

## Supporting Information

# Homointerface covalent organic framework membranes for efficient desalination

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# 1. Methods

## 1.1 Materials

All chemicals and solvents were commercially available and used as received. 2,5-Diaminobenzenesulfonic acid (DABA) was purchased from Saen chemical technology (Shanghai) Co., Ltd. 1,3,5-Triformylphloroglucinol (TFP) was supplied by Jilin Chinese Academy of Sciences-Yanshen Technology Co., Ltd. p-Phenylenediamine (PA), 1,3,5-trimethylbenzene and acetic acid were provided by Shanghai Aladdin Biochemical Technology Co., Ltd. Hydrazine hydrate ( $\text{N}_2\text{H}_4$ ) was obtained from Tianjin Jiangtian chemical technology co., Ltd. Sodium sulfate ( $\text{Na}_2\text{SO}_4$ ), magnesium sulfate ( $\text{MgSO}_4$ ), magnesium chloride ( $\text{MgCl}_2$ ) and sodium chloride ( $\text{NaCl}$ ) were obtained from Tianjin Kemiou Chemical Reagent Co. Ltd. Dopamine hydrochloride was purchased from Shanghai Macklin Biochemical Co., Ltd. Tris(hydroxymethyl) aminomethane-hydrochloric acid buffer (Tris-HCl, 50.0 mM, pH=8.5) was purchased from Nanjing Senbeijia Biotech Co., Ltd. Deionized water (DI) was obtained from a Milli-Q reverse osmosis system (Millipore, US). Polyacrylonitrile supports (PAN, MWCO=100 kDa) were purchased from Lanjing Membrane Technology Co., Ltd (Shandong, China).

## 1.2 Characterization

**SEM.** The surface and cross-sectional morphologies of the membranes were conducted by a field emission scanning electron microscopy (Hitachi S4800, Japan). All the membrane samples were sputtered with Au by a Q150T turbo-pumped sputter coater with the current of 25 mA for 120 s before SEM characterization.

**TEM.** The cross-sectional images of the membranes were obtained by a transmission electron microscopy (JEOL-2100F, Japan). Membrane sample was embedded by epoxy resin and then cut into slices (about 100 nm in thickness) by an ultramicrotome (Leica Ultracut R). Then, a microgrid copper was used to load these slice-shaped samples.

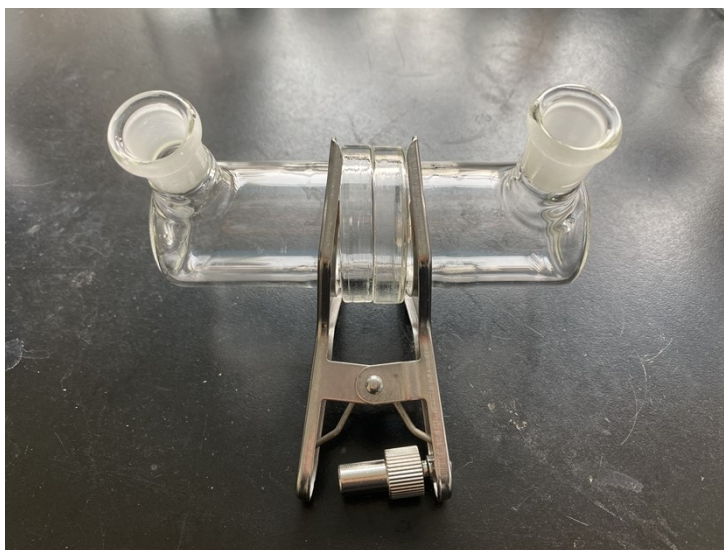
**AFM.** Three-dimensional morphology of the surface of COF membranes was characterized by Atomic force

microscopy (AFM, NTEGRA spectra, Russia).

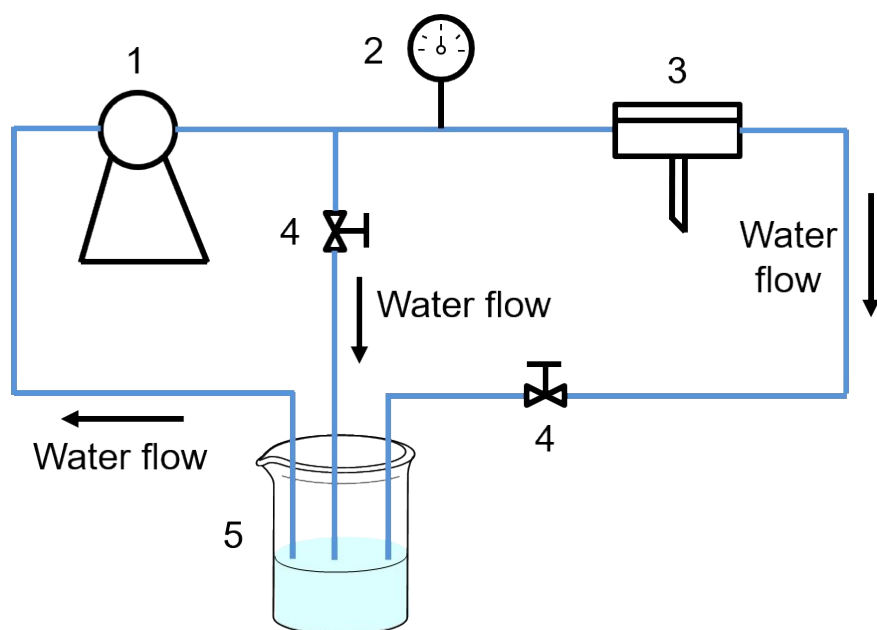
**Contact angle measurements.** Contact angles of the membranes were measured with a deionized water droplet of 3.0  $\mu\text{L}$  on membrane surface adopting a contact angle goniometer (JC2000C Contact Angle Meter, China). A high-speed camera was used to capture the instantaneous water contact angle of the membranes. Five membrane samples were measured and the values reported were averaged.

**Zeta potential.** The surface zeta potential of the membranes was performed by a SurPASS electrokinetic analyzer (Anton Paar KG, Austria) at room temperature ( $25 \pm 1$  °C). For each measurement, the membrane samples ( $1\text{ cm} \times 0.5\text{ cm}$ ) were attached to a holder to contact with 1  $\text{mmol L}^{-1}$  KCl solution ( $\text{pH}=6.0 \pm 0.2$ ). Each membrane sample was tested at least five times to obtain an average value.

## 2. Figures



**Fig. S1** Home-made diffusion cell.



1 Pump; 2 Pressure gage; 3 Membrane cell;  
4 valve; 5 Feed tank

**Fig. S2** Cross-flow filtration system for evaluating desalination performance of the membranes. Flow velocity: 45

L h<sup>-1</sup>, driving pressure: 5.0 bar, effective filtration area: 3.14 cm<sup>2</sup>.

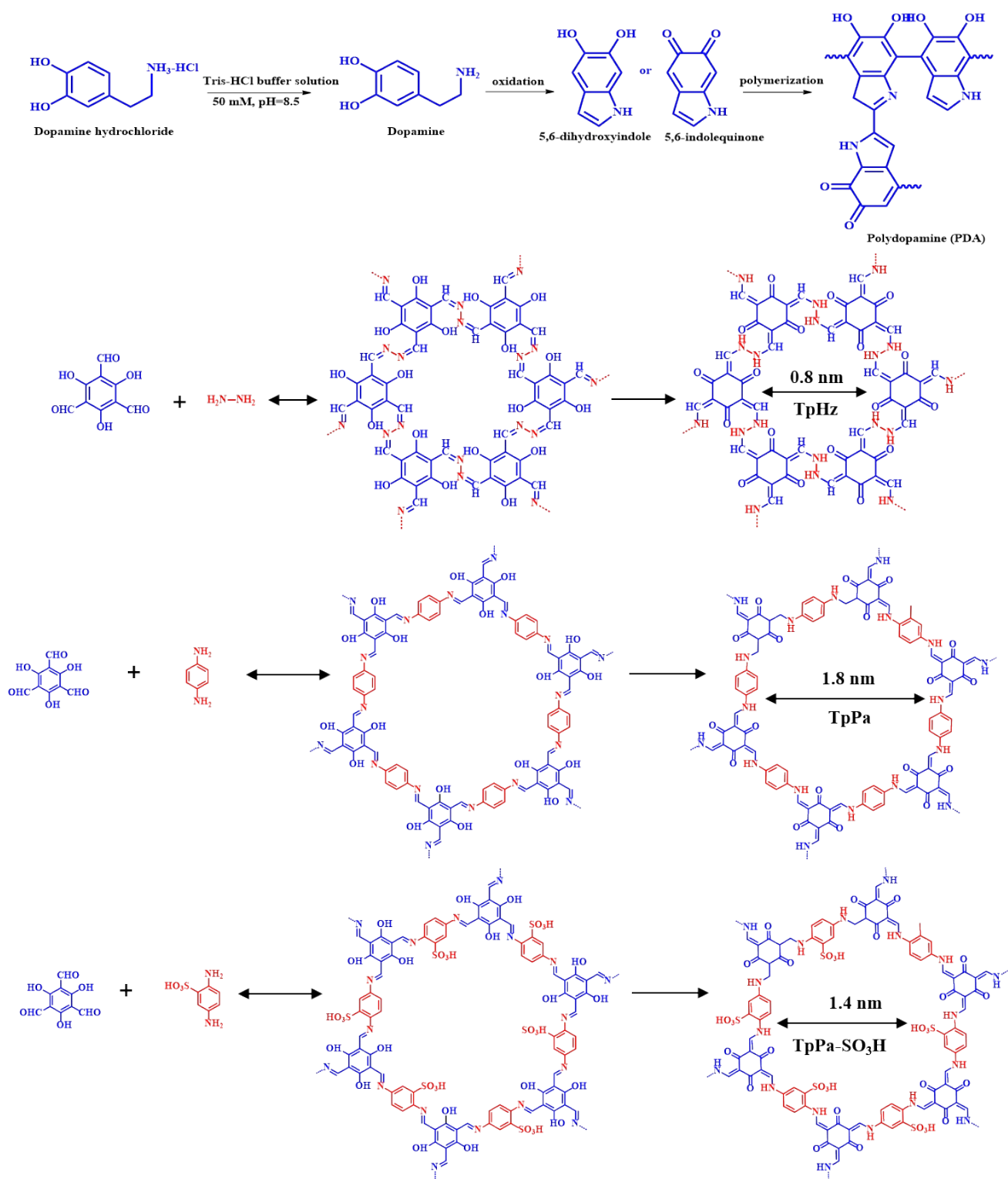
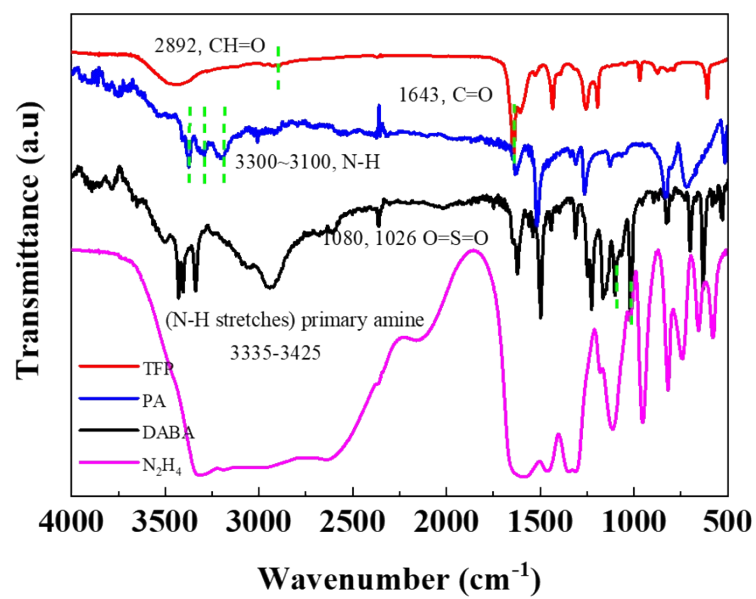
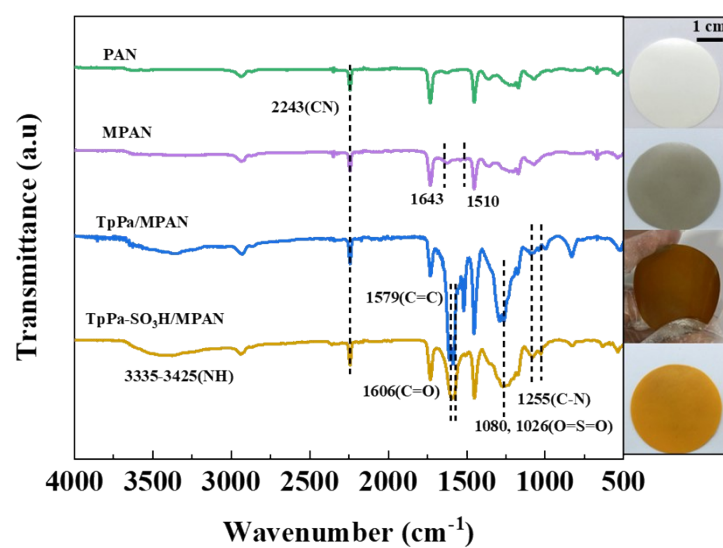


