

## Supplementary Information

October 21, 2021

### 1 Two-body potential

Here we reproduce the definition of the two body potential  $V_B^{xy}(r)$  between a linker  $x$  and a linker  $y$  shown in<sup>1,2</sup>:

$$V_B^{xy}(r) = \frac{4\lambda_x\lambda_y\lambda_m \operatorname{kei}(q_0r)}{\pi^2(\lambda_m^2 + \lambda_x\lambda_y + \lambda_x\lambda_m + \lambda_y\lambda_m) - 16\lambda_x\lambda_y \operatorname{kei}(q_0r)} \left( \pi\Delta l_x\Delta l_y + \left( \frac{2\Delta l_x^2\lambda_x}{\lambda_x + \lambda_m} + \frac{2\Delta l_y^2\lambda_y}{\lambda_y + \lambda_m} \right) \operatorname{kei}(q_0r) \right) + \frac{1}{2} \log \left( 1 - \frac{16\lambda_x\lambda_y \operatorname{kei}(q_0r)^2}{\pi^2(\lambda_x + \lambda_m)(\lambda_y + \lambda_m)} \right). \quad (1)$$

Here,  $\Delta l_i = h_0 - l_i$ ,  $q_0$  describes the inverse of the correlation length, and  $\lambda_m$  is the effective spring stiffness of the membrane, related to the inverse of the fluctuation amplitude of the unbound membrane<sup>3</sup>.

When assuming a tensionless membrane of stiffness  $\kappa$ , residing in a minimum of a potential with a curvature  $\gamma$ ,  $q_0 = 1/\sqrt[4]{\kappa/\gamma}$  and,  $\lambda_m = 8\sqrt{\kappa\gamma}$ . Analogous equations can exist for membranes with tension<sup>3</sup>.

In the simulations, the particles are prevented from residing at the same lattice site, which in the potential has the effect of a hard-core repulsion below a distance given by the lattice spacing  $a$ . Therefore we modify the potential as follows

$$V_2^{xy}(r) = \begin{cases} \infty & r \leq a \\ V_B^{xy}(r) & r > a. \end{cases} \quad (2)$$

### 2 Expected coordination number

In the random system containing  $N_1$  links of type 1 and  $A - 1 - (N_1 - 1) = A - N_1$  lattice sites that are empty or of type 2, the average probability to find a type 1 bond on any one lattice site is

$$\frac{N_1 - 1}{A - 1}, \quad (3)$$

and therefore the average number of neighbours is given by

$$n_b^e = \frac{4(N_1 - 1)}{A - 1}. \quad (4)$$

Another way to see this is by considering the average number of neighbours by the probability of finding  $n$  neighbours ( $p(n)$ ) on the available 1 to 4 neighbouring lattice sites  $\sum_{i=1}^4 n \cdot p(n)$ , which can be calculated by the probability of drawing either a type 1 bond or not from the total number of bonds,

$$n_b^e = 1 \cdot \frac{\binom{N_1-1}{1} \binom{A-N_1}{3}}{\binom{A-1}{4}} + 2 \cdot \frac{\binom{N_1-1}{2} \binom{A-N_1}{2}}{\binom{A-1}{4}} + 3 \cdot \frac{\binom{N_1-1}{3} \binom{A-N_1}{1}}{\binom{A-1}{4}} + 4 \cdot \frac{\binom{N_1-1}{4} \binom{A-N_1}{0}}{\binom{A-1}{4}} = \frac{4(N_1 - 1)}{A - 1}. \quad (5)$$

### 3 Monte Carlo Simulations

Benefitting from a recently developed coarse-grained Monte Carlo simulation framework<sup>2,4</sup>, we conduct Monte Carlo simulations on the growth of adhesion domains with multiple binding pairs to mimic the current experiments as closely as possible. Specifically, the GUV and SLB membranes are discretized into square lattices with lattice constant  $a$ . Periodic boundary conditions are considered here. A growing contact zone with an increasing radius is located in the middle of the simulation box, while the DNA linkers on the free region between GUV and SLB membranes maintain constant density, providing the reservoir reconstructing the appropriate statistical ensemble. The simulation system evolves in time by alternate execution of diffusion, stochastic binding/unbinding as well as growth steps. The mobility of freely diffusive DNA linkers is treated as a random walk with time step  $\Delta t$ . Here,  $D_{\text{DNA}}$  represents the diffusion coefficient of DNA constructs which is taken to be the same for all DNA lengths because they all have the same membrane attachments. The forming and breaking of intermittent bridges between the two membranes follow a coarse-grained kinetics<sup>5</sup>. DNA linkers can switch from an intra- to an inter-membrane configuration only if the opposing lattice site is empty, while breaking the bridge places the intra-membrane DNA construct on the SLB or the vesicle with equal probability. The effective association/dissociation rates coupling to the thermodynamic properties of membranes can be analytically obtained. In addition, the contact zone radius will increase at a slow

