

## **Electronic Supplementary Information (ESI) for**

### **Photoelectrochemical sandwich immunoassay of CYFRA21-1 based on In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub> type-II heterojunction and CdS quantum dots-polydopamine nanospheres labeling**

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## Reagents and instruments

FTO conductive glass was commercially obtained from Zhuhai Kaivo Optoelectronic Technology Co., Ltd. Sodium tungstate ( $\text{Na}_2\text{WO}_4 \cdot 2\text{H}_2\text{O}$ ), indium nitrate ( $\text{In}(\text{NO}_3)_3 \cdot 4.5\text{H}_2\text{O}$ ), cadmium chloride ( $\text{CdCl}_2 \cdot 2.5\text{H}_2\text{O}$ ), citric acid, CS, sodium sulfide ( $\text{Na}_2\text{S} \cdot 9\text{H}_2\text{O}$ ), ascorbic acid (AA), hydrochloric acid (HCl) and ammonia water ( $\text{NH}_3 \cdot \text{H}_2\text{O}$ ) were commercially obtained from the National Chemical Reagents Company of China. Thioglycolic acid (TGA), N-hydroxysuccinimide (NHS), 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC), dopamine hydrochloride (DA), BSA and GLD were supplied from Sigma-Aldrich. CYFRA21-1 and CYFRA21-1 antibody (Ab) were purchased from Shanghai Linc-Bio Science Co., Ltd. Human immunoglobulin G (IgG), human interleukin-6 (IL-6), procalcitonin (PCT) and cardiac troponin I (cTnI) were purchased from Beijing Bioss Co., Ltd. All other chemicals are of analytical grade or higher quality. The detection and washing buffers used for the immunoassay were 0.1 M phosphate buffered solution (PBS, pH 7.4, 0.1 M  $\text{KH}_2\text{PO}_4$ - $\text{K}_2\text{HPO}_4$ ) and 0.01 M PBS (pH 7.4), respectively. All solutions in the experiments were prepared with Milli-Q ultrapure water (Millipore,  $\geq 18 \text{ M}\Omega \text{ cm}$ ). Normal human serum samples were kindly donated by Hunan Normal University Hospital.

The PEC experiments were performed on a CHI660E electrochemical workstation (Shanghai Chenhua Instrument Co., Ltd.) equipped with a xenon lamp (300-1000 nm wavelength range and  $100 \text{ mW cm}^{-2}$  optical power density). Cyclic voltammetry (CV), electrochemical impedance spectroscopy (EIS) and Mott-Schottky barrier experiments were conducted on a CHI660E electrochemical workstation using a three-electrode system, with FTO or its modified electrode as the working electrode (effective surface area  $0.25 \text{ cm}^2$ ), a KCl-saturated calomel electrode (SCE) as the reference electrode, and a platinum disk

electrode (3 mm in diameter) as the counter electrode. The potentials are reported versus SCE, except otherwise specified. Scanning electron microscopy (SEM) characterizations were performed on a FEI Helios NanoLab 600i scanning electron microscope equipped with an Oxford energy dispersive X-ray spectroscopy (EDX) detector for elemental analysis. A TECNAI F-30 transmission electron microscopy (TEM) instrument was used for morphology characterization. An Ultima IV X-ray diffraction (XRD) instrument was used for crystal analysis. X-ray photoelectron spectroscopy (XPS) was recorded on a Thermo Scientific K-Alpha instrument. UV-vis diffuse reflectance spectra (DRS, with BaSO<sub>4</sub> as a reference) and ultraviolet-visible (UV-vis) absorption spectra were obtained on a UV-2600i UV-vis spectrophotometer. Fourier transform infrared (FT-IR) spectrum was collected on a IRTracer 100 FT-IR spectrophotometer (KBr pellet method). CS film-thickness was evaluated on an Accrion/Nanofilm EP4SE ellipsometer.

### **Synthesis of polydopamine nanospheres (PDA NSs) and CdS QDs**

The self-polymerization of dopamine was carried out in a water/ethanol/ammonia mixed solution to synthesize PDA NSs with a size of ca. 160 nm.<sup>1</sup> Briefly, 2 mL  $\text{NH}_3 \cdot \text{H}_2\text{O}$ , 40 mL ethanol and 90 mL ultrapure water were mixed with gentle stirring for 30 min at 30 °C. 0.5 g dopamine hydrochloride was dissolved in 10 mL ultrapure water, which was then quickly injected into the above water/ethanol/ammonia mixture. The color of the dispersion immediately turned light yellow and finally dark brown. After continuous stirring for 24 h, the precipitate was collected by centrifugation and washed three times with ultrapure water.