Fig. S3 Reaction route of polydopamine and COFs.



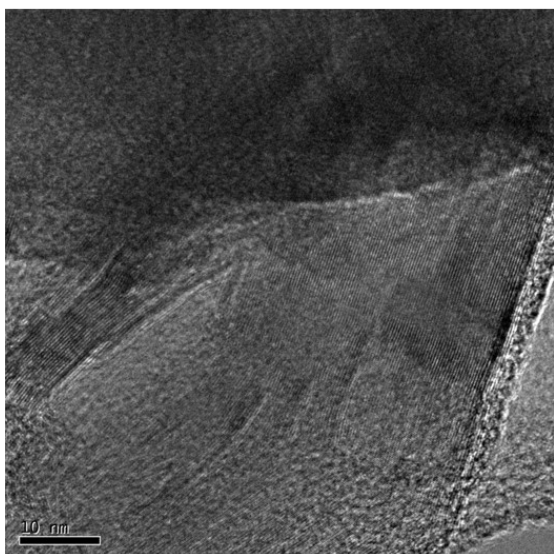
**Fig. S4** FTIR spectrum of the building monomers.



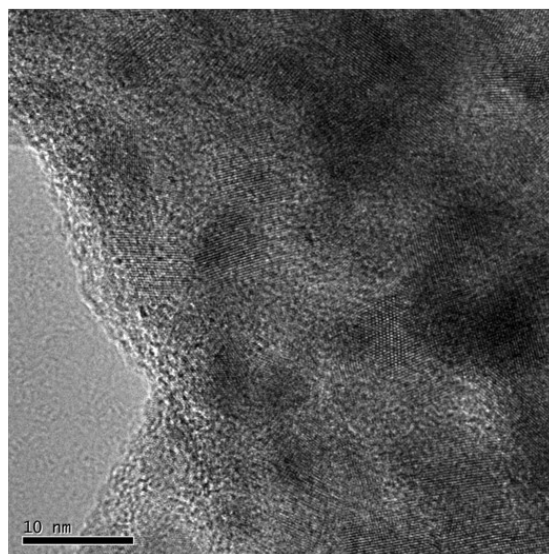
**Fig. S5** ATR-FTIR spectrum of the supports and COF membranes with single layer. From top to bottom: PAN support, MPAN support, TpPa/MPAN and TpPa-SO<sub>3</sub>H/MPAN membranes.



