## **Supporting Information**

## Template-free assembly of Cu,N-codoped hollow carbon nanospheres

## as low-cost and highly efficient peroxidase nanozymes

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**Fig. S1.** XPS survey scans of CuNHCN calcinated at 600°C, 700 °C and 800 °C. Insets: contents of C, N, O and Cu.



Fig. S2. SEM images of the (a) hollow Cu-PmPD nanospheres and (b) CuNHCN.



Fig. S3. FT-IR spectrum of the hollow Cu-PmPD nanospheres



**Fig. S4.** FT-IR spectrum of the interior parts of the Cu-PmPD nanospheres (the interior parts was obtained according to the literature<sup>1,2</sup>. Briefly, the interior parts was dissolved in the reaction solvent, which could be obtained through centrifugation and freeze drying)



Fig. S5. TEM images of the (a) NC and (b)CuN



**Fig. S6.** N<sub>2</sub> adsorption-desorption isotherm curves and pore size distribution curves of (a) CuC,(b) NC, (c) CuNHCN.



**Fig. S7.** Effects of (a) pH, (b) temperature, (c) concentration and (d) stability of CuNHCN under optimized reaction conditions. Error bar shows the standard deviation of three independent measurements.



Fig. S8 The storage performance of CuNHCN under optimized reaction conditions



Fig.S9. Influence of the mass ratio of mPD to CuCl<sub>2</sub> on the peroxidase-like catalytic performance



Fig.S10. Influence of the temperature on the peroxidase-like catalytic performance of CuNHCN



**Fig. S11.** The influence of different concentrations of ROS scavengers (NaN<sub>3</sub>, PBQ and AA) on the  $A_{652}$  of the CuNHCN/TMB/H<sub>2</sub>O<sub>2</sub> system. Reaction condition: 0.15 mM TMB, 0.1 mM H<sub>2</sub>O<sub>2</sub>, 25 mg L<sup>-1</sup> catalyst, pH 3.5 acetate buffer(0.2 M), 4 min reaction at 40 °C.



Fig. S12. The influence of different substrates (5 mM fructose, 5 mM sucrose, 5 mM lactose, 5 mM Maltose or 0.5 mM glucose) on the  $A_{652}$  of the glucose oxidase/TMB/CuNHCN system. Error bar shows the standard deviation of three independent measurements.



**Fig. S13.** The influence of different substrates (metal ion(K<sup>+</sup>, Na<sup>+</sup>,Ca<sup>2+</sup>) (5 mM), sulfite (5 mM), nitrite (0.5 mM), chloride(0.5 mM), alanine (Ala) (5 mM), glutamic acid (Glu) (5 mM), glycine (Gly) (5 mM), lysine (Lys) (5 mM), tyrosine (Tyr) (5 mM), Cysteine (Cys) (0.5 mM), glutathione (GSH) (0.5 mM), Cys+NEM, GSH+NEM, uric acid (UA) (0.5 mM), vitamin E (0.5 mM) and AA (0.5 mM)) on the A<sub>652</sub> of the H<sub>2</sub>O<sub>2</sub>/TMB/CuNHCN system. Error bar shows the standard deviation of three independent measurements.

Material —	$K_{\rm m}$ (mM)		V <sub>max</sub> (10 <sup>-8</sup> M/s)		Defe
	TMB	$H_2O_2$	TMB	$H_2O_2$	· Keis
Au NPs /Graphene-	0.38	26.4	18.3	15.41	[3]
Au NPs/PVP-GNs	2.63	104	13.04	11.98	[3]
Cu/Au/Pt TNPs	0.15	2.34	7.33	136.5	[4]
β-CD-Cu-NCs	0.543	32.87	43.4	45.2	[4]
Hollow CuS nanocubes	1.62	0.94	16.64	2.55	[6]
Au loaded nanoporous Fe <sub>2</sub> O <sub>3</sub>	0.049	138 5	5 882	4 770	[7]
nanozymes	0.049	150.5	5.002	4.770	[,]
Pt hollow nanodendrites	0.81	6.9	1.2	9.9	[8]
Fe <sub>3</sub> O with aptamer	0.088	30.24	0.406	0.342	[9]
Au-Ni/g-C <sub>3</sub> N4	0.16	4.47	2.34	6.16	[10]
Co <sub>3</sub> O <sub>4</sub> nanocrystals	0.49	1.90	16	12.7	[11]
HRP	0.434	3.70	10.0	8.71	[12]
CuNHCN	0.0655	0.918	5.85	42.84	This work

**Table S1** Comparison of the CuNHCN based on colorimetry method for the determination of glucose and AA.

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