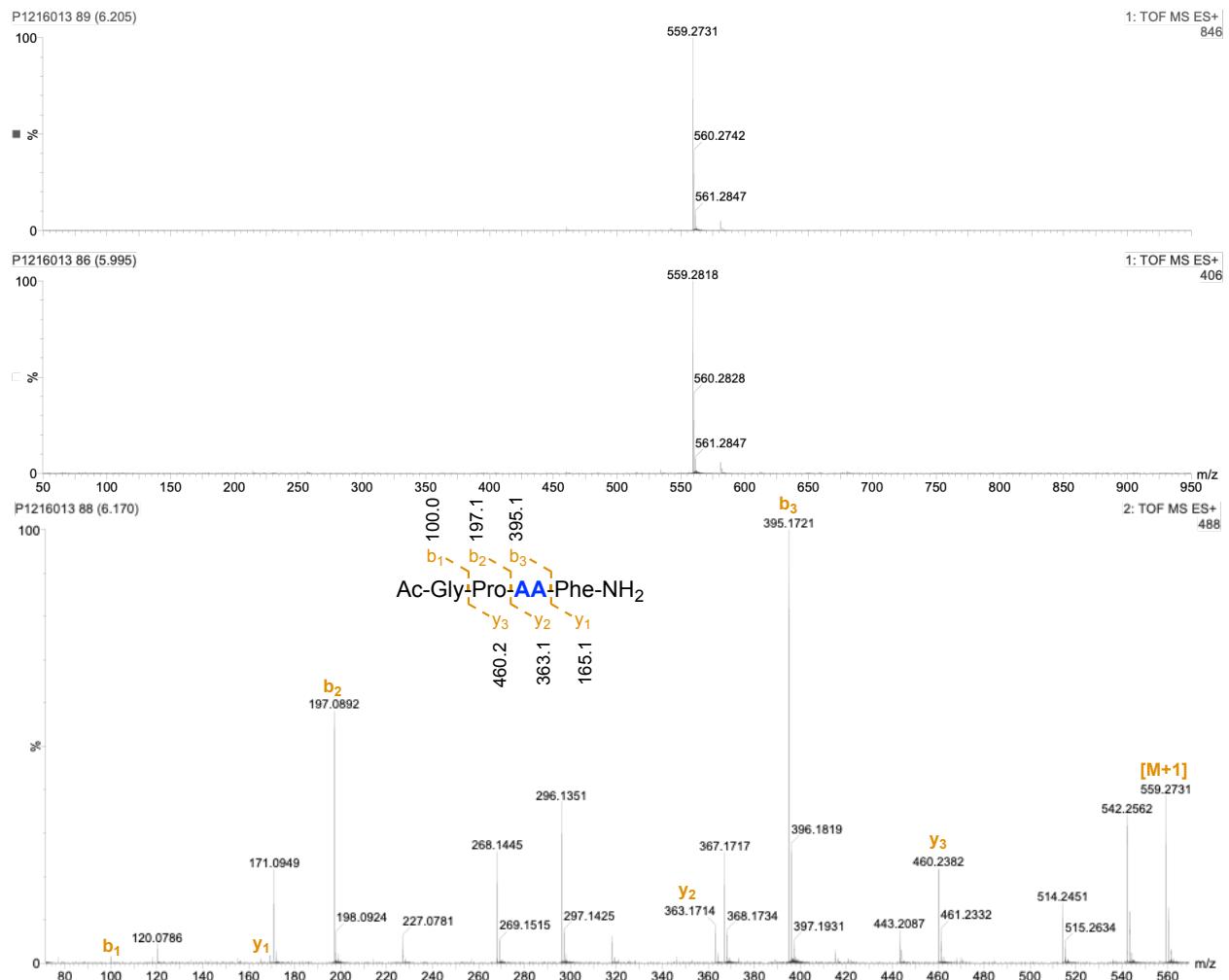


3B': MW = 558.6, Purity = 56.9%, Yield = 3.3% [0.043 mg]

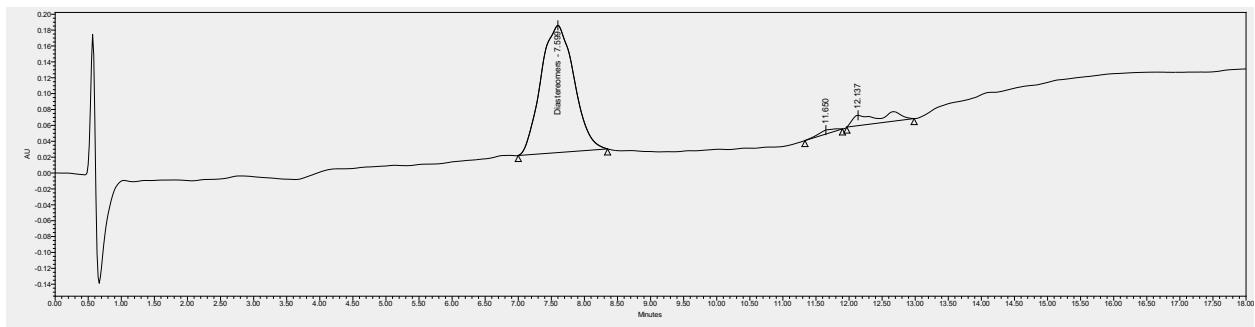
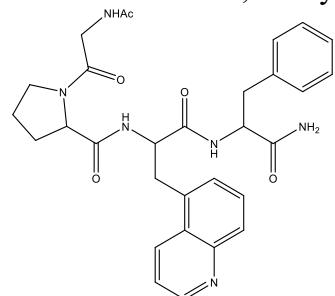


	Name	Retention Time	Area	% Area
1	Diastereomers	7.567	1905181	56.87
2		11.694	417853	12.47
3		12.339	1026994	30.66

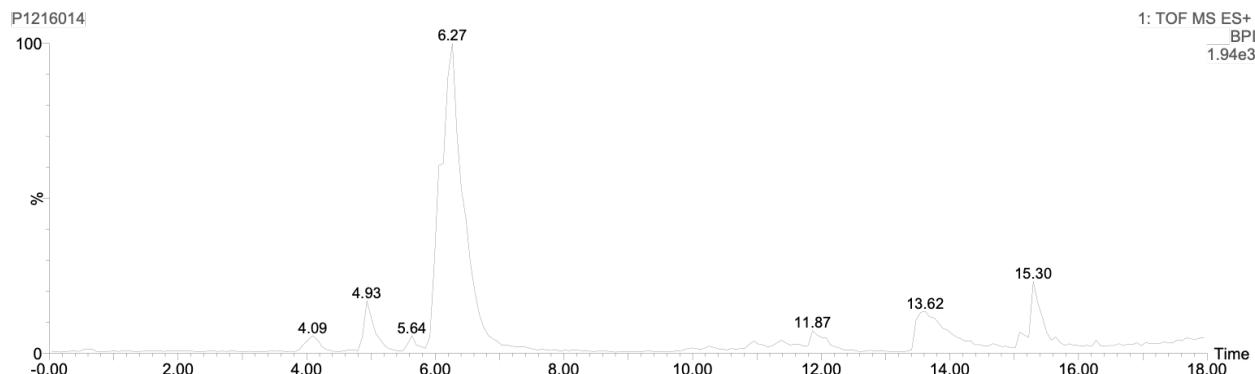


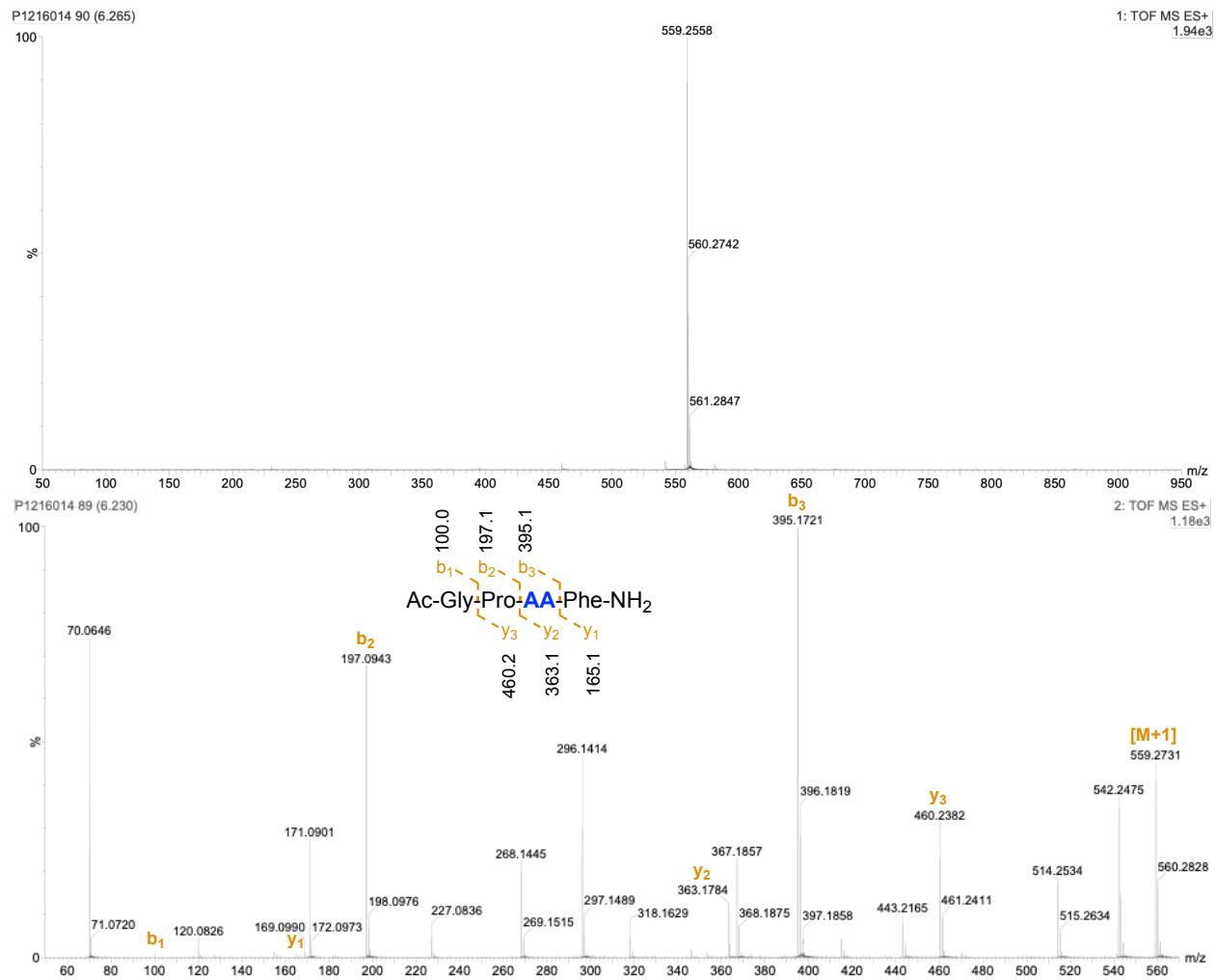


4B': MW = 558.6, Purity = 91.1%, Yield = 9.7% [0.13 mg]

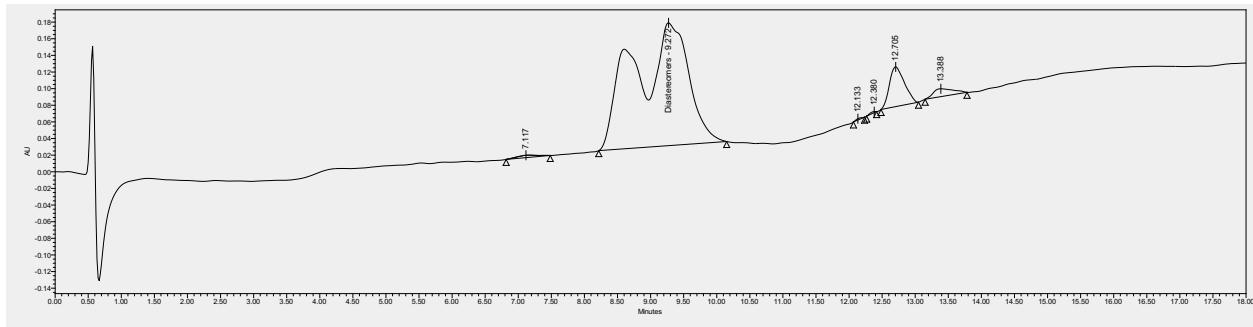
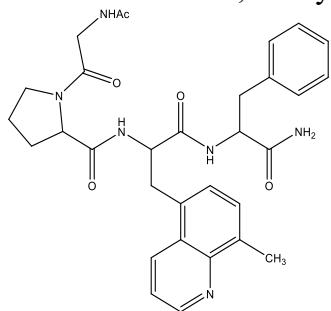


	Name	Retention Time	Area	% Area
1	Diastereomers	7.599	5585025	91.09
2		11.650	86251	1.41
3		12.137	460252	7.51

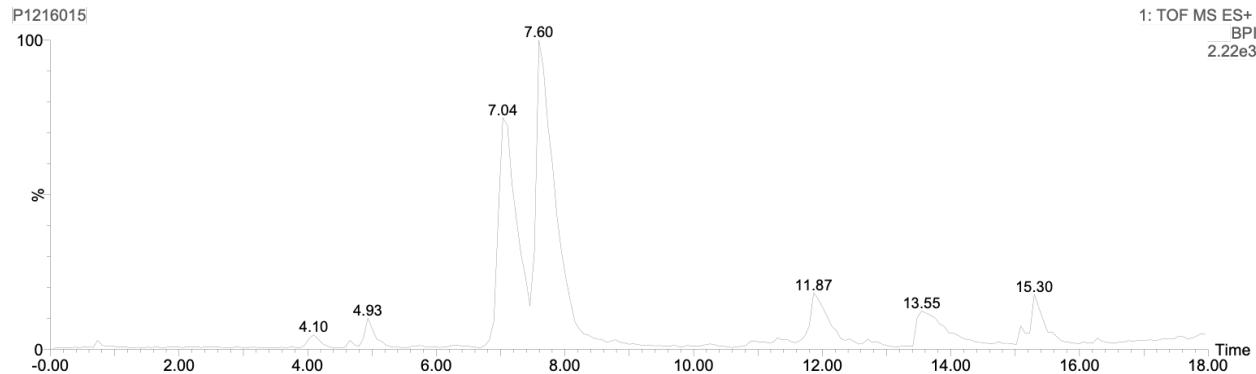


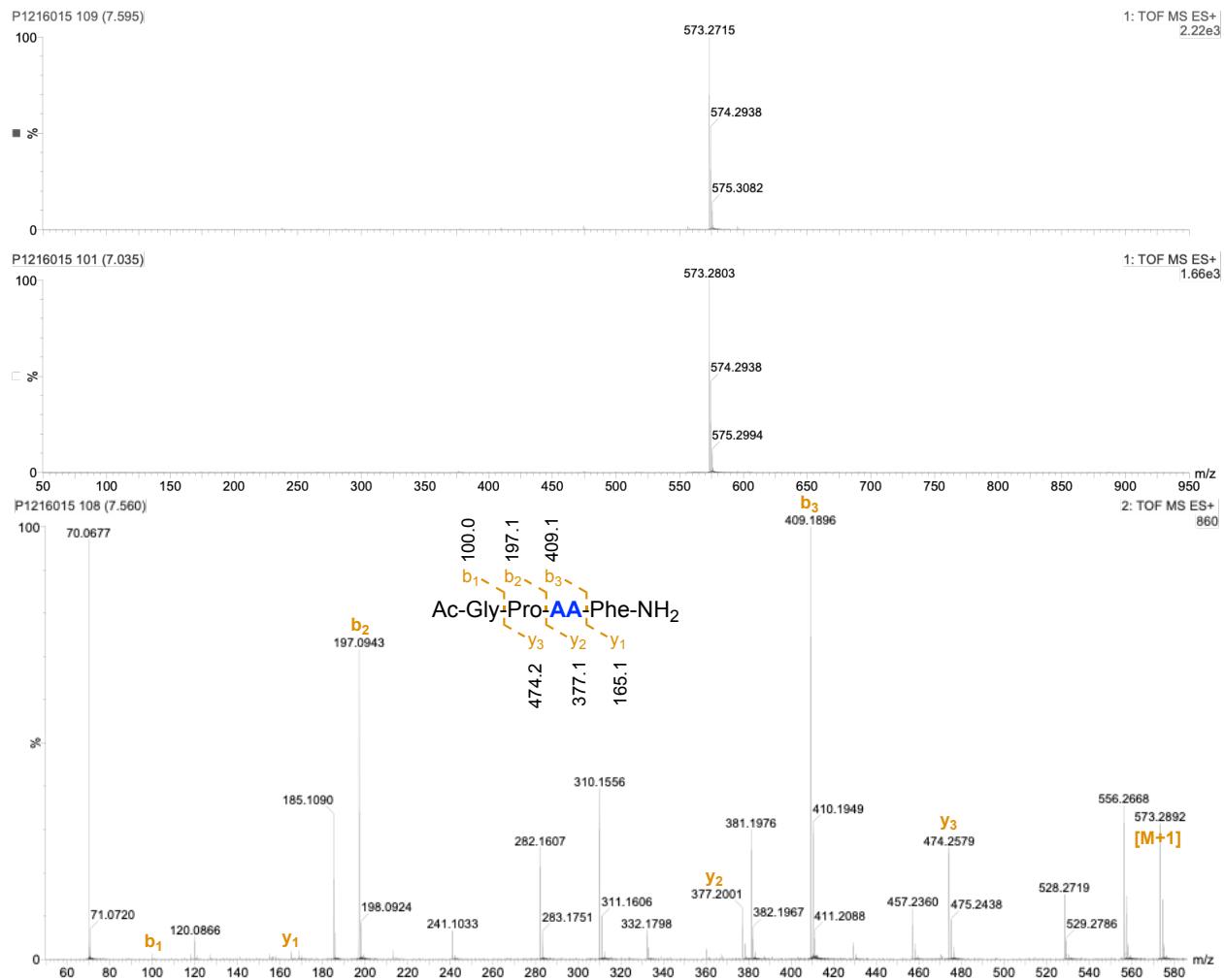


5B': MW = 572.7, Purity = 89.0%, Yield = 14.4% [0.19 mg]

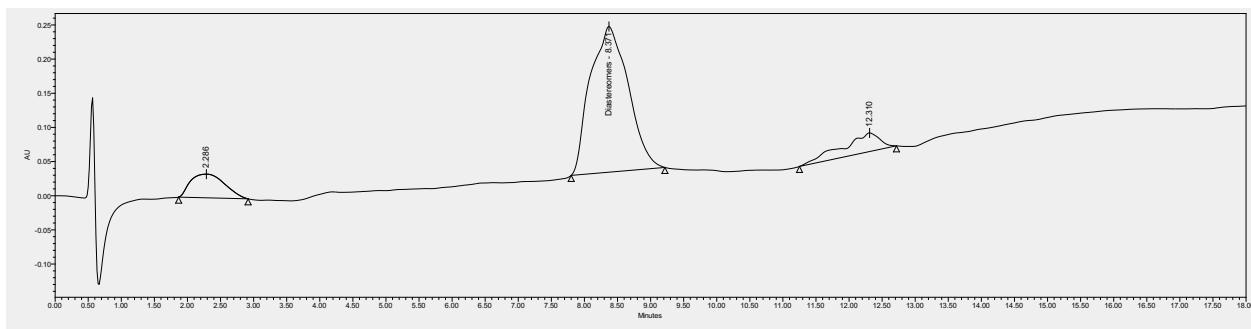
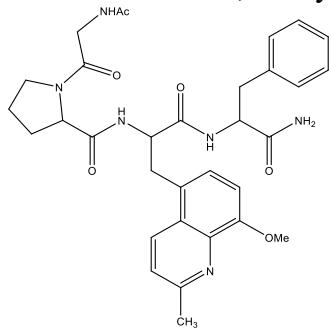


	Name	Retention Time	Area	% Area
1		7.117	59826	0.64
2	Diastereomers	9.272	8307687	88.95
3		12.133	9705	0.10
4		12.380	10083	0.11
5		12.705	754383	8.08
6		13.388	197753	2.12

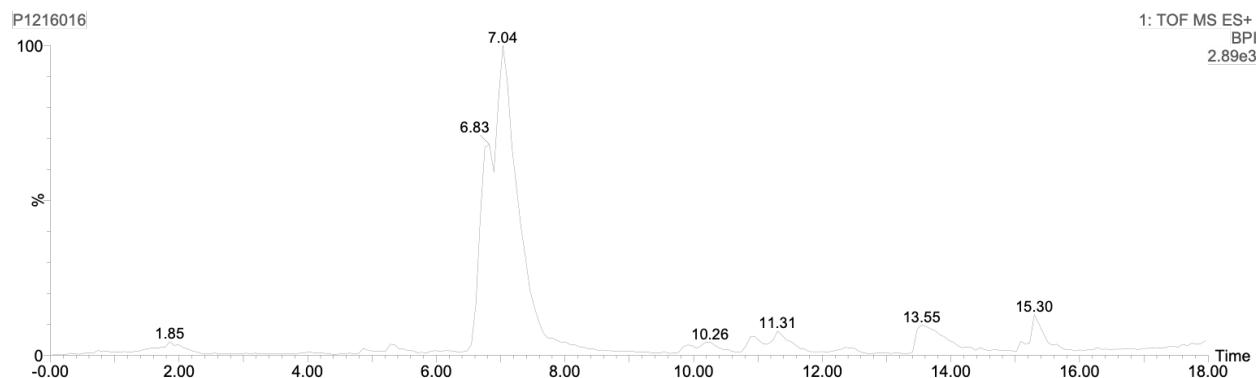


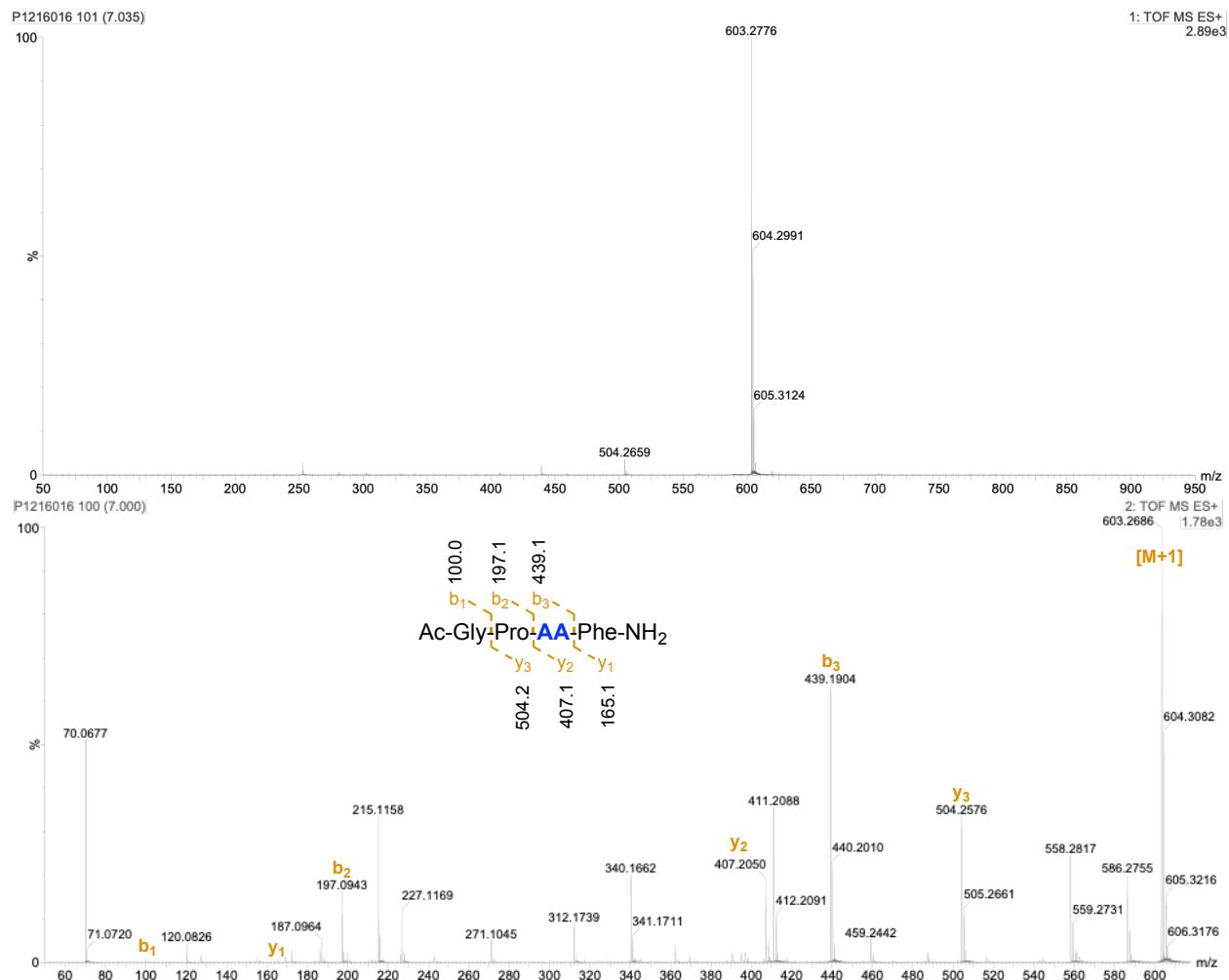


6B': MW = 602.7, Purity = 78.2%, Yield = 14.8% [0.21 mg]

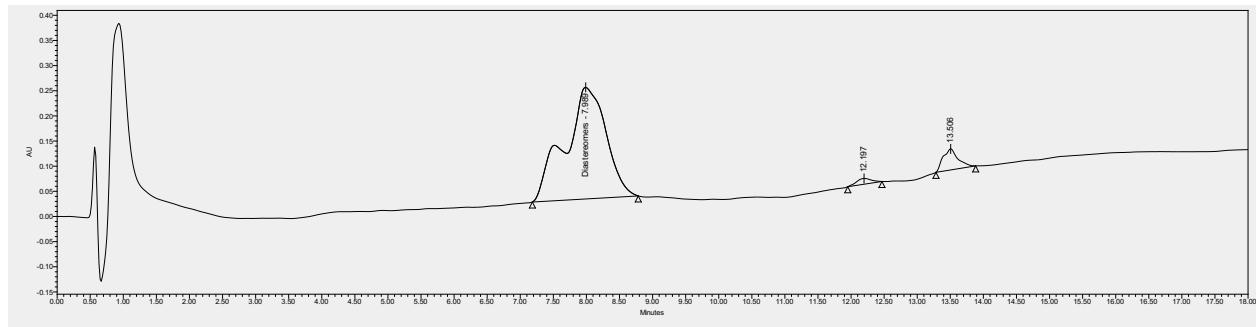
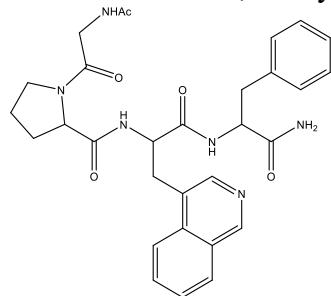


	Name	Retention Time	Area	% Area
1		2.286	1244568	11.45
2	Diastereomers	8.371	8500843	78.24
3		12.310	1119486	10.30

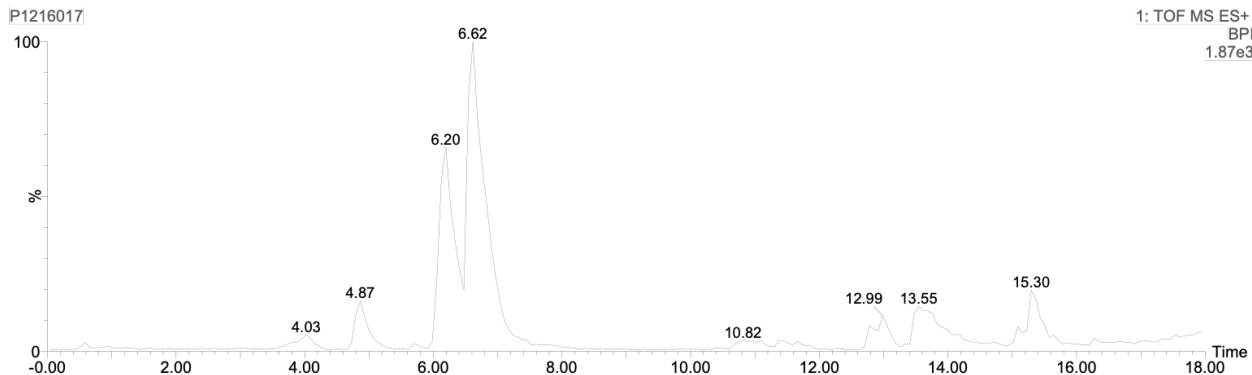


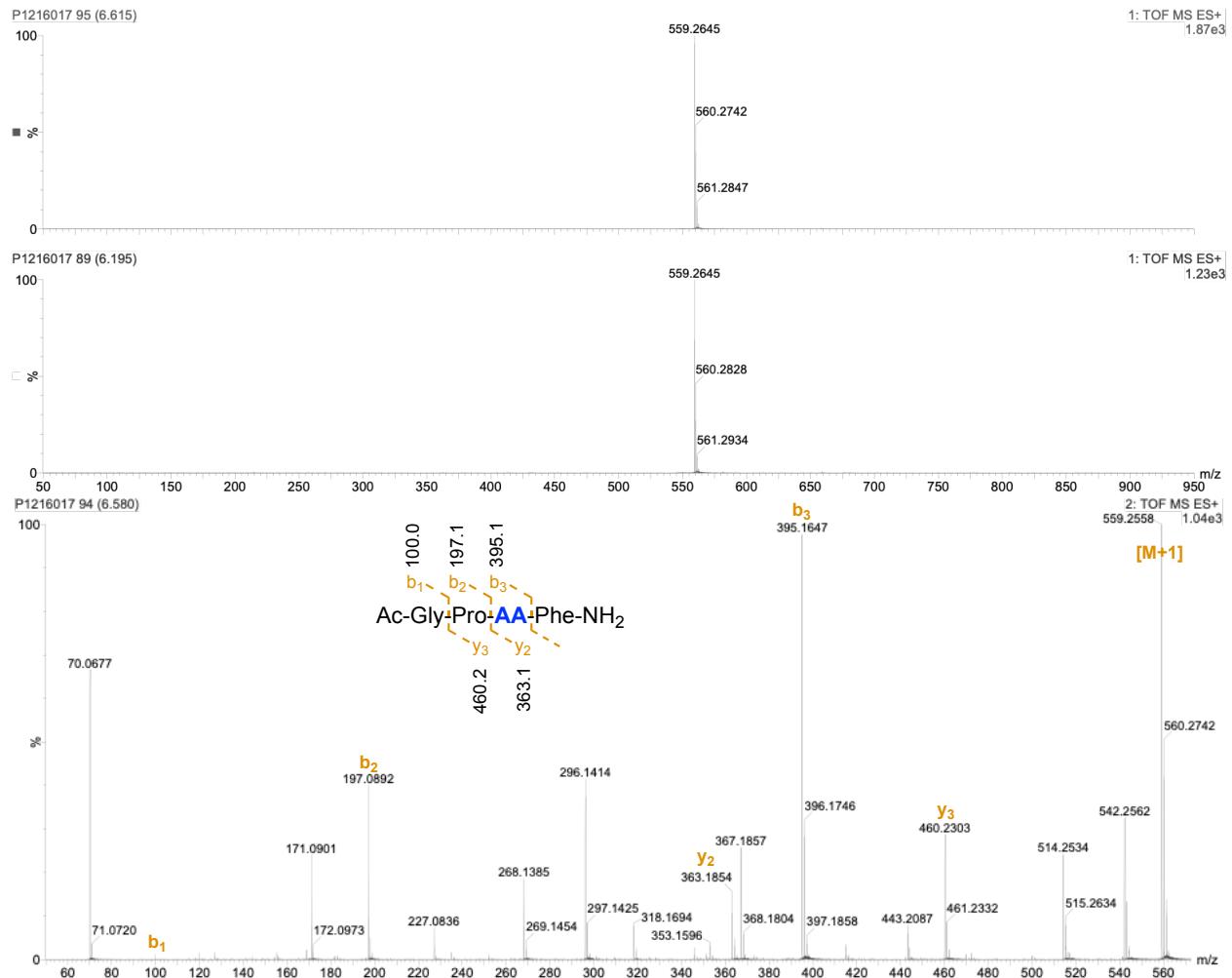


7B': MW = 558.6, Purity = 92.0%, Yield = 16.7% [0.22 mg]

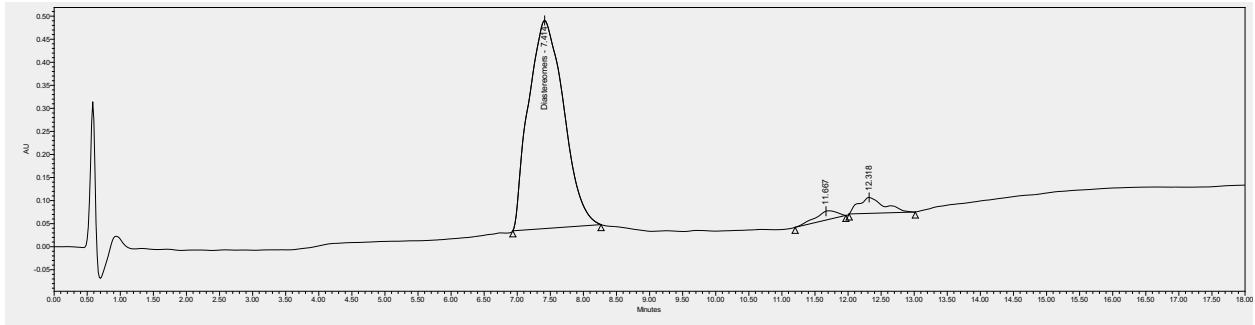
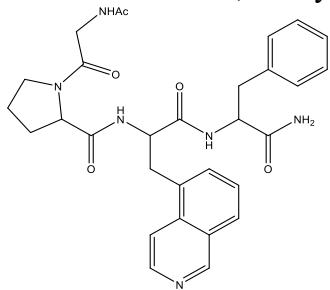


	Name	Retention Time	Area	% Area
1	Diastereomers	7.989	9599552	91.98
2		12.197	179405	1.72
3		13.506	657199	6.30

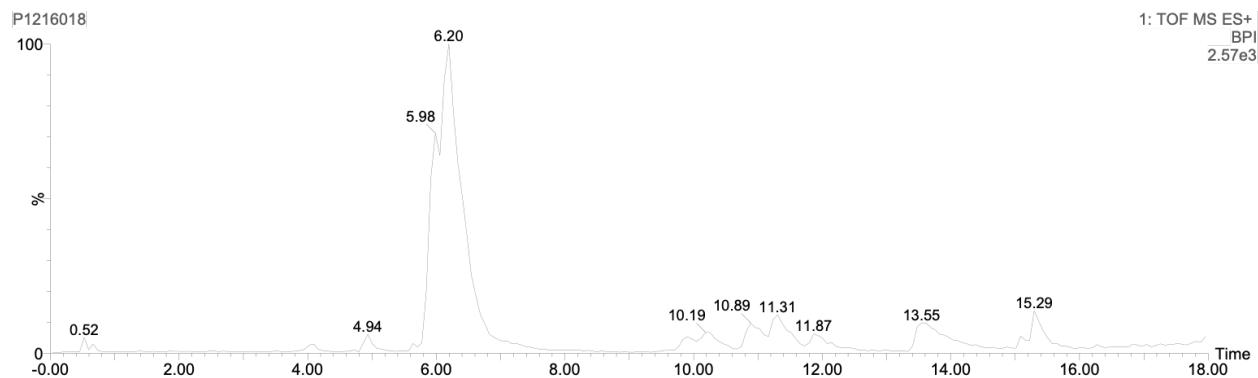


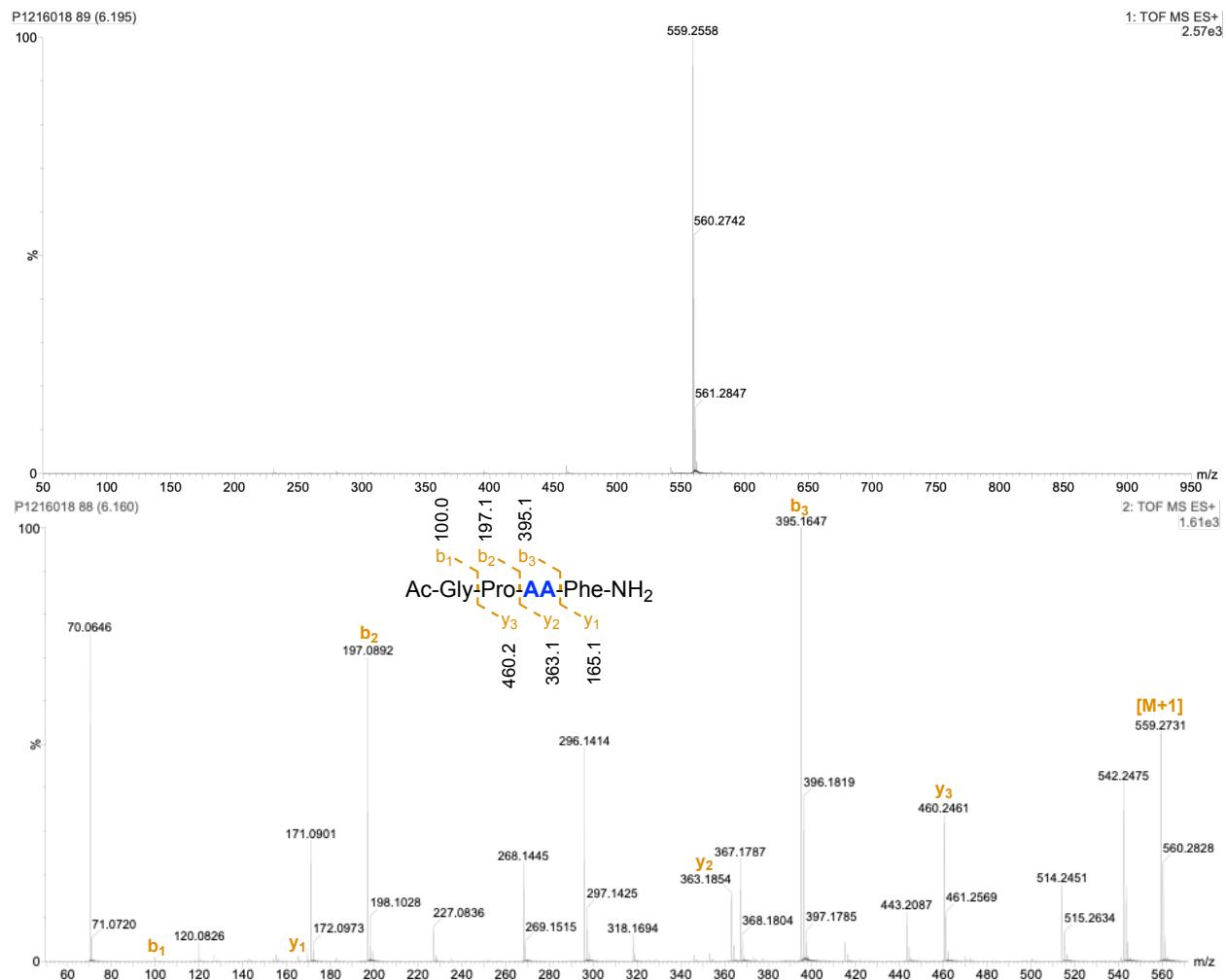


8B': MW = 558.6, Purity = 92.3%, Yield = 28.8% [0.38 mg]

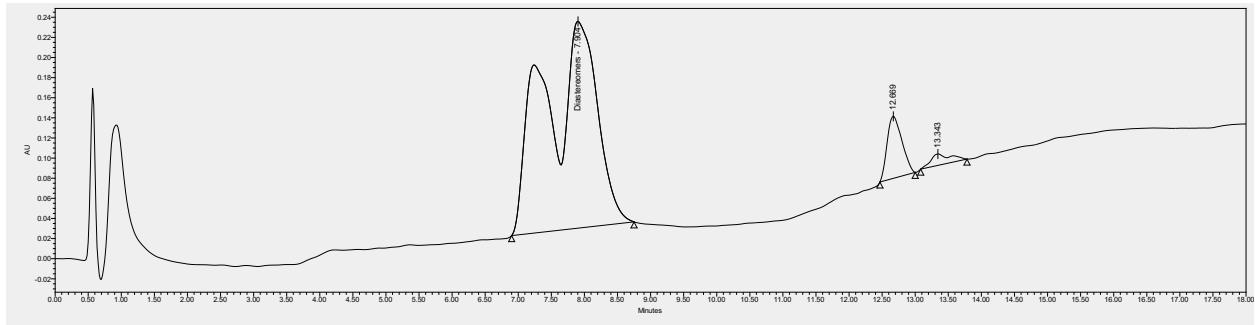
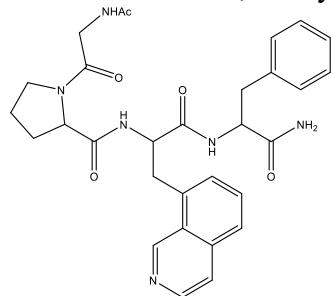


	Name	Retention Time	Area	% Area
1	Diastereomers	7.414	16593572	92.32
2		11.667	436159	2.43
3		12.318	944441	5.25

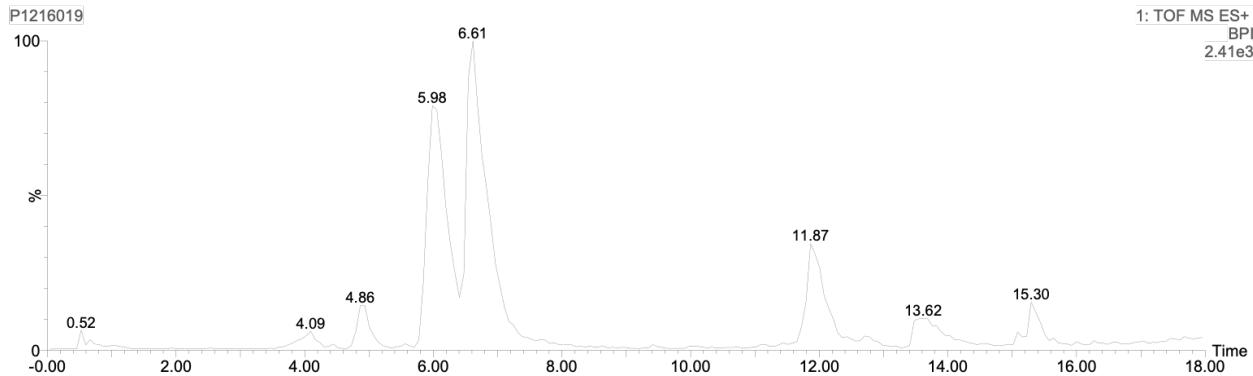


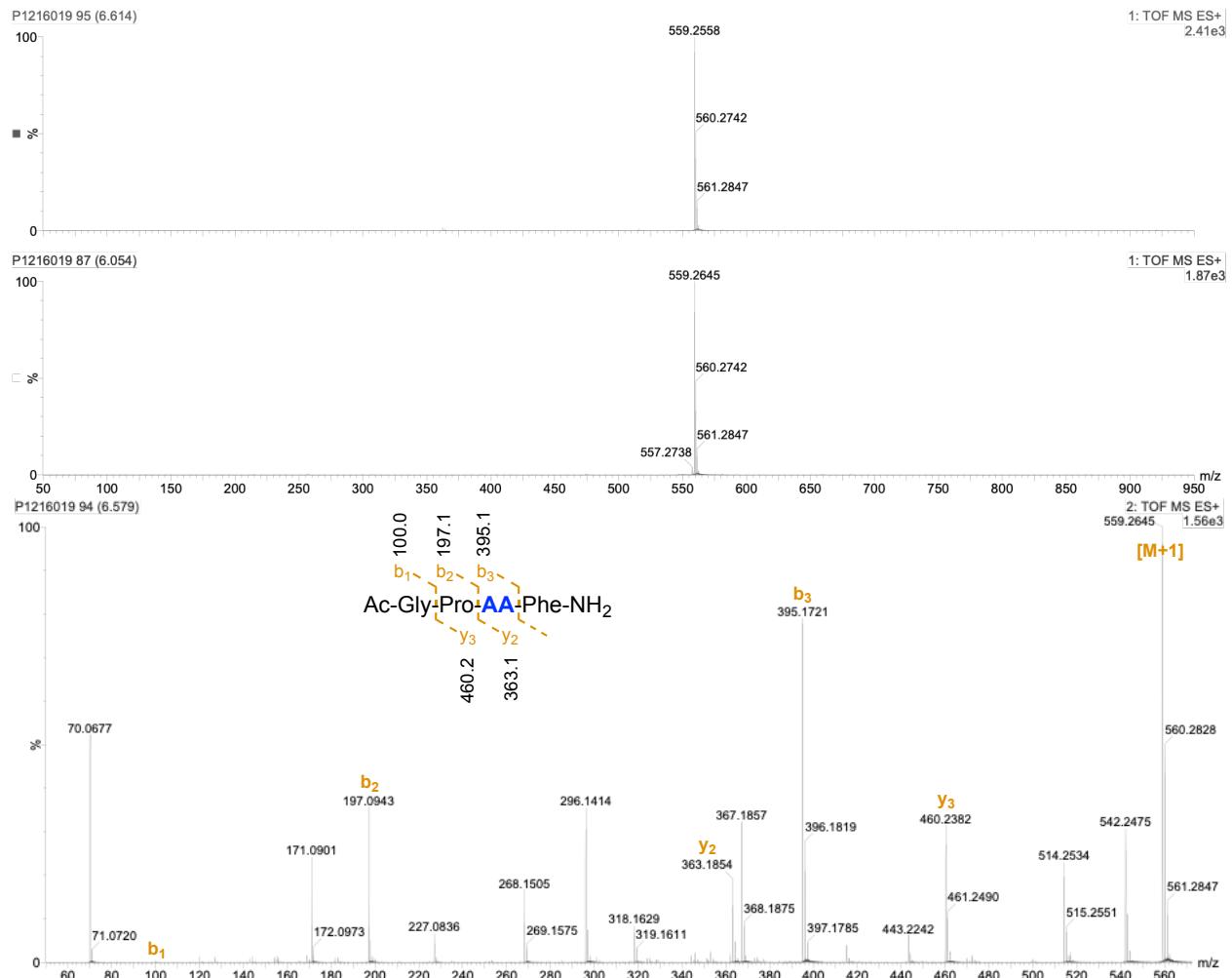


9B': MW = 558.6, Purity = 90.4%, Yield = 19.5% [0.25 mg]

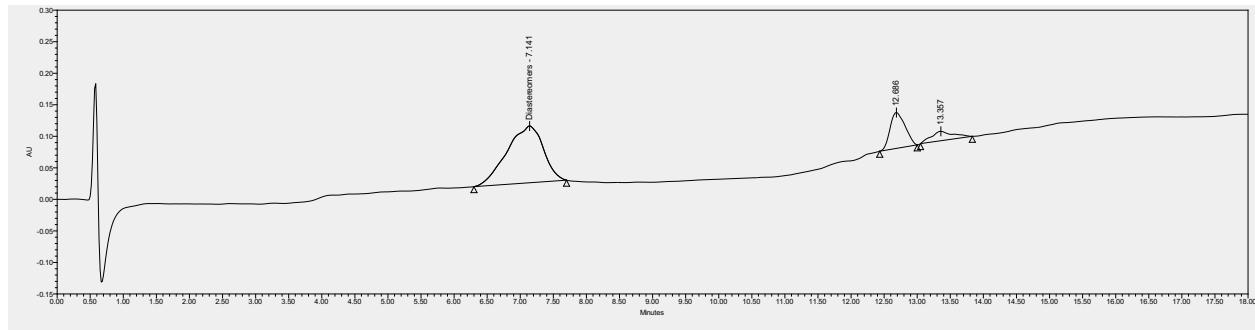
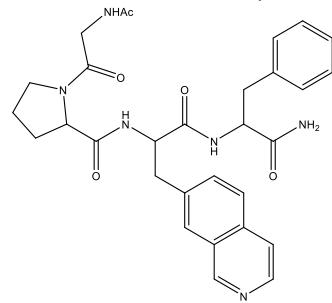


	Name	Retention Time	Area	% Area
1	Diastereomers	7.904	11205820	90.38
2		12.669	960905	7.75
3		13.343	231676	1.87

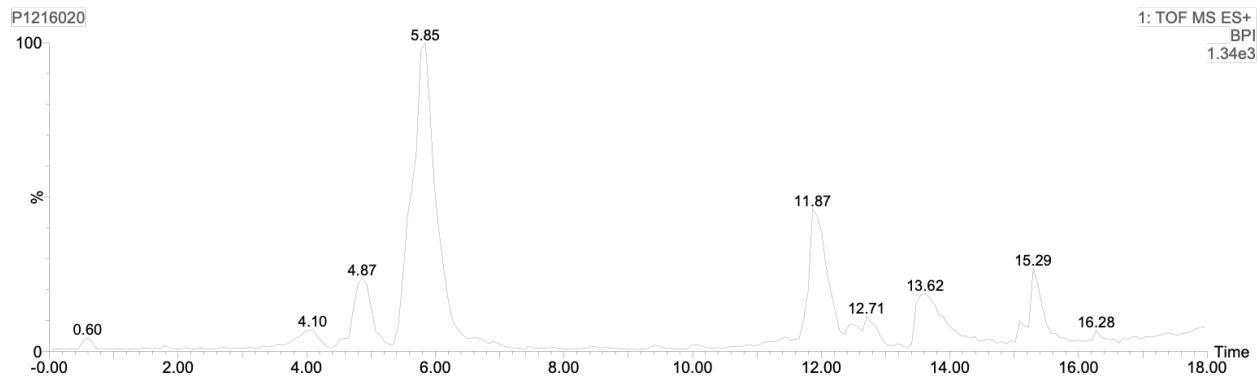


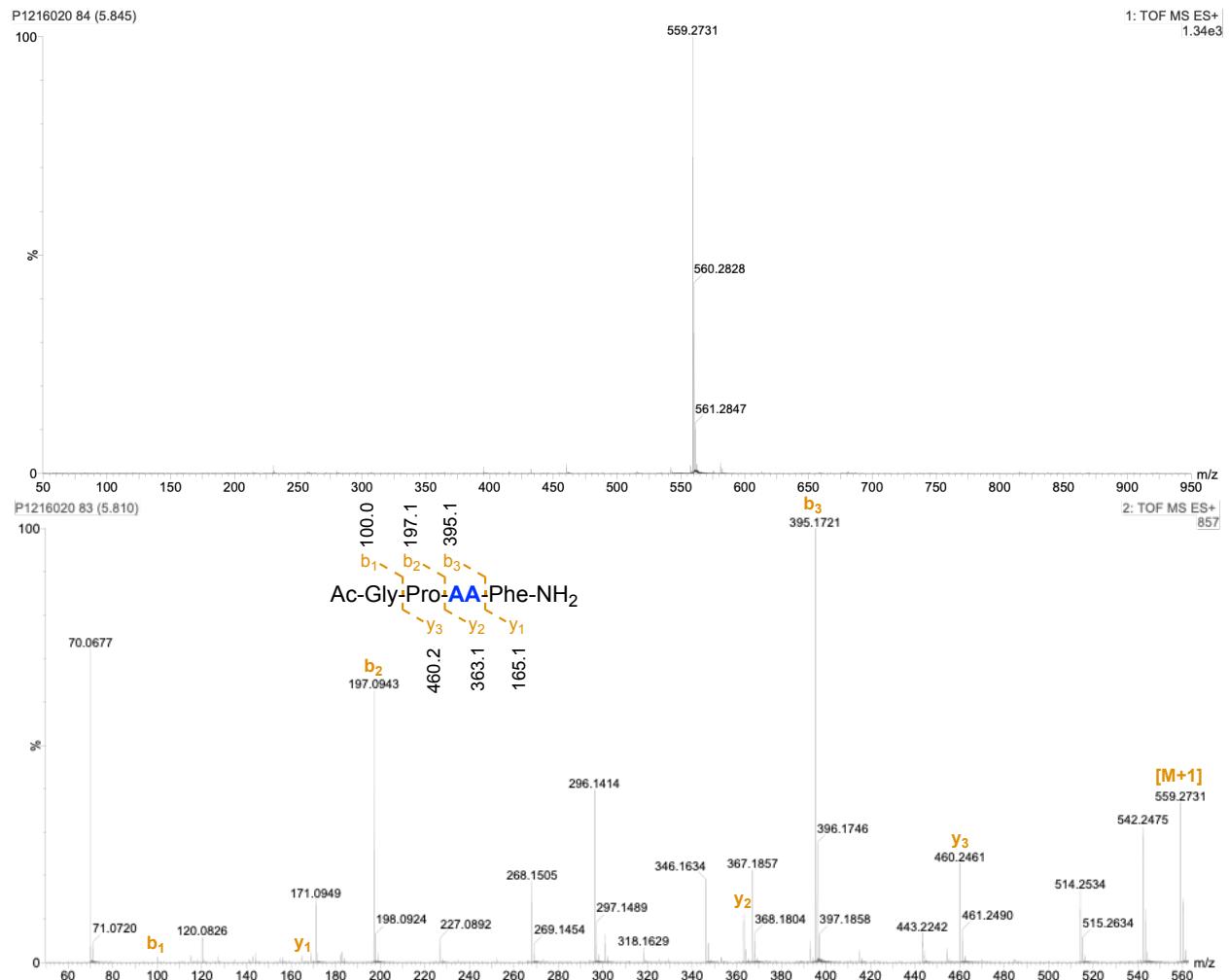


10B': MW = 558.6, Purity = 73.6%, Yield = 6.0% [0.078 mg]

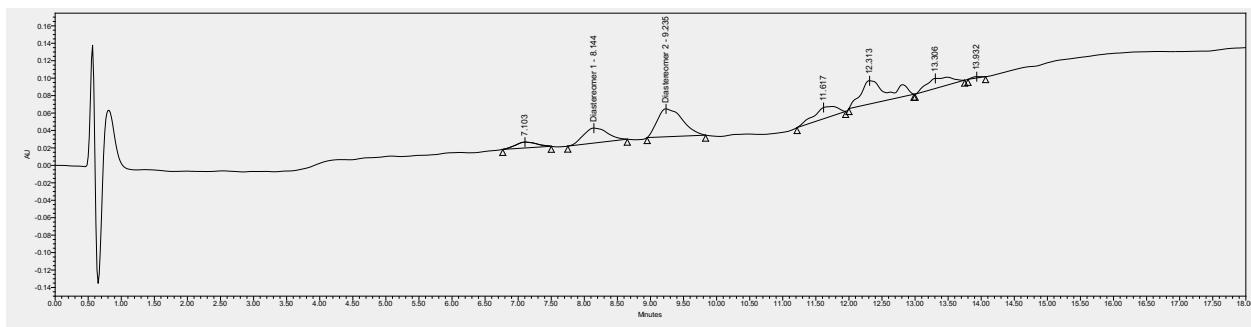
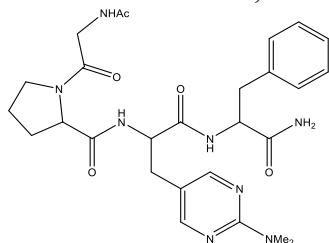


	Name	Retention Time	Area	% Area
1	Diastereomers	7.141	3459832	73.60
2		12.686	898364	19.11
3		13.357	342932	7.29

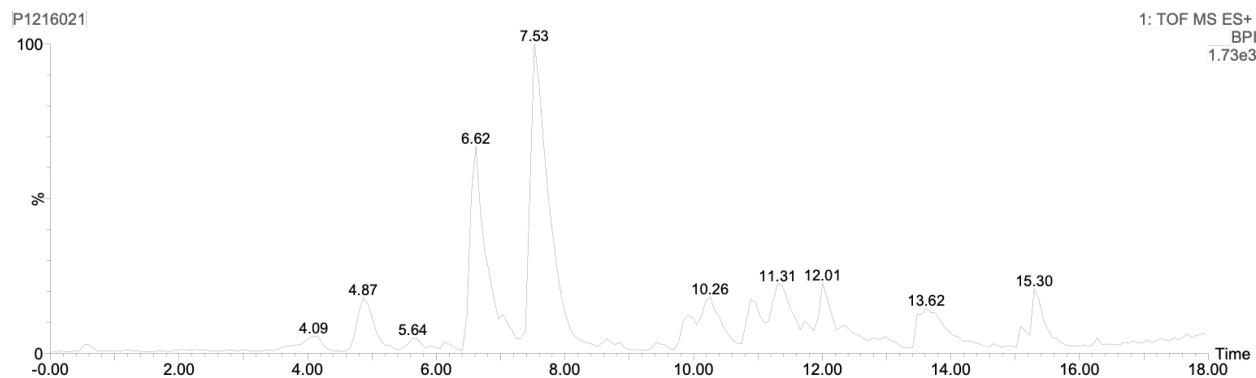


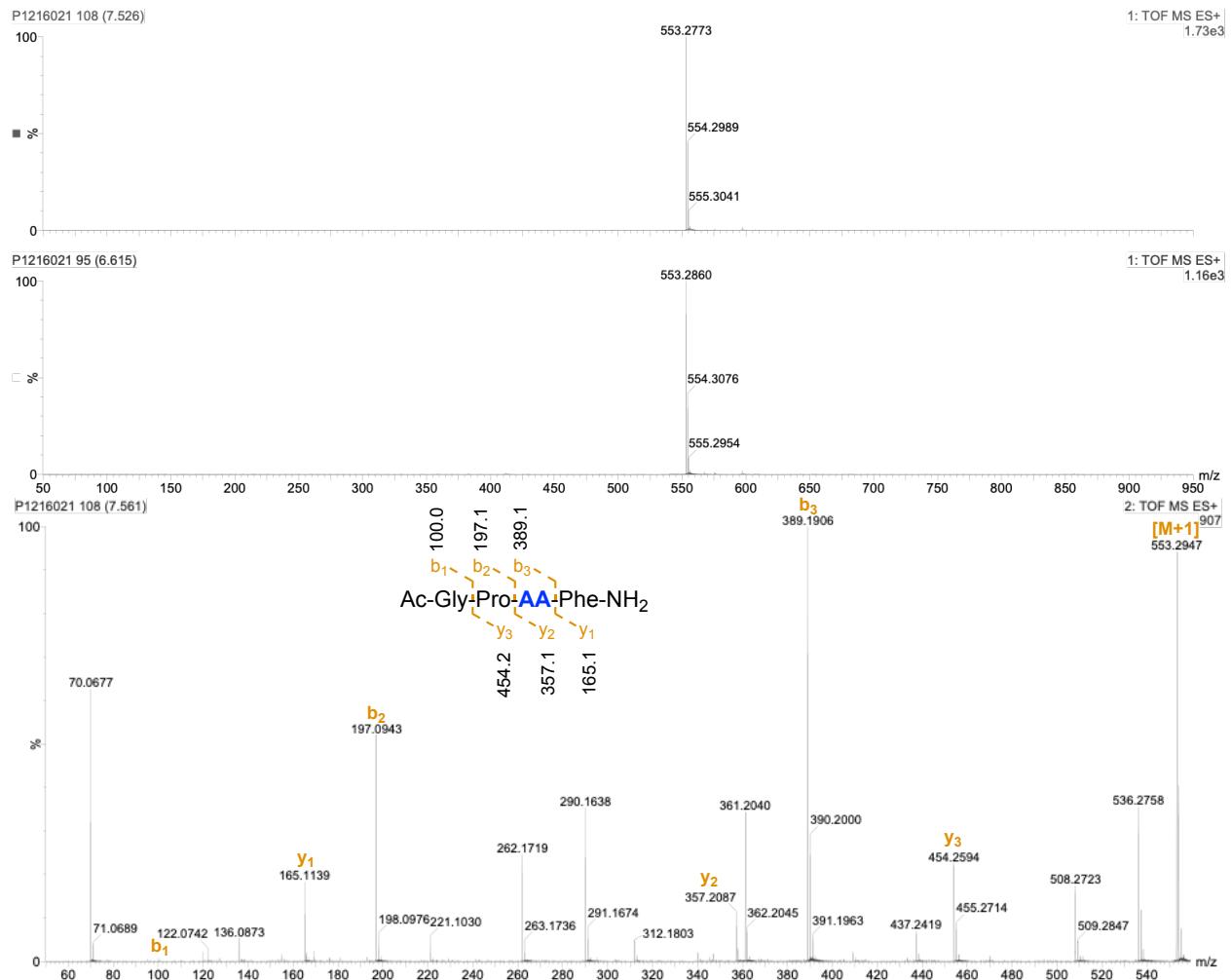


11B': MW = 552.6, Purity = 46.0%, Yield = 4.6% [0.059 mg]

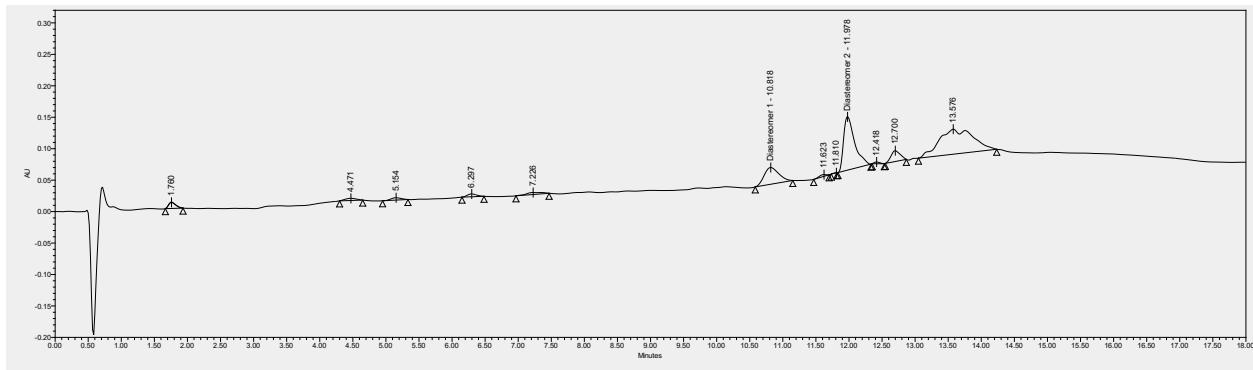
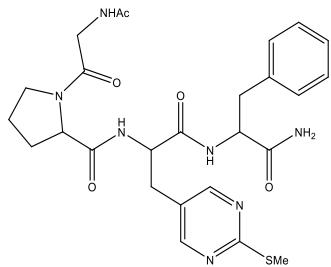


	Name	Retention Time	Area	% Area
1		7.103	150863	5.40
2	Diastereomer 1	8.144	436926	15.64
3	Diastereomer 2	9.235	847343	30.33
4		11.617	310463	11.11
5		12.313	741239	26.53
6		13.306	291918	10.45
7		13.932	14989	0.54

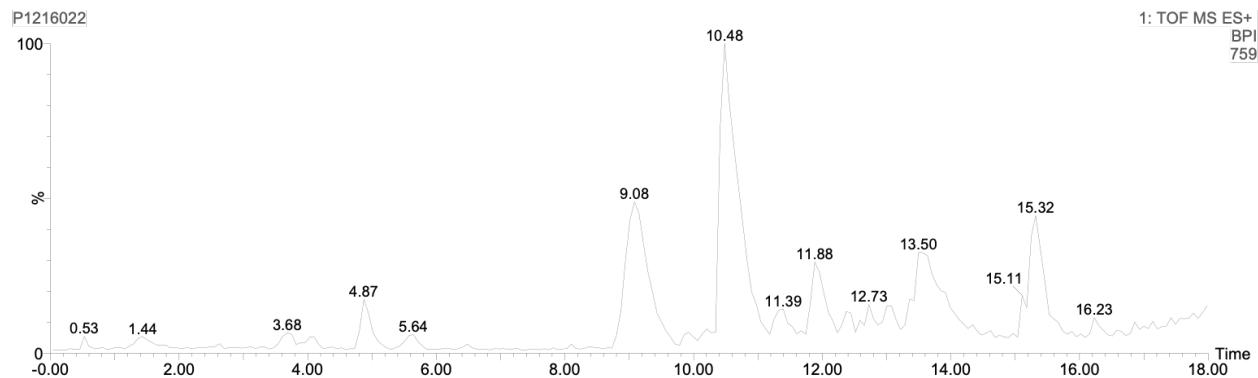


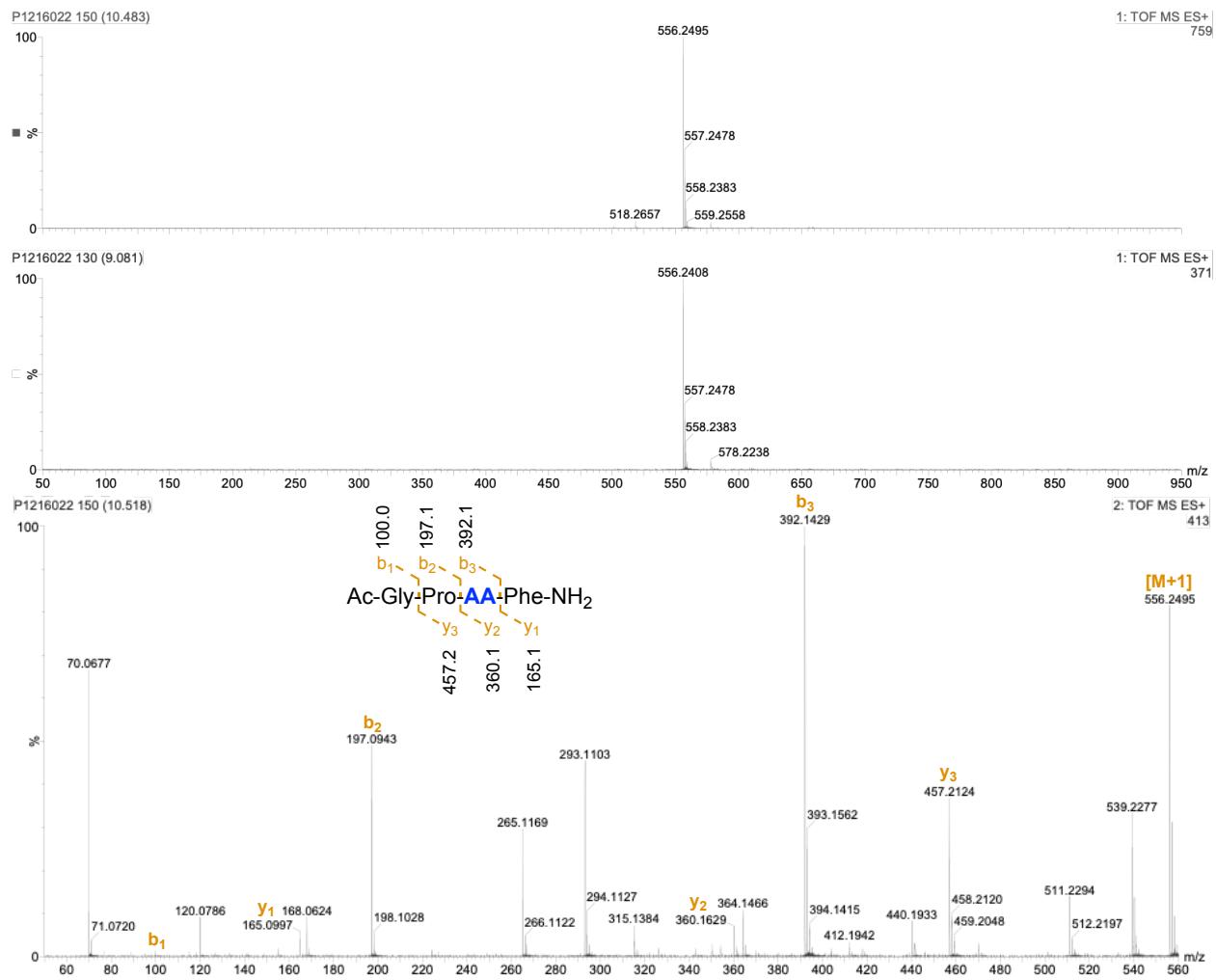


12B': MW = 555.6, Purity = 42.8%, Yield = 4.0% [0.052 mg]

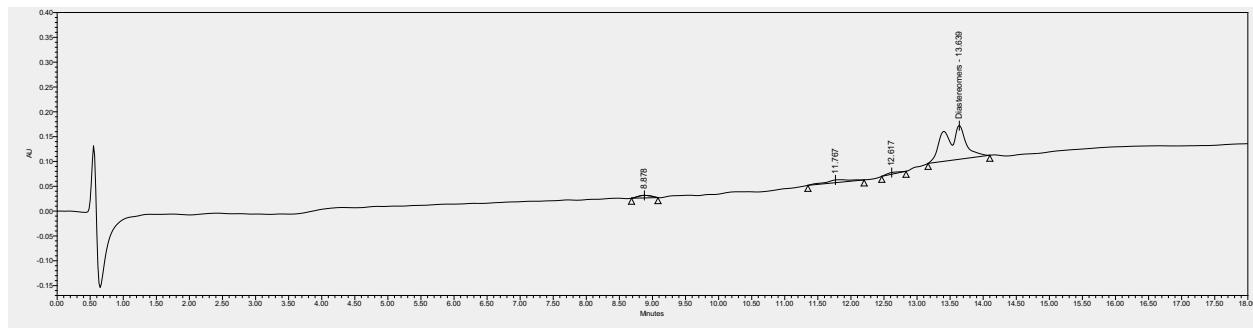
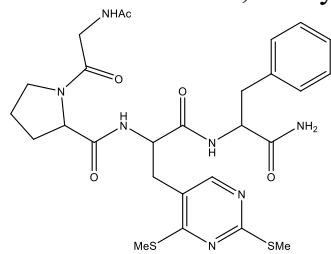


	Name	Retention Time	Area	% Area
1		1.760	73132	2.24
2		4.471	41225	1.26
3		5.154	40381	1.24
4		6.297	44364	1.36
5		7.226	49957	1.53
6	Diastereomer 1	10.818	410558	12.56
7		11.623	22421	0.69
8		11.810	1905	0.06
9	Diastereomer 2	11.978	989484	30.27
10		12.418	16523	0.51
11		12.700	165337	5.06
12		13.576	1413453	43.24

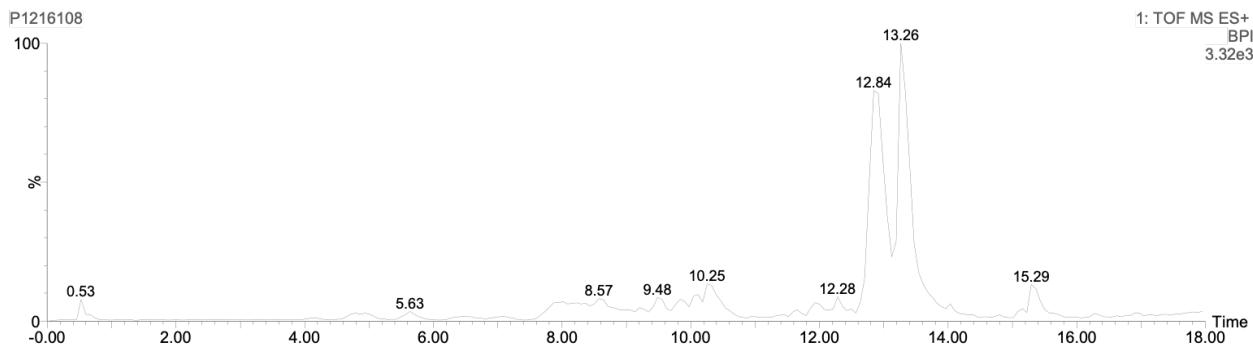


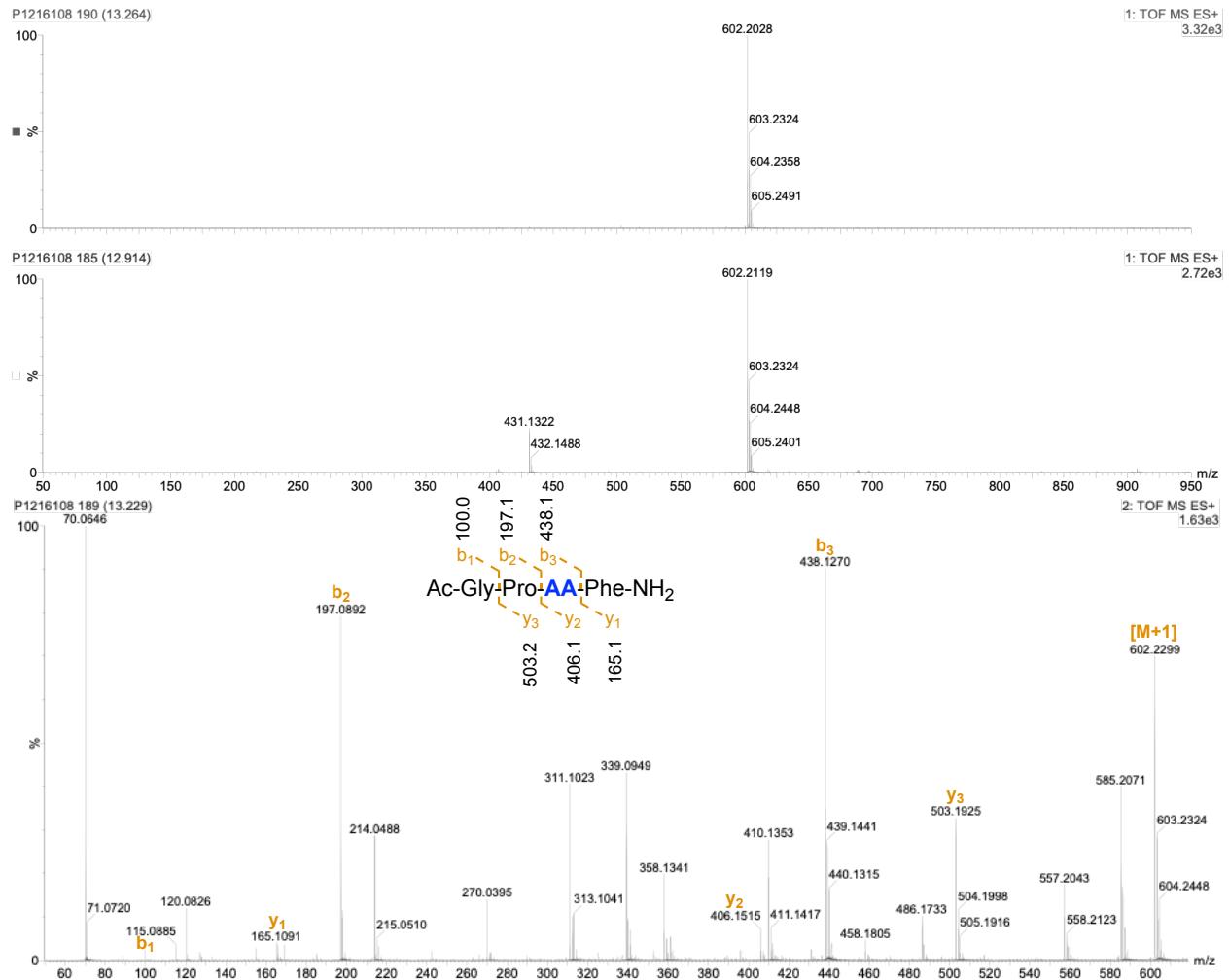


1C': MW = 601.7, Purity = 86.1%, Yield = 5.5% [0.078 mg]

