

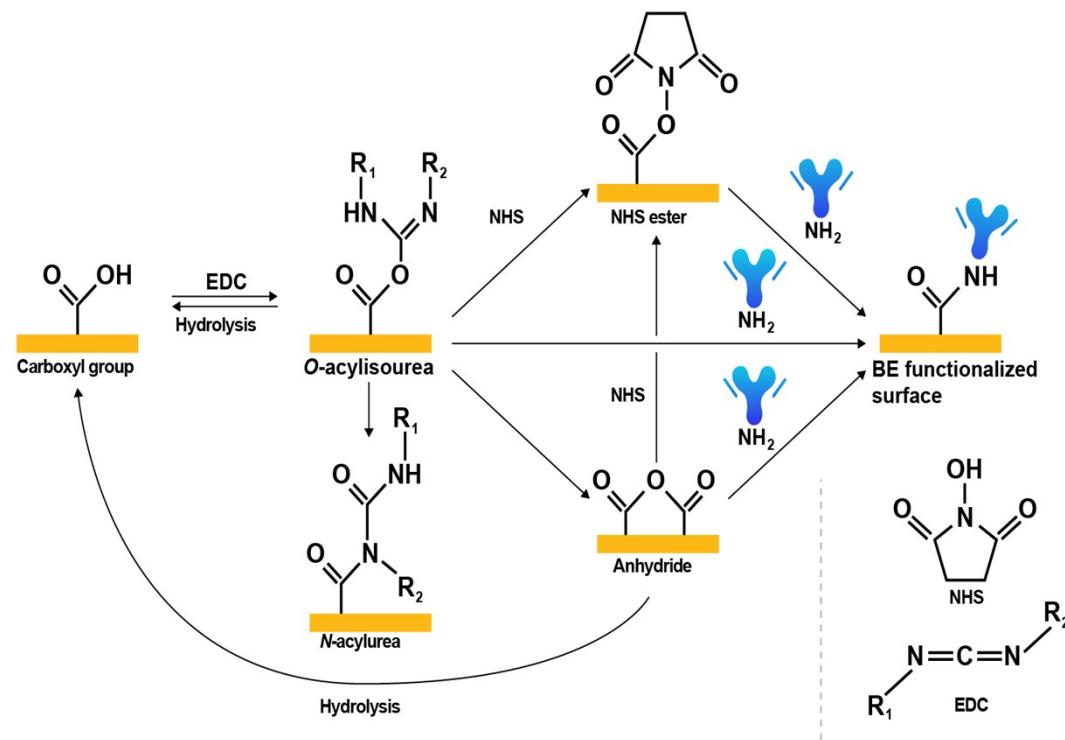
Supplementary material to

## **Biorecognition Antifouling Coatings in Complex Biological Fluids: A Review of Functionalization Aspects**

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**Figure S1:** Scheme of functionalization reactions using EDC/NHS.

**Table S1:** The effect of functionalization on fouling from bodily fluids for a selection of antifouling (ultra-low and low fouling) platforms relevant for the review

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionaliz ation [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Poly(carboxybetaine) platforms</b>									
pCBAA	HBP <sup>a]</sup> (100%)	10	2.73	EDC/NHS	hydrolysis (PBS, pH 7.4, 50min)	anti-Ecoli / anti-Salmonella	18.54	SPR	<sup>1</sup>
	HBP <sup>a]</sup> (100%)	7	< 0.2 (below LOD)	EDC/NHS	hydrolysis (buffer pH 8-9, 21 min)	anti-Salmonella	2.6 ± 1.1	SPR	<sup>2</sup>
	HBP (50%)						1.2 ± 0.4		
	HBP <sup>a]</sup> (100%)	10	~ 6	EDC/NHS	hydrolysis (10 mM Na <sub>2</sub> CO <sub>3</sub> + 300 mM NaCl, pH 10, 10 min)	anti-TSH	~ 9	SPRi	<sup>3</sup>
	HBS <sup>b]</sup> (100%)	10	~ 0 (below LOD)			anti-ALCAM			
						anti-HCG			
Two-layer hierarchical architecture of highly dense layer of pCBAA and second loose layer of pCBAA on top	HBS <sup>b]</sup> (100%)	10	1.6	EDC/NHS	hydrolysis (10 mM Na <sub>2</sub> CO <sub>3</sub> + 300 mM NaCl, pH 10, 10 min)	anti-TSH antibody	0.5	SPR	<sup>5</sup>
	HBP <sup>a]</sup> (100%)	10	< 5	EDC/NHS	hydrolysis (10 mM Na <sub>2</sub> CO <sub>3</sub> + 300 mM NaCl, pH 10, 10 min, followed by 10 mM sodium acetate, pH 5)	anti-TSH antibody	< 5	SPR	<sup>6</sup>
pCBAA hydrogel thin films with carboxybetaine diacrylamide crosslinker	HBS <sup>b]</sup> (100%)	10	4.7 ± 1.4 (extracted from plot)	EDC/NHS	hydrolysis followed by possible covalent interaction with GLY	anti-TSH antibody	5.9 ± 1.7 (extracted from plot)	SPR	<sup>7</sup>