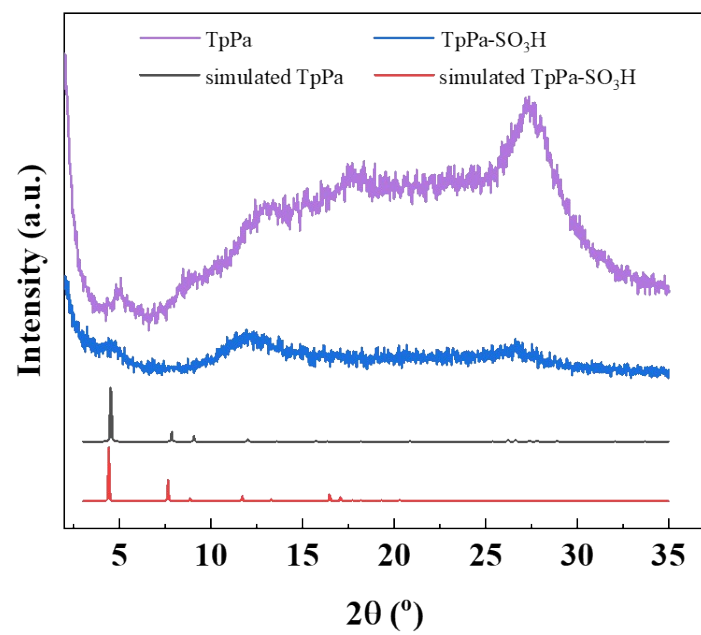
TpPa



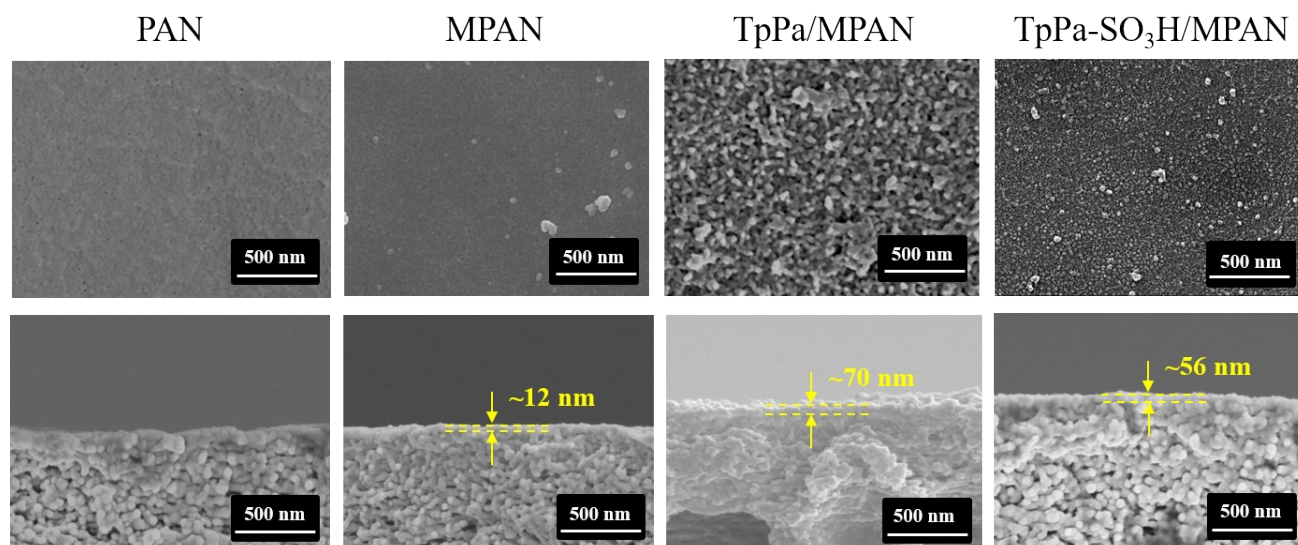
TpPa-SO<sub>3</sub>H



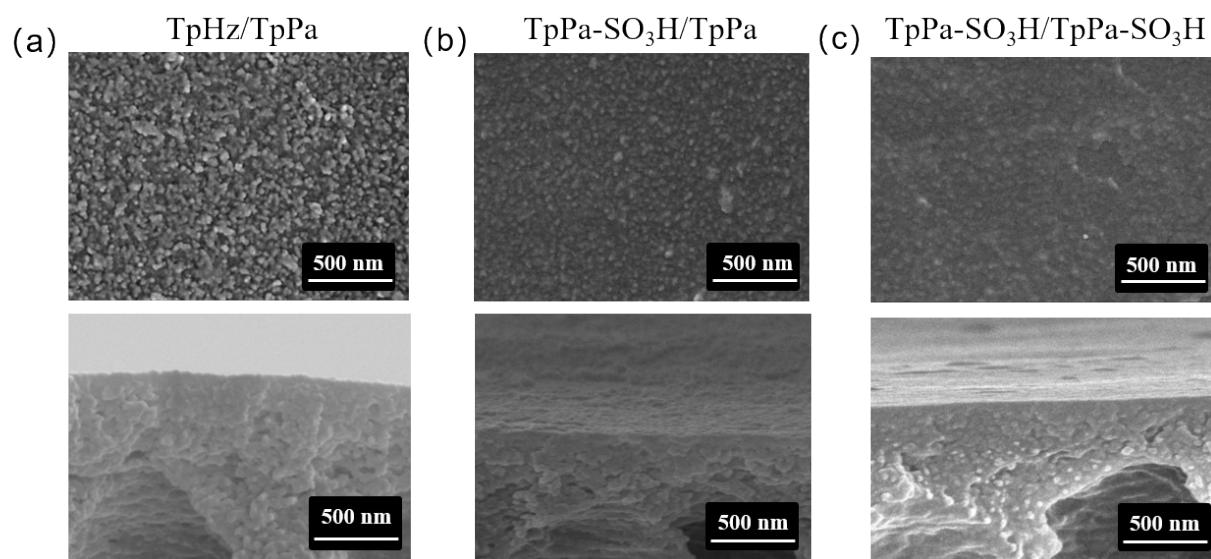
**Fig. S6** High-resolution TEM of TpPa and TpPa-SO<sub>3</sub>H nanofilms.



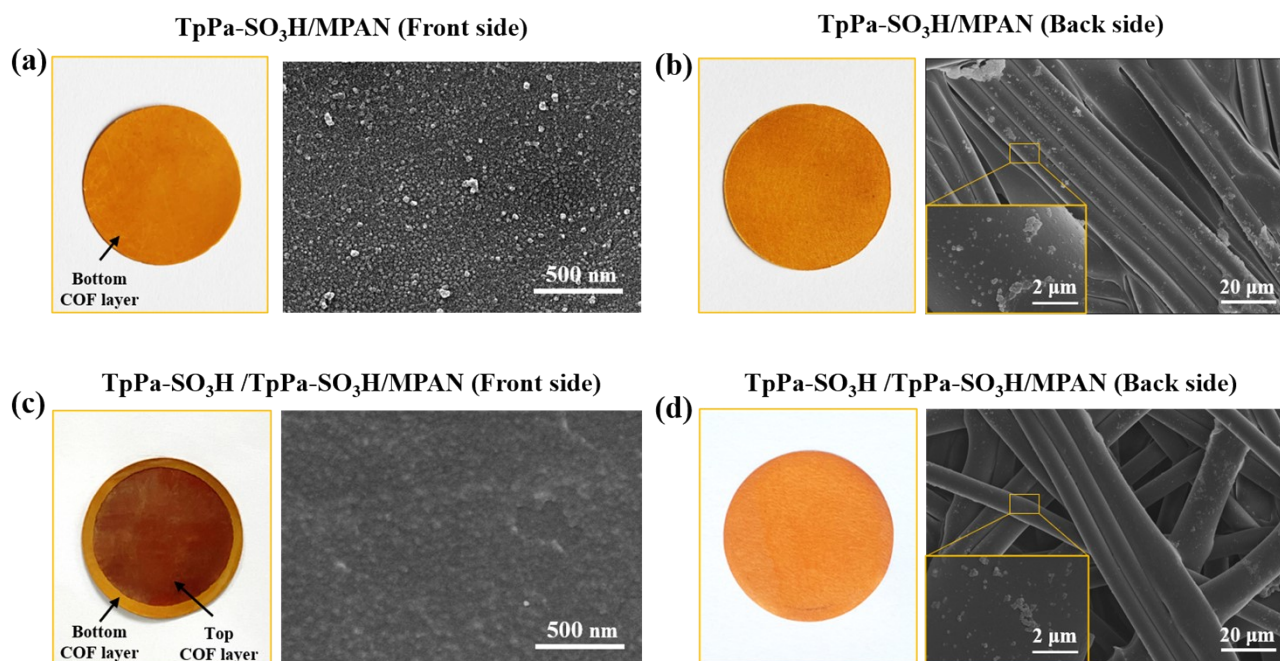
**Fig. S7** XRD pattern of the single layer COF membranes and the simulated COFs.



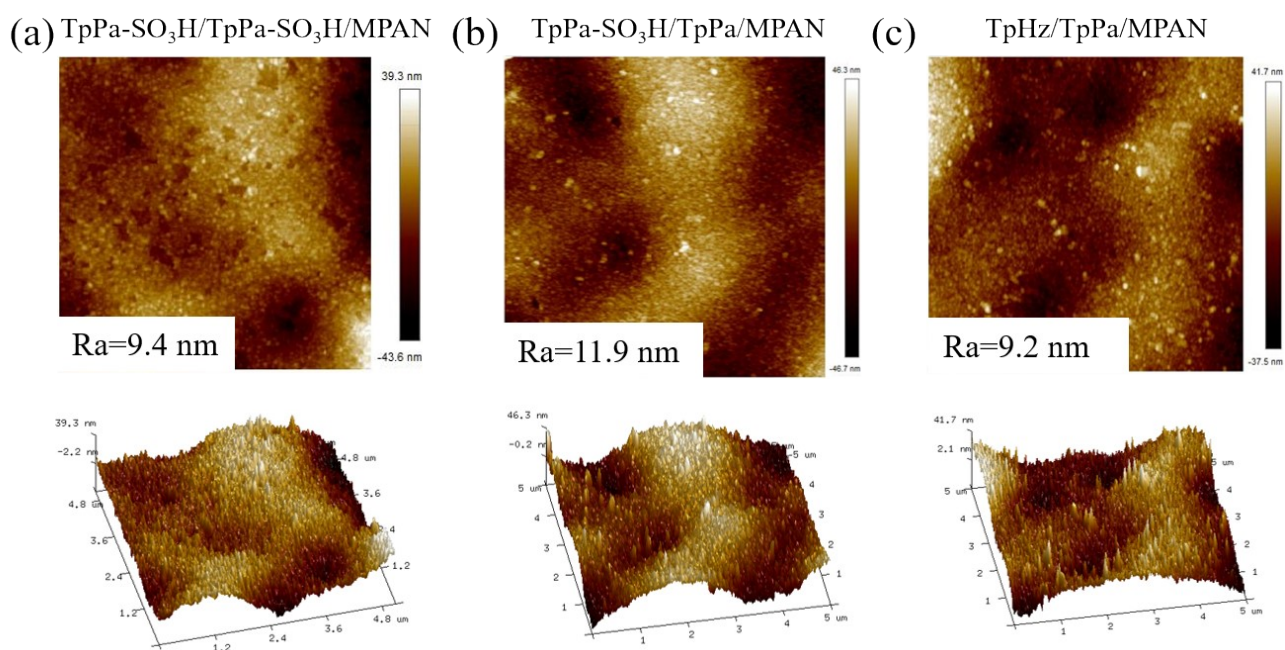
**Fig. S8** Surface and cross-sectional SEM images of the membranes.



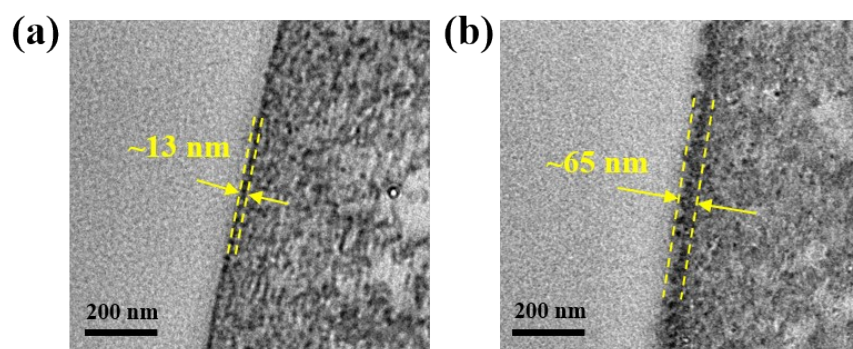
**Fig. S9** Surface and cross-sectional SEM images of the bilayer COF membranes.



**Fig. S10** The digital photographs (left) and surface SEM images (right) of the top side of TpPa-SO<sub>3</sub>H/MPAN (a) and TpPa-SO<sub>3</sub>H/TpPa-SO<sub>3</sub>H/MPAN (c). The digital photographs (left) and surface SEM images (right) of the bottom side of TpPa-SO<sub>3</sub>H/MPAN (b) and TpPa-SO<sub>3</sub>H/TpPa-SO<sub>3</sub>H/MPAN (d). Insets show an image at a higher magnification.