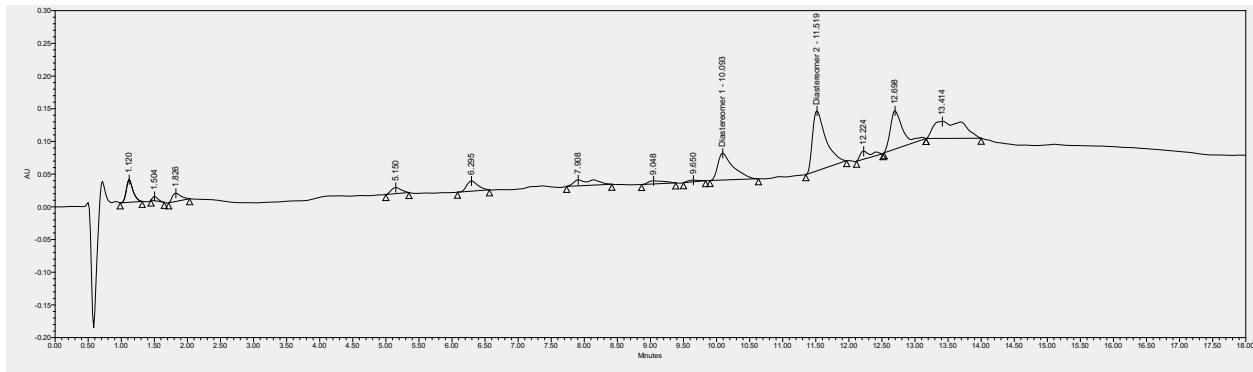
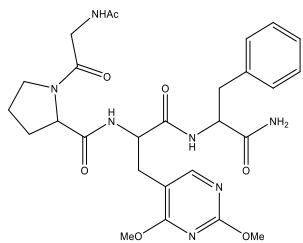


	Name	Retention Time	Area	% Area
1		8.878	72076	4.02
2		11.767	133926	7.47
3		12.617	42934	2.39
4	Diastereomers	13.639	1543967	86.12

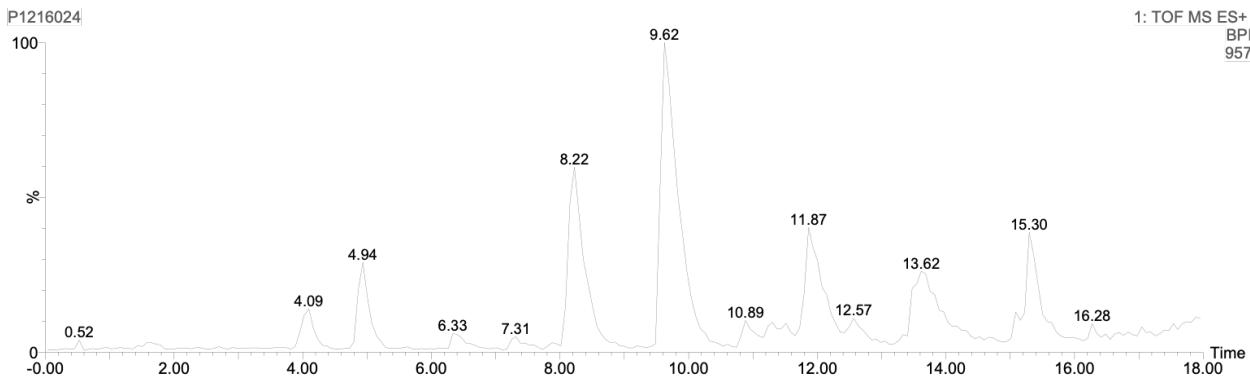


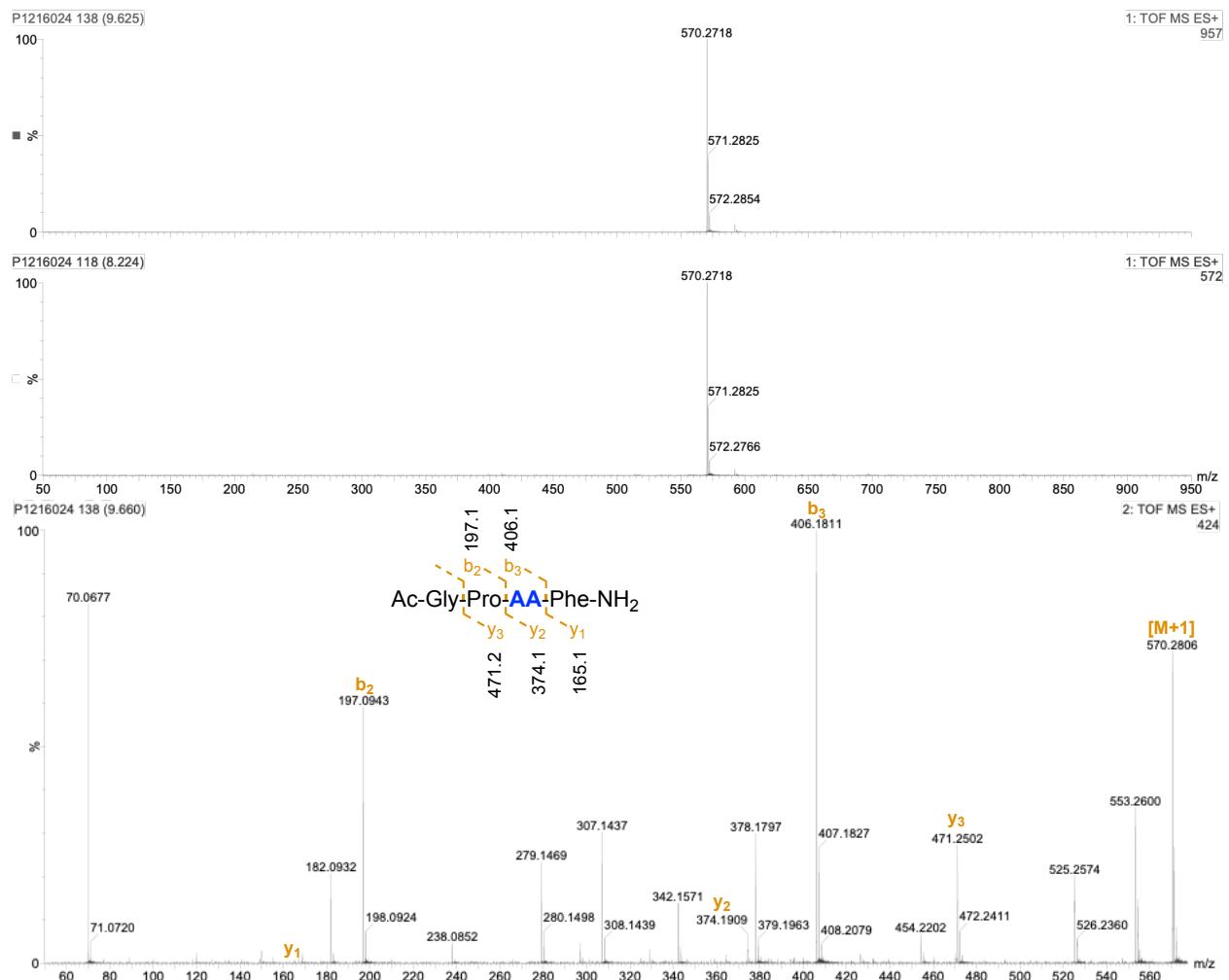


2C': MW = 569.6, Purity = 41.7%, Yield = 2.7% [0.036 mg]

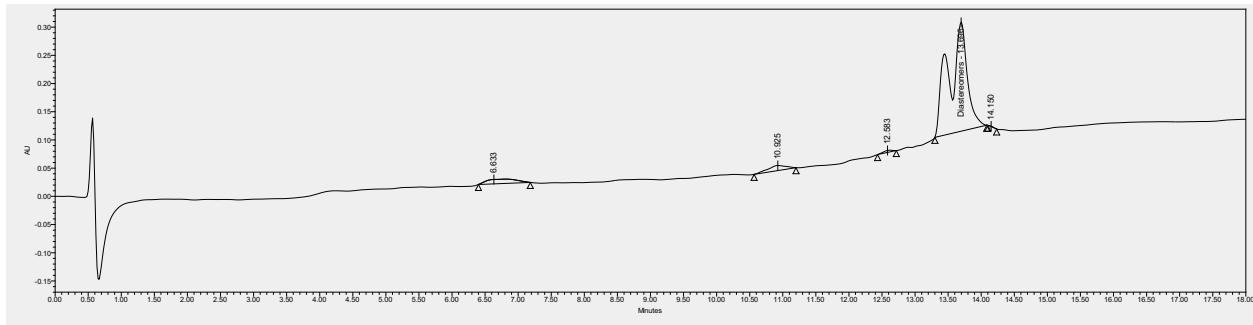
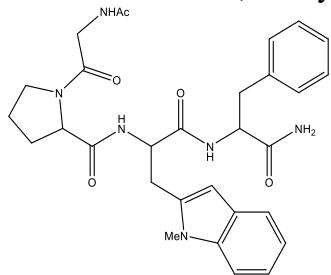


	Name	Retention Time	Area	% Area
1		1.120	244153	5.22
2		1.504	35955	0.77
3		1.826	116045	2.48
4		5.150	98509	2.11
5		6.295	184294	3.94
6		7.908	198591	4.25
7		9.048	81194	1.74
8		9.650	30191	0.65
9	Diastereomer 1	10.093	660702	14.13
10	Diastereomer 2	11.519	1289024	27.58
11		12.224	139175	2.98
12		12.698	756886	16.19
13		13.414	839561	17.96

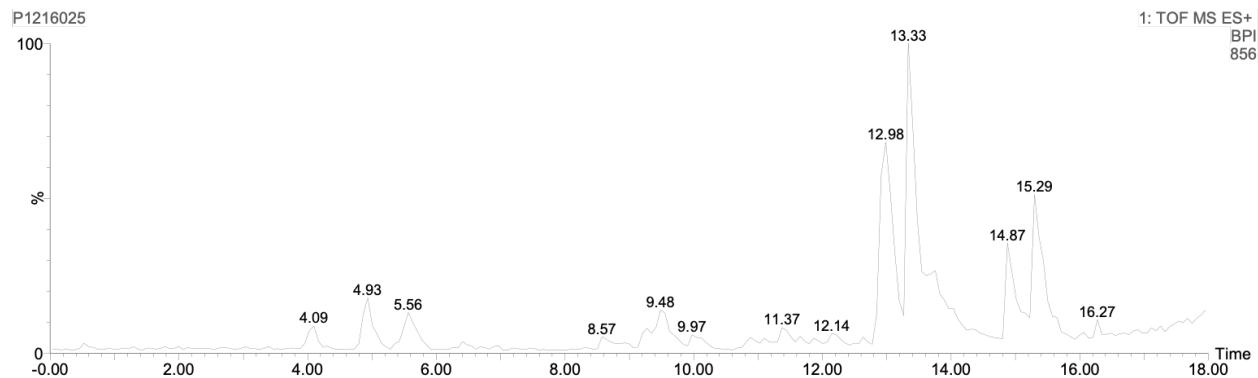


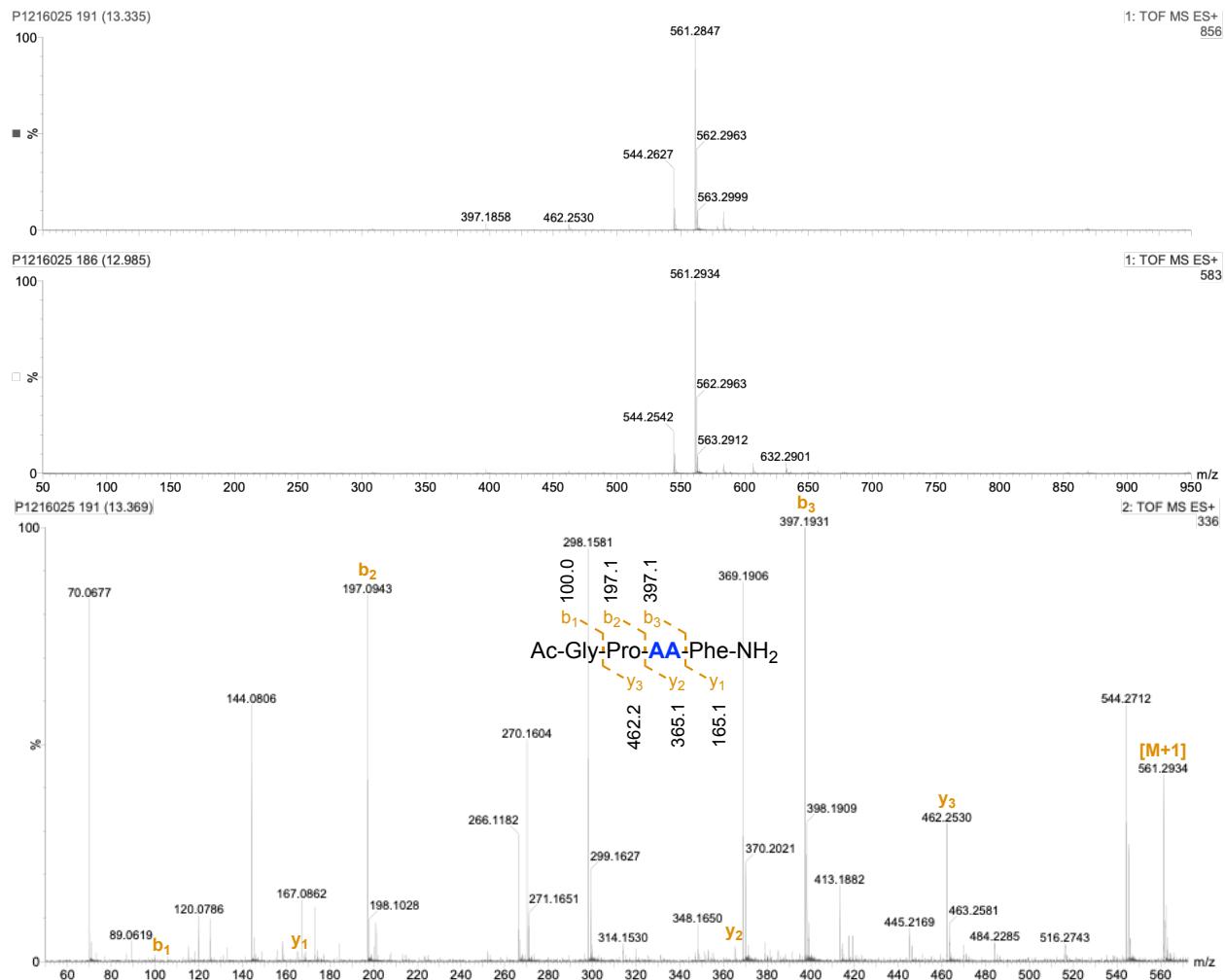


3C': MW = 560.7, Purity = 89.0%, Yield = 3.6% [0.047 mg]

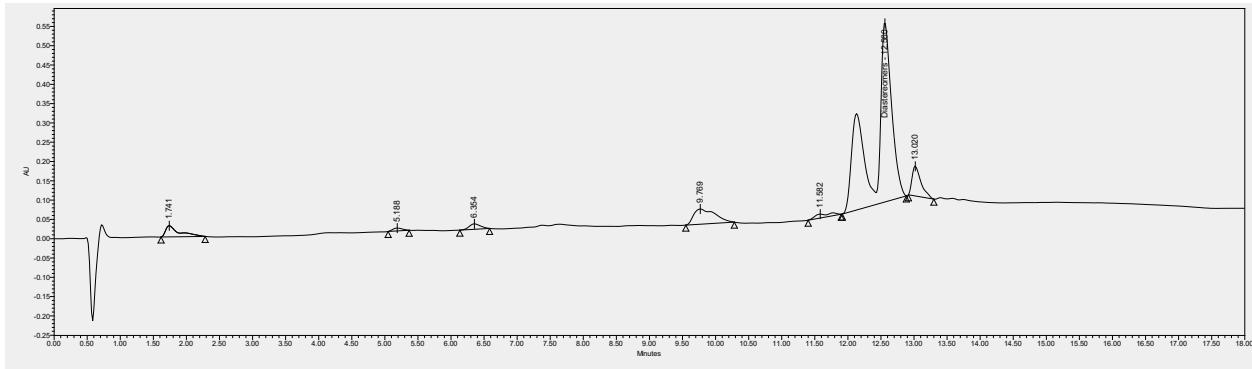
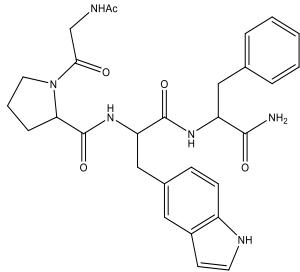


	Name	Retention Time	Area	% Area
1		6.633	246352	5.86
2		10.925	178094	4.24
3		12.583	32669	0.78
4	Diastereomers	13.696	3739460	89.00
5		14.150	4847	0.12

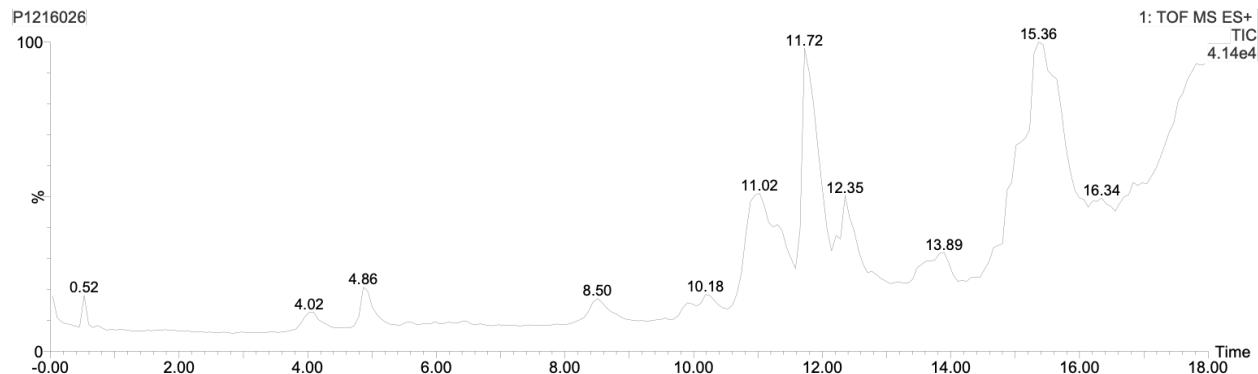


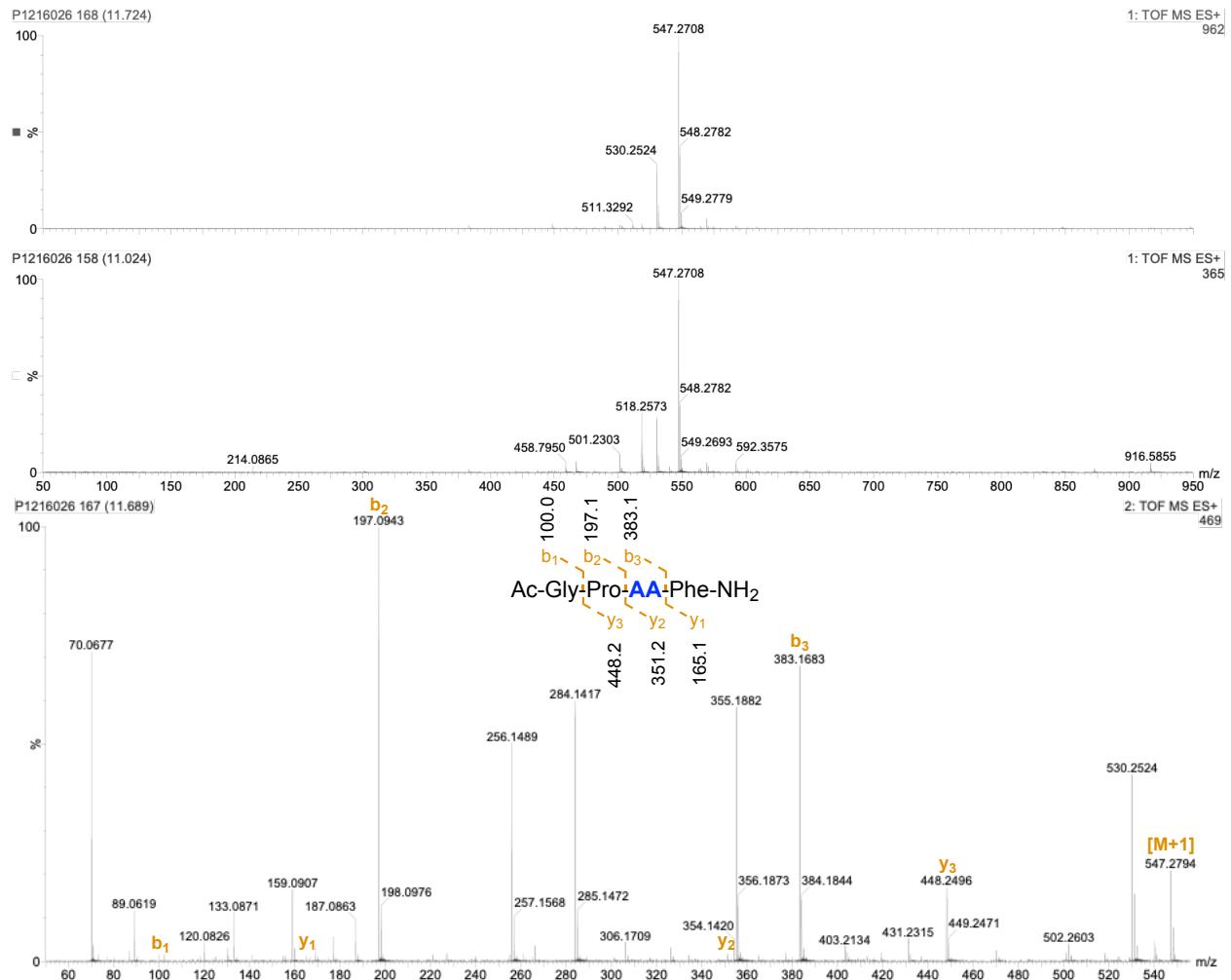


4C': MW = 546.6, Purity = 77.8%, Yield = 6.5% [0.083 mg]

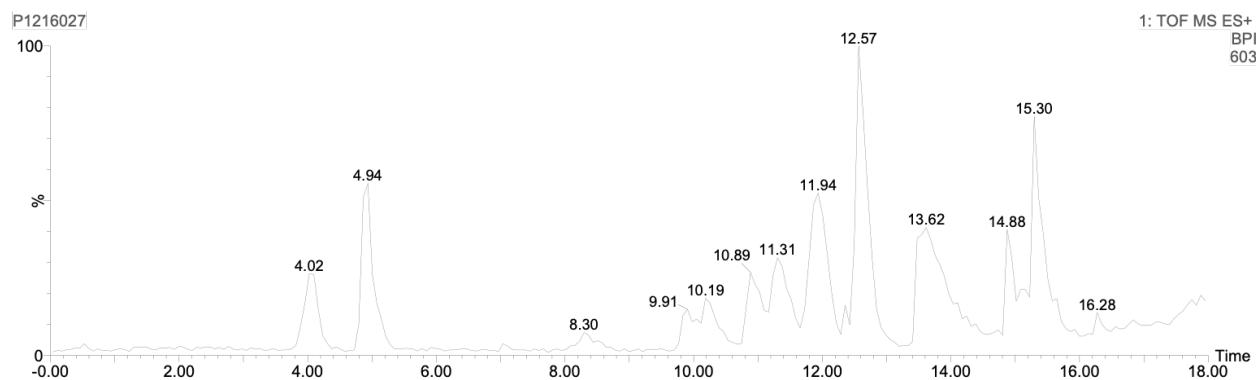
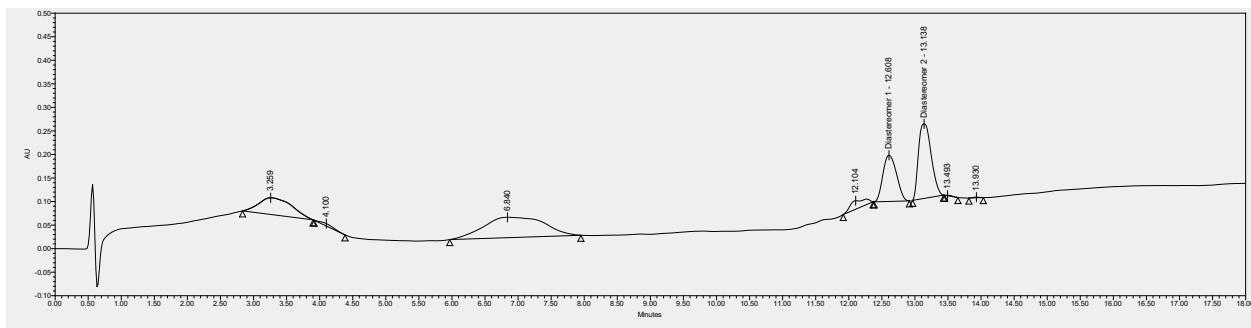
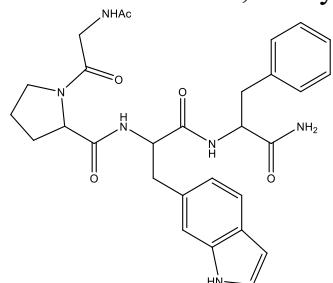


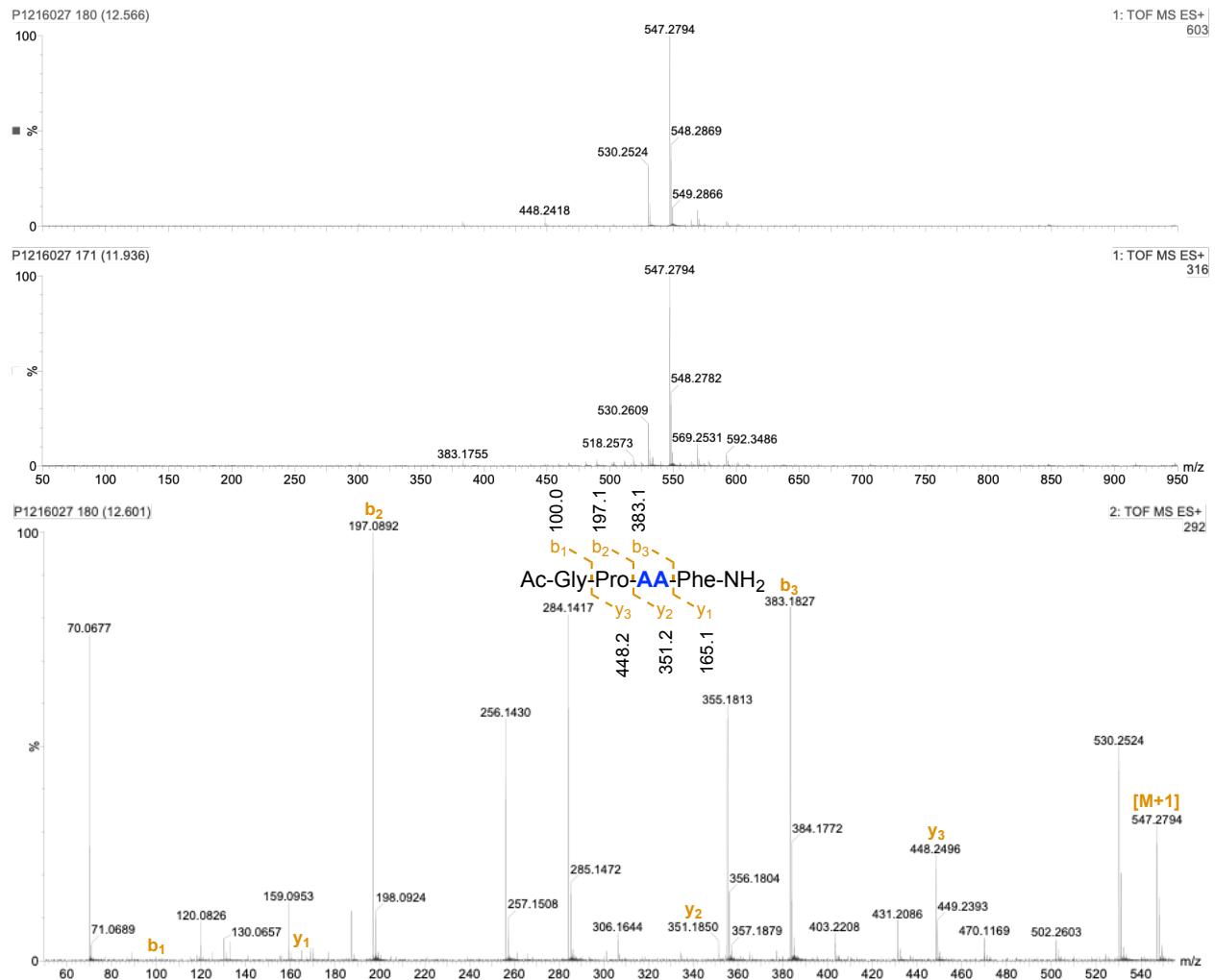
	Name	Retention Time	Area	% Area
1		1.741	415418	3.79
2		5.188	75320	0.69
3		6.354	167137	1.52
4		9.769	878128	8.00
5		11.582	170220	1.55
6	Diastereomers	12.560	8538789	77.83
7		13.020	726476	6.62



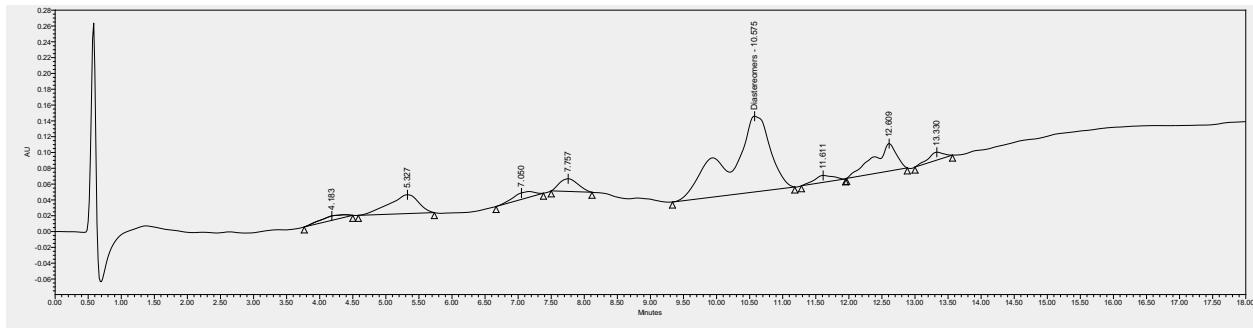
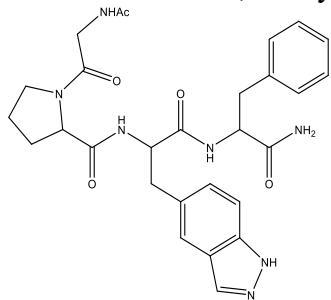


5C': MW = 546.6, Purity = 54.2%, Yield = 9.6% [0.12 mg]

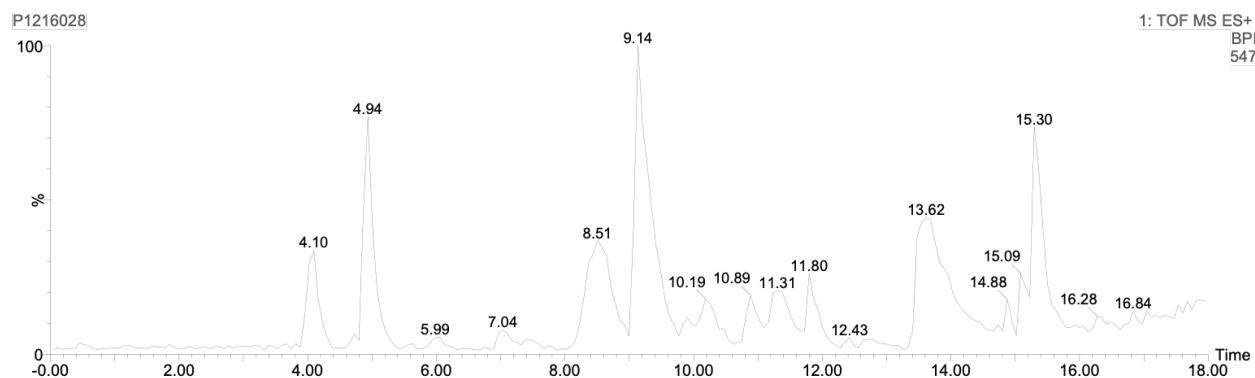


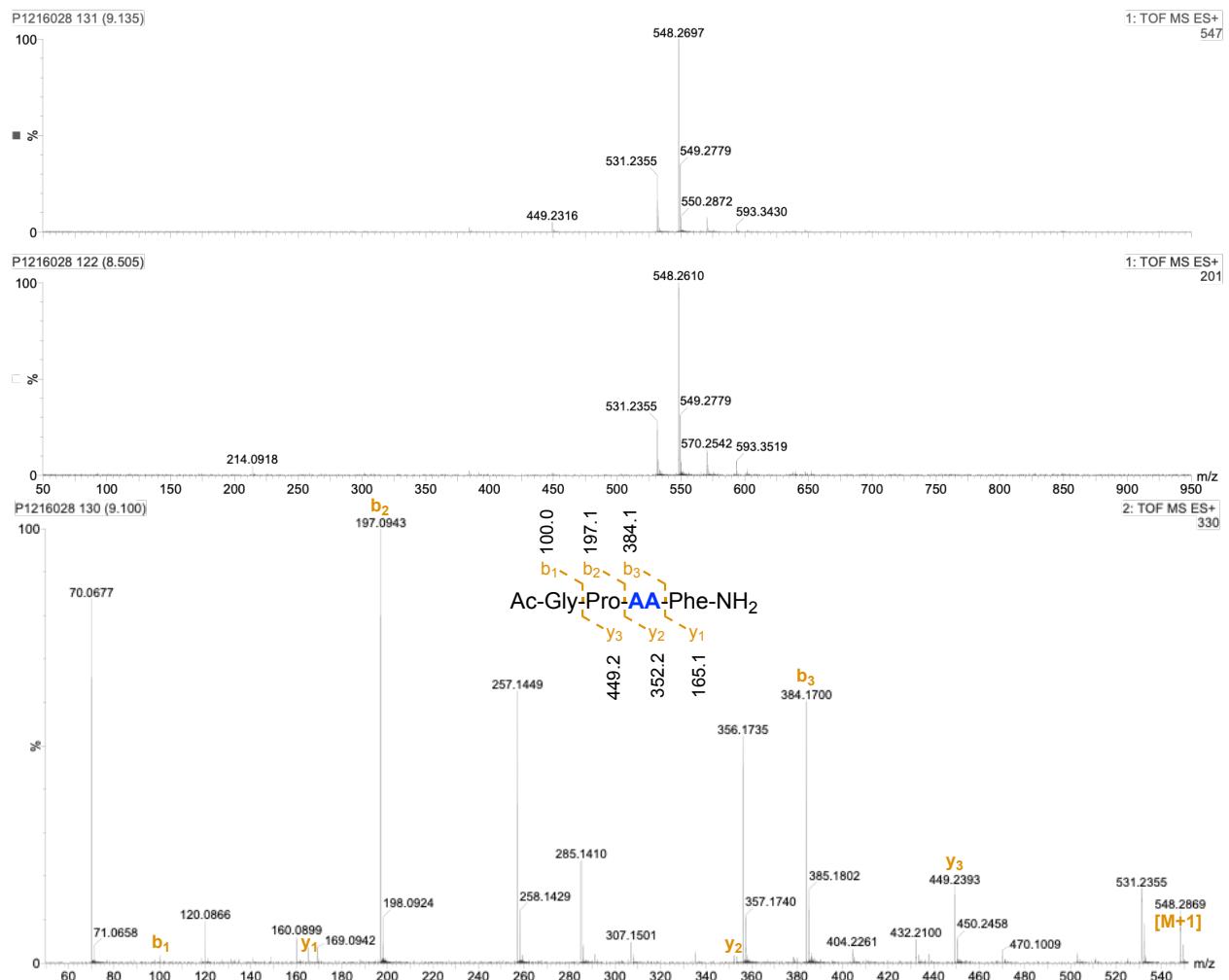


6C': MW = 547.6, Purity = 62.0%, Yield = 6.2% [0.080 mg]

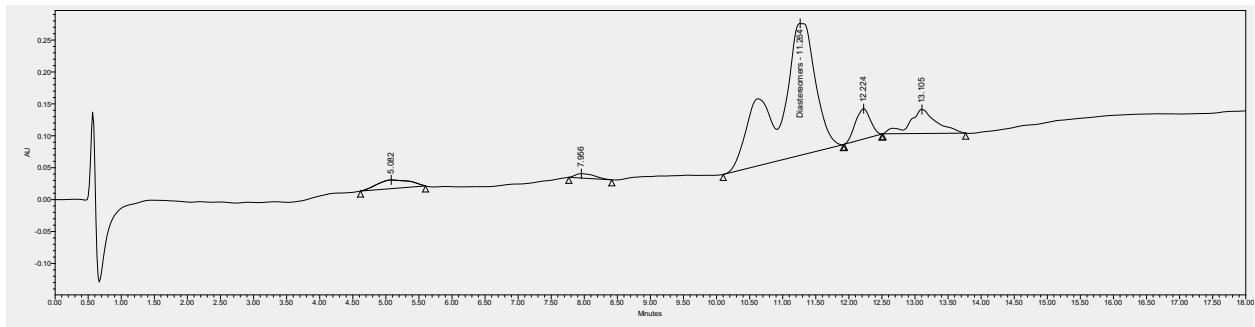
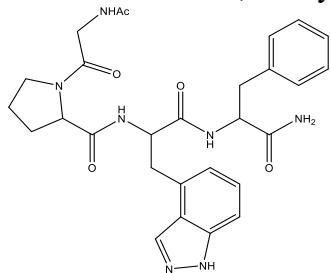


	Name	Retention Time	Area	% Area
1		4.183	142485	2.09
2		5.327	744279	10.93
3		7.050	179673	2.64
4		7.757	315844	4.64
5	Diastereomers	10.575	4224271	62.02
6		11.611	173919	2.55
7		12.609	865817	12.71
8		13.330	164924	2.42

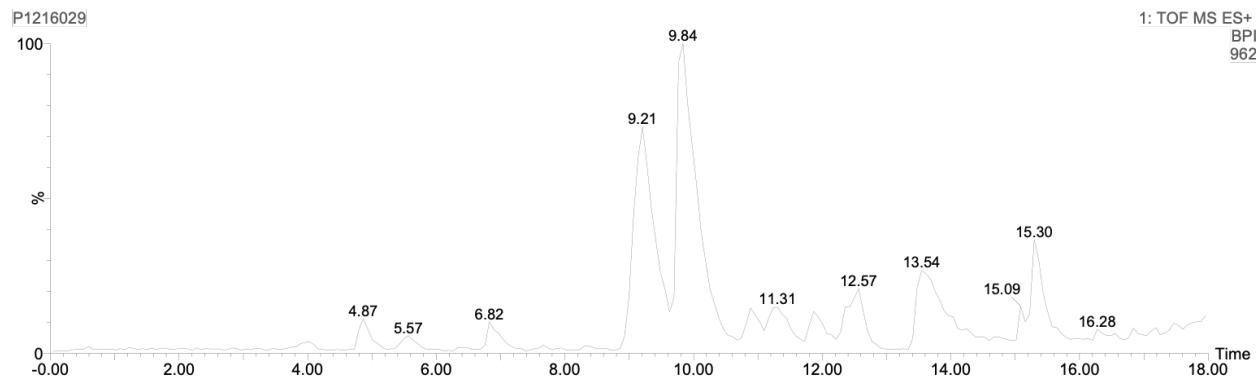


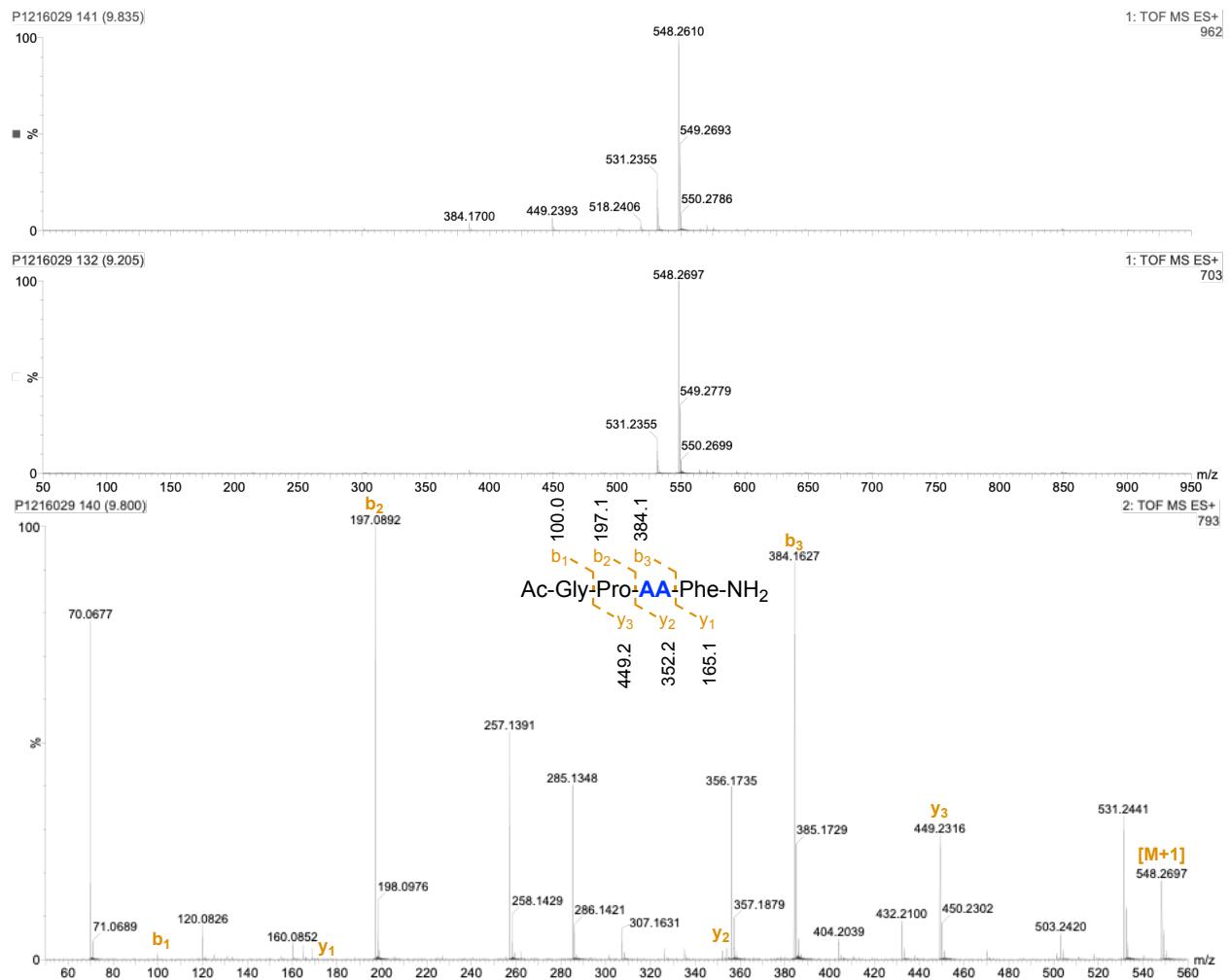


7C': MW = 547.6, Purity = 77.9%, Yield = 12.7% [0.16 mg]

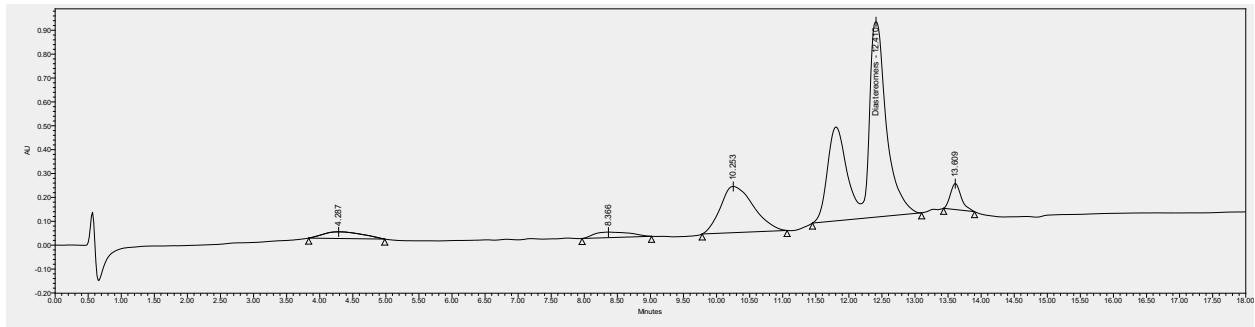
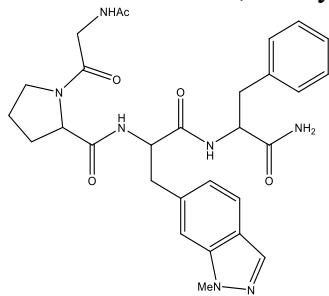


	Name	Retention Time	Area	% Area
1		5.082	466551	4.23
2		7.956	143479	1.30
3	Diastereomers	11.264	8597511	77.92
4		12.224	755364	6.85
5		13.105	1070331	9.70

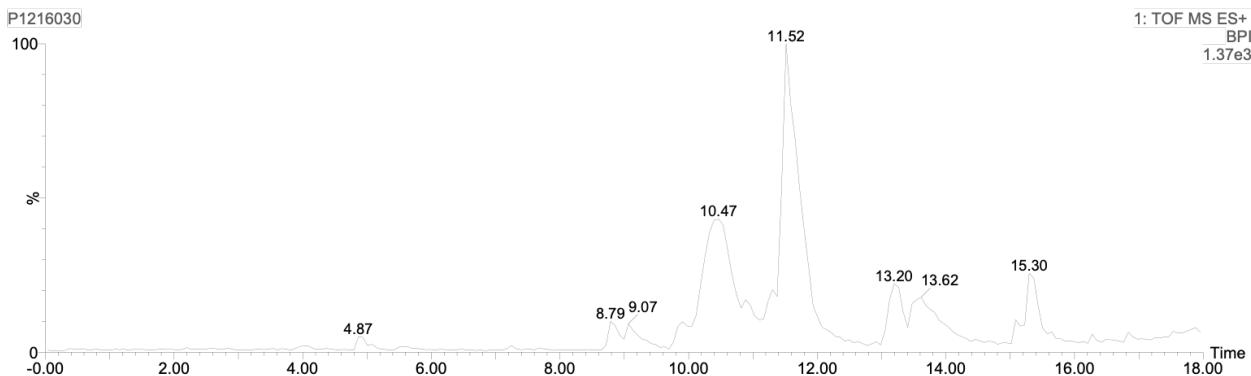


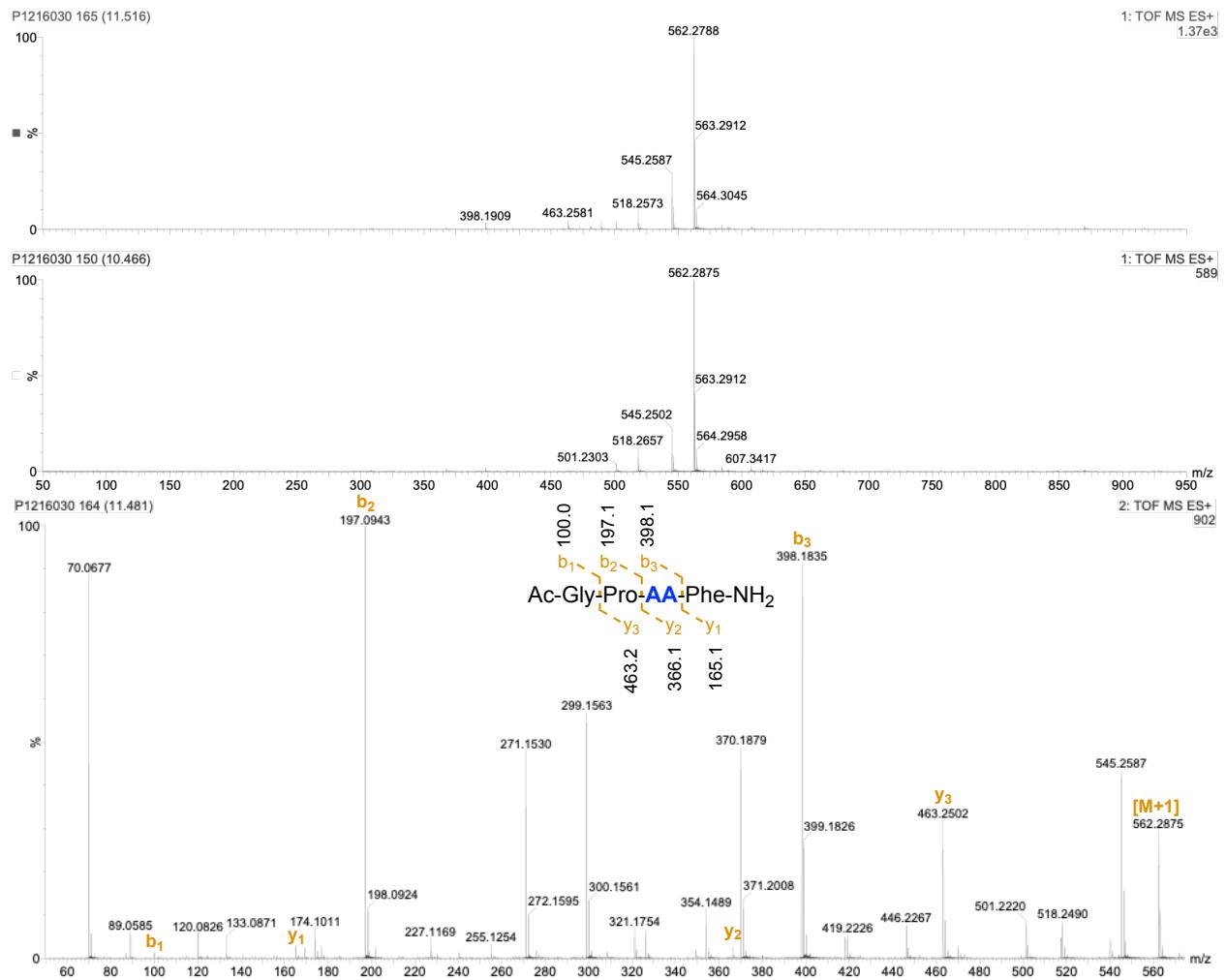


8C': MW = 561.6, Purity = 69.2%, Yield = 33.1% [0.43 mg]

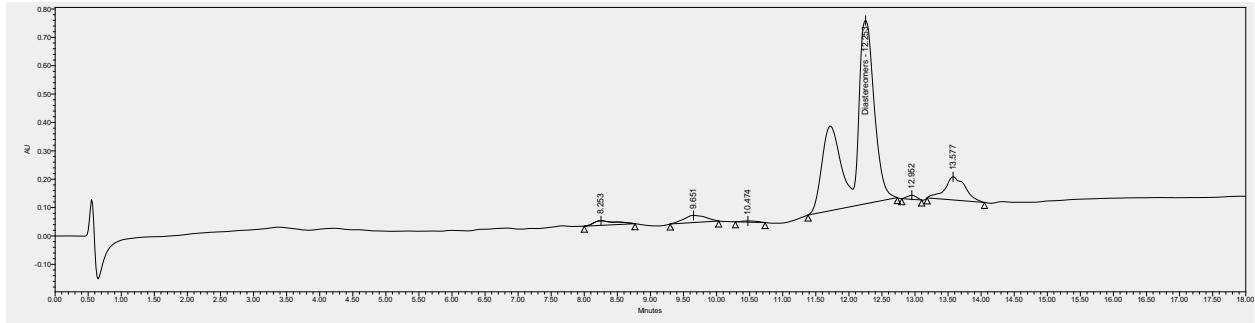
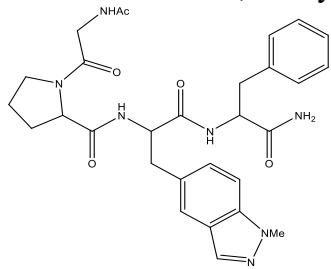


	Name	Retention Time	Area	% Area
1		4.287	1154892	3.56
2		8.366	887289	2.74
3		10.253	6703786	20.69
4	Diastereomers	12.410	22416973	69.19
5		13.609	1237419	3.82

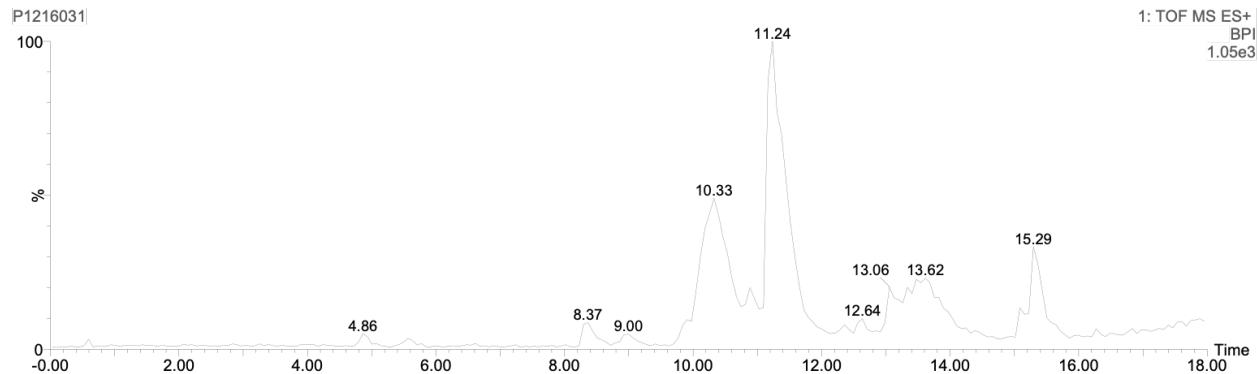


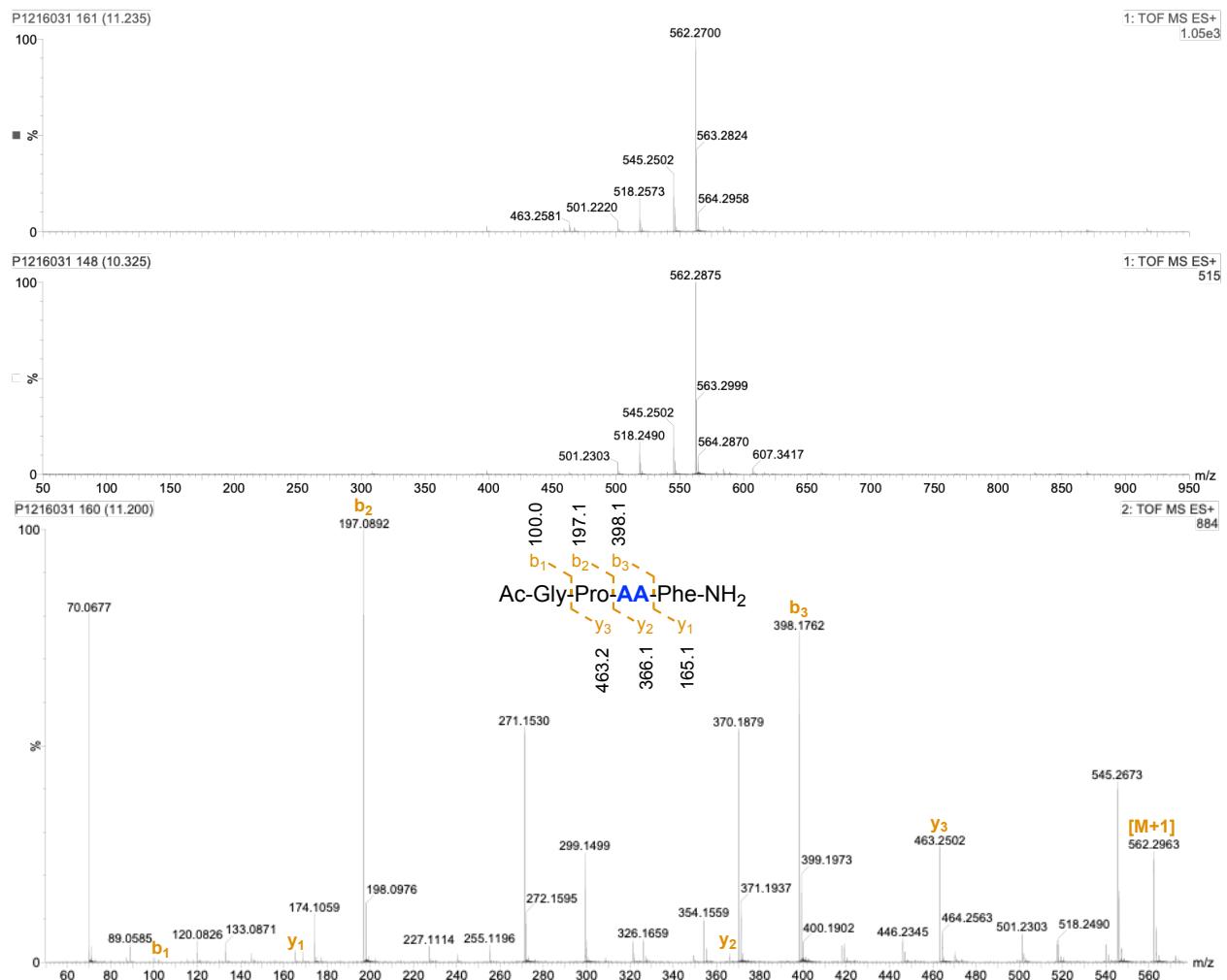


9C': MW = 561.6, Purity = 84.6%, Yield = 23.8% [0.31 mg]

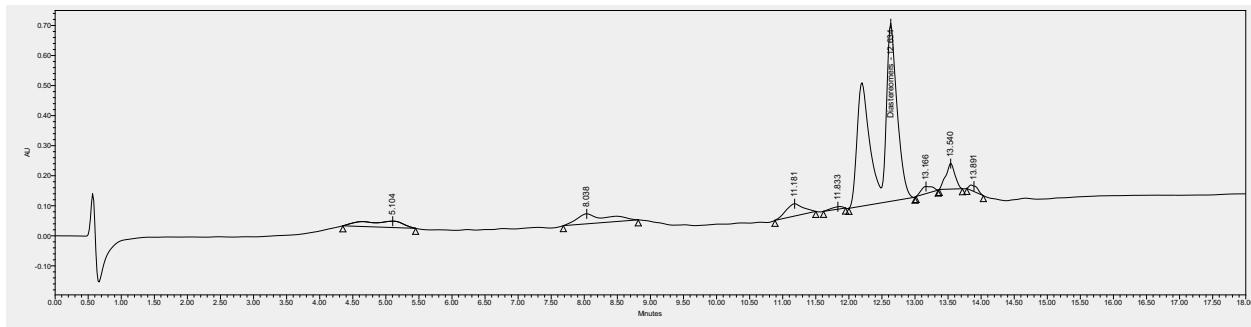
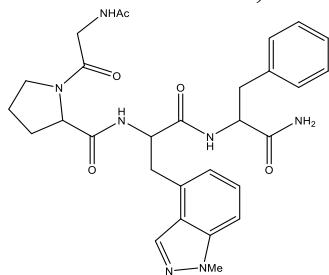


	Name	Retention Time	Area	% Area
1		8.253	377277	1.98
2		9.651	587998	3.09
3		10.474	69300	0.36
4	Diastereomers	12.253	16085566	84.60
5		12.952	141046	0.74
6		13.577	1752051	9.21

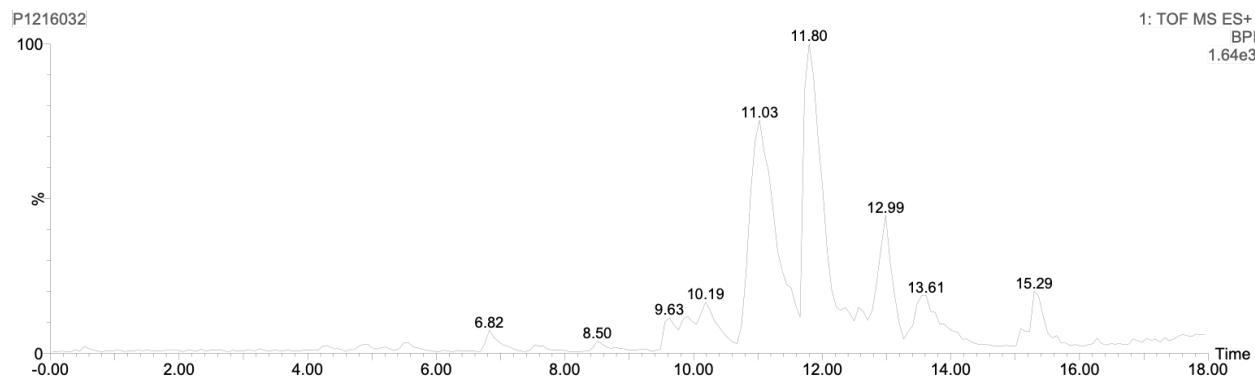


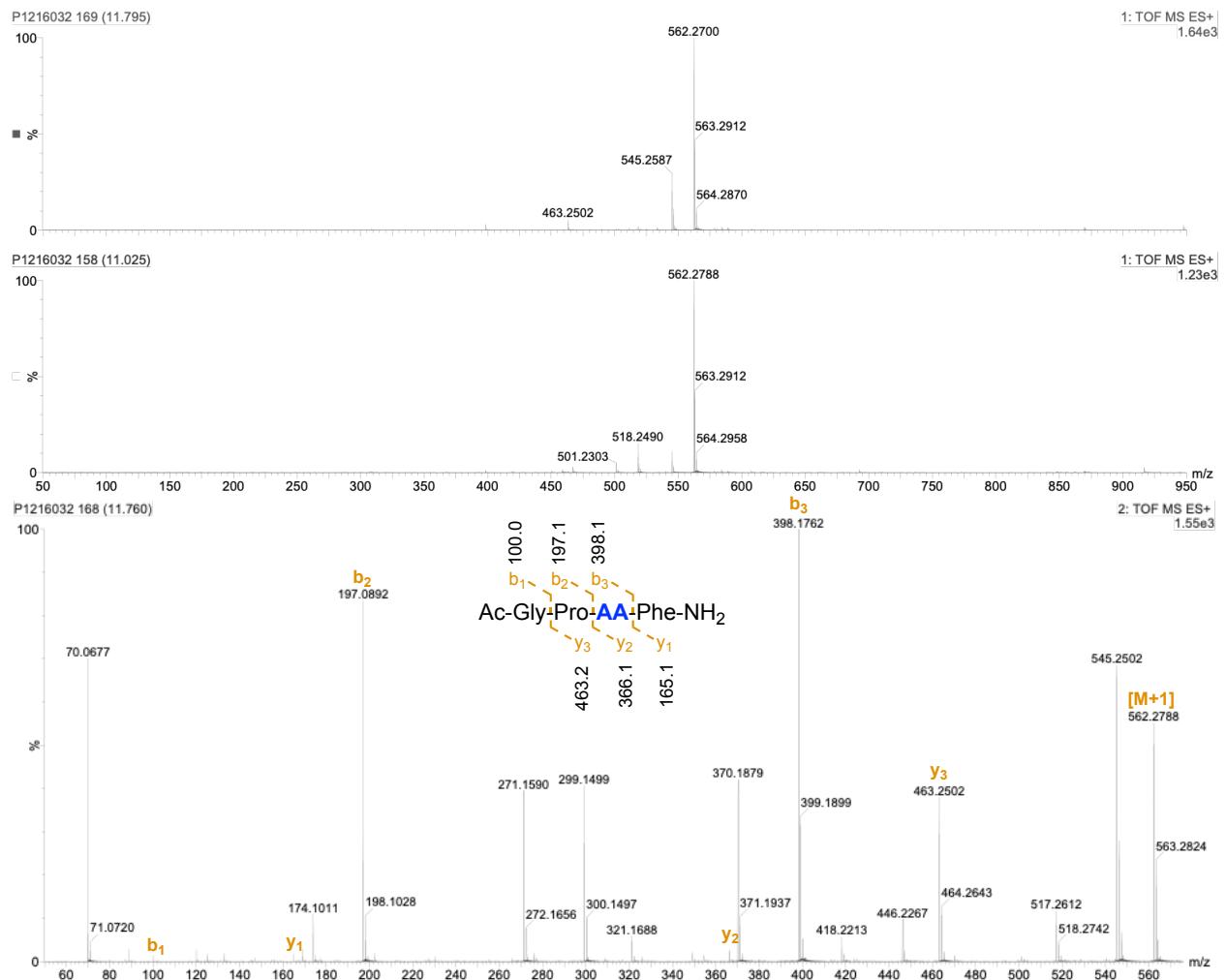


10C': MW = 561.6, Purity = 74.9%, Yield = 14.2% [0.19 mg]

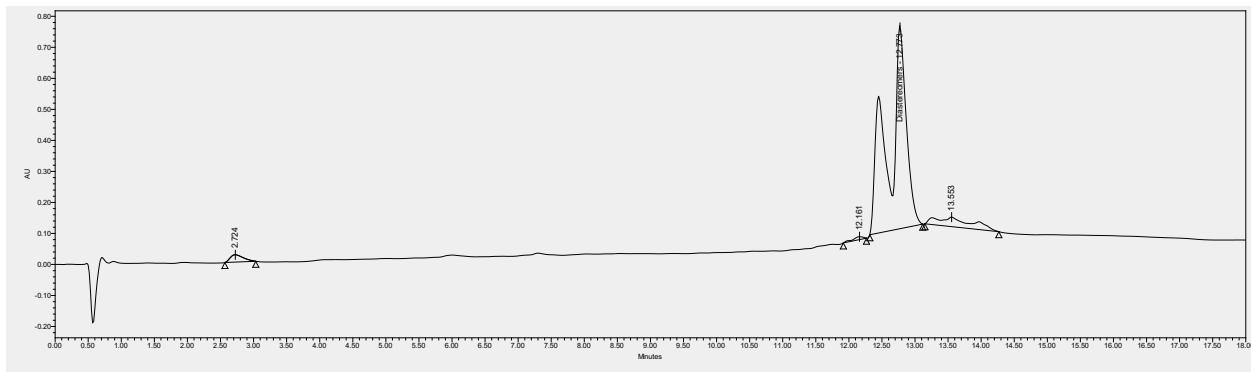
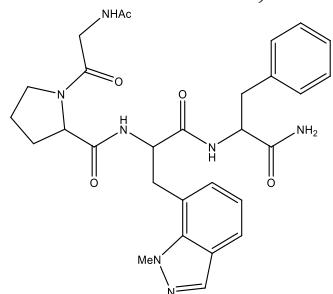


	Name	Retention Time	Area	% Area
1		5.104	861030	5.37
2		8.038	1060676	6.61
3		11.181	741179	4.62
4		11.833	100574	0.63
5	Diastereomers	12.634	12009543	74.87
6		13.166	282876	1.76
7		13.540	815690	5.09
8		13.891	168922	1.05

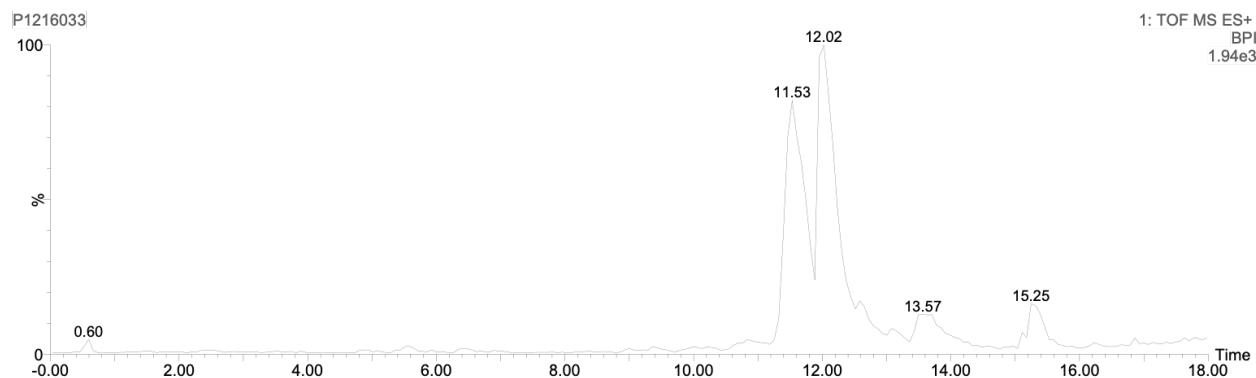


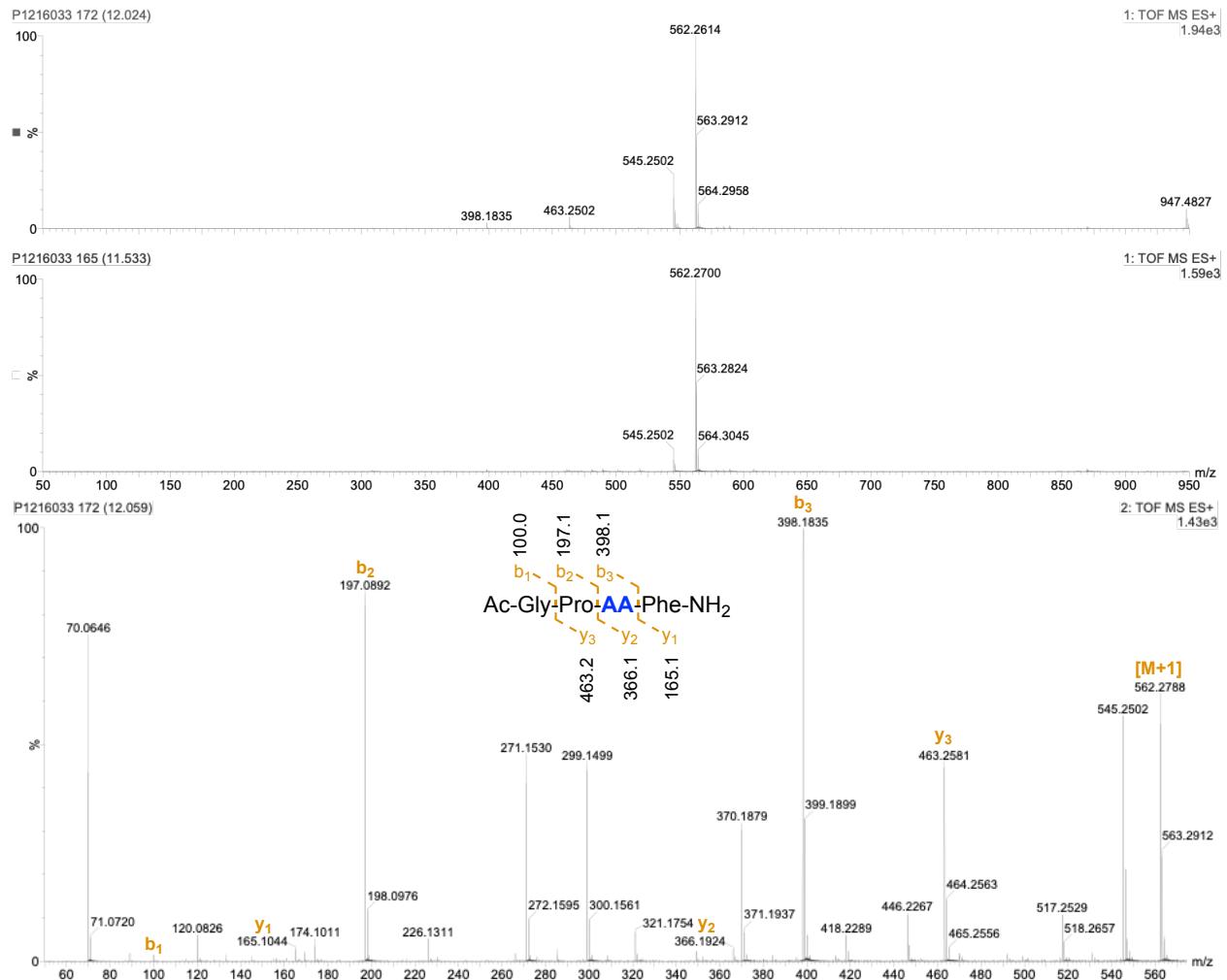


11C': MW = 561.6, Purity = 88.2%, Yield = 17.5% [0.23 mg]

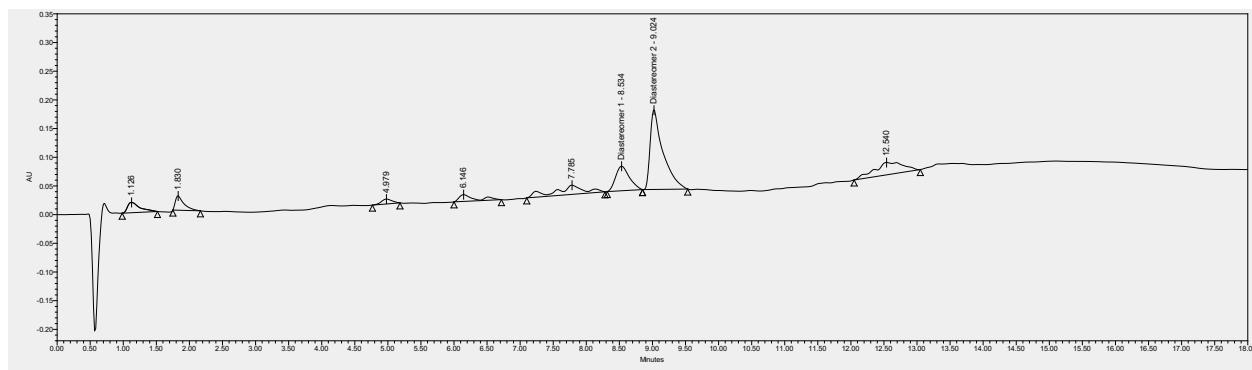
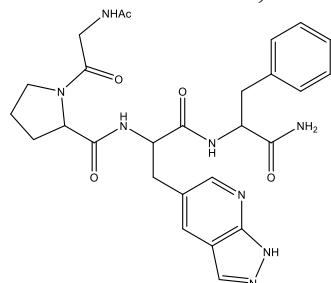


