

1 *Supporting information*

2 **A dual-functional fluorescence biosensor based on enzyme-**  
3 **involved catalytic hairpin assembly for the detection of**  
4 **APE1 and miRNA-21**

5 Xiaoyong Lu<sup>a</sup>, Dan Li<sup>b</sup>, Zewei Luo<sup>b,\*</sup> and Yixiang Duan<sup>a,\*</sup>

6 *<sup>a</sup>Research Center of Analytical Instrumentation, Key Laboratory of Bio-Resource and*  
7 *Eco-Environment, Ministry of Education, College of Life Sciences, Sichuan University,*  
8 *Chengdu 610064, Sichuan, P.R. China.*

9 *<sup>b</sup>Research Center of Analytical Instrumentation, Key Laboratory of Synthetic and*  
10 *Natural Functional Molecule Chemistry of Ministry of Education, College of*  
11 *Chemistry & Materials Science, Northwest University, Xi'an 710069, Shaanxi, P.R.*  
12 *China.*

13 *\*Corresponding author, E-mail: zwluo@nwu.edu.cn (Z. Luo), yduan@scu.edu.cn (Y.*  
14 *Duan)*

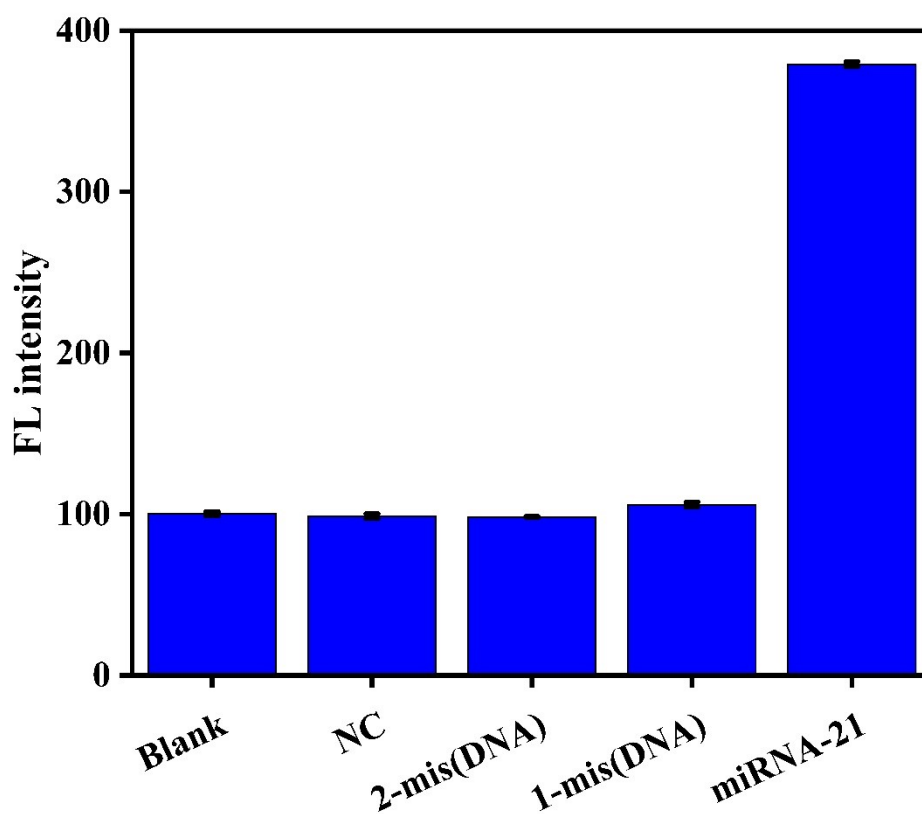
15

16

17

18 **Condition optimization**

19 For the optimization of APE1 inactivation time, 9.6 μL of 1 μM H1, 6 μL of 0.05  
20 U/μL APE1 and 82.8 μL of 1× CutSmart buffer were first incubated at 37 °C for 20  
21 minutes, and treated at 80 °C for different times. Then, 9.6 μL of 1 μM H2 and 12 μL  
22 of 500 nM miRNA-21 were added into the above system, and incubated at 37 °C for  
23 30 minutes. For the optimization of CHA reaction time, 9.6 μL of 1 μM H1, 6 μL of  
24 0.05 U/μL APE1 and 82.8 μL of 1× CutSmart buffer were first incubated at 37 °C for  
25 20 minutes, and treated at 80 °C for 5 minutes. Then, 9.6 μL of 1 μM H2 and 12 μL of  
26 500 nM miRNA-21 were added into the above system, and incubated at 37 °C for  
27 different times. To optimize the final concentration of H1 and H2, 9.6 μL of different  
28 concentrations of H1, 6 μL of 0.05 U/μL APE1 and 82.8 μL of 1× CutSmart buffer  
29 were first incubated at 37 °C for 20 minutes, and treated at 80 °C for 5 minutes. Then,  
30 9.6 μL of different concentrations of H2 and 12 μL of 500 nM miRNA-21 were added  
31 into the above system, and incubated at 37 °C for 50 minutes. All the resulting  
32 solutions were measured with fluorescent intensity immediately.