<sup>a)</sup> Human Blood Plasma; <sup>b)</sup> Human Blood Serum

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionaliz ation [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Poly(carboxybetaine) platforms</b>									
p(CBMAA)	HBP <sup>a]</sup> (100%)	10	11.1 ± 2.8	EDC/NHS	hydrolysis (10 mM sodium borate + 10 mM imidazole + 10 mM NaCl, pH 8, 30min)	anti-Salmonella	25.4 ± 4.1	SPR	8
p(CBMAA 7.5 mol%-co-HPMAA)			0				2.6 ± 1.2		
p(CBMAA 15 mol%-co-HPMAA)	HBP <sup>a]</sup> (100%)	10	2.9 ± 0.8	EDC/NHS	hydrolysis (10mM sodium borate buffer +150 mM NaCl+10mM imidazole)	anti-Salmonella	11.6 ± 2.8	SPR	9
	HBP <sup>a]</sup> (100%)		4.9 ± 2.0	EDC/NHS	1M glycine	anti-Salmonella NH2-ON-probes	6.6 ± 3.6 4.9 ± 2.3	SPR	8
p(CBMAA 30 mol%-co-HPMAA)	HBS <sup>b]</sup> (10%)	10	0	EDC/NHS	hydrolysis (PBS, 90 min)	Hepatitis B surface antigen	0	SPR	10
	SLV10% <sup>c)</sup>	10	below LOD	EDC/NHS	hydrolysis (PBS, 90 min)	Hepatitis B surface antigen not measured	below LOD not measured	SPR	11
	SLV100% <sup>d)</sup>		below LOD						
	HBP <sup>a]</sup> (100%)		10.0 ± 3.5		hydrolysis (10 mM sodium borate + 10mM imidazole + 10mM NaCl, pH 8, 30min)	anti-Salmonella antibody	23.4 ± 3.9	SPR	8

<sup>a)</sup> Human Blood Plasma; <sup>b)</sup> Human Blood Serum; <sup>c)</sup> Saliva (10%, supernatant after centrifugation); <sup>d)</sup> Saliva (10%, supernatant after centrifugation)