	Name	Retention Time	Area	% Area
1		2.724	327793	2.43
2		12.161	105623	0.78
3	Diastereomers	12.773	11878770	88.20
4		13.553	1156521	8.59

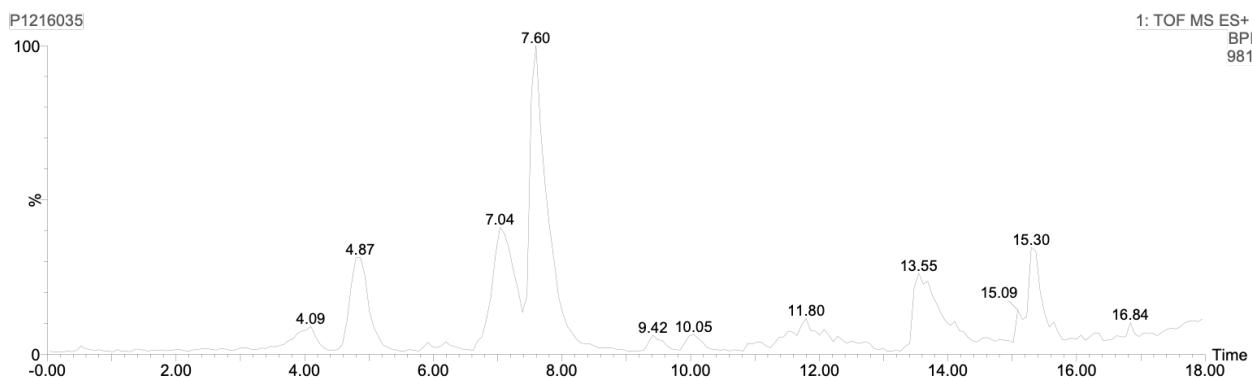


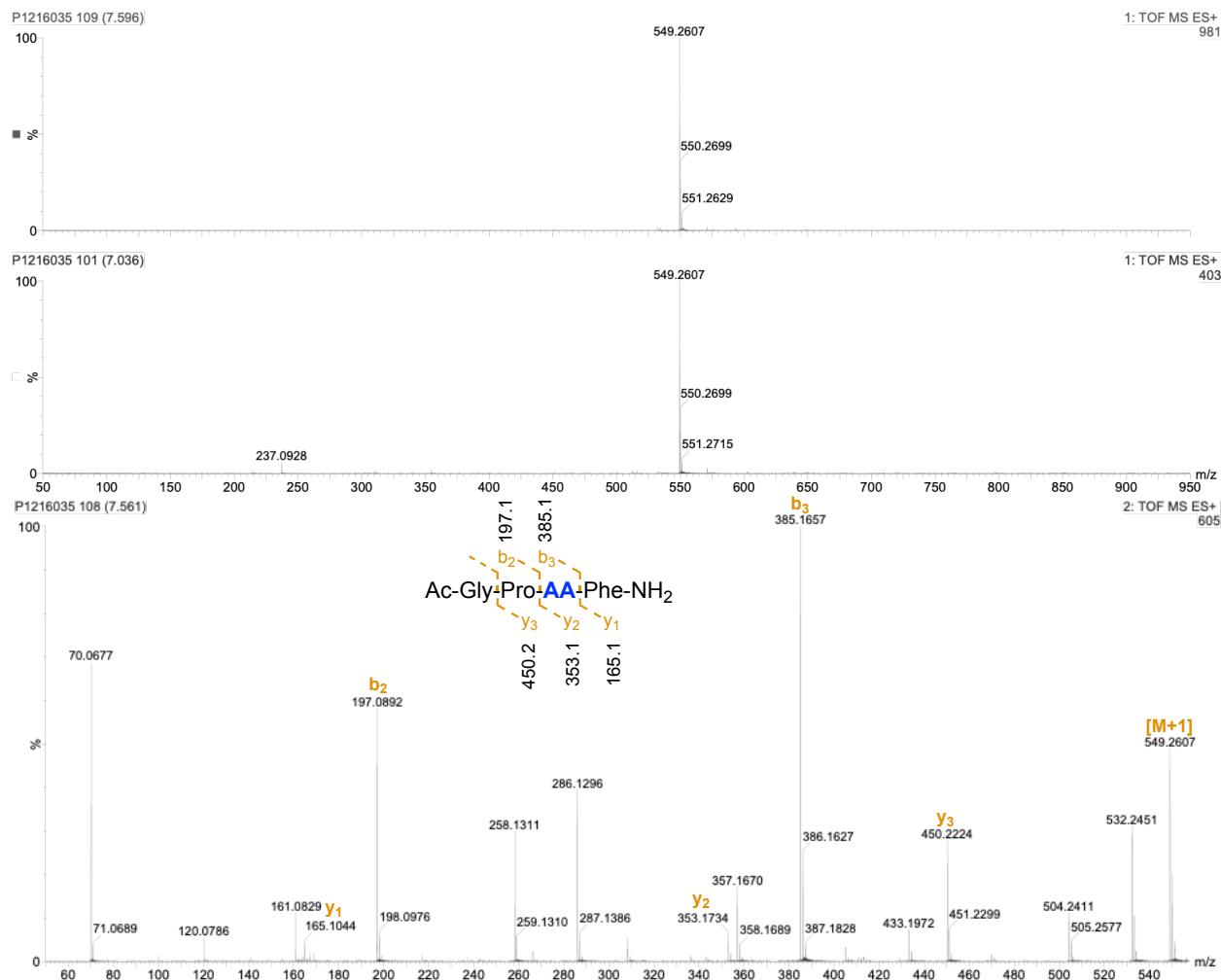


12C': MW = 548.6, Purity = 57.3%, Yield = 3.0% [0.038 mg]

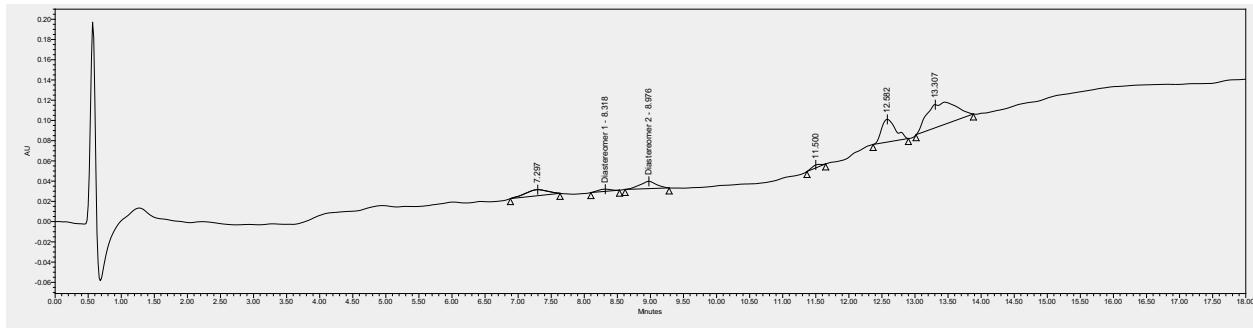
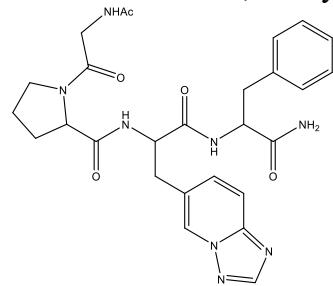


	Name	Retention Time	Area	% Area
1		1.126	235936	5.41
2		1.830	225089	5.16
3		4.979	96237	2.21
4		6.146	194459	4.46
5		7.785	473400	10.85
6	Diastereomer 1	8.534	618425	14.17
7	Diastereomer 2	9.024	1879938	43.08
8		12.540	640590	14.68

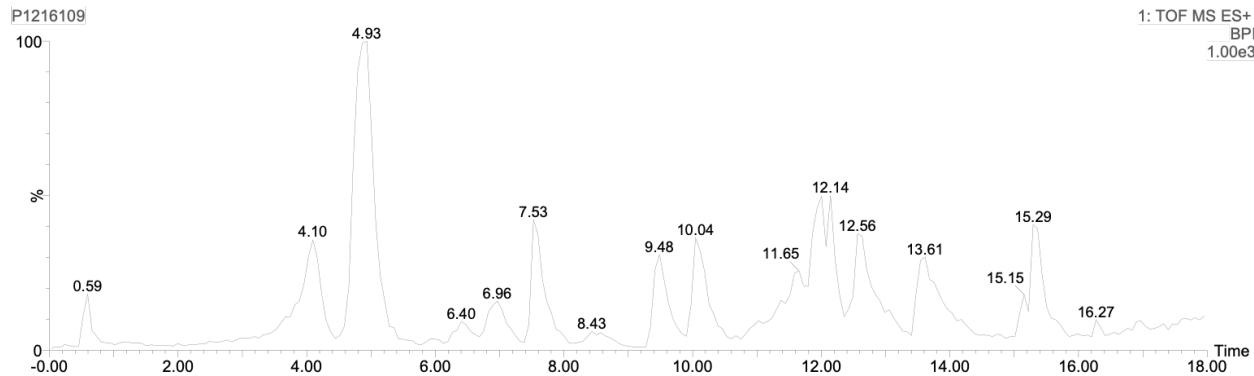


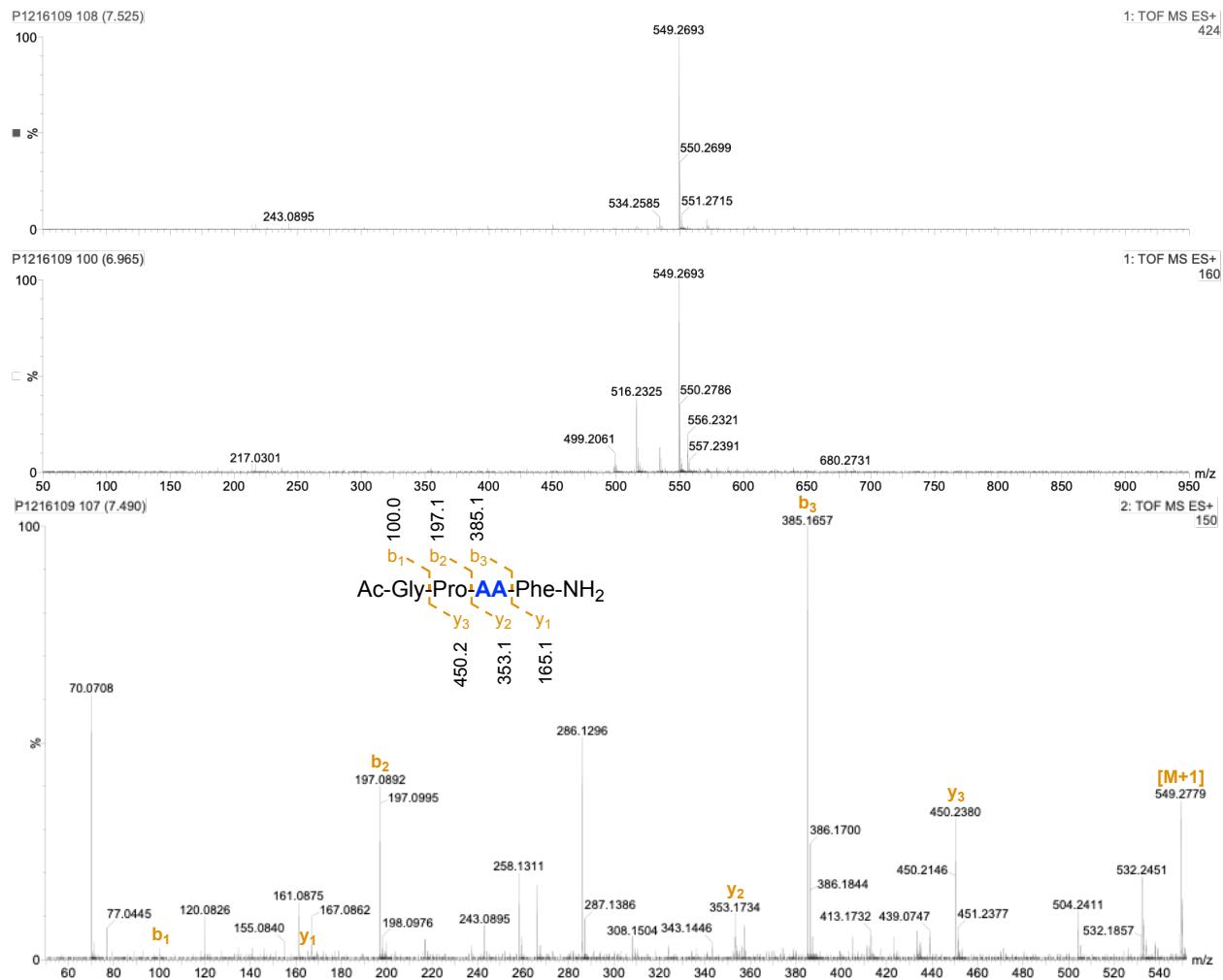


1D': MW = 548.6, Purity = 11.5%, Yield = 0.23% [0.003 mg]

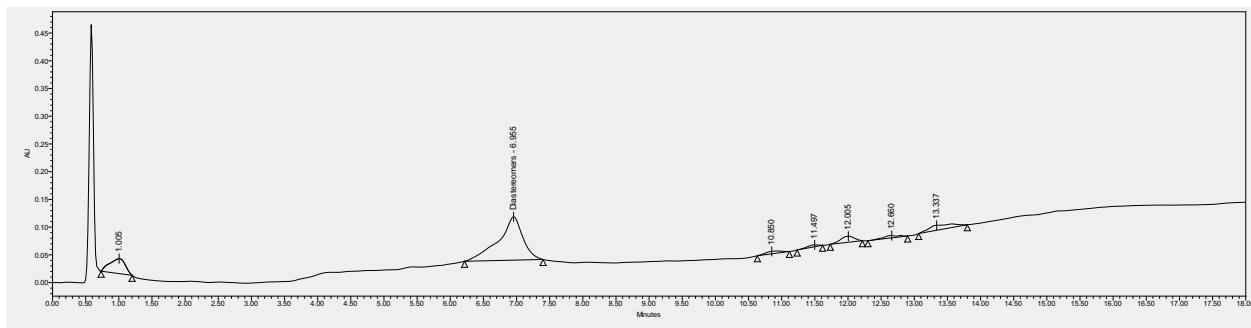
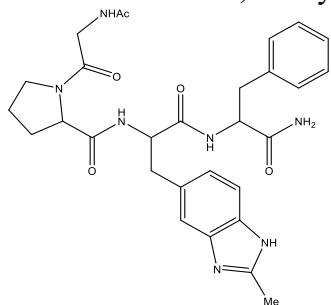


	Name	Retention Time	Area	% Area
1		7.297	139899	10.29
2	Diastereomer 1	8.318	29730	2.19
3	Diastereomer 2	8.976	126595	9.31
4		11.500	26953	1.98
5		12.582	331263	24.36
6		13.307	705304	51.87

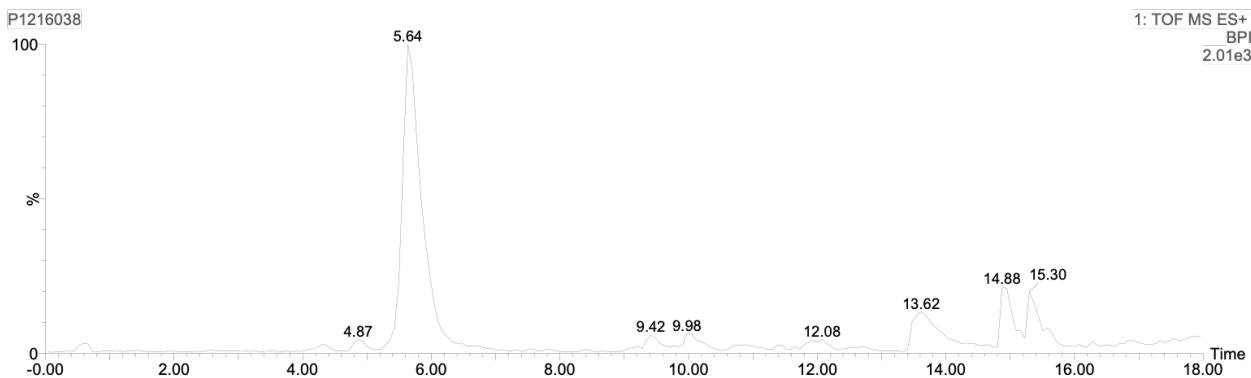


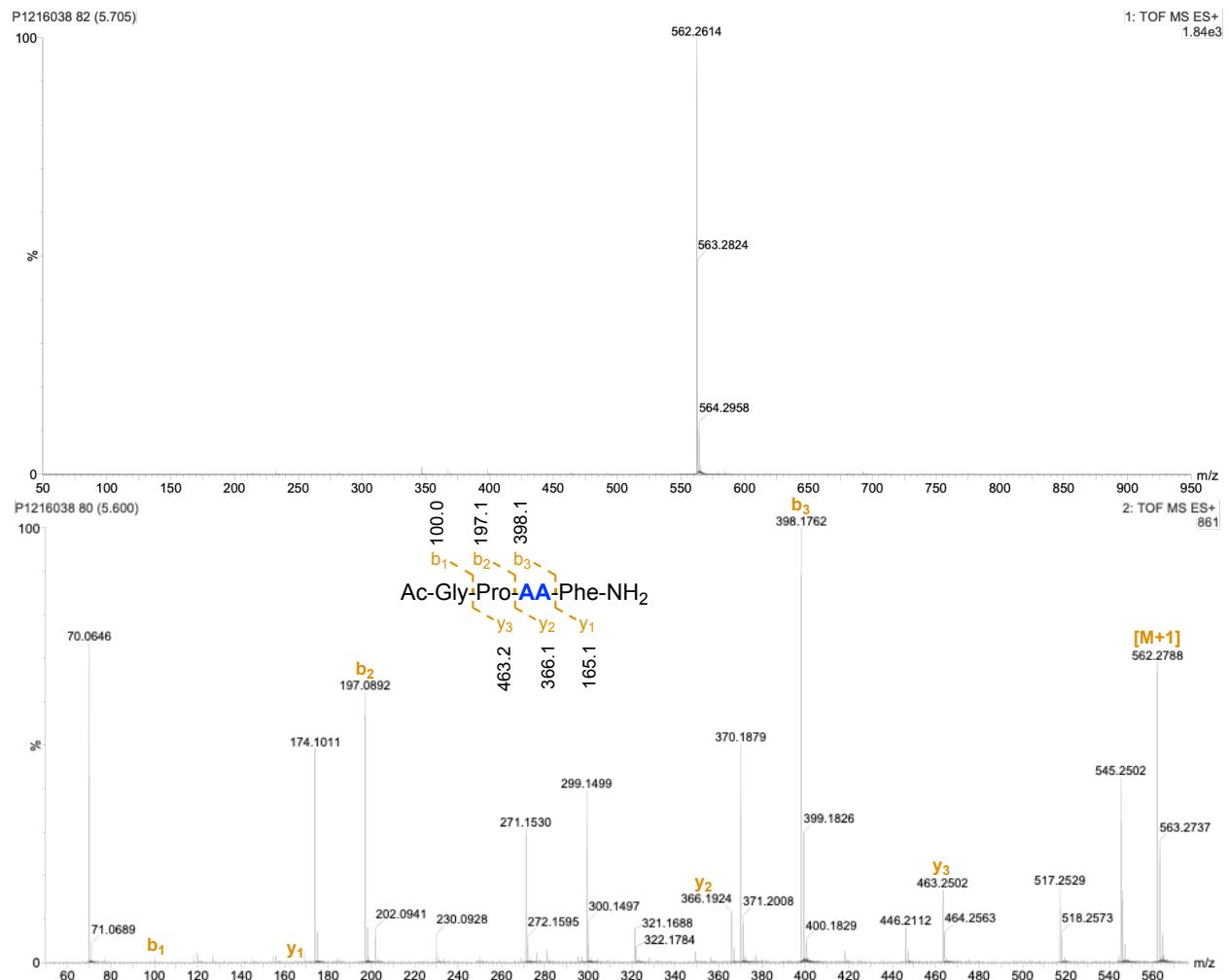


2D': MW = 561.6, Purity = 66.5%, Yield = 3.0% [0.039 mg]

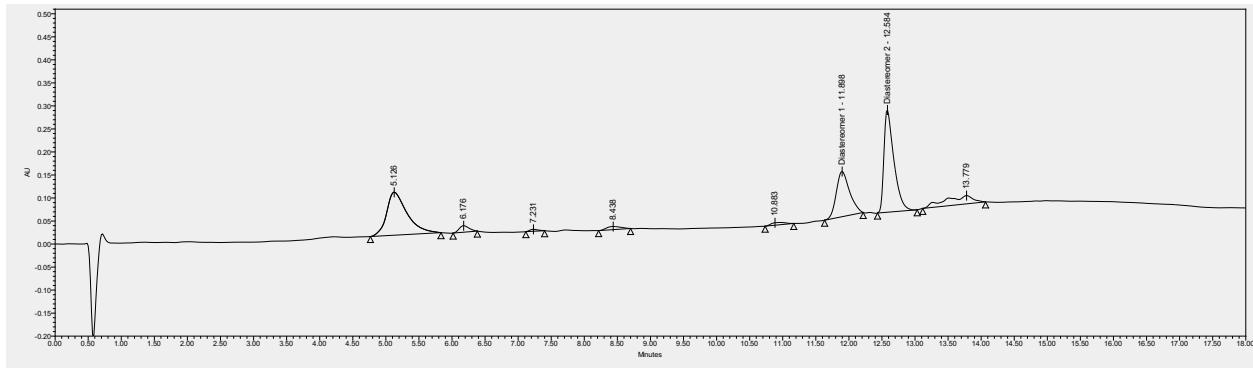
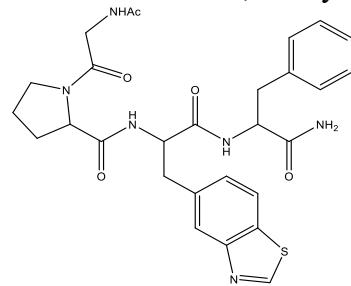


	Name	Retention Time	Area	% Area
1		1.005	437775	14.58
2	Diastereomers	6.955	1997507	66.54
3		10.850	72370	2.41
4		11.497	42924	1.43
5		12.005	161425	5.38
6		12.660	67307	2.24
7		13.337	222700	7.42

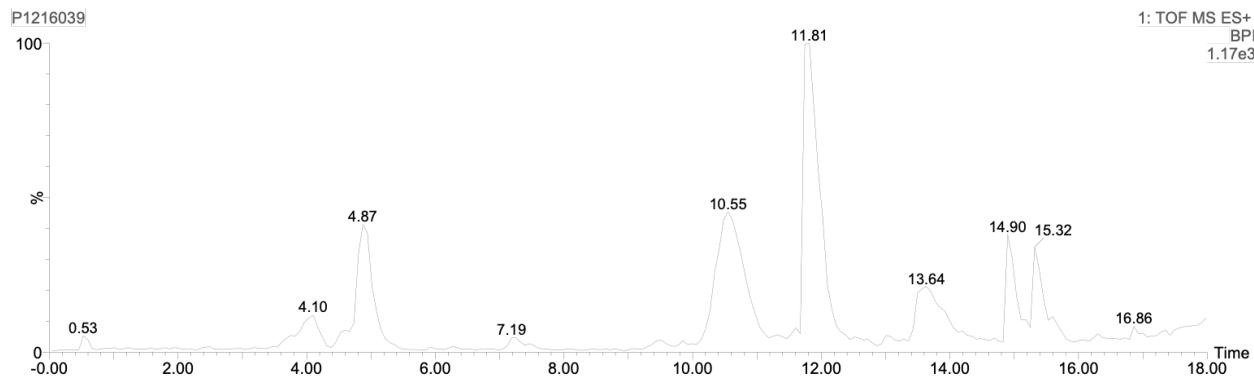


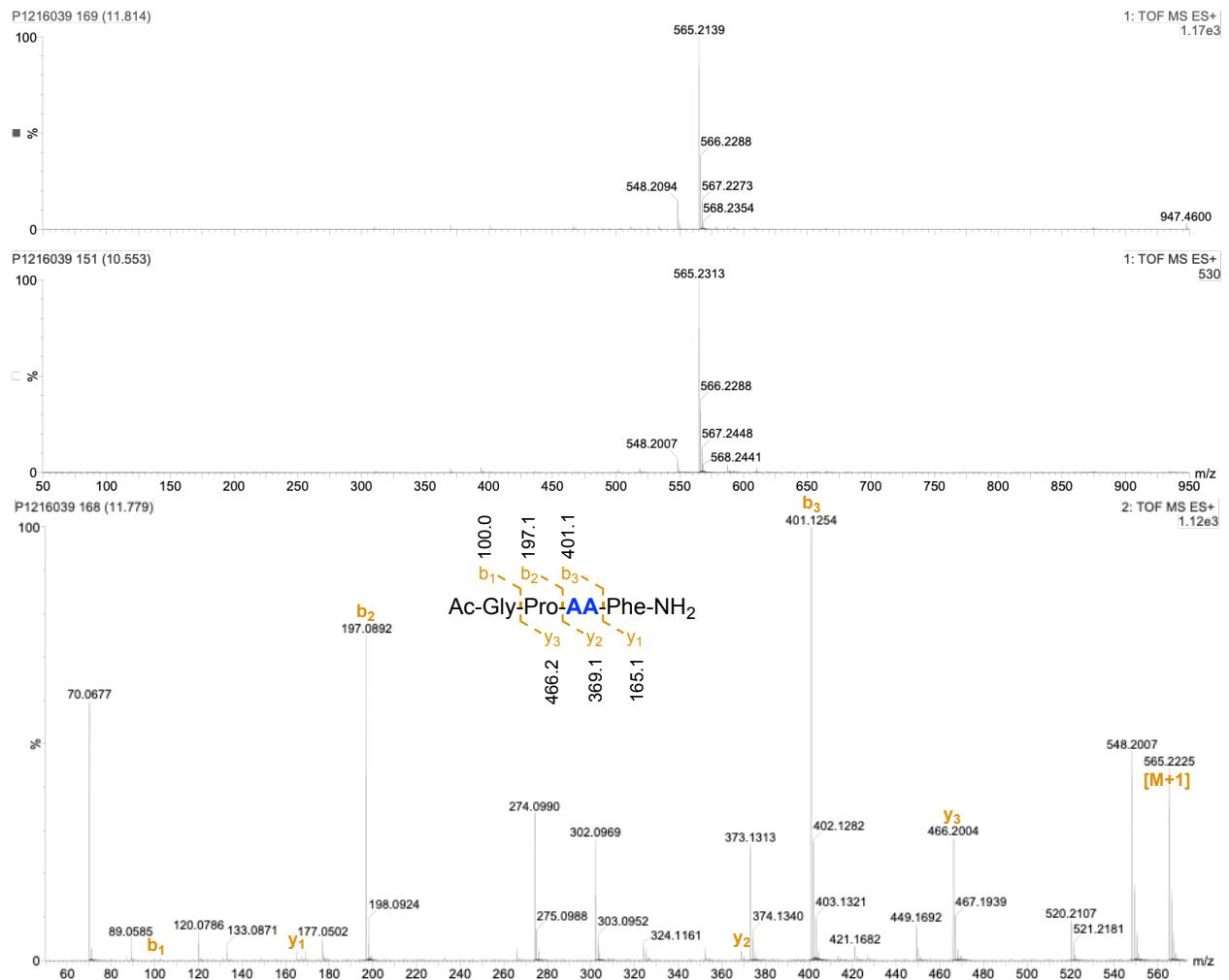


3D': MW = 564.7, Purity = 55.7%, Yield = 5.0% [0.066 mg]

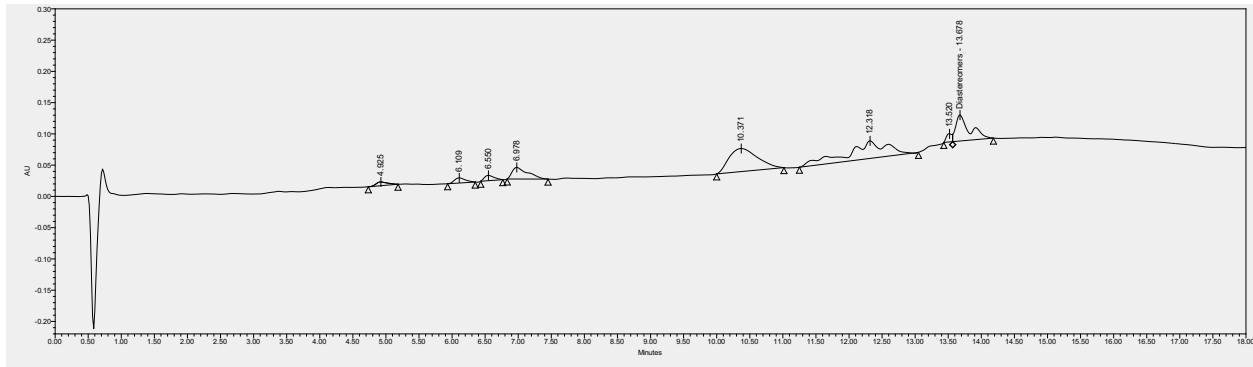
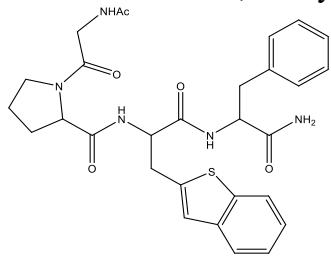


	Name	Retention Time	Area	% Area
1		5.126	2011698	30.35
2		6.176	147197	2.22
3		7.231	37430	0.56
4		8.438	104811	1.58
5		10.883	78565	1.19
6	Diastereomer 1	11.898	1369881	20.67
7	Diastereomer 2	12.584	2319147	34.99
8		13.779	559313	8.44

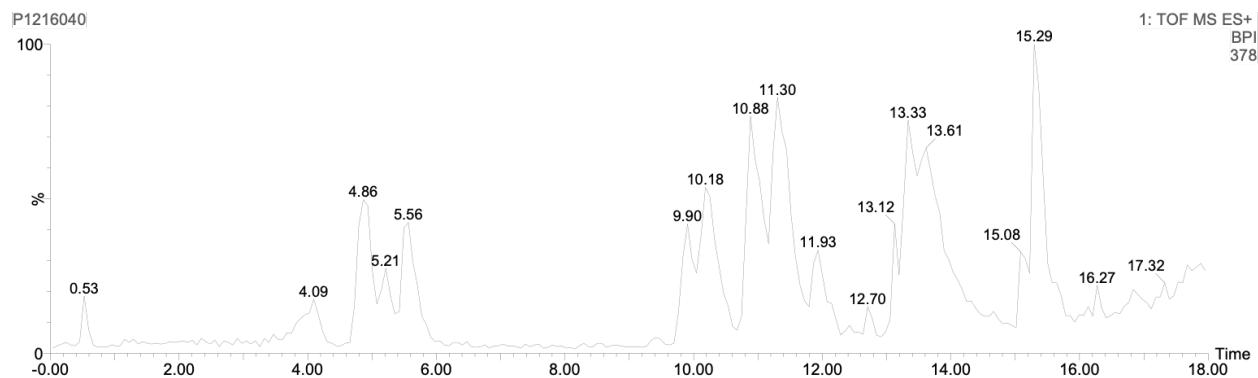


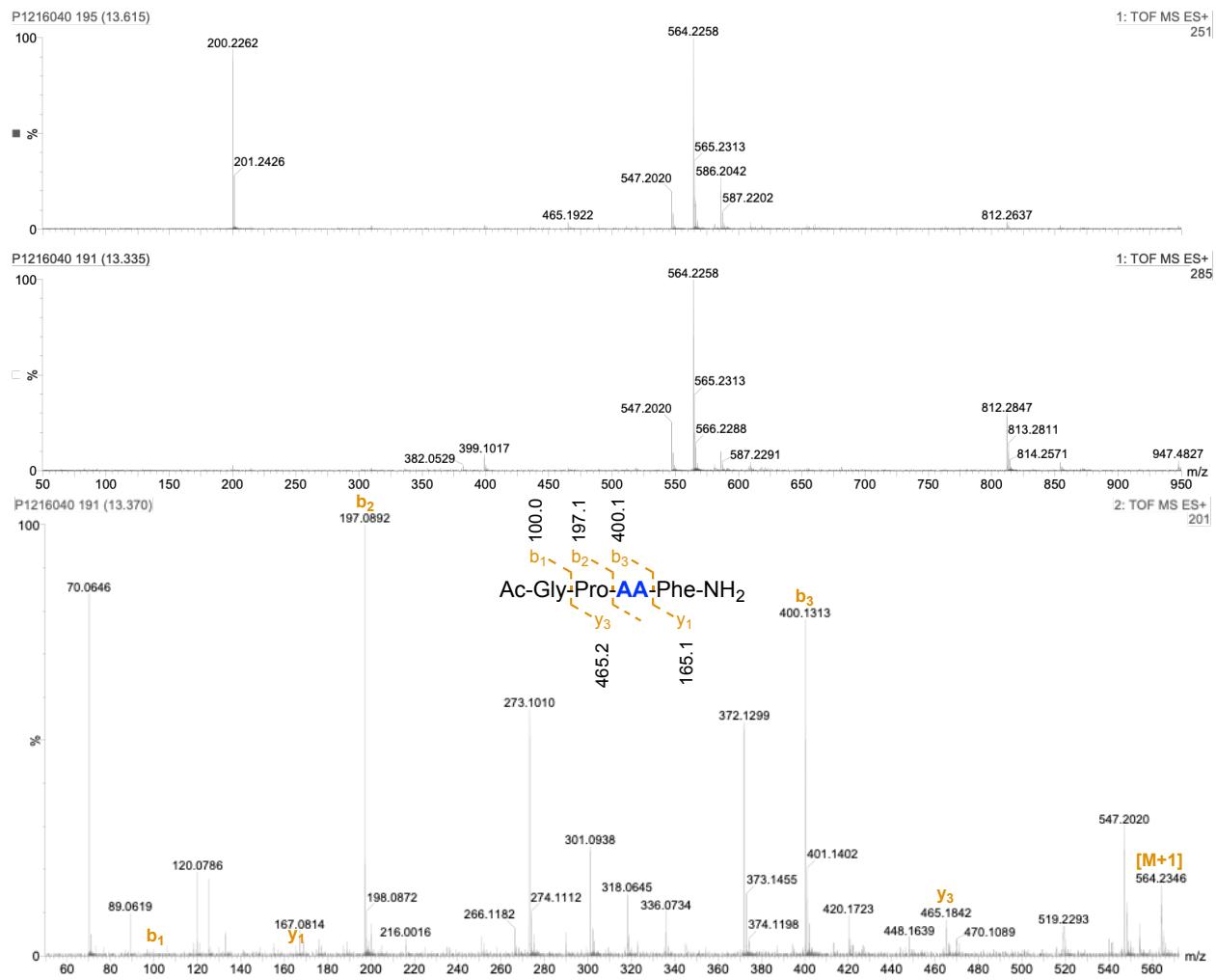


4D': MW = 563.7, Purity = 17.1%, Yield = 0.73% [0.010 mg]

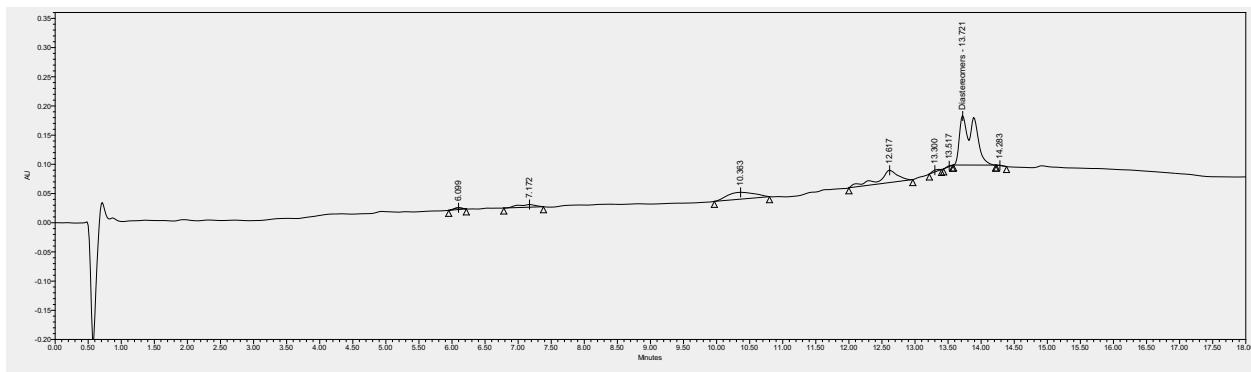
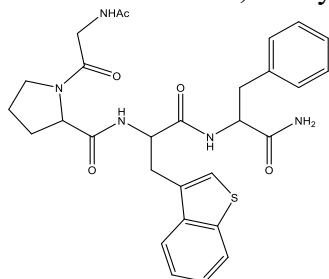


	Name	Retention Time	Area	% Area
1		4.925	72683	2.07
2		6.109	94316	2.68
3		6.550	87350	2.48
4		6.978	308128	8.76
5		10.371	1108002	31.51
6		12.318	1167326	33.20
7		13.520	77965	2.22
8	Diastereomers	13.678	600755	17.08

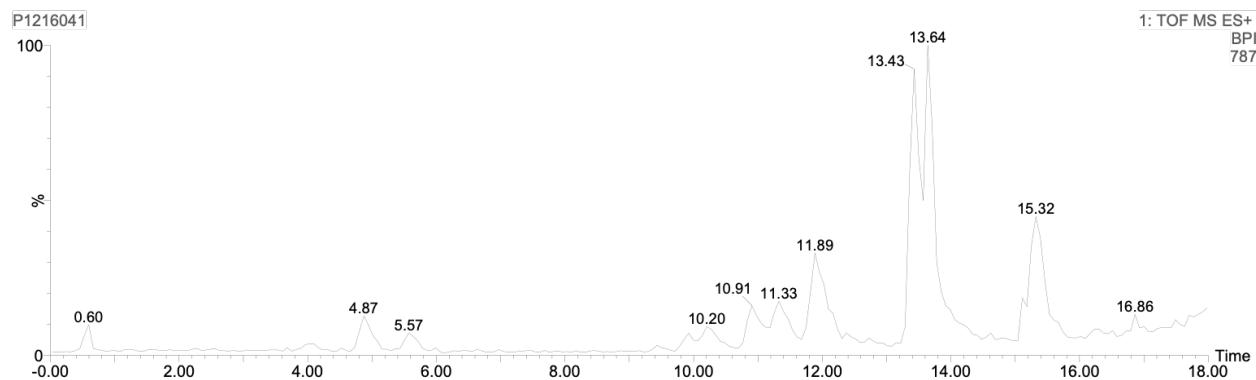


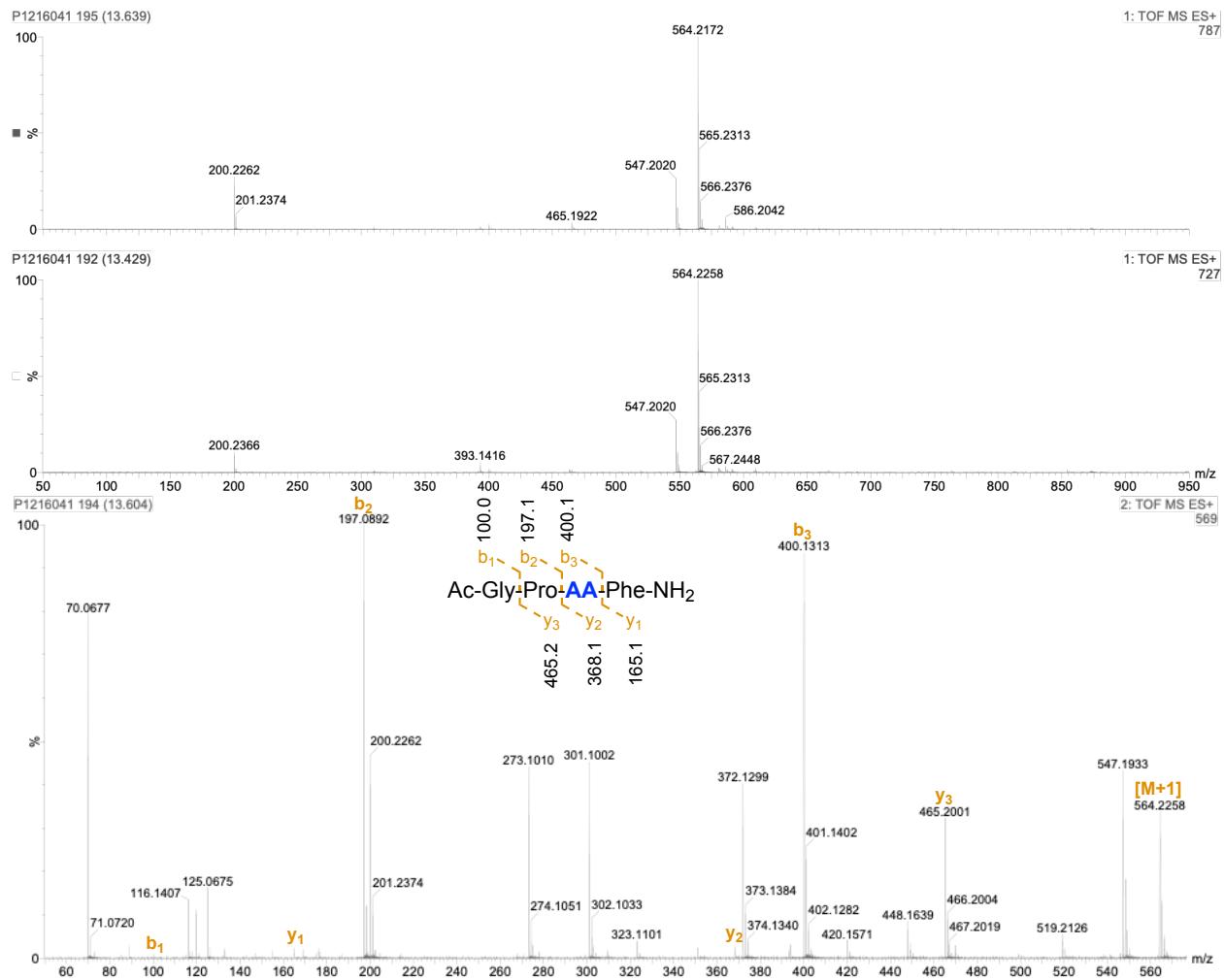


5D': MW = 563.7, Purity = 59.6%, Yield = 1.6% [0.021 mg]

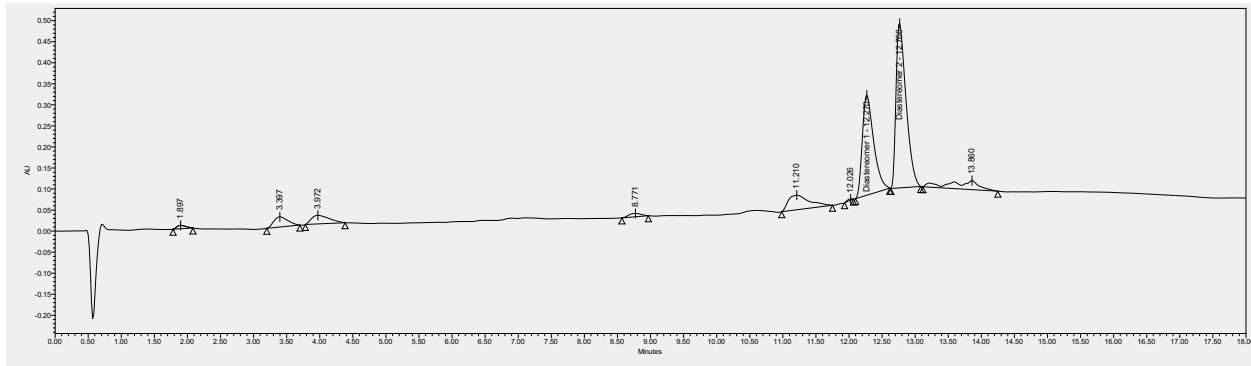
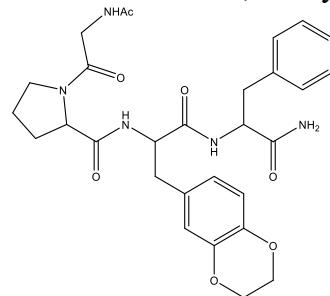


	Name	Retention Time	Area	% Area
1		6.099	25698	1.14
2		7.172	89723	3.98
3		10.363	340361	15.08
4		12.617	425364	18.85
5		13.300	21002	0.93
6		13.517	7119	0.32
7	Diastereomers	13.721	1344578	59.59
8		14.283	2625	0.12

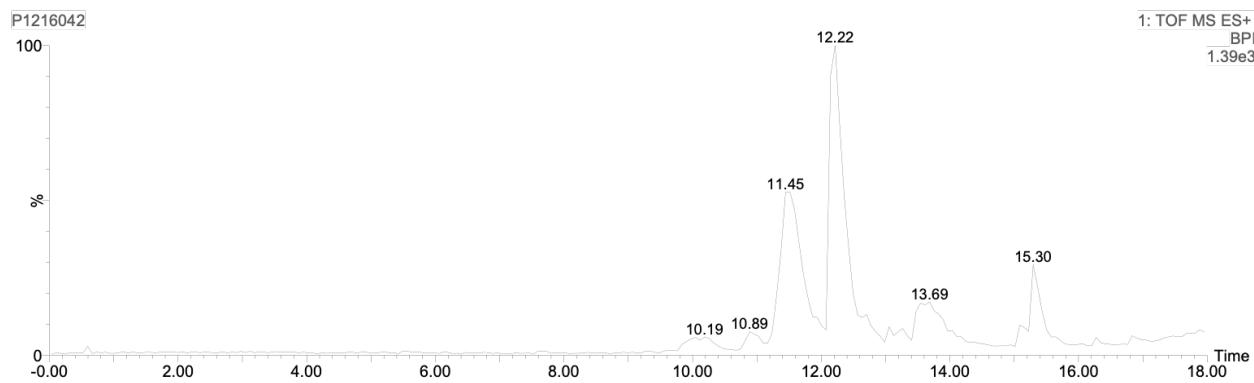


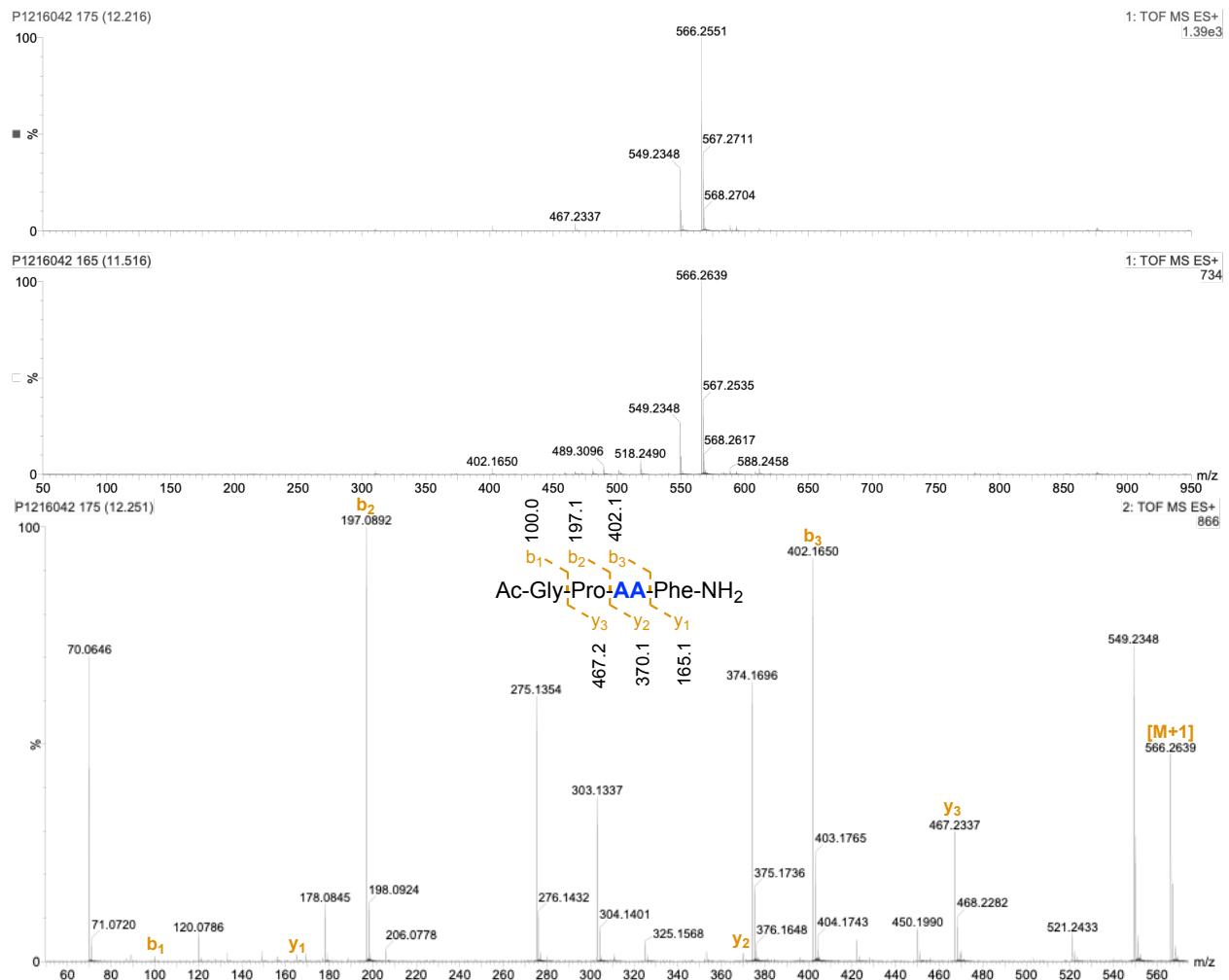


6D': MW = 565.6, Purity = 75.0%, Yield = 18.7% [0.25 mg]

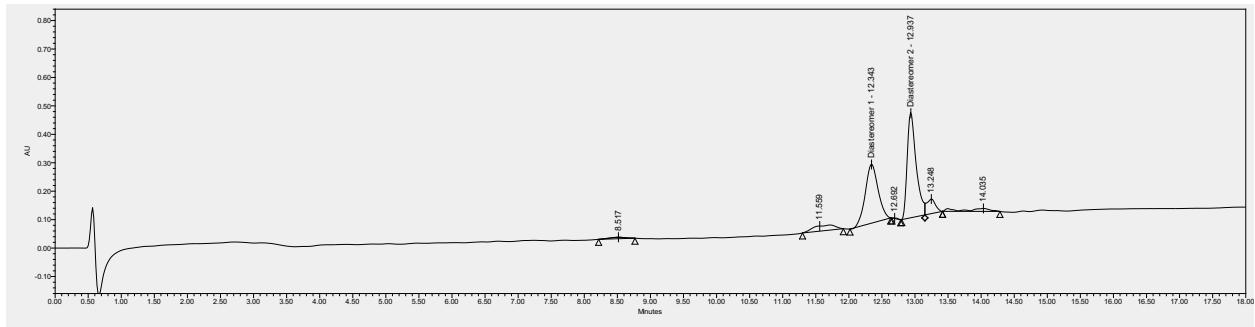
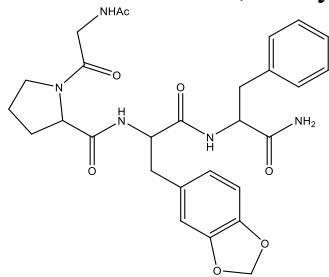


	Name	Retention Time	Area	% Area
1		1.897	77417	0.84
2		3.397	358664	3.87
3		3.972	379586	4.10
4		8.771	104258	1.13
5		11.210	769781	8.31
6		12.026	18190	0.20
7	Diastereomer 1	12.270	2835935	30.61
8	Diastereomer 2	12.766	4110785	44.37
9		13.860	609721	6.58

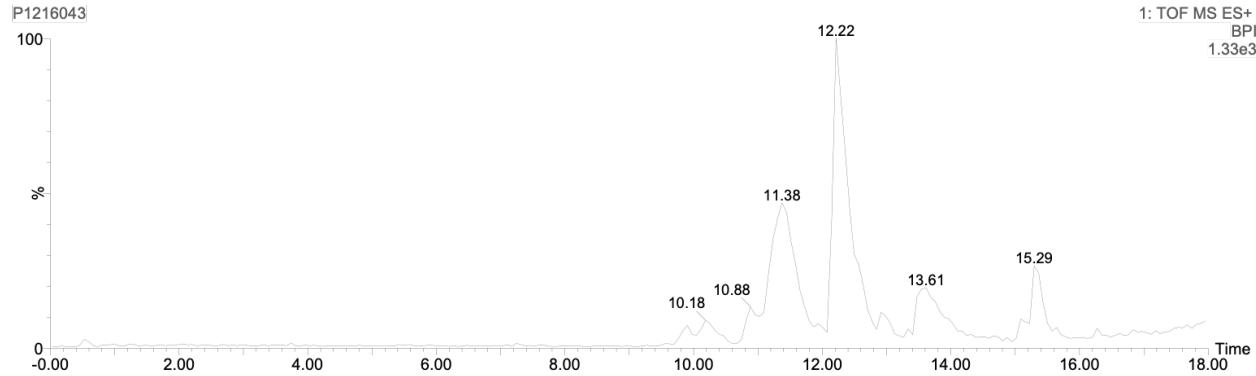


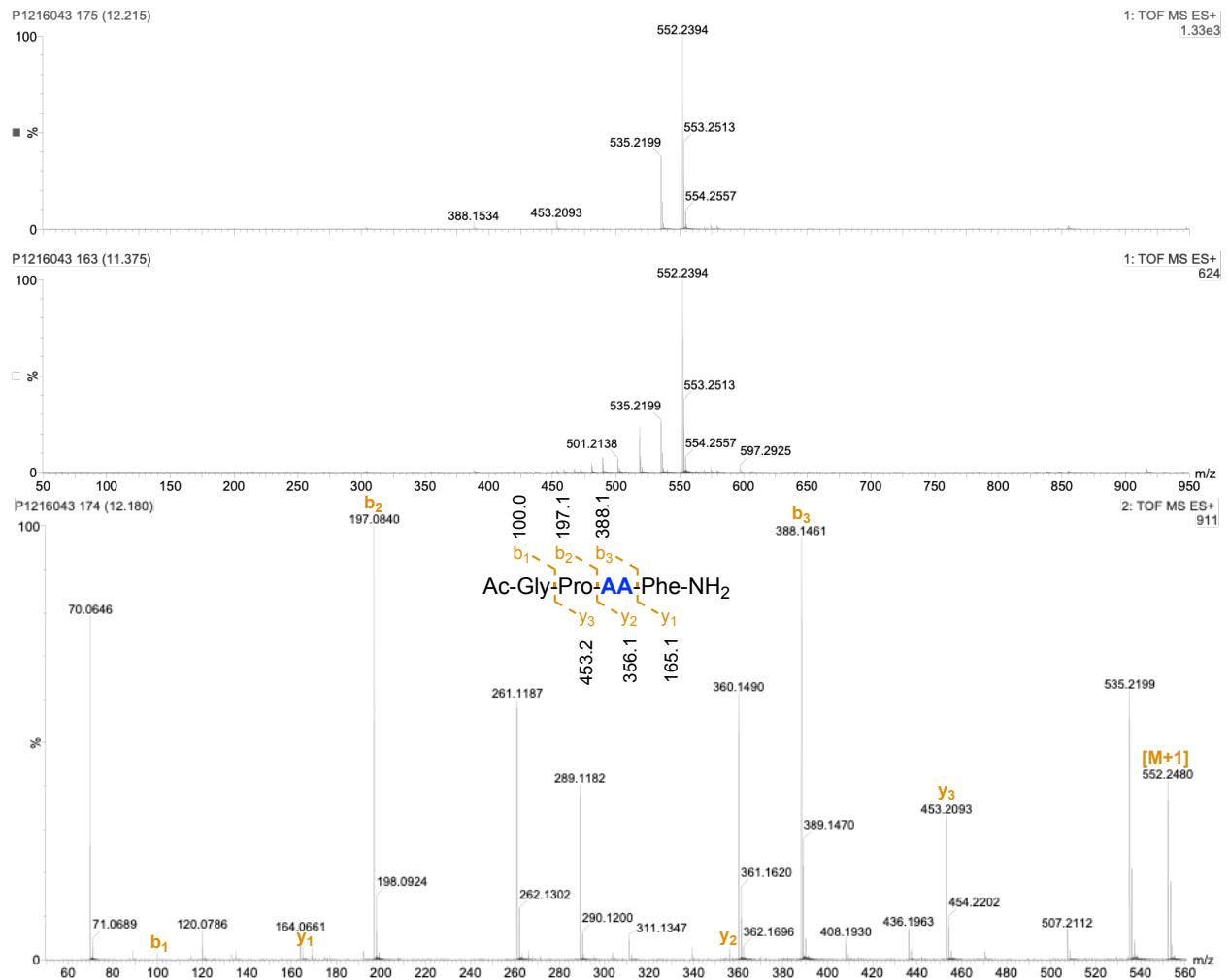


7D': MW = 551.6, Purity = 82.8%, Yield = 16.8% [0.22 mg]

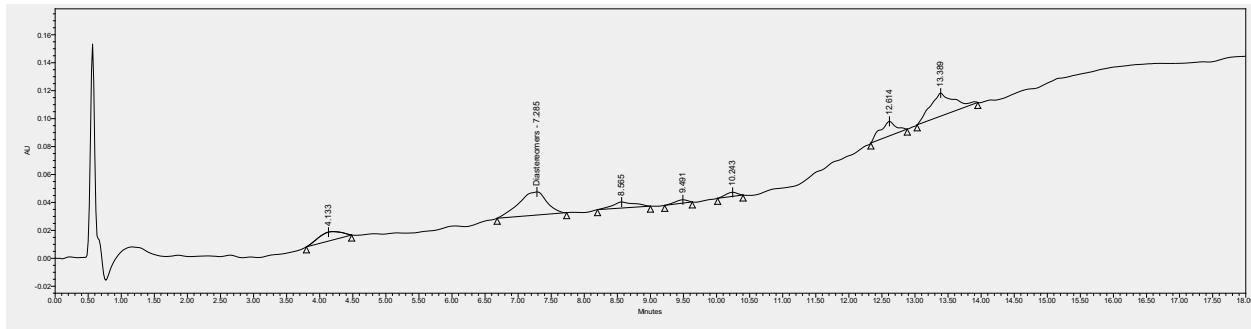
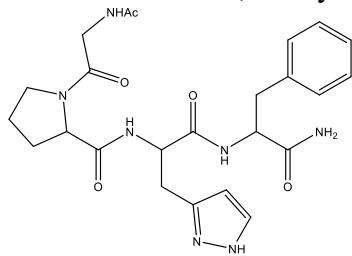


	Name	Retention Time	Area	% Area
1		8.517	88885	1.17
2		11.559	433533	5.73
3	Diastereomer 1	12.343	2801638	37.03
4		12.692	13378	0.18
5	Diastereomer 2	12.937	3459808	45.73
6		13.248	485206	6.41
7		14.035	282896	3.74

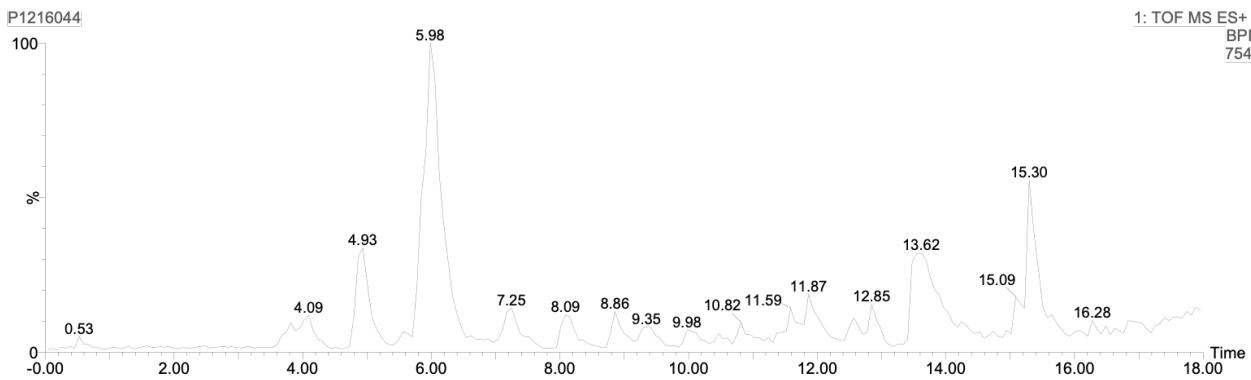


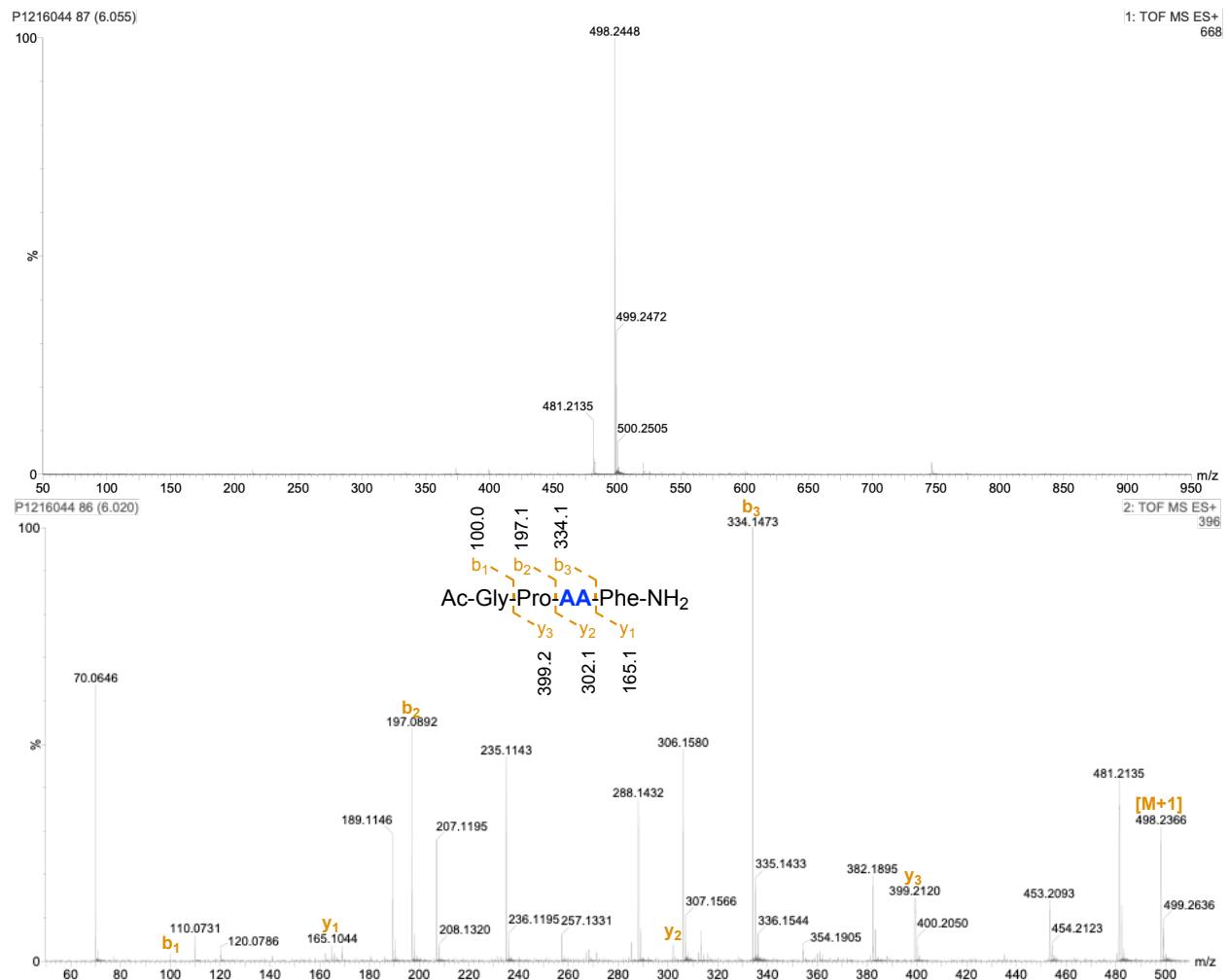


8D': MW = 497.6, Purity = 35.3%, Yield = 1.3% [0.015 mg]

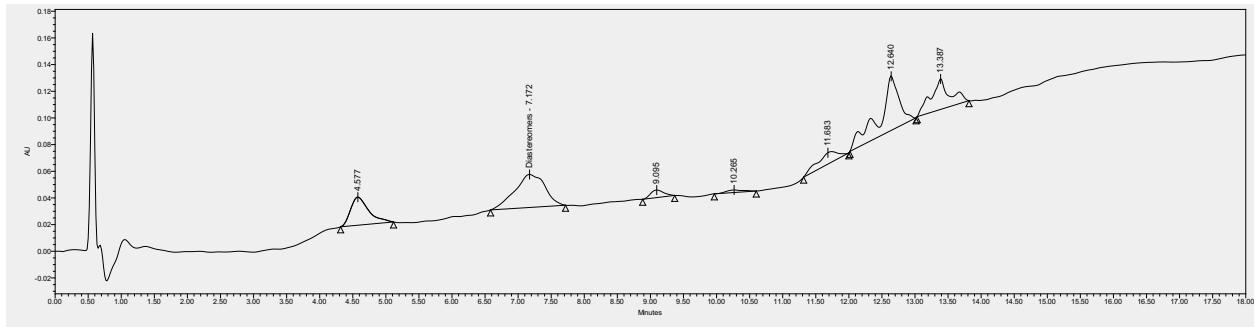
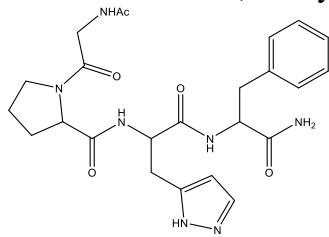


	Name	Retention Time	Area	% Area
1		4.133	153453	11.01
2	Diastereomers	7.285	492081	35.31
3		8.565	99977	7.17
4		9.491	31815	2.28
5		10.243	36572	2.62
6		12.614	176606	12.67
7		13.389	403133	28.93

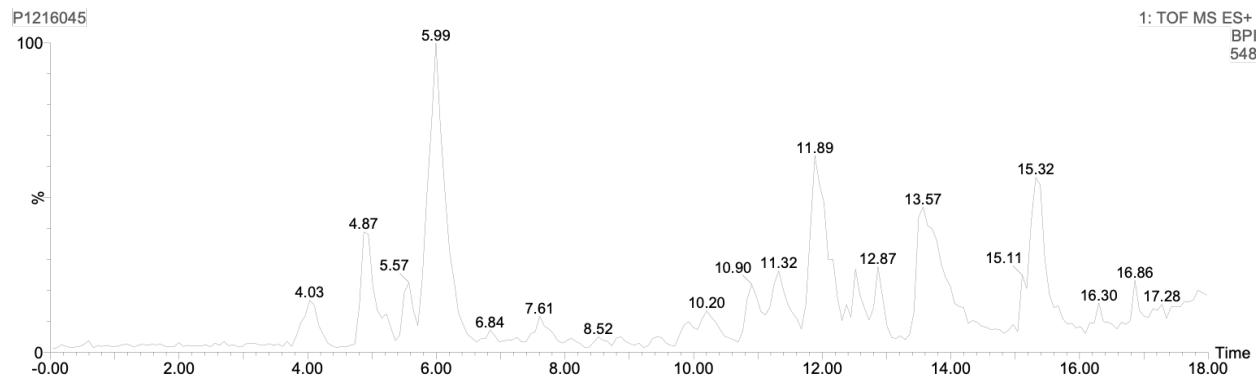


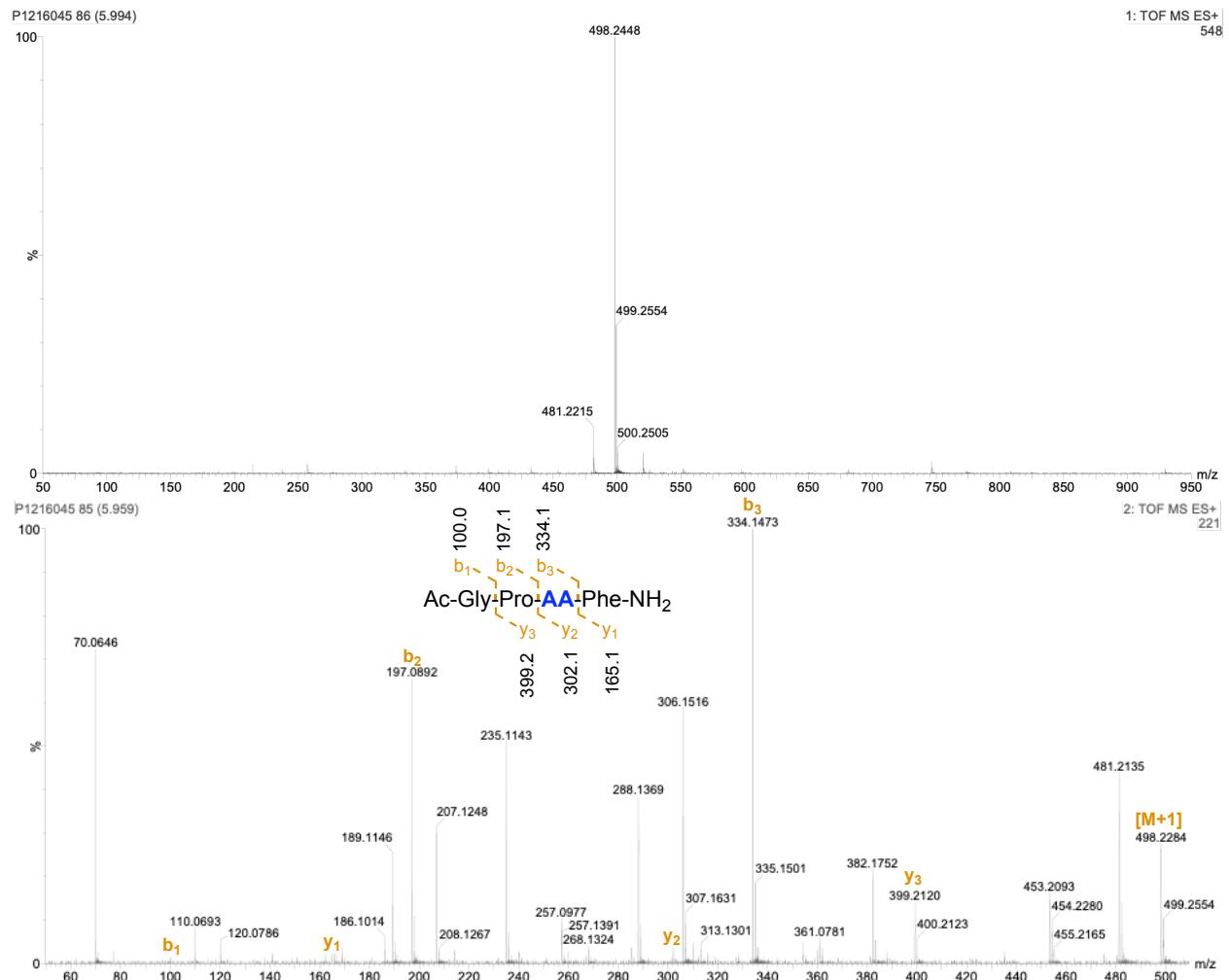


9D': MW = 497.6, Purity = 28.3%, Yield = 2.1% [0.025 mg]

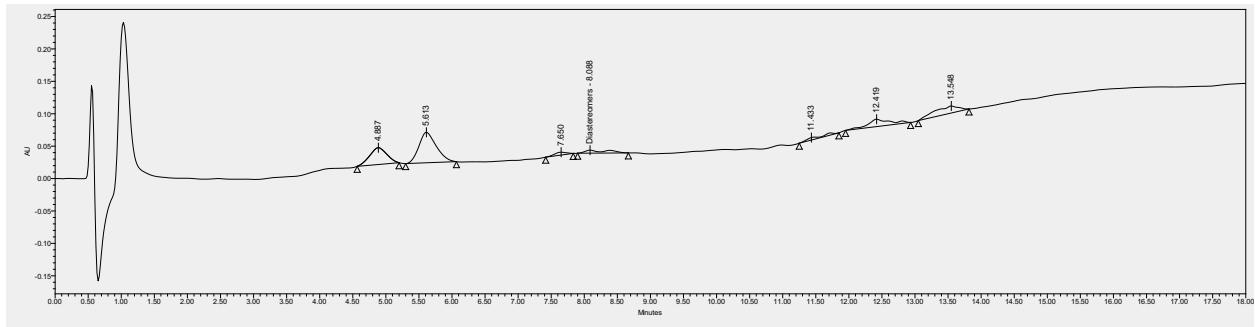
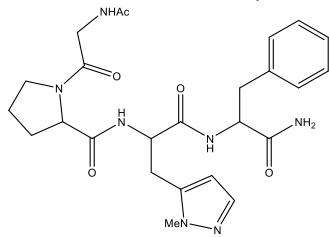


	Name	Retention Time	Area	% Area
1		4.577	403774	14.53
2	Diastereomers	7.172	785457	28.27
3		9.095	78609	2.83
4		10.265	34383	1.24
5		11.683	187582	6.75
6		12.640	835474	30.07
7		13.387	453567	16.32