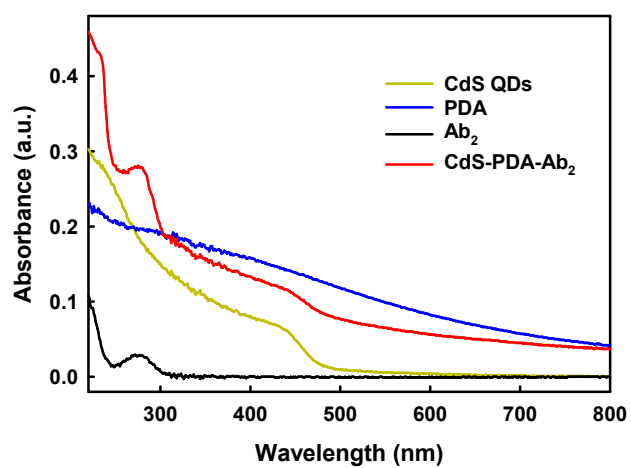
TGA-modified CdS QDs were synthesized according to the reported method.<sup>2</sup> 250  $\mu\text{L}$  TGA was added to a three-necked flask containing 50 mL 10 mM aqueous  $\text{CdCl}_2$  under nitrogen atmosphere and solution-stirring condition, and the solution pH was adjusted to pH 11 with 1.0 M NaOH, followed by adding 5 mL 0.1 M aqueous  $\text{Na}_2\text{S}$ . After nitrogen treatment for 4 h, the CdS QDs were deposited with acetone and redispersed in ultrapure water to prepare a 1 mg  $\text{mL}^{-1}$  CdS QDs dispersion. The CdS QDs dispersion was stored in a 4 °C refrigerator away from light.

**Quantitative verification of the presence of  $\text{In}_2\text{O}_3$  and  $\text{WO}_3$  on the  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  electrode from the element atomic percentage data in Fig. 1D**

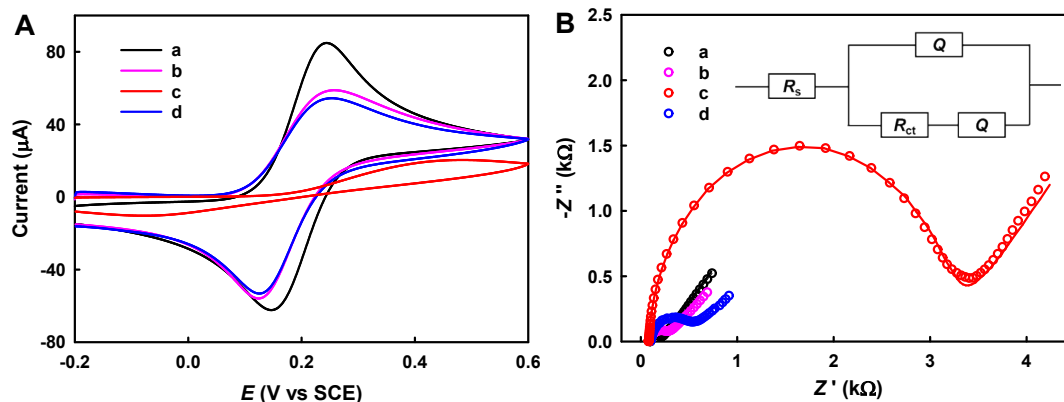
FTO conductive glass consists of  $\text{SiO}_2$  substrate and F-doped  $\text{SnO}_2$ . In the EDX of the  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  electrode shown in Fig. 1D, the atomic percentages are 14.01% for Sn element and 10.51% for Si element, and thus the calculated total atomic percentage of O element in  $\text{SnO}_2$  and  $\text{SiO}_2$  should be  $14.01\% \times 2 + 10.51\% \times 2 = 49.04\%$ . The total atomic percentage of oxygen element is 66.68% in Fig. 1D. After deducting the calculated total atomic percentage of O element in  $\text{SnO}_2$  and  $\text{SiO}_2$  (49.04%), the atomic percentage of the remaining O element, which should logically be equivalent to the total atomic percentage of O element in  $\text{In}_2\text{O}_3$  and  $\text{WO}_3$ , is  $x_{\text{O}(-)} = 66.68\% - 49.04\% = 17.64\%$ .