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionaliz ation [ng/cm <sup>2</sup> ]	Probing method	Ref.
p(SBMAA 3 mol%-co-CBMAA 15 mol%-co-HPMAA 82 mol%)	HBP <sup>a)</sup> (100%)	10	5.3 ± 1.1	EDC/NHS	1M AEAA	anti-bacterial antibody	10.9 ± 3.2	SPR	12
PEG(3)-Dendrimer carboxybetaine	HBP <sup>a)</sup> (10%)	30	2.40	EDC/NHS	blocking with tris(hydroxymethyl)amino methane hydrochloride (0.5 M)	anti-ARG1 N- terminus antibody	~175 (extracted from plot)	SPRi	13
Catechol <sub>2</sub> -pCBMA <sub>2</sub>	HBP <sup>a)</sup> (100%)	10	8.9 ± 3.4	EDC/NHS	hydrolysis (buffer, pH 8-9, 21 min)	anti-ALCAM	9.5 ± 4.1	SPR	14
	HBS <sup>b)</sup> (100%)		11.0 ± 5.0				NOT SHOWN		
DOPA <sub>2</sub> -pCBMA <sub>2</sub>	HBS <sup>b)</sup> (100%)	10	11.7 ± 3.0	EDC/NHS	hydrolysis (10 mM sodium phosphate + 300 mM NaCl, pH 8.2, 10 min)	anti-ALCAM	55 (ALCAM naturally occurs in plasma)	SPR	15
	HBP <sup>a)</sup> (100%)		6.3 ± 0.9			not tested	not tested		
DOPA <sub>2</sub> -pCBMA <sub>2</sub>	FBS <sup>e)</sup> (centrifuge- filtered)	5	~2 (extracted from plot)	EDC/NHS	hydrolysis (10 mM borate buffer + 0.75 M NaCl, pH 8.2, 10 min)	mouse monoclonal anti-ALCAM antibody	~31 (extracted from plot)	Suspended Micro- channel Resonator	16
DOPA-pCBMA	HBP <sup>a)</sup> (100%)	15	~ 11 pm (extracted from plot)	EDC/NHS	hydrolysis (10 mM HEPES, 300 mM NaCl, pH 8.2)	Immunoglob ulin control antibodies	~54 pm (extracted from plot)	Silicon microring resonator	17
<b>Platforms with end hydroxyl group</b>									
pHEMA	HBP <sup>a)</sup> (100%)	10	16.2	DSC/DMAP in DMF	ethanolamine	anti-Ecoli O157 or anti- Salmonella	61.7	SPR	1

<sup>a)</sup> Human Blood Plasma; <sup>b)</sup> Human Blood Serum; <sup>e)</sup> Fetal Bovine Serum

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecogniti on element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Platforms with end hydroxyl group</b>									
p(HPMAA)	HBP <sup>a)</sup> (100%)	15	< 0.03 (SPR detection limit)	DSC/DMAP	hydrolysis (PBS, 24 h)	Ab against peptide- glycan- polysaccha- ride antigen unique to group <i>Gstreptococ cus</i>	< 0.03 (SPR detection limit)	SPR	18
<b>PEG platforms</b>									
Mixed SAMs of C11-EG6-COOH and C11-EG4-OH	HBP <sup>a)</sup> (100%)	10	147	EDC/NHS	ethanolamine	anti-Ecoli O157 or anti- Salmonella	135	SPR	1
			147			anti-Ecoli O157 or anti- Salmonella, BSA blocked surface	23		
DNA probe and (CH <sub>2</sub> ) <sub>15</sub> -COOH with covalently bound BSA	HBP <sup>a)</sup> (10%)	10	5	DNA hybridiza- tion	none	anti-ALCAM conjugated with DNA	2.9 ± 0.6	SPRi	19
						anti-hCG conjugated with DNA probe	3 ± 1		

<sup>a)</sup> Human Blood Plasma

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecogniti on element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>PEG platforms</b>									
poly(MeOEGMA-b-GMA) + NaN3	HBP <sup>a)</sup> (100%)	15	32 (extracted from plot)	catalyst- free SPAAC reaction	none	biotin conjugated to dibenzocycl ooctyne	88 (extracted from plot)	SPR	20
Structured polymer brush p(MeOEGMA-b-GMA) + NaN3 as antifouling and DBCO-biotin as functionalizable layer			88 (extracted from plot)			streptavidin and biotinylated mouse anti- IgG	25 (extracted from plot)		
methyl ether methacrylate (MeOEGMA) end-group nucleophilic substitution with azide			17 (extracted from plot)			biotin conjugated to dibenzocycl ooctyne	18 (extracted from plot)		
poly(MeOEGMA)-DBCO-biotin brush			18 (extracted from plot)			streptavidin and biotinylated mouse anti- IgG	23 (extracted from plot)		