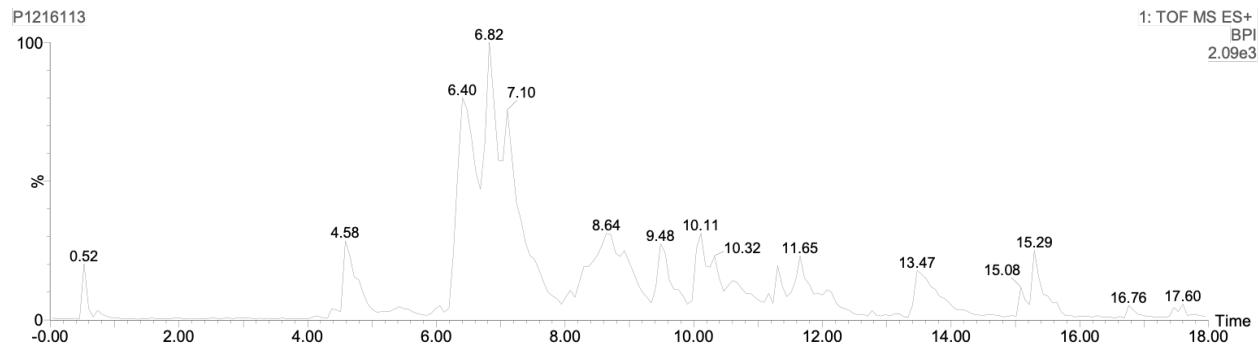


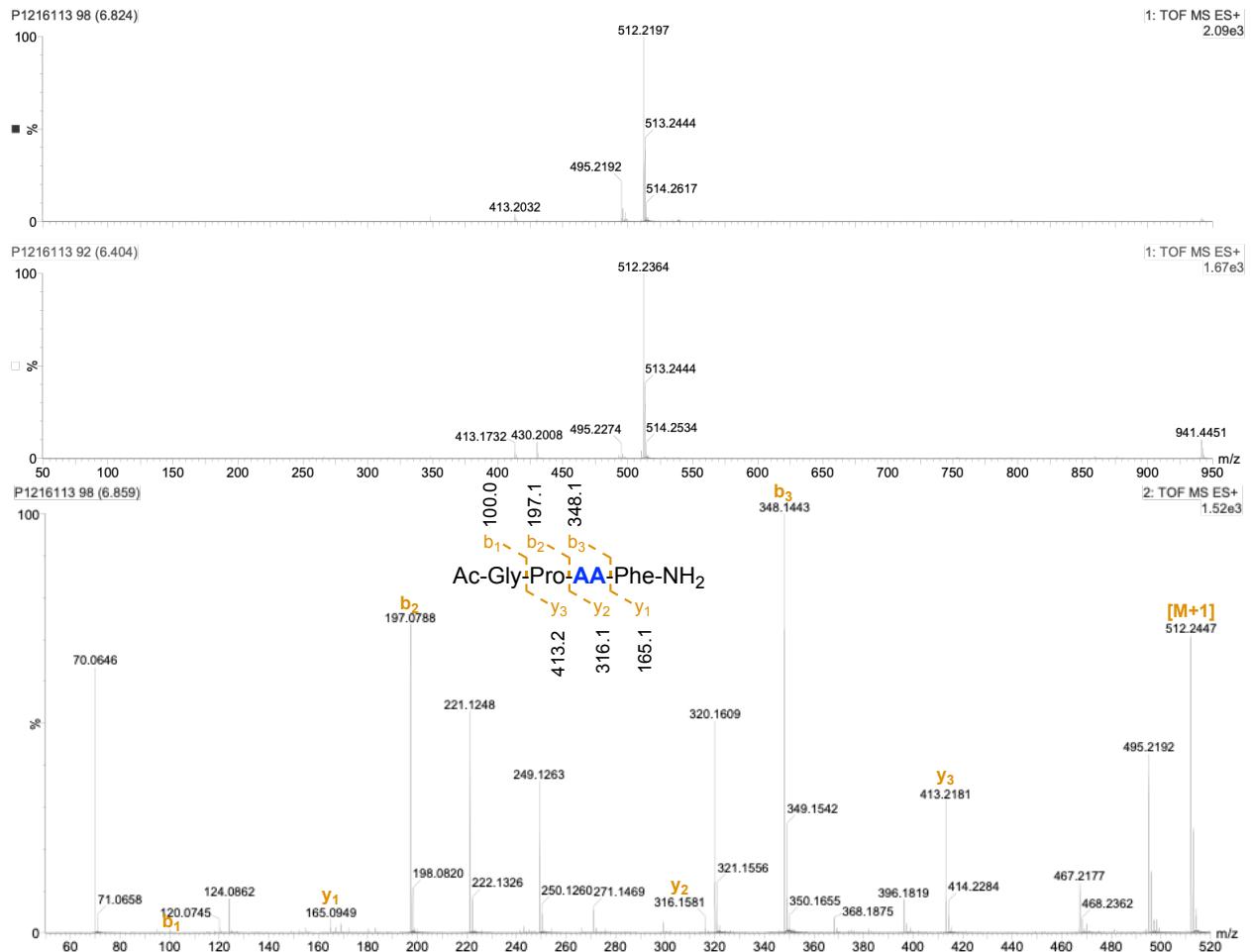


10D': MW = 511.6, Purity = 5.2%, Yield = 0.29% [0.004 mg]

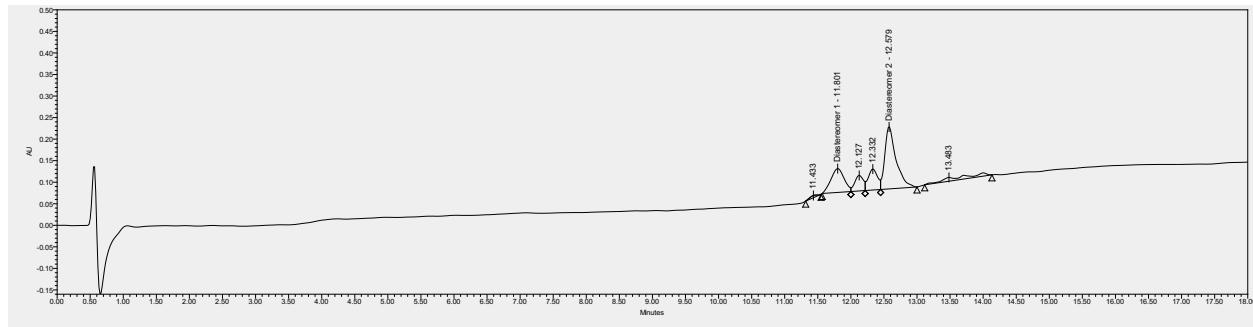
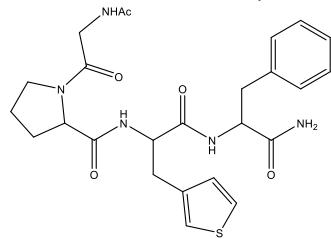


	Name	Retention Time	Area	% Area
1		4.887	468431	22.07
2		5.613	859604	40.49
3		7.650	55995	2.64
4	Diastereomers	8.088	109378	5.15
5		11.433	71822	3.38
6		12.419	260055	12.25
7		13.548	297654	14.02

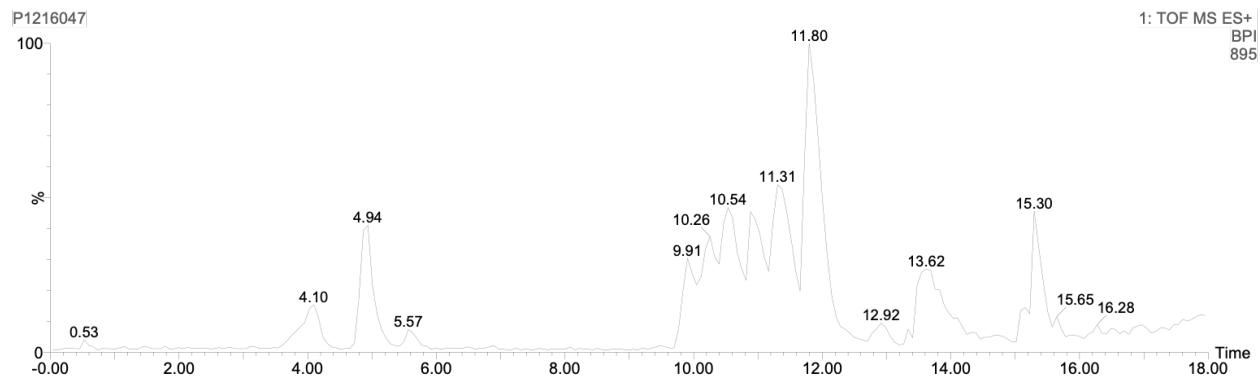


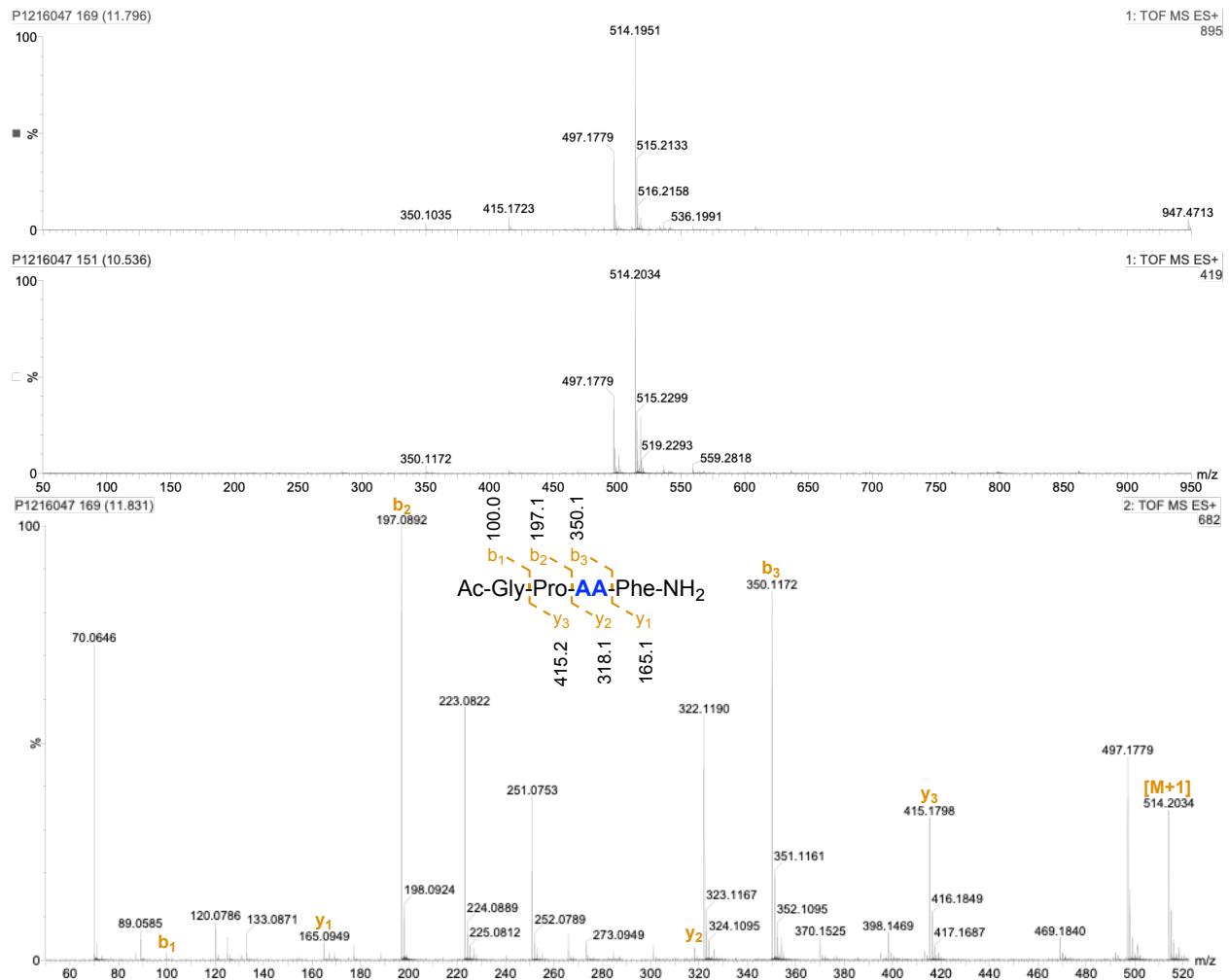


11D': MW = 513.6, Purity = 68.8%, Yield = 7.5% [0.090 mg]

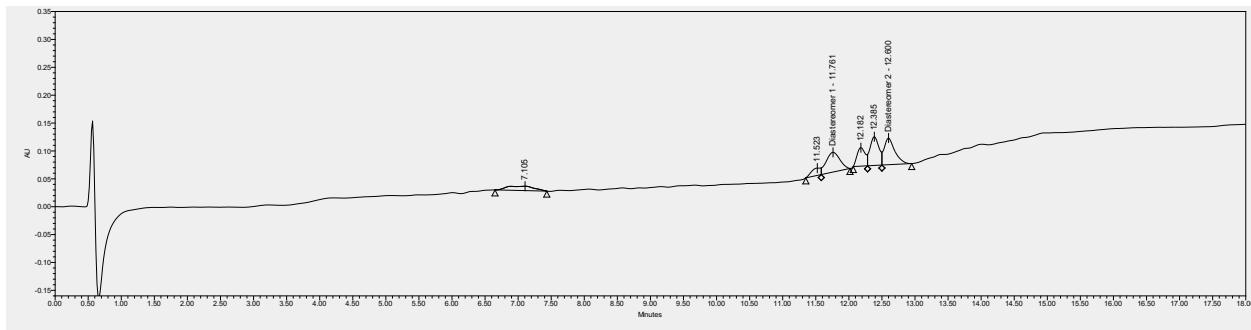
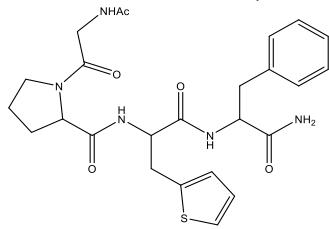


	Name	Retention Time	Area	% Area
1		11.433	38481	1.08
2	Diastereomer 1	11.801	765423	21.44
3		12.127	324157	9.08
4		12.332	480241	13.45
5	Diastereomer 2	12.579	1691680	47.39
6		13.483	269945	7.56

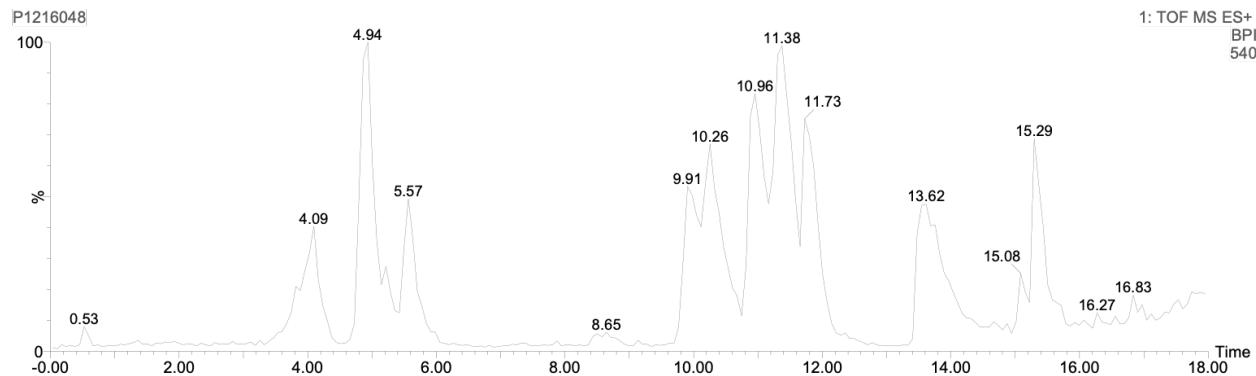


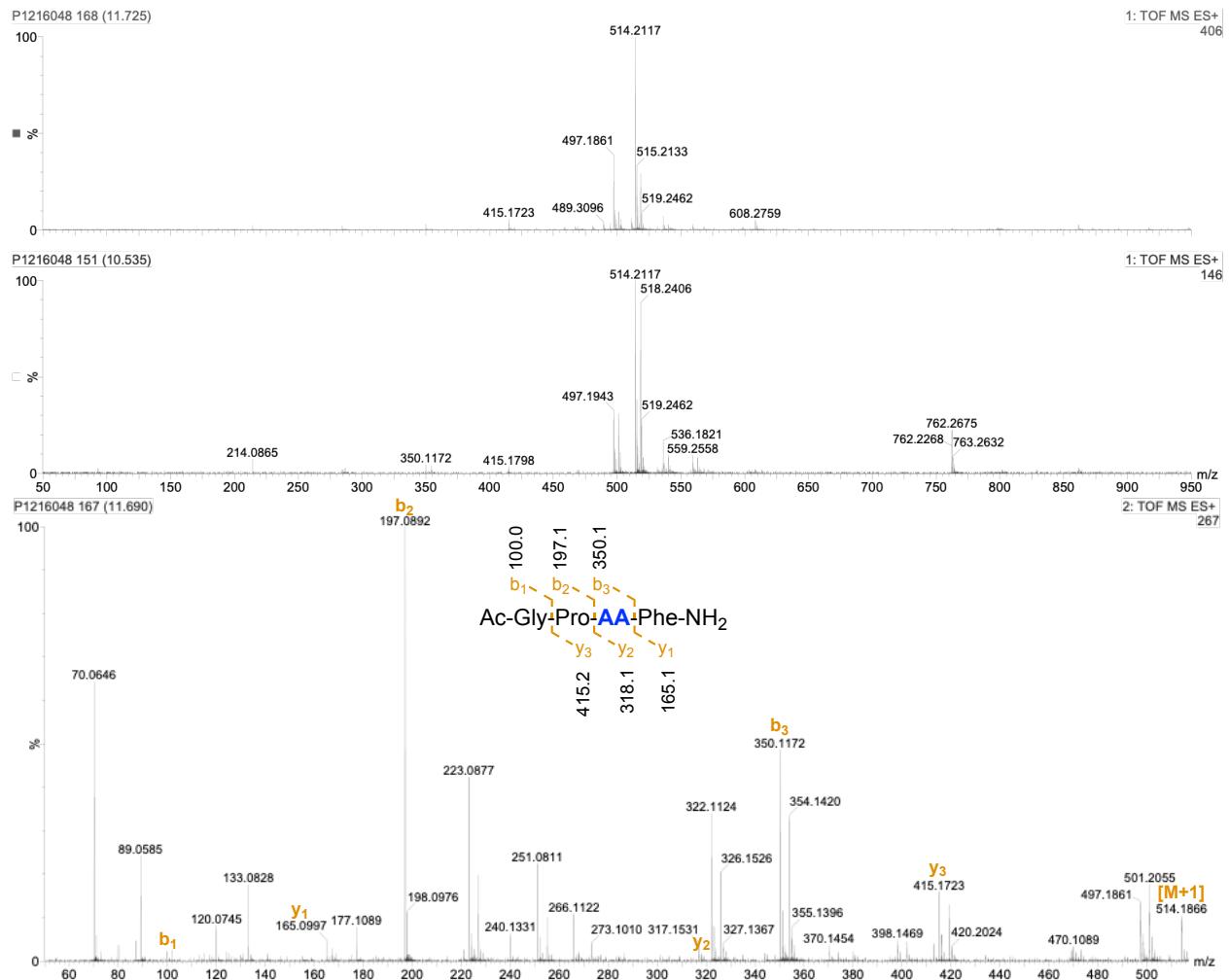


12D': MW = 513.6, Purity = 48.4%, Yield = 3.2% [0.038 mg]

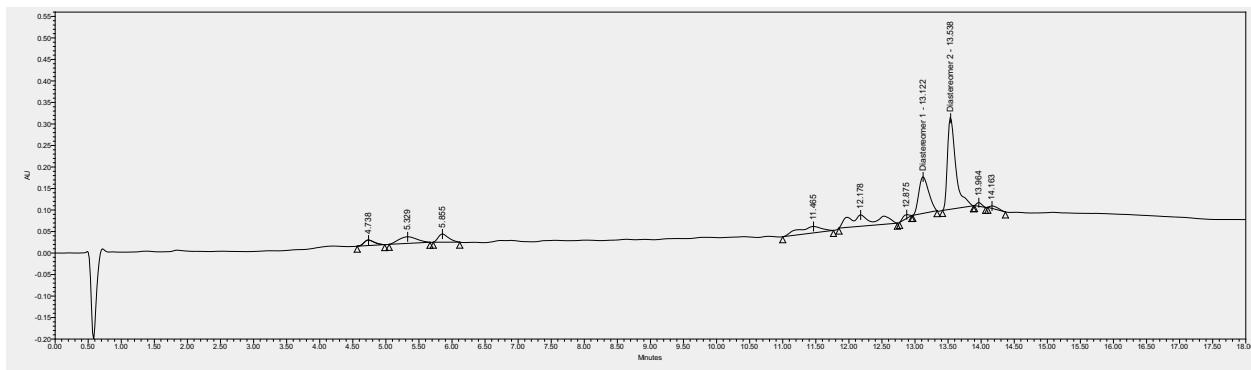
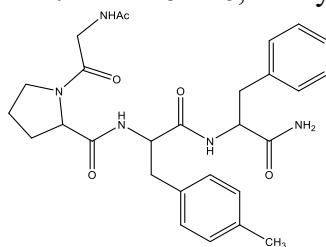


	Name	Retention Time	Area	% Area
1		7.105	221663	10.23
2		11.523	129609	5.98
3	Diastereomer 1	11.761	512350	23.64
4		12.182	287272	13.26
5		12.385	480605	22.18
6	Diastereomer 2	12.600	535542	24.71

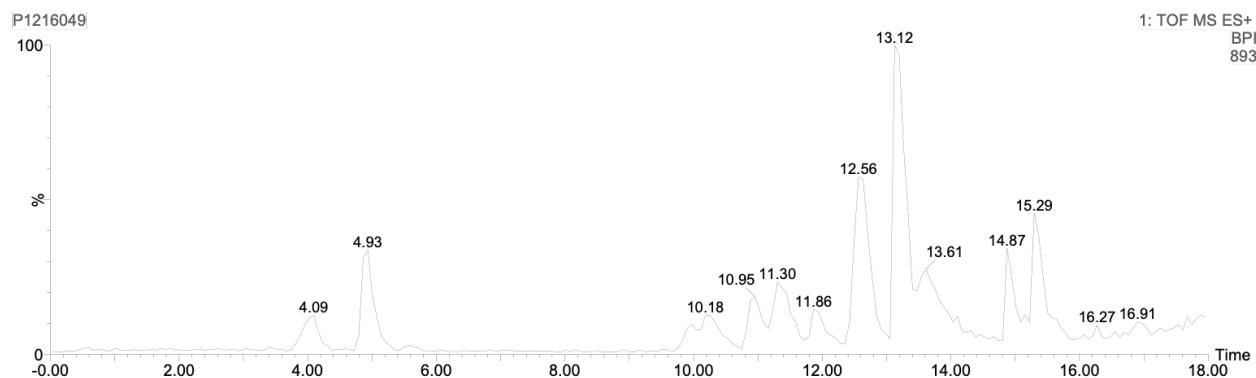


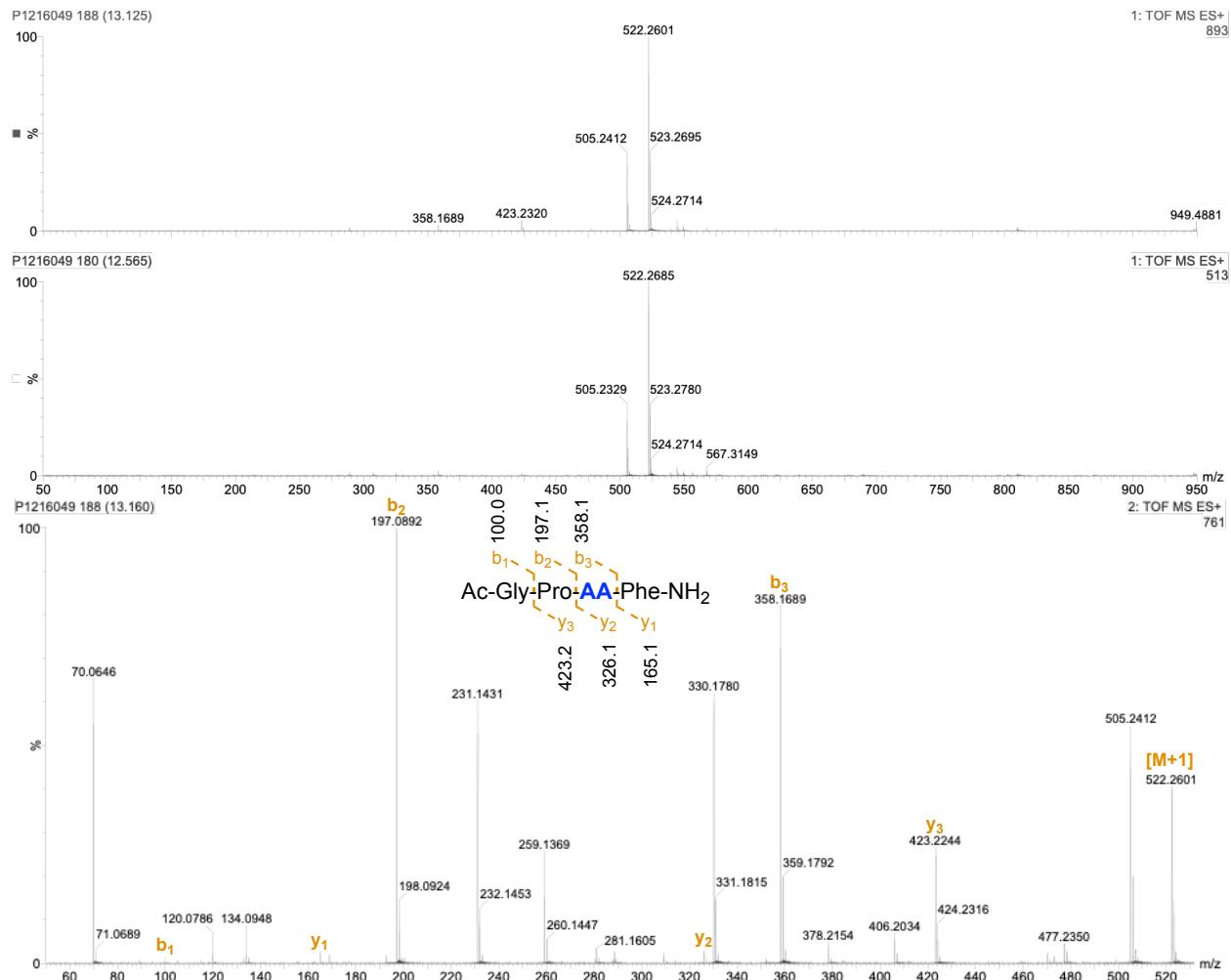


1E': MW = 521.6, Purity = 58.4%, Yield = 5.9% [0.072 mg]

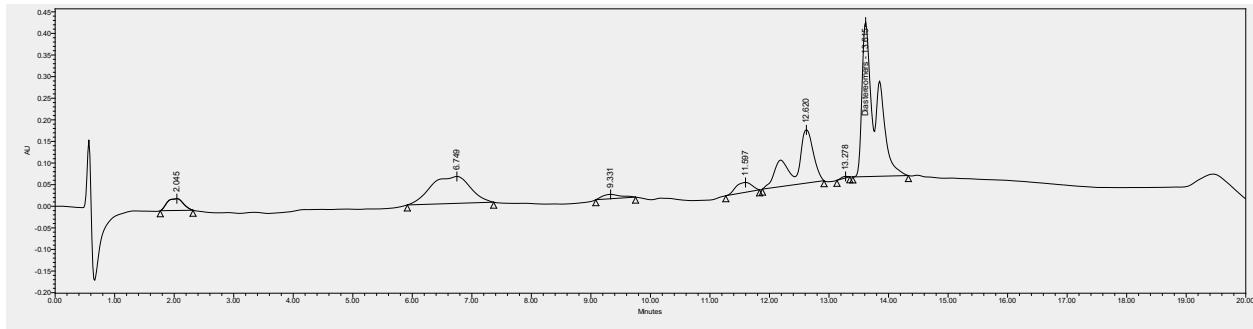
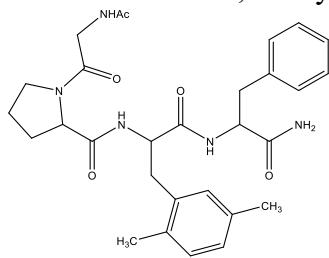


	Name	Retention Time	Area	% Area
1		4.738	138340	2.94
2		5.329	295104	6.28
3		5.855	206634	4.39
4		11.465	390770	8.31
5		12.178	763660	16.24
6		12.875	60693	1.29
7	Diastereomer 1	13.122	878147	18.68
8	Diastereomer 2	13.538	1867833	39.73
9		13.964	46627	0.99
10		14.163	53918	1.15

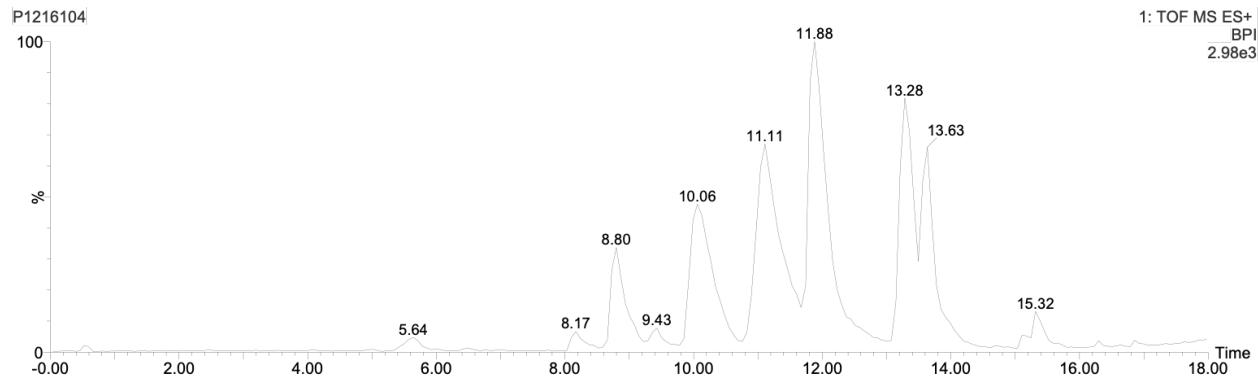


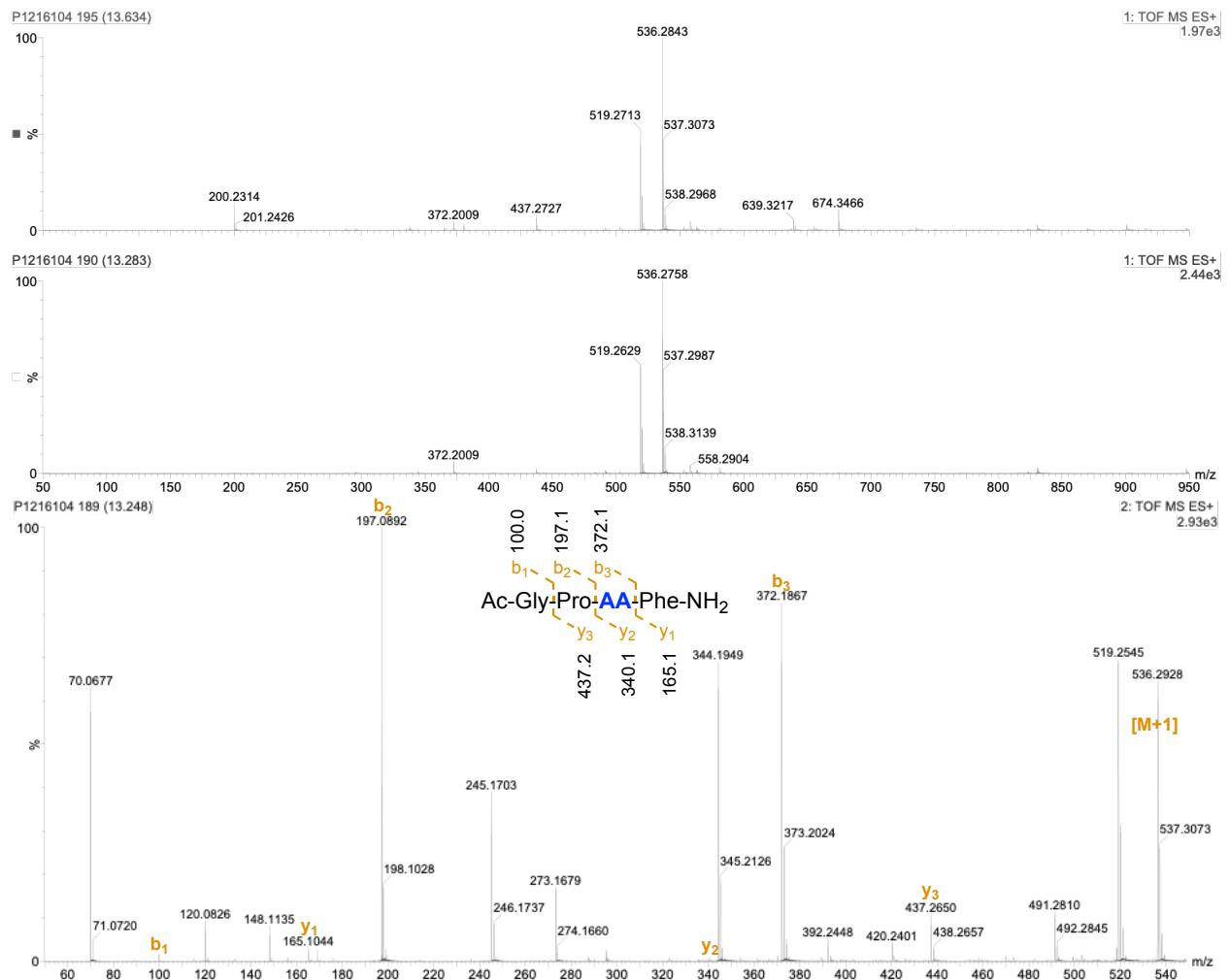


2E': MW = 535.7, Purity = 45.8%, Yield = 15.7% [0.20 mg]

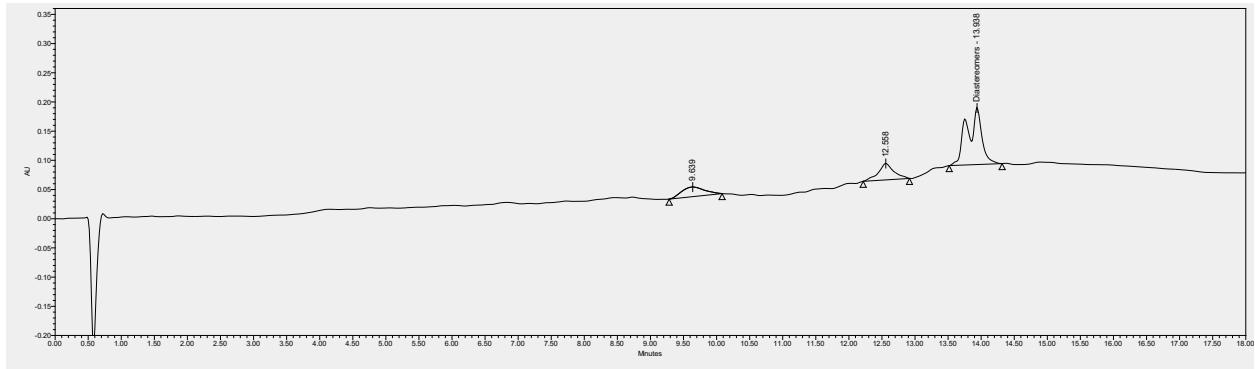
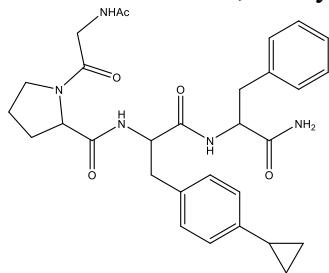


	Name	Retention Time	Area	% Area
1		2.045	515344	4.05
2		6.749	2875647	22.57
3		9.331	219245	1.72
4		11.597	435764	3.42
5		12.620	2824625	22.17
6		13.278	29275	0.23
7	Diastereomers	13.615	5840347	45.84

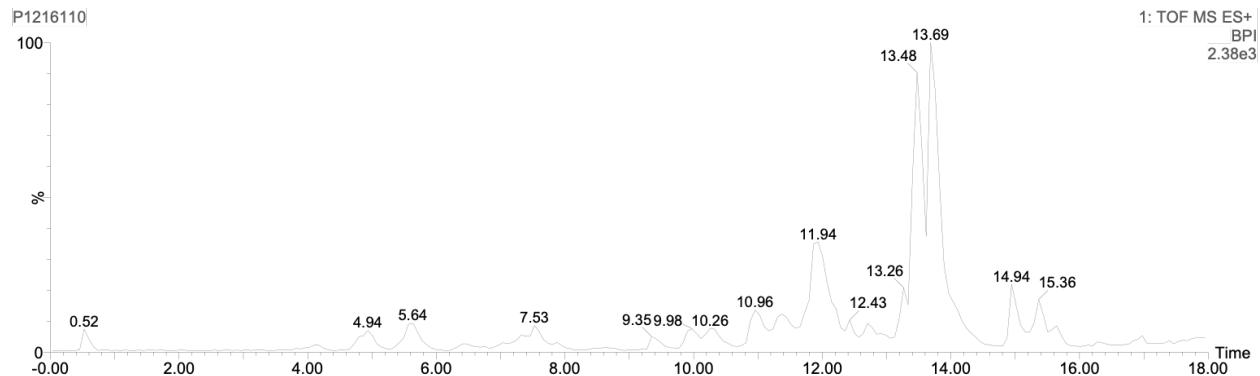


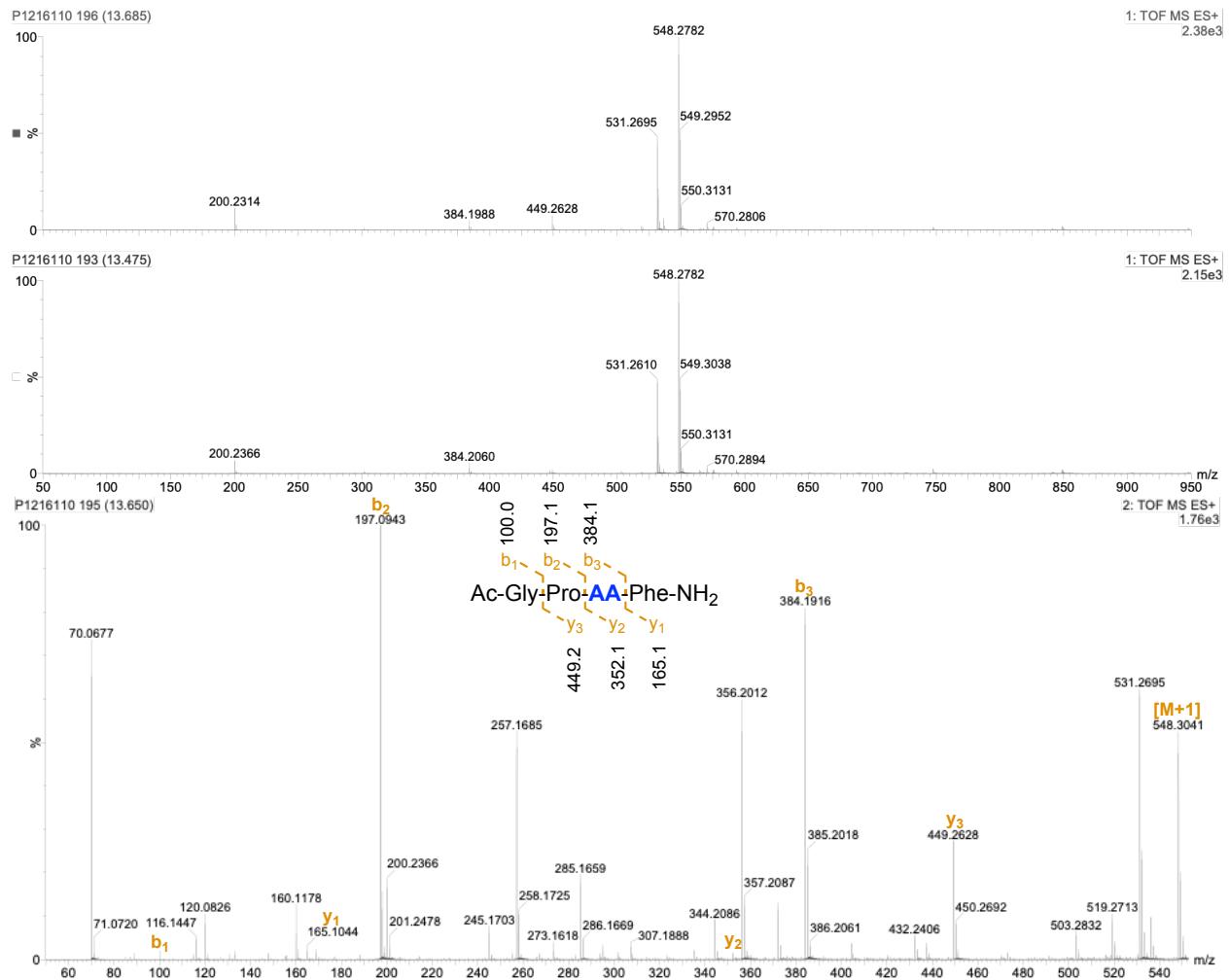


3E': MW = 547.6, Purity = 63.9%, Yield = 3.3% [0.042 mg]

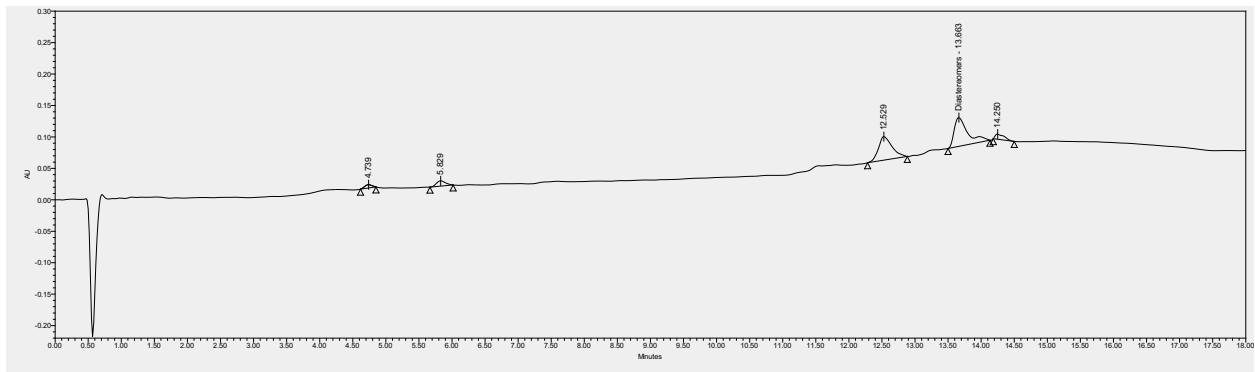
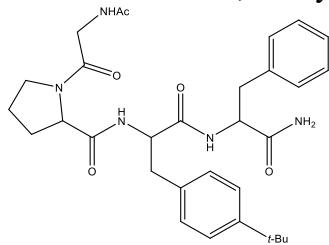


	Name	Retention Time	Area	% Area
1		9.639	402424	16.87
2		12.558	459959	19.28
3	Diastereomers	13.938	1523338	63.85

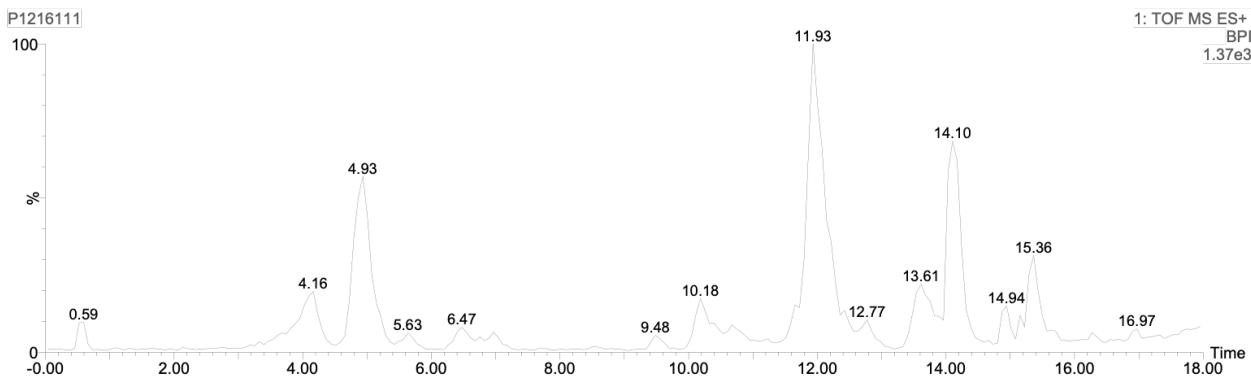


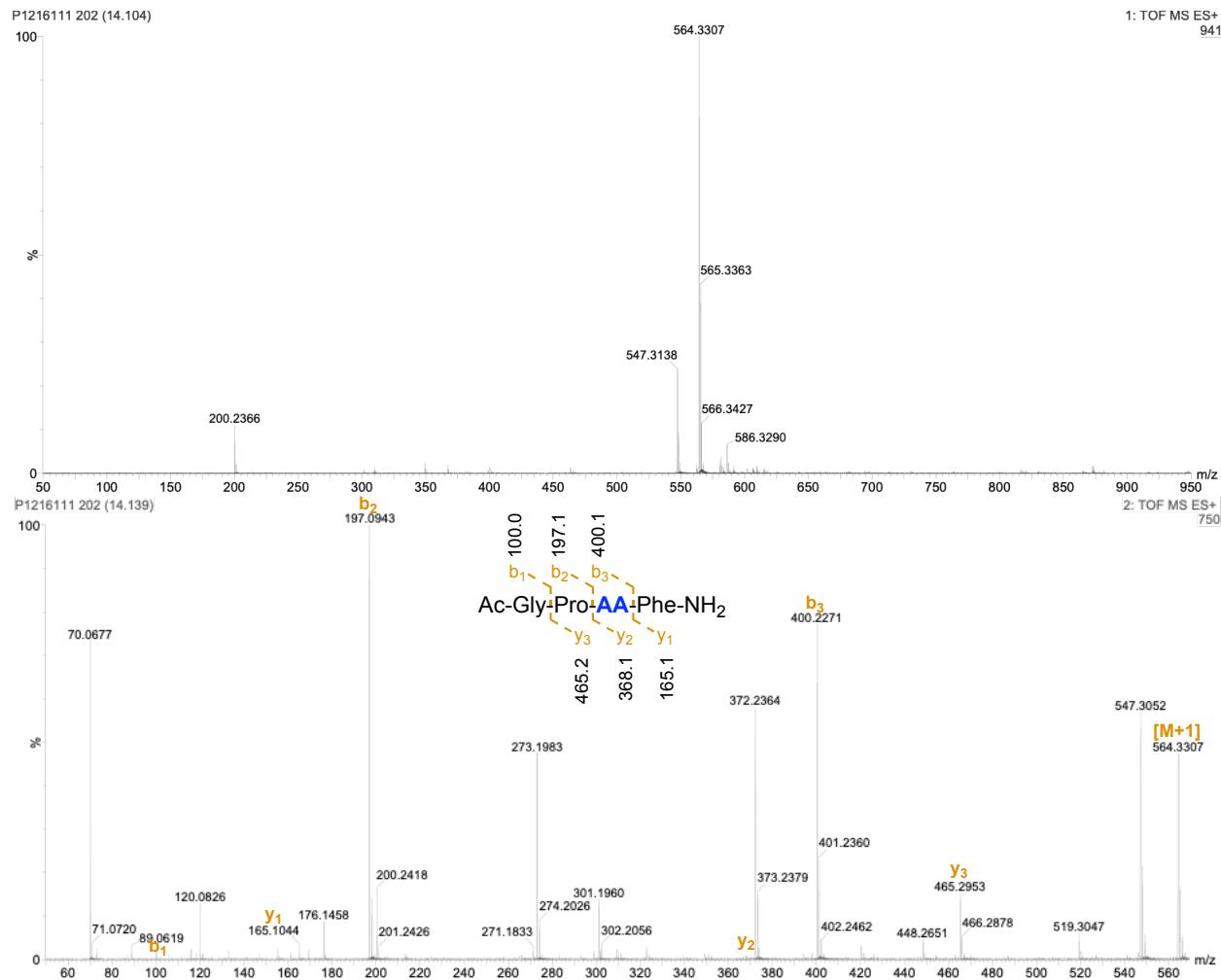


4E': MW = 563.7, Purity = 46.7%, Yield 1.4% [0.018 mg]

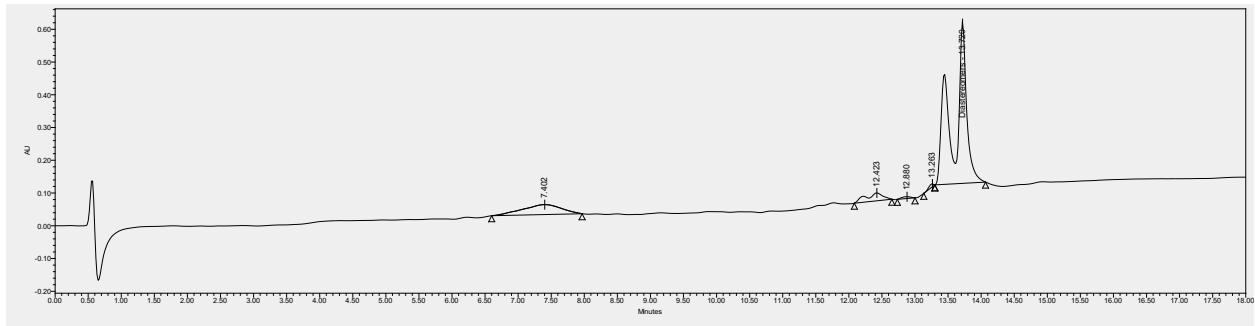
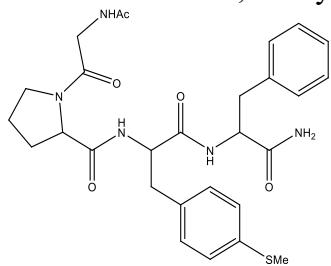


	Name	Retention Time	Area	% Area
1		4.739	40503	2.94
2		5.829	80196	5.82
3		12.529	534366	38.80
4	Diastereomers	13.663	642632	46.66
5		14.250	79493	5.77

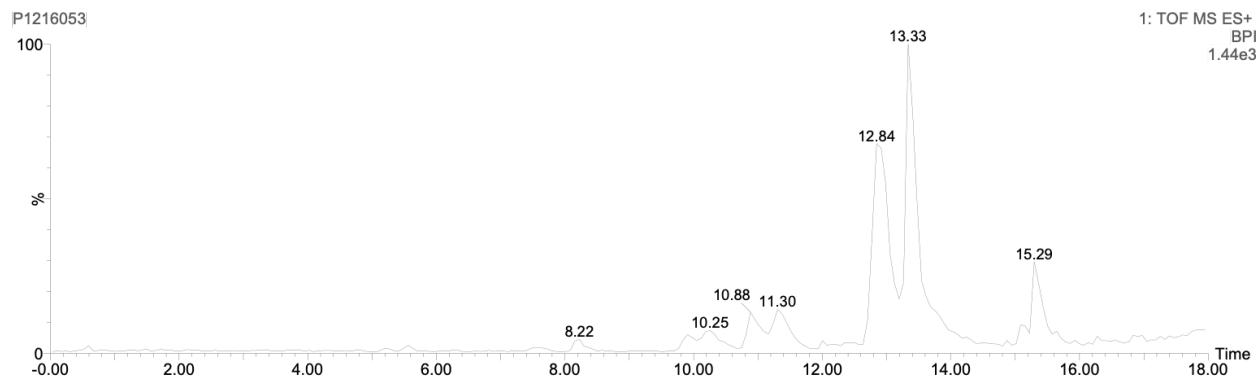


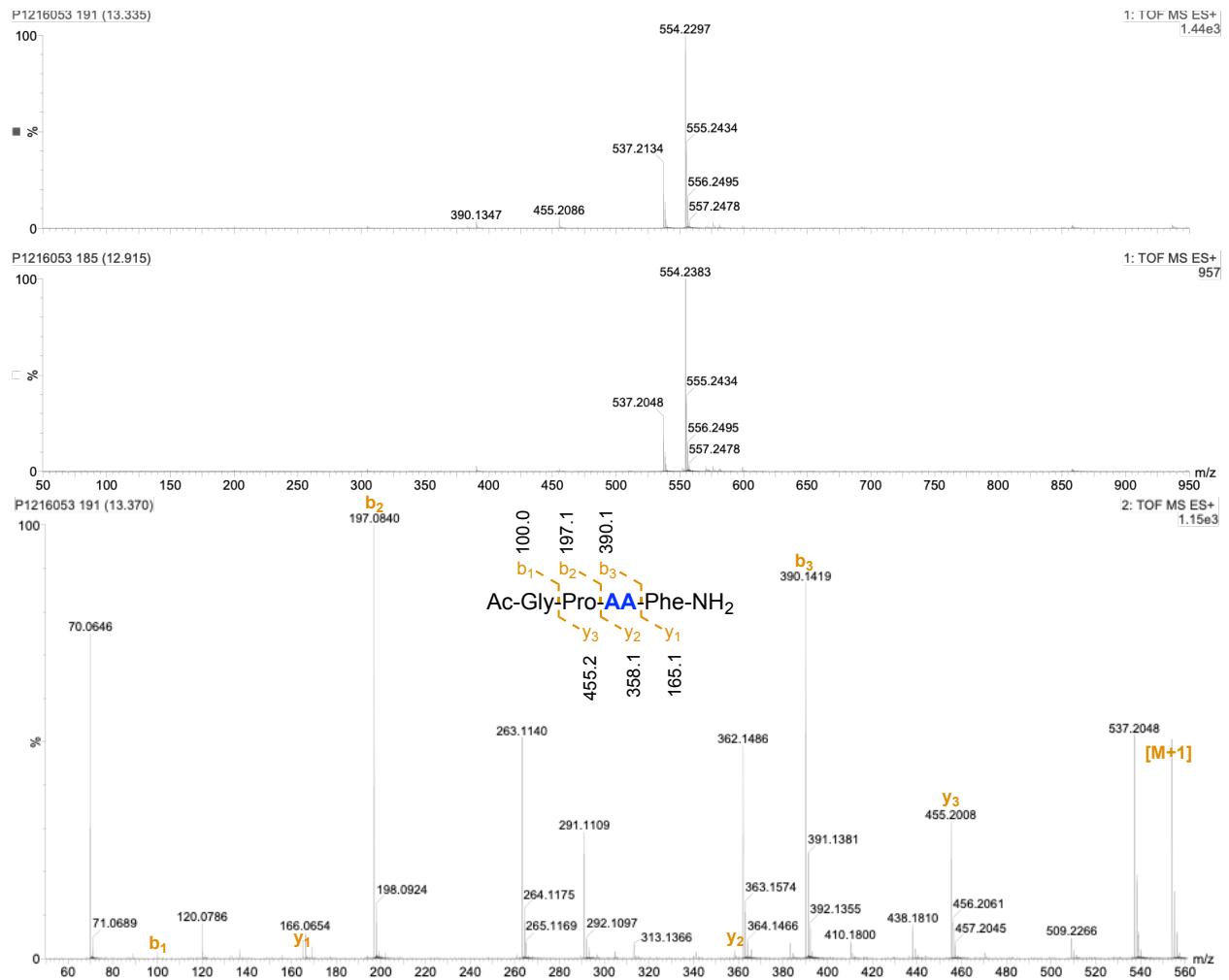


5E': MW = 553.7, Purity = 78.7%, Yield = 17.7% [0.23 mg]

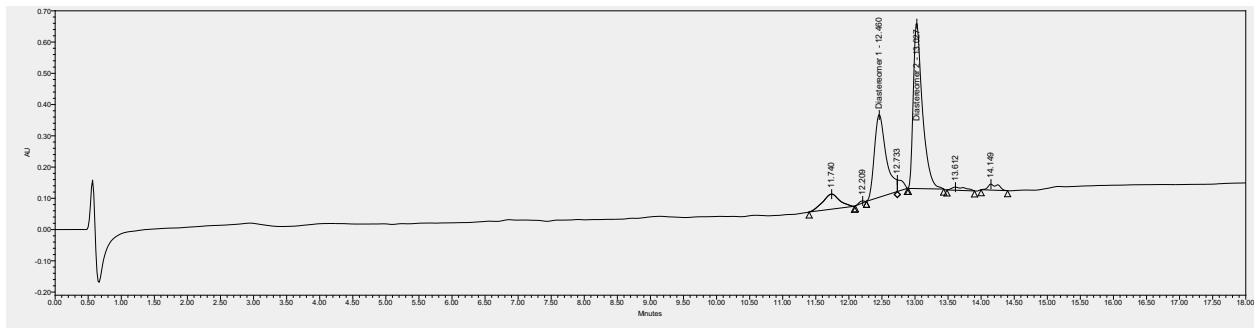
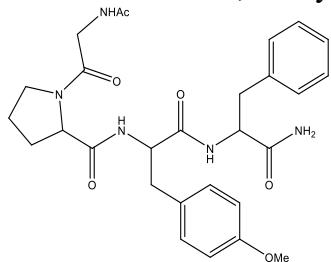


	Name	Retention Time	Area	% Area
1		7.402	1243595	14.86
2		12.423	436623	5.22
3		12.880	55570	0.66
4		13.263	50532	0.60
5	Diastereomers	13.720	6581170	78.65

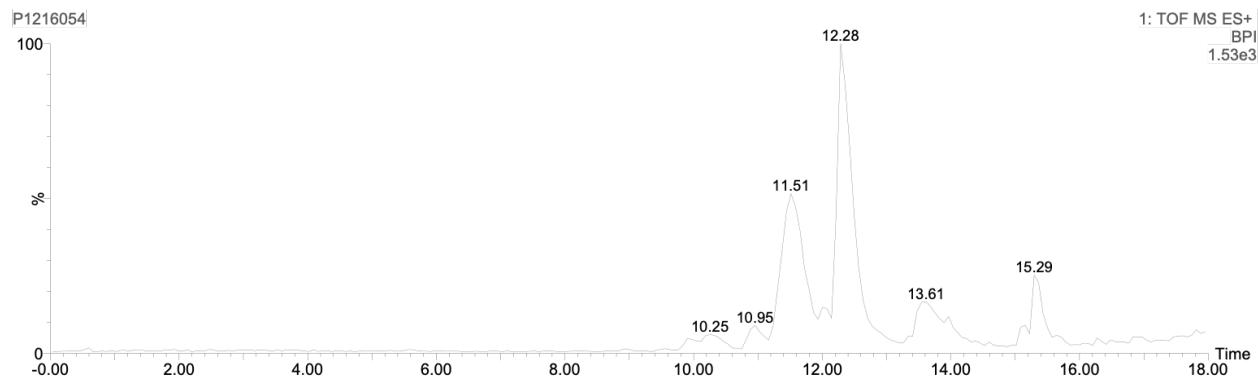


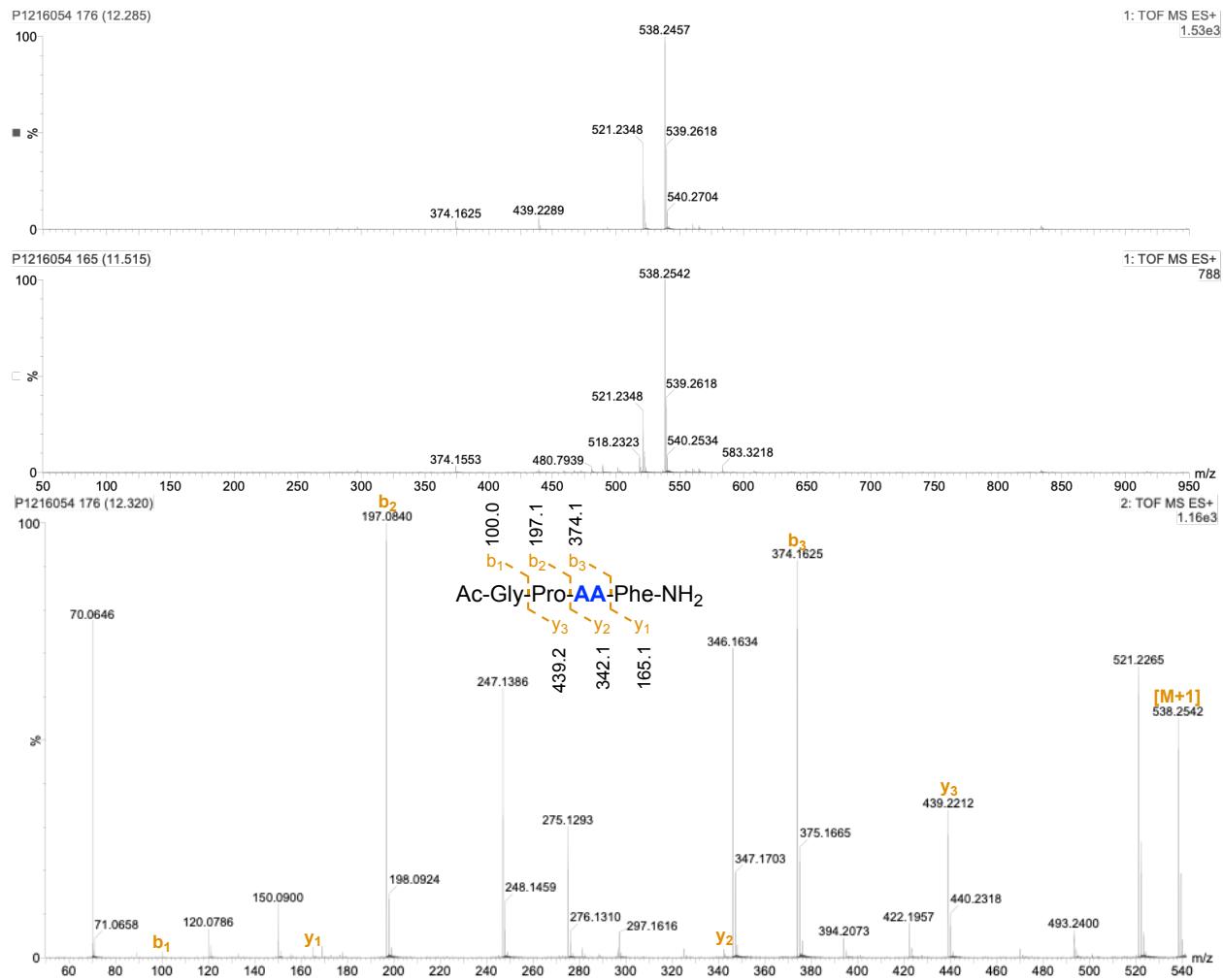


6E': MW = 537.6, Purity = 84.9%, Yield = 22.4% [0.28 mg]

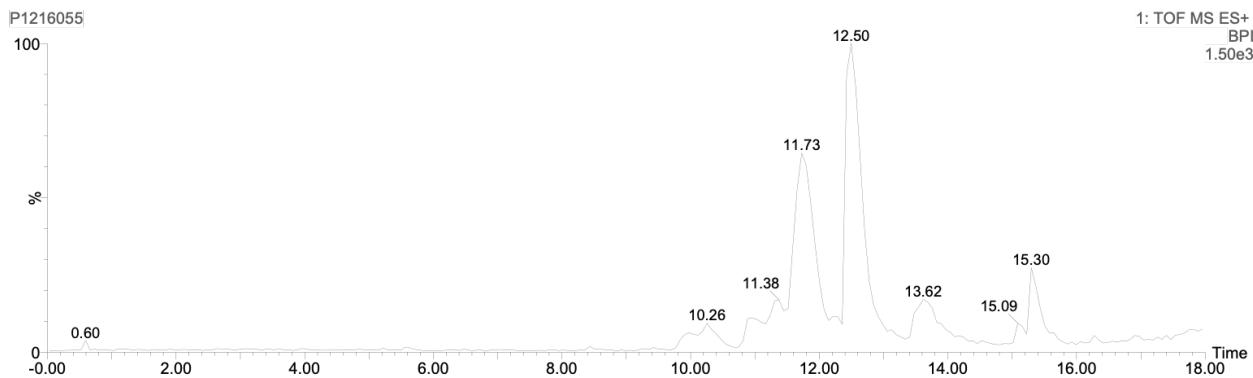
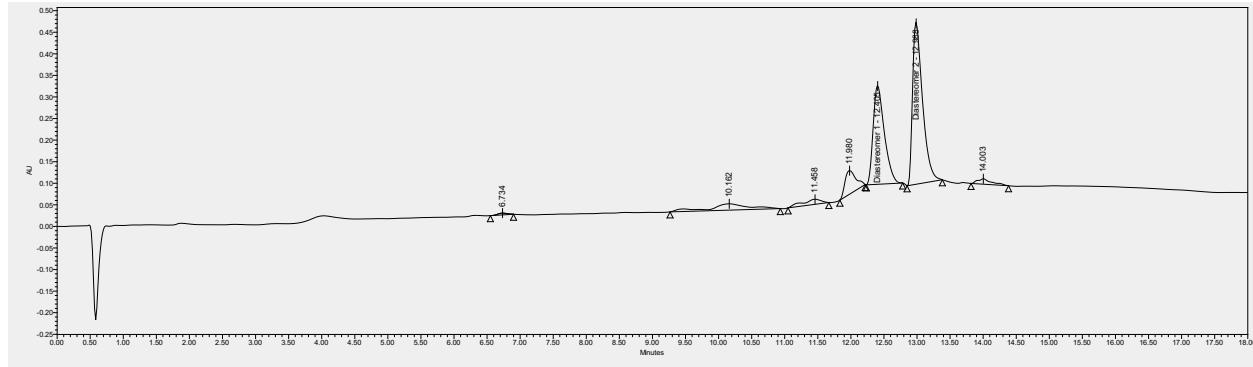
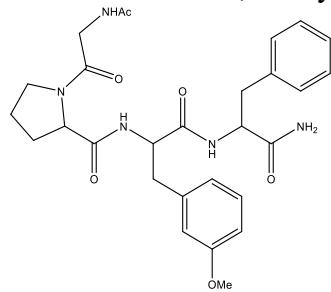


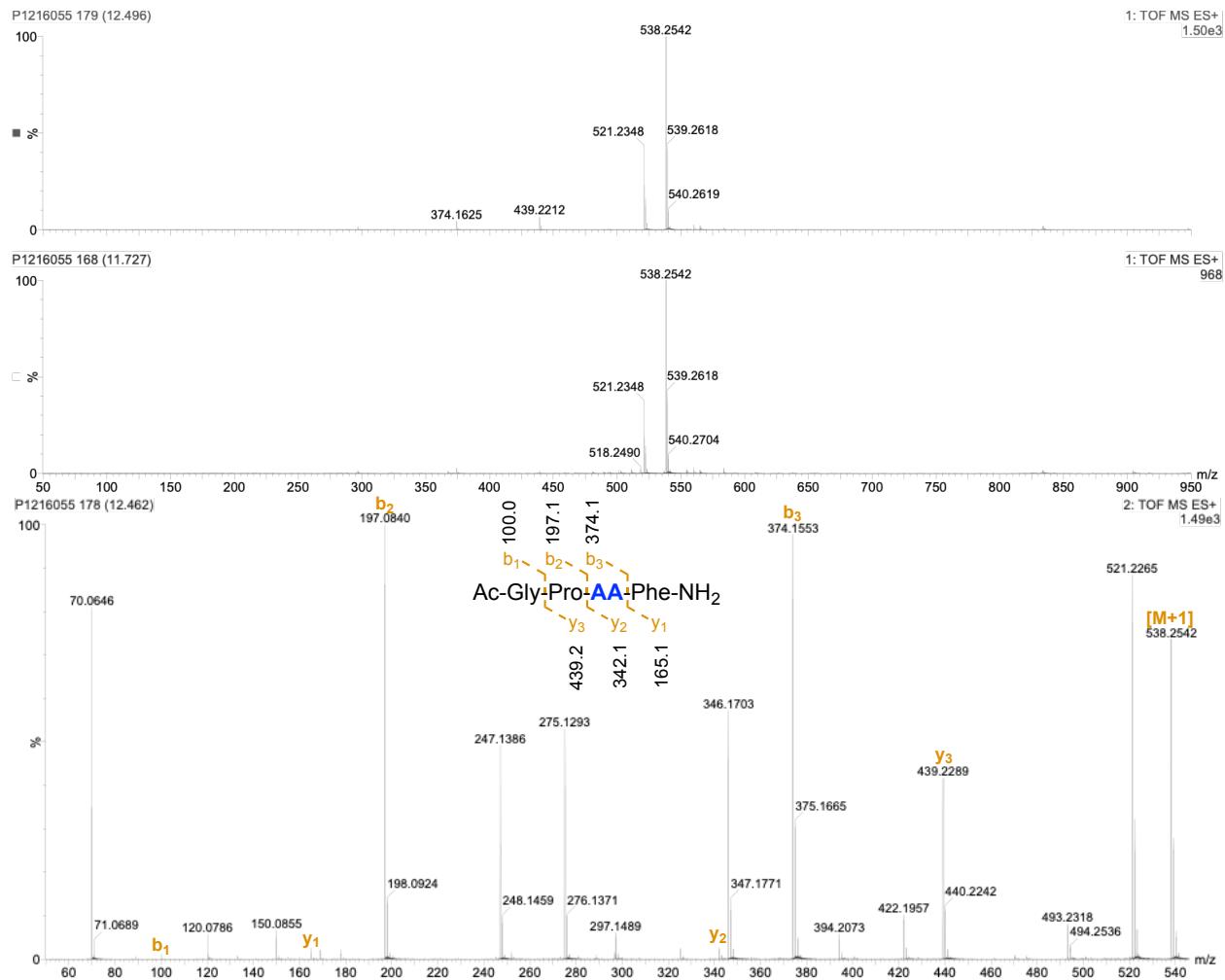
	Name	Retention Time	Area	% Area
1		11.740	862232	8.79
2		12.209	39611	0.40
3	Diastereomer 1	12.460	3251919	33.14
4		12.733	214576	2.19
5	Diastereomer 2	13.027	5079842	51.76
6		13.612	153208	1.56
7		14.149	212322	2.16



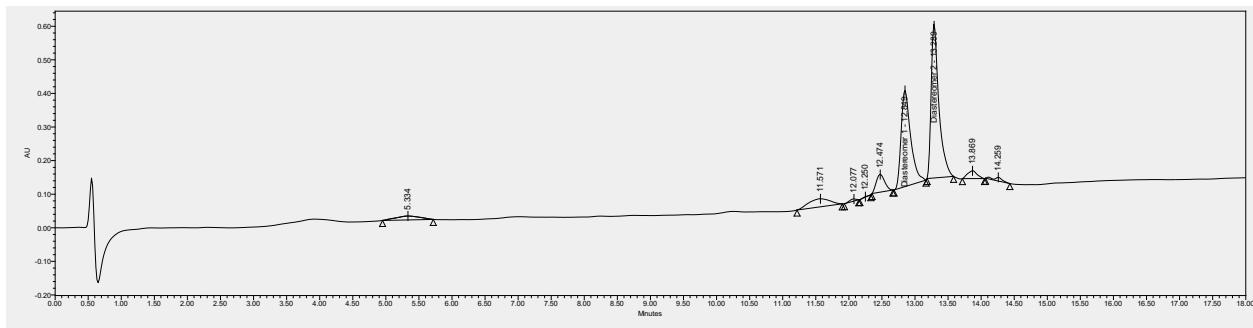
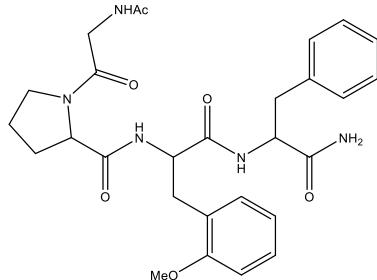


7E': MW = 537.6, Purity = 79.6%, Yield = 14.2% [0.18 mg]

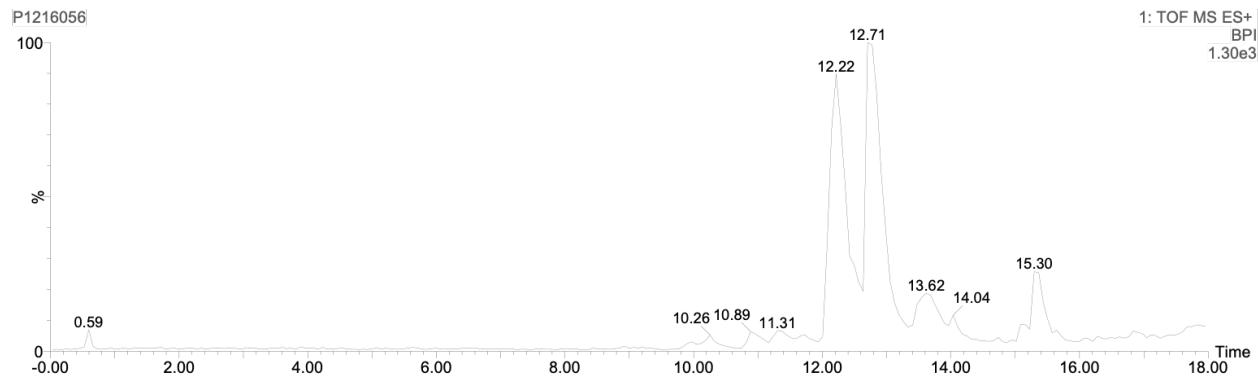


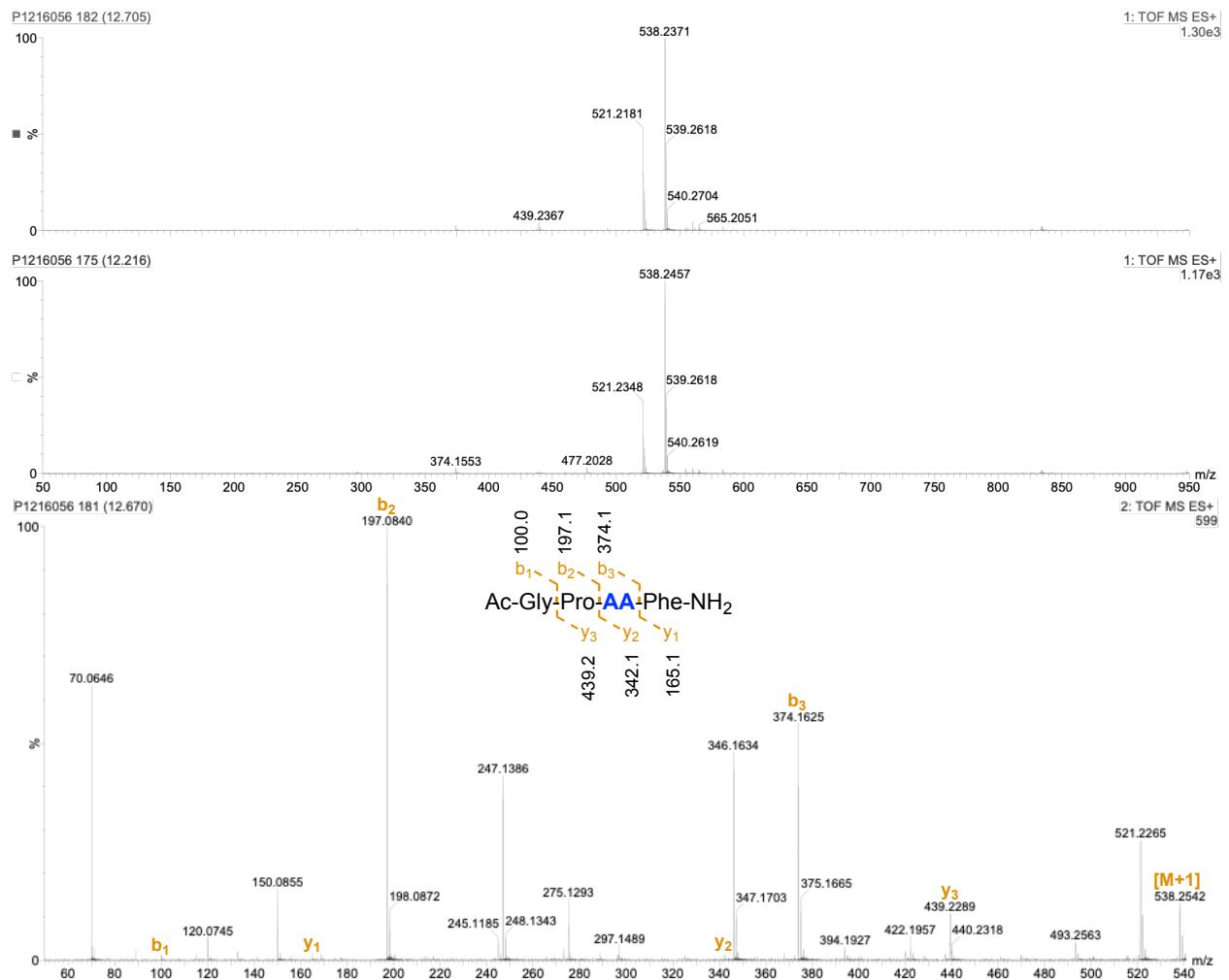


8E': MW = 537.6, Purity = 79.5%, Yield = 17.7% [0.22 mg]