In Fig. 1D, the atomic percentages are 2.60% for In element and 3.95% for W element, thus the calculated total atomic percentage of O element in  $\text{In}_2\text{O}_3$  and  $\text{WO}_3$  should be  $x_{\text{O}(\text{In,W})} = 2.60\% \times 3/2 - 3.95\% \times 3 = 15.75\%$ .

Considering the complexity of the actual experimental factors (such as the interference of possible trace impurities), the above calculated  $x_{\text{O}(-)}$  and  $x_{\text{O}(\text{In,W})}$  agree well with each other, indicating that  $\text{WO}_3$  and  $\text{In}_2\text{O}_3$  indeed coexist on the  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  electrode.

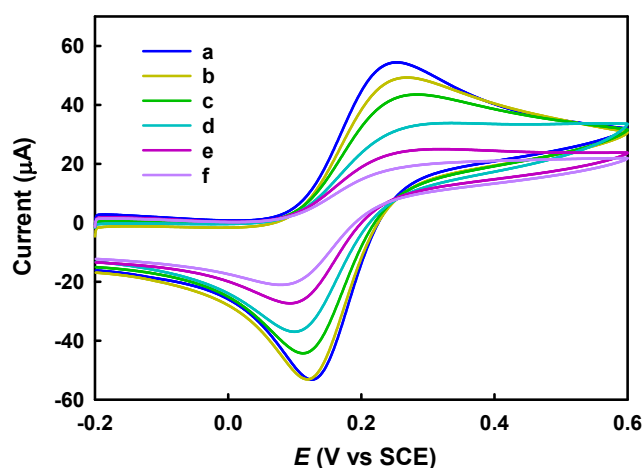


**Fig. S1** UV-vis absorption spectra of CdS QDs, PDA, Ab<sub>2</sub> and CdS-PDA-Ab<sub>2</sub>.

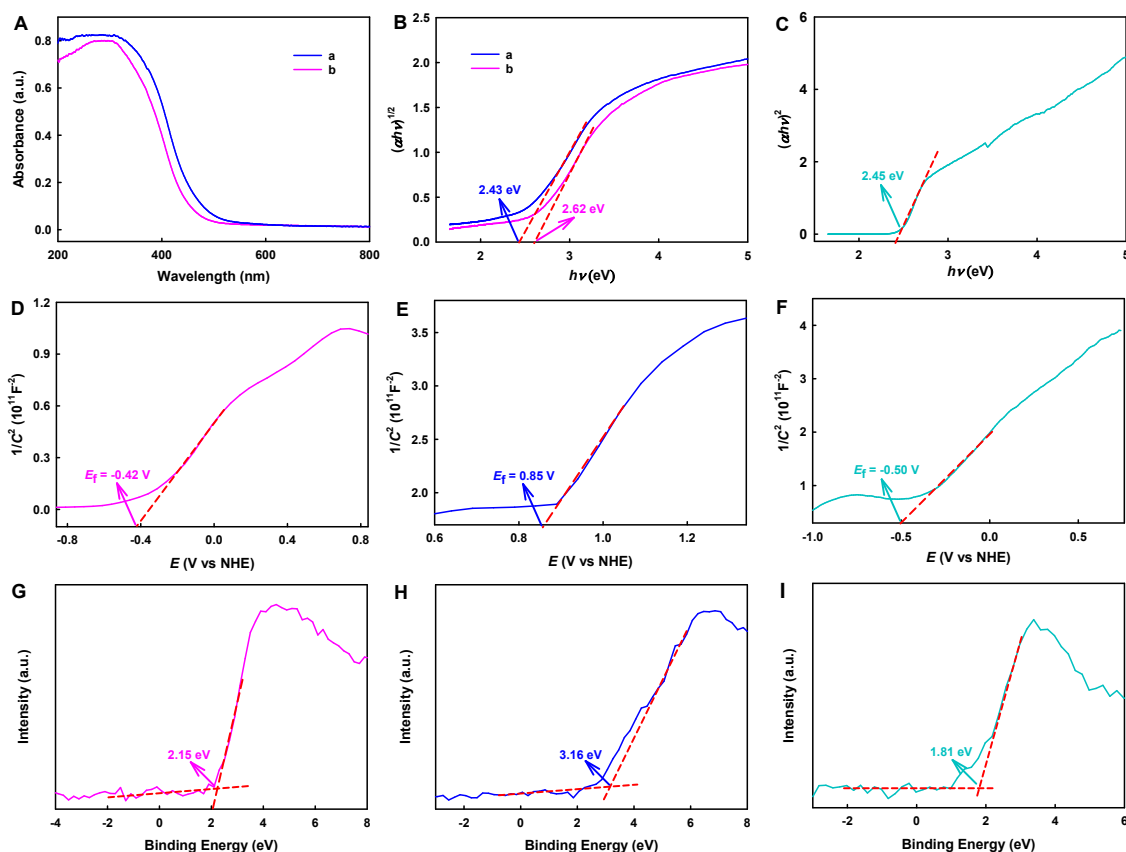


**Fig. S2** CV (A, 50 mV s<sup>-1</sup> scan rate) and EIS (B) curves of FTO (a), In<sub>2</sub>O<sub>3</sub>/FTO (b), WO<sub>3</sub>/FTO (c) and In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO (d) electrodes. Both CV and EIS were performed in 0.01 M PBS (pH 7.4) containing 2.0 mM K<sub>4</sub>[Fe(CN)<sub>6</sub>] and 0.1 M Na<sub>2</sub>SO<sub>4</sub>. EIS experiment: 100 kHz~0.01 Hz, 0.20 V bias (the formal potential of [Fe(CN)<sub>6</sub>]<sup>3-/4-</sup> redox couple), 200 s quiet time (to ensure a concentration ratio of 1:1 ([Fe(CN)<sub>6</sub>]<sup>3-</sup> versus [Fe(CN)<sub>6</sub>]<sup>4-</sup>) near the electrode surface). Circles: experimental, curves: fitted to the Randles equivalent circuit (inset). The CV oxidation peak current values of the electrodes are ordered as FTO (84.6 μA) > In<sub>2</sub>O<sub>3</sub>/FTO (59.0 μA) > In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO (54.4 μA) > WO<sub>3</sub>/FTO (20.6 μA).