<sup>a)</sup> Human Blood Plasma

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecogniti on element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
DNA tetrahedron probes	cell lysate (100%)	30	N/A (platform itself s functional)	none	none	DNA-probe	0	SPR	21
	WB <sup>f)</sup> (5%)						0		
	red cells (9.85e8 CFU/mL)						5.6 ± 1.4		
	HBP <sup>a)</sup> (100%)						8.0 ± 2.1		
	HBS <sup>b)</sup> (100%)						7.7 ± 1.2		
	HBP <sup>a)</sup> (100%)	30	N/A (platform itself s functional)	none	none	DNA-probe	41.5 ± 4.0	SPR	21
	HBS <sup>b)</sup> (100%)						68.2 ± 7.5		
Ionic liquid SAM (1-(carboxymethyl)-3-(mercaptododecyl)-imidazoliumbromide)	HBS <sup>b)</sup> (100%)	not defined	99	NHS/EDC	EA	anti-human IgG antibody	45	SPR	22
Biotynylated poly(acrylic acid)-(ethylene oxide)n monolayer adsorbed on top of multilayer poly(ethylene imine)-[(poly(styrene sulfonate)/poly(allylamine hydrochloride)] <sub>3</sub>	FBS <sup>e)</sup> (10%)	10	< 1	none	none	streptavidin - biotinylated antiovalbum in	<1	QCM	23
1-(carboxyethyl)-3-(12-mercaptododecyl)-1H-imidazolium bromide [(HS) <sup>12</sup> C <sub>12</sub> (COOH) <sup>5</sup> C <sub>5</sub> im] <sup>+</sup> Br <sup>-</sup> ionic liquid	Breast cancer cell lysates - MCF-7 and SK-BR-3 (50%)	20	Irreversible fouling 191 ± 45, real time nonspecific fouling 6 ± 4	NHS	cell lysate	polyclonal rabbit anti- HER2	Irreversible fouling not shown, real time nonspecific fouling 61 ± 8	SPR	24

<sup>a)</sup> Human Blood Plasma; <sup>b)</sup> Human Blood Serum; <sup>c)</sup> Fetal Bovine Serum; <sup>d)</sup> Whole Blood

(Continues)

**Table S1** (Continuation)

Platform	Medium (bodily fluid)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecogniti on element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
IgG-molecular imprinted hydrogel based on acrylamide, methylene N,N'-bis (acrylamide) and N-isopropylacrylamide crosslinked monomers	FBS <sup>e)</sup> (50%)	30	Data not shown for FBS <sup>e)</sup> (50%)	Imprinted hydrogel	none	Cavity for IgG detection	Charge-transfer resistance change of 2.063%	EIS	25
peptide CPPPNQNQNQNQDHWRGVV A mixed with CPPP-PNQNQNQNQ	FBS <sup>e)</sup> (100%)	30	N/A (platform itself is functional)	none	none	last part of peptide HWRGWVA selectively binds Fc region of human IgG	Signal suppression ~1%	DPV	26
peptide CPPPNQNQNQNQDHWRGVV A mixed with CPPP-PNQNQNQNQ	FBS <sup>e)</sup> (100%)	180	N/A (platform itself is functional)	non	none	last part of peptide HWRGWVA selectively binds Fc region of human IgG	Signal suppression of ~11%	DPV	26
Mixed SAM from peptide EKEKEKE-PPPPC, ATP binding aptamer 5'-SH-(CH <sub>2</sub> ) <sub>6</sub> -ACCTGGGGAGTATTGCGGAGG AAGGT-3' and 6-mercaptophexanol	HBP <sup>a)</sup> (10%)	not defined	N/A (platform itself is functional)	none	none	SH-aptamer self-assembled with antifouling peptide	Charge-transfer resistance change of ~10% (extracted from plot)	EIS	27
Mixed SAM from PEG, ATP binding aptamer 5'-SH-(CH <sub>2</sub> ) <sub>6</sub> -ACCTGGGGAGTATTGCGGAGG AAGGT-3' and 6-mercaptophexanol	HBP <sup>a)</sup> (10%)	not defined	N/A (platform itself is functional)	none	none	SH-aptamer simultaneously self-assembled with thiolated PEG	Charge-transfer resistance change of ~64 (extracted from plot)	EIS	28

<sup>a)</sup> Human Blood Plasma; <sup>e)</sup> Fetal Bovine Serum

**Table S2:** The effect of functionalization on fouling from foodstuff for a selection of antifouling (ultra-low and low fouling) platforms relevant for the review.