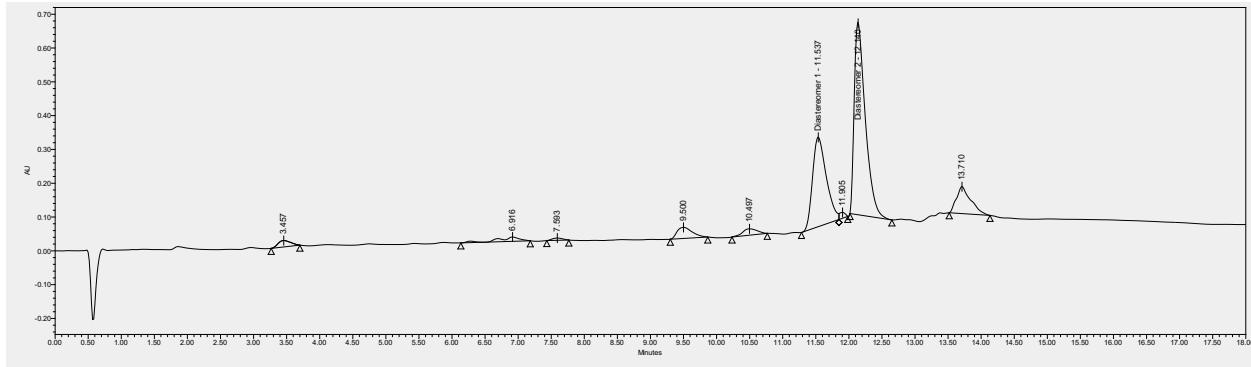
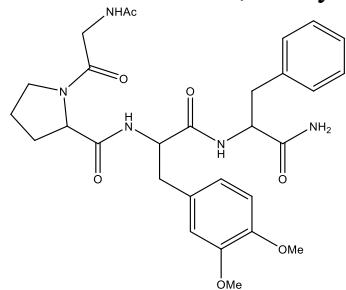


	Name	Retention Time	Area	% Area
1		5.334	297174	3.59
2		11.571	545449	6.59
3		12.077	46009	0.56
4		12.250	6909	0.08
5		12.474	468999	5.66
6	Diastereomer 1	12.849	2873822	34.70
7	Diastereomer 2	13.289	3710645	44.80
8		13.869	231876	2.80
9		14.259	102051	1.23

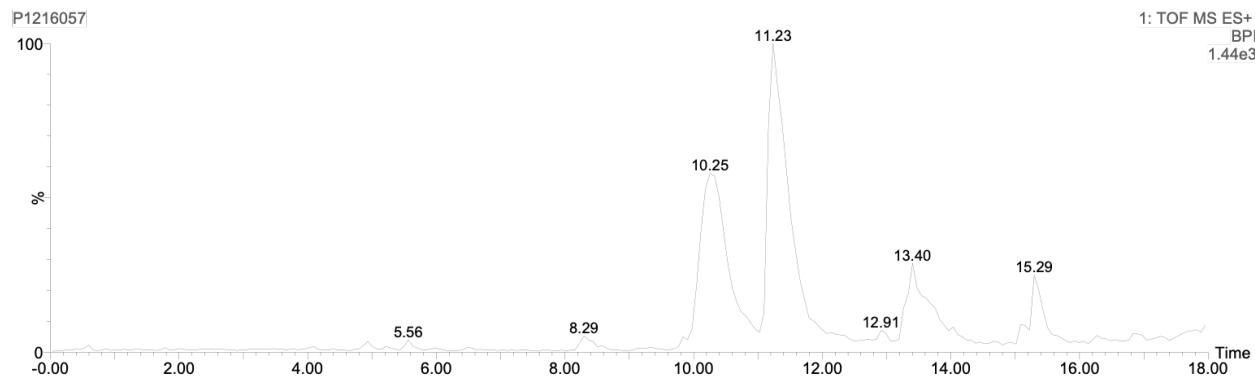


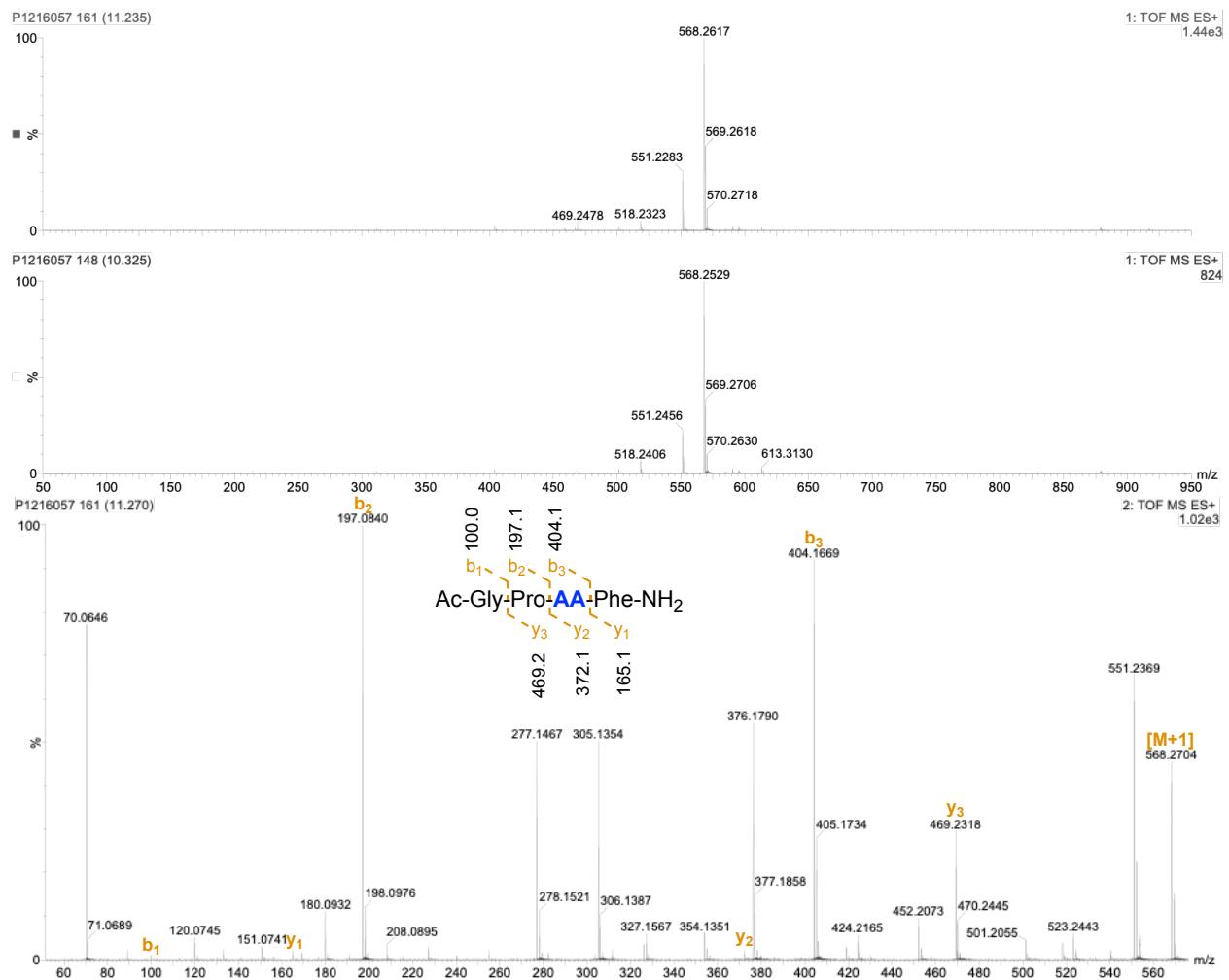


9E': MW = 567.6, Purity = 79.2%, Yield = 22.4% [0.30 mg]

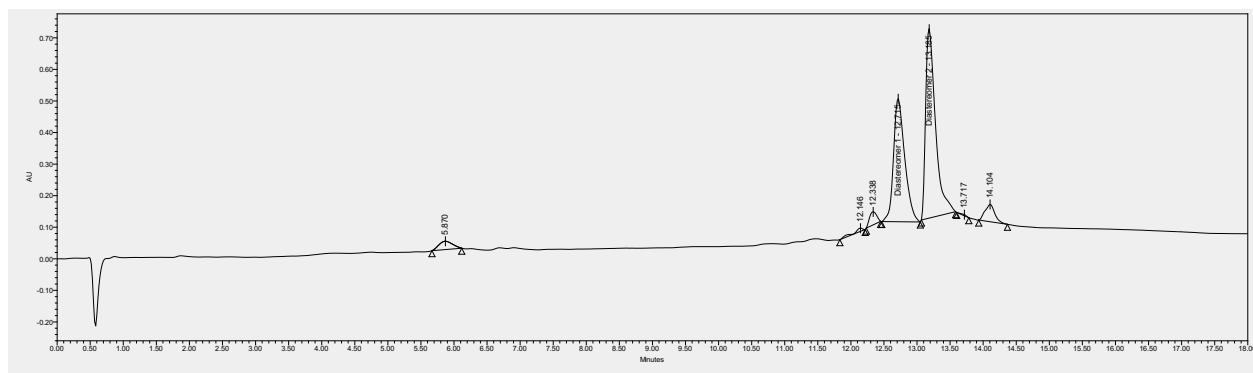
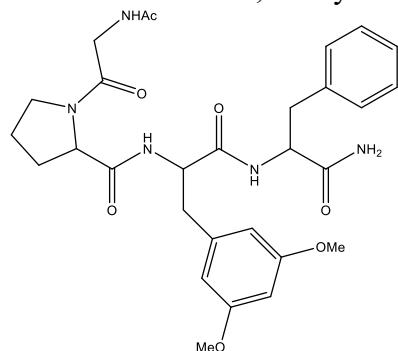


	Name	Retention Time	Area	% Area
1		3.457	252982	1.92
2		6.916	286735	2.18
3		7.593	74817	0.57
4		9.500	525492	4.00
5		10.497	289317	2.20
6	Diastereomer 1	11.537	3814306	29.01
7		11.905	104615	0.80
8	Diastereomer 2	12.140	6598771	50.19
9		13.710	1199691	9.13

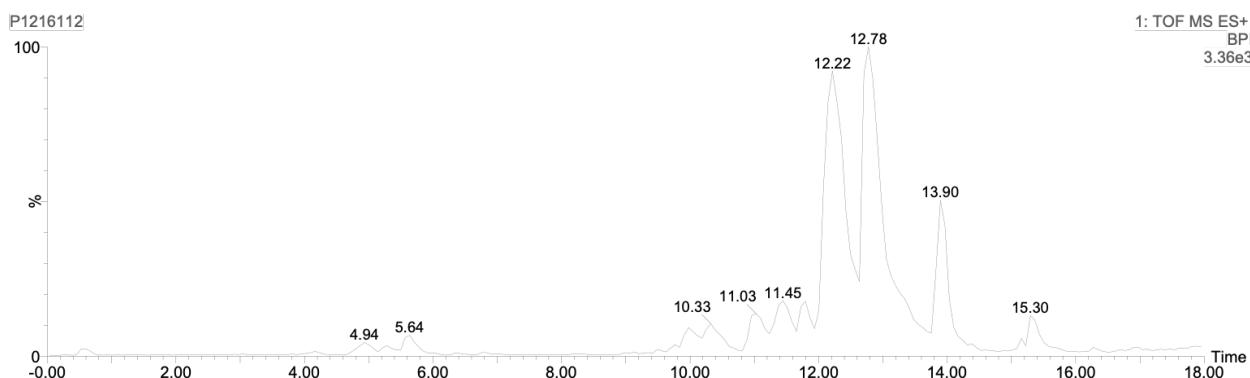


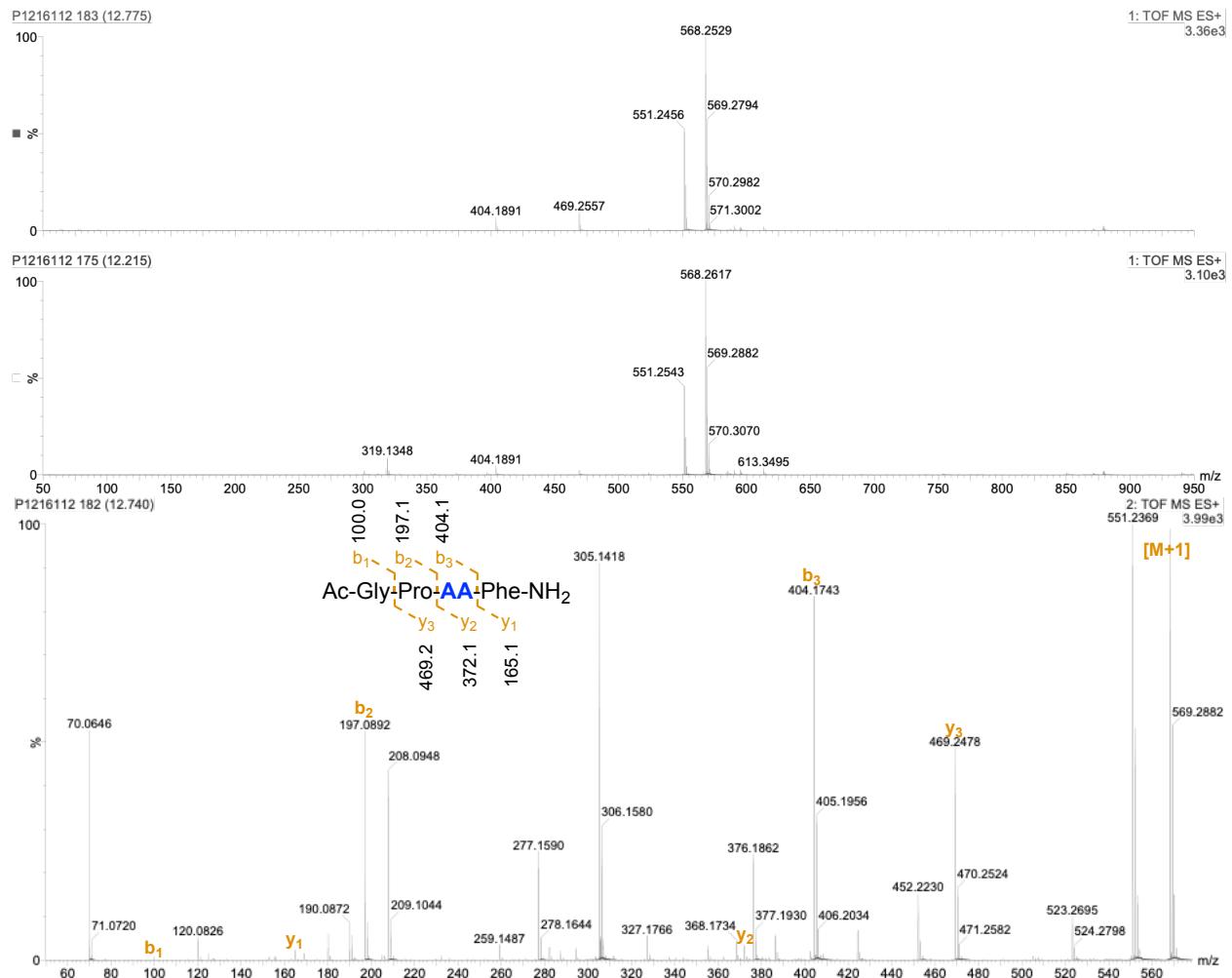


10E': MW = 567.6, Purity = 88.7%, Yield = 23.3% [0.31 mg]

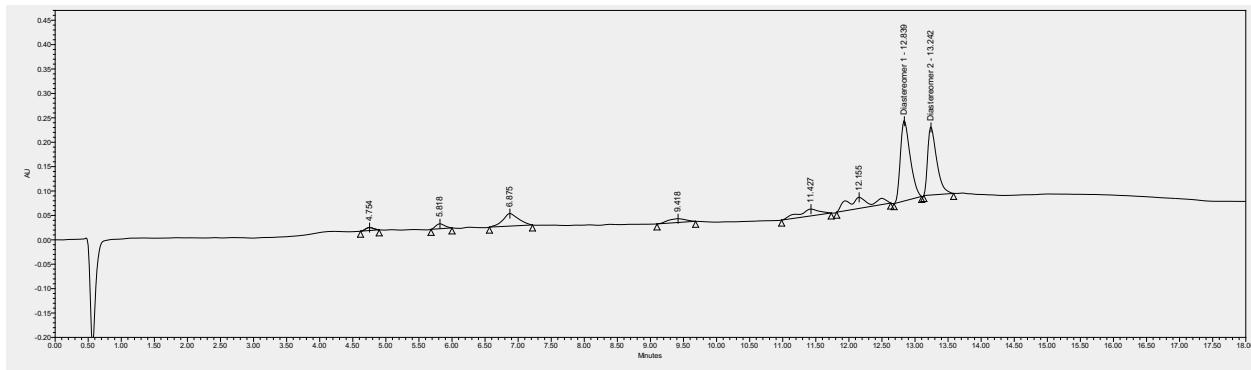
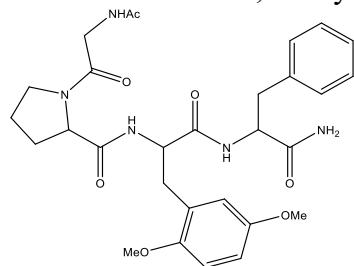


	Name	Retention Time	Area	% Area
1		5.870	380960	3.12
2		12.146	102226	0.84
3		12.338	313936	2.57
4	Diastereomer 1	12.715	4598051	37.71
5	Diastereomer 2	13.185	6217556	50.99
6		13.717	20466	0.17
7		14.104	560347	4.60

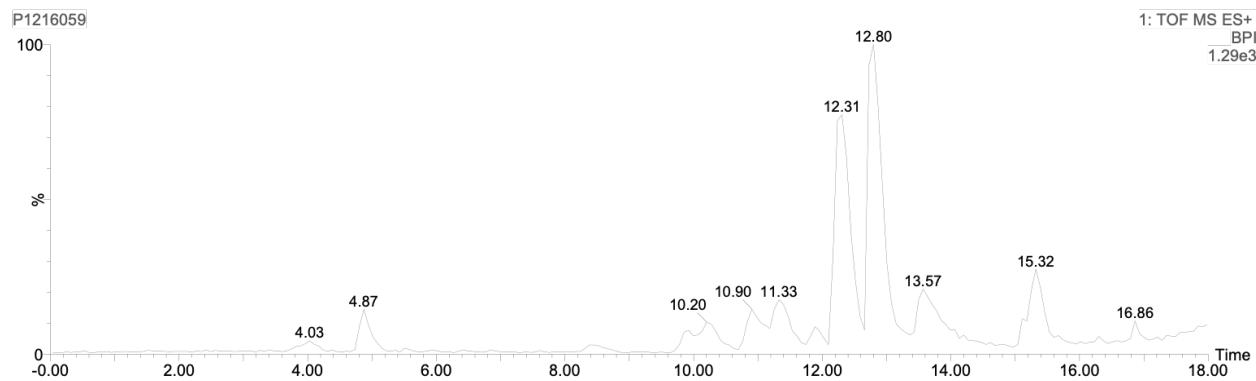


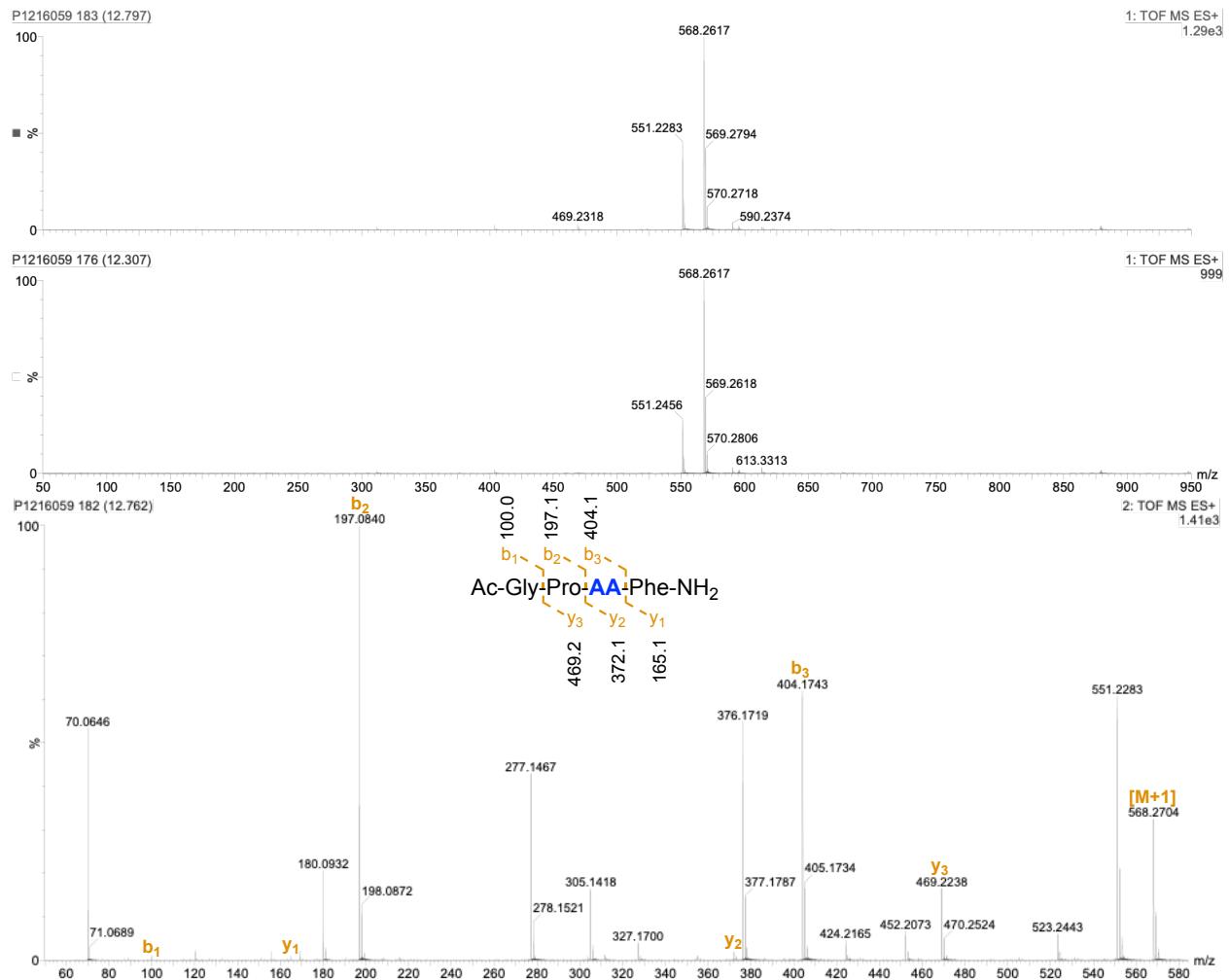


11E': MW = 567.6, Purity = 65.2%, Yield = 6.5% [0.086 mg]

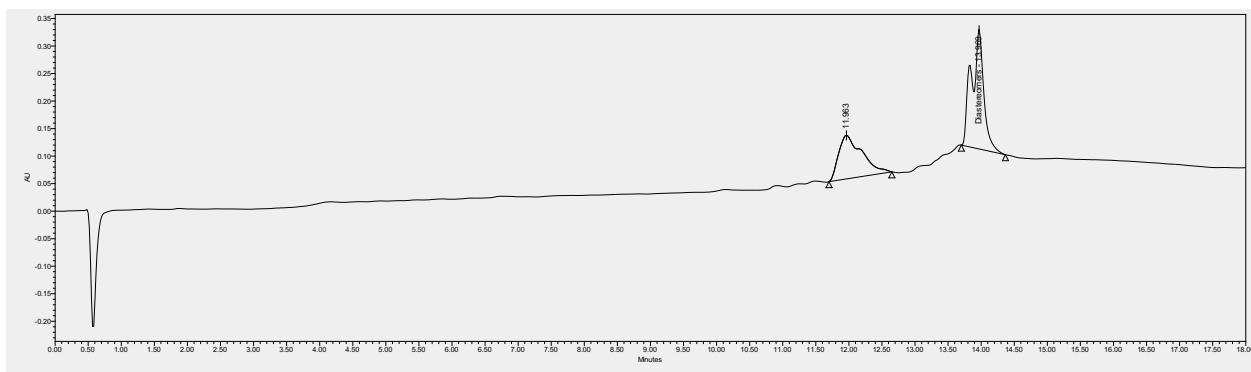
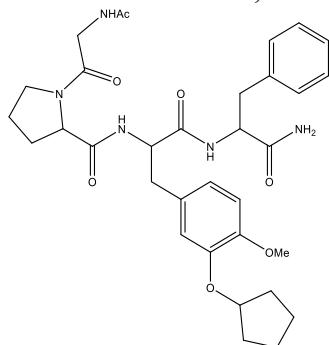


	Name	Retention Time	Area	% Area
1		4.754	51671	1.12
2		5.818	96192	2.08
3		6.875	425233	9.21
4		9.418	146780	3.18
5		11.427	310613	6.73
6		12.155	576944	12.50
7	Diastereomer 1	12.839	1690409	36.61
8	Diastereomer 2	13.242	1318978	28.57

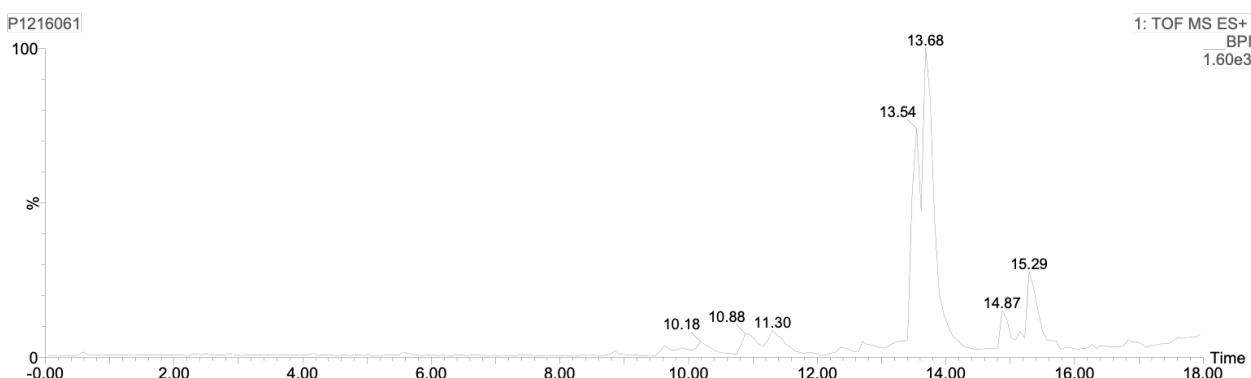


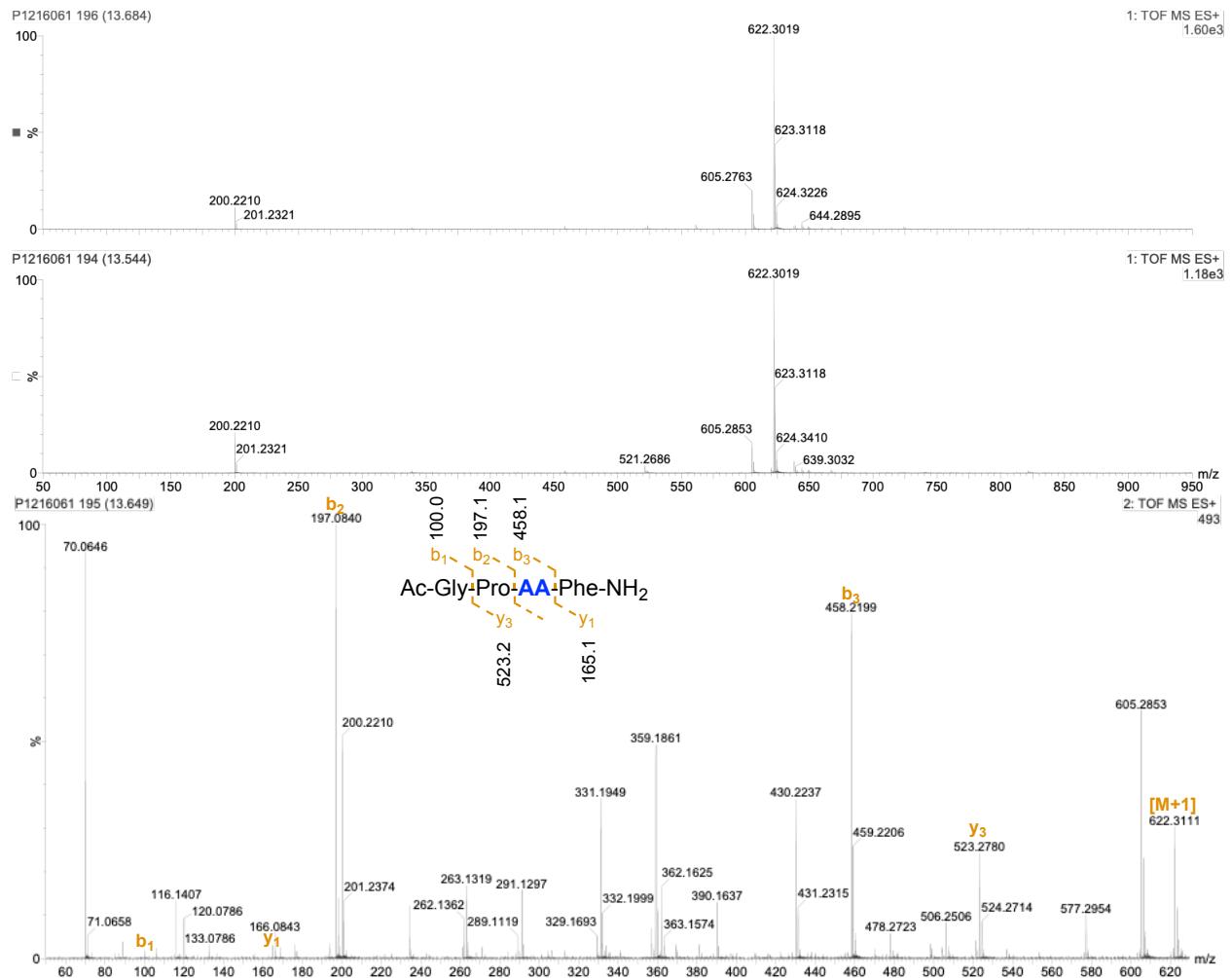


12E': MW = 621.7, Purity = 59.1%, Yield = 6.0% [0.087 mg]

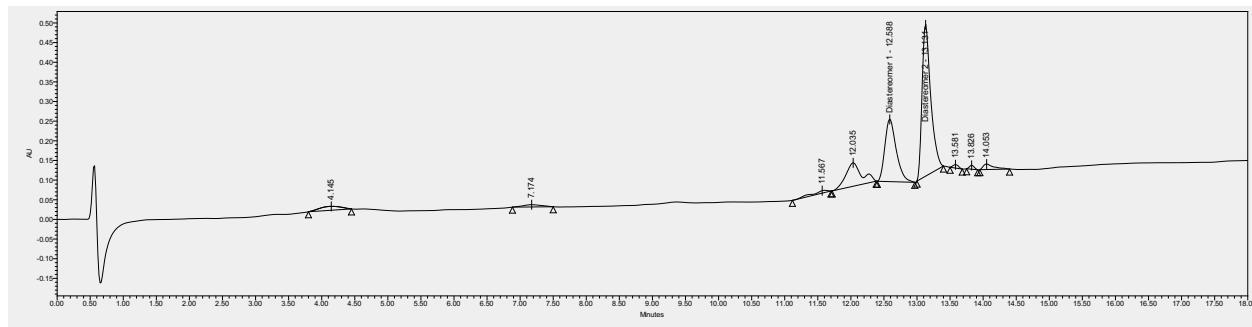
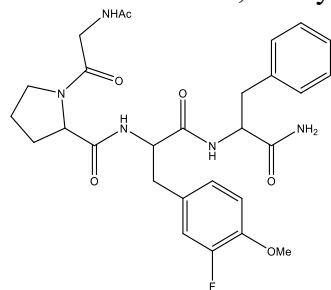


	Name	Retention Time	Area	% Area
1		11.963	1935350	40.90
2	Diastereomers	13.969	2796381	59.10

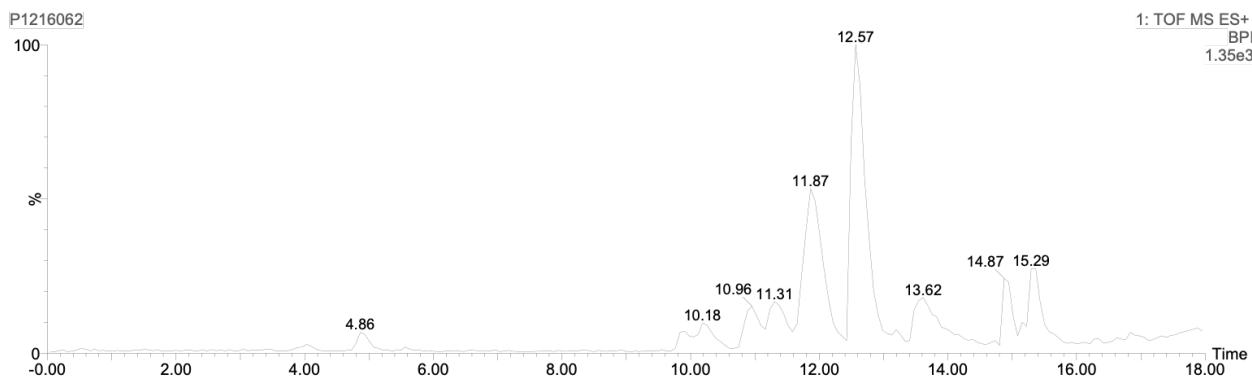


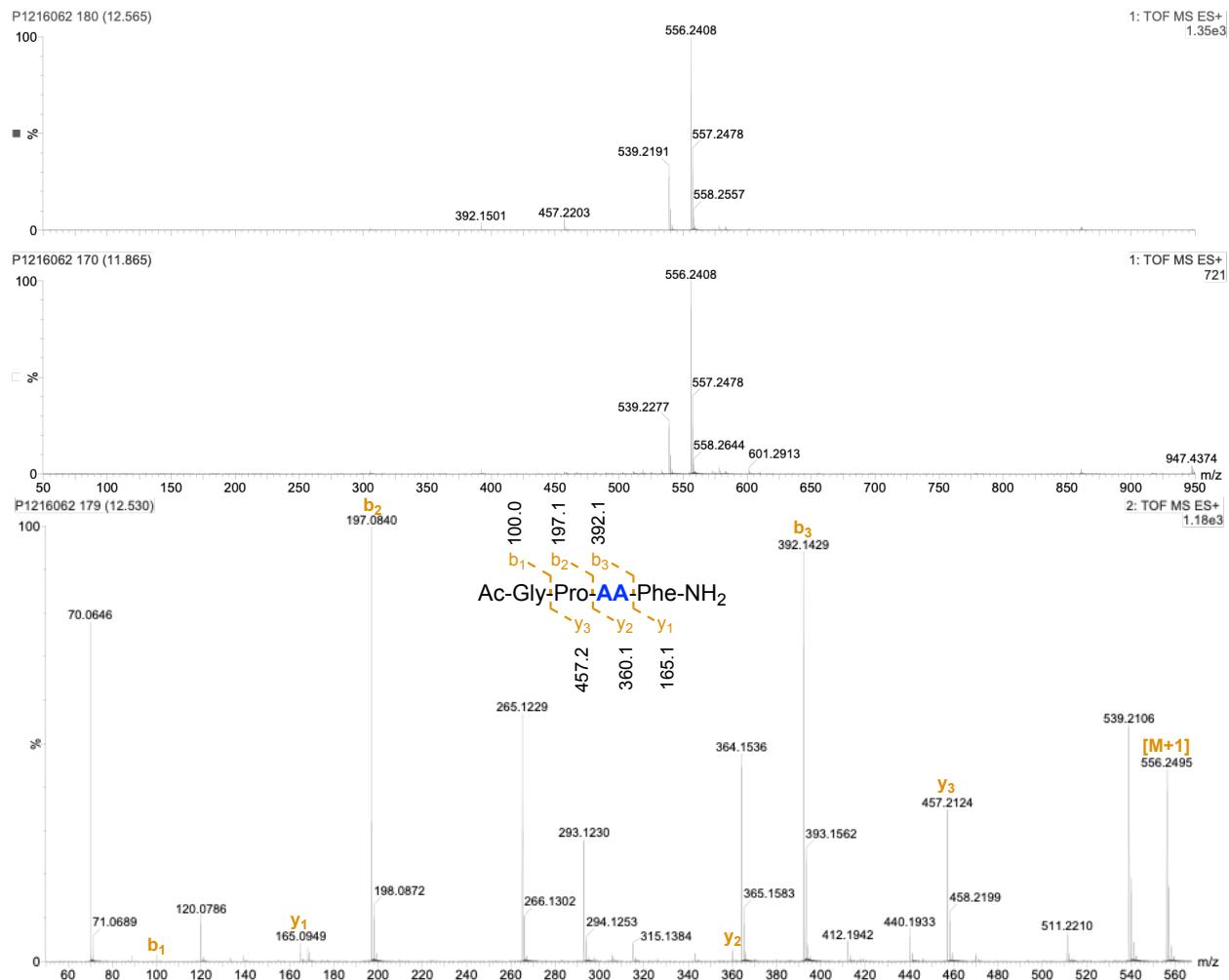


1F': MW = 555.6, Purity = 76.0%, Yield = 14.6% [0.19 mg]

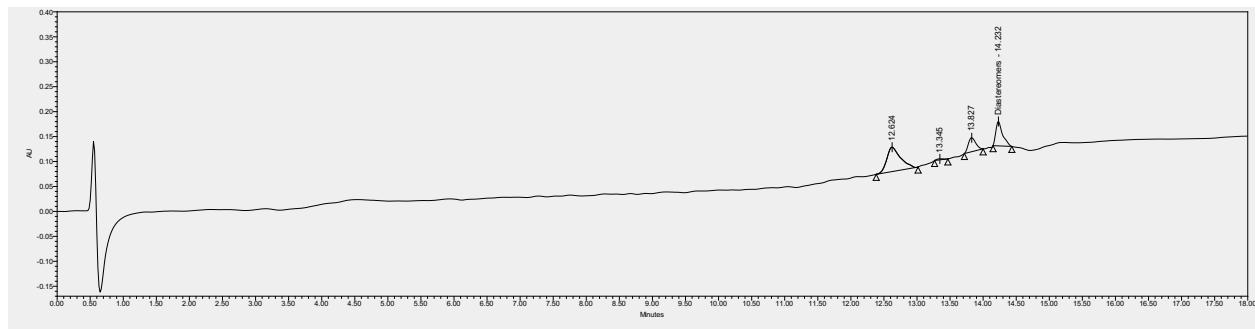
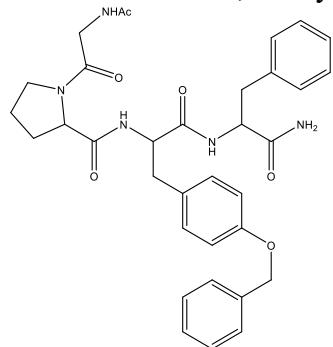


	Name	Retention Time	Area	% Area
1		4.145	230687	3.24
2		7.174	114243	1.60
3		11.567	112673	1.58
4		12.035	987867	13.87
5	Diastereomer 1	12.588	1899373	26.67
6	Diastereomer 2	13.131	3519544	49.42
7		13.581	50403	0.71
8		13.826	49737	0.70
9		14.053	157003	2.20

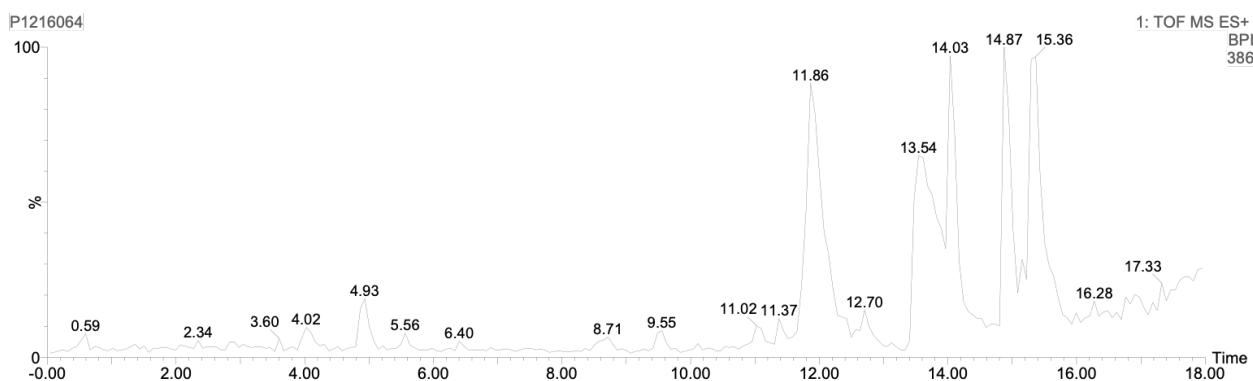


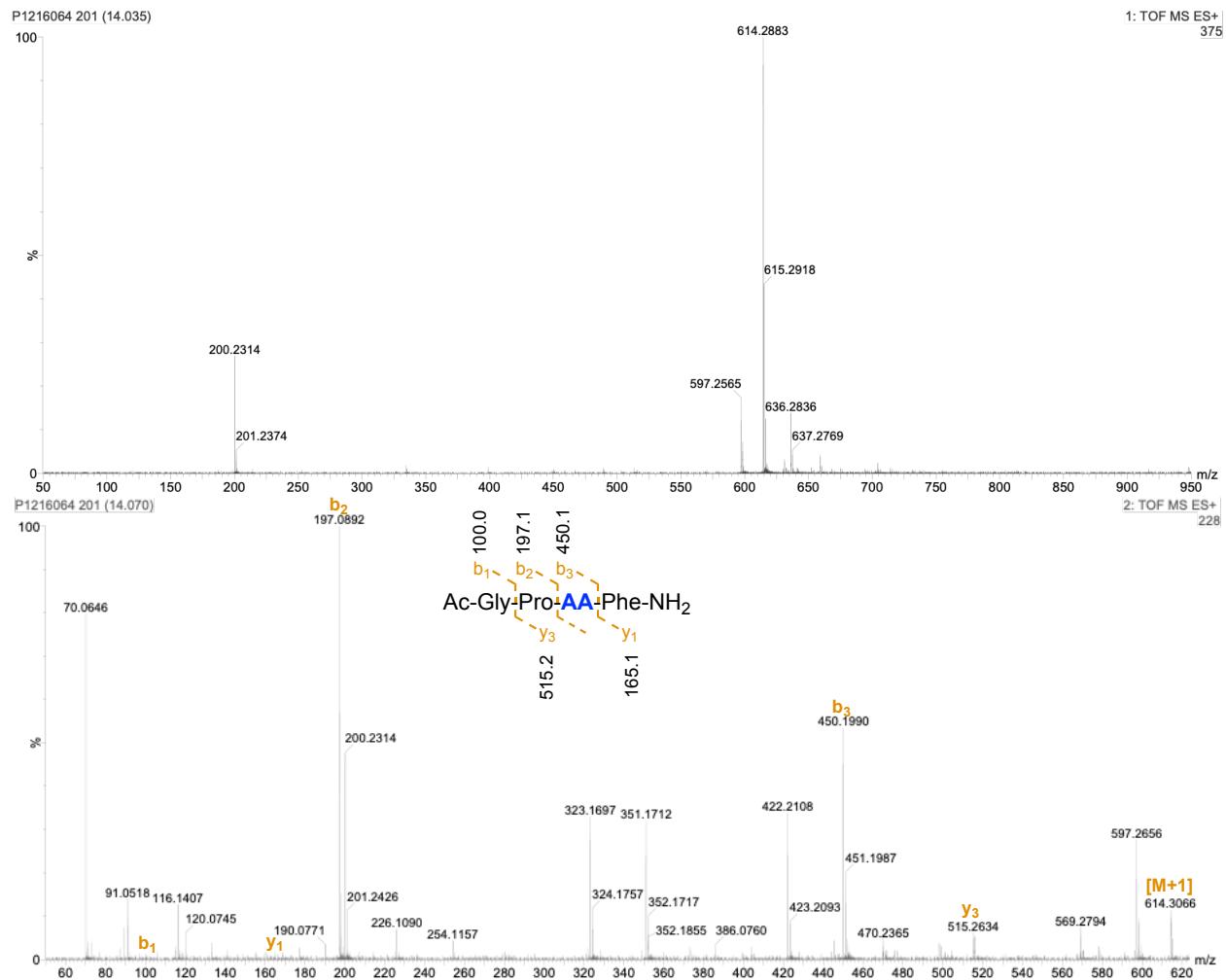


2F': MW = 613.7, Purity = 27.0%, Yield = 0.70% [0.010 mg]

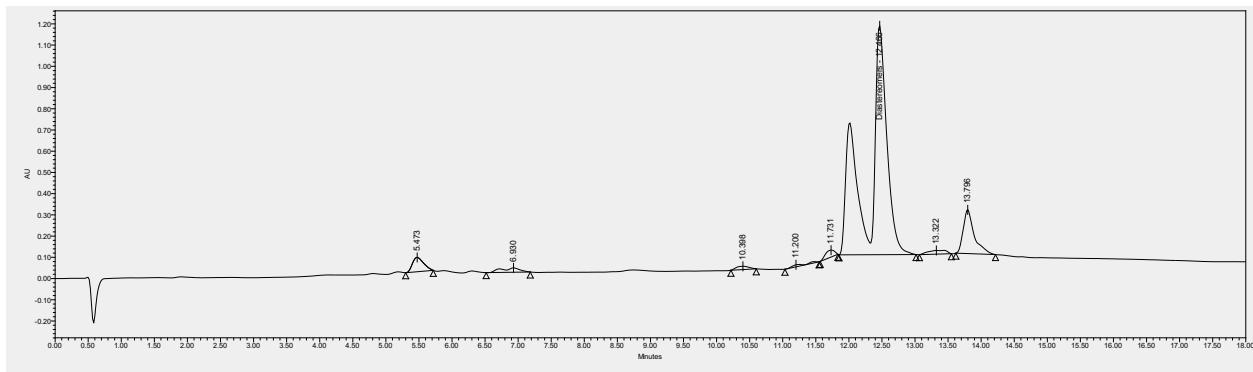
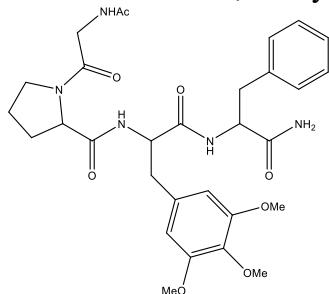


	Name	Retention Time	Area	% Area
1		12.624	701605	54.72
2		13.345	14100	1.10
3		13.827	220174	17.17
4	Diastereomers	14.232	346206	27.00

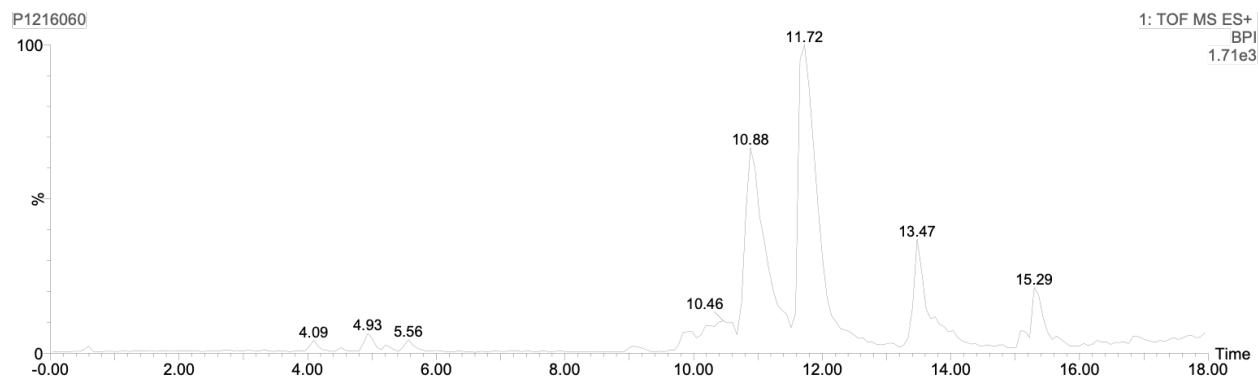


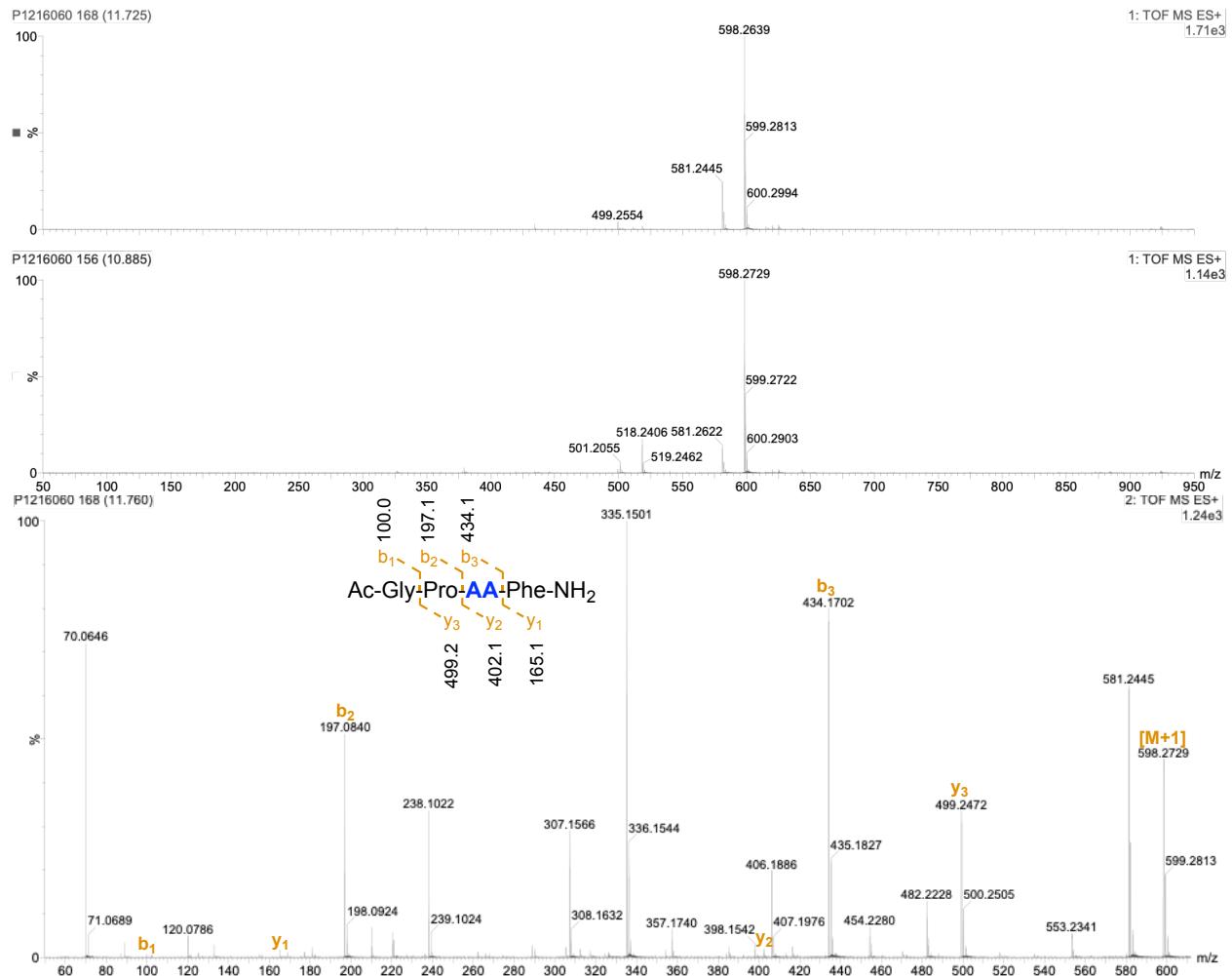


3F': MW = 597.7, Purity = 81.7%, Yield = 44.1% [0.61 mg]

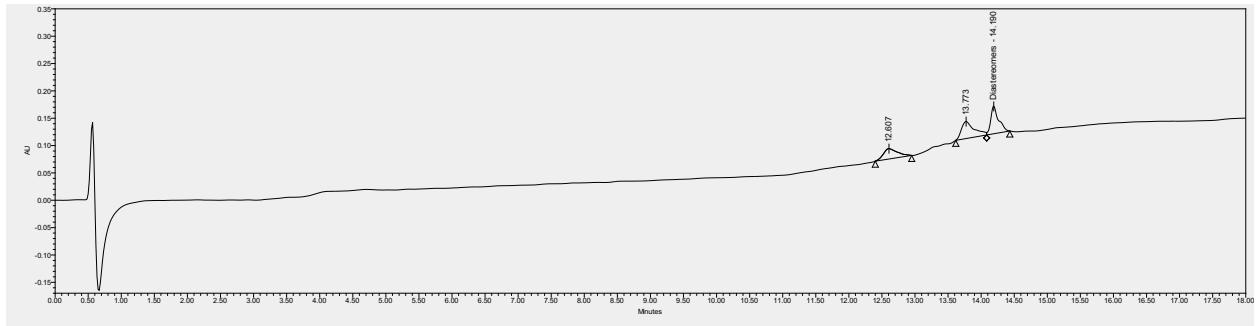
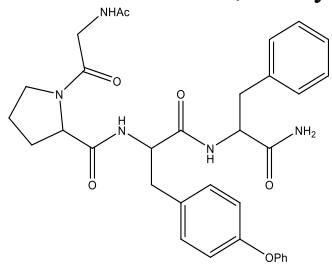


	Name	Retention Time	Area	% Area
1		5.473	796739	3.17
2		6.930	406399	1.62
3		10.398	210541	0.84
4		11.200	137232	0.55
5		11.731	326916	1.30
6	Diastereomers	12.466	20501442	81.69
7		13.322	341290	1.36
8		13.796	2375528	9.47

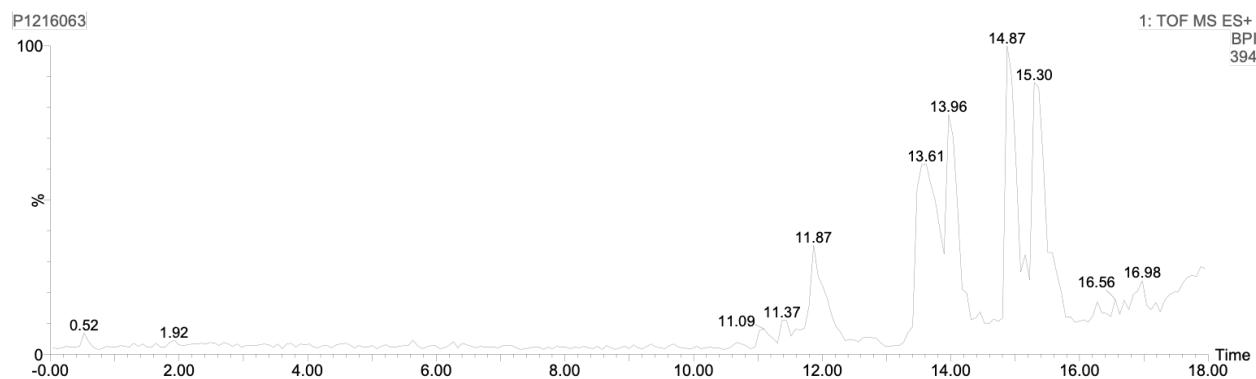


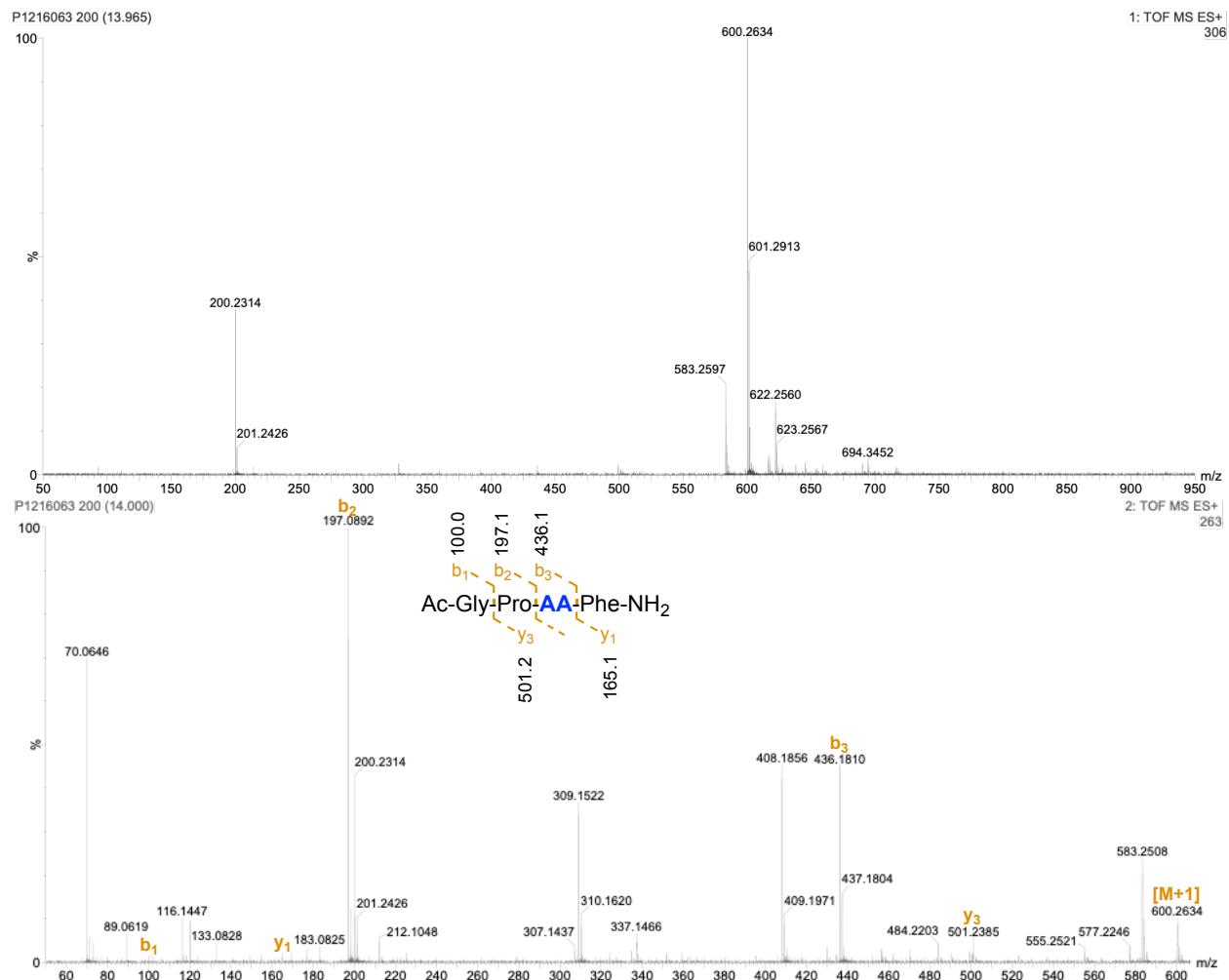


4F': MW = 599.7, Purity = 37.2%, Yield = 0.84% [0.012 mg]

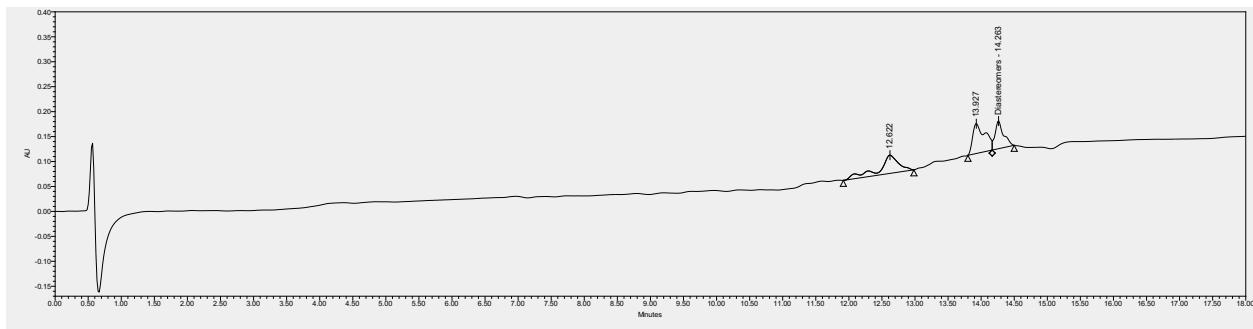
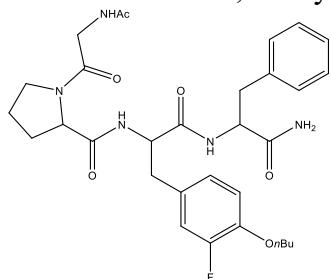


	Name	Retention Time	Area	% Area
1		12.607	281560	25.25
2		13.773	418266	37.52
3	Diastereomers	14.190	415074	37.23

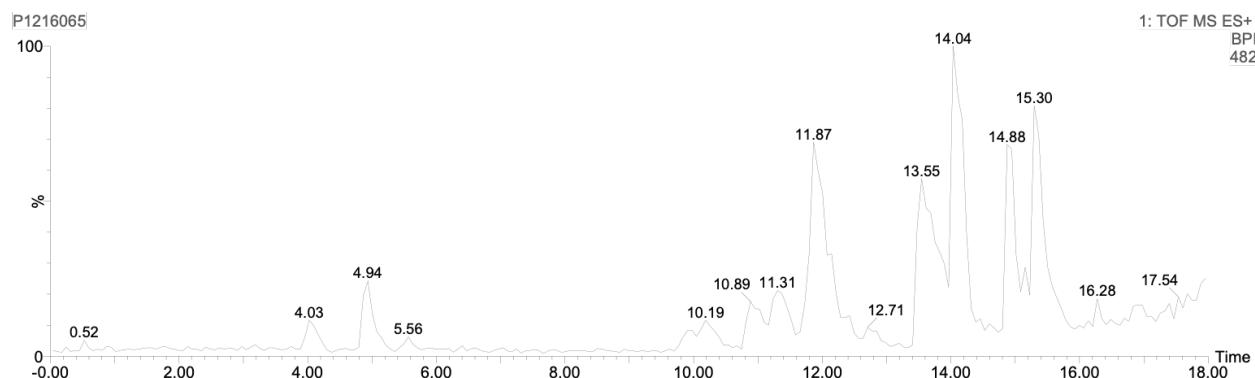


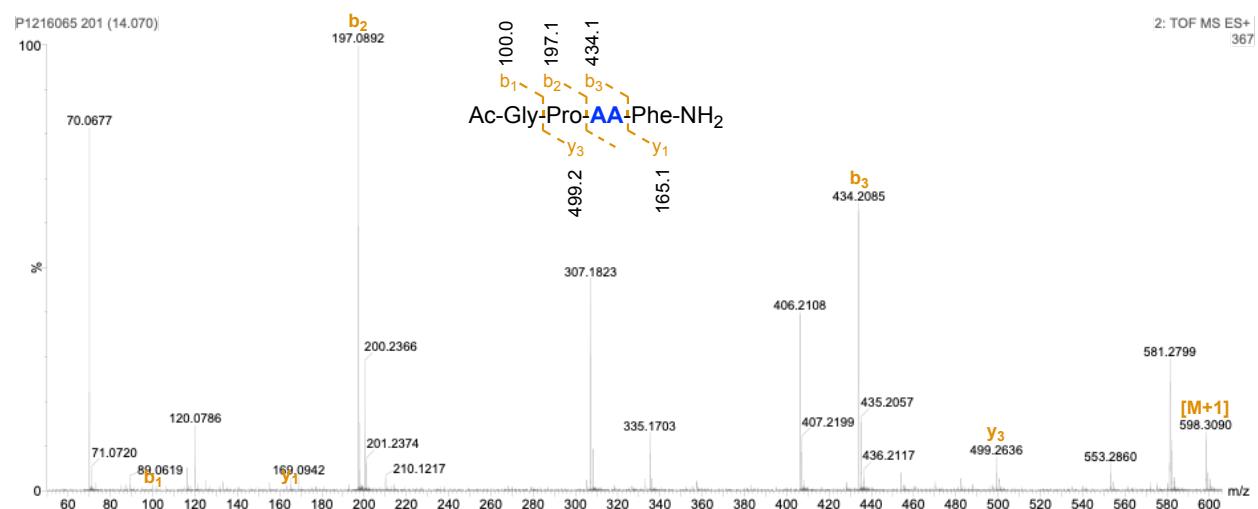
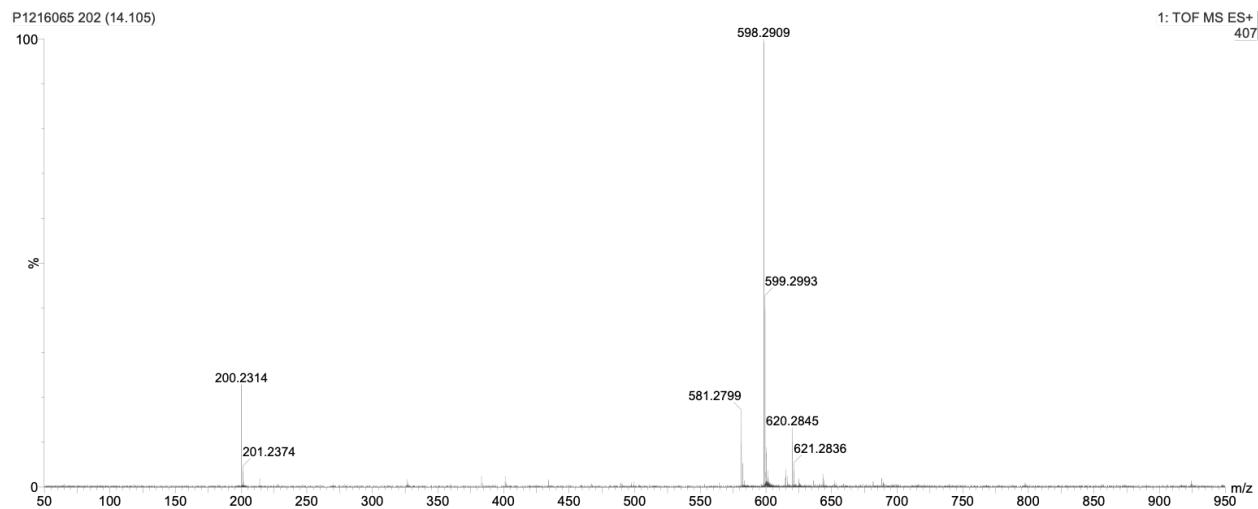


5F': MW = 597.7, Purity = 24.9%, Yield = 1.3% [0.019 mg]

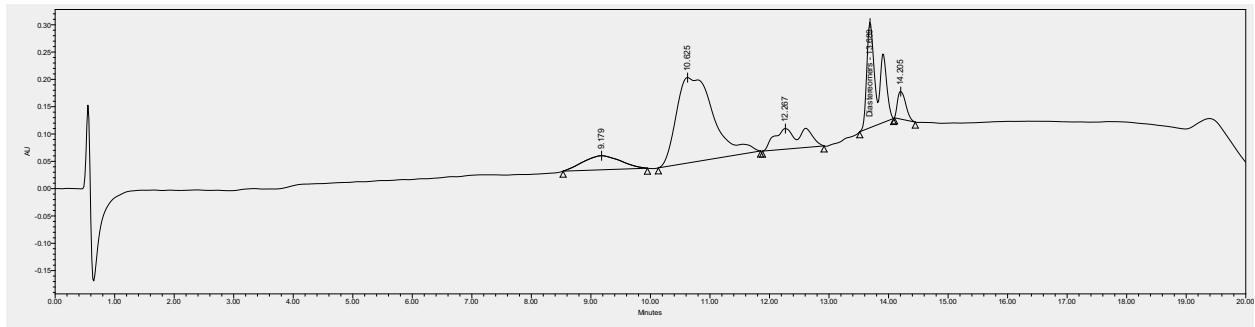
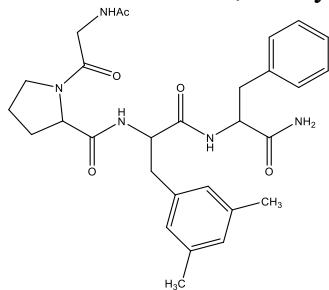


	Name	Retention Time	Area	% Area
1		12.622	742529	36.91
2		13.927	768435	38.20
3	Diastereomers	14.263	500821	24.89

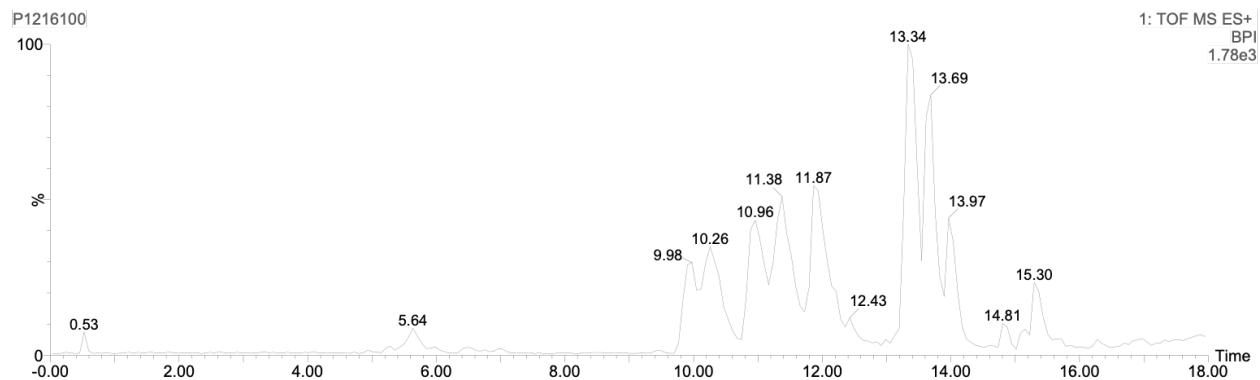


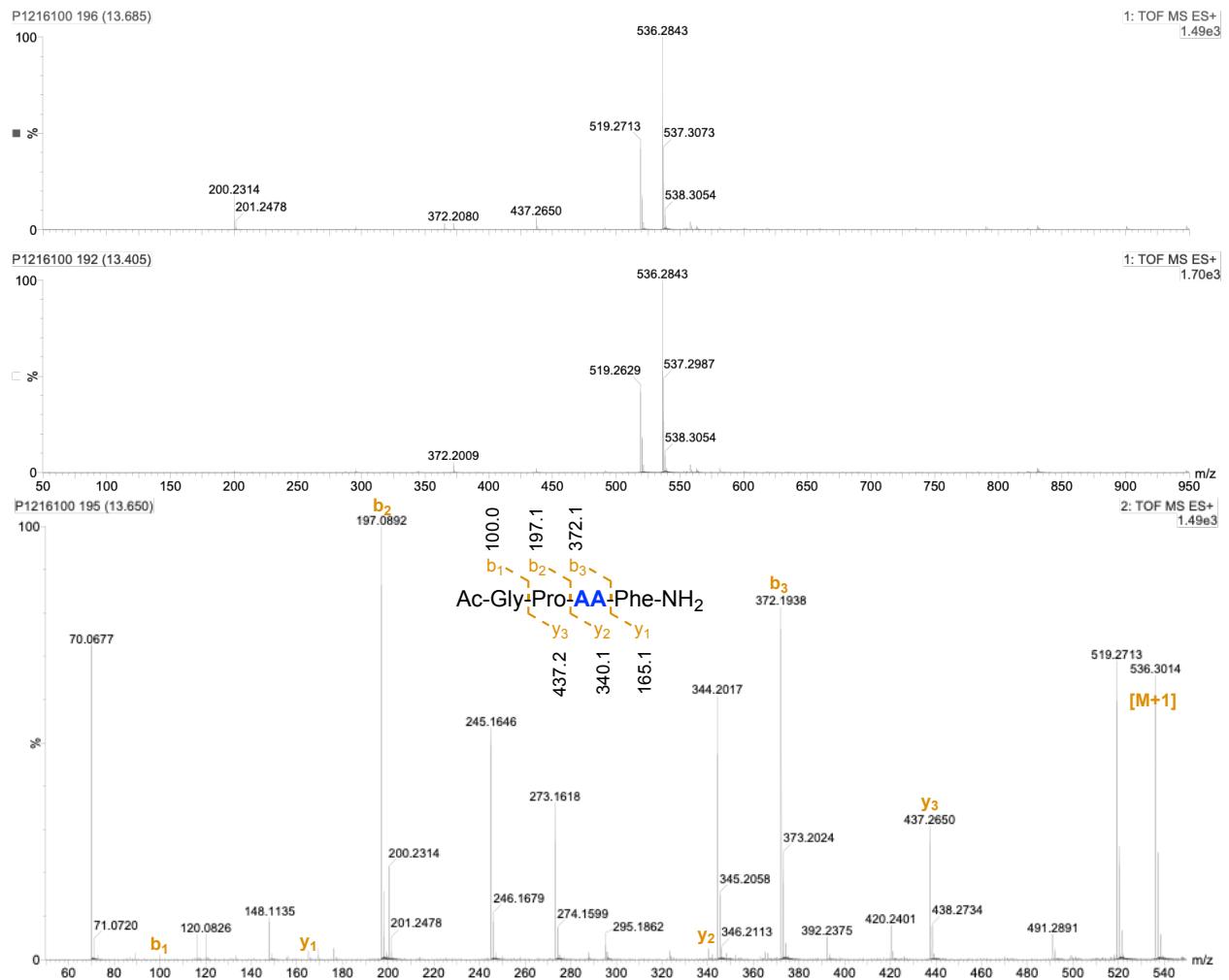


6F': MW = 535.7, Purity = 21.0%, Yield = 6.9% [0.087 mg]