**Fig. S3** CV curves on  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (a),  $\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (b),  $\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (c),  $\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (d),  $\text{Ag}/\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (e) and  $\text{BSA}/\text{CdS-PDA-Ab}_2/\text{Ag}/\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  (f) electrodes in 0.01 M PBS (pH 7.4) containing 2.0 mM  $\text{K}_4[\text{Fe}(\text{CN})_6]$  and 0.1 M  $\text{Na}_2\text{SO}_4$ . Scan rate:  $50 \text{ mV s}^{-1}$ . The CV oxidation peak current values are ordered as  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $54.4 \mu\text{A}$ ) >  $\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $48.8 \mu\text{A}$ ) >  $\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $43.2 \mu\text{A}$ ) >  $\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $33.9 \mu\text{A}$ ) >  $\text{Ag}/\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $25.2 \mu\text{A}$ ) >  $\text{BSA}/\text{CdS-PDA-Ab}_2/\text{Ag}/\text{BSA}/\text{Ab}_1/\text{GLD}/\text{CS}/\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  ( $19.7 \mu\text{A}$ ).

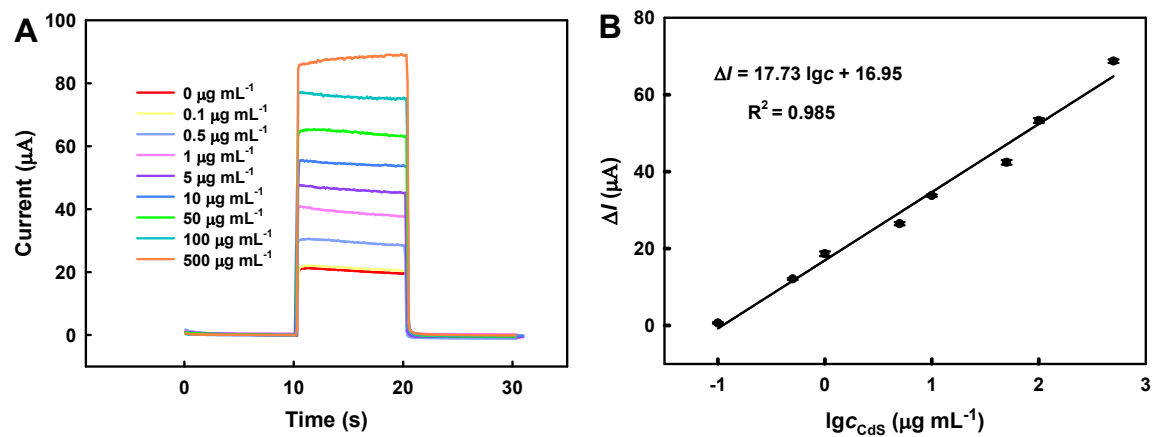


**Fig. S4** UV-vis diffuse reflectance spectra (A) and Tauc plots (B) of  $\text{WO}_3$  (a) and  $\text{In}_2\text{O}_3$  (b). Tauc plot of CdS QDs (C). Mott-Schottky plots of  $\text{In}_2\text{O}_3$  (D),  $\text{WO}_3$  (E) and CdS QDs (F). XPS-VB images of  $\text{In}_2\text{O}_3$  (G),  $\text{WO}_3$  (H) and CdS QDs (I). Mott-Schottky experiments: 1000 Hz frequency, 0.2 M aqueous  $\text{Na}_2\text{SO}_4$ .

