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Au nanoclusters-decorated WO₃ nanorods for ultrasensitive photoelectrochemical sensing of Hg²⁺

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Instrumentation

Transmission electron microscopy (TEM) studies were performed on a TECNAI F-30 highresolution transmission electron microscope. Scanning electron microscopy (SEM) studies were performed on a Hitachi S4800 scanning electron microscope. X-ray diffraction (XRD) pattern was obtained on a PANalytical X'pert Pro X-ray diffractometer using Cu Ka radiation, operating at 40 kV and 30 mA, with a scan rate of 4° min⁻¹. X-ray photoelectron spectroscopy (XPS) measurement was investigated using a PHI QUANTUM 2000 X-ray photoelectron spectroscopic instrument. Ultraviolet-Visible (UV-Vis) spectra were obtained on U3310 UV-Vis spectrometer (Hitachi, Japan). Fourier transform infrared (FTIR) spectrum was collected on Nicolet Nexus 670 Fourier transform infrared spectrometer. All electrochemical experiments were carried out on the CHI760E electrochemical workstation. All photoelectrochemical experiments were carried out on electrochemical workstation (ZENNIUM, ZAHNER-elecktrik GmbH & Co. KG, Germany) equipped with a controlledintensity modulated-photospectroscopy setup (CIMPS, PP211, ZAHNERelecktrik GmbH & Co. KG, Germany). A white light lamp (WLC02, ZAHNER-elecktrik GmbH & Co. KG, Germany) with a visible-light filter (400-700 nm) was used as the light source.

Chemicals

Sodium tungstate, sodium chloride, mercury chloride, copper chloride, iron chloride, zinc chloride, cobalt chloride, potassium chloride, chromium chloride, calcium chloride, cadmium chloride, nickel chloride, hydrochloric acid and ascorbic acid (AA) were purchased from Sinophem Chemical Reagents Co. LTD. (Shanghai, China). Glutathione was purchased from Aladdin. HAuCl₄·3H₂O, 4-chloro-1-naphthol were purchased from Sigma Aldrich. All

reagents are analytical pure and and used without further purification. Milli-Q ultrapure water

(Millipore, $\geq 18 \text{ M}\Omega \text{ cm}$) was used throughout.



Fig. S1 FT-IR spectrum of AuNCs.

b **a** 0.30 С WO₃ WO₃ AuNCs 0.25 AuNCs Absorbance / a.u. 0.20 (ahv)^{1/2}/a.u. (ahv)? / a.u. 0.15 0.10 0.05 E_g=2.77 eV E_a=2.90 eV 0.00 200 2.2 2.4 2.6 2.8 3.0 3.2 3.4 3.6 3.8 300 400 500 600 700 800 1.5 2.0 3.0 3.5 4.0 2.5 Wavelength / nm hv/eV hv/eV d 10 е f 0.6 WO₃ – AuNCs/ITO – ITO -1.12 -1 8 LUM 0.4 Potential / V vs NHE C⁻² / F⁻²x10¹⁰ Current / mA 6 0 HOMO 1.78 V -0.26 СВ AuNCs 0.2 4 1 -WO₃ 0.0 2 1.78 HON -0.162 V 2 0 -0.2 2.51 -0.6 -0.4 -0.2 0.0 0.2 0.4 0.6 0 2 -1 1 3. Potential V vs NHE Potential / V vs NHE

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Fig. S2 (a) UV-vis diffuse reflectance spectra of WO₃ and AuNCs. (b) Plot of $(\alpha hv)^{1/2}$ versus hv for WO₃. (c) $(\alpha hv)^2$ versus hv for AuNCs. (d) Mott-Schottky plot of WO₃. (e) Cyclic voltammogram of AuNCs in degassed acetonitrile solution containing 0.1 M tetrabutyl ammonium perchlorate. (f) Illustration of energy-levels of AuNCs and WO₃.



Fig. S3 Effects of weight ratios of WO₃ to AuNCs in WO₃/AuNCs on transient photocurrents of the prepared WO₃/AuNCs.



Scheme S1 (a) Charge transfer pathway of WO₃/AuNCs/ITO after 4-CN + H_2O_2 incubation. (b) Charge transfer pathway of WO₃/AuNCs/ITO after Hg^{2+} incubation and then 4-CN + H_2O_2 incubation.



Fig. S4 Transient photocurrents of $WO_3/AuNCs/ITO$ after the incubation with different concentrations of Hg^{2+} .



Fig. S5 Effects of (a) Hg^{2+} inhibition time, (b) 4-CN concentration, (c) H_2O_2 concentration, and (d) precipitation reaction time on photocurrent responses for detecting 10 nM Hg^{2+} .



Fig. S6 Storage stability of the PEC sensor for detection of 10 nM $\mathrm{Hg^{2+}}$.

Technique	Linear range	Detection limit	Ref.
	(pM)	(pM)	
Electrochemistry	1.0×10 ⁴ -1.0×10 ⁶	5.8×10 ³	1
Electrochemistry	1.0×10 ² -1.3×10 ⁵	30	2
Electrochemistry	5.0×10 ² -5.0×10 ⁶	2.3×10 ²	3
Colorimetry	3.0×10 ³ -5.0×10 ⁵	3.0×10 ³	4
Colorimetry	2.0×10 ⁴ -2.0×10 ⁷	5.0×10 ³	5
Fluorescence	10-1.0×10 ⁵	4.5	6
Fluorescence	100-1.0×10 ⁵	65	7
Fluorescence	1.0×10 ³ -5.0×10 ⁶	3.5×10 ²	8
Fluorescence	1.0×10 ⁵ -6.0×10 ⁶	3.8×10 ³	9
Fluorescence	0-1.6×10 ³	1.1×10 ²	10
Surface-enhanced Raman scattering	1-10 ³	0.4	11
Surface-enhanced Raman scattering	100-10 ⁴	30	12
Photoelectrochemistry	1.0-2.0×10 ³	1.0	13
Photoelectrochemistry	1.0-5.0×10 ³	0.5	14
Photoelectrochemistry	1.0-5.0×10 ⁴	0.2	This work

Table S1. The comparison of the determination of Hg²⁺ in the literature.

Original	Added	Detected	Recovery (%)	RSD
(pM)	(pM)	(pM)		
52.00	10.00	58.30	94.0	3.3
	100.0	146.0	96.0	4.7
	1000	1092	104	1.1

Table S2. Results of determination of Hg²⁺ in a river water sample.

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