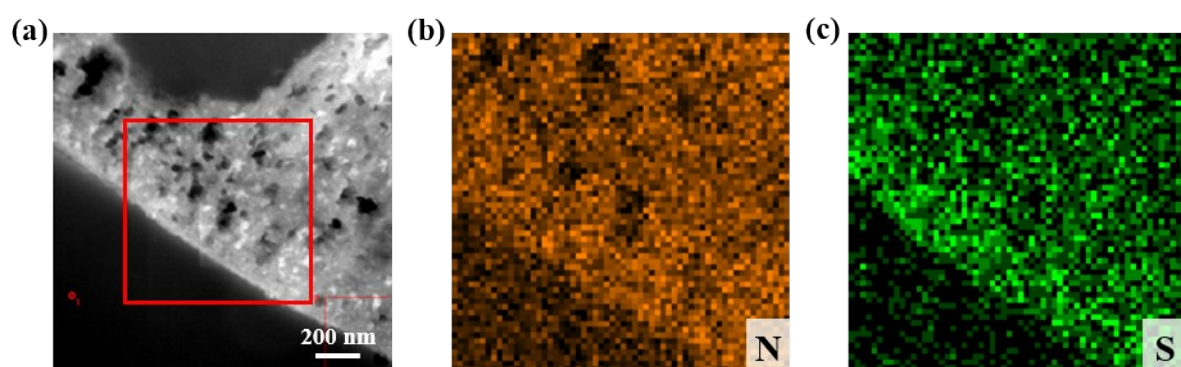


**Fig. S11** AFM topography of bilayer COF membranes. Scanning area: 5 μm by 5 μm.



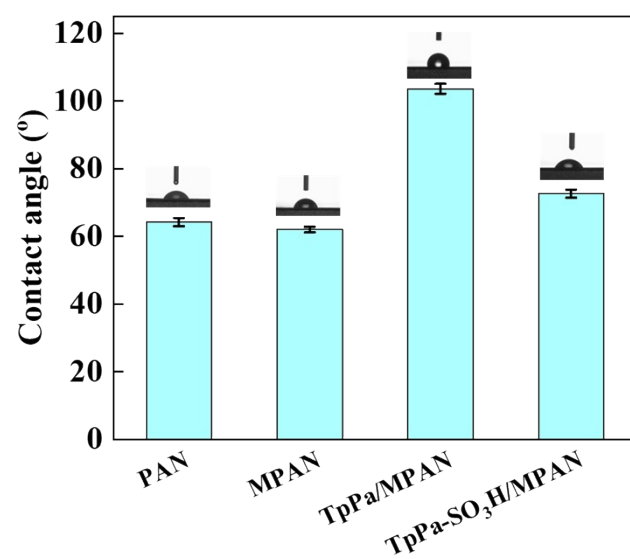
**Fig. S12** Cross-sectional TEM images of (a) MPAN support, (b) TpPa/MPAN membrane.



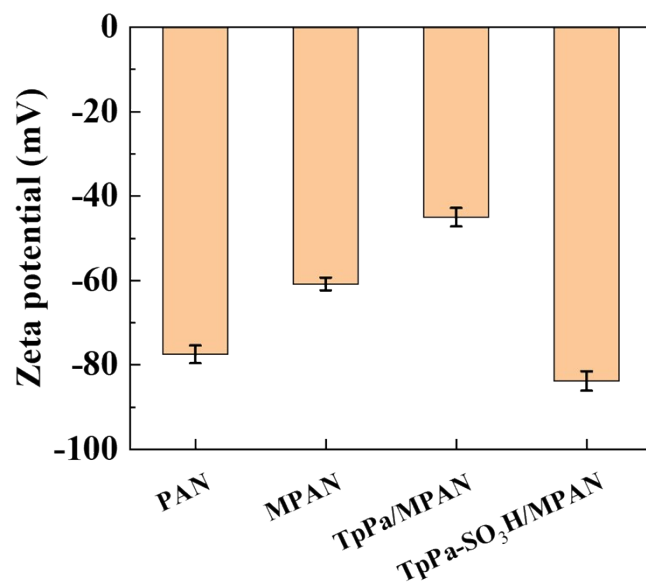


**Fig. S13** (a) Dark-field cross-sectional TEM images and corresponding element maps of (b) N, (c) S of TpPa-SO<sub>3</sub>H/MPAN membrane.

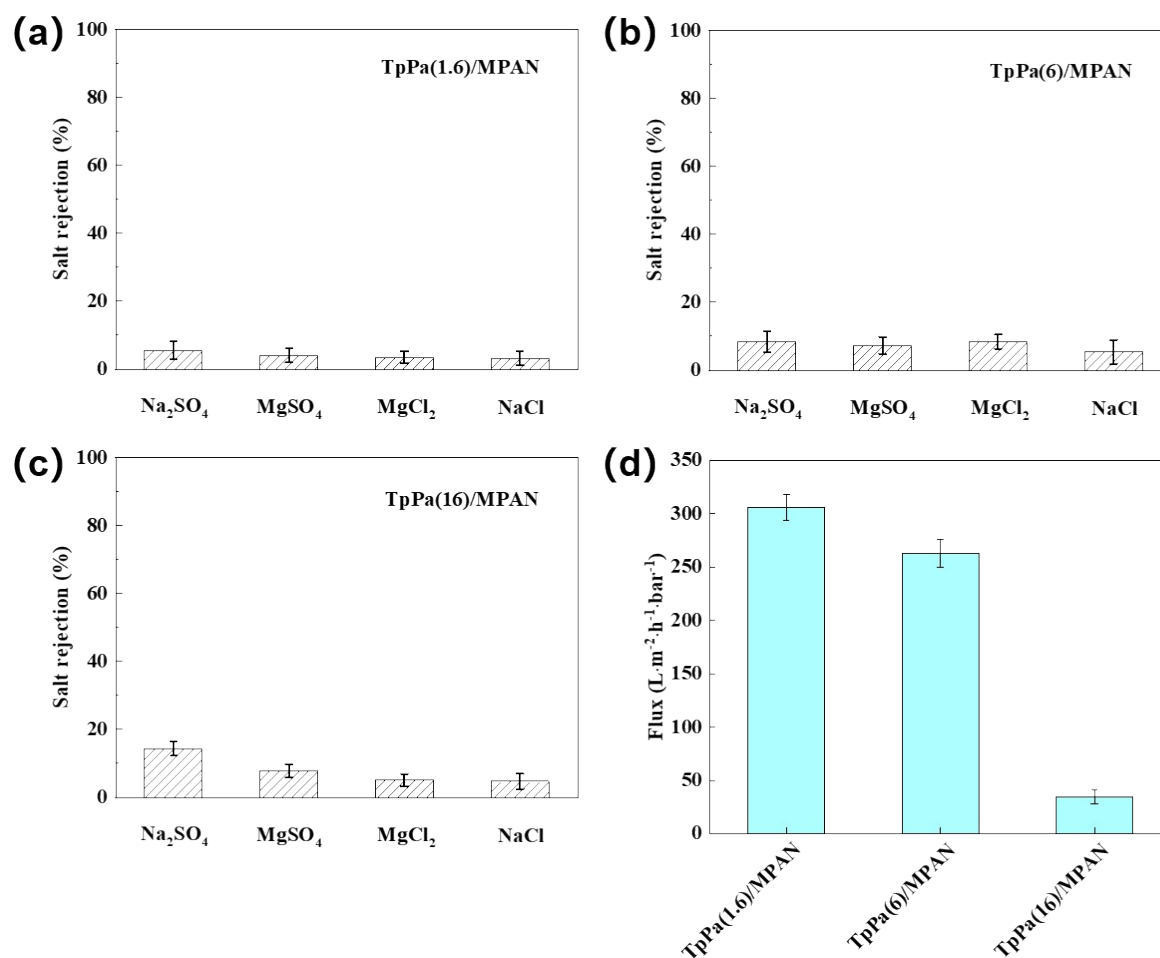




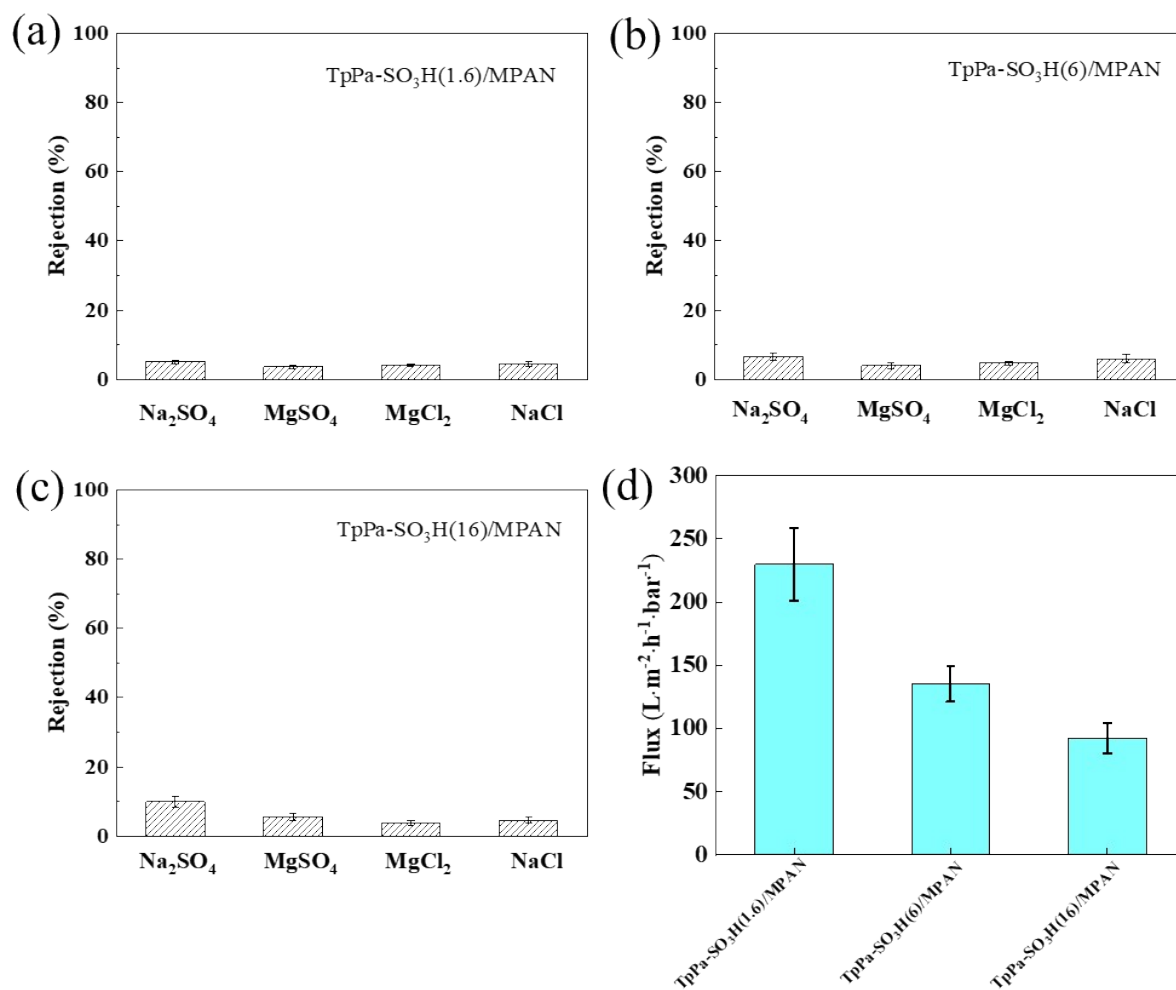
**Fig. S14** Water contact angle of the supports and COF membranes with single layer.