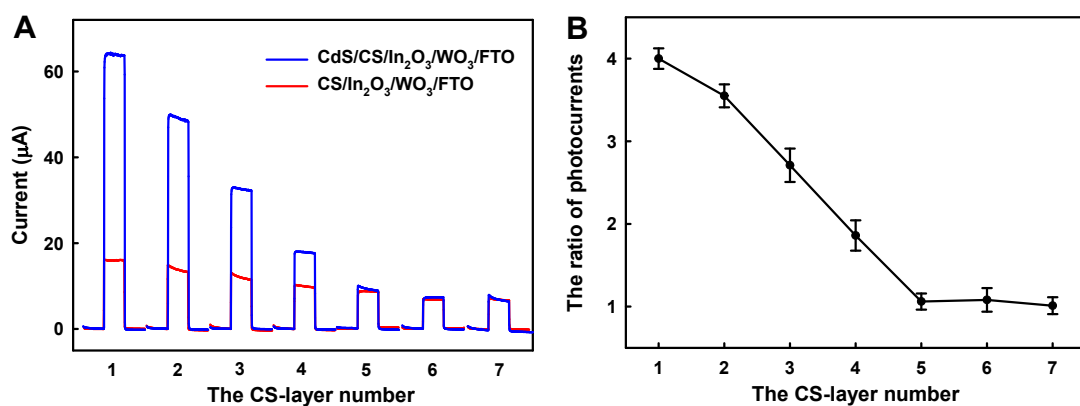
Here, the forbidden band widths ( $E_g$ ) of  $\text{In}_2\text{O}_3$ ,  $\text{WO}_3$  and CdS QDs can be calculated according to the Tauc equation,  $\alpha h\nu = A (h\nu - E_g)^{m/2}$ , where  $\alpha$  is the absorption coefficient or the absorbance,  $\nu$  in  $\text{s}^{-1}$  is the incident photon frequency,  $E_g$  in eV is the band gap,  $h$  is Planck's constant ( $6.63 \times 10^{-34}$  J s),  $A$  is a constant, and  $m$  is a variable determined by the transition characteristics of the semiconductor ( $m=1$  for a direct semiconductor and  $m = 4$  for an indirect semiconductor).<sup>3</sup> As shown in Fig. S4B and C, the  $E_g$  values of  $\text{In}_2\text{O}_3$  and  $\text{WO}_3$  as indirect semiconductors is obtained as 2.62 eV and 2.43 eV from the plot of  $(\alpha h\nu)^{1/2}$  versus  $h\nu$ , respectively, and the  $E_g$  of CdS QDs as a direct semiconductor is obtained as 2.45 eV