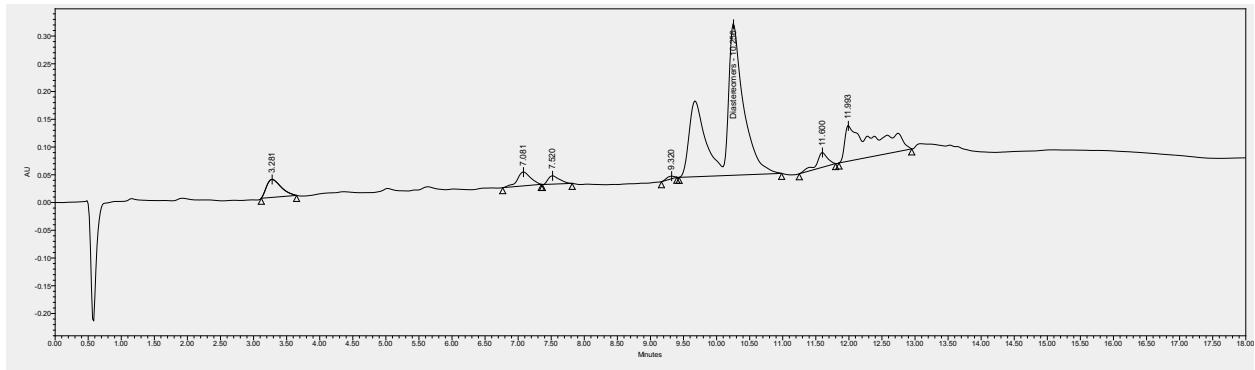
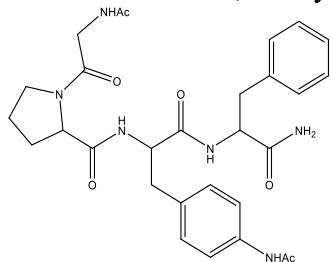


	Name	Retention Time	Area	% Area
1		9.179	1131627	9.22
2		10.625	6760693	55.06
3		12.267	1315720	10.72
4	Diastereomers	13.689	2582297	21.03
5		14.205	488816	3.98

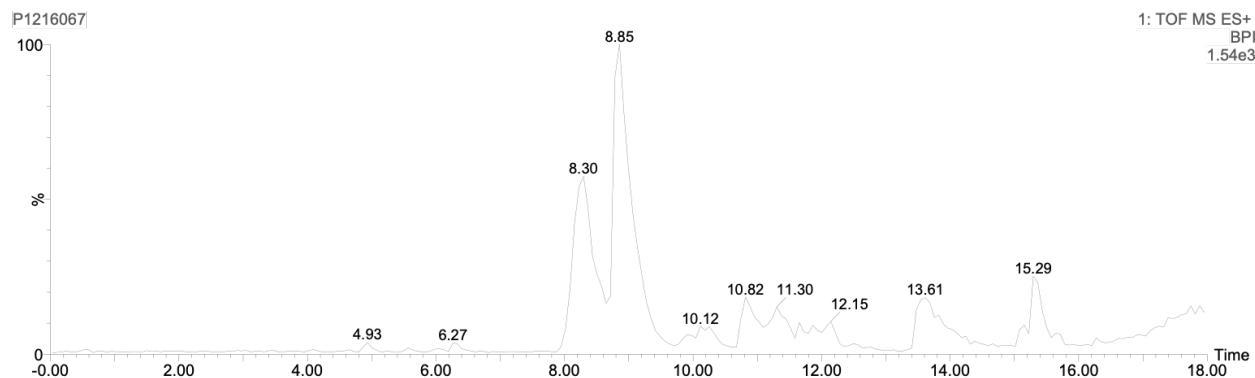


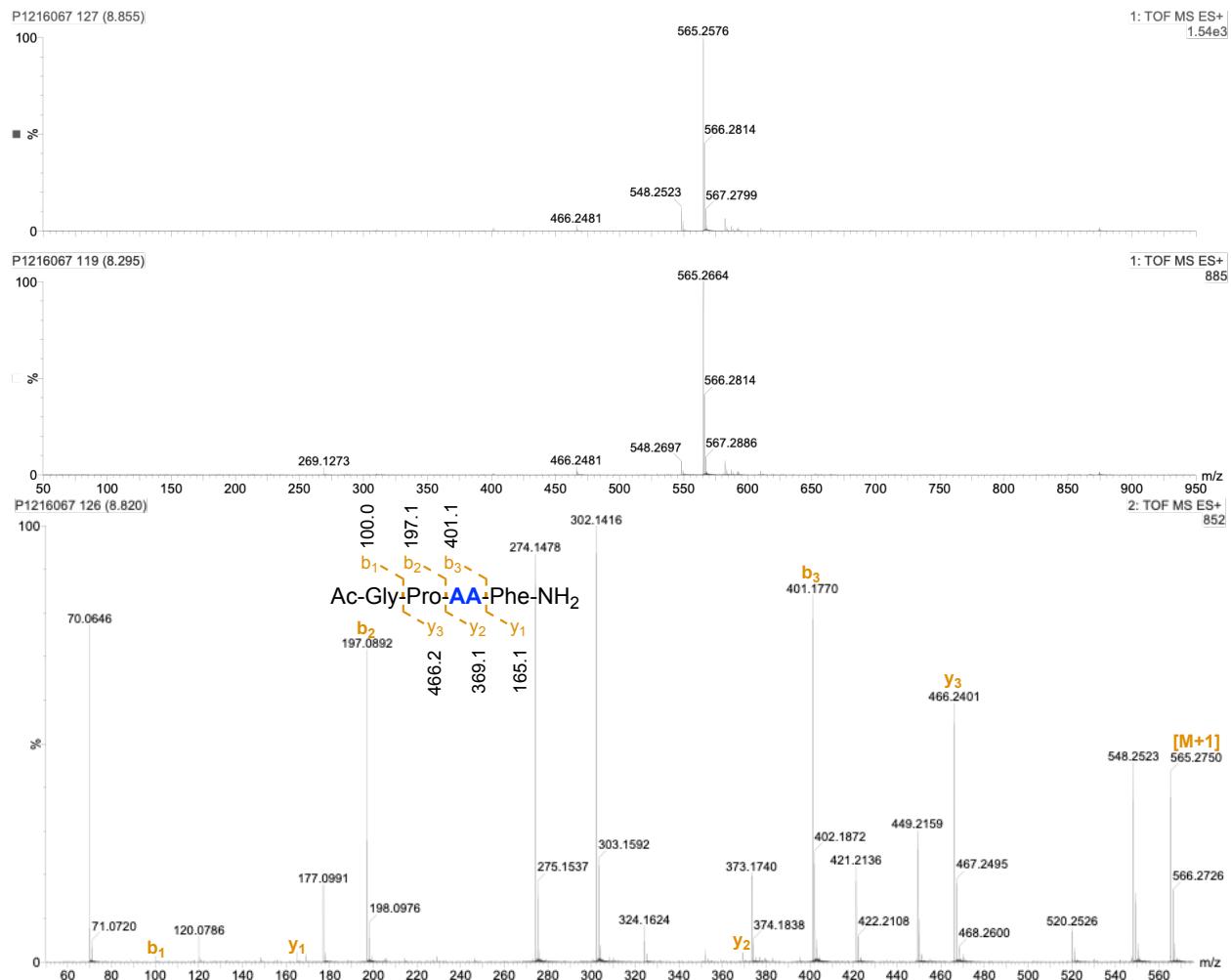


7F': MW = 564.6, Purity = 65.7%, Yield = 14.1% [0.19 mg]

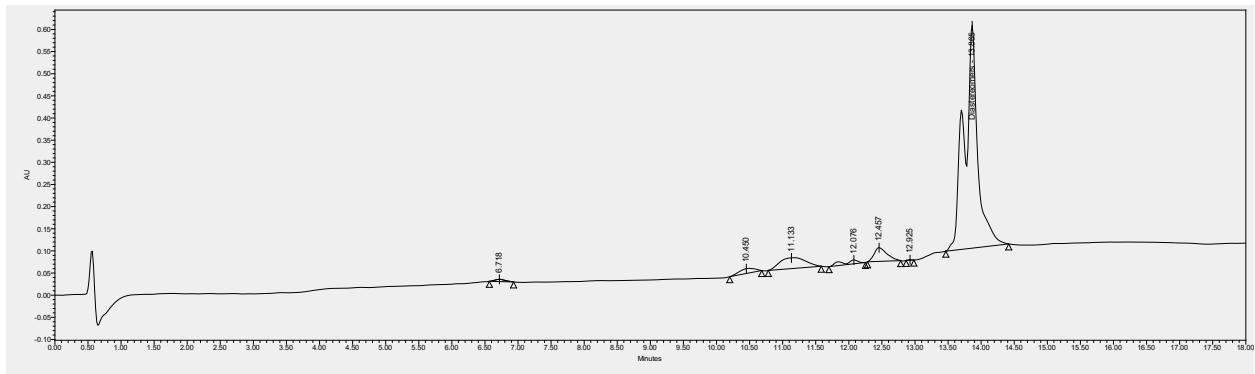
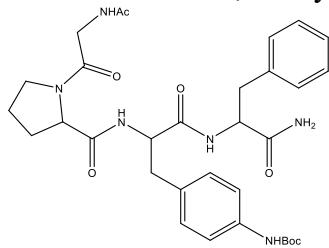


	Name	Retention Time	Area	% Area
1		3.281	496098	4.95
2		7.081	353073	3.53
3		7.520	179813	1.80
4		9.320	39917	0.40
5	Diastereomers	10.256	6577453	65.68
6		11.600	310559	3.10
7		11.993	2057287	20.54

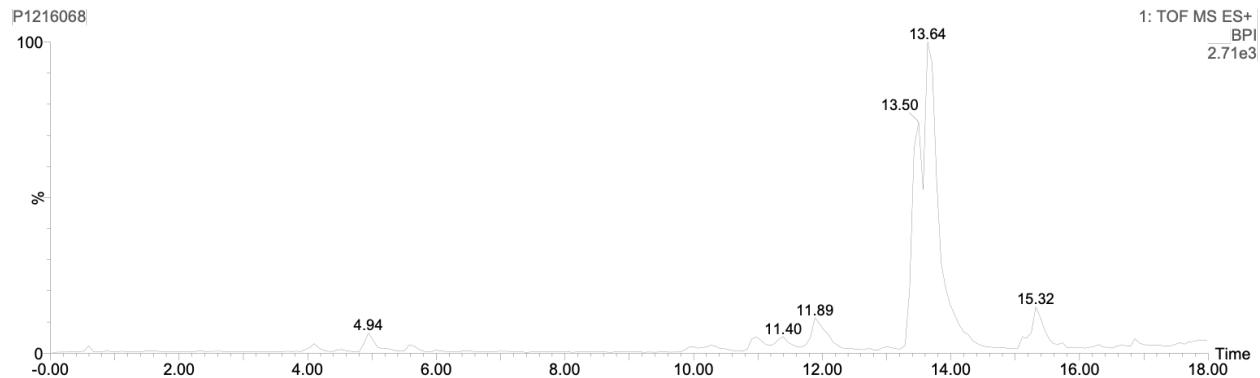


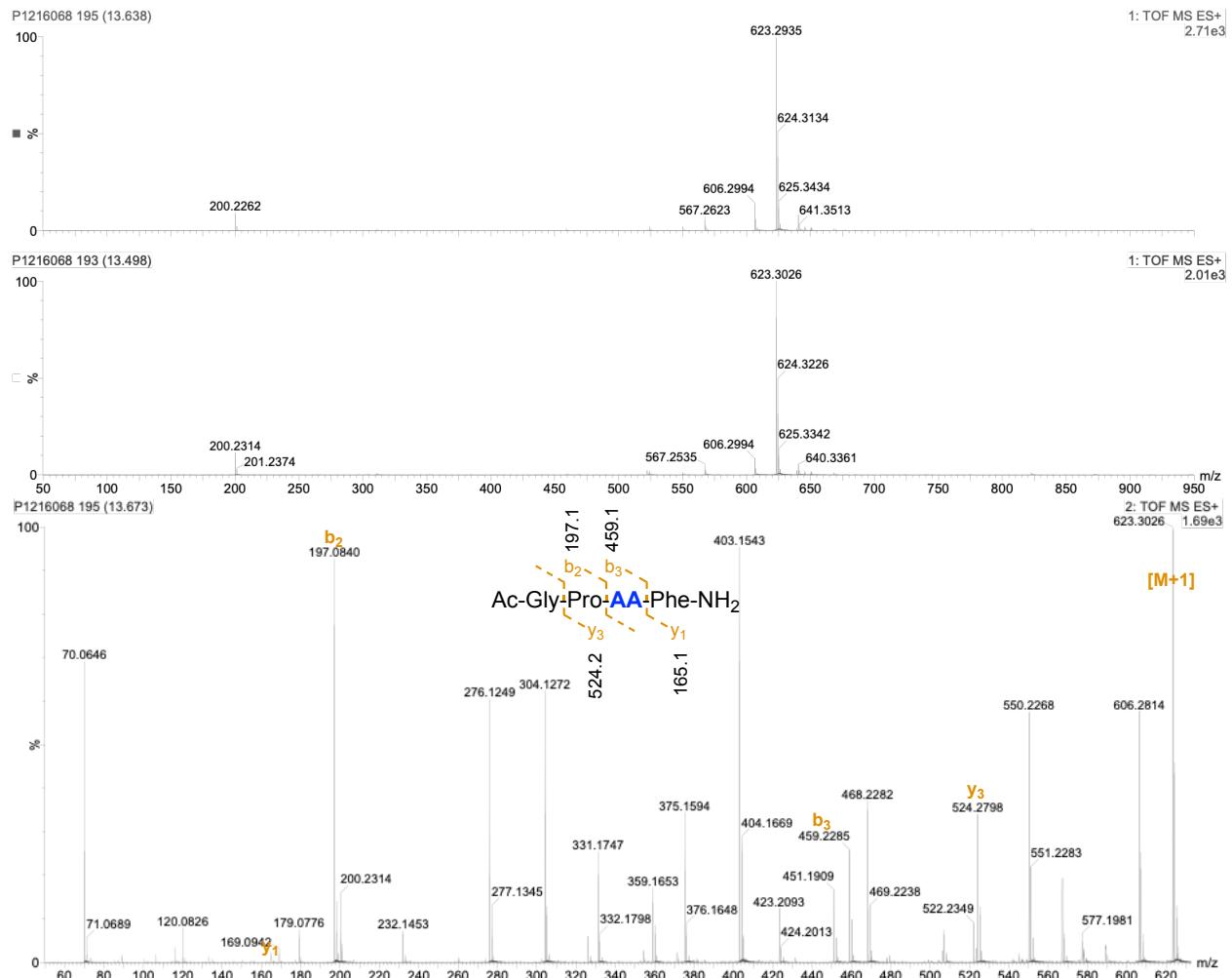


8F': MW = 622.7, Purity = 82.8%, Yield = 21.0% [0.31 mg]

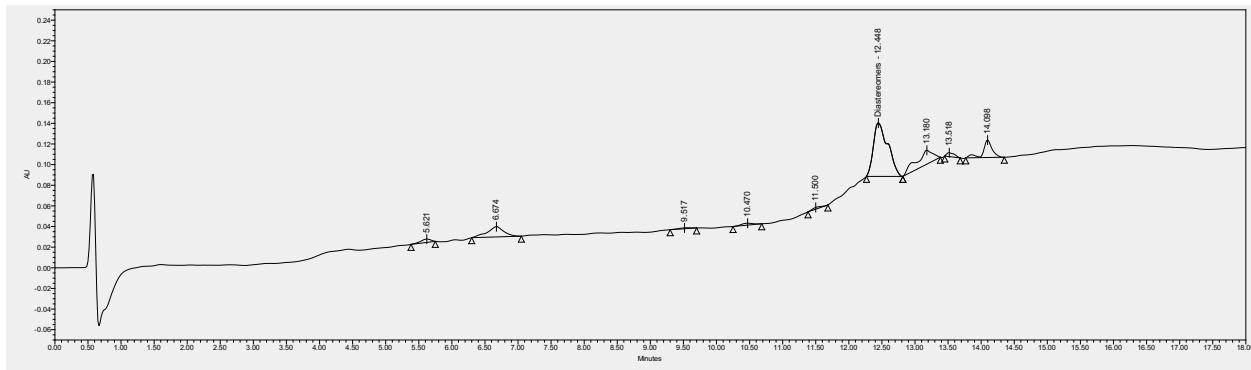
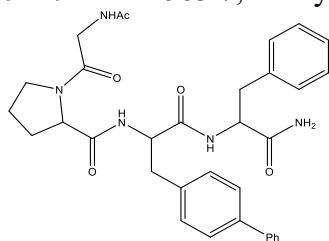


	Name	Retention Time	Area	% Area
1		6.718	56341	0.66
2		10.450	180159	2.10
3		11.133	666593	7.77
4		12.076	148717	1.73
5		12.457	417768	4.87
6		12.925	4582	0.05
7	Diastereomers	13.865	7109923	82.83

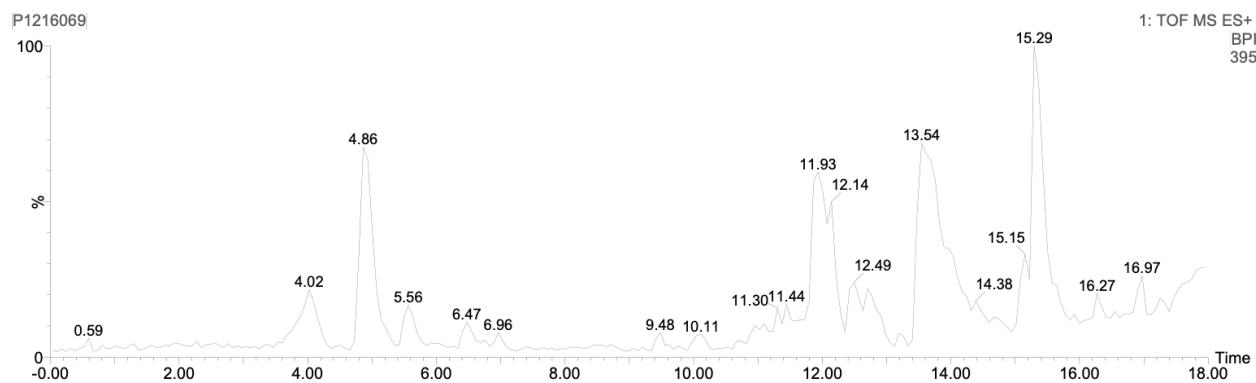


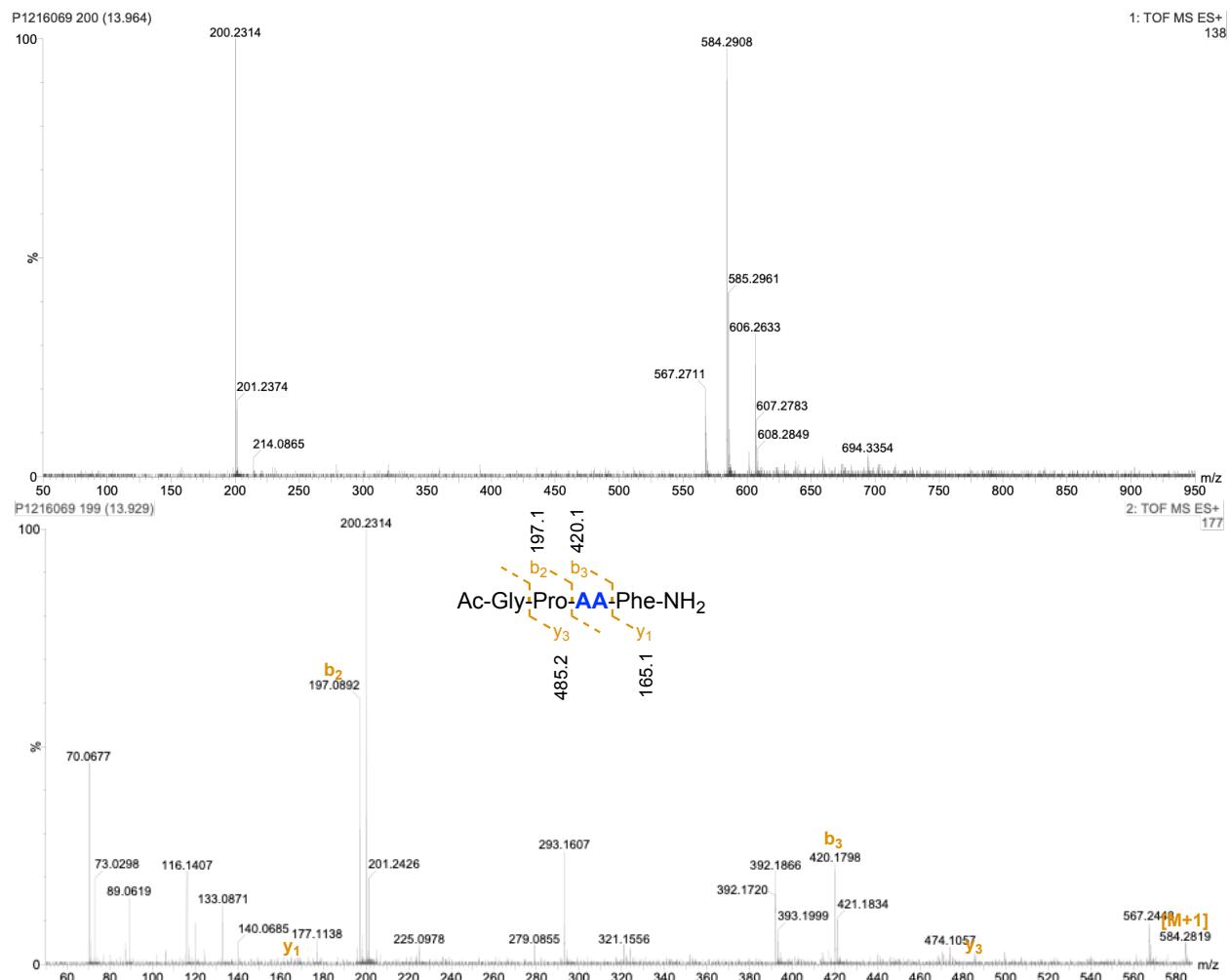


9F': MW = 583.7, Purity = 53.4%, Yield = 1.2% [0.017 mg]

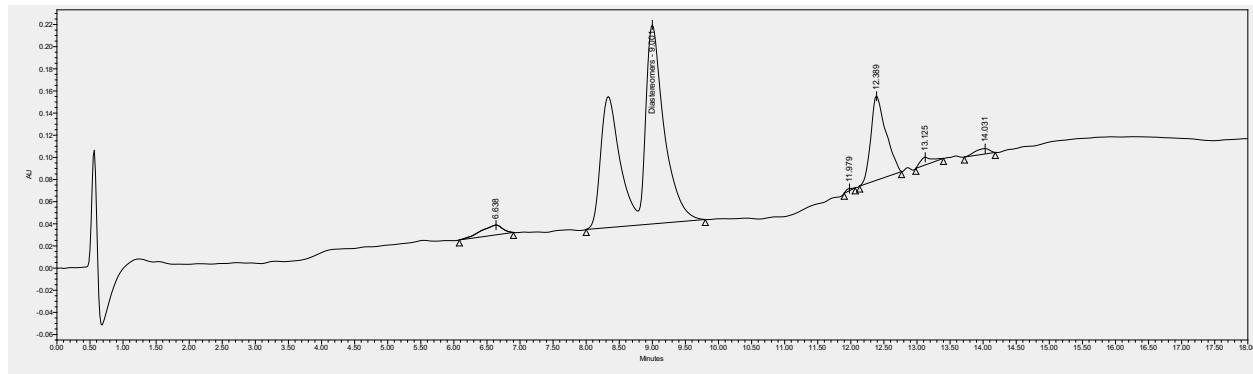
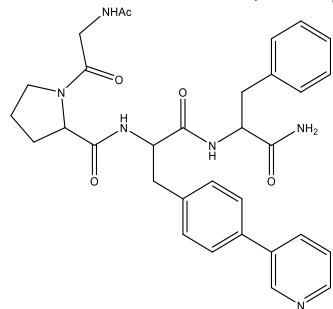


	Name	Retention Time	Area	% Area
1		5.621	35000	2.37
2		6.674	174226	11.82
3		9.517	11326	0.77
4		10.470	22624	1.53
5		11.500	14982	1.02
6	Diastereomers	12.448	787254	53.41
7		13.180	239573	16.25
8		13.518	30960	2.10
9		14.098	158116	10.73

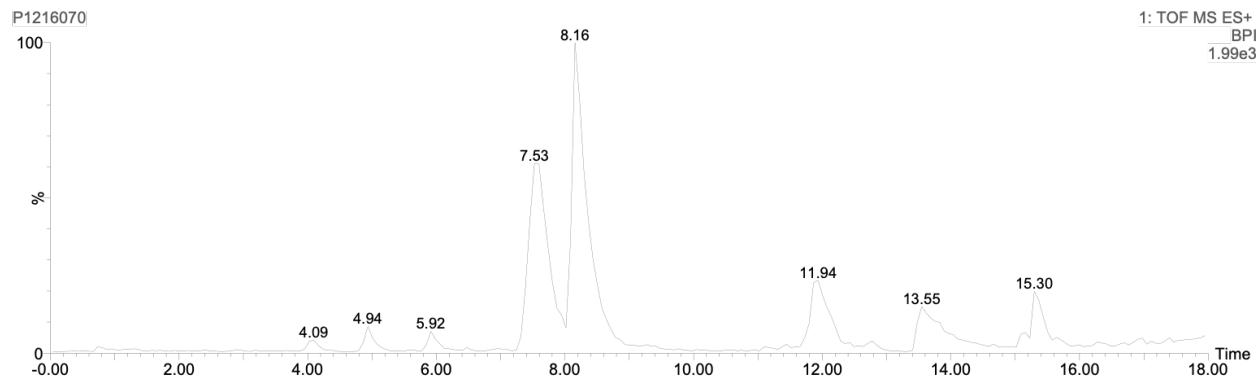


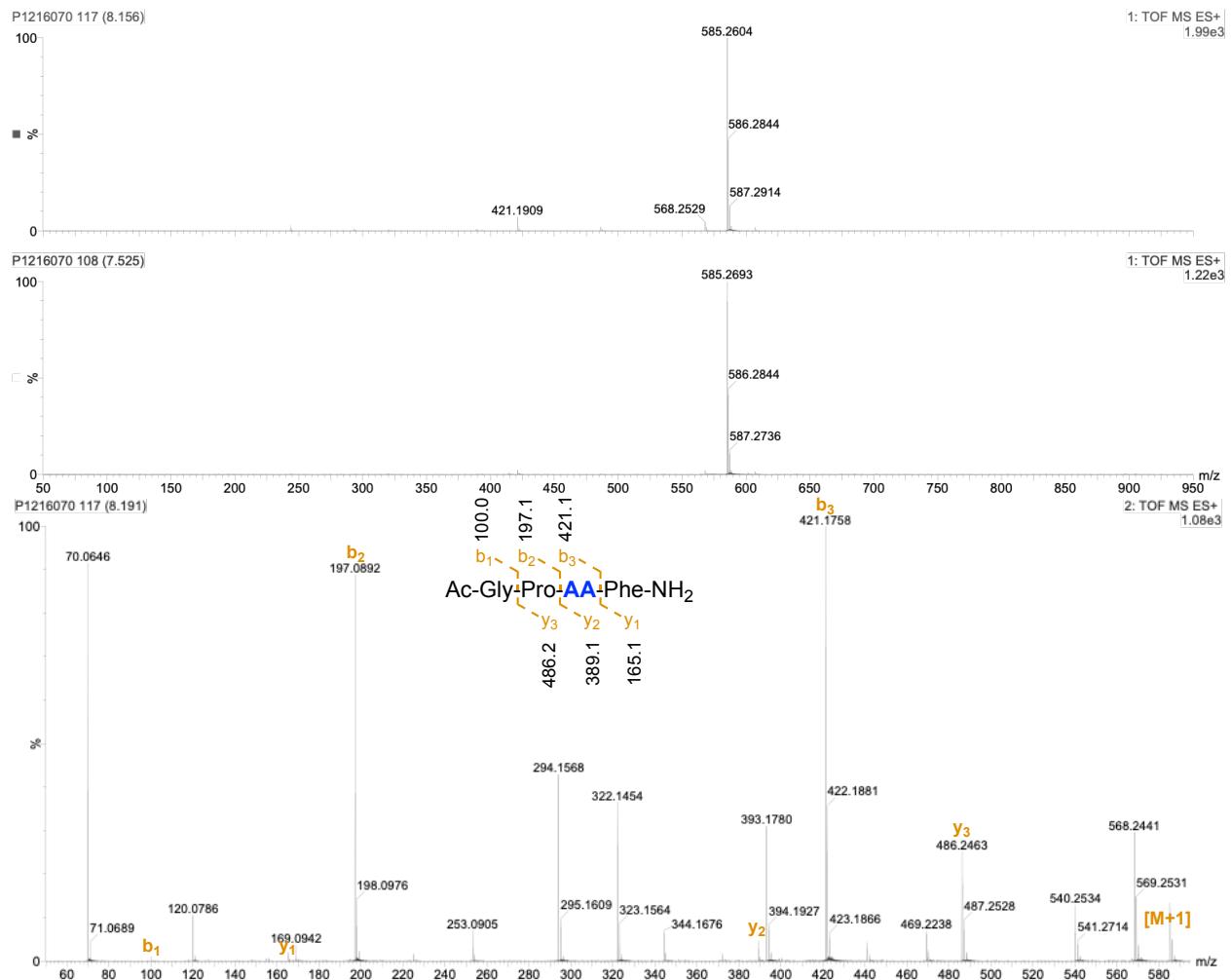


10F': MW = 584.7, Purity = 78.8%, Yield = 16.0% [0.22 mg]

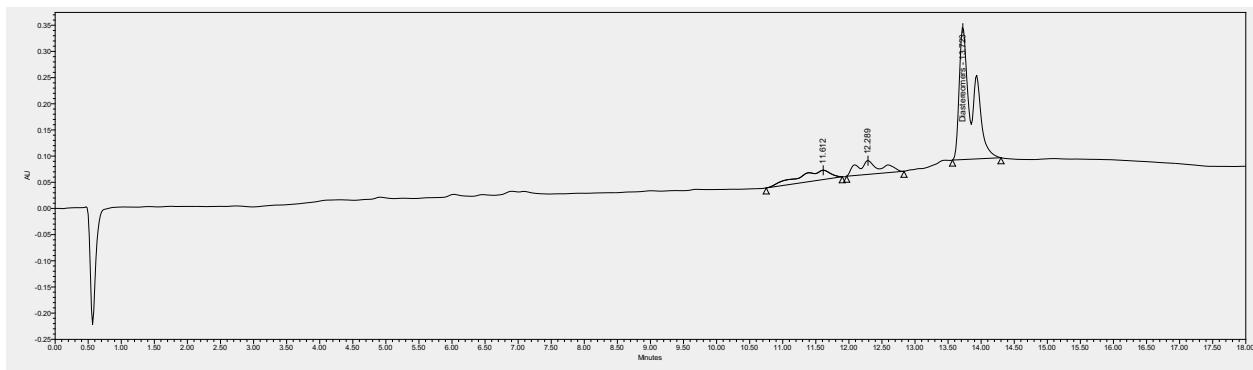
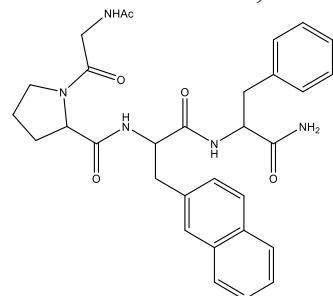


	Name	Retention Time	Area	% Area
1		6.638	203748	2.74
2	Diastereomers	9.001	5849060	78.67
3		11.979	12251	0.16
4		12.389	1207018	16.23
5		13.125	84923	1.14
6		14.031	77733	1.05

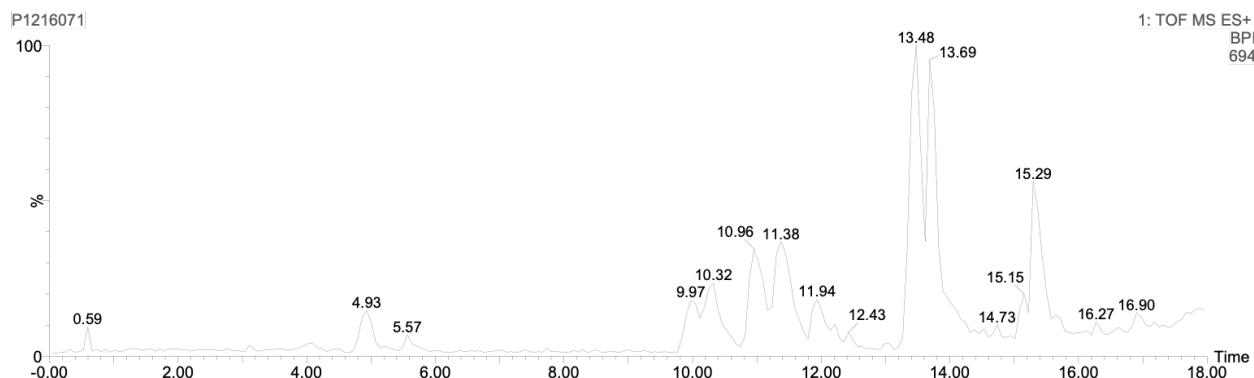


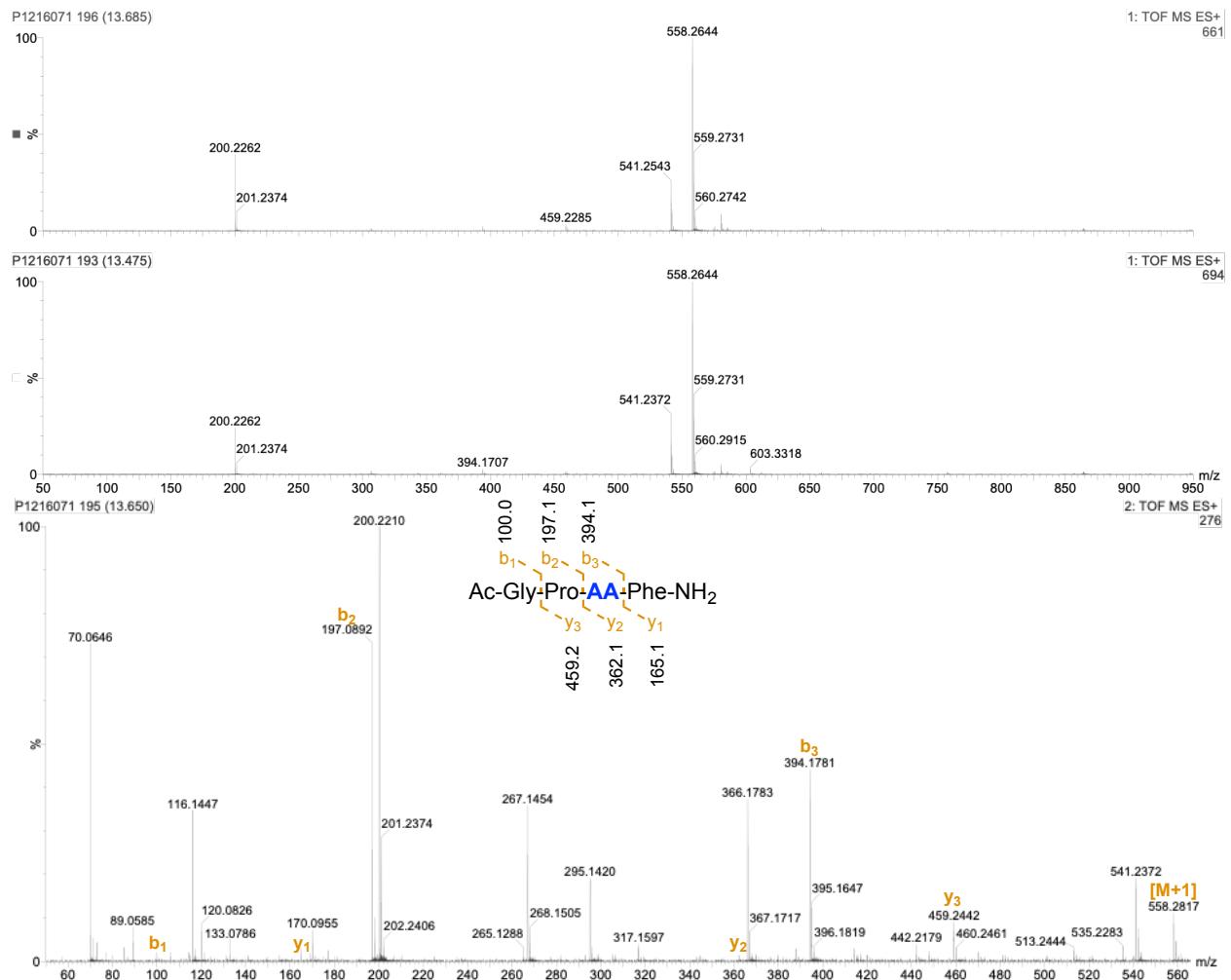


11F': MW = 557.7, Purity = 72.8%, Yield = 3.4% [0.044 mg]

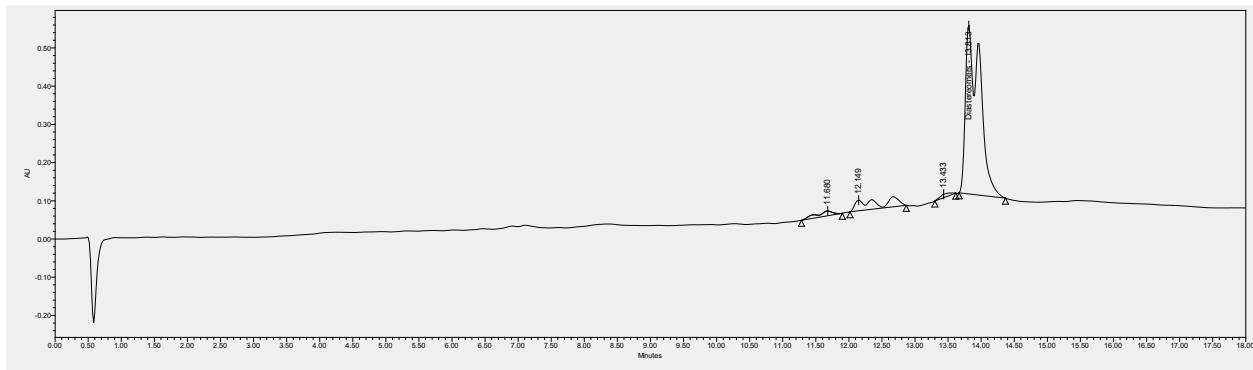
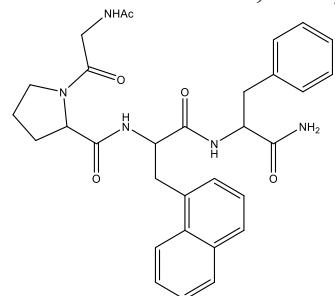


	Name	Retention Time	Area	% Area
1		11.612	663090	13.73
2		12.289	650249	13.46
3	Diastereomers	13.723	3517431	72.81

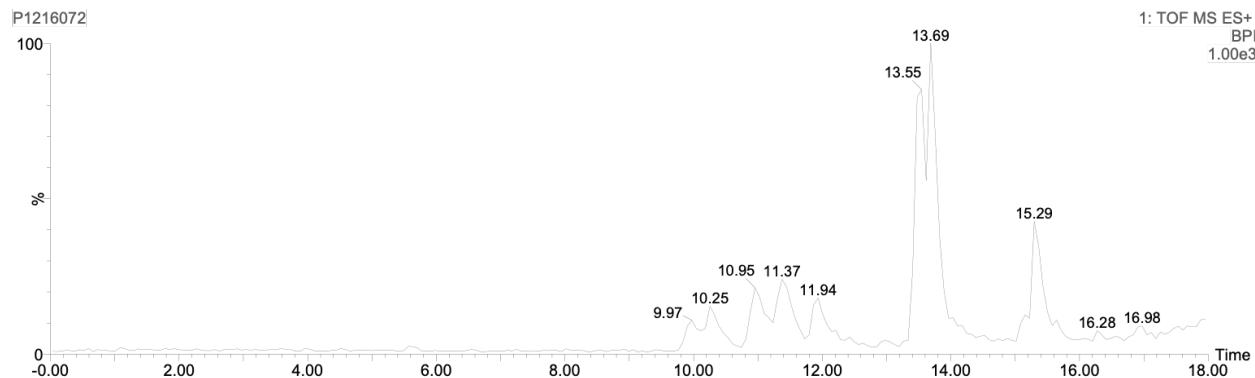


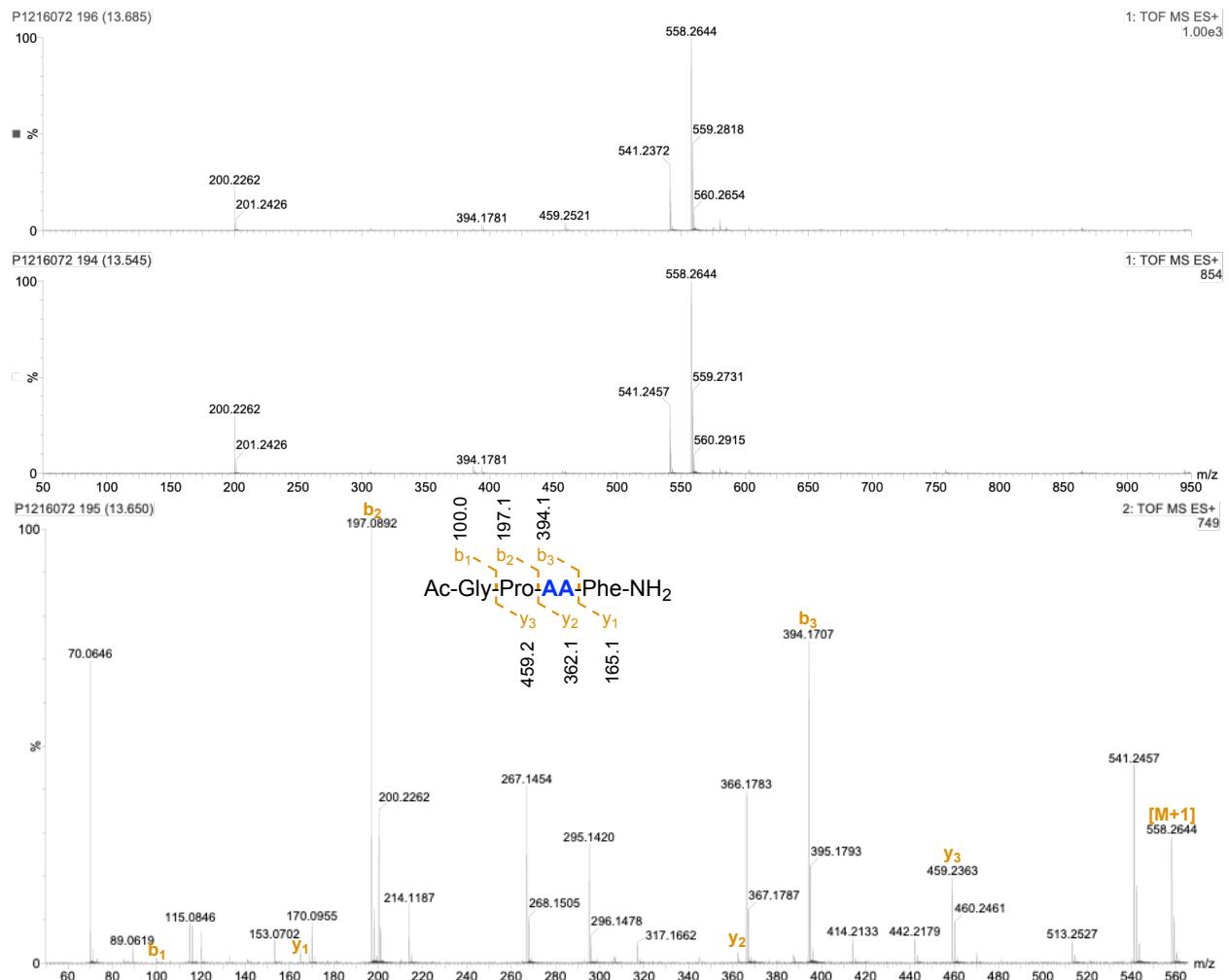


12F': MW = 557.7, Purity = 86.1%, Yield = 6.5% [0.084 mg]

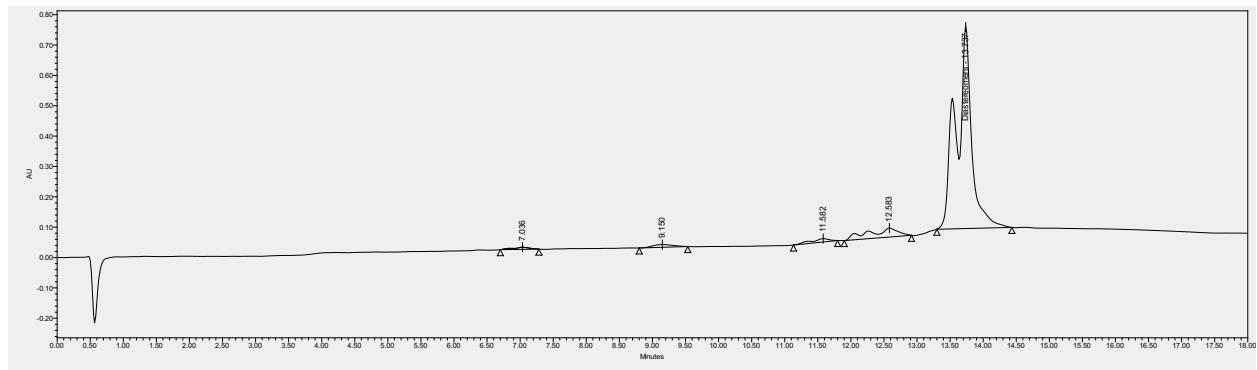
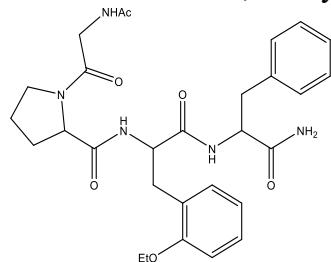


	Name	Retention Time	Area	% Area
1		11.680	227134	2.89
2		12.149	767459	9.76
3		13.433	100495	1.28
4	Diastereomers	13.813	6768724	86.07

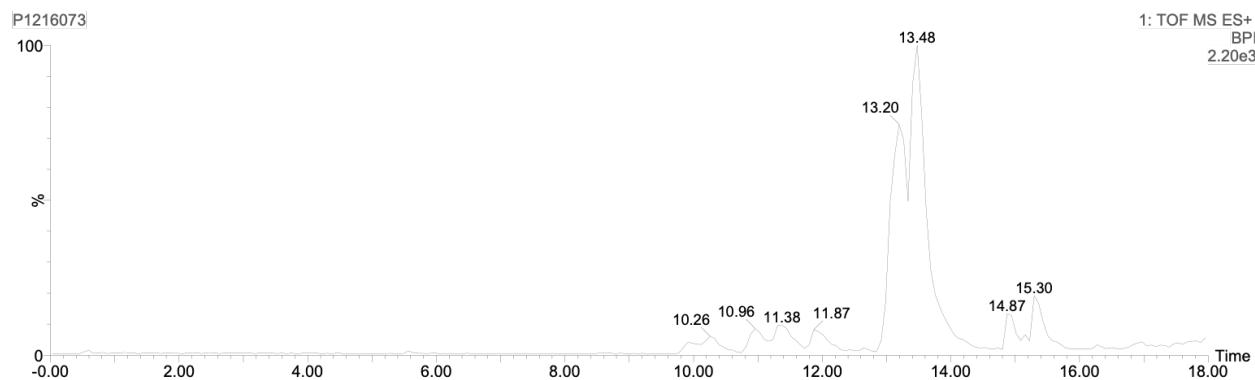


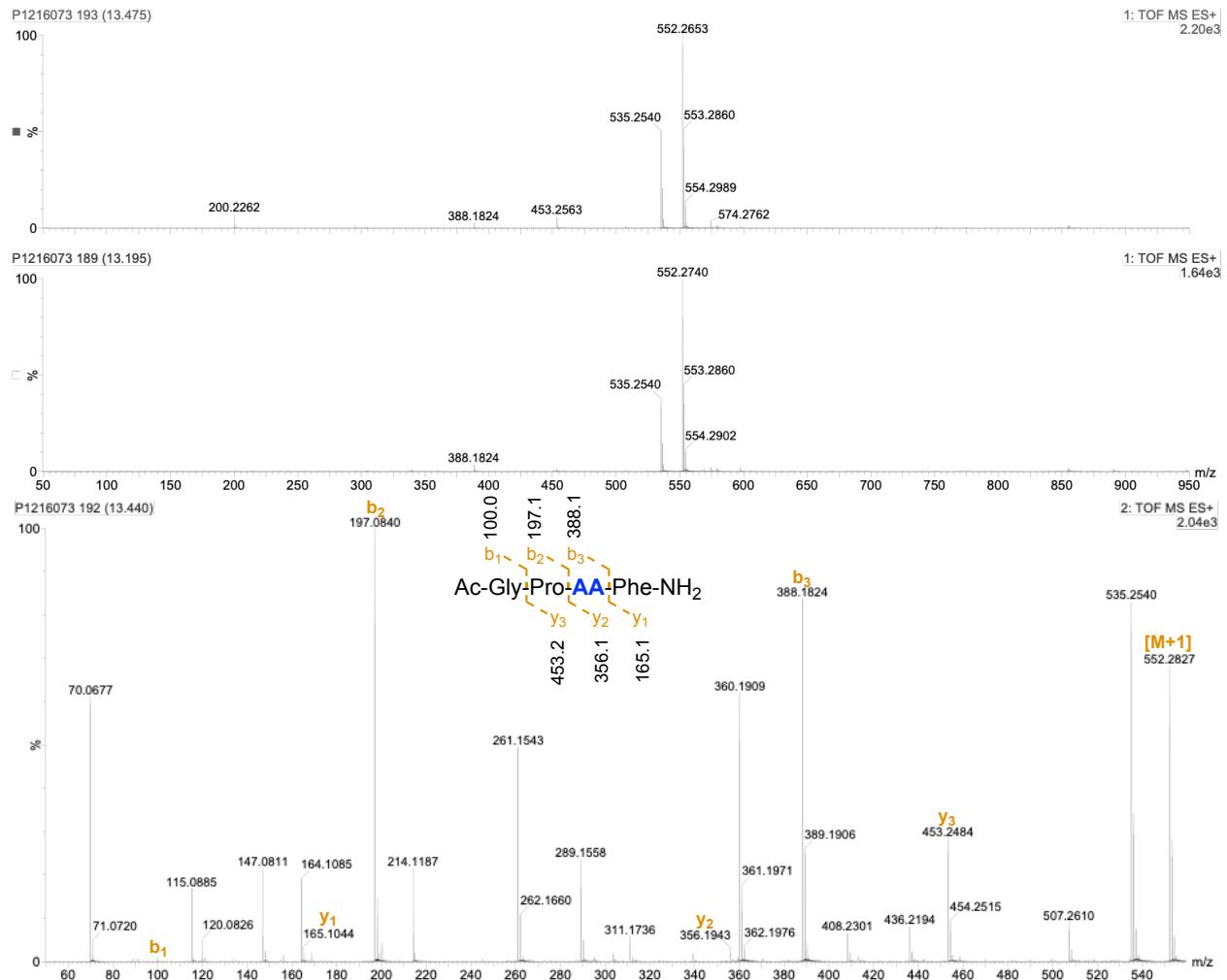


1G': MW = 551.6, Purity = 87.9%, Yield = 23.1% [0.30 mg]

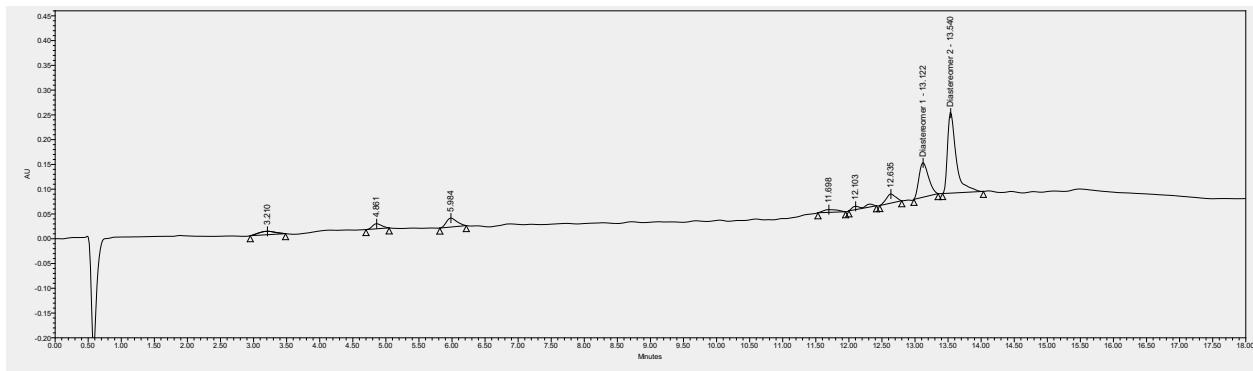
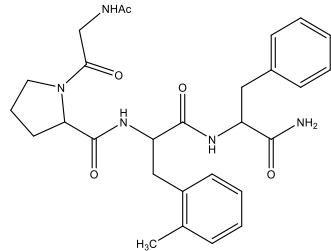


	Name	Retention Time	Area	% Area
1		7.036	125645	1.03
2		9.150	194099	1.59
3		11.582	218590	1.79
4		12.583	941999	7.70
5	Diastereomers	13.737	10750959	87.90

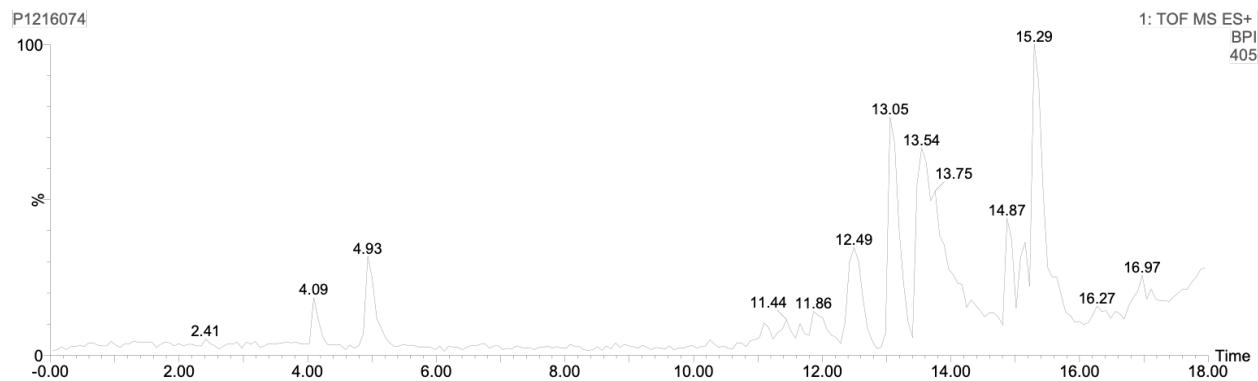


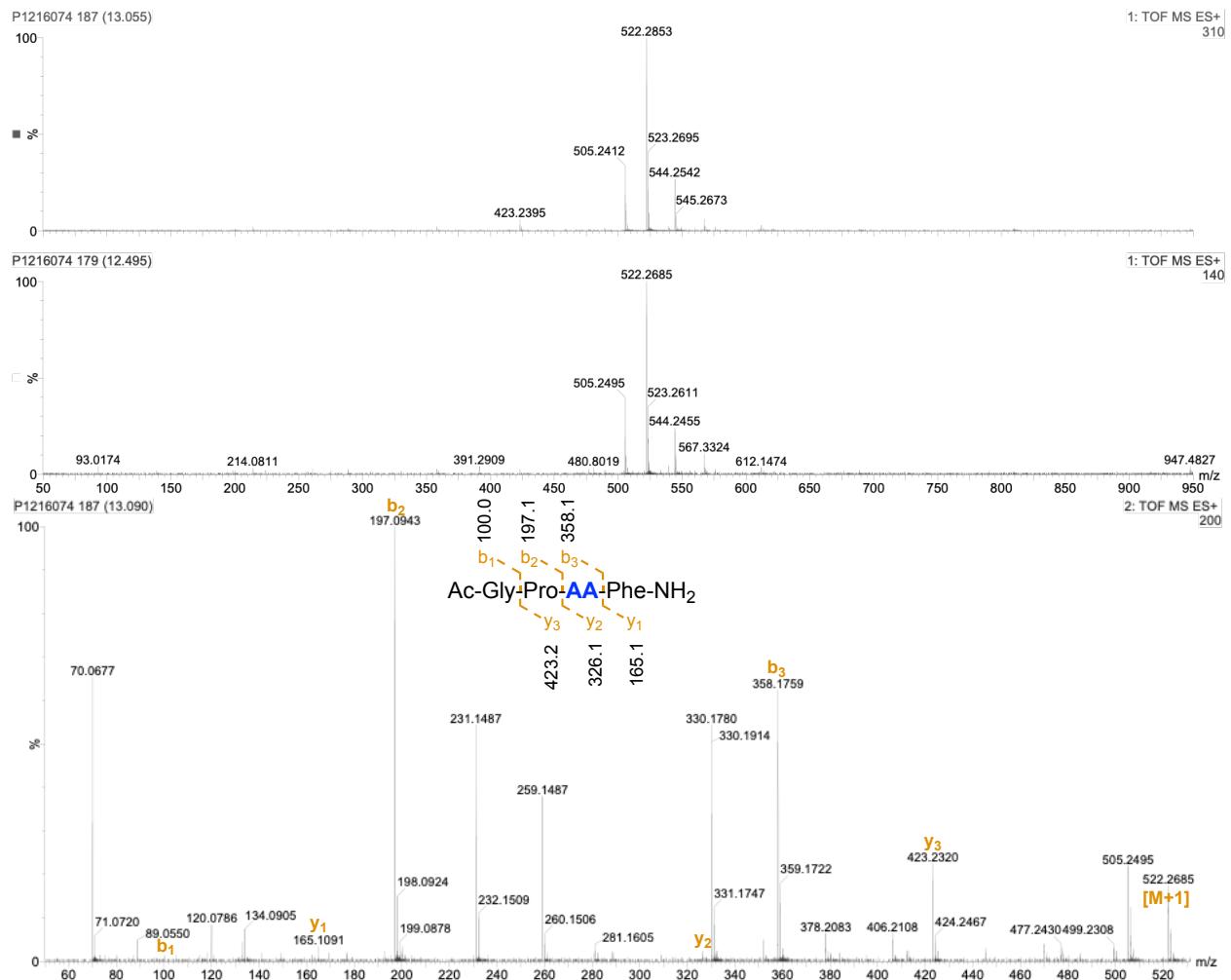


2G': MW = 521.6, Purity = 73.5%, Yield = 4.7% [0.057 mg]

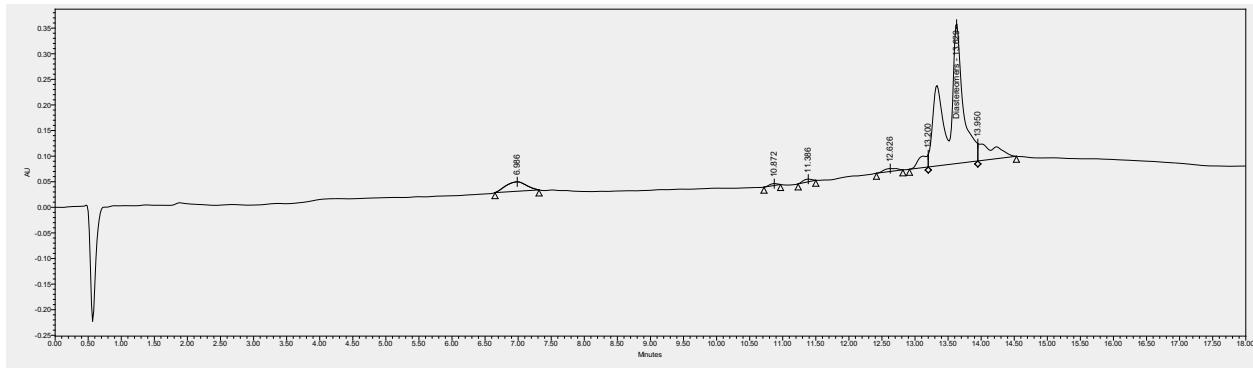
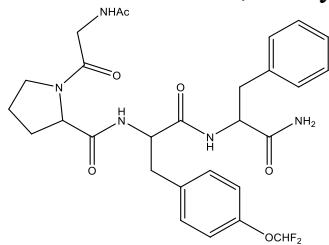


	Name	Retention Time	Area	% Area
1		3.210	123987	4.18
2		4.861	100252	3.38
3		5.984	200970	6.77
4		11.698	84061	2.83
5		12.103	96460	3.25
6		12.635	181686	6.12
7	Diastereomer 1	13.122	700423	23.61
8	Diastereomer 2	13.540	1479393	49.86

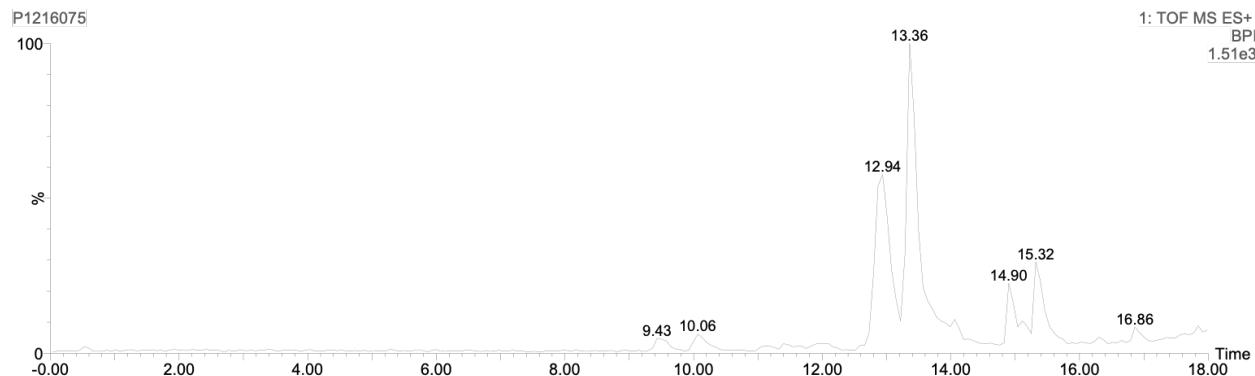


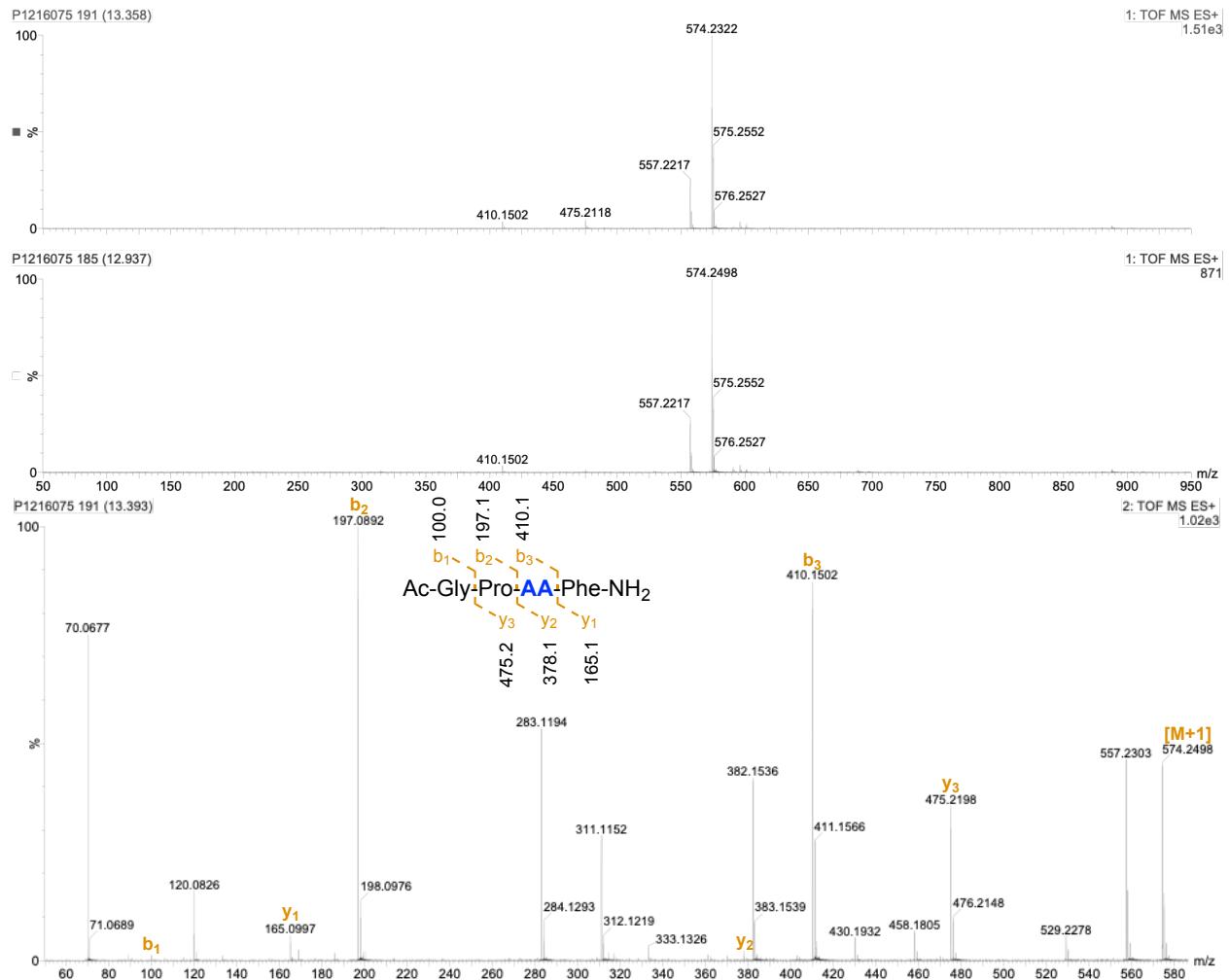


3G': MW = 573.6, Purity = 76.9%, Yield = 10.0% [0.13 mg]

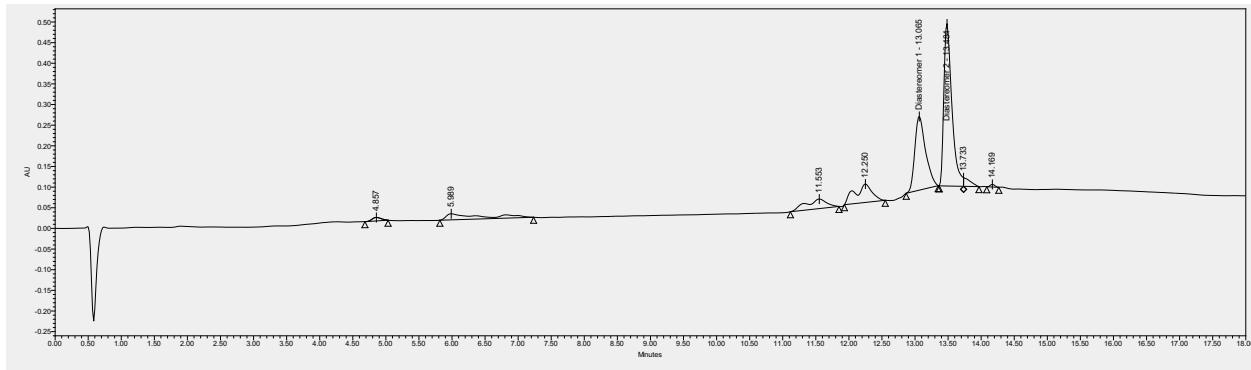
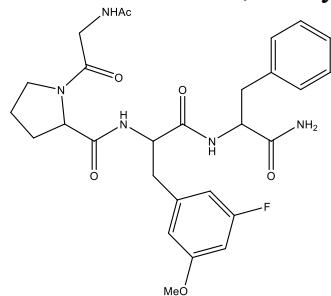


	Name	Retention Time	Area	% Area
1		6.986	402354	6.64
2		10.872	32296	0.53
3		11.386	50046	0.83
4		12.626	79204	1.31
5		13.200	231030	3.81
6	Diastereomers	13.629	4656666	76.86
7		13.950	607219	10.02

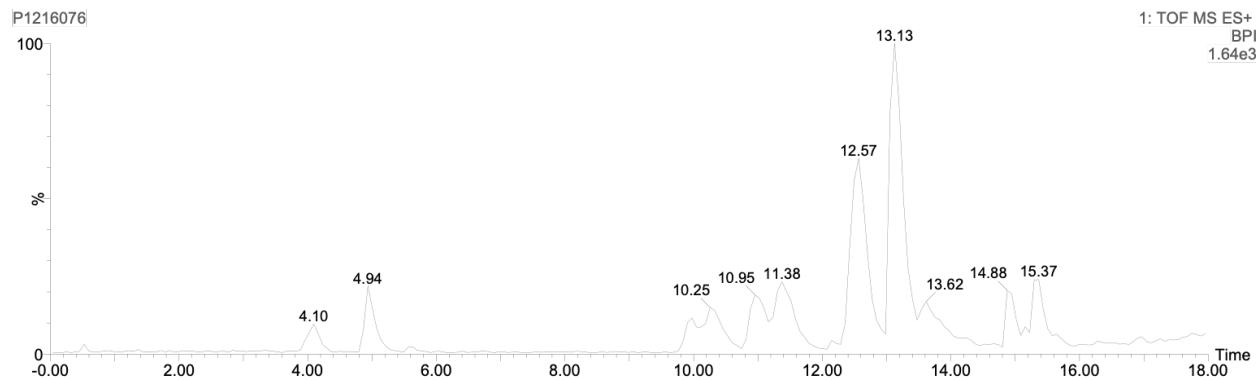


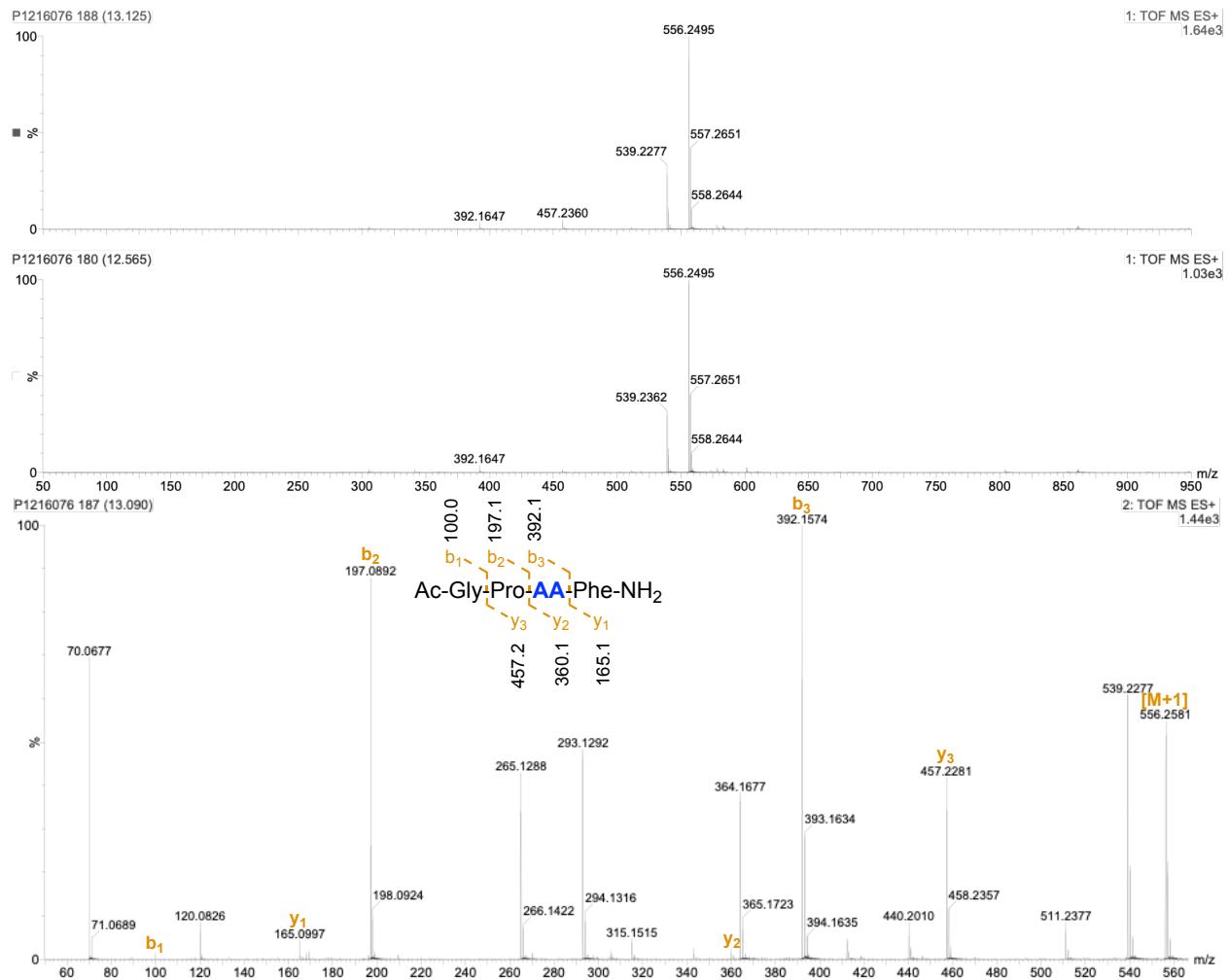


4G': MW = 555.6, Purity = 71.7%, Yield = 11.5% [0.15 mg]