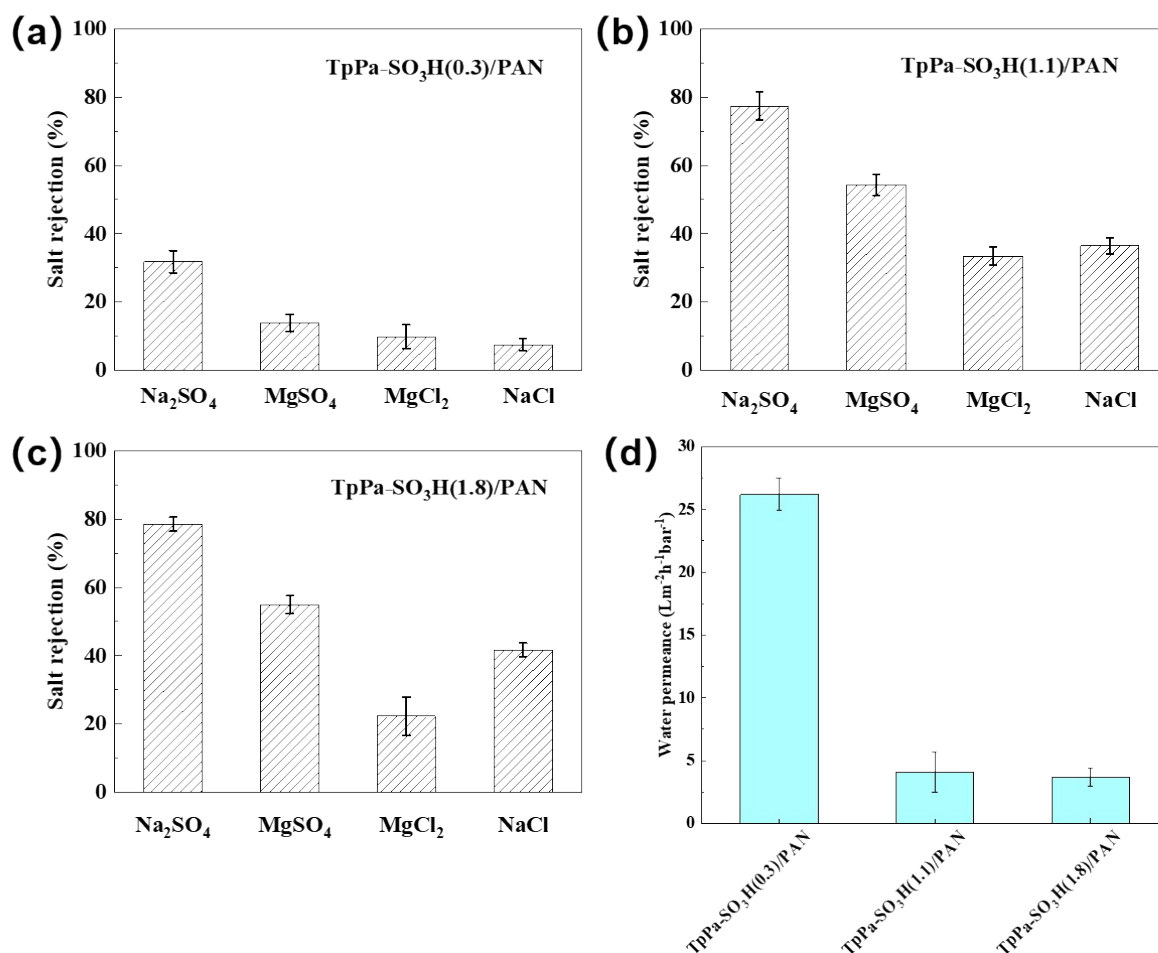
**Fig. S15** Zeta potential of the supports and COF membranes with single layer.



**Fig. S16** Salt rejection of (a-c) and water flux (d) of TpPa/MPAN membranes prepared *via in situ* growth technique at room temperature.

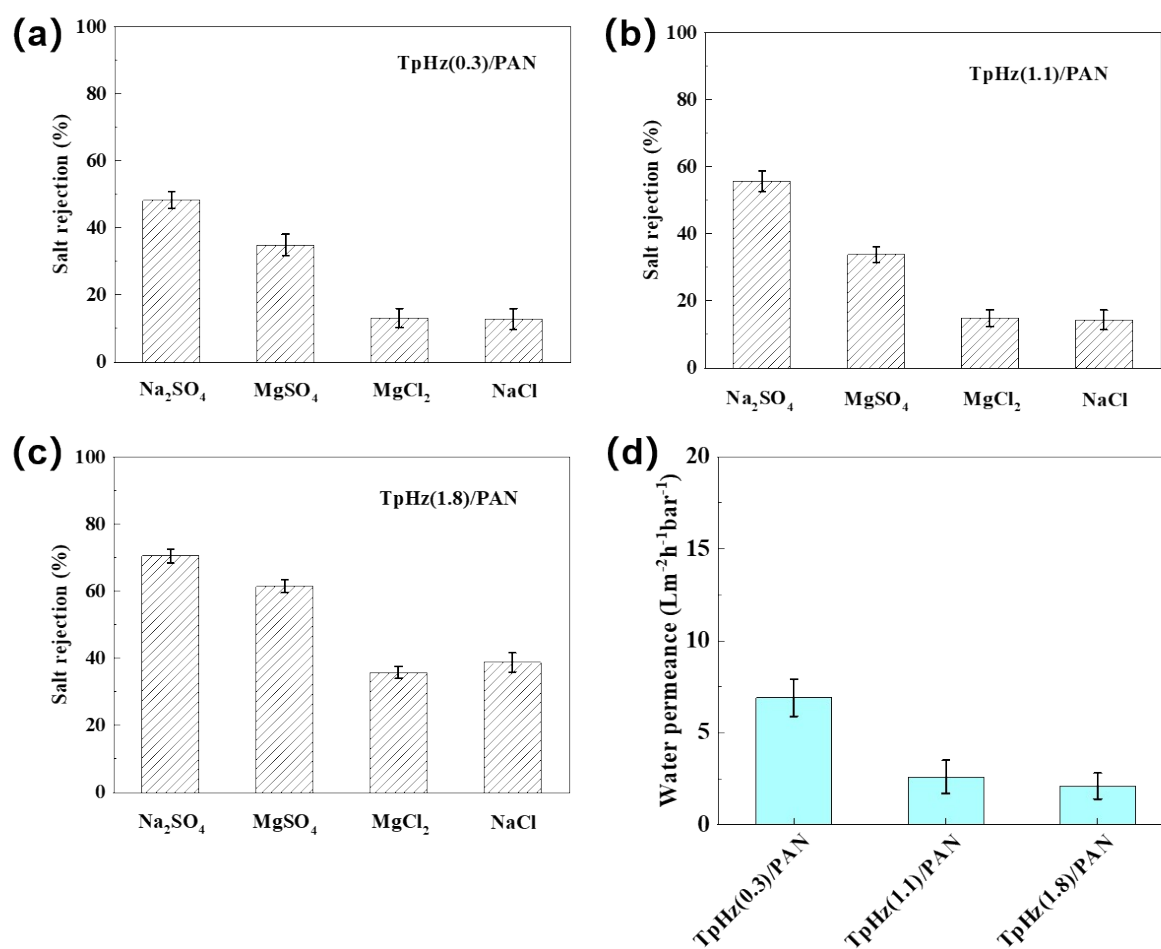


**Fig. S17** Salt rejection (a-c) and water flux (d) of TpPa-SO<sub>3</sub>H/MPAN membranes prepared *via in situ* growth technique at room temperature.



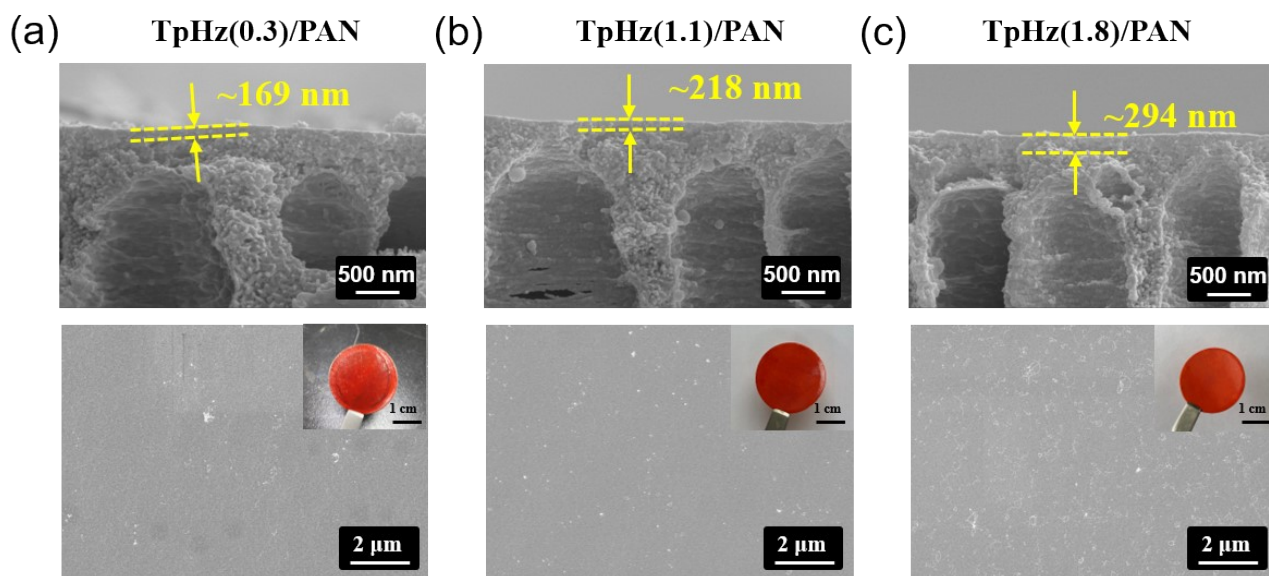
**Fig. S18** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of TpPa-SO<sub>3</sub>H/PAN

membranes prepared by counter-diffusion approach under room temperature.