1 Figure S1 Selectivity of the biosensor for miRNA-21 against other miRNA-21 mimics (DNA 2 strands). From left to right: blank; NC, non-complimentary miRNA-21 mimics; 2-mis, 2 nt 3 mismatched miRNA-21 mimics; 1-mis, 1 nt mismatched miRNA-21 mimics.

Method	Target	Detection strategy	LOD	Linear range	
fluorescence	APE1	real time-qPCR	0.1 U/mL	0.1-5 U/mL	1
fluorescence	APE1	DNA nanoprobe	0.02 U/mL	0.02-2 U/mL	2
fluorescence	APE1	host-guest interaction	0.05 U/mL	0.05-5 U/mL	3
fluorescence	APE1	tetrahedral DNA nanostructure	0.01 U/mL	0.01-1 U/mL	4
colorimetric	APE1	G-quadruplex-hemin DNazymes	1.8 U/mL	2.2-22.5 U/mL	5
fluorescence	APE1	enzyme involved CHA	0.016 U/mL	0.05-1 U/mL	This work
fluorescence	miRNA-21	MnO <sub>2</sub> nanosheets and CHA	0.33 nM	1-50 nM	6
colorimetric	miRNA-21	graphene/gold-nanoparticle <sup>2</sup>	3.2 nM	10-980 nM	7
fluorescence	miRNA-21	graphene/gold - nanoparticle	4.5 nM	0-300 nM	8
fluorescence	miRNA-21	surface acoustic wave	0.19 nM	0.5-5 nM	9

electrochemical	miRNA-21	gold nanoparticle/reduced graphene oxide	12 nM	-	10
fluorescence	miRNA-21	enzyme involved CHA	0.25 nM	2.5-40 nM	This work

1  
2  
3  
4  
5

**Table S1** Comparison of different sensors for APE1 or miRNA-21 detection.

## 6 References

- 7 1 S. Fang, L. Chen and M. Zhao, *Anal Chem*, 2015, 87, 11952-11956.  
8 2 F. Li, Q. Xie, Y. Qin, C. Tong, B. Liu and W. Wang, *Anal Biochem*, 2021, 633,  
9 114394.  
10 3 M. Zhou, S. Qin, Z. Feng, C. Song, H. Zhang, W. Li, Q. Wang, J. Liu, J. Huang,  
11 X. Yang and K. Wang, *Chinese Chem Lett*, 2018, 29, 973-976.  
12 4 T. Zhou, R. Luo, Y. Li, J. Fan, Y. Hu, C. Tong, B. Liu and D. Li, *Sensor Actuat*  
13 *B-Chem*, 2020, 317, 128203.  
14 5 X. Huang, Z. He, K. Zhou, H. Zhi and J. Yang, *Analyst*, 2021, 146, 7476-7482.  
15 6 W. Ouyang, Z. Liu, G. Zhang, Z. Chen, L. Guo, Z. Lin, B. Qiu and G. Chen, *Anal*  
16 *Methods*, 2016, 8, 8492-8497.  
17 7 H. Zhao, Y. Qu, F. Yuan and X. Quan, *Anal Methods*, 2016, 8, 2005-2012.  
18 8 M. Hong, H. Sun, L. Xu, Q. Yue, G. Shen, M. Li, B. Tang and C.-Z. Li, *Anal*  
19 *Chim Acta*, 2018, 1021, 129-139.  
20 9 G. Celik Cogal, P. K. Das, G. Yurdabak Karaca, V. R. Bhethanabotla and A.  
21 Uygun Oksuz, *ACS Appl. Bio Mater*, 2021, 4, 7932-7941.  
22 10 H. Torul, E. Yarali, E. Eksin, A. Ganguly, J. Benson, U. Tamer, P.  
23 Papakonstantinou and A. Erdem, *Biosensors-Basel*, 2021, 11, 236.  
24  
25