according to the plot of  $(ah\nu)^2$  versus  $h\nu$ .<sup>4,5</sup>

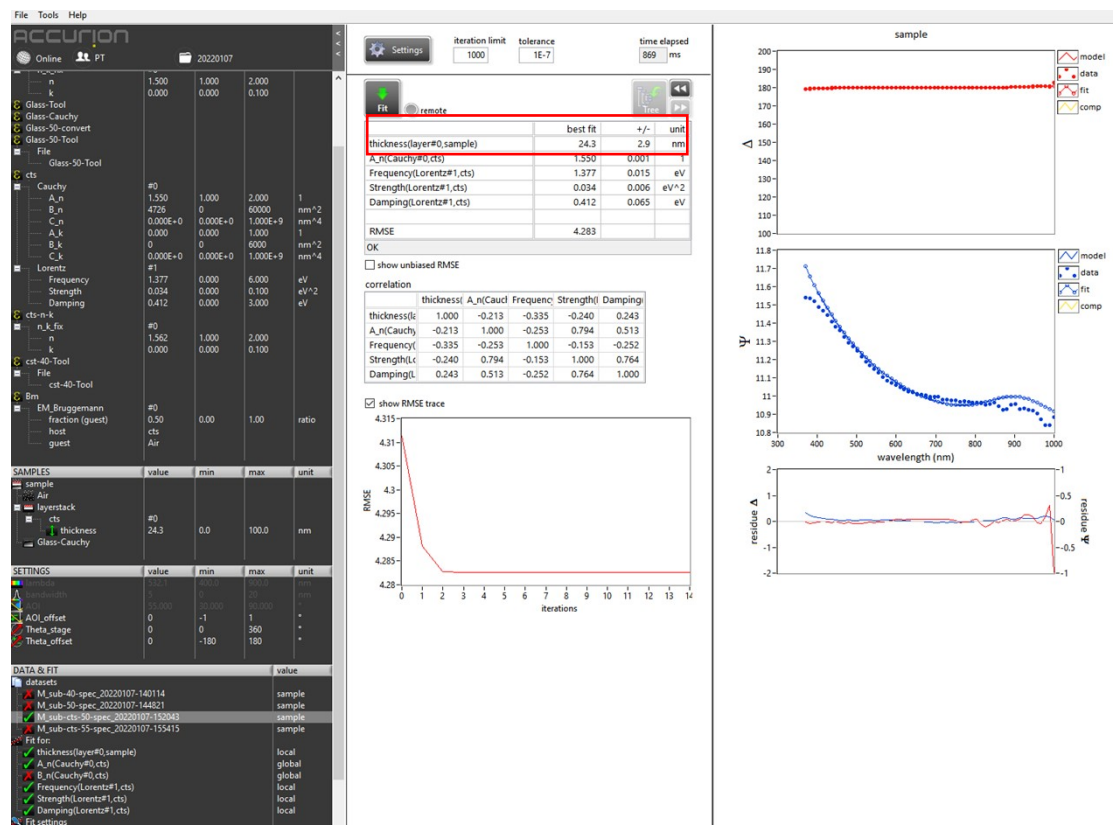
Taking  $\text{In}_2\text{O}_3$  as an example, we give the calculation details for the band structures. First, the  $E_g$  of  $\text{In}_2\text{O}_3$  is calculated from the UV-vis DRS of  $\text{In}_2\text{O}_3$  and the Tauc equation. As shown by curve b in Fig. S4A,  $\text{In}_2\text{O}_3$  has corresponding absorbance values at wavelengths from 200 to 800 nm. The frequency ( $\nu$ ) corresponding to a wavelength from 200 nm to 800 nm is equal to the speed of light (ca.  $3 \times 10^8$  m  $\text{s}^{-1}$ ) divided by the wavelength. Since  $\text{In}_2\text{O}_3$  is an indirect semiconductor,  $m$  is 4. In the Tauc equation,  $A$  is a constant whose value has no effect on the calculation result. Therefore, the absorbance value ( $\alpha$  value),  $\nu$  value,  $h$  ( $6.63 \times 10^{-34}$  J s) and  $m$  ( $= 4$ ) obtained at each wavelength can be substituted into the Tauc equation to obtain the Tauc plot of  $\text{In}_2\text{O}_3$  (Fig. S4B). For example, we obtain  $\alpha = 0.400$  and  $\nu = 7.50 \times 10^{14}$   $\text{s}^{-1}$  at 400 nm, thus  $h\nu = (6.63 \times 10^{-34}$  J s)  $\times$  ( $7.5 \times 10^{14}$   $\text{s}^{-1}$ ) =  $4.97 \times 10^{-19}$  J = 3.11 eV ( $x$ -axis), and  $(ah\nu)^{1/2} = (0.400 \times 3.11$  eV) $^{1/2} = 1.12$  ( $y$ -axis) in Fig. 4B.  $E_g$  can be obtained by making the intersection of the tangent and the abscissa. Second, according to the literature,<sup>6,7</sup> the  $E_{\text{VB}}$  of  $\text{In}_2\text{O}_3$  can be obtained by combining the XPS-VB method and linear extrapolation, as shown in Fig. S4G. Finally, substituting the  $E_g$  value of 2.62 eV and the  $E_{\text{VB}}$  value of 2.15 eV into the equation  $E_g = E_{\text{VB}} - E_{\text{CB}}$  gives  $E_{\text{CB}} = 2.15$  eV - 2.62 eV = -0.47 eV. The calculations of the band structures of  $\text{WO}_3$  and CdS QDs are similar to those of  $\text{In}_2\text{O}_3$ .