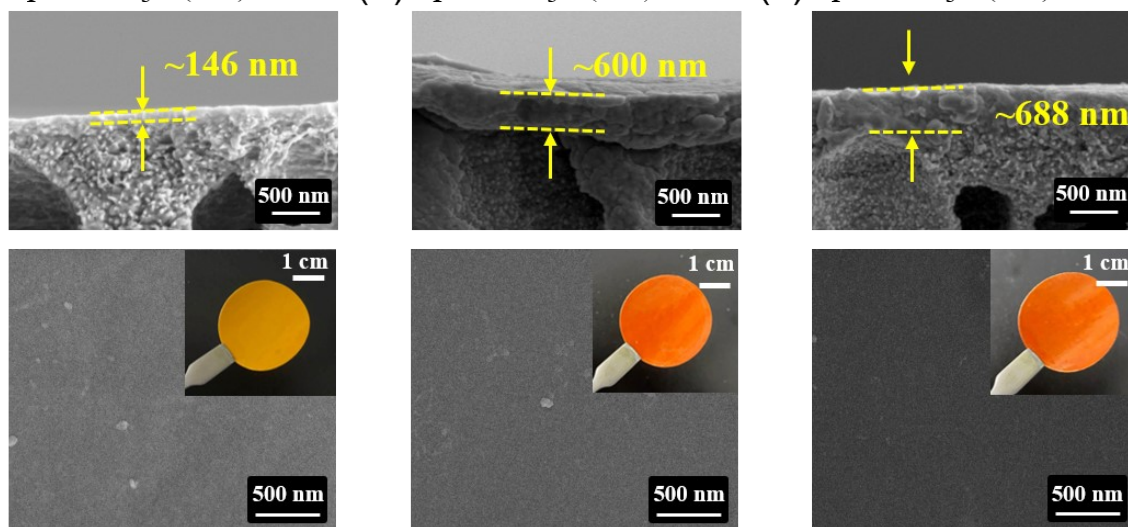
**Fig. S19** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of TpHz/PAN

membranes prepared by counter-diffusion approach under room temperature.



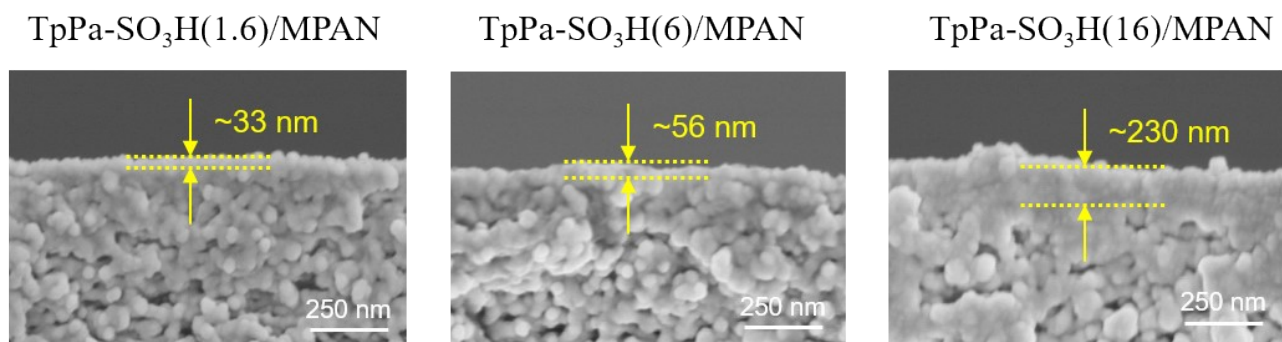
**Fig. S20** SEM images of the TpHz/PAN membranes prepared via counter-diffusion approach under room temperature.

(a) TpPa-SO<sub>3</sub>H(0.3)/PAN (b) TpPa-SO<sub>3</sub>H(1.1)/PAN (c) TpPa-SO<sub>3</sub>H(1.8)/PAN



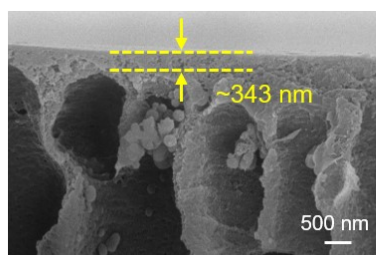
**Fig. S21** SEM images of the TpPa-SO<sub>3</sub>H/PAN membranes prepared via counter-diffusion approach under room temperature.



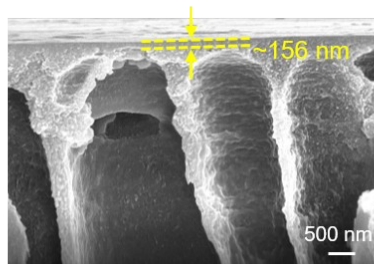


**Fig. S22** Cross-sectional SEM images of the TpPa-SO<sub>3</sub>H/MPAN membranes prepared via *in situ* growth under room temperature.

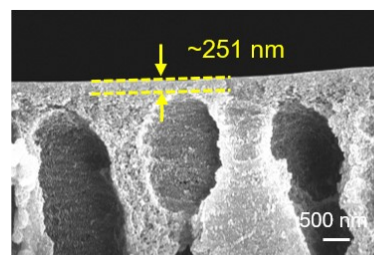
TpPa-SO<sub>3</sub>H(1.1)/TpPa-SO<sub>3</sub>H(1.6)



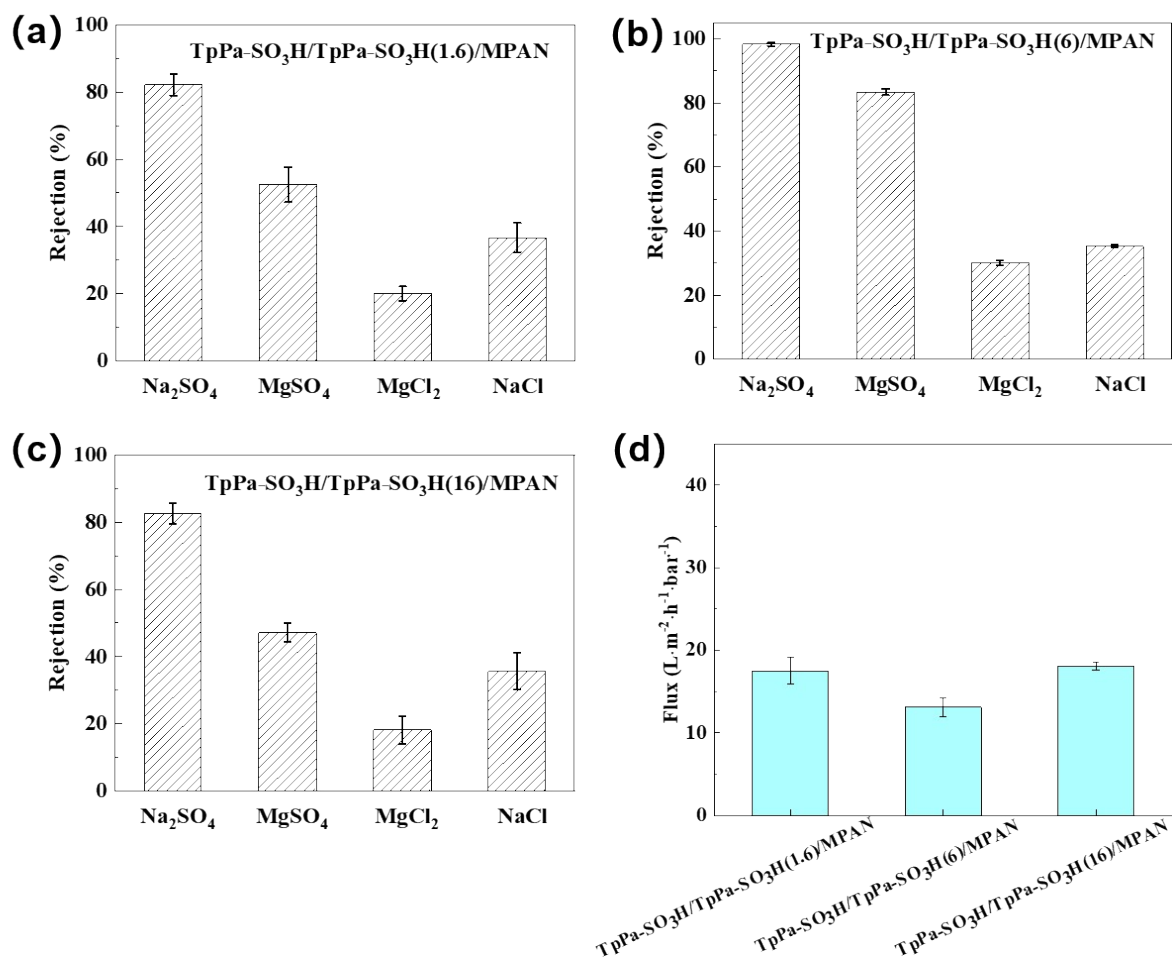
TpPa-SO<sub>3</sub>H(1.1)/TpPa-SO<sub>3</sub>H(6)