Platform	Medium (foodstuff)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Poly(carboxybetaine) platforms</b>									
pCBAA	milk (100%)	10min	0.83	EDC/NHS	hydrolysis (PBS. pH 7.4. 50min)	anti-Salmonella	1.81	SPR	<sup>1</sup>
	milk (100%)	10	0.2		hydrolysis (10 mM sodium borate buffer +150 mM NaCl+10 mM imidazole)	anti-Salmonella	4.1	SPR	<sup>9</sup>
	spinach (10%)		2.8				16.8		
	cucumber (10%)		0.25				8.1		
	hamburger (10%)		0				2.2		
	lettuce (10%)		0.14				4.3		
	cucumber (10%)	15	<0.2 (below LOD)	EDC/NHS	hydrolysis (PBS, pH 10)	anti-Ecoli	6.2 ± 1.7	SPR	<sup>29</sup>
	hamburger (10%)						3.4 ± 1.3		
	orange extract (100%)	10	1.45	EDC/NHS	hydrolysis (PBS pH 7.4, 50 min)	anti-Ecoli / anti-Salmonella	1.4	SPR	<sup>1</sup>
	tomato extract (100%)		1.05				0.06		
	cucumber extract (100%)		1.8				3.09		
p(CBMAA)	milk (100%)	10	0.6	EDC/NHS	hydrolysis (10 mM sodium borate buffer +150 mM NaCl + 10 mM imidazole)	anti-Salmonella	20.4	SPR	<sup>9</sup>
	spinach (10%)		2.74				12.2		
	cucumber (10%)		2.1				3.4		
	hamburger (10%)		2.2				4.5		
	lettuce (10%)		2.4				4.9		

(Continues)

**Table S2** (Continuation)

Platform	Medium (foodstuff)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionalizat ion [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Poly(carboxybetaine) platforms</b>									
p(CBMAA 7.5 mol%-co- HPMAA)	milk (100%)	10	0	EDC/NHS	hydrolysis (10 mM sodium borate buffer +150 mM NaCl + 10 mM imidazole)	anti-Salmonella	0.6	SPR	9
	spinach (10%)		0				1.1		
	cucumber (10%)		0				0		
	hamburger (10%)		0				0		
	lettuce (10%)		0				0		
p(CBMAA 15 mol%-co- HPMAA)	milk (100%)	10	0	EDC/NHS	hydrolysis (10 mM sodium borate buffer +150 mM NaCl + 10 mM imidazole)	anti-Salmonella	1.7	SPR	9
	spinach (10%)		0.5				3.4		
	cucumber (10%)		0.3				0.05		
	hamburger (10%)		0				0		
	lettuce (10%)		0.2				0		
p(CBMAA 30 mol%-co- HPMAA)	milk (100%)	10	0	EDC/NHS	hydrolysis (10 mM sodium borate buffer +150 mM NaCl + 10 mM imidazole)	anti-Salmonella	6.8	SPR	9
	spinach (10%)		1.2				4.2		
	cucumber (10%)		1.4				2.8		
	hamburger (10%)		0				2.5		
	lettuce (10%)		1.4				3.5		

(Continues)

**Table S2** (Continuation)

Platform	Medium (foodstuff)	Exposition time [min]	Fouling before functionalizatio n [ng/cm <sup>2</sup> ]	Method of functionaliz ation	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after functionaliz ation [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>Platforms with end hydroxyl group</b>									
pHEMA	orange (100%)	10	1.6	DSC/DMAP in DMF	EA	anti-Ecoli O157 or anti- Salmonella	10.5	SPR	1
	tomato (100%)		2.6				14.5		
	cucumber (100%)		5.3				26.7		
	milk (100%)		5.7				23.6		
	fresh-whole fat milk (100%)	30	~0	DSC/DMAP in DMF	hydrolysis (PBS, pH 7.4, 24h)	anti- Cronobacter (ex situ immobilizatio n)	~0	SPR	30
	whole-fat milk from powder (Laktino) (10%)		~0				~0		
	infant formulation from powder (Sunar™) (10%)		~0				~0		

(Continues)