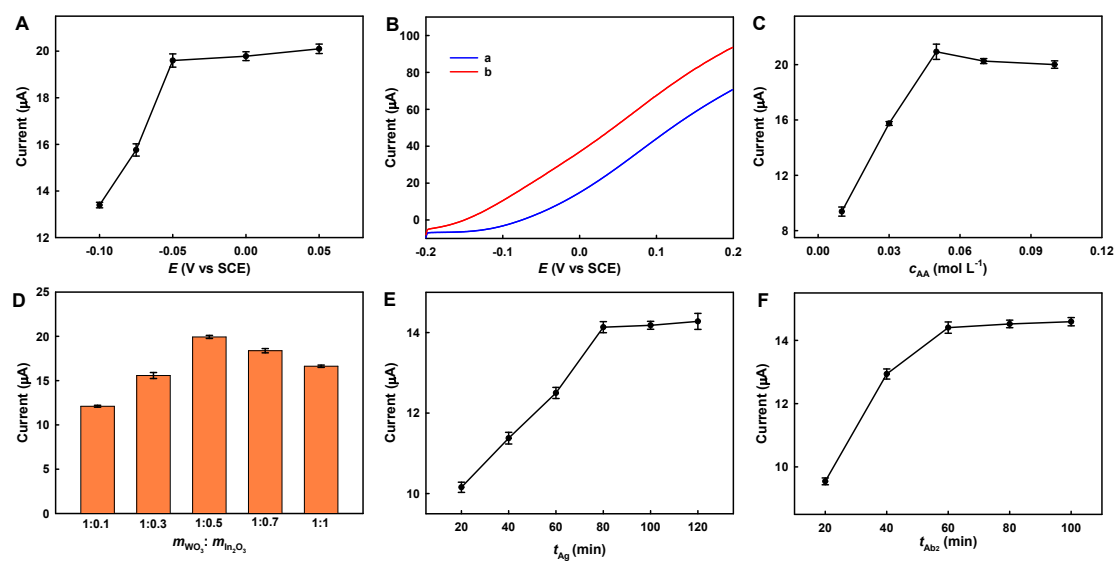
**Fig. S5** (A) Photocurrent responses on  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$  electrode after cast-coating  $20 \mu\text{L}$  CdS QDs at different concentrations. (B) Corresponding calibration curve. Light on and off at a 10-s interval,  $-0.05$  V bias,  $0.1$  M PBS (pH 7.4) containing  $0.05$  M AA.



**Fig. S6** Photocurrent of dried CdS/CS/In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO and dried CS/In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO electrodes versus number of CS layers (A). Photocurrent ratio of dried CdS/CS/In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO to dried CS/In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO versus number of CS layers (B). Cast-coating 6 μL of 1% acetic acid containing 0.05% CS and/or 20 μL of 0.5 mg mL<sup>-1</sup> CdS QDs for each modification, -0.05 V bias, 0.1 M PBS (pH 7.4) containing 0.05 M AA.



