Supporting Information

Amyloid-Like Assembly Converting Commercial Proteins to Water-Insoluble Adsorbents with Ultrahigh Adsorption Capacity and Ultrafast Adsorption Rate for Uranium Extraction

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Molecular dynamics simulations:

Structure optimization of BSA: All-atom crystal structure of BSA analyzed by Bujacz et al. was obtained from RCSB Protein Data Bank [S1-2]. The coordinates of cocrystallized water molecules were deleted before simulations and the disulfide bonds of BSA were broken to adapt to the role of TCEP. First, BSA was solvated in a rectangular box ($160 \times 81 \times 108 \text{ Å}^3$) using the SPC/E water model [S3] and neutralized by sodium cations. The conjugate gradient method [S4-5] was used to pre-equalize the system for eliminating the excessive stress in initial structures. After energy minimization, a 300 ps position-restrained NPT process was performed to relax the solvent around the protein at 298 K, during which the temperature and pressure coupling were performed using a vrescale thermostat [S6] and Berendsen barostat [S7]. A 200 ns production simulation was then performed under NPT ensemble with Parrinello-Rahman barostat [S8].

Simulated ions adsorption: BSA and counterions were extracted from the asequilibrated box and placed in the center of an empty rectangular box $(200 \times 100 \times 150 \text{ Å}^3)$. Then 38 Na⁺, 70 UO₂²⁺, 70 Fe³⁺, 70 Mg²⁺, 70 Cu²⁺, 70 Co²⁺, 70 Zn²⁺, 140 NO₃⁻, 808 Cl⁻, and 40000 water molecules were randomly placed in the box using Packmol program [S9] for providing homogeneous electrolyte solution. The parameters of energy minimization, position-restrained simulation, and production simulation are the same as described above.

During simulations, three-dimensional periodic boundary conditions (PBC) were used to avoid the influence of the box boundary during simulations. The cut-off distance of nonbonded interactions is 13 Å and the long-range electrostatic interactions were calculated by the particle-mesh Ewald (PME) method [S10]. All simulations were based on Amber99sb-ildn force field [S11] and carried out by using the GROMACS 2018.6 software package [S12]. Parameters of UO_2^{2+} and NO_3^{-} were taken from Shenoy et al. [S13], while other metal cations and anions were based on Merz's parameters [S14-16]. All visualization structures are provided by VMD 1.9.3 software [S17]. Radial distribution functions (RDF) $g(r)_{A-B}$ defined by equation S1:

$$g(\mathbf{r})_{A-B} = \frac{1}{\left\langle \rho_B \right\rangle} \frac{1}{N_A} \sum_{i=1}^{N_A} \sum_{j=1}^{N_B} \frac{\delta\left(r_{ij} - r\right)}{4\pi r^2}$$

was utilized to calculate the coordination number of $UO_2^{2^+}$, with $\langle \rho_B \rangle$ the particle density of type B averaged over all spheres around particles A. Notably, RDF calculated in this work is between the center of mass of $UO_2^{2^+}$ (B in the equation) and van der Waals surface of BSA (A in equation).

Adsorbent	Adsorption	Equilibrium	Conditions	Ref.
	capacity (mg/g)	time		
BSA-TCEP	1064	2 h	V = 2 L, m = 5 mg, FR = 1 L/h	This work
PIDO NF	951	550 min	V = 5 L, m = 15 mg, FR = 5 L/min	18
MS@PIDO/Alg	539.19	24 h	V = 5 L, m = 60 mg, FR = 5 L/min	19
AO-HNTs	295.66	13 h	V = 1 L, m = 10 mg, FR = 1 L/h, T =	20
			25 °C, pH = 8	
SSUP fiber	12.51	30 min	V = 30 mL, m = 10 mg	21
PAO hydrogel	718	600 h	V = 1 L, m = 10 mg	22
CP-PAO	350	160 h	V = 2 L, m = 10 mg	23
PPA@MISS-	307.3	60 min		24
PAF-1				
SMON-PAO	1158.16	30 h	V = 5 L, m = 10 mg	25
fibers				
UiO-66-3C4N	200	8 h	V = 1 L, m = 5 mg, pH = 8	26
H-ABP fiber	302	24 h	V = 1 L, m = 20 mg, pH = 8	27
UiO-66	340	14 h	V = 1 L, m = 10 mg, pH = 6	28
Zn ²⁺ /PAO	664	36 h	V = 2 L, m = 10 mg	29
RGO/PDA/oxime	605	6 h	V = 500 mL, m = 5 mg, T = 25 °C,	30
			pH = 5	

 Table S1. Comparison of adsorption performances with other state-of-the-art adsorbents.

V is the volume of the solution; m is the mass of the adsorbents; FR is flow rate; T is the temperature.

	Pse	eudo-secon	d-order model	Pseudo-first-order model		
Concentration	R ²	q _e	k_2 (g/mg/min)	\mathbb{R}^2	q _e	k_1 (min ⁻¹)
1 ppm	0.99509	314.47	7.2×10^{-4}	0.92283	310.77	0.065
4 ppm	0.99975	671.14	4.68×10^{-4}	0.90125	608.94	0.4521
8 ppm	0.99928	1074.61	$1.18 imes 10^{-4}$	0.84129	907.27	0.3112
16 ppm	0.98949	2036.37	1.25 × 10 ⁻⁵	0.9385	1727	0.0195

Table S2. Parameters of Pseudo-first-order and Pseudo-second-order adsorption kineticsfor BSA-based adsorbents.



Fig. S1 Self-made flow system for testing adsorption performance.



Fig. S2 The curvilinear regression between uranium concentration and absorbance.



Fig. S3 Self-made circulating water flow system for dynamic adsorption.



Fig. S4 FT-IR spectra of pristine BSA and BSA-based adsorbents.



Fig. S5 XPS spectra of pristine BSA and BSA-based adsorbents.

BON TOED		
BOATOLI	Element	Wt %
	С	60.77
	Ν	23.02
	0	16.25
P o	U	0
1		
0 1 2 3 4	5 6 7	8 9 keV
BSA-TCEP-U		
BSA-TCEP-U	Element	Wt %
BSA-TCEP-U	Element C	Wt % 30.65
BSA-TCEP-U	Element C N	Wt % 30.65 11.06
BSA-TCEP-U	Element C N O	Wt % 30.65 11.06 17.87
BSA-TCEP-U	Element C N O U	Wt % 30.65 11.06 17.87 40.42
BSA-TCEP-U	Element C N O U	Wt % 30.65 11.06 17.87 40.42

Fig. S6 EDS results of the adsorbents before and after adsorption.

Fig. S7 Adsorption capacity the protein-based adsorbents prepared by different proteins. The initial concentration of uranium is 8 ppm.

Fig. S8 Plots of t/q_t as a function of reaction time, showing the reactions followed by pseudo-second-order kinetic model.

Fig. S9 The kinetics of uranium adsorption with an initial concentration of 100 ppb.

Fig. S10 Adsorption capacity of the adsorbents at different temperatures. The initial concentration of uranium is 8 ppm.

Fig. S11 FT-IR spectra of the adsorbents before adsorption, after adsorption and after desorption, respectively.

Fig. S12 Mass changes of modified sponges with different deposition times.

Fig. S13 SEM images of MS (a) without and (a) with BSA coatings after contacting with *Escherichia coli*, respectively. SEM images of MS (c) without and (d) with BSA coatings after contacting with *Staphylococcus aureus*, respectively.

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