constant rate, 0.001 per time step, from initially zero to the final state with  $R = 30$ , so that the concentration of bridges is always in equilibrium with the instantaneous area of contact between the two membranes. The further details of our Monte Carlo simulations are described in Ref. <sup>2,4</sup>. Using the representative system parameters summarized in Table 1 of the manuscript, we can perform a series of simulations and understand the DNA linker mediated membrane adhesion.

## 4 Membrane parameters

The full list of parameters used in the simulation is shown in Table 1. The simulation box is made from  $N$  cells with lattice constant  $a$ . The contact zone grows at a constant rate  $\Delta R/\Delta t$  that  $\Delta R$  is the incremental radius of contact zone at each time step,  $\Delta t$ . The diffusion coefficients of both short and long free DNA constructs are represented by  $D_{\text{DNA}}$ . The immobilization of DNA bridges is considered.  $C_S^0$  and  $C_L^0$  stand for the initial concentrations of short and long DNA constructs, respectively.  $C_{BS}$  and  $C_{BL}$  are the initial concentrations of short and long DNA bridges. The interaction between single chol and its binding site in the bilayer is associated with the insertion energy  $E_i$ , interaction range  $\alpha_S$ ,  $\alpha_L$  and intrinsic reaction rate  $k_{0S}$ ,  $k_{0L}$ . Due to common design of the sticky ends,  $E_i$  is identical for the short and long linker. The remaining parameters are explained in the table caption in the main text.

**Table 1** Full list of membrane parameters used in the simulation.

| Quantity                                 | Symbol                           | Value   |
|--|----------------------------------|---|
| Lattice constant                         | $a$                              | 10 nm   |
| Number of cells                          | $N$                              | $80 \times 80$  |
| Growth rate of the contact zone          | $\Delta R/\Delta t$              | 0.05 nm   |
| Diffusion coefficient of $L_S$ and $L_L$ | $D_{\text{DNA}}$                 | $0.16 \mu\text{m}^2/\text{s}$                         |
| Diffusion coefficient of bridges         |                                  | 0   |
| Initial concentration of DNA             | $C_S^0 + C_L^0$                  | 50%   |
| Initial concentration of bridges         | $C_{BS}, C_{BL}$                 | 0   |
| Time step                                | $\Delta t = a^2/4D_{\text{DNA}}$ | $156.2 \mu\text{s}$                                   |
| Membrane bending rigidity                | $\kappa$                         | $30 k_B T$  |
| Potential curvature                      | $\gamma$                         | $2.17 \times 10^5 k_B T/\mu\text{m}^4$ <sup>6,7</sup> |
| Potential height                         | $h_0$                            | $50 \text{ nm}$ <sup>6,7</sup>                        |
| Temperature                              | $T$                              | 300 K   |
| Fluctuations amplitude                   | $\sigma_0$                       | $7 \text{ nm}$ <sup>6,7</sup>                         |
| DNA linker elastic modulus               | $\lambda_S, \lambda_L$           | $3 \times 10^4 k_B T/\mu\text{m}^2$ <sup>8</sup>      |
| DNA linker length                        | $l_S$                            | $8 \text{ nm}$ <sup>8</sup>                           |
|  | $l_L$                            | $15 \text{ nm}$ <sup>8</sup>                          |
| Insertion energy                         | $E_i$                            | $6 k_B T$ <sup>9</sup>                                |
| Interaction range                        | $\alpha_S = \alpha_L$            | $1 \text{ nm}$ <sup>4</sup>                           |
| Intrinsic reaction rate                  | $k_{0S} = k_{0L}$                | $1000/\text{s}$ <sup>10</sup>                         |

## References

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