**Fig. S7** Nonlinear fitting of the ellipsometric data to obtain the thickness of a single-layer CS film.

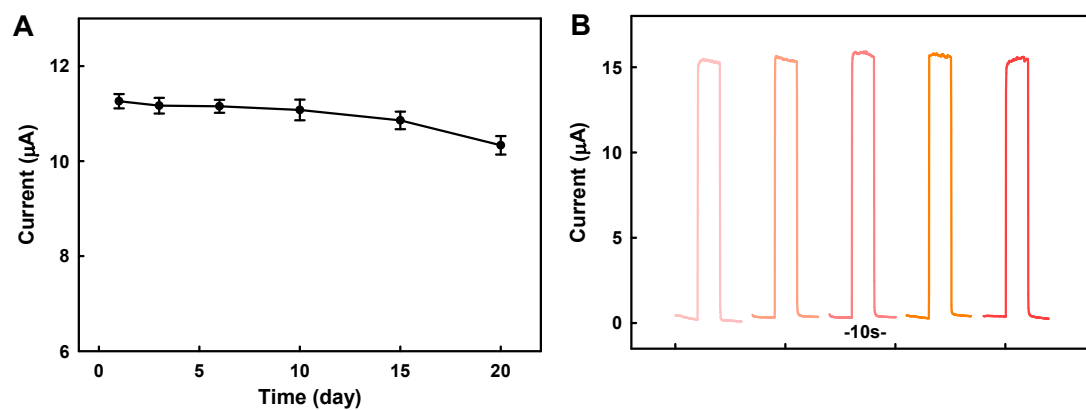


**Fig. S8** Optimization of bias potential (A). LSV curves (B, 50 mV s<sup>-1</sup>) on In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO electrode under light off (a) or light on (b) condition. Optimization of AA concentration (C). Photocurrent response of In<sub>2</sub>O<sub>3</sub> and WO<sub>3</sub> versus WO<sub>3</sub>-In<sub>2</sub>O<sub>3</sub> mass ratio (D). Optimization of 100 pg mL<sup>-1</sup> Ag incubation time (E) and BSA/CdS-PDA-Ab<sub>2</sub> incubation time (F). Solution: 0.1 M PBS (pH 7.4) containing AA. Bias = -0.05 V, AA concentration (c<sub>AA</sub>) = 0.05 M, WO<sub>3</sub>-In<sub>2</sub>O<sub>3</sub> mass ratio (m<sub>WO<sub>3</sub></sub>:m<sub>In<sub>2</sub>O<sub>3</sub></sub>) = 1:0.5, CYFRA21-1 incubation time (t<sub>Ag</sub>) = 80 min, and BSA/CdS-PDA-Ab<sub>2</sub> incubation time (t<sub>Ab<sub>2</sub></sub>) = 60 min, except for the parameter under optimization.

As shown in Fig. S8A, the photocurrent of the In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO electrode increases with the positive shift of the bias from -0.1 V to -0.05 V, but the photocurrent is almost stable with the further positive shift of the bias to 0.05 V. Fig. S8B shows the LSV curves of In<sub>2</sub>O<sub>3</sub>/WO<sub>3</sub>/FTO electrode under the condition of switching on or off the lamp. When the potential is greater than -0.05 V, the dark background current generated solely by the electrolysis becomes larger and larger, resulting in the decrease of signal-to-background ratio. Therefore, a bias of -0.05 V (vs SCE) is selected for PEC measurements. In Fig. S8C, the

photocurrent increases when the concentration of ascorbic acid ( $c_{AA}$ ) is changed from 0.01 M to 0.05 M, and the photocurrent is almost unchanged with the further increase of  $c_{AA}$ , indicating that 0.05 M AA is sufficient to clear the holes of  $\text{In}_2\text{O}_3/\text{WO}_3/\text{FTO}$ . Therefore, 0.05 M AA is selected as the optimal holes scavenger concentration. As shown in Fig. S8D, the maximum photocurrent can be obtained when the mass ratio of  $\text{WO}_3$  to  $\text{In}_2\text{O}_3$  ( $m_{\text{WO}_3}:m_{\text{In}_2\text{O}_3}$ ) is 1:0.5, probably because other ratios have more electron-hole recombination centers. Furthermore, optimizing the incubation time of Ag and BSA/CdS-PDA- $\text{Ab}_2$  ( $t_{\text{Ag}}$  and  $t_{\text{Ab}_2}$ ) is beneficial to improving the accuracy of the sensor and the experimental efficiency. In Fig. S8E, the photocurrent increases with the increase of  $t_{\text{Ag}}$ , and the photocurrent is almost unchanged when  $t_{\text{Ag}}$  is greater than 80 min, indicating that 80 min is sufficient for the specific Ag- $\text{Ab}_1$  immuno-recognition. As shown in Fig. S8F, the photocurrent also increases with the increase of  $t_{\text{Ab}_2}$ , and the photocurrent appears stable after 60 min. Therefore, 80 min and 60 min are selected as the optimal incubation time for Ag and BSA/CdS-PDA- $\text{Ab}_2$ , respectively.





**Fig. S9** Long-term stability of PEC immuno-electrode after storage at 4 °C in a refrigerator (A, 1 pg mL<sup>-1</sup> Ag). Reproducibility of five PEC immuno-electrodes fabricated in batch (B, 500 pg mL<sup>-1</sup> Ag). -0.05 V bias, 0.1 M PBS (pH 7.4) with 0.05 M AA.

**Table S1** Performance comparison for the analysis of CYFRA21-1

Technique	Linear range (ng mL <sup>-1</sup> )	LOD (pg mL <sup>-1</sup> )	Ref.
Lateral flow immunoassay	1.3 ~ 480	160	8
Electrochemiluminescence	0.001 ~ 350	0.3	9
Electrochemistry	0.0005 ~ 50	0.39	10
Electrochemistry	0.25 ~ 800	100	11
Electrochemistry	0.01 ~ 50	10	12
Electrochemistry	0 ~ 12	240	13
Electrochemistry	0.000625 ~ 0.01	0.625	14
PEC	0.0005 ~ 50	0.16	15
PEC	0.0001~ 4	0.03	16
PEC	0.01 ~ 100	2.5	17
PEC	0.0001 ~ 50	0.056	This work

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