**Table S2** (Continuation)

Platform	Medium (foodstuff)	Exposition time [min]	Fouling before funciona- lization [ng/cm <sup>2</sup> ]	Method of funciona- lization	Method of deactivation of residual intermediates (if relevant)	Biorecognitio n element	Fouling after funciona- lization [ng/cm <sup>2</sup> ]	Probing method	Ref.
<b>PEG platforms</b>									
Mix of HS-C11-EG6-COOH and HS-C11-EG4-OH	orange (100%)	10	21.12	EDC/NHS	ethanolamine	anti-Ecoli O157 or anti- Salmonella	2.6	SPR	1
	tomato (100%)		9.13				7.51		
	cucumber (100%)		51.9				18.52		
	milk (100%)		154.25				6.88		
	orange (100%)		21.12			anti-Ecoli O157 or anti- Salmonella. BSA blocked surface	2.28		
	tomato (100%)		9.13				2.64		
	cucumber (100%)		51.9				4.4		
	milk (100%)		154.25				6.24		
<b>Other platforms</b>									
Hyaluronic acid-grafted Au	soybean milk (10mg/mL. centrifuge- filtered 0.22um)	10	0.6	EDC/NHS	1M ethanolamine	anti-BSA	0.67	SPR	31
	cow milk (100%. centrifuge- filtered 0.22um)		9.8				17		

## References

1. H. Vaisocherova, V. Sevcu, P. Adam, B. Spackova, K. Hegnerova, A. D. Pereira, C. Rodriguez-Emmenegger, T. Riedel, M. Houska, E. Brynda and J. Homola, *Biosensors & bioelectronics*, 2014, **51**, 150-157.
2. H. Vaisocherova, W. Yang, Z. Zhang, Z. Cao, G. Cheng, M. Piliarik, J. Homola and S. Jiang, *Analytical Chemistry*, 2008, **80**, 7894-7901.
3. N. D. Brault, A. D. White, A. D. Taylor, Q. Yu and S. Jiang, *Analytical Chemistry*, 2013, **85**, 1447-1453.
4. W. Yang, H. Xue, W. Li, J. Zhang and S. Jiang, *Langmuir*, 2009, **25**, 11911-11916.
5. N. D. Brault, H. S. Sundaram, C.-J. Huang, Y. Li, Q. Yu and S. Jiang, *Biomacromolecules*, 2012, **13**, 4049-4056.
6. C.-J. Huang, Y. Li and S. Jiang, *Analytical Chemistry*, 2012, **84**, 3440-3445.
7. Y. N. Chou, F. Sun, H. C. Hung, P. Jain, A. Sinclair, P. Zhang, T. Bai, Y. Chang, T. C. Wen, Q. M. Yu and S. Y. Jiang, *Acta Biomaterialia*, 2016, **40**, 31-37.
8. H. Lísalová, E. Brynda, M. Houska, I. Víšová, K. Mrkvová, X. C. Song, E. Gedeonová, F. Surman, T. Riedel, O. Pop-Georgievski and J. Homola, *Analytical Chemistry*, 2017, **89**, 3524-3531.
9. H. Vaisocherová-Lísalová, F. Surman, I. Víšová, M. Vala, T. Špringer, M. L. Ermini, H. Šípová, P. Šedivák, M. Houska, T. Riedel, O. Pop-Georgievski, E. Brynda and J. Homola, *Analytical Chemistry*, 2016, **88**, 10533-10539.
10. T. Riedel, F. Surman, S. Hageneder, O. Pop-Georgievski, C. Noehammer, M. Hofner, E. Brynda, C. Rodriguez-Emmenegger and J. Dostálek, *Biosensors and Bioelectronics*, 2016, **85**, 272-279.
11. T. Riedel, S. Hageneder, F. Surman, O. Pop-Georgievski, C. Noehammer, M. Hofner, E. Brynda, C. Rodriguez-Emmenegger and J. Dostálek, *Analytical Chemistry*, 2017, **89**, 2972-2977.
12. M. Forinová, A. Pilipenco, I. Víšová, N. S. Lynn, J. Dostálek, H. Mašková, V. Hönig, M. Palus, M. Selinger, P. Kočová, F. Dyčka, J. Štěrba, M. Houska, M. Vrabcová, P. Horák, J. Anthi, C.-P. Tung, C.-M. Yu, C.-Y. Chen, Y.-C. Huang, P.-H. Tsai, S.-Y. Lin, H.-J. Hsu, A.-S. Yang, A. Dejneka and H. Vaisocherová-Lísalová, *ACS applied materials & interfaces*, 2021, **13**, 60612-60624.
13. R. D'Agata, N. Bellassai, M. C. Giuffrida, A. M. Aura, C. Petri, P. Kögler, G. Vecchio, U. Jonas and G. Spoto, *Talanta*, 2021, **221**, 121483.
14. C. Gao, G. Li, H. Xue, W. Yang, F. Zhang and S. Jiang, *Biomaterials*, 2010, **31**, 1486-1492.
15. N. D. Brault, C. Gao, H. Xue, M. Piliarik, J. Homola, S. Jiang and Q. Yu, *Biosensors and Bioelectronics*, 2010, **25**, 2276-2282.
16. M. G. von Muhlen, N. D. Brault, S. M. Knudsen, S. Jiang and S. R. Manalis, *Analytical Chemistry*, 2010, **82**, 1905-1910.
17. J. T. Kirk, N. D. Brault, T. Baehr-Jones, M. Hochberg, S. Jiang and D. M. Ratner, *Biosensors & bioelectronics*, 2013, **42**, 100-105.
18. C. Rodriguez-Emmenegger, E. Brynda, T. Riedel, M. Houska, V. Šubr, A. Bologna Alles, E. Hasan, J. E. Gautrot and W. T. S. Huck, *Macromolecular Rapid Communications*, 2011, **32**, 952-957.
19. M. Piliarik, M. Bocková and J. Homola, *Biosensors and Bioelectronics*, 2010, **26**, 1656-1661.
20. V. Parrillo, A. de los Santos Pereira, T. Riedel and C. Rodriguez-Emmenegger, *Analytica Chimica Acta*, 2017, **971**, 78-87.
21. W. Nie, Q. Wang, L. Zou, Y. Zheng, X. Liu, X. Yang and K. Wang, *Analytical Chemistry*, 2018, DOI: 10.1021/acs.analchem.8b02686.
22. M. Ratel, A. Provencher-Girard, S. S. Zhao, J. Breault-Turcot, J. Labrecque-Carbonneau, M. Branca, J. N. Pelletier, A. R. Schmitzer and J.-F. Masson, *Analytical Chemistry*, 2013, **85**, 5770-5777.
23. J. Davila, D. Toulemon, T. Garnier, A. Garnier, B. Senger, J.-C. Voegel, P. J. Mésini, P. Schaaf, F. Boulmedais and L. Jierry, *Langmuir*, 2013, **29**, 7488-7498.