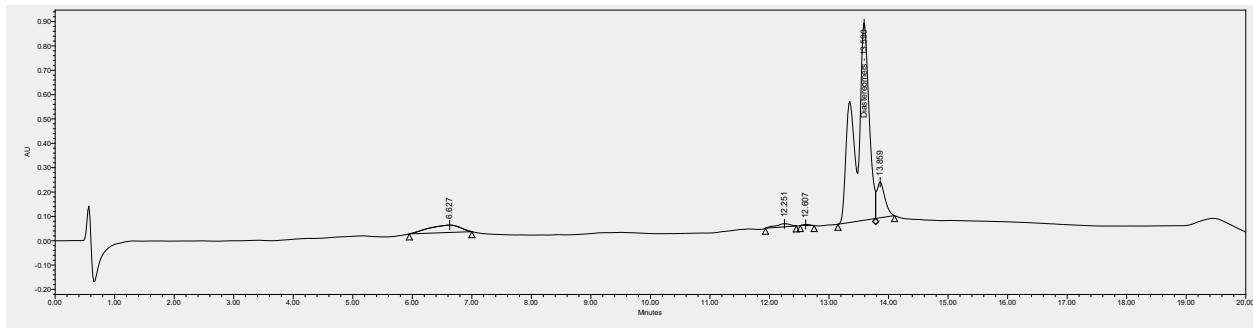
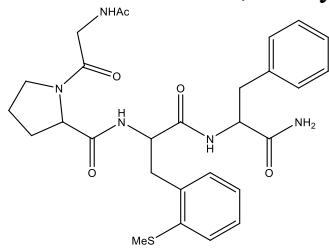


	Name	Retention Time	Area	% Area
1		4.857	80885	1.09
2		5.989	531549	7.14
3		11.553	512758	6.89
4		12.250	804752	10.81
5	Diastereomer 1	13.065	2017411	27.09
6	Diastereomer 2	13.484	3319915	44.58
7		13.733	148709	2.00
8		14.169	30776	0.41

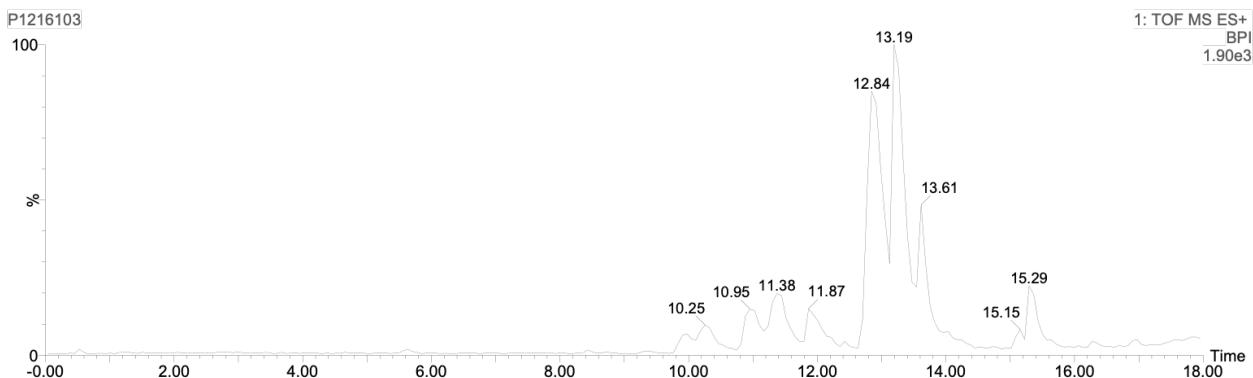


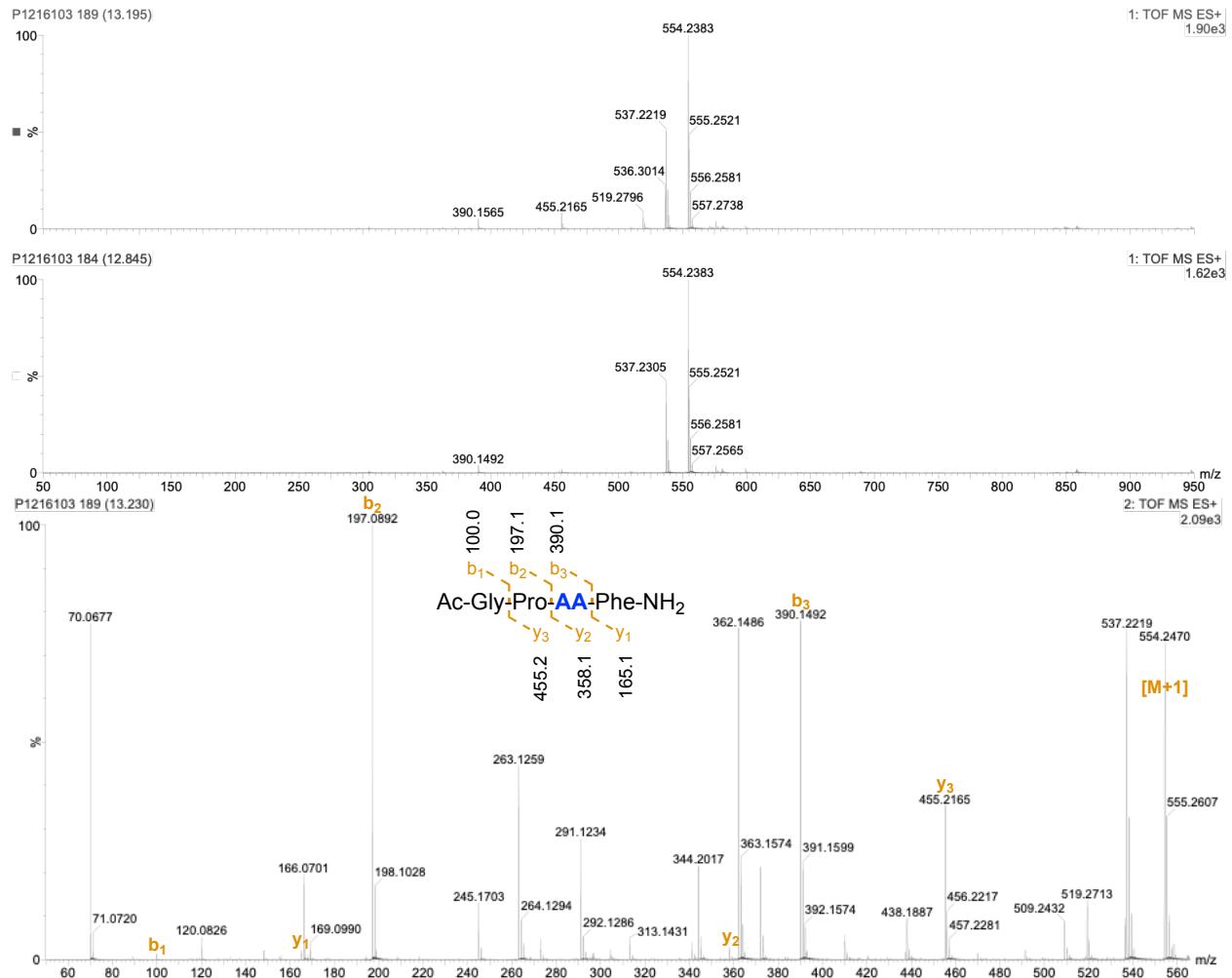


5G': MW = 553.7, Purity = 82.4%, Yield = 35.2% [0.45 mg]

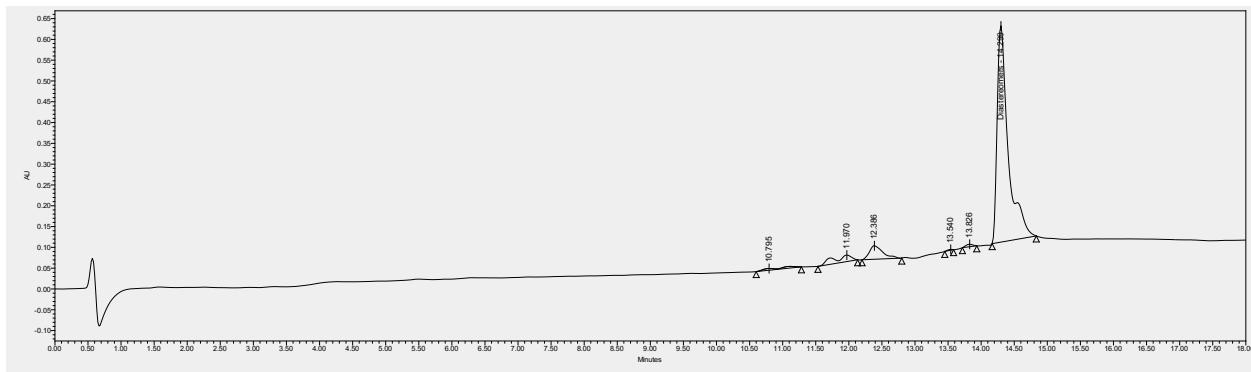
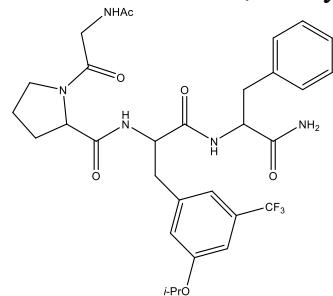


	Name	Retention Time	Area	% Area
1		6.627	1125799	7.09
2		12.251	220958	1.39
3		12.607	38009	0.24
4	Diastereomers	13.590	13083460	82.40
5		13.859	1409523	8.88

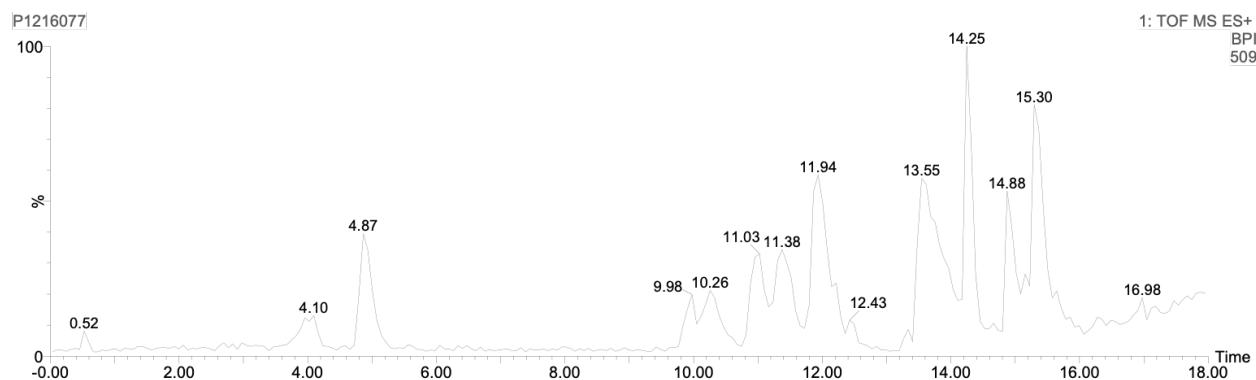


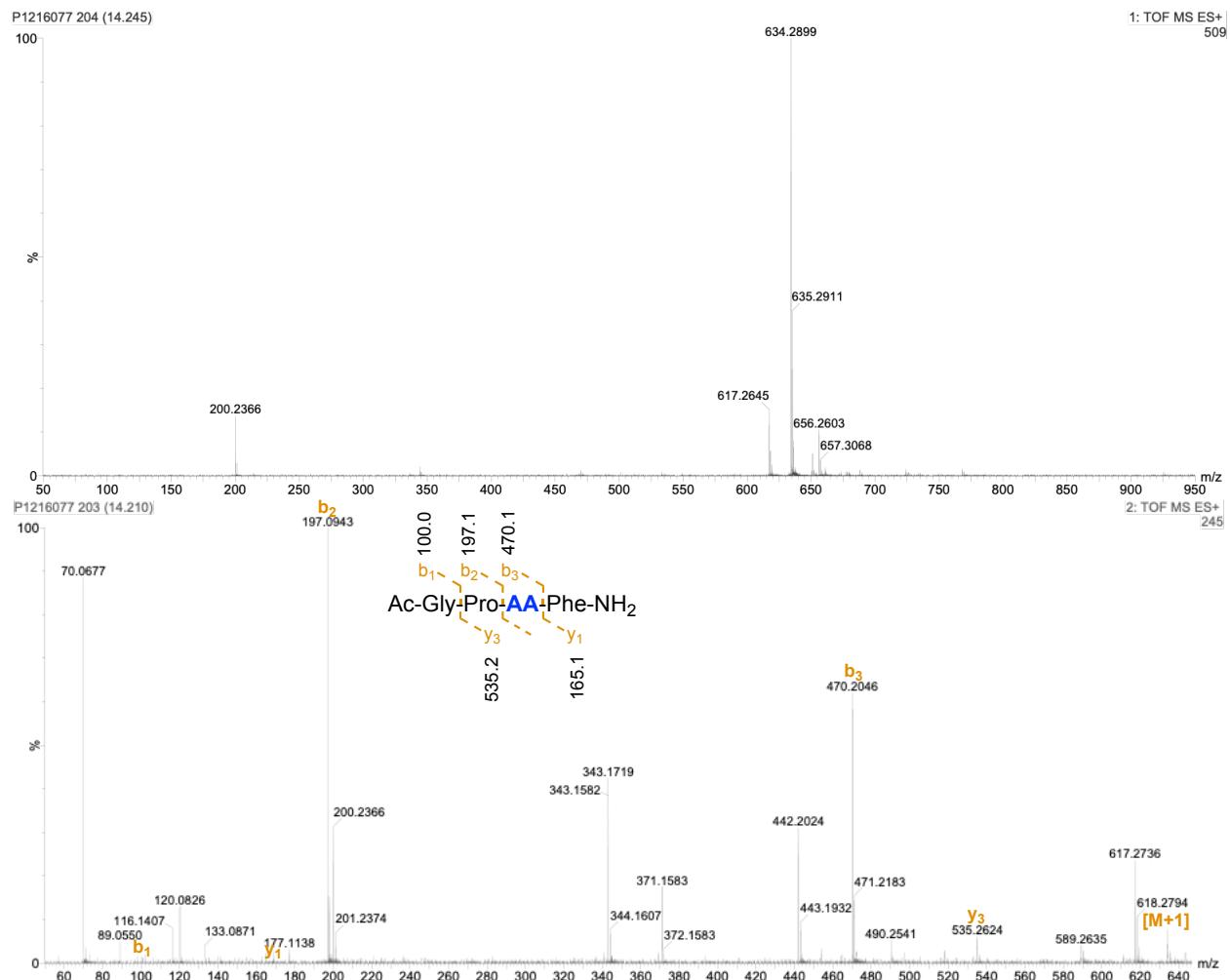


6G': MW = 633.7, Purity = 86.7%, Yield = 17.8% [0.26 mg]

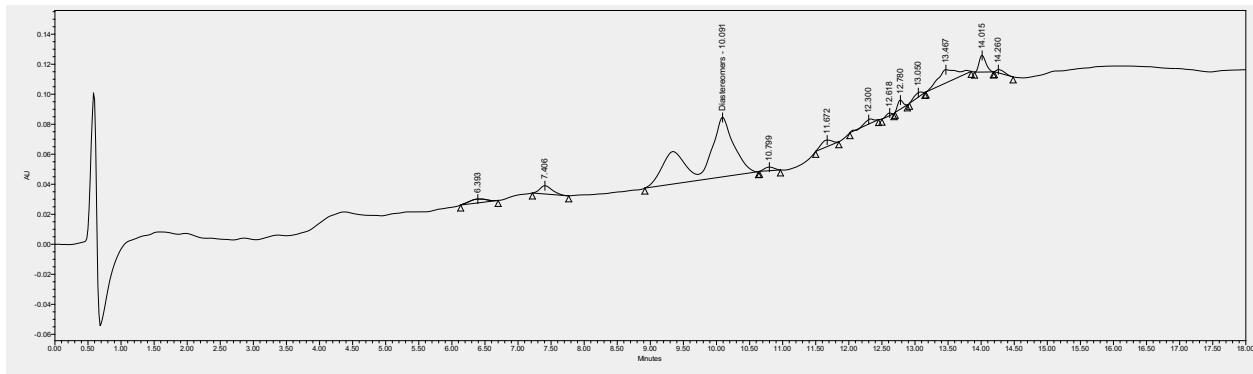
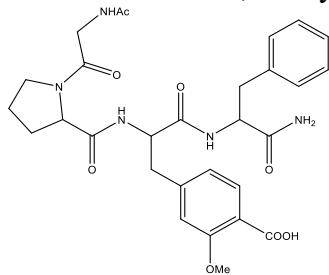


	Name	Retention Time	Area	% Area
1		10.795	104645	1.51
2		11.970	307900	4.44
3		12.386	458808	6.61
4		13.540	10775	0.16
5		13.826	43811	0.63
6	Diastereomers	14.299	6010272	86.65

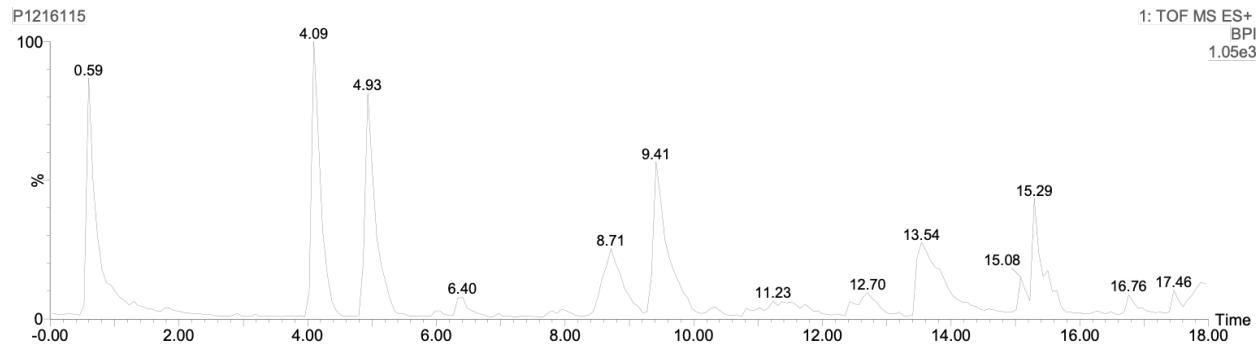


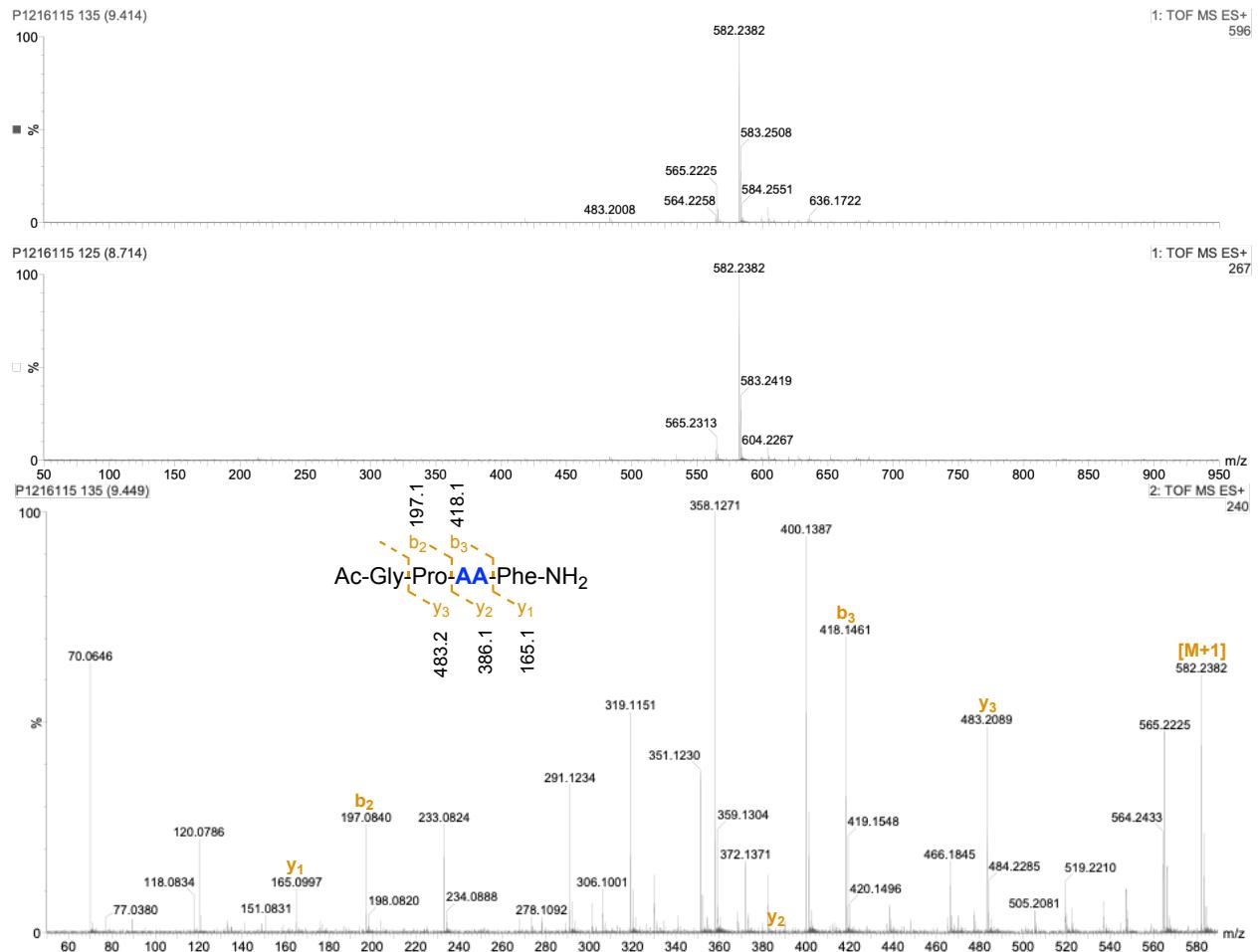


7G': MW = 581.6, Purity = 71.2%, Yield = 4.2% [0.056 mg]

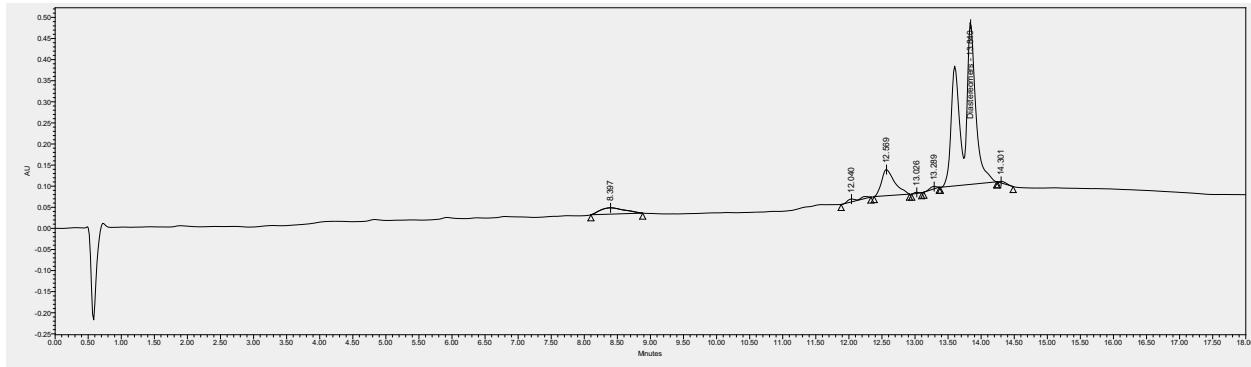
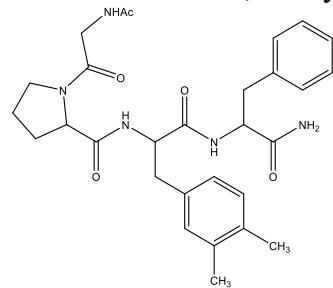


	Name	Retention Time	Area	% Area
1		6.393	46395	2.35
2		7.406	72858	3.68
3	Diastereomers	10.091	1408601	71.22
4		10.799	24918	1.26
5		11.672	50999	2.58
6		12.300	30770	1.56
7		12.618	9877	0.50
8		12.780	33803	1.71
9		13.050	25043	1.27
10		13.467	175686	8.88
11		14.015	79714	4.03
12		14.260	19027	0.96

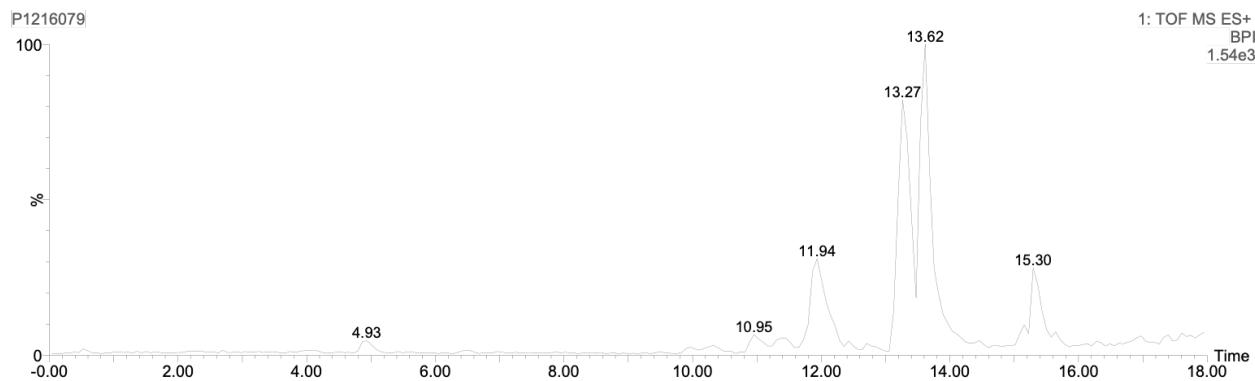


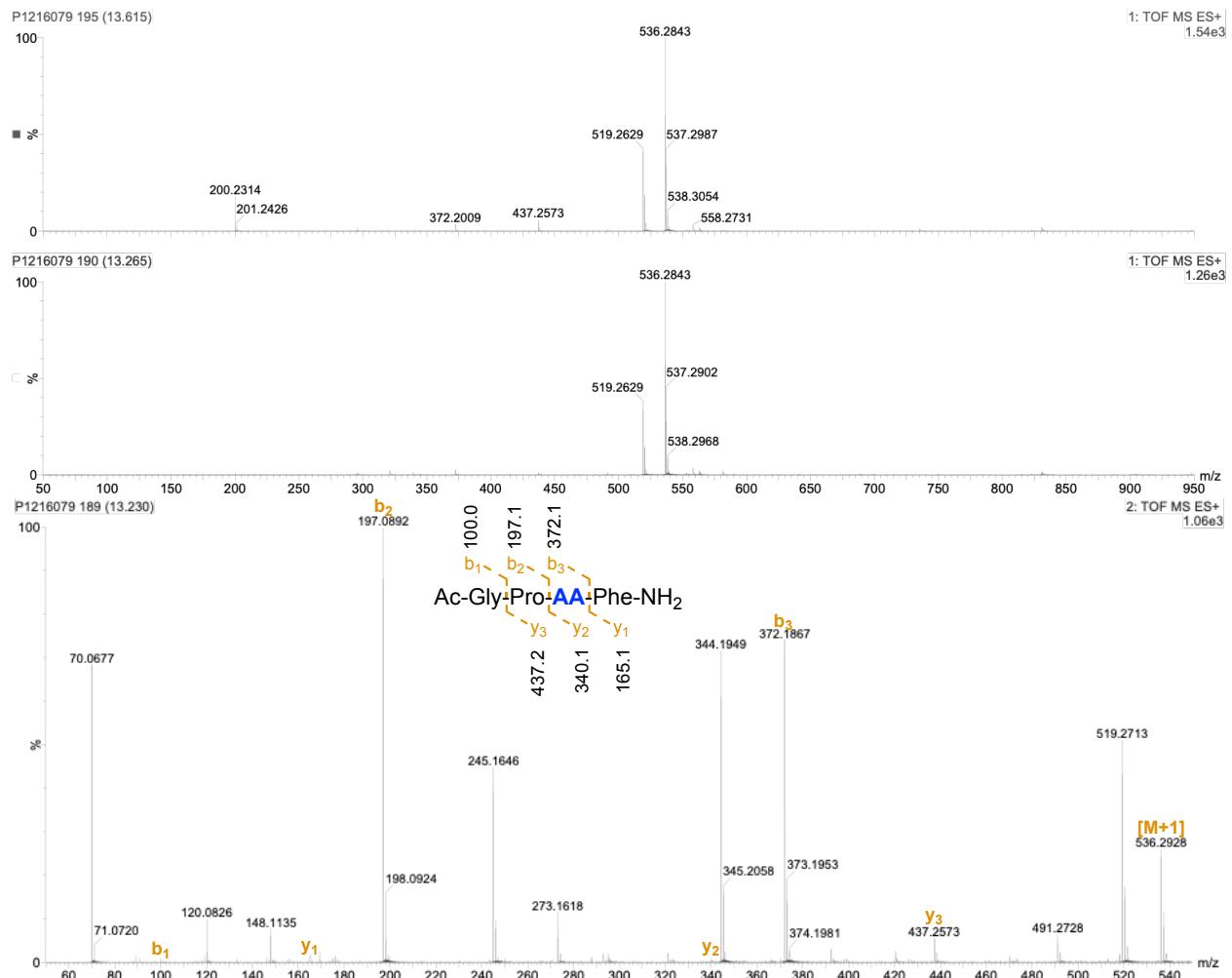


8G': MW = 535.6, Purity = 80.6%, Yield = 12.7% [0.16 mg]

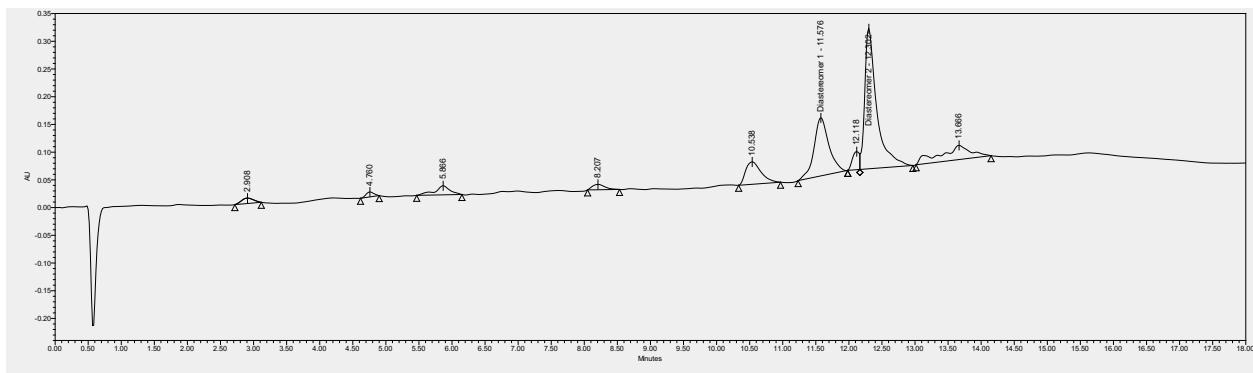
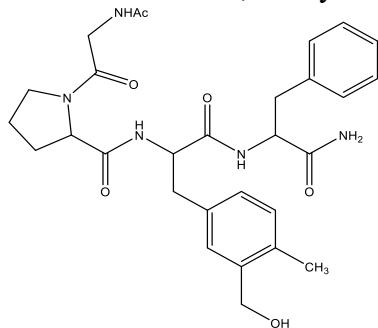


	Name	Retention Time	Area	% Area
1		8.397	389218	5.32
2		12.040	88768	1.21
3		12.569	853896	11.67
4		13.026	13814	0.19
5		13.289	42808	0.59
6	Diastereomers	13.840	5900062	80.63
7		14.301	29067	0.40

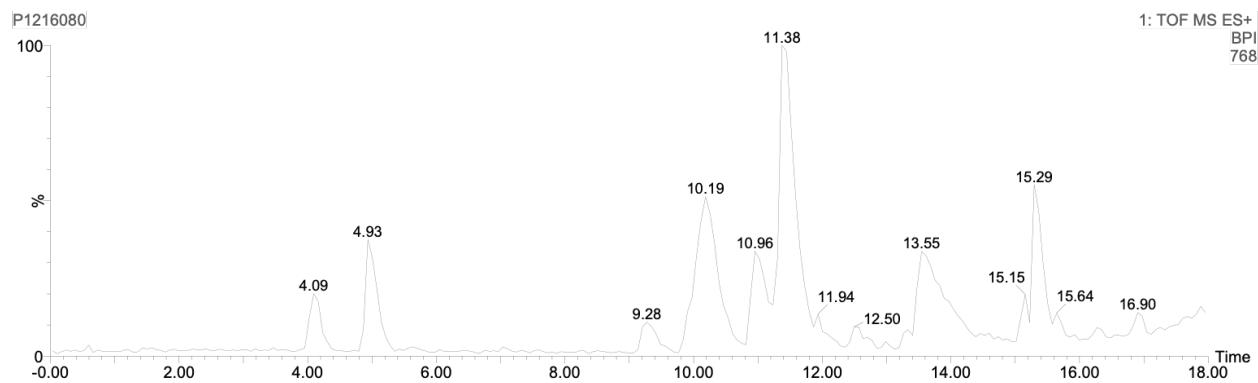


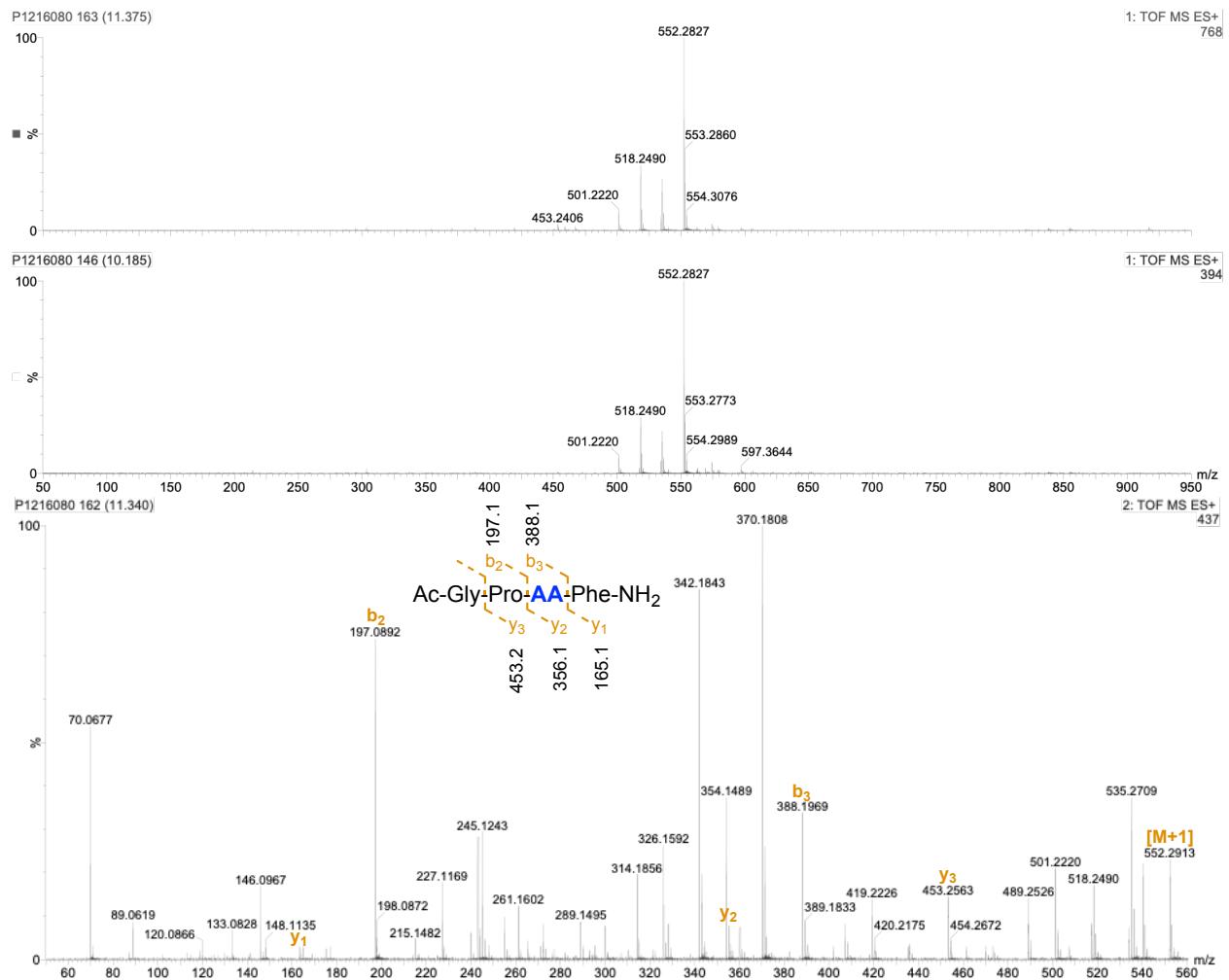


9G': MW = 551.6, Purity = 67.7%, Yield = 10.4% [0.13 mg]

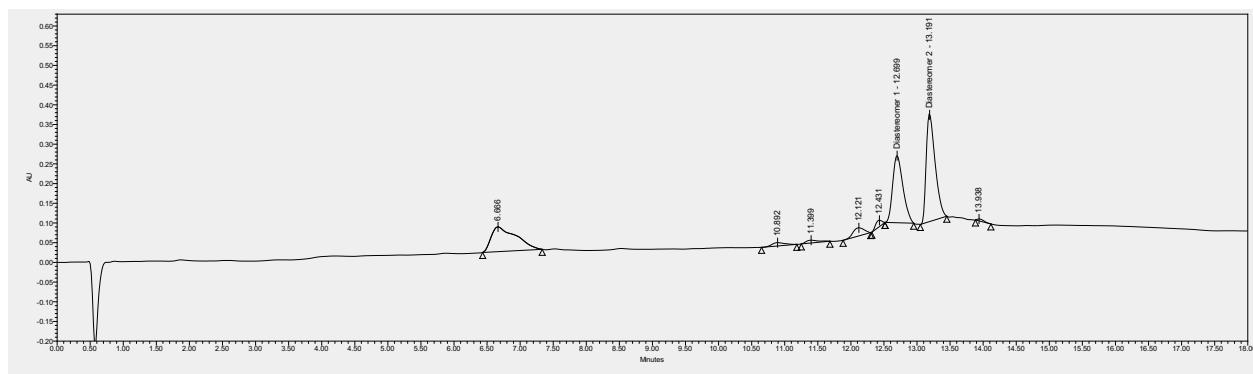
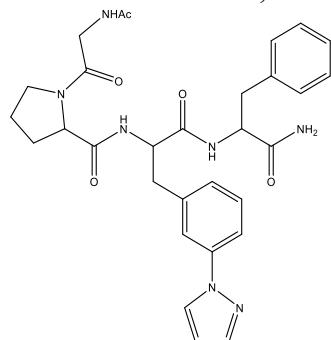


	Name	Retention Time	Area	% Area
1		2.908	123856	1.74
2		4.760	74142	1.04
3		5.866	261269	3.67
4		8.207	129627	1.82
5		10.538	654864	9.20
6	Diastereomer 1	11.576	1641749	23.06
7		12.118	234860	3.30
8	Diastereomer 2	12.302	3179205	44.66
9		13.666	818575	11.50

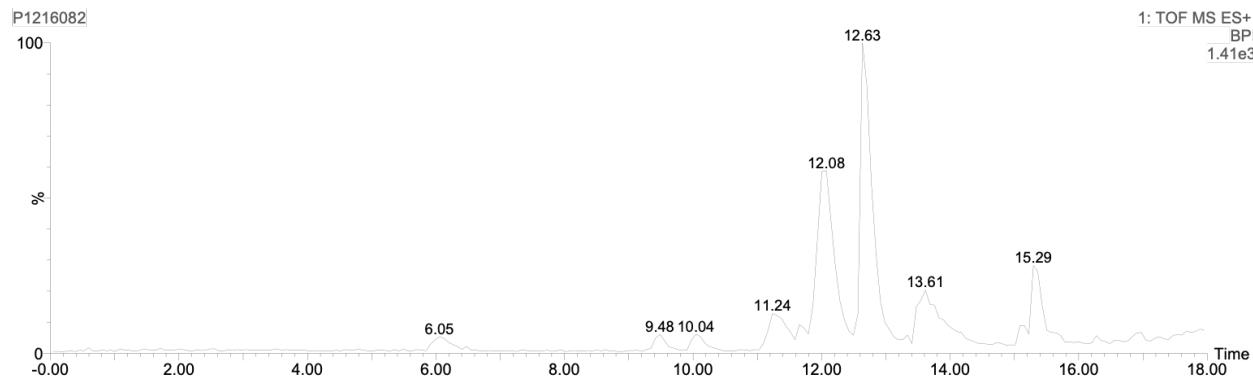


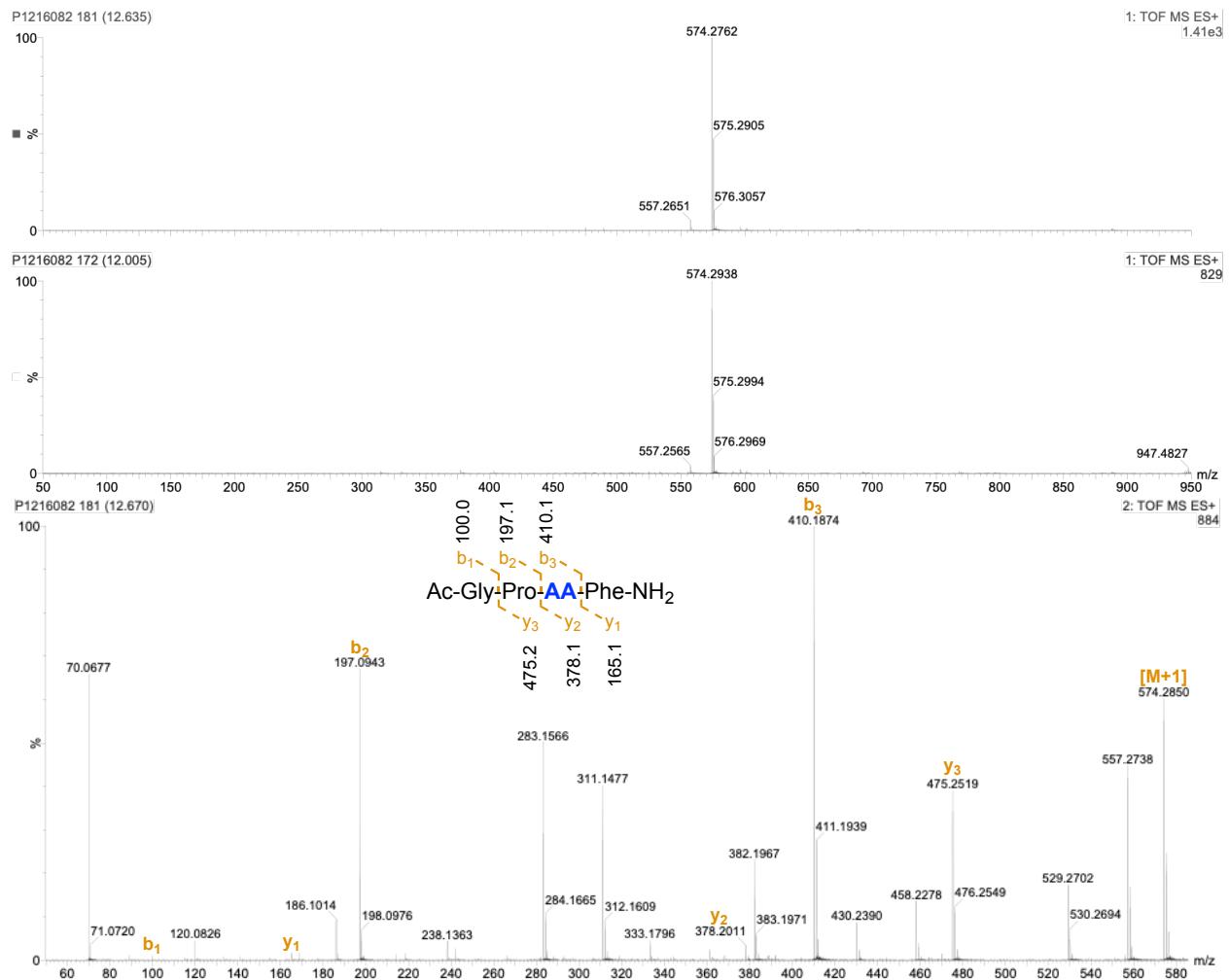


10G': MW = 572.7, Purity = 66.2%, Yield = 7.1% [0.095 mg]

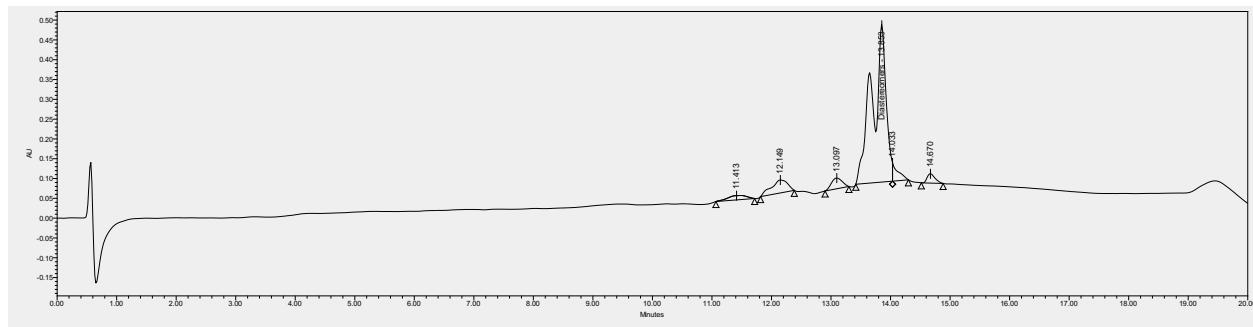
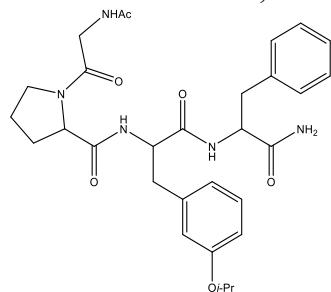


	Name	Retention Time	Area	% Area
1		6.666	1639589	24.72
2		10.892	117137	1.77
3		11.399	93340	1.41
4		12.121	243962	3.68
5		12.431	111953	1.69
6	Diastereomer 1	12.699	1834583	27.66
7	Diastereomer 2	13.191	2557181	38.55
8		13.938	35747	0.54

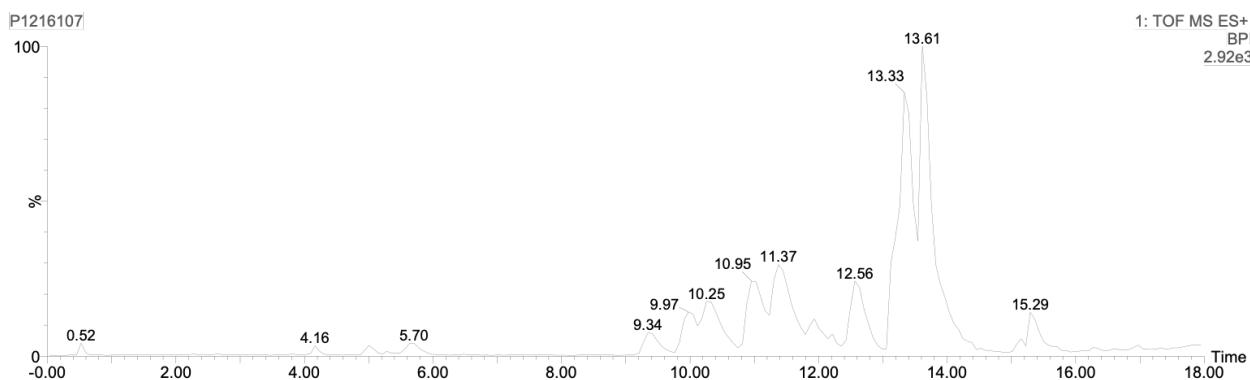


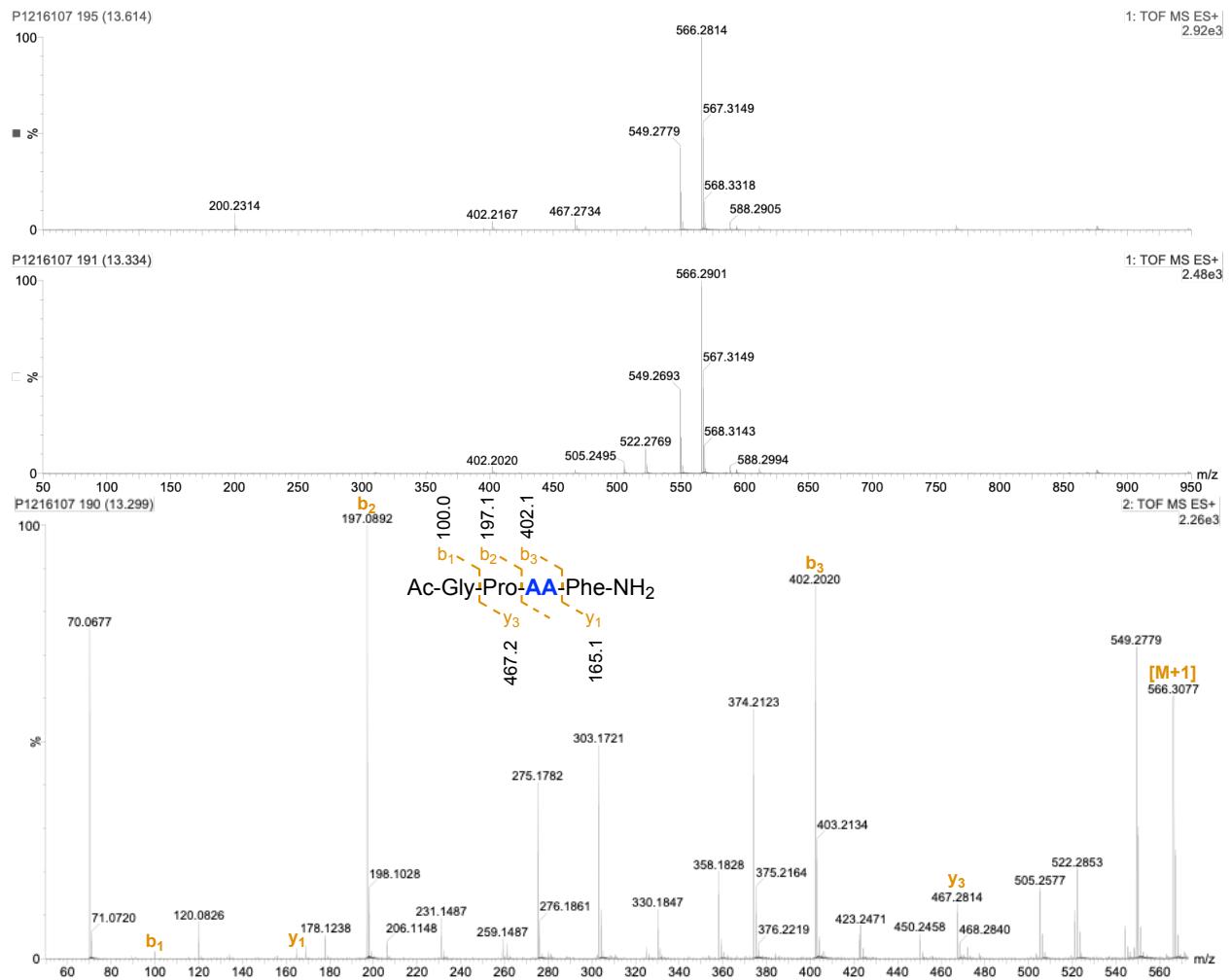


11G': MW = 565.7, Purity = 78.9%, Yield = 17.3% [0.23 mg]

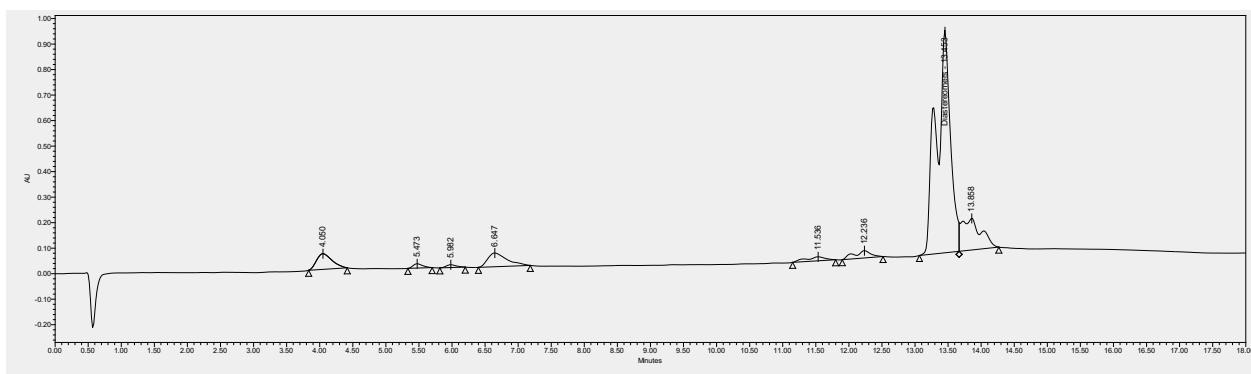
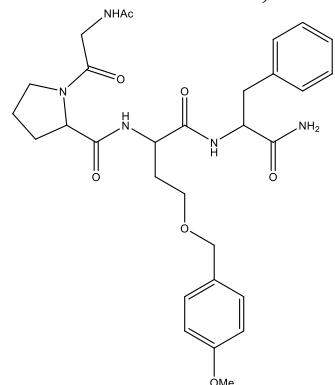


	Name	Retention Time	Area	% Area
1		11.413	239892	2.94
2		12.149	626876	7.68
3		13.097	342532	4.20
4	Diastereomers	13.853	6441616	78.92
5		14.033	296190	3.63
6		14.670	215468	2.64

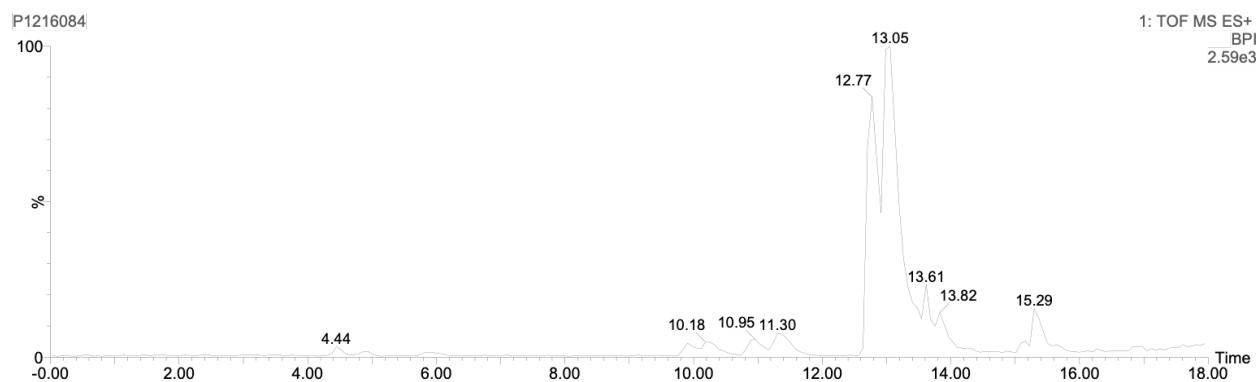


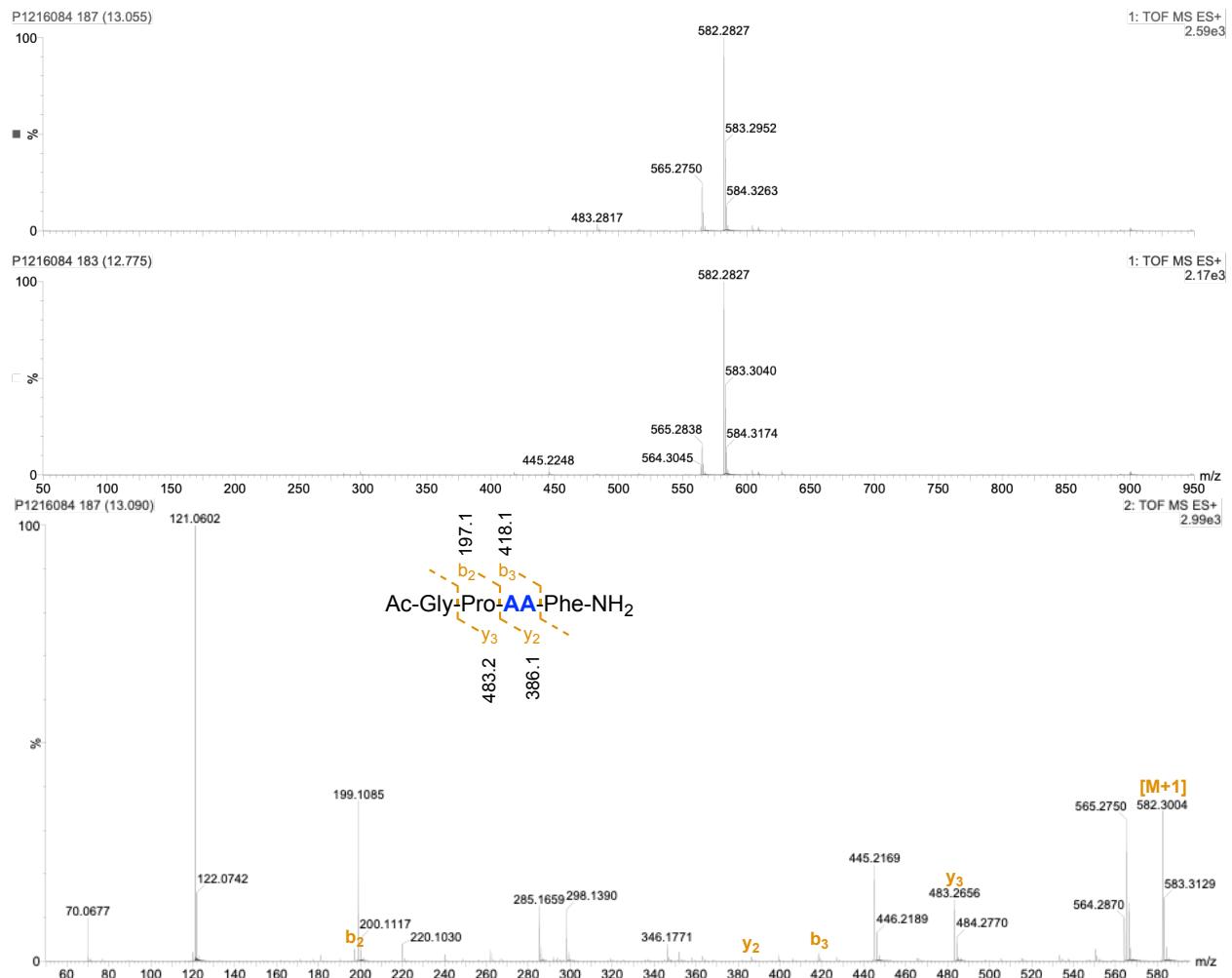


12G': MW = 581.7, Purity = 69.2%, Yield = 27.7% [0.38 mg]

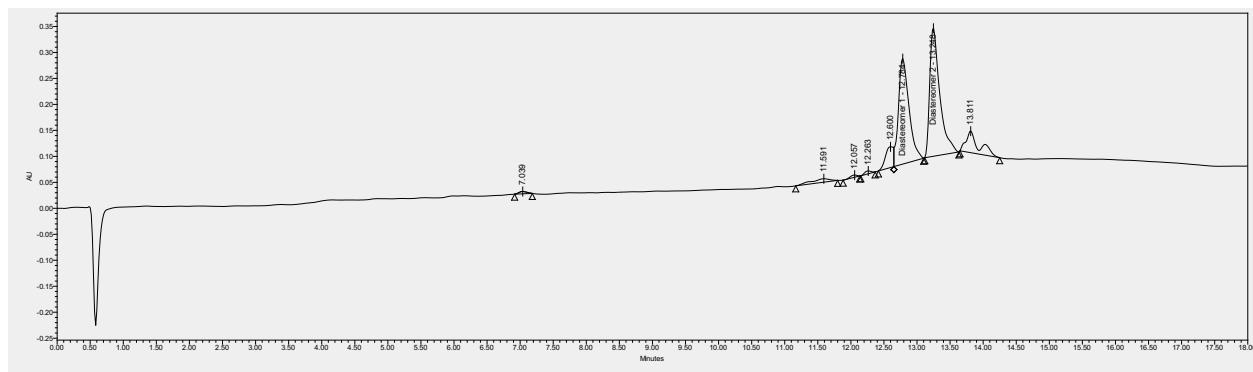
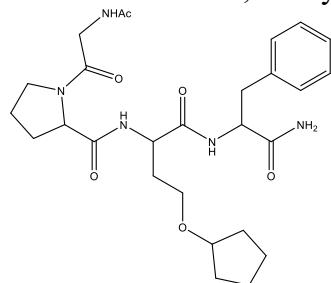


	Name	Retention Time	Area	% Area
1		4.050	1000590	5.36
2		5.473	171377	0.92
3		5.982	117155	0.63
4		6.647	1089826	5.83
5		11.536	324520	1.74
6		12.236	506107	2.71
7	Diastereomers	13.453	12923853	69.18
8		13.858	2548276	13.64

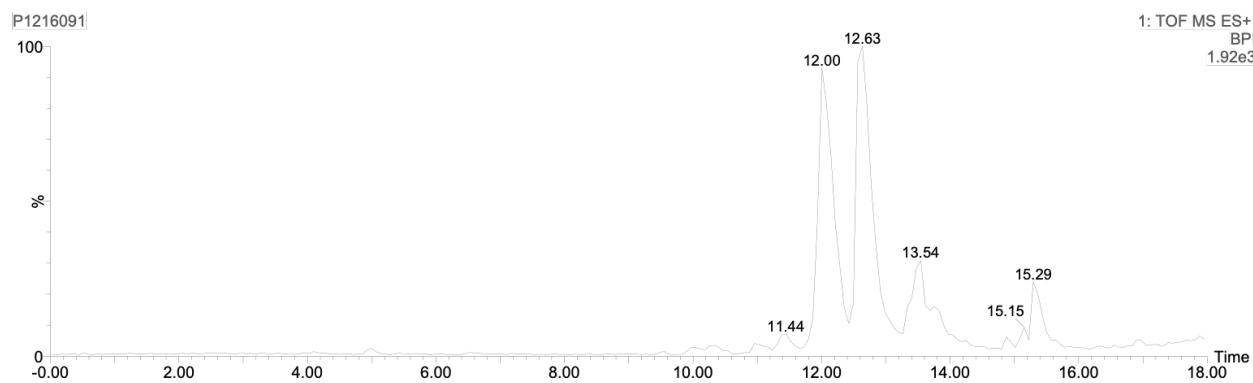


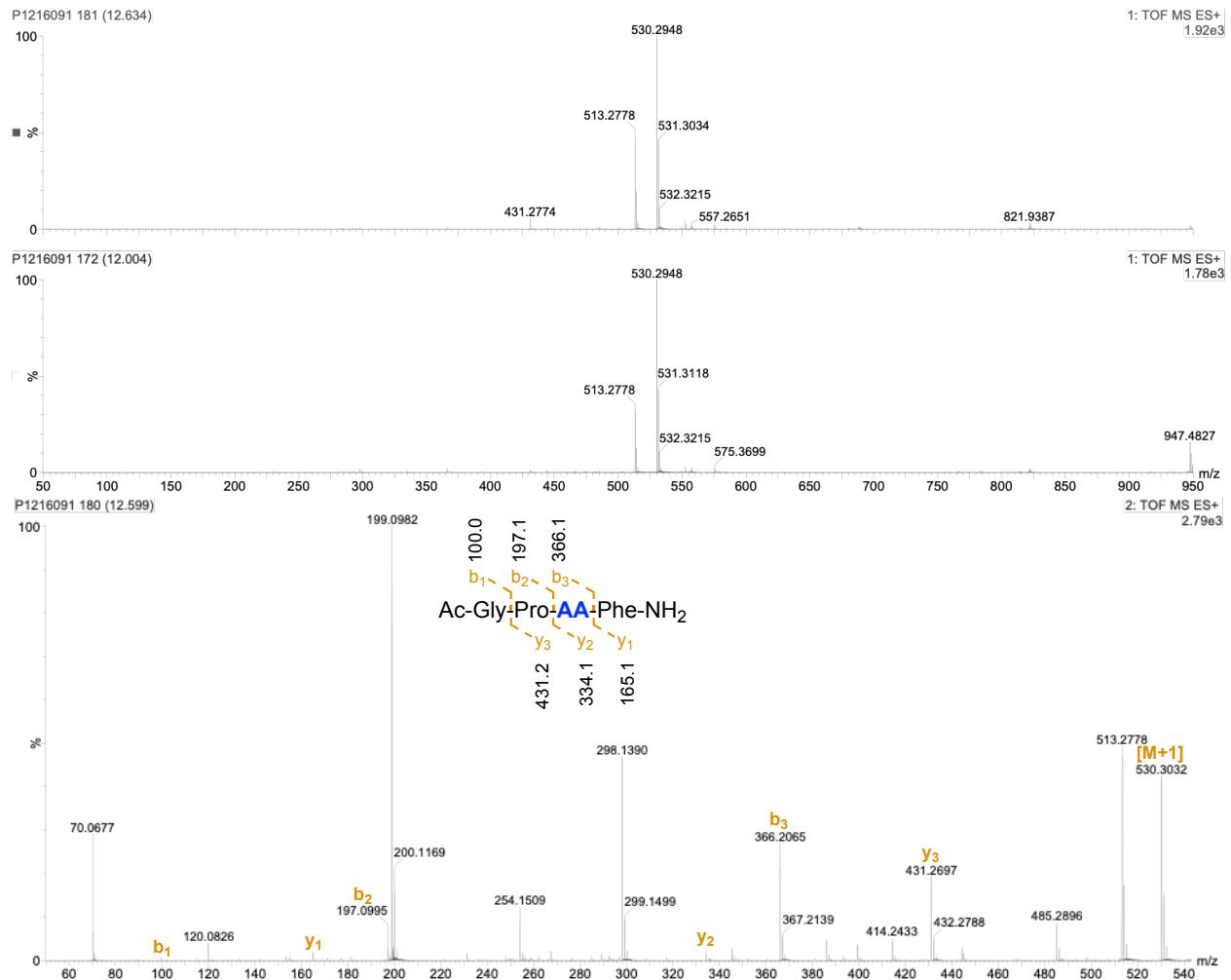


1H': MW = 529.6, Purity = 81.1%, Yield = 15.6% [0.19 mg]

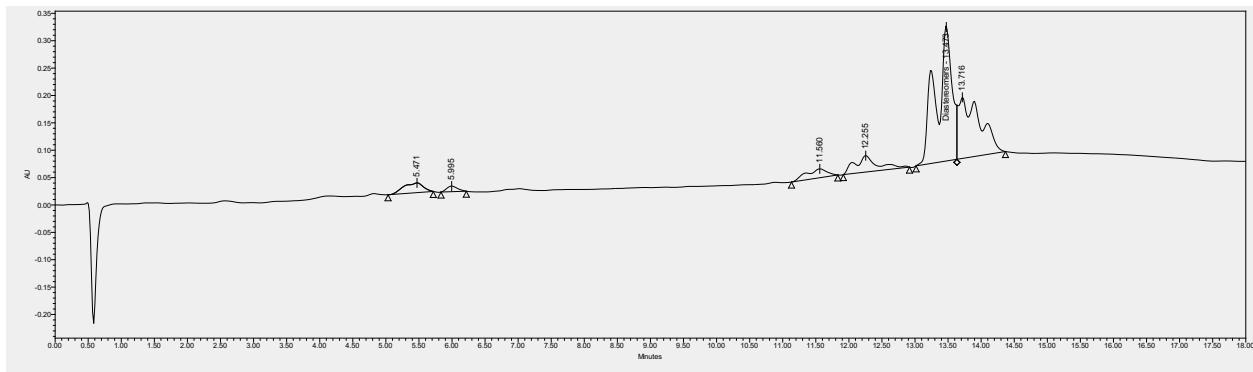
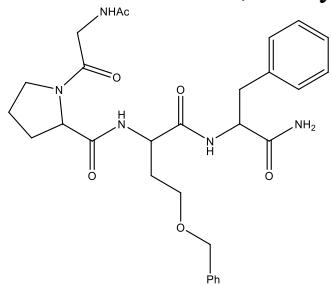


	Name	Retention Time	Area	% Area
1		7.039	37683	0.63
2		11.591	138896	2.31
3		12.057	31112	0.52
4		12.263	38015	0.63
5		12.600	332277	5.53
6	Diastereomer 1	12.784	2261479	37.66
7	Diastereomer 2	13.248	2609071	43.45
8		13.811	556291	9.26

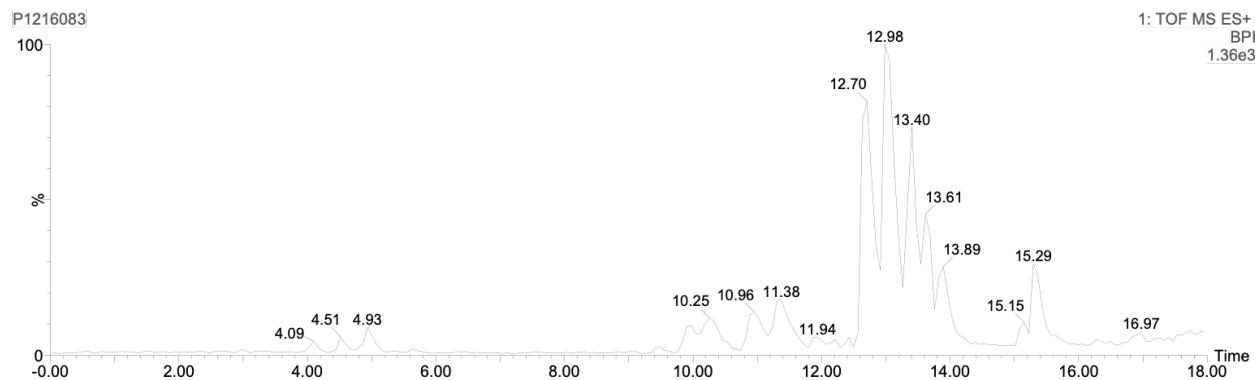


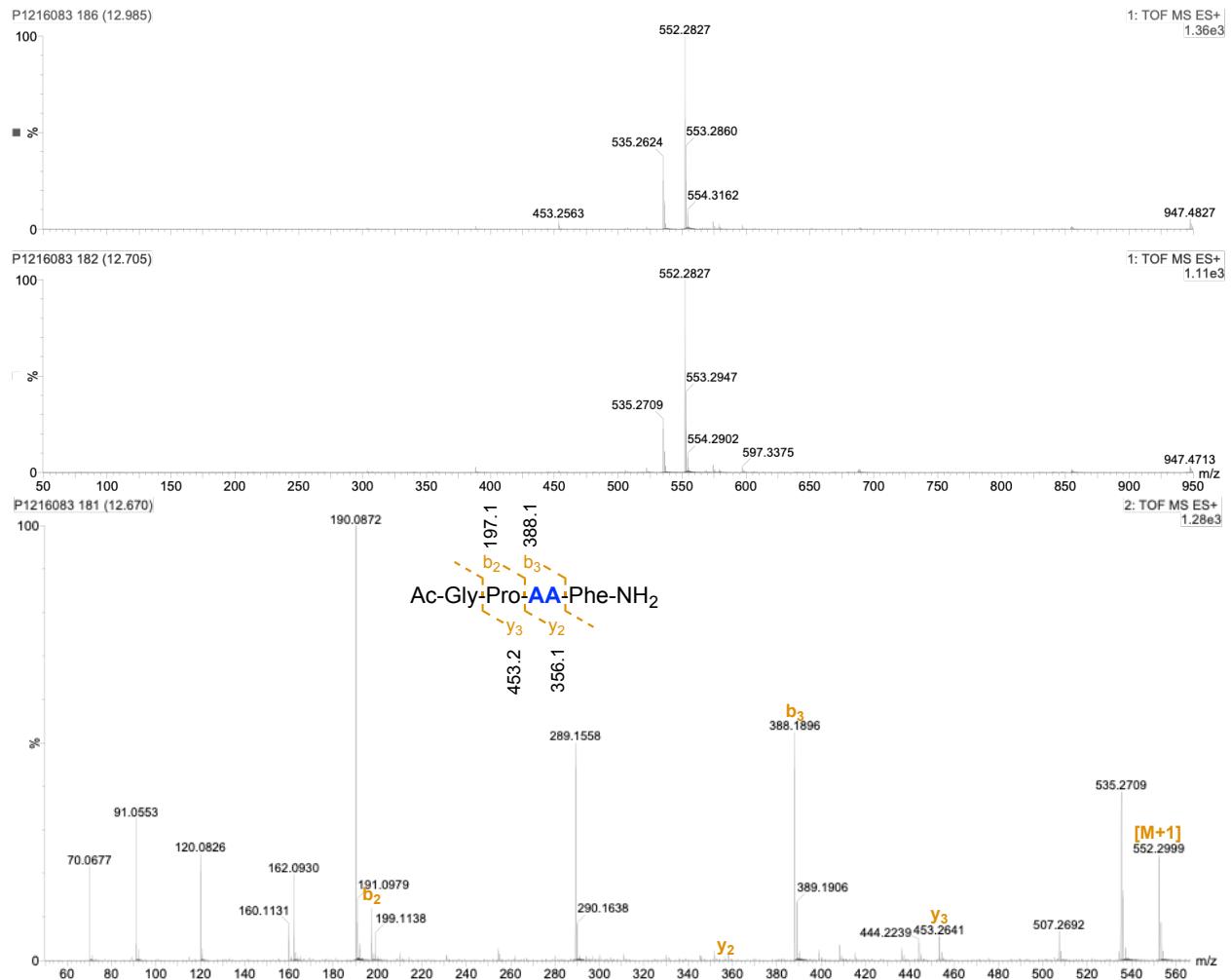


2H': MW = 551.6, Purity = 49.7%, Yield = 8.7% [0.11 mg]