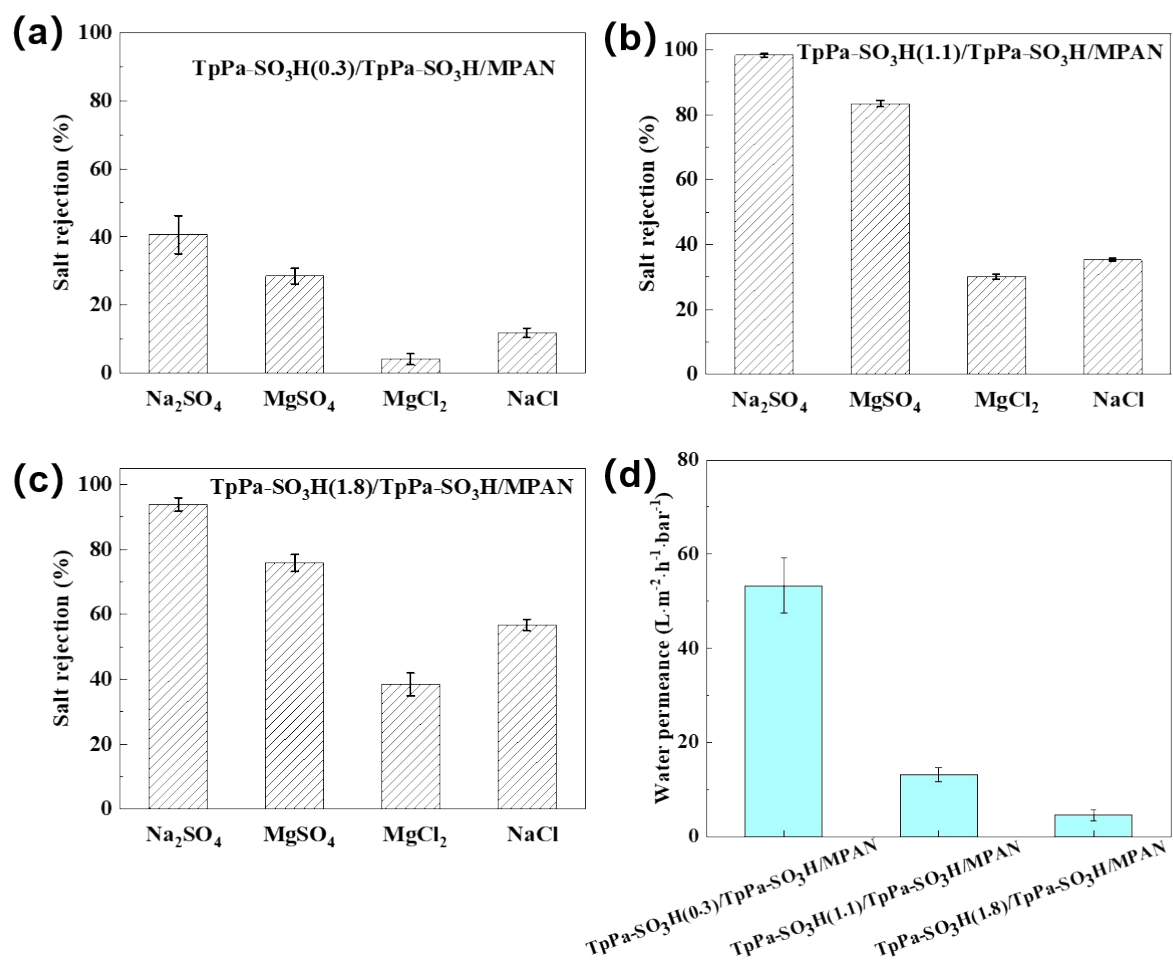
TpPa-SO<sub>3</sub>H(1.1)/TpPa-SO<sub>3</sub>H(16)



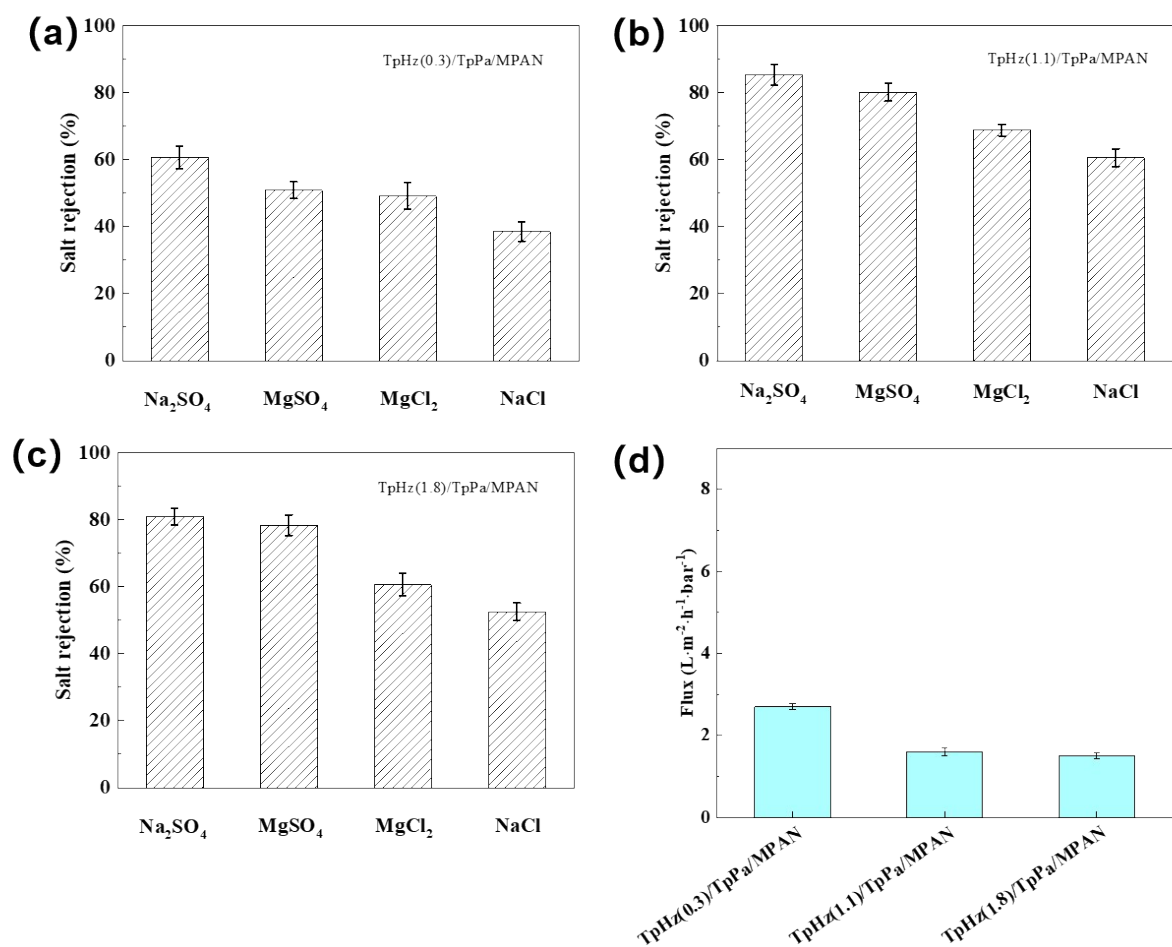
**Fig. S23** Cross-sectional SEM images of the homointerface TpPa-SO<sub>3</sub>H/TpPa-SO<sub>3</sub>H/MPAN membranes.



**Fig. S24** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of homointerface TpPa-SO<sub>3</sub>H(1.1)/TpPa-SO<sub>3</sub>H/MPAN membranes.

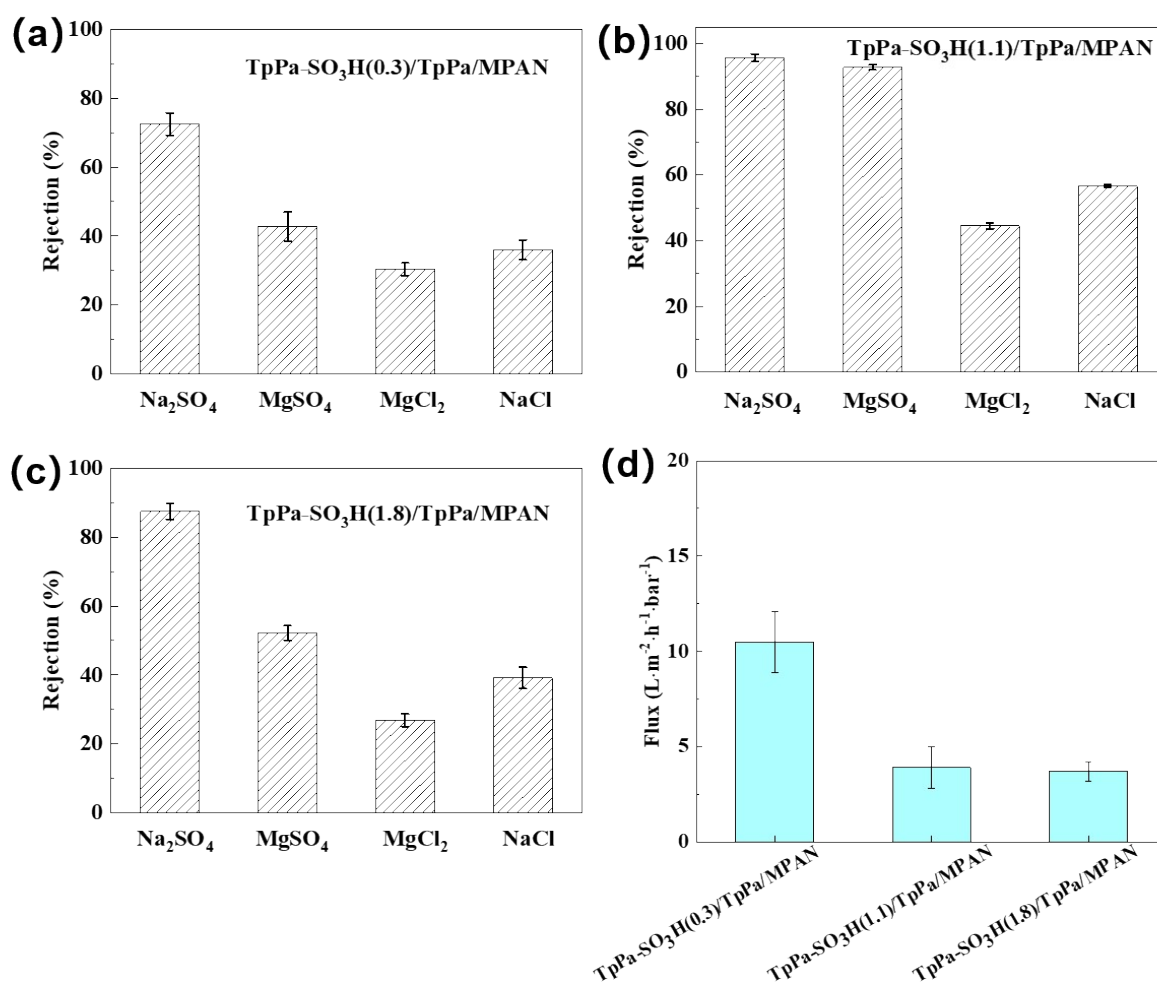


**Fig. S25** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of homointerface TpPa-SO<sub>3</sub>H/TpPa-SO<sub>3</sub>H(6)/MPAN membranes.