24. A. Aubé, S. Campbell, A. R. Schmitzer, A. Claing and J.-F. Masson, *Analyst*, 2017, **142**, 2343-2353.
25. M. Cui, Y. H. Gong, M. G. Du, K. Wang, T. D. Li, X. L. Zhu, S. Wang and X. L. Luo, *Sens. Actuator B-Chem.*, 2021, **337**, 9.
26. N. Liu, N. Hui, J. J. Davis and X. Luo, *ACS sensors*, 2018, DOI: 10.1021/acssensors.8b00318.
27. G. Wang, X. Su, Q. Xu, G. Xu, J. Lin and X. Luo, *Biosensors and Bioelectronics*, 2018, **101**, 129-134.
28. G. Wang, Q. Xu, L. Liu, X. Su, J. Lin, G. Xu and X. Luo, *ACS applied materials & interfaces*, 2017, **9**, 31153-31160.
29. H. Vaisocherová-Lísalová, I. Víšová, M. L. Ermini, T. Špringer, X. C. Song, J. Mrázek, J. Lamačová, N. Scott Lynn, P. Šedivák and J. Homola, *Biosensors and Bioelectronics*, 2016, **80**, 84-90.
30. C. Rodriguez-Emmenegger, O. A. Avramenko, E. Brynda, J. Skvor and A. Bologna Alles, *Biosensors and Bioelectronics*, 2011, **26**, 4545-4551.
31. X. Liu, R. Huang, R. Su, W. Qi, L. Wang and Z. He, *ACS applied materials & interfaces*, 2014, **6**, 13034-13042.