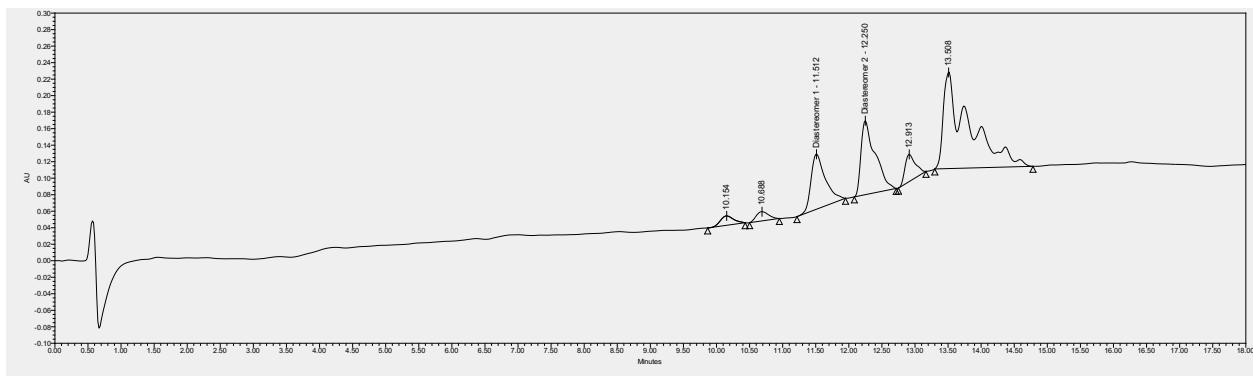
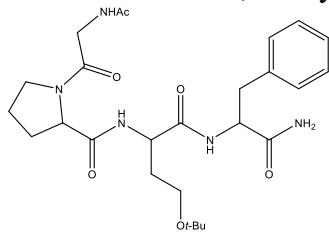


	Name	Retention Time	Area	% Area
1		5.471	375224	4.60
2		5.995	117249	1.44
3		11.560	361290	4.43
4		12.255	692913	8.49
5	Diastereomers	13.473	4056649	49.71
6		13.716	2557082	31.34

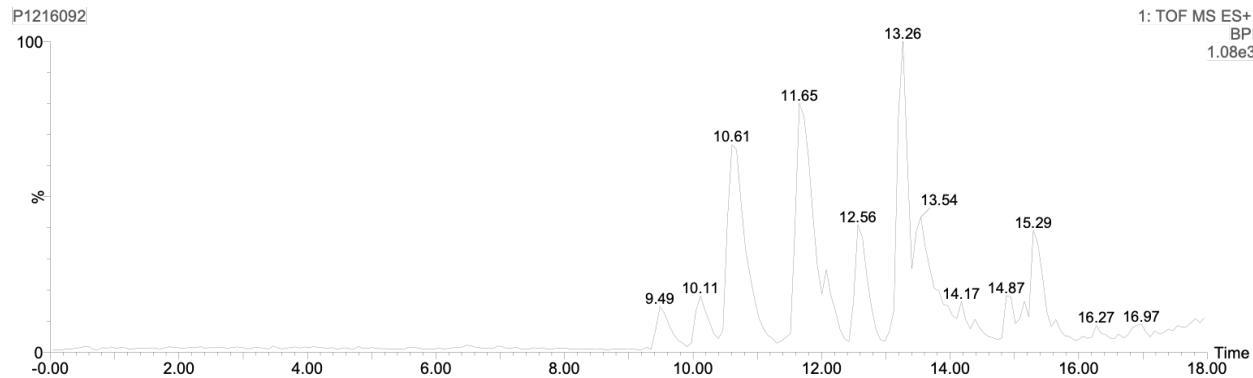


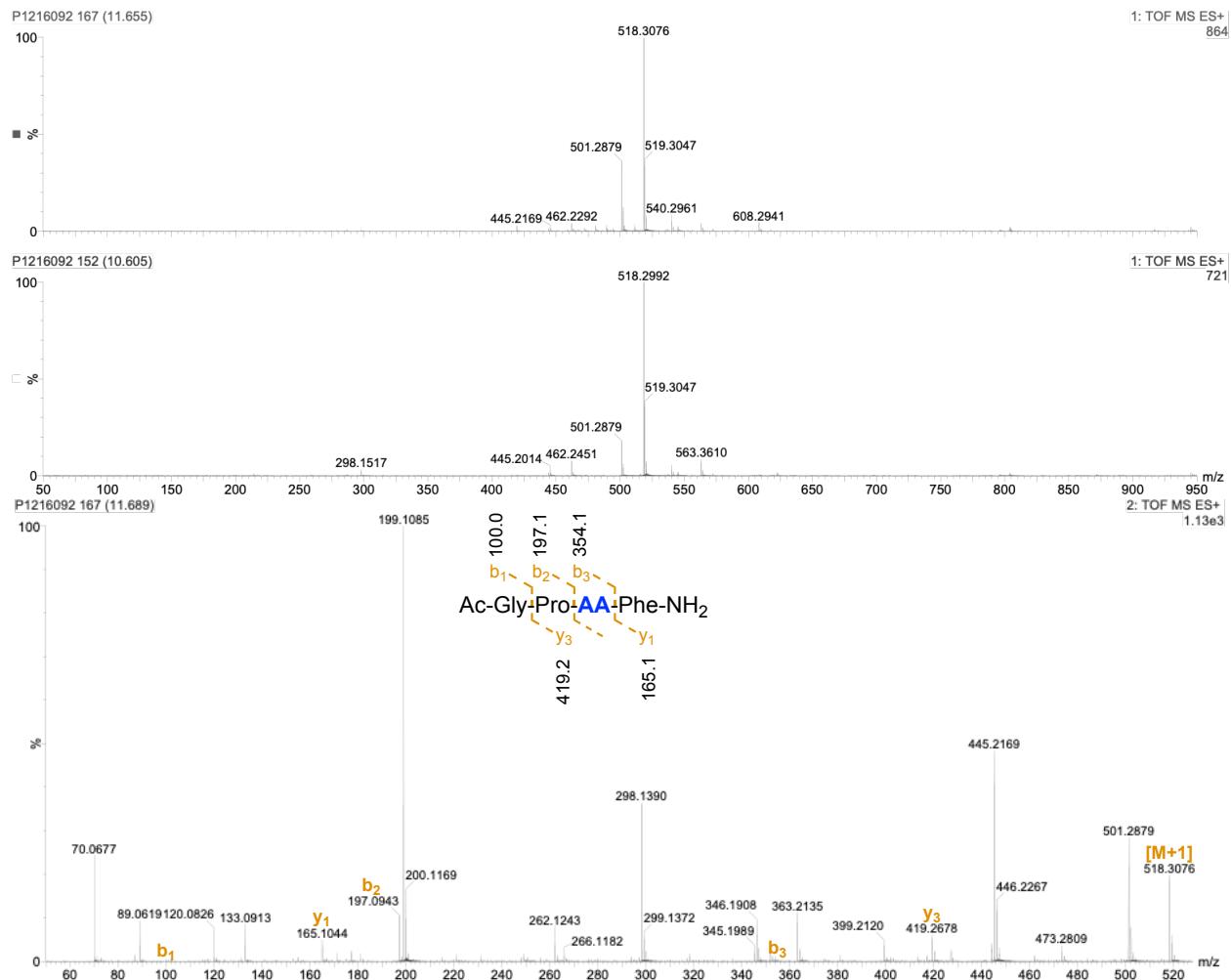


3H': MW = 517.6, Purity = 38.1%, Yield = 10.3% [0.12 mg]

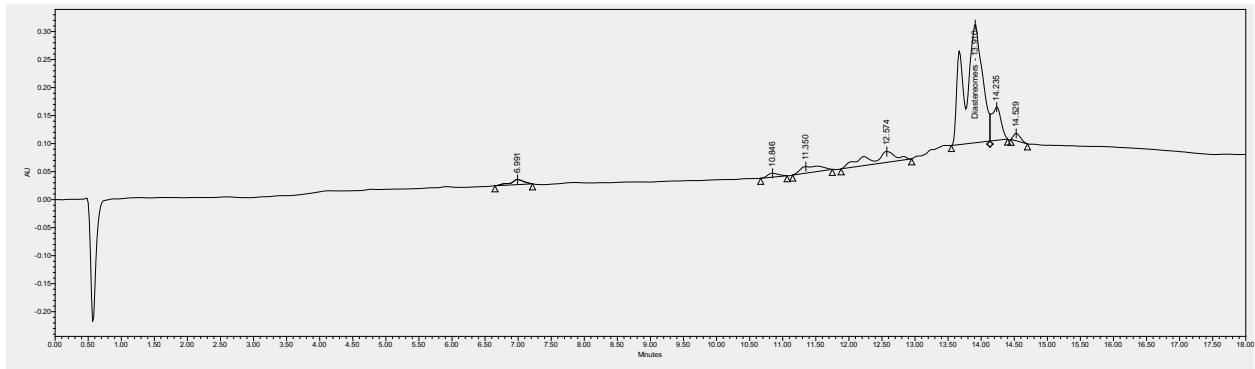
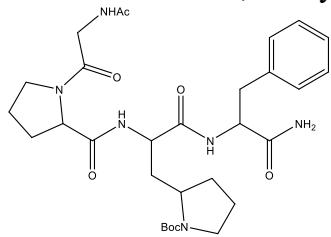


	Name	Retention Time	Area	% Area
1		10.154	160113	2.60
2		10.688	147322	2.39
3	Diastereomer 1	11.512	1010961	16.43
4	Diastereomer 2	12.250	1330428	21.63
5		12.913	360610	5.86
6		13.508	3142800	51.08

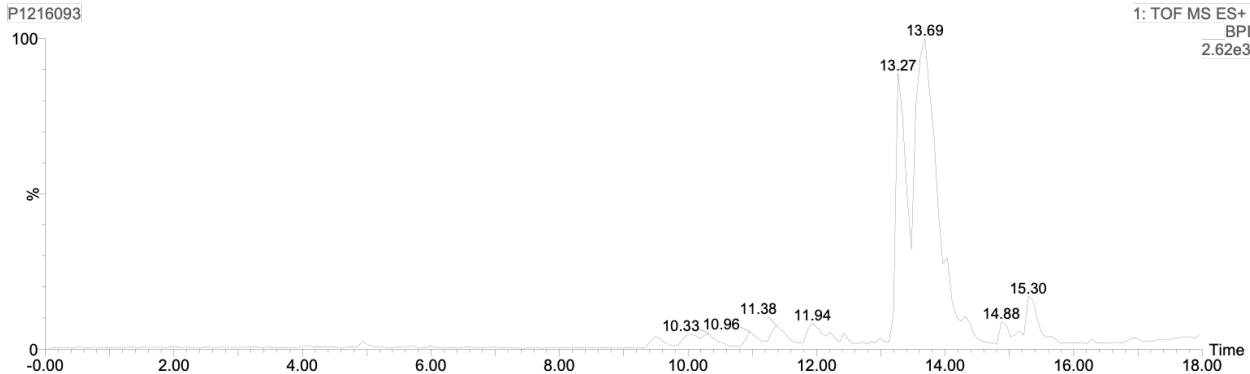


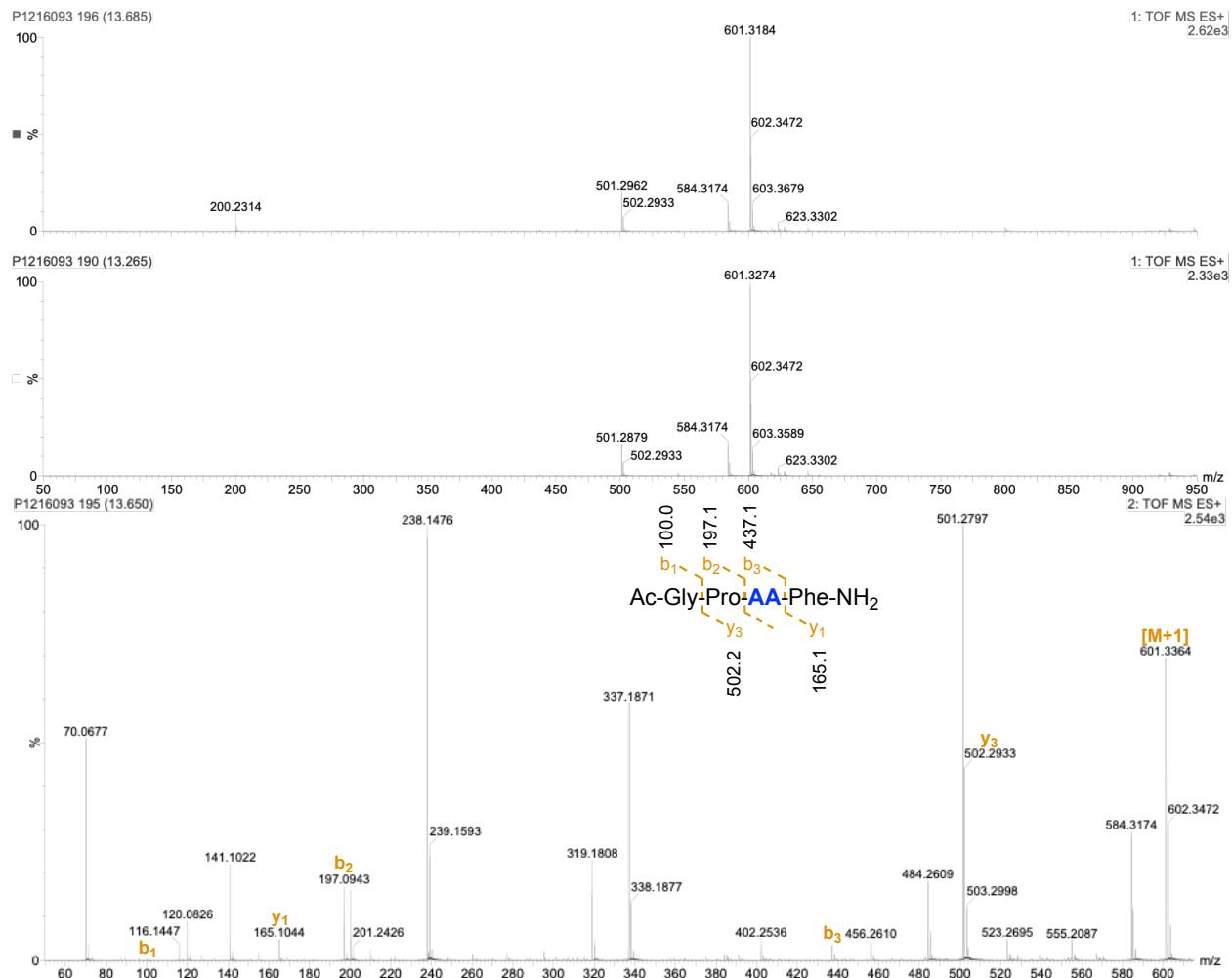


4H': MW = 600.7, Purity = 69.7%, Yield = 12.7% [0.18 mg]

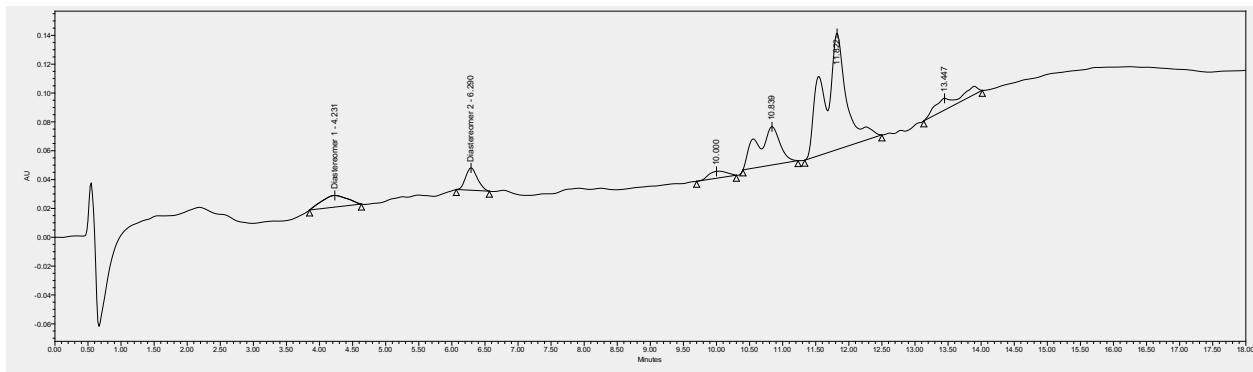
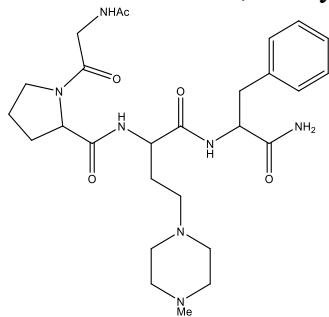


	Name	Retention Time	Area	% Area
1		6.991	127744	2.24
2		10.846	80606	1.41
3		11.350	248652	4.36
4		12.574	585778	10.27
5	Diastereomers	13.910	3972502	69.65
6		14.235	590580	10.36
7		14.529	97281	1.71

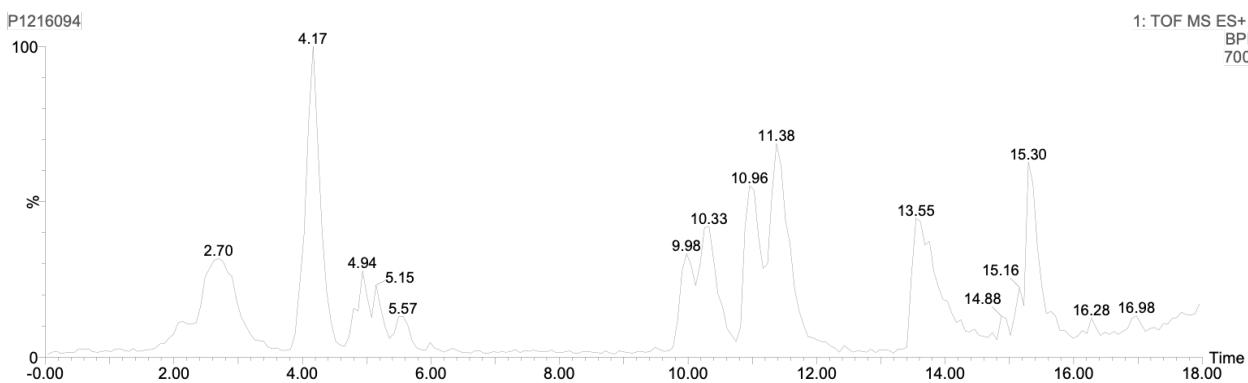


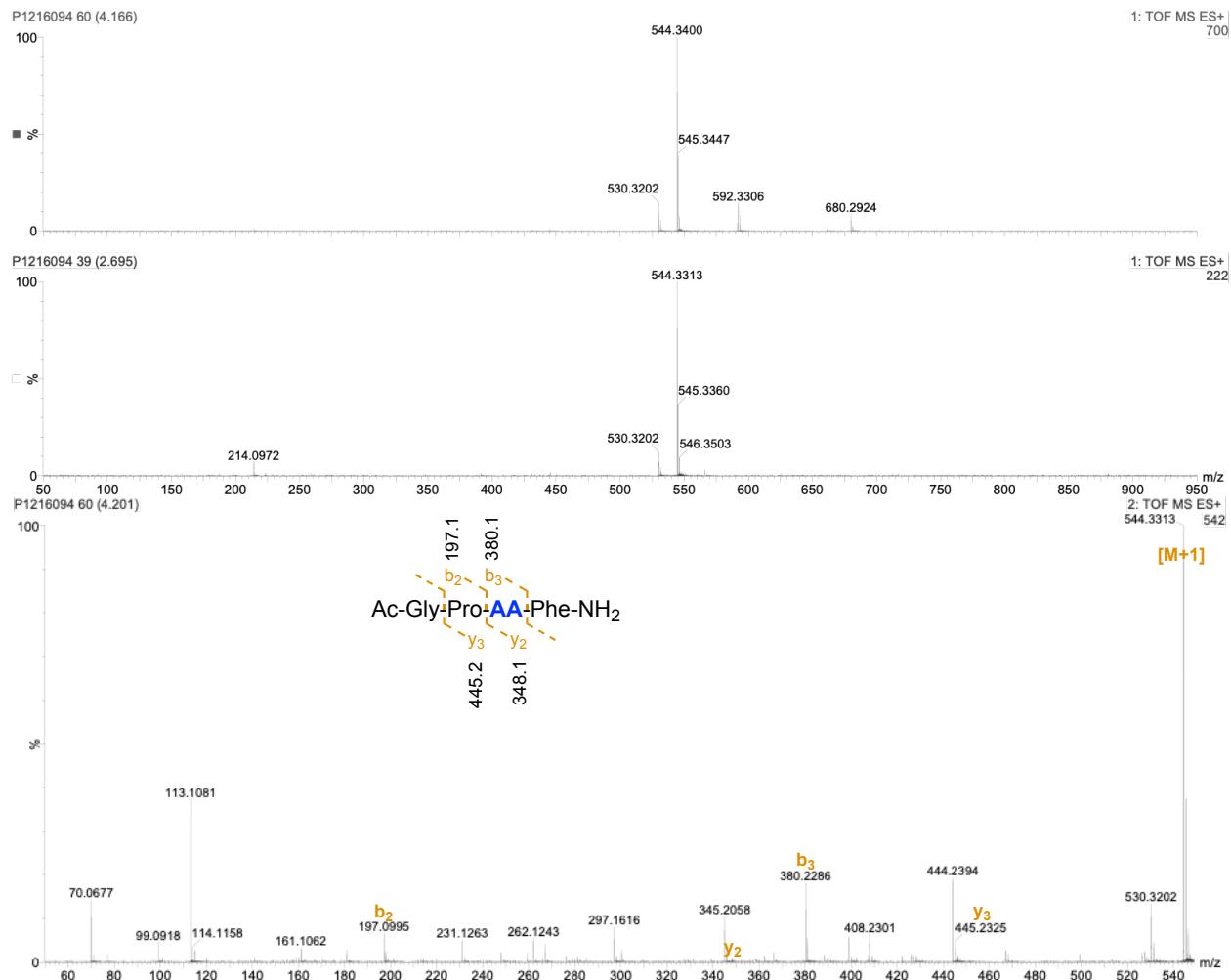


5H': MW = 543.7, Purity = 12.4%, Yield = 2.3% [0.029 mg]

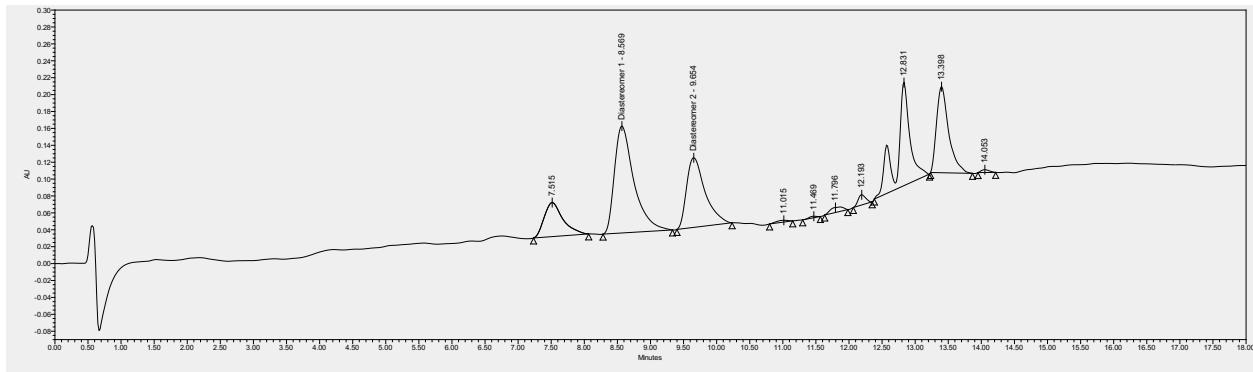
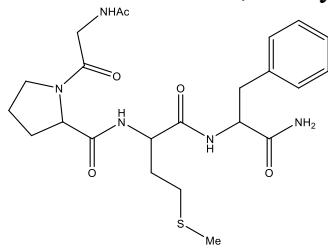


	Name	Retention Time	Area	% Area
1	Diastereomer 1	4.231	218721	6.61
2	Diastereomer 2	6.290	192620	5.82
3		10.000	90059	2.72
4		10.839	644331	19.48
5		11.822	1923578	58.14
6		13.447	239013	7.22

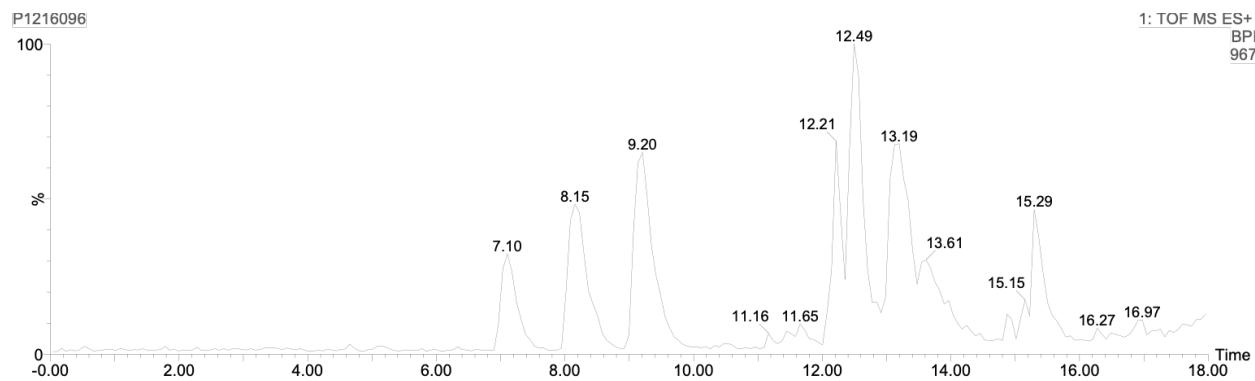


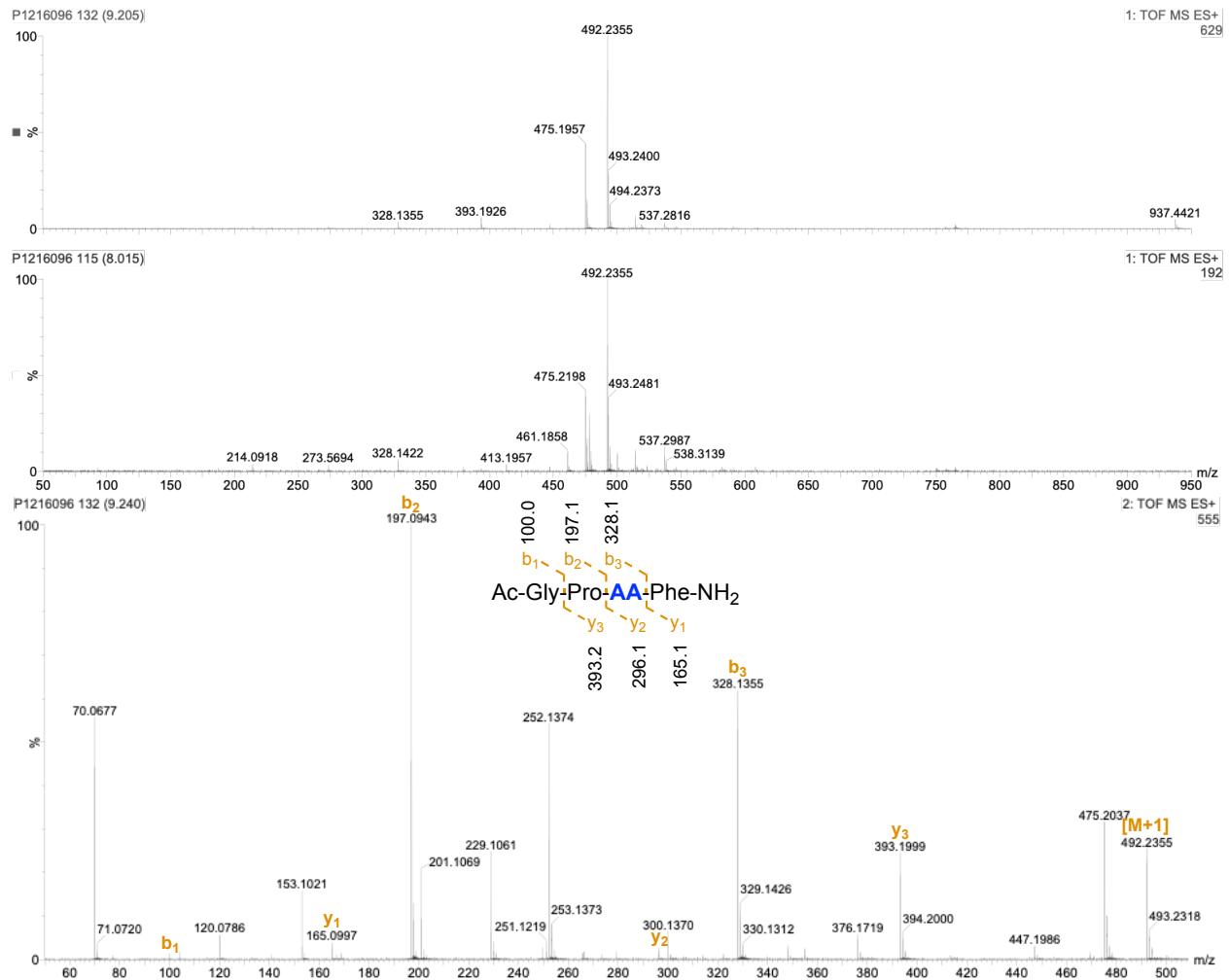


6H': MW = 491.6, Purity = 51.1%, Yield = 16.8% [0.19 mg]

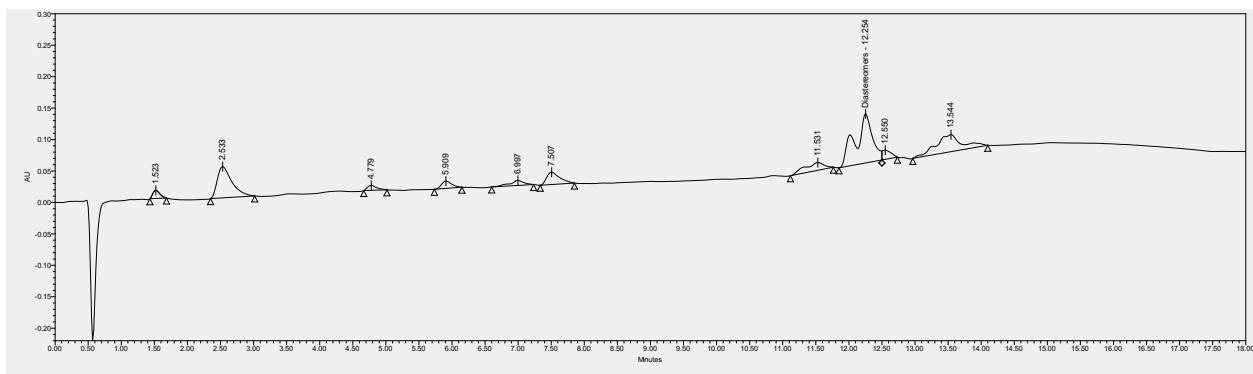
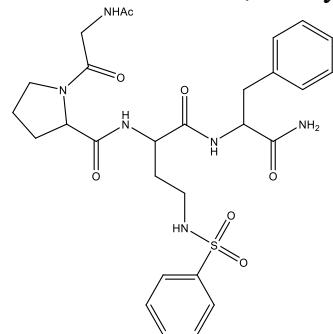


	Name	Retention Time	Area	% Area
1		7.515	796380	9.71
2	Diastereomer 1	8.569	2587021	31.56
3	Diastereomer 2	9.654	1604621	19.57
4		11.015	27217	0.33
5		11.469	14936	0.18
6		11.796	79494	0.97
7		12.193	95854	1.17
8		12.831	1663683	20.29
9		13.398	1300215	15.86
1		14.053	28795	0.35
0				

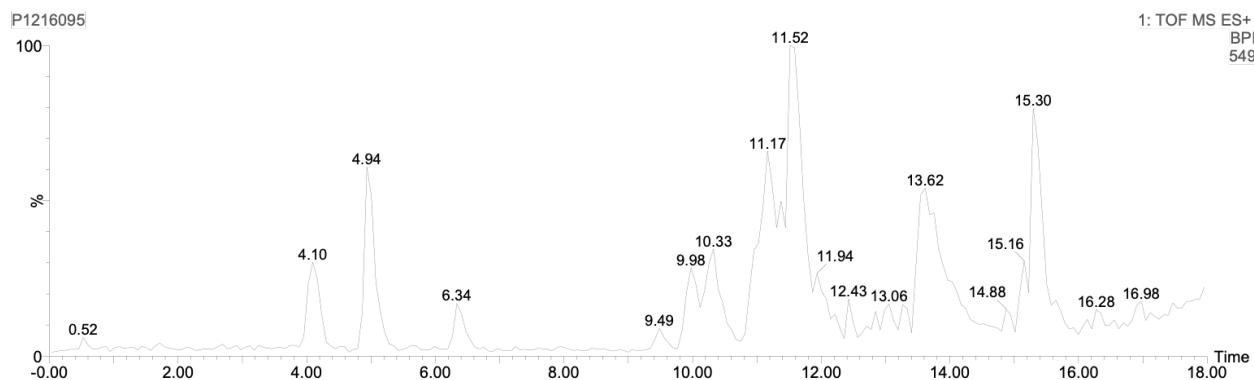


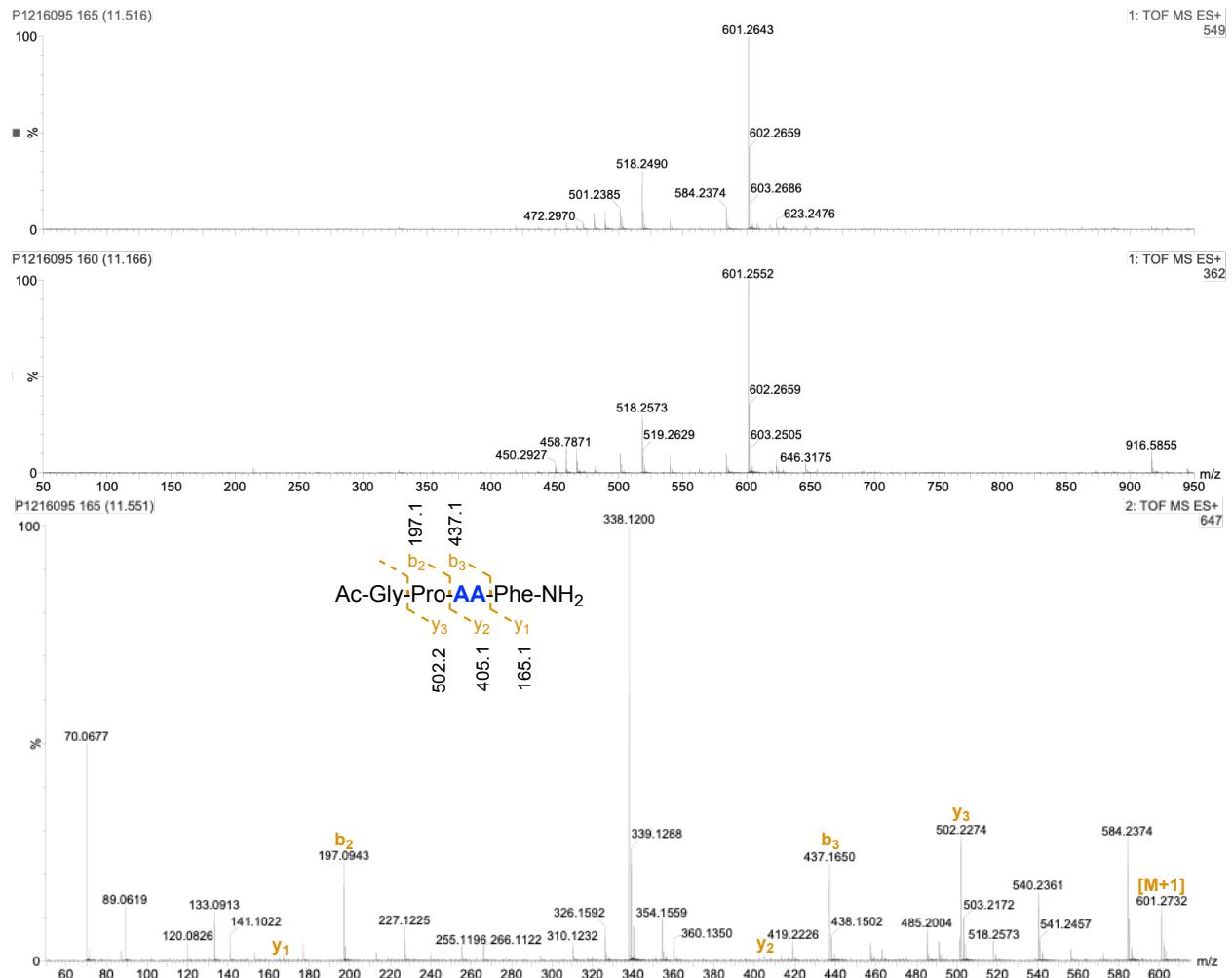


7H': MW = 600.7, Purity = 35.2%, Yield = 3.0% [0.042 mg]

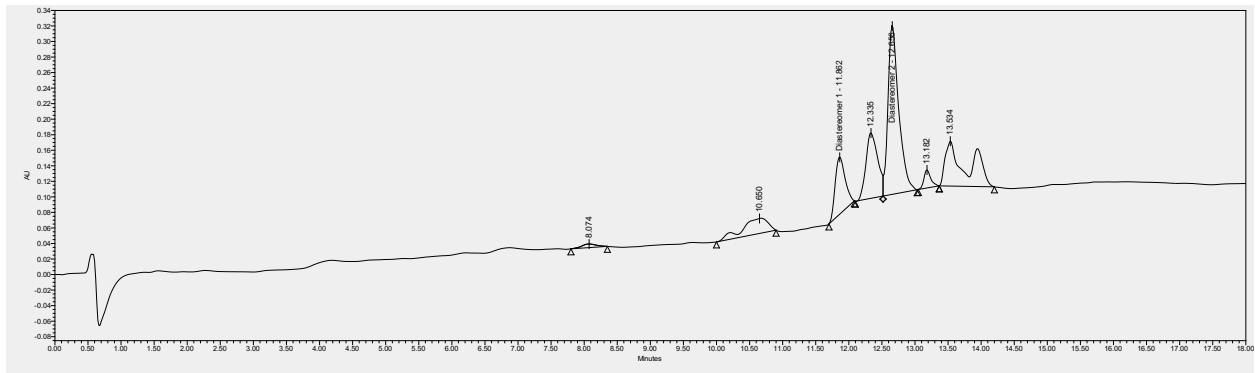
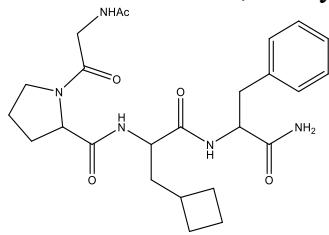


	Name	Retention Time	Area	% Area
1		1.523	95469	2.40
2		2.533	763594	19.18
3		4.779	76113	1.91
4		5.909	124079	3.12
5		6.997	116795	2.93
6		7.507	268668	6.75
7		11.531	262732	6.60
8	Diastereomers	12.254	1400654	35.19
9		12.550	130642	3.28
10		13.544	742011	18.64

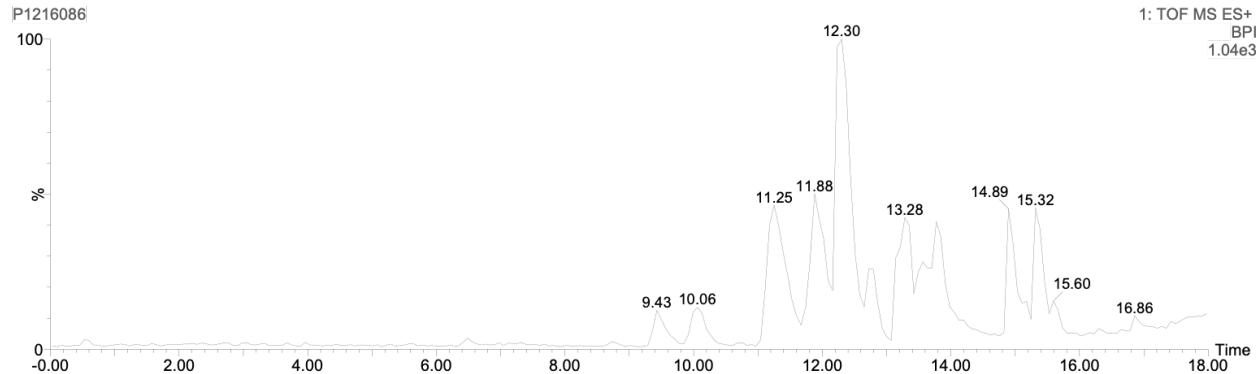


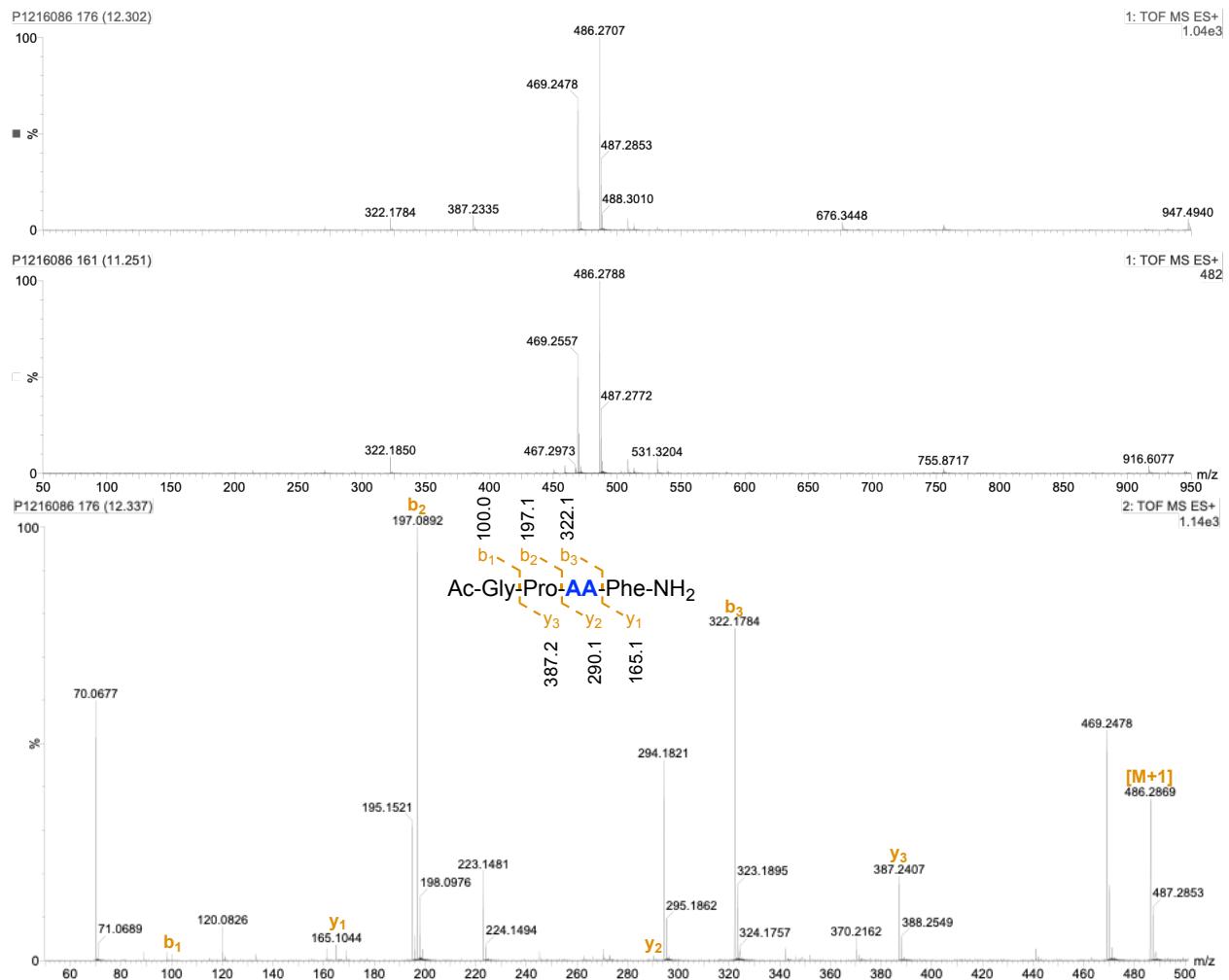


8H': MW = 485.6, Purity = 50.9%, Yield = 14.2% [0.16 mg]

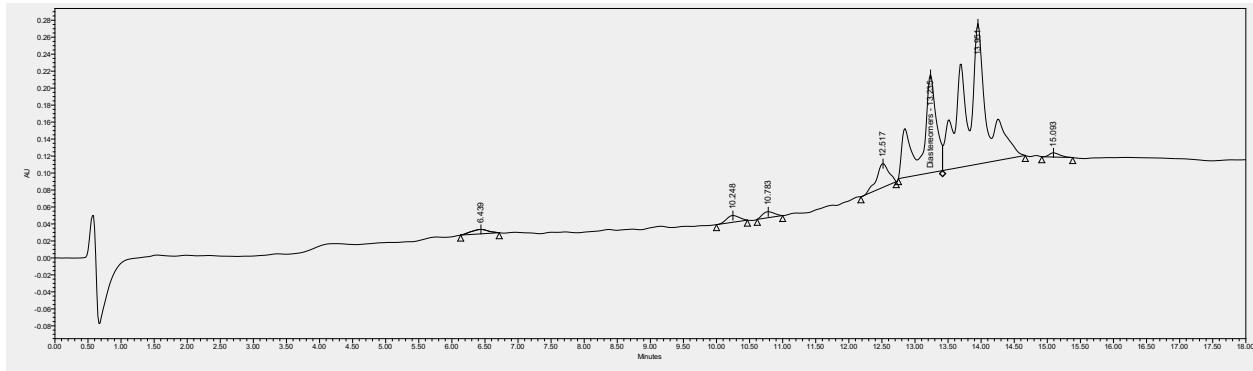
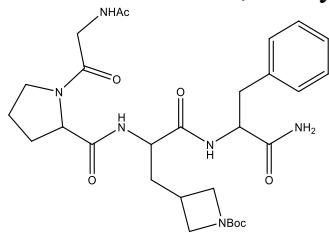


	Name	Retention Time	Area	% Area
1		8.074	72196	1.14
2		10.650	542601	8.53
3	Diastereomer 1	11.862	771342	12.13
4		12.335	1045789	16.45
5	Diastereomer 2	12.656	2462397	38.73
6		13.182	166563	2.62
7		13.534	1297561	20.41

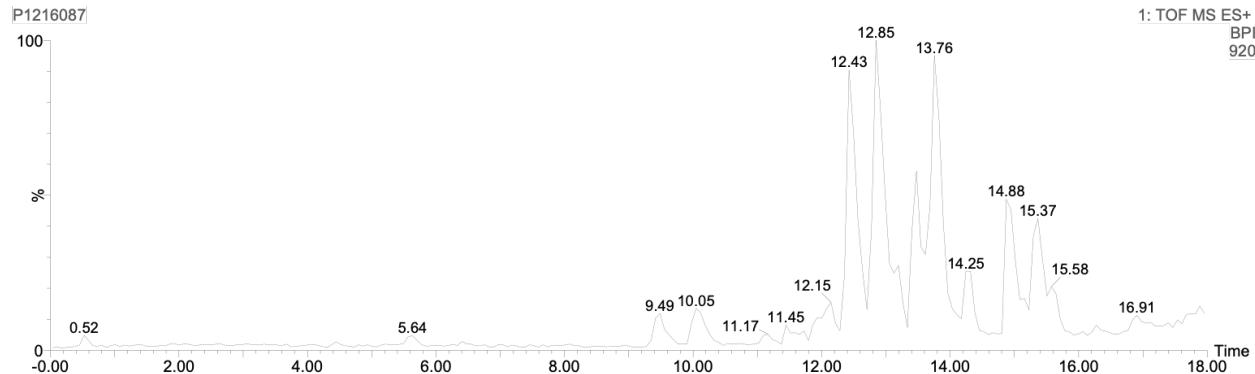


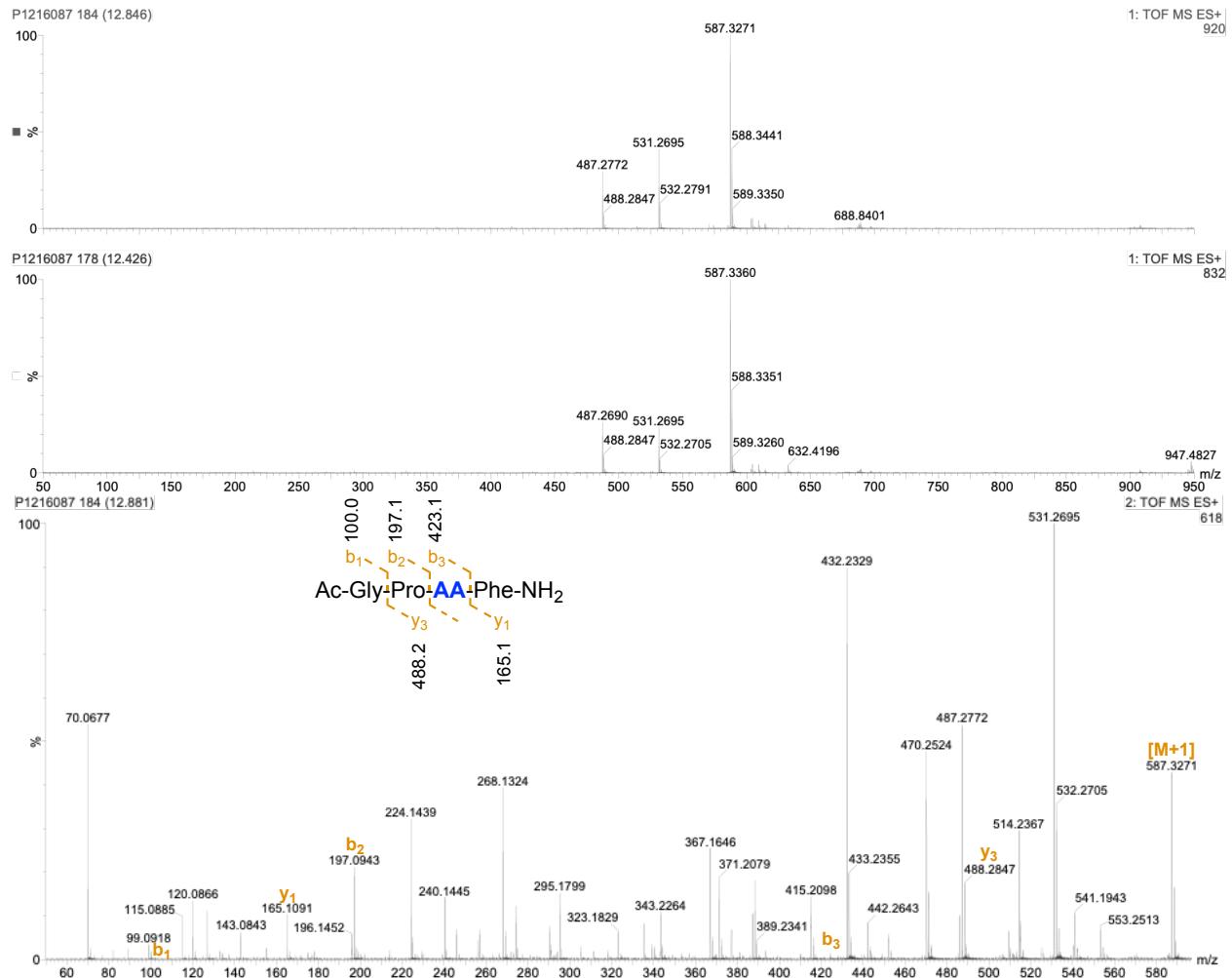


9H': MW = 586.7, Purity = 28.5%, Yield = 8.2% [0.11 mg]

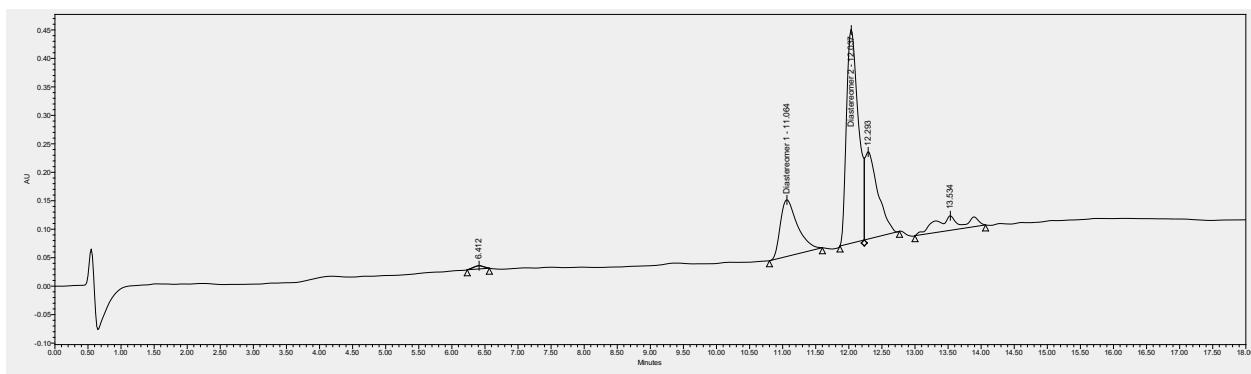
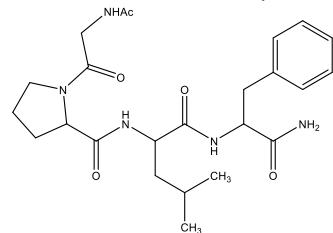


	Name	Retention Time	Area	% Area
1		6.439	95558	1.46
2		10.248	105831	1.62
3		10.783	87893	1.35
4		12.517	365325	5.59
5	Diastereomers	13.235	1863748	28.54
6		13.951	3955380	60.57
7		15.093	56305	0.86

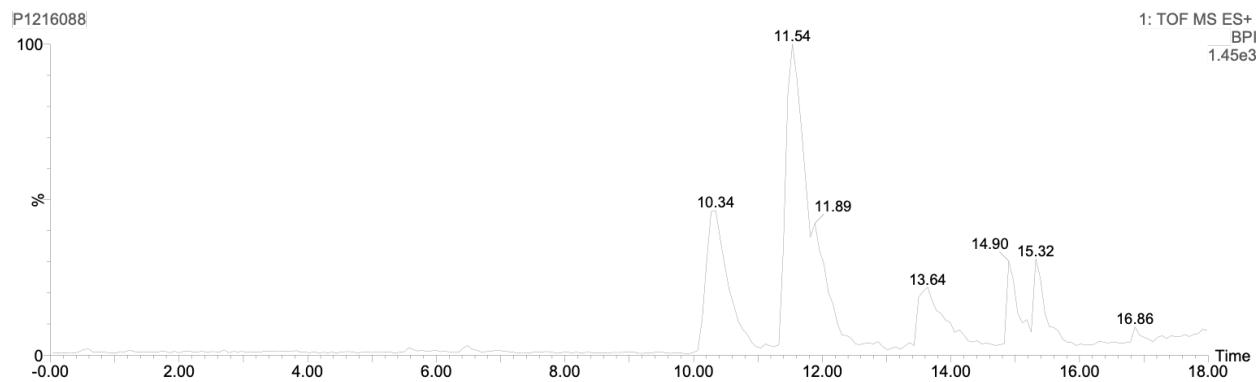


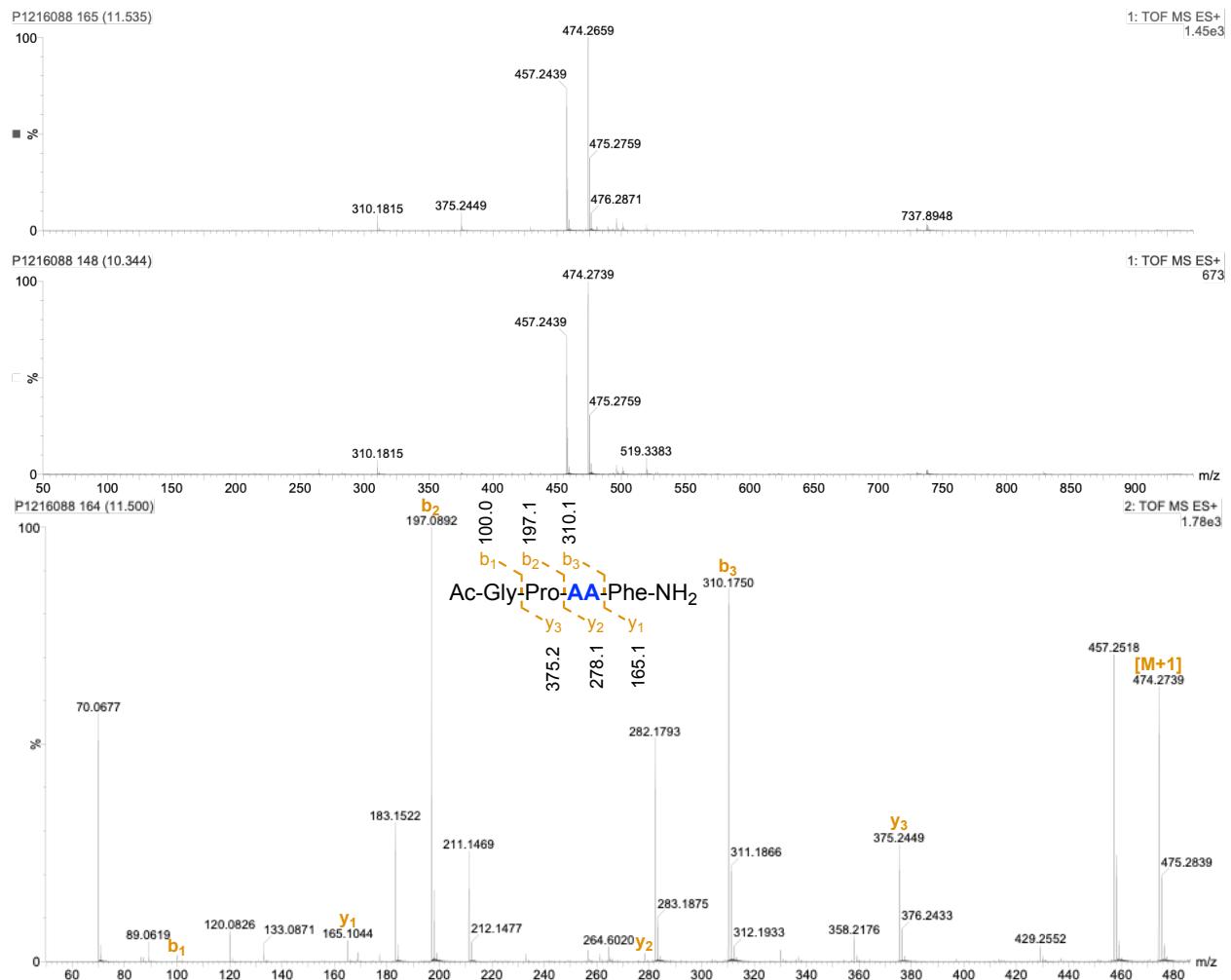


10H': MW = 473.6, Purity = 68.9%, Yield = 28.7% [0.32 mg]

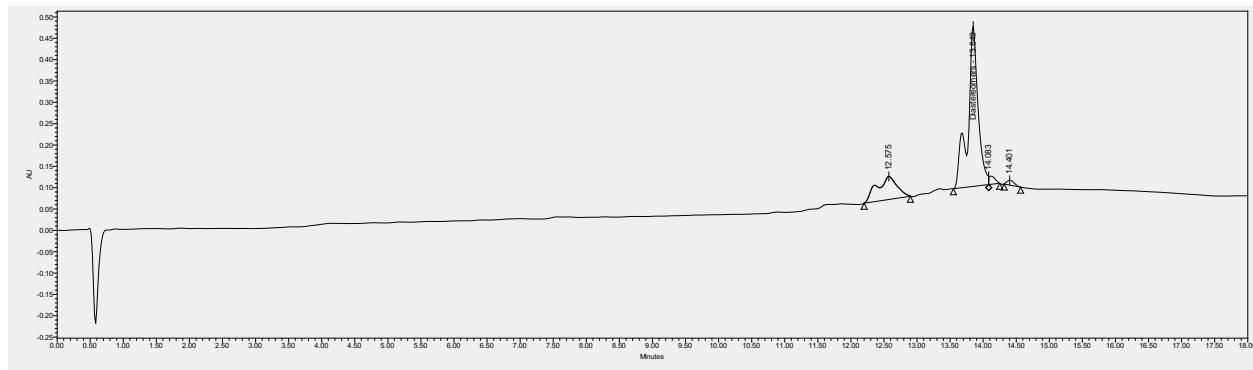
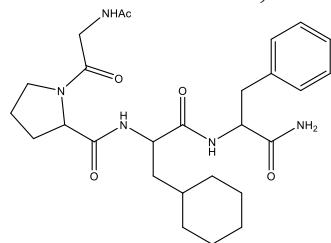


	Name	Retention Time	Area	% Area
1		6.412	62010	0.66
2	Diastereomer 1	11.064	1801359	19.06
3	Diastereomer 2	12.037	4712137	49.87
4		12.293	2113029	22.36
5		13.534	760558	8.05

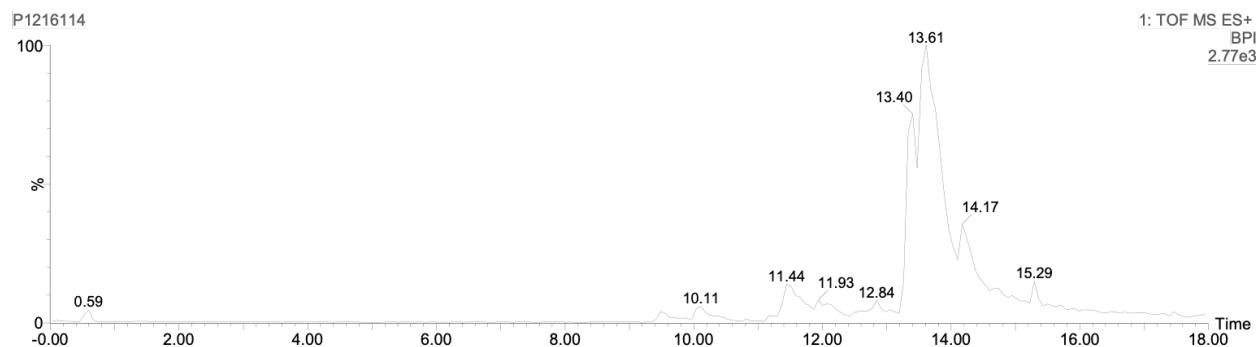


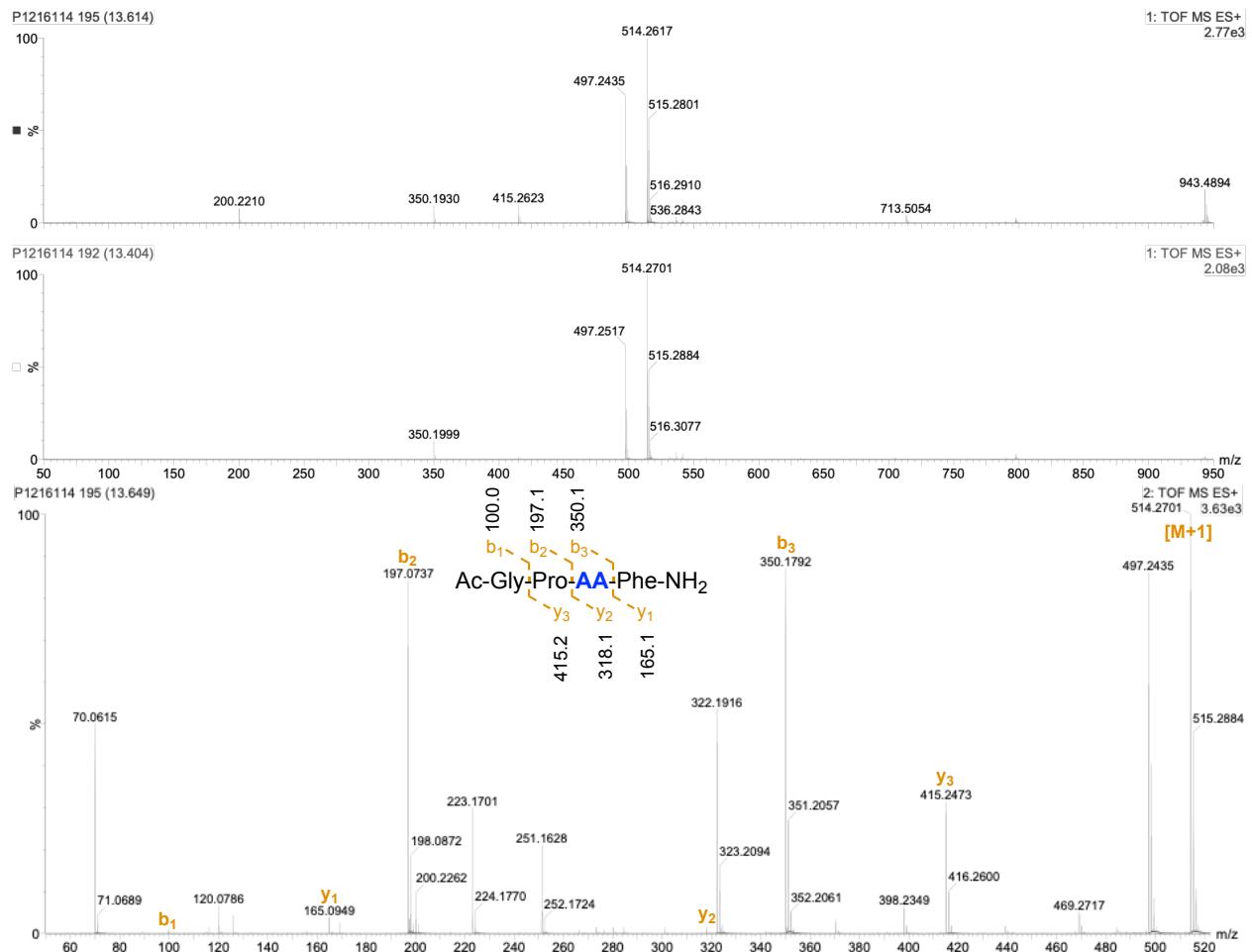


11H': MW = 513.6, Purity = 75.1%, Yield = 13.1% [0.16 mg]

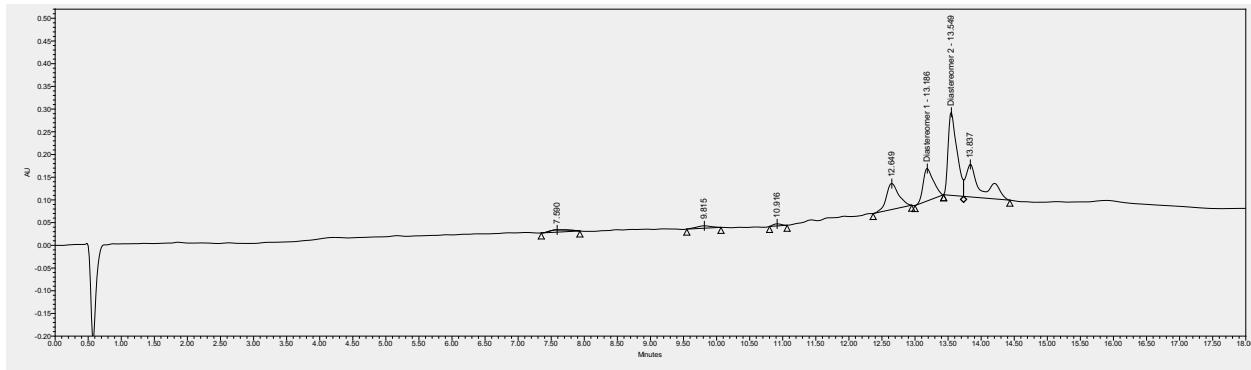
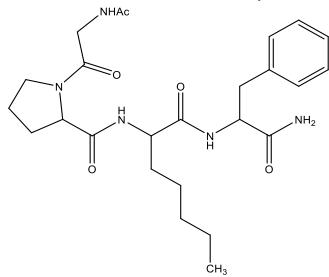


	Name	Retention Time	Area	% Area
1		12.575	1147553	21.10
2	Diastereomers	13.849	4083102	75.07
3		14.083	121884	2.24
4		14.401	86797	1.60

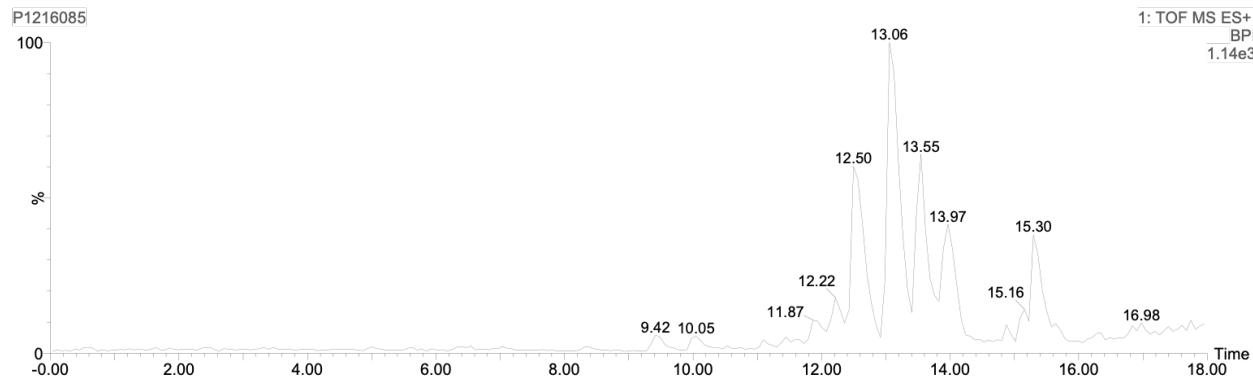


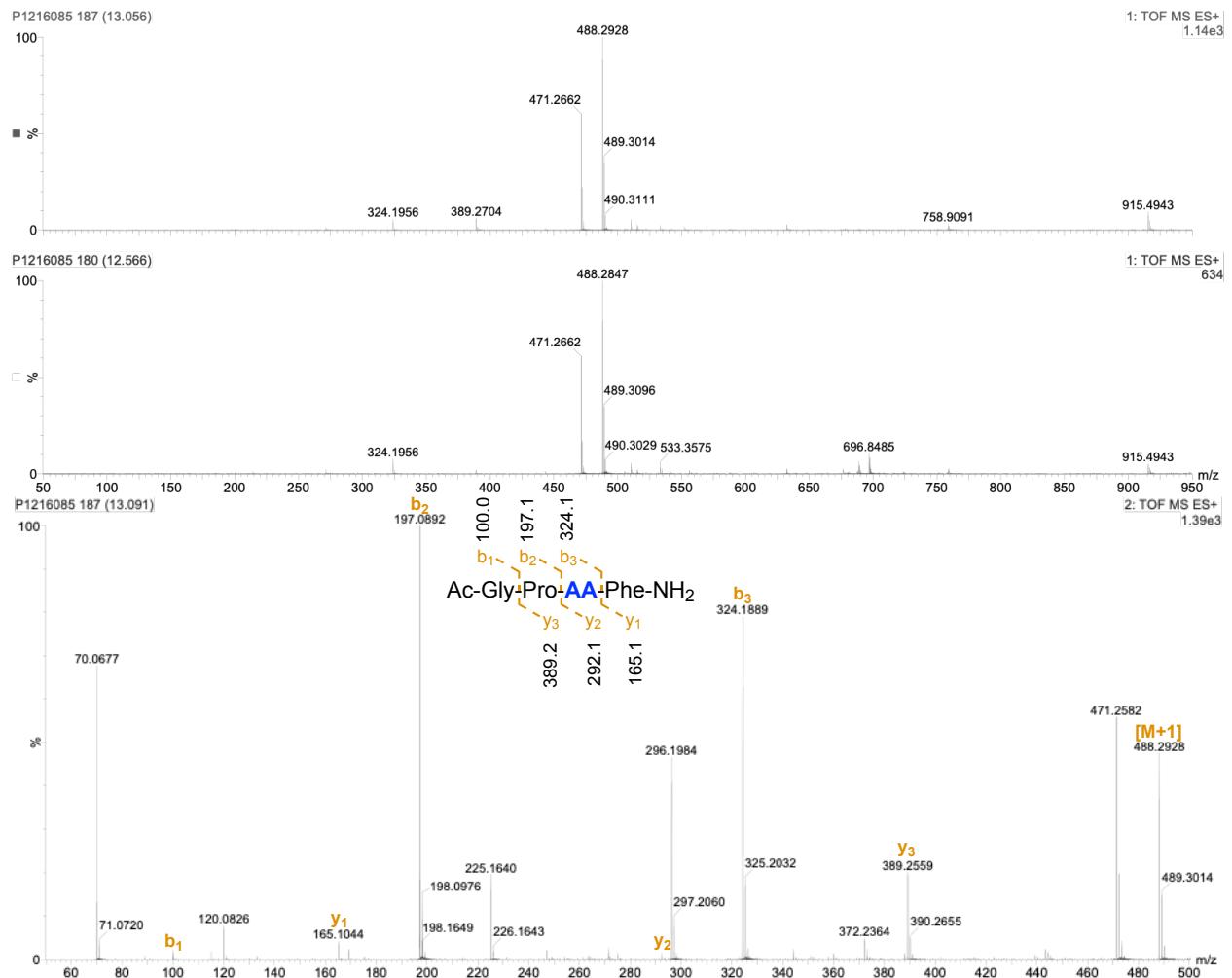


12H': MW = 487.6, Purity = 53.6%, Yield = 7.9% [0.090 mg]



	Name	Retention Time	Area	% Area
1		7.590	102030	2.21
2		9.815	82593	1.79
3		10.916	40026	0.87
4		12.649	747634	16.22
5	Diastereomer 1	13.186	773407	16.78
6	Diastereomer 2	13.549	1697330	36.83
7		13.837	1165701	25.29





References

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- 2 P. M. Morrison, P. J. Foley, S. L. Warriner, M. E. Webb, *Chem. Commun.* 2015, **51**, 13470–13473.
- 3 B. J. H. Kuipers, H. Gruppen, *J. Agric. Food Chem.* 2007, **55**, 5445–5451.
- 4 G. Torsi, G. Chiavari, C. Laghi, A. M. Asmundsdottir, F. Fagioli, R. Vecchietti, *J. Chromatogr. A* 1989, **482**, 207–214.