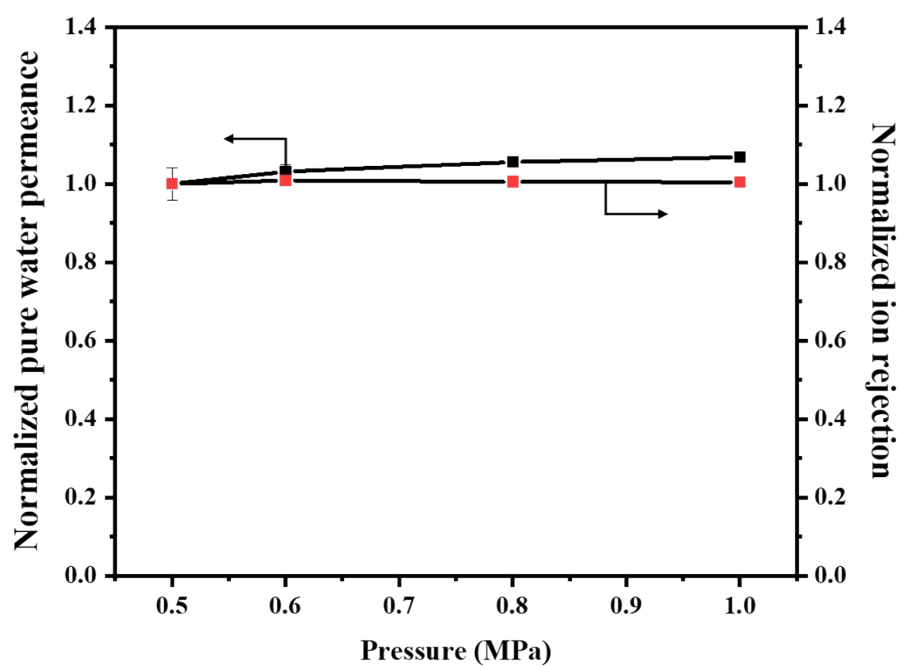


**Fig. S26** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of homointerface

TpHz/TpPa(6)/MPAN membranes.



**Fig. S27** Effect of monomer concentration on (a-c) salt rejection and (d) water permeance of homointerface TpPa-SO<sub>3</sub>H/TpPa(6)/MPAN membranes.



**Fig. S28** The water permeance and  $\text{Na}_2\text{SO}_4$  rejection of the bilayer COF membranes (TpPa-SO<sub>3</sub>H/TpPa-SO<sub>3</sub>H/MPAN) under high pressure.

### 3. Tables

**Table S1.** Single layer COF membranes prepared with various monomer concentrations.

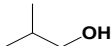
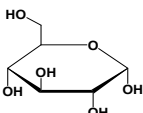
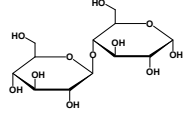
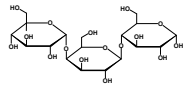
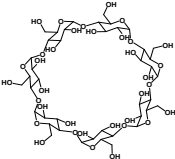
Membrane	PDA deposition time (h)	COF building monomers		
		PA concentration (mmol L <sup>-1</sup> )	DABA concentration (mmol L <sup>-1</sup> )	TFP concentration (mmol L <sup>-1</sup> )
TpPa(1.6)/MPAN	1	2.4	/	1.6
TpPa(6)/MPAN	1	9	/	6
TpPa(16)/MPAN	1	24	/	16
TpPa- SO <sub>3</sub> H(1.6)/MPAN	1	/	2.4	1.6
TpPa-SO <sub>3</sub> H(6)/MPAN	1	/	9	6
TpPa-SO <sub>3</sub> H(16)/MPAN	1	/	24	16



**Table S2.** Bilayer COF membranes prepared with different building monomers.

Membrane	Monomer concentration of COF			Monomer concentration of COF		
	bottom layer (mmol L <sup>-1</sup> )			top layer (mmol L <sup>-1</sup> )		
	PA	DABA	TFP	N <sub>2</sub> H <sub>4</sub>	DABA	TFP
TpHz(0.3)/TpPa(6)/MPAN	9.0	/	6.0	0.5	/	0.3
TpHz(1.1)/TpPa(6)/MPAN	9.0	/	6.0	1.7	/	1.1
TpHz(1.8)/TpPa(6)/MPAN	9.0	/	6.0	2.8	/	1.8
TpPa-SO <sub>3</sub> H(0.3)/TpPa(6)/MPAN	9.0	/	6.0	/	0.5	0.3
TpPa-SO <sub>3</sub> H(1.1)/TpPa(6)/MPAN	9.0	/	6.0	/	1.7	1.1
TpPa-SO <sub>3</sub> H(1.8)/TpPa(6)/MPAN	9.0	/	6.0	/	2.8	1.8
TpPa-SO <sub>3</sub> H(1.1)/TpPa-SO <sub>3</sub> H(1.6)/MPAN(1#)	/	2.4	1.6	/	1.7	1.1
TpPa-SO <sub>3</sub> H(1.1)/TpPa-SO <sub>3</sub> H(6)/MPAN(2#)	/	9.0	6.0	/	1.7	1.1
TpPa-SO <sub>3</sub> H(1.1)/TpPa-SO <sub>3</sub> H(16)/MPAN(3#)	/	24	16	/	1.7	1.1
TpPa-SO <sub>3</sub> H(0.3)/TpPa-SO <sub>3</sub> H(6)/MPAN	/	9.0	6.0	/	0.5	0.3
TpPa-SO <sub>3</sub> H(1.1)/TpPa-SO <sub>3</sub> H(6)/MPAN	/	9.0	6.0	/	1.7	1.1
TpPa-SO <sub>3</sub> H(1.8)/TpPa-SO <sub>3</sub> H(6)/MPAN	/	9.0	6.0	/	2.8	1.8

**Table S3.** Spherical neutral solutes with various molecular weights were selected for rejection tests.

Solutes	Molecular structure	Molecular Weight (Da)	Stokes radius (nm) <sup>2</sup>	Rejection (%)		
				TpHz/TpPa /MPAN	TpPa-SO <sub>3</sub> H/TpPa-MPAN	TpPa-SO <sub>3</sub> H/TpPa-SO <sub>3</sub> H-MPAN
Isobutanol		74.1	0.279	28.4±1.4	15.2±1.8	12.0±2.0
Glucose		180.1	0.358	60.3±2.1	53.5±2.2	45.4±2.6
Sucrose		342.2	0.462	83.2±3.2	78.0±3.1	77.0±3.3
Raffinose		504.4	0.584	89.0±2.7	86.1±1.2	85.1±1.7
β-cyclodextrin		1135.0	0.742	95.7±2.4	96.4±3.5	93.0±2.5

A series of spherical neutral solutes (isobutanol, glucose, sucrose, raffinose and β-cyclodextrin) were used to estimate the MWCO of the bilayer COF membranes. The rejection rate of 90% is regarded as the MWCO of the membrane. Each solution (100 ppm) was recycled by a home-made cross-flow apparatus at 5.0 bar. The rejection of the solutes was calculated based on equation (1) below, where the concentrations of the permeate ( $C_p$ ) and feed ( $C_f$ ) were measured by a total organic carbon (TOC, HTY-CT1000A, China) analyzer.

$$R = \frac{C_f - C_p}{C_f} \times 100\% \quad (1)$$

The average pore size of the bilayer COF membranes equals the Stokes radius of the spherical solute when the rejection rate reaches 50%. The aperture distribution is expressed as a probability density function and is derived from equation 2 below [1-3]:

$$\frac{dR(r_p)}{dr_p} = \frac{1}{r_p \ln \sigma_p \sqrt{2\pi}} \exp \left[ -\frac{(\ln r_p - \ln \mu_p)^2}{2(\ln \sigma_p)^2} \right] \quad (2)$$

where  $r_p$  refers to Stokes radius of the spherical neutral solute,  $\sigma_p$  refers to the ratio of solute radius at  $R_p=84.13\%$  to

50.0%,  $\mu_p$  refers to the average pore size calculated from Stokes radius of the solutes, respectively.

**Table S4.** Performance comparison of various COF membranes towards salt rejection.

Membrane	salt	Permeance (L m <sup>-2</sup> h <sup>-1</sup> MPa <sup>-1</sup> )	Rejection (%)	Operation pressure (MPa)	Ref.
FS-COM-1	Na <sub>2</sub> SO <sub>4</sub>	38.6	90-95	0.1	[4]
IISERP-COOH- COF1	Na <sub>2</sub> SO <sub>4</sub>	5	96.3	0.2	[5]
TpHz/PES	Na <sub>2</sub> SO <sub>4</sub>	40.5	58.3	0.4	[6]
COF-LZU1	Na <sub>2</sub> SO <sub>4</sub>	760	3.2	0.5	[7]
COF-LZU1/HPAN	Na <sub>2</sub> SO <sub>4</sub>	442	63.6	0.2	[8]
TpPa-1/HPAN	Na <sub>2</sub> SO <sub>4</sub>	418.5	15	0.1	[9]
COF-LZU1/PES	Na <sub>2</sub> SO <sub>4</sub>	800	10.8	0.2	[10]
ACOF-1	Na <sub>2</sub> SO <sub>4</sub>	5.6	95.7	0.3	[11]
TpHz/TpPa/MPA N	Na <sub>2</sub> SO <sub>4</sub>	16	85.4	0.5	
TpPa-SO <sub>3</sub> H/TpPa /MPAN	Na <sub>2</sub> SO <sub>4</sub>	39	95.7	0.5	This work
TpPa-SO <sub>3</sub> H/TpPa- SO <sub>3</sub> H/MPAN	Na <sub>2</sub> SO <sub>4</sub>	131	98.3	0.5	

## 4. References

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