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## **Supplementary Materials**

## An *in situ* spontaneously-forming micelle-hydrogel system with programable release for sequential therapy of anaplastic thyroid cancer

Xi Yang<sup>1,#</sup>, Lingyun Zhang<sup>2,3,#</sup>, Lingnan Zheng<sup>1</sup>, Yan Wang<sup>4</sup>, Ling Gao<sup>1</sup>, Rui Luo<sup>4</sup>, Xinchao Li<sup>4</sup>, Changyang Gong<sup>4</sup>, Han Luo<sup>2\*</sup>, Qinjie Wu<sup>4,\*</sup>

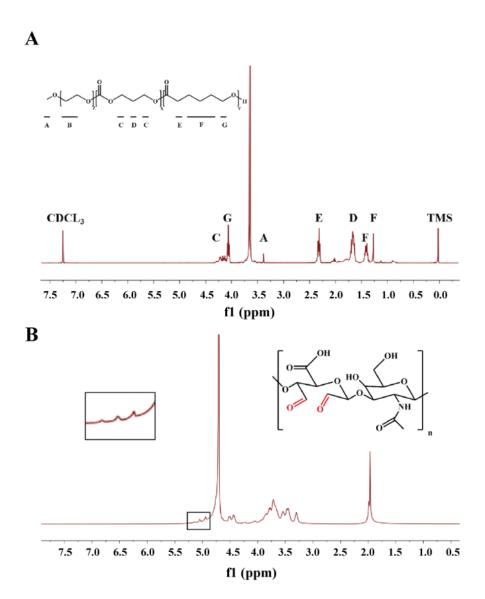
- <sup>1</sup> Department of Medical Oncology, Cancer Center, West China Hospital, Sichuan University, Chengdu 610041, P. R. China
- <sup>2</sup> Department of Thyroid and Parathyroid Surgery, Laboratory of thyroid and parathyroid disease, Frontiers Science Center for Disease-related Molecular Network, West China Hospital, Sichuan University, Chengdu 610041, P. R. China
- West China School of Medicine, Sichuan University, Chengdu 610041, P. R. China
  State Key Laboratory of Biotherapy and Cancer Center, West China Hospital, Sichuan University, Chengdu 610041, P. R. China
- \* To whom correspondence should be addressed (Q Wu and H Luo). E-mail: cellwqj@163.com and luohan-hx@scu.edu.cn.

<sup>&</sup>lt;sup>#</sup> These authors contributed equally to this work.

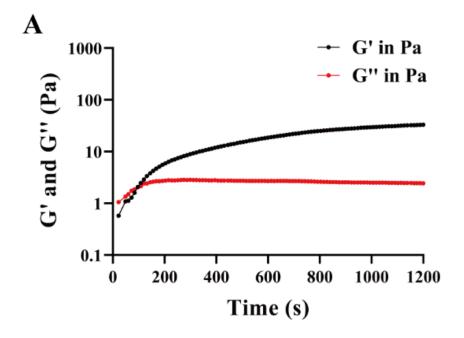
## Supplementary Table 1. Characteristics of ATC cell lines C643 and 8305C

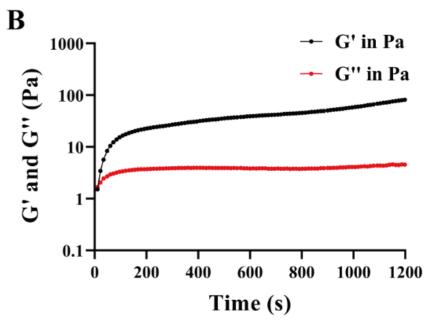
| Cell Line             | C643          | 8305C         |
|-----------------------|---------------|---------------|
| Year of establishment | 1987          | 1993          |
| Patient Age           | 76            | 67            |
| Patient Gender        | Male          | Male          |
| Derivation            | Primary Tumor | Primary Tumor |
| BRAF                  | -             | p.V600*       |
| HRAS                  | p.G13R*       | -             |
| TP53                  | p.R248Q       | p.R273C       |
| TERT promoter         | c124C>T       | c146C>T       |

<sup>\*</sup> Key genetic driver.



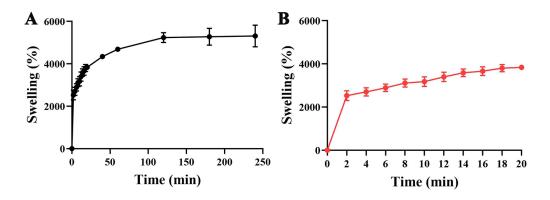
Supplementary Figure 1. <sup>1</sup>*H*-NMR spectrum of MPEG–P(CL-*ran*-TMC) in CDCl<sub>3</sub> (A) and AHA (B).



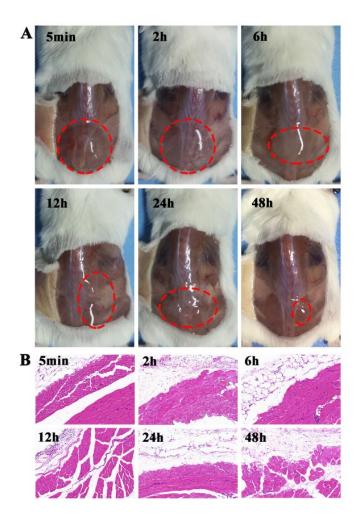


Supplementary Figure 2. Rheological analysis for the drug loaded NOCC/AHA hydrogel.

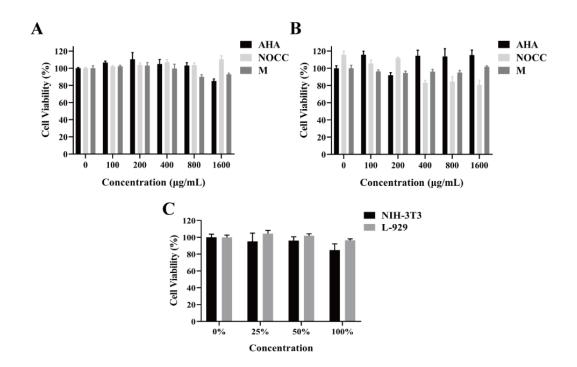
Time-dependence of storage modulus (G'), loss modulus (G") for the drug loaded NOCC/AHA hydrogel. (A) DDP-hydrogel; (B)  $P_m$ -hydrogel.



**Supplementary Figure 3**. (A&B) Swelling kinetics of NOCC/AHA hydrogel in the different time.

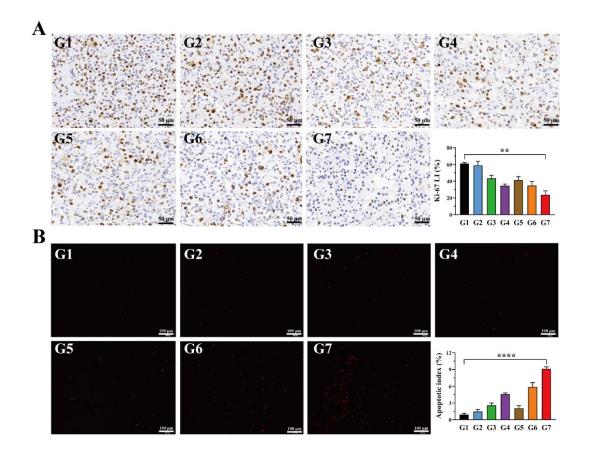


Supplementary Figure 4. *In vivo* degradation and biocompatibility of the NOCC-AHA hydrogel.(A) Gross observation of degradation assay in the different time; (B) Histological observations of biocompatibility assay in the different time.



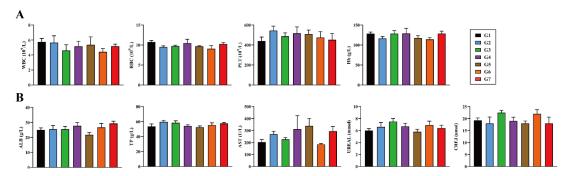
Supplementary Figure 5. Effect of AHA, NOCC, MPEG-P(CL-ran-TMC) and NOCC/AHA hydrogel extracts on cell viability measured by MTT assay.

(A) Cytotoxicity on NIH-3T3 and (B) L929 cells after 2-day incubation with AHA, NOCC or MPEG–P(CL-ran-TMC) at different concentrations; (C) Cytotoxicity on NIH-3T3 cells and L929 cells after 2-day incubation with the NOCC/AHA hydrogel extracts. Data were presented as mean  $\pm$  SD (n = 3).



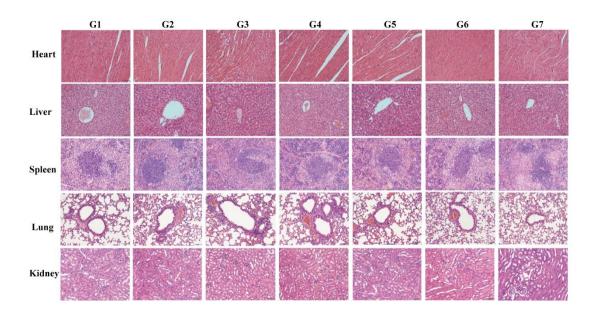
Supplementary Figure 6. Ki-67 immunohistochemical and TUNEL immunofluorescent staining of 8305C tumors.

(A) Representative Ki-67 immunohistochemical images of 8305C tumors and mean Ki-67 LI for each group (G1: NS i.t., G2: Micelle-hydrogel i.t., G3:  $P_m$  + DDP i.v., G4:  $P_m$  + DDP i.t., G5:  $P_m$ -hydrogel i.t., G6: DDP-hydrogel i.t., G7: iMHS i.t.). Scale bar = 50  $\mu$ m; (B) Representative TUNEL immunofluorescent images of 8305C tumors and mean apoptotic index for each group. Scale bar = 100  $\mu$ m. LI: labelling index.

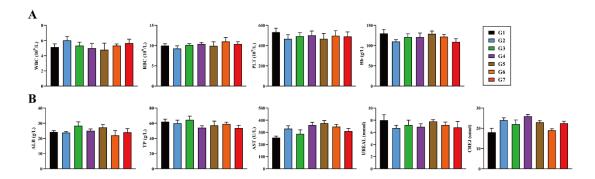


Supplementary Figure 7. Complete blood count and serum biochemistry study of the C643 subcutaneous tumor-bearing mice.

(A) Complete blood count including white blood cell (WBC), red blood cell (RBC), hemoglobin (Hb) and platelet (PLT) of each group; (B) Serum chemistry profile including total protein (TP), albumin (ALB), aspartate transaminase (AST), blood urea (UREAL), creatinine (CREJ) of each group.

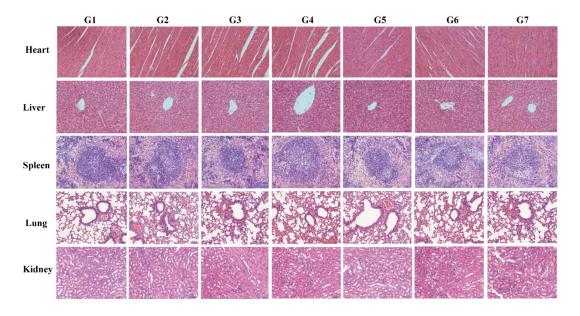


Supplementary Figure 8. Representative hematoxylin-eosin staining images of major organs (heart, lung, liver, spleen, and kidney) of each group in the C643 subcutaneous tumor-bearing mice.



Supplementary Figure 9. Complete blood count and serum biochemistry study of the 8305C subcutaneous tumor-bearing mice.

(A) Complete blood count including white blood cell (WBC), red blood cell (RBC), hemoglobin (Hb) and platelet (PLT) of each group; (B) Serum chemistry profile including total protein (TP), albumin (ALB), aspartate transaminase (AST), blood urea (UREAL), creatinine (CREJ) of each group.



Supplementary Figure 10. Representative hematoxylin-eosin staining images of major organs (heart, lung, liver, spleen, and kidney) of each group in the 8305C subcutaneous tumor